Design, Synthesis, and Biological Activities of Vibsanin B Derivatives: A New Class of HSP90 C-terminal Inhibitors

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Compd.	MCF-7	SMMC-7721	A-549	HL-60	SW480	
6	2.85 ± 0.03	3.25 ± 0.14	3.46 ± 0.21	2.64 ± 0.06	2.65 ± 0.17	
7	0.24 ± 0.01	0.21 ± 0.01	0.14 ± 0.01	0.62 ± 0.01	-	
8	4.51 ± 0.09	4.63 ± 0.23	3.75 ± 0.12	3.05 ± 0.23	2.95 ± 0.16	
9	12.58 ± 0.43	8.67 ± 0.14	12.35 ± 0.93	3.15 ± 0.07	12.89 ± 0.24	
10	16.08 ± 0.10	14.88 ± 0.63	20.89 ± 0.47	10.15 ± 0.54	17.49 ± 0.29	
ViB-4-OH	17.38 ± 2.38	16.37 ± 0.59	18.65 ± 0.34	15.90 ± 0.61	19.67 ± 1.37	
DDP ^a	16.80 ± 0.46	9.58 ± 0.24	14.09 ± 0.76	1.69 ± 0.03	25.46 ± 0.72	

Table S1. Antiproliferative Activity of Compounds 6 – 10 and ViB-4-OH (IC₅₀, μ M)

^{*a*} cisplatin was used as positive control.

	Status of embryonic development ^b								
Compd	4 μM		$8 \ \mu M$				16 µM		
•	LoD ^c	SC ^c	BM ^c	LoD	SC	BM	LoD	SC	BM
ViB	(20/20)	(20/20)	(20/20)	(20/20)	(20/20)	(19/20)	15/18	9/15	1/15
	live	yes	yes	live	yes	yes	live	yes	yes
12f	(20/20)	(20/20)	(20/20)	(20/20)	(20/20)	(19/20)	20/20	20/20	16/20
	live	yes	yes	live	yes	yes	live	yes	yes
7	(20/20)	(20/20)	(20/20)	(20/20)	(20/20)	(19/20)	9/17	6/9	0/17
	live	yes	yes	live	yes	yes	live	yes	yes

Table S2. Primary toxicity of vibsanin B and its derivatives 7 and 12f on zebrafish embryo ^a

^{*a*} All zebrafish embryos used in this work were bred in a standard condition of 28.5 °C egg water. 15-20 embryos were put into the one well of 12 well-plate and treated with vibsanin B (ViB) derivatives 7 and **12f** at concentration of 4, 8, 16 μ M, for 12 hours, respectively;

^b The primary toxicity of vibsanin B and its derivatives 7 and **12f** on zebrafish embryo (3 days post fertilization) was determined by observation of the embryonic phenotypes including systemic circulation and body movement;

^c LoD (live or dead), SC (systemic circulation), BM (body movement).

Compd. Solubility		Absorption ^c	PSA^{d}	Hepatotoxicity ^e	Rat Oral LD_{50}^{f}
7	1	1	64.347	-9.8619	0.1762
12f	2	1	71.052	-9.9846	0.7611

Table S3. In silico drug-like profiles of vibsanin B derivatives^{*a*}

^{*a*} Predicted properties were performed on ADMET and Toxicity Prediction modules of Discovery Studio 4.0. ^{*b*} aqueous solubility level in water at 25 °C. 0 (extremely low), 1 (very low), 2 (low), 3 (good), 4 (optimal). ^{*c*} human intestinal absorption (HIA) after oral administration. 0 (good), 1 (moderate), 2 (poor), 3 (very poor). ^{*d*} PSA: fast polar surface area (compounds with 10 or fewer rotatable bonds and PSA below 140 Å would have a higher chance of being orally bioavailable). ^{*e*} hepatotoxicity (< -0.4095: nontoxicity, > -0.4905: toxicity). ^{*f*} the unit is g/kg (body weight).

Mutants	Primers: Forward primers(F), Reverse primers(R)
HSP90β-NTD	F: 5'-ATAAGAATGCGGCCGCGCCTGAGGAAGTGCACCATGG-3'
(P2-S234)	R: 5'-GCTCTAGACTATTTCTCTTCCTCTGCCTCAT-3'
HSP90β-MD	F: 5'-ATAAGAATGCGGCCGCGAGCGGTAAGGATAAGAAGAA-3'
(S261-K552)	R: 5'- GCTCTAGACTACTTCTTCTTCTCCTCCTCAT-3'
HSP90β-CTD	F: 5'-ATAAGAATGCGGCCGCGGAGGAGGAGAAGAAGAAGAAGAT-3'
(E547-D724)	R: 5'- GCTCTAGACTAATCGACTTCTTCCATGCGAG-3'

TableS4. Primers for pFlag-CMV-3/4-HSP90b-NTD, MD, CTD constructs.

TableS5. Primers for pET28a(+)-His-HSP90β-CTD construct.

Mutants	Primers: Forward primers (F), Reverse primers (R)
НЅР90β-СТД (Е547-Д724)	F: 5'-GGAATTCCATATGGAGGAGGAGAAGAAGAAG-3'
	R: 5'-ACGCGTCGACCTAATCGACTTCTTCCAT-3'

	Novobiocin	12a	7	12f	8f	11	ViB
IC ₅₀	339±5.33	1.13±0.05	1.31±0.21	0.91±0.17	2.96±0.35	0.11±0.03	4.24±0.11

Table S6. IC₅₀ (μ M) of each compound in luciferase refolding assay.

HSP90β-NTD, MD, CTD cDNA sequences:

HSP90β-NTD(P2-K234):

HSP90β-MD(S261-K552):

 ACATCTCTGTCAGAGTATGTTTCTCGCATGAAGGAGACACAGAAGTCCATCTATTAC ATCACTGGTGAGAGCAAAGAGCAGGTGGCCAACTCAGCTTTTGTGGAGCGAGTGCG GAAACGGGGGCTTCGAGGTGGTATATATGACCGAGCCCATTGACGAGTACTGTGTGC AGCAGCTCAAGGAATTTGATGGGAAGAAGAAGACG GAGCTGCCTGAGGATGAGGAGGAGGAGAAGAAGAAG

HSP90β-CTD(E547-D724):

GAGGAGGAGAAGAAGAAGAAGATGGAAGAGAGAGGAGGAAAGGTTTGAGAACCTCTGCA AGCTCATGAAAGAAATCTTAGATAAGAAGGTTGAGAAGGTGACAATCTCCAATAGA CTTGTGTCTTCACCTTGCTGCATTGTGACCAGCACCTACGGCTGGACAGCCAATATG GAGCGGATCATGAAAGCCCAGGCACTTCGGGACAACTCCACCATGGGCTATATGAT GGCCAAAAAGCACCTGGAGATCAACCCTGACCACCCCATTGTGGAGACGCTGCGGC AGAAGGCTGAGGCCGACAAGAATGATAAGGCAGTTAAGGACCTGGTGGTGCTGCTG TTTGAAACCGCCCTGCTATCTTCTGGCTTTTCCCTTGAGGATCCCCAGACCCACTCCA ACCGCATCTATCGCATGATCAAGCTAGGTCTAGGTATTGATGAAGATGAAGTGGCA GCAGAGGAACCCAATGCTGCAGTTCCTGATGAGATCCCCCTCTCGAGGGCGATGA GGATGCGTCTCGCATGGAAGAAGTCGAT



Full-length NTD MD CTD

Figure S1. Vibsanin B targets Hsp90 CTD. (A) Lysates of 293T cells expressing Hsp90 β mutants including Hsp90 β -NTD, MD and CTD were incubated with 30 μ M Biotin-ViB, respectively, followed by pull down assay with strepavidin-argrose beads. The precipitates were analyzed by western blot assay against Flag antibodies. (B). Recombinant His-Hsp90 β -CTD protein was incubated with Biotin-ViB, ViB, ViB-4-OH or 7 for 2 hours, and the mixtures were blotted with antibodies against His or biotin. (C) Structures of Biotin-ViB and Biotin-linker.



Figure S2. Predicted binding mode of 6 (A), 7 (B), 9 (C), 10 (D) at the interface of the C-terminal domain of Chain B (light cyan) and Chain A (light yellow) of the yeast Hsp90 (2cg9). Ligands colored by element type (C yellow, O red, F cyan, polar H white), key residues colored by element type (C green, O red, N blue, polar H white), key interactions were showed in red dot line. Docking was performed using AutoDock v4.01.



Figure S3. Solid state conformations of compounds 13c and 8d. (A) BC conformation of 13c, red arrow is side view, gray arrow is bottom view, crystalline structure CCDC number: 981550; (B) CT conformation of 8d, red arrow is side view, gray arrow is top view, crystalline structure CCDC number: 1517432.



Figure S4. Status of embryonic development after treatment with ViB, 7, and $12f(0, 4, 8, 16 \mu M)$ for 12 h at 3dfp.

The conformational instability of vibsanin B derivatives and its influence on NMR analysis and antiproliferative activity:

Note 1. ViB exists in two major conformers (BC type and CT type)¹ which are resulted from its 11-membered ring. In NMR test, these two conformers could be always detected whether at high temperature (> 60 °C) or low temperature (< 0 °C). In this study, more conformationally stable CT type is about 2.8-fold (25 °C) to BC type which was consistent with the reference.² The conformational isomerization in ¹³ C NMR test is not so obvious as that in ¹H NMR test, there are usually 3-9 carbon signals more than the real ones. This phenomenon makes the NMR spectra of ViB derivatives seem lack of purity. For convenience, only the CT type NMR data of ViB homologues are reported,³ so does this work.



Figure **S5**. ¹H NMR spectra of vibsanin B at 25 °C. Red arrows are the signals of BC type, blue stars are the signals of CT type, the ratio of CT : BC is **2.8 : 1**



Figure S6. ¹³ C NMR spectra of vibsanin B at 25 °C. Red stars are the signals of BC type.

Note 2. The conformational instability of ViB could be improved by modification of its C7 hydroxyl group and C8 side chain. We found that C7 hydroxyl group acetylated compound 8d exists in only CT conformer (see X-ray analysis Figure S4) (Figure S7), and the improvement was also found in C7 hydroxyl group methylated compound 8e, which the ratio of CT:BC is 5.5 : 1. Similar observation was found in the ¹H NMR spectra of compounds 8f and 28f. Obviously, enhancing the conformational stability did not improve the antiproliferative activity (8f, 28f), suggesting that the conformational instable ligands (like compounds 7, 11, 12a – 12f, etc. in the present study) could also bind to the target protein well and then exhibit their biological functions like conformationally rigid ligands.



Figure S7. ¹H NMR spectra of 8d at 25 °C. The ratio of CT : BC is 1:0



Figure S8. ¹H NMR spectra of 8e at 25 °C. The ratio of CT : BC is 5.5 : 1



Figure S9. Solution stability of vibsanin B derivatives. (A) Thermostability of ViB and 12f in toluene (110 °C and 60 °C) and deionized water (100 °C); (B) Stability of ViB, 12f, 8f, and 8d in PBS pH 7.4 at 37.0 °C for 12 h, Δ % represents decomposition percentage, Δ % < 4 represents good solution stability. The decomposition behaviors of the compounds were measured by HPLC. Data for the percentage of remaining compound are expressed as values from one experiment for A or as means ± SD from three independent experiments for B.



Figure S10. HPLC traces for solution stability of vibsanin B derivatives. (A) Stability of **12f** (0.1 M) in DMSO (4 °C) for 4 weeks; (B) Thermostability of **8f** (0.1 M) in toluene (110 °C) for 30 min; (100 °C); (C) Stability of ViB (0.1 M) in deionized water (100 °C) for 10 min.

Procedure for preparation of Biotin-ViB:

Biotin-ViB. Biotin-ViB was prepared according to the literature,⁴ and was obtained as a colorless oil. ¹H NMR (800 MHz, CDCl₃) δ 7.70 (d, J = 12.7 Hz, 1H), 6.61 (d, J = 16.3 Hz, 1H), 6.11 – 6.03 (m, 2H), 5.89 (s, 1H), 5.78 (s, 1H), 5.67 (d, J = 16.1 Hz, 1H), 5.50 (d, J = 9.2 Hz, 1H), 5.43 (s, 1H), 5.17 (dd, J = 16.1, 9.2 Hz, 1H), 5.10 (t, J = 6.8 Hz, 1H), 4.92 (d, J = 11.9 Hz, 1H), 4.70 (d, J = 11.9 Hz, 1H), 4.51 (ddt, J = 17.5, 13.1, 6.5 Hz, 4H), 4.36 – 4.31 (m, 1H), 4.27 – 4.17 (m, 2H), 3.94 – 3.88 (m, 2H), 3.86 (t, J = 5.2 Hz, 2H), 3.65 (p, J = 7.4 Hz, 3H), 3.63 – 3.58 (m, 5H), 3.56 (d, J = 17.2 Hz, 1H), 3.47 (d, J = 17.2 Hz, 1H), 3.14 (dd, J = 12.0, 7.3 Hz, 1H), 2.92 – 2.87 (m, 1H), 2.72 (t, J = 9.8 Hz, 1H), 2.31 (t, J = 7.4 Hz, 2H), 2.17 (s, 3H), 1.91 (s, 3H), 1.67 (s, 3H), 1.58 (s, 3H), 1.40 (s, 3H), 1.02 (s, 3H); ¹³C NMR (200 MHz, CDCl₃) δ 200.6, 173.5, 171.6, 166.4, 163.4, 157.8, 156.6, 145.4, 145.0, 138.2, 132.6, 131.7, 128.1, 124.2, 124.1, 123.3, 116.1, 80.5, 73.3, 70.5, 70.5, 69.5, 69.2, 66.9, 63.3, 61.9, 60.1, 55.4, 50.3, 49.9, 44.6, 43.7, 40.6, 40.5, 38.9, 33.8, 28.3, 28.2, 27.5, 25.7, 24.7, 23.1, 22.8, 20.4, 17.9, 17.7; MS (ESI): [M+H]⁺ calcd for C₄₆H₆₉N₆O₁₁S, 913; found, 913; [M+Na]⁺ calcd for C₄₆H₆₈N₆O₁₁SNa, 935; found, 935.

Procedure for preparation of Biotin-linker:



Compound S2. To a solution of bromo methylacetate (**S1**) (120 mg) in THF (10 mL) was added NaI (180 mg), the resulting mixture was stirred at 0 °C for 10 min, then propargyl amine (50 mg in 2 mL THF) was added, the mixture was stirred at room temperature for 4 h after consumption of the starting materials. The reaction was quenched by adding water (2 mL). Then the mixture was portioned by adding ethyl acetate (20 mL), and the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄. The solvent was evaporated under vacuum and the residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 4:1 v/v) to afford compound **S2** (80 mg, 80%) as a yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 3.72 (s, 3H), 3.50 (s, 2H), 3.47 (d, *J* = 2.4 Hz,

2H), 2.22 (t, J = 2.3 Hz, 1H), 1.76 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 172.5, 81.2, 72.1, 52.0, 49.1, 37.8; MS (ESI): [M+H]⁺ calcd for C₆H₁₀NO₂, 128; found, 128.

Biotin-linker. To a solution of compound S2 (10 mg) and compound S3 ⁴ (47 mg) in DMF/H₂O (2.4 mL, 5:1, v/v) were added CuSO₄ (2 mg) and ascorbic acid (1.4 mg) successively. The mixture was stirred at the room temperature under nitrogen for 2.5 h after consumption of the starting materials. Then water (2 mL) was added to quench the reaction, and the aqueous phase was extracted with EtOAc (10 mL×3) for three times. The combined organic layers were washed with brine (10 mL × 3), dried over Na₂SO₄, and evaporated *in vacuo*. The resulting crude product was subjected to silica gel chromatography (DCM/MeOH 10:1, v/v) to give Biotin-linker (37 mg, 88% yield) as a colorless oil. ¹H NMR (800 MHz, CDCl₃) δ 7.72 (s, 1H), 6.06 (s, 1H), 5.47 (s, 1H), 4.52 (d, *J* = 29.3 Hz, 3H), 4.31 (s, 1H), 4.21 (m, 2H), 3.94 (brs, 2H), 3.86 (brs, 2H), 3.72 (s, 3H), 3.66 (t, *J* = 4.5 Hz, 3H), 3.62 (s, 5H), 3.48 (brs, 2H), 3.14 (s, 1H), 2.89 (d, *J* = 8.8 Hz, 1H), 2.74 (d, *J* = 11.7 Hz, 1H), 2.34 (t, *J* = 6.9 Hz, 3H), 1.77 – 1.68 (m, 2H), 1.65 (m, 4H), 1.43 (m, 3H); ¹³C NMR (200 MHz, CDCl₃) δ 173.8, 172.6, 163.7, 145.9, 123.3, 70.5, 70.4, 69.6, 69.2, 63.4, 62.0, 60.2, 55.7, 52.1, 50.3, 49.8, 44.1, 40.7, 33.8, 28.4, 28.3, 24.8; HRMS (ESI): [M+H]⁺ calcd for C₂₂H₃₇N₆O₇S, 529.2444; found, 529.2454.

Procedure for preparation of vibsanin B derivatives:

18-O-acetyl vibsanin B (8). To a stirred solution of ViB (0.2 g) and TEA (200 μ L) in DCM (5 mL) was added Ac₂O (100 μ L) and DMAP (6 mg), the mixture was stirred at room temperature for 4 h. The solvent was evaporated under vacuum and the residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 10:1 v/v) to afford compound **8** (215 mg, 98%) as a colorless oil: $[\alpha]^{17}_{D}$ +43 (*c* 2.0, CH₂Cl₂). ¹H NMR (500 MHz, CDCl₃) δ 6.46 (d, *J* = 16.4 Hz, 1H), 6.06 (d, *J* = 16.4 Hz, 1H), 6.02 (dd, *J* = 13.2, 3.8 Hz, 1H), 5.79 (s, 1H), 5.63 (d, *J* = 16.1 Hz, 1H), 5.36 (d, *J* = 9.2 Hz, 1H), 5.20 (dd, *J* = 16.2, 9.2 Hz, 1H), 5.09 (t, *J* = 7.0 Hz, 1H), 4.85 (d, *J* = 12.1 Hz, 1H), 4.59 (d, *J* = 12.3 Hz, 1H), 2.20 (s, 3H), 2.05 (s, 3H), 1.94 (s, 3H), 1.68 (s, 3H), 1.58 (s, 3H), 1.37 (s, 3H), 1.03 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 200.5, 170.9, 167.6, 159.7, 155.1, 145.5, 138.6, 132.0, 131.9, 128.6, 124.3, 123.6, 115.4, 82.1, 74.4, 66.3, 44.8, 40.8, 39.1, 27.7, 25.8, 23.3, 23.1, 21.0, 20.7, 18.7, 17.8; HRMS (ESI): [M+Cl]⁻ calcd for C₂₇H₃₈O₆Cl, 493.2362; found, 493.2359.

7,18-di-O-acetyl vibsanin B (8a). To a stirred solution of ViB (0.2 g) and TEA (670 μ L) in DCM (10 mL) was added Ac₂O (500 μ L) and DMAP (0.6 g), the mixture was stirred at room temperature for 16 h. DCM (10 mL) and water (10 mL) were added to the mixture, the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 20:1 v/v) to afford compound 8a (216 mg, 90%) as a white amorphous powder: $[\alpha]^{23}_{D}$ +21 (*c* 3.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 7.01 (d, *J* = 16.4 Hz, 1H), 6.19 (d, *J* = 16.4 Hz, 1H), 6.03 (dd, *J* = 12.9, 3.5 Hz, 1H), 5.97 (d, *J* = 9.1 Hz, 1H), 5.74 (d, *J* = 16.0 Hz, 1H), 5.74 (s, 1H), 5.15 (dd, *J* = 16.2, 9.1 Hz, 1H), 5.09 (t, *J* = 6.6 Hz, 1H), 4.84 (d, *J* = 12.1 Hz, 1H), 4.56 (d, *J* = 12.1 Hz, 1H), 2.18 (s, 3H), 2.02 (s, 3H), 1.93 (s, 3H), 1.68 (s, 3H), 1.58 (s, 3H), 1.56 (s, 3H), 1.02 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 200.4, 170.8, 169.8, 165.3, 158.2, 152.1, 146.7, 138.4, 132.0, 131.0, 124.2, 123.9, 122.8, 115.8, 81.5, 76.0, 66.5, 44.8, 40.9, 38.9, 27.7, 25.8, 23.3, 22.9, 21.9, 20.8, 20.5, 17.8, 16.7. HRMS (ESI): [M-H]⁻ calcd for C₂₉H₃₉O₇, 499.2701; found, 499.2687.

7,18-di-O-methyl vibsanin B (**8b**) and 18-O-methyl vibsanin B (**8c**). To a stirred solution of ViB (50 mg) in MeI (2 mL) was added freshly prepared silver oxide (560 mg), the mixture was sealed and stirred at 40 °C for 24 h. The mixture was passed a pad of celite, the filter cake was washed by

DCM (5 mL \times 3). The solvent was concentrated under vacuum and the residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 30:1 – 10:1 v/v) to afford compound **8b** (16 mg, 30%) and **8c** (31 mg, 60%) as colorless oil.

8b, $[\alpha]^{20}{}_{D}$ +51 (*c* 2.5, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 6.49 (d, *J* = 16.4 Hz, 1H), 6.11 (d, *J* = 16.4 Hz, 1H), 5.93 (dd, *J* = 12.5, 2.9 Hz, 1H), 5.73 (s, 1H), 5.67 (d, *J* = 16.2 Hz, 1H), 5.49 (d, *J* = 9.1 Hz, 1H), 5.14 (dd, *J* = 16.2, 9.2 Hz, 1H), 5.08 (t, *J* = 6.8 Hz, 1H), 4.21 (d, *J* = 11.3 Hz, 1H), 3.87 (d, *J* = 11.3 Hz, 1H), 3.33 (s, 3H), 3.13 (s, 3H), 2.17 (s, 3H), 1.90 (s, 3H), 1.67 (s, 3H), 1.57 (s, 3H), 1.39 (s, 3H), 1.01 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 201.5, 165.5, 157.4, 154.0, 145.6, 140.8, 132.0, 131.9, 129.7, 124.4, 123.8, 116.2, 78.9, 78.3, 74.8, 58.8, 51.6, 44.7, 40.7, 38.9, 27.6, 25.8, 23.3, 22.9, 20.5, 17.8, 14.2; HRMS (ESI): $[M+H]^+$ calcd for C₂₇H₄₁O₅, 445.2949; found, 445.2952.

8c, $[α]^{23}_{D}$ +40 (*c* 4.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.54 (d, *J* = 16.3 Hz, 1H), 6.05 (d, *J* = 16.3 Hz, 1H), 5.93 (dd, *J* = 12.8, 3.7 Hz, 1H), 5.78 (s, 1H), 5.66 (d, *J* = 16.2 Hz, 1H), 5.38 (d, *J* = 9.1 Hz, 1H), 5.19 (dd, *J* = 16.2, 9.2 Hz, 1H), 5.09 (t, *J* = 6.9 Hz, 1H), 4.18 (d, *J* = 11.6 Hz, 1H), 3.90 (d, *J* = 11.6 Hz, 1H), 3.35 (s, 3H), 2.19 (s, 3H), 2.06 – 2.01 (m, 1H), 1.93 (s, 3H), 1.67 (s, 3H), 1.57 (s, 3H), 1.36 (s, 3H), 1.02 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 201.8, 167.5, 159.6, 154.7, 145.7, 140.9, 131.9, 129.2, 128.8, 124.3, 123.4, 115.4, 81.9, 74.8, 74.5, 58.9, 44.7, 40.7, 39.0, 27.8, 25.8, 23.3, 23.0, 20.6, 18.6, 17.8; HRMS (ESI): $[M+Na]^+$ calcd for C₂₆H₃₈O₅Na, 453.2611; found, 453.2614.

18-O-triethylsilyl vibsanin B (13a). To a stirred solution of ViB (0.5 g) and TEA (330 μL) in DCM (10 mL) was added TESC1 (300 μL), the mixture was stirred at room temperature for 6 h. The solvent was evaporated under vacuum and the crude product was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 15:1 v/v) to afford compound **13a** (623 mg, 98% yield) as a white foam: $[\alpha]^{17}_{\text{D}}$ +68 (*c* 2.1, CH₂Cl₂). ¹H NMR (500 MHz, CDCl₃) δ 6.57 (d, *J* = 16.3 Hz, 1H), 6.05 (d, *J* = 16.3 Hz, 1H), 5.92 (d, *J* = 9.3 Hz, 1H), 5.79 (s, 1H), 5.68 (d, *J* = 16.2 Hz, 1H), 5.36 (d, *J* = 9.1 Hz, 1H), 5.19 (dd, *J* = 16.2, 9.1 Hz, 1H), 5.09 (d, *J* = 7.1 Hz, 1H), 4.44 (d, *J* = 12.9 Hz, 2H), 4.15 (dd, *J* = 19.4, 12.5 Hz, 1H), 2.20 (s, 3H), 1.94 (s, 3H), 1.68 (s, 3H), 1.59 (s, 3H), 1.37 (s, 3H), 1.03 (s, 3H), 0.96 (m, 9H), 0.60 (m, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 202.3, 167.4, 159.5, 154.9, 145.8, 143.3, 131.8, 128.9, 126.7, 124.4, 123.2, 115.5, 82.0, 74.5, 64.5, 44.4, 40.6, 38.9, 27.7, 25.8,

23.2, 23.0, 20.6, 18.6, 17.8, 6.9, 4.5; HRMS (ESI): $[M+Na]^+$ calcd for $C_{31}H_{50}O_5SiNa$, 553.3320; found, 553.3324.

18-O-t-butyldimethylsilyl vibsanin B (13b). To a stirred solution of ViB (0.5 g) and imidazole (160 mg) in DCM (10 mL) was added TBSC1 (270 mg) and DAMP (15 mg), the mixture was stirred at room temperature for 6 h. 10 mL of DCM and 10 mL of water were added to the mixture, the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The crude product was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 15:1 v/v) to afford compound **13b** (640 mg, 100% yield) as a white foam: $[\alpha]^{17}{}_{\rm D}$ +61 (*c* 1.0, CH₂Cl₂). ¹H NMR (500 MHz, CDCl₃) *δ* 6.56 (d, *J* = 16.3 Hz, 1H), 6.04 (d, *J* = 16.3 Hz, 1H), 5.95 – 5.86 (m, 1H), 5.78 (s, 2H), 5.67 (d, *J* = 16.2 Hz, 1H), 5.35 (d, *J* = 9.1 Hz, 1H), 5.18 (dd, *J* = 16.2, 9.1 Hz, 1H), 5.09 (t, *J* = 7.0 Hz, 1H), 4.42 (d, *J* = 12.8 Hz, 1H), 4.16 (d, *J* = 12.8 Hz, 1H), 2.20 (s, 3H), 1.94 (s, 3H), 1.68 (s, 3H), 1.58 (s, 3H), 1.37 (s, 3H), 1.02 (s, 3H), 0.91 (s, 9H), 0.09 (s, 6H). ¹³C NMR (125 MHz, CDCl₃) *δ* 202.3, 167.4, 159.4, 155.0, 145.9, 143.3, 131.8, 128.9, 126.7, 124.4, 123.2, 115.5, 82.0, 74.5, 64.9, 44.3, 40.6, 38.9, 27.7, 26.0, 25.8, 23.2, 23.0, 20.6, 20.5, 18.6, 18.4, 17.8, -3.5, -5.3; HRMS (ESI): [M+Na]⁺ calcd for C₃₁H₅₀O₅SiNa, 553.3320; found, 553.3323.

18-O-t-butyldiphenylsilyl vibsanin B $(13c)^{3a}$. To a solution of ViB (0.4 g), imidazole (140 mg) and DMAP (24 mg) in DCM (10 mL) was added TBDPSCl (390 μ L), the mixture was stirred at room temperature for 24 h. The solvent was evaporated under vacuum and the residue was purified by flash chromatography on silica gel and further recrystallization (MeOH) to afford compound **13c** as a colorless bulk crystal: $[\alpha]^{17}_{D}$ +63 (*c* 1.2, CH₂Cl₂).

7-O-acetyl vibsanin B (8d). To a solution of compound 13a (50 mg) and and TEA (670 μ L) in DCM (10 mL) was added Ac₂O (500 μ L) and DMAP (0.6 g), the mixture was stirred at room temperature for 16 h. DCM (10 mL) and water (10 mL) were added to the mixture, the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The crude product was dissolved in AcOH/THF/H₂O (8 mL, 5/2/1, v/v), the resulting mixture was stirred at RT for 4 h. Ethyl acetate (10 mL) was added, the organic layer was washed with NaHCO₃ saturated aqueous solution (10 mL × 3), brine (10 mL × 1), dried over Na₂SO₄, and concentrated under vacuum.

acetate 6:1 v/v) to afford compound **8d** (38 mg, 96%) as white powder, which further recrystallized from (petroleum ether/ethyl acetate 10:1 v/v) to yield needle crystal: $[\alpha]^{25}_{D}$ +100 (*c* 2.0, CH₂Cl₂). m.p. 210 -211 °C. ¹H NMR (600 MHz, CDCl₃) δ 7.01 (d, *J* = 16.4 Hz, 1H), 6.18 (d, *J* = 16.4 Hz, 1H), 6.02 (dd, *J* = 12.9, 3.6 Hz, 1H), 5.96 (d, *J* = 9.1 Hz, 1H), 5.73 (d, *J* = 16.0 Hz, 1H), 5.73 (s, 1H), 5.14 (dd, *J* = 16.2, 9.1 Hz, 1H), 5.08 (t, *J* = 6.9 Hz, 1H), 4.83 (d, *J* = 12.1 Hz, 1H), 4.55 (d, *J* = 12.1 Hz, 1H), 2.18 (s, 3H), 2.01 (s, 3H), 1.93 (s, 3H), 1.67 (s, 3H), 1.57 (s, 3H), 1.01 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 200.4, 170.8, 169.8, 165.3, 158.1, 152.0, 146.6, 138.3, 132.0, 131.0, 124.2, 122.8, 115.8, 81.5, 76.0, 66.5, 44.8, 40.8, 38.8, 27.6, 25.8, 23.3, 22.9, 21.9, 20.8, 20.5, 17.8, 16.7; HRMS (ESI): [M+Cl]⁻ calcd for C₂₇H₃₈O₆Cl, 493.2362; found, 493.2378.

7-O-methyl vibsanin B (*8e*). To a solution of compound **13a** (50 mg) in MeI (5 mL) was added freshly prepared silver oxide (220 mg), the mixture was sealed and stirred at 40 °C for 24 h. The mixture was passed a pad of celite, the filter cake was washed by DCM (5 mL × 3). The solvent was concentrated under vacuum. Similar to the synthesis of **8d**, the TES group was removed under the same condition to afford compound **8e** (13 mg, 32%, 65% based on the recovery of ViB 20 mg) as colorless oil: $[\alpha]^{23}_{D}$ +50 (*c* 3.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.44 (d, *J* = 16.5 Hz, 1H), 6.14 (d, *J* = 16.5 Hz, 1H), 5.99 (dd, *J* = 12.9, 3.9 Hz, 1H), 5.74 (s, 1H), 5.67 (d, *J* = 16.2 Hz, 1H), 5.46 (d, *J* = 9.2 Hz, 1H), 5.15 (dd, *J* = 16.2, 9.2 Hz, 1H), 5.09 (t, *J* = 6.8 Hz, 1H), 4.40 (d, *J* = 12.4 Hz, 1H), 4.21 (d, *J* = 12.5 Hz, 1H), 3.12 (s, 3H), 2.18 (s, 3H), 2.06 (d, *J* = 13.8 Hz, 1H), 1.90 (s, 3H), 1.68 (s, 3H), 1.58 (s, 3H), 1.39 (s, 3H), 1.02 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 202.2, 165.5, 157.6, 153.5, 145.6, 142.4, 132.0, 131.9, 130.2, 124.3, 123.8, 116.1, 78.8, 78.4, 65.4, 51.6, 44.9, 40.7, 38.9, 27.6, 25.8, 23.2, 22.9, 20.5, 17.8, 14.1; HRMS (ESI): [M+Na]⁺ calcd for C₂₆H₃₈O₅Na, 453.2611; found, 453.2610.

7-methylene vibsanin B (8f). To a solution of compound 13b (50 mg) in pyridine (5 mL) was added dropwise of SOCl₂ (200 μ L) at 0 °C, the mixture was stirred at 0 °C for 0.5 h. The reaction was then quenched by adding of water (0.2 mL) and ethylacetate (10 mL), the organic layer was washed with CuSO₄ saturated aqueous solution (10 mL × 3), brine (5 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The mixture was passed a pad of celite, the filter cake was washed by DCM (5 mL × 3). The crude product was dissolved in THF (4 mL), TBAF (100 μ L, 1 M in THF) to this solution at 0 °C, the resulting mixture was stirred at RT for 2 h. Ethyl acetate (10 mL) was added, the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 5:1 v/v) to afford compound **8f** (31 mg, 82%) as colorless oil: $[\alpha]^{17}_{D}$ -152 (*c* 0.2, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.98 (d, *J* = 16.1 Hz, 1H), 6.28 (d, *J* = 16.1 Hz, 1H), 6.05 (dd, *J* = 12.7, 4.0 Hz, 1H), 5.90 (d, *J* = 8.9 Hz, 1H), 5.80 (m, 1H), 5.74 (d, *J* = 16.3 Hz, 1H), 5.38 (s, 1H), 5.27 (dd, *J* = 16.3, 9.0 Hz, 1H), 5.18 (d, *J* = 1.7 Hz, 1H), 5.11 (t, *J* = 7.1 Hz, 1H), 4.39 (dd, *J* = 12.2, 4.1 Hz, 1H), 4.19 (d, *J* = 12.5 Hz, 1H), 2.20 (s, 3H), 1.94 (s, 3H), 1.68 (s, 3H), 1.59 (s, 3H), 1.42 (td, *J* = 12.8, 5.2 Hz, 1H), 1.05 (s, 3H); ¹H NMR (100 MHz, CDCl₃) δ 201.6, 165.3, 158.8, 150.6, 143.5, 142.2, 131.9, 130.5, 130.2, 127.0, 124.5, 124.3, 115.5, 110.7, 74.8, 65.8, 41.0, 39.0, 31.1, 27.7, 25.8, 23.4, 22.9, 20.5, 17.8; HRMS (ESI): [M+Na]⁺ calcd for C₂₅H₃₄O₄Na, 421.2349; found, 421.2351.

Synthesis of Compound 14. To a solution of ViB (20 mg) in AcOH (2 mL), was added zinc powder (6 mg), the resulting suspension was stirred vigorously at RT for 12 h. The mixture was then passed a short pad of celite, the filter cake was washed with ethyl acetate (5 mL × 3), the combined organic layers were washed with NaHCO₃ saturated aqueous solution (5 mL × 3), brine (10 mL × 1), dried over Na₂SO₄, and concentrated under vacuum. The residue was purified by chromatography on silica gel (petroleum ether/ethyl acetate 4:1 – 2:1 v/v) to afford compound 14 (14 mg, 70%) as colorless oil: $[\alpha]^{17}_{D}$ +33 (*c* 2.1, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 5.79 (t, *J* = 8.4 Hz, 1H), 5.69 (s, 1H), 5.59 (d, *J* = 15.7 Hz, 1H), 5.25 (d, *J* = 8.5 Hz, 1H), 5.14 (dd, *J* = 15.7, 8.5 Hz, 1H), 5.04 (t, *J* = 5.7 Hz, 1H), 4.35 (d, *J* = 11.7 Hz, 1H), 4.18 (d, *J* = 12.8 Hz, 1H), 3.05 (brs, 1H), 2.72 (brs, 2H), 2.62 (brs, 1H), 2.14 (s, 3H), 2.00 (brs, 1H), 1.89 (s, 3H), 1.64 (s, 3H), 1.54 (s, 3H), 1.42 (td, *J* = 12.6, 4.2 Hz, 1H), 1.30 (td, *J* = 12.6, 5.4 Hz, 1H), 1.24 (s, 3H), 0.96 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 207.2, 166.5, 158.2, 146.5, 144.2, 132.4, 131.7, 124.5, 122.4, 115.9, 81.1, 74.1, 65.9, 42.2, 38.9, 38.8, 37.1, 33.7, 29.8, 27.7, 25.8, 24.3, 23.1, 20.5, 17.7; HRMS (ESI): [M+Cl]⁻ calcd for C₂₅H₃₈O₅Cl, 453.2413; found, 453.2409.

18-methoxymethoxyl vibsanin B (15). To a solution of ViB (20 mg) and DIPEA (24 μ L) in DCM (2 mL) at 0 °C, was added MOMCl (8 μ L), the reaction was allowed to warm to RT and stir for 2 h. The reaction was then quenched by adding of water (0.2 mL) and DCM (10 mL), the organic layer

was washed with brine (5 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 15:1 v/v) to afford compound **15** (20 mg, 90%) as slightly yellow oil: $[\alpha]^{20}_{D}$ +55 (*c* 6.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.55 (d, *J* = 16.3 Hz, 1H), 6.07 (d, *J* = 16.3 Hz, 1H), 5.95 (dd, *J* = 12.9, 3.9 Hz, 1H), 5.78 (s, 1H), 5.66 (d, *J* = 16.2 Hz, 1H), 5.38 (d, *J* = 9.1 Hz, 1H), 5.20 (dd, *J* = 16.2, 9.1 Hz, 1H), 5.09 (t, *J* = 6.9 Hz, 1H), 4.63 – 4.58 (m, 2H), 4.37 (d, *J* = 11.5 Hz, 1H), 4.02 (d, *J* = 11.5 Hz, 1H), 3.33 (s, 3H), 2.20 (s, 3H), 1.94 (s, 3H), 1.68 (s, 3H), 1.58 (s, 3H), 1.37 (s, 3H), 1.04 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 201.7, 167.5, 159.8, 154.6, 145.7, 140.8, 132.0, 129.7, 128.9, 124.3, 123.4, 115.4, 96.2, 81.9, 74.6, 69.5, 55.6, 44.8, 40.7, 39.0, 27.8, 25.8, 23.3, 23.1, 20.7, 18.6, 17.8; HRMS (ESI): [M+Na]⁺ calcd for C₂₇H₄₀O₆Na, 483.2723; found, 483.2727.

18-O-p-nitrobenzoyl vibsanin B (*16*). To a solution of ViB (40 mg) and TEA (16 μL) in DCM (2 mL) at 0 °C, was added *p*-nitrobenzoylchloride (20 mg), the reaction was allowed to warm to RT and stirred for 30 min. The reaction was then quenched by adding of water (0.2 mL) and DCM (10 mL), the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 12:1 v/v) to afford compound **16** (52 mg, 95%) as colorless oil: $[α]^{23}_{D}$ +58 (*c* 3.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 8.26 (m, 2H), 8.17 (m, 2H), 6.69 (d, *J* = 16.4 Hz, 1H), 6.13 (m, 2H), 5.80 (s, 1H), 5.67 (d, *J* = 16.1 Hz, 1H), 5.34 (d, *J* = 9.2 Hz, 1H), 5.24 (dd, *J* = 16.1, 9.2 Hz, 1H), 5.17 (d, *J* = 11.9 Hz, 1H), 5.08 (t, *J* = 7.0 Hz, 1H), 4.81 (d, *J* = 11.9 Hz, 1H), 2.21 (s, 1H), 1.95 (s, 3H), 1.67 (s, 3H), 1.58 (s, 3H), 1.39 (s, 3H), 1.07 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 200.4, 167.9, 164.4, 160.5, 156.2, 150.7, 145.3, 137.8, 135.0, 132.8, 132.1, 131.1 (×2), 128.2, 124.1, 123.8 (×2), 123.7, 115.1, 82.7, 74.4, 68.1, 44.7, 40.8, 39.2, 27.9, 25.8, 23.3, 23.1, 20.7, 18.8, 17.9; HRMS (ESI): [M+Na]⁺ calcd for C₃₂H₃₉NO₈Na, 588.2568; found, 588.2566.

18-azido vibsanin B (17). To a solution of ViB (50 mg) in DMF/CCl₄ (5 mL, 1:4, v/v) was added NaN₃ (16 mg) and PPh₃ (63 mg), the reaction was warmed to 40 °C and stirred for 2 h. The reaction was then quenched by adding of water (1 mL) and ethyl acetate (10 mL), the organic layer was washed with brine (10 mL \times 3), dried over Na₂SO₄, and concentrated under vacuum. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 10:1 v/v) to afford

compound **17** (50 mg, 96%) as yellow foam: $[\alpha]^{20}_{D}$ +126 (*c* 2.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.46 (d, *J* = 16.4 Hz, 1H), 6.08 (d, *J* = 16.5 Hz, 1H), 5.98 (dd, *J* = 12.9, 4.0 Hz, 1H), 5.78 (d, *J* = 1.1 Hz, 2H), 5.66 (d, *J* = 16.2 Hz, 1H), 5.36 (d, *J* = 9.2 Hz, 1H), 5.21 (dd, *J* = 16.2, 9.2 Hz, 1H), 5.10 (t, *J* = 7.0 Hz, 1H), 4.15 (d, *J* = 13.8 Hz, 1H), 3.93 (d, *J* = 13.7 Hz, 1H), 2.20 (s, 3H), 1.94 (s, 3H), 1.68 (s, 3H), 1.58 (s, 3H), 1.38 (s, 3H), 1.04 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 200.5, 167.5, 159.9, 154.4, 145.4, 138.0, 132.1, 131.6, 128.5, 124.2, 123.7, 115.3, 81.9, 74.3, 54.5, 44.9, 40.6, 39.0, 27.8, 25.8, 23.2, 23.0, 20.7, 18.6, 17.8; HRMS (ESI): [M+Na]⁺ calcd for C₂₅H₃₅N₃O₄Na, 464.2520; found, 464.2519.

Triazol **18**. To a solution of **17** (20 mg) in DMF/H₂O (5 mL, 4:1, v/v) was added propargyl alcohol (3 μ L), sodium ascorbate (1 mg) and CuSO₄ (0.2 mg), the resulting mixture was sonicated for 30 min until the yellow solids were not precipitated any more. Water (2 mL) and ethyl acetate (20 mL) were added to the mixture, and the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 1:1 v/v) to afford compound **18** (20 mg, 88%) as white amorphous powder: [α]¹⁷_D +80 (*c* 1.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 7.50 (s, 1H), 6.19 (dd, *J* = 12.9, 3.8 Hz, 1H), 5.92 (d, *J* = 16.4 Hz, 1H), 5.75 (s, 1H), 5.55 (s, 1H), 5.25 (d, *J* = 14.5 Hz, 1H), 5.20 (d, *J* = 9.2 Hz, 1H), 5.14 – 5.08 (m, 2H), 5.03 (d, *J* = 14.3 Hz, 1H), 4.78 (d, *J* = 13.2 Hz, 1H), 4.66 (d, *J* = 13.1 Hz, 1H), 2.17 (s, 3H), 1.92 (s, 3H), 1.69 (s, 3H), 1.59 (s, 3H), 1.39 (td, *J* = 12.8, 5.2 Hz, 1H), 1.24 (s, 3H), 1.01 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 200.5, 167.3, 159.8, 154.9, 148.7, 144.9, 137.8, 133.5, 132.1, 127.7, 124.1, 123.9, 123.3, 115.3, 81.7, 73.9, 67.8, 56.3, 44.8, 40.9, 39.0, 27.8, 25.9, 23.2, 23.1, 20.7, 18.3, 17.9; HRMS (ESI): [M+H]⁺ calcd for C₂₈H₄₀N₃O₅, 498.2962; found, 498.2966.

Xanthate **19**. To a solution of ViB (40 mg) in THF (2 mL) at -40 °C was added a suspension of NaH (4 mg) in THF (1 mL), the mixture was stirred for 5 min at the same temperature, freshly distilled CS₂ (7 μ L) was added and the reaction was kept stirring for 30 min at -40 °C. To this solution, MeI (8 μ L) was added, and the reaction was allowed to warm to RT and stirred for 10 h. After completion, water (1 mL) and ethyl acetate (20 mL) were added to the mixture, the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The

residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 4:1 v/v) to afford compound **19** (35 mg, 72%) as colorless oil: $[\alpha]^{20}_{D}$ +53 (*c* 2.5, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 6.52 (d, *J* = 16.4 Hz, 1H), 6.14 (dd, *J* = 12.8, 3.9 Hz, 1H), 6.08 (d, *J* = 16.4 Hz, 1H), 5.78 (s, 1H), 5.64 (d, *J* = 16.1 Hz, 1H), 5.34 (d, *J* = 9.4 Hz, 2H), 5.24 – 5.16 (m, 2H), 5.09 (t, *J* = 6.9 Hz, 1H), 2.52 (s, 3H), 2.20 (s, 3H), 1.93 (s, 3H), 1.68 (s, 3H), 1.58 (s, 3H), 1.37 (s, 3H), 1.04 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 215.3, 200.2, 167.5, 159.8, 155.3, 145.4, 137.5, 134.1, 132.1, 128.3, 124.2, 123.6, 115.4, 82.0, 75.0, 74.7, 74.3, 44.7, 40.8, 39.0, 27.8, 25.8, 23.3, 23.0, 20.6, 18.6, 17.8; HRMS (ESI): [M+Na]⁺ calcd for C₂₇H₃₈O₅S₂, 529.2053; found, 529.2054.

4R-hydroxyl vibsanin B (20a) and 4S-hydroxyl vibsanin B (20b). To a solution of ViB (0.1 g) and CeCl₃·7H₂O (9 mg) in MeOH (5 mL) at 0 °C, was added portionwise of NaBH₄ (27 mg), the resulting mixture was stirred for 2 h at 0 °C. After completion, NH₄Cl saturated aqueous solution (1 mL) and ethyl acetate (20 mL) were added, the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The residue was purified by chromatography on silica gel (petroleum ether/ethyl acetate 3:1 – 2:1, v/v) to afford compound **20a** (48 mg, 48%) as white foam and compound **20b** (47 mg, 47%) as colorless wax.

20a: $[\alpha]^{23}_{D}$ +41 (*c* 2.0, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 5.82 – 5.75 (m, 1H), 5.74 (s, 1H), 5.59 (dd, *J* = 12.3, 3.7 Hz, 1H), 5.55 – 5.46 (m, 1H), 5.36 (d, *J* = 15.7 Hz, 1H), 5.14 (d, *J* = 9.2 Hz, 1H), 5.07 (t, *J* = 8.9 Hz, 3H), 4.28 (d, *J* = 12.1 Hz, 1H), 4.15 (d, *J* = 12.1 Hz, 1H), 2.16 (s, 3H), 1.90 (s, 3H), 1.64 (s, 3H), 1.54 (s, 3H), 1.26 (s, 3H), 1.02 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 167.7, 158.9, 147.7, 141.1, 140.4, 131.6, 129.4, 125.6, 124.6, 121.2, 115.7, 86.8, 75.8, 73.4, 66.7, 42.0, 40.7, 38.3, 27.7, 25.8, 23.4, 23.1, 22.7, 20.6, 17.8; HRMS (ESI): [M+Cl]⁻ calcd for C₂₅H₃₈O₅Cl, 453.2413; found, 453.2433.

20b: $[\alpha]^{23}{}_{D}$ +27 (*c* 3.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 5.76 – 5.73 (m, 1H), 5.71 (d, *J* = 5.5 Hz, 1H), 5.68 (d, *J* = 5.3 Hz, 1H), 5.66 (d, *J* = 4.7 Hz, 1H), 5.43 (dd, *J* = 16.7, 1.1 Hz, 1H), 5.35 – 5.31 (m, 1H), 5.07 (d, *J* = 3.6 Hz, 1H), 4.87 (d, *J* = 4.2 Hz, 1H), 4.12 (s, 1H), 2.16 (d, *J* = 0.9 Hz, 2H), 2.09 – 2.05 (m, 1H), 1.91 (d, *J* = 0.9 Hz, 2H), 1.66 (s, 3H), 1.56 (s, 3H), 1.30 (d, *J* = 7.6 Hz, 3H), 1.05 (s, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 166.9, 158.2, 147.0, 144.2, 132.8, 131.6, 127.4,

124.7, 121.9, 120.8, 116.0, 76.9, 70.6, 69.3, 66.9, 43.6, 40.9, 38.3, 27.7, 25.8, 23.4, 23.0, 20.6, 20.1, 17.8; HRMS (ESI): [M+Cl]⁻ calcd for C₂₅H₃₈O₅Cl, 453.2413; found, 453.2433.

Methyl oxime **22**. To a solution of mixed **20a** and **20b** (0.1 g) in DCM (7 mL) was added MnO₂ (0.21 g), the resulting suspension was stirred at RT for 2 h. The mixture was then passed a short pad of celite, the filter cake was washed with DCM (5 mL × 3), and the solvent was concentrated under vacuum. The residue was purified by flash chromatography on silica gel (petroleum ether/ethyl acetate 4:1, v/v) to afford compound **21** (85 mg, 89%) as yellow foam. ¹H NMR (400 MHz, CDCl₃) δ 9.42 (t, *J* = 3.0 Hz, 1H), 6.63 (dd, *J* = 12.7, 4.5 Hz, 1H), 5.80 (dd, *J* = 15.6, 9.6 Hz, 1H), 5.74 (s, 1H), 5.32 (dd, *J* = 15.7, 3.4 Hz, 2H), 5.28 – 5.16 (m, 1H), 5.05 (d, *J* = 11.9 Hz, 1H), 5.00 (d, *J* = 9.1 Hz, 1H), 4.53 (d, *J* = 11.1 Hz, 1H), 4.48 (d, *J* = 10.9 Hz, 1H), 2.17 (s, 3H), 1.91 (s, 3H), 1.67 (s, 3H), 1.57 (s, 3H), 1.29 (s, 3H), 1.16 (s, 3H); MS ESI (*m/z*): 439 [M+Na]⁺

To a solution of compound **21** (20 mg) in 1.1 mL of pyridine/AcOH (10/1, v/v) was added *N*-methoxyhydroxylamine hydrochloride (8 mg), the reaction was allowed to warm to 40 °C and stirred for 12 h. After completion, 2 mL of water and 10 mL of ethyl acetate were added, the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum to yield crude oxime intermediate. To a solution of this crude product in 2 mL of DCM was added TEMPO (0.8 mg) and PIDA (15 mg), the resulting mixture was stirred at RT for 2 h. The solvent was concentrated under vacuum, and the residue was purified by chromatography on silica gel (petroleum ether/ethyl acetate 4:1, v/v) to afford compound **22** (13 mg, 60% over 2 steps) as colorless oil: $[\alpha]^{17}_{\rm D}$ +20 (*c* 3.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 7.71 (s, 1H), 6.49 (d, *J* = 16.4 Hz, 1H), 6.07 (d, *J* = 16.4 Hz, 1H), 6.03 (dd, *J* = 13.0, 4.1 Hz, 1H), 5.78 (s, 1H), 5.59 (d, *J* = 16.1 Hz, 1H), 5.32 (d, *J* = 9.2 Hz, 1H), 5.17 (dd, *J* = 16.1, 9.2 Hz, 1H), 5.09 (t, *J* = 7.0 Hz, 1H), 3.83 (s, 3H), 2.20 (s, 3H), 1.94 (s, 3H), 1.68 (s, 3H), 1.58 (s, 3H), 1.38 (s, 3H), 1.03 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 198.4, 167.6, 160.0, 156.4, 149.5, 148.7, 145.4, 138.6, 134.6, 132.1, 128.5, 124.1, 115.3, 82.6, 74.6, 62.3, 44.9, 41.0, 39.4, 27.8, 25.8, 23.3, 20.7, 20.5, 18.8, 17.8; MS ESI (*m*/*z*): 444 [M+H]⁺

18-aldehyde vibsanin B (9)^{3a}. A 50 mL round bottom flask was charged with ViB (1 g) and DCM-DMSO (18 mL, 5 : 1, v/v), IBX (1.7 g) was added in. The mixture was stirred for 3 h at room

temperature. The mixture was passed through short pad of Celite. The solvent was concentrated under vacuum and the residue was purified by flash chromatography on silica gel (petroleum ether : ethyl acetate = 10 : 1, v/v) to afford the product as a colorless foam (yields: 95%): $[\alpha]^{20}_{D}$ +48 (*c* 2.2, CH₂Cl₂).

General Procedure for Synthesis of Compounds $23a - 23c^{3a}$. A solution of compound 7 (42 mg) in DCM (1 mL), the appropriate secondary amine (2 equiv.) and NaBH(OAc)₃ (42 mg) was stirred at RT. The progress of the reactions was monitored by TLC. The solvent was evaporated under vacuum and the mixture was purified by column chromatography on silica gel.

18-carboxyl vibsanin B (10). To a solution of compound **9** (0.2 g), 2-methyl-2-butene (1 mL) and NaH₂PO₄ (0.25 g) in *t*BuOH/H₂O (4 mL, 3/1, v/v) was added portionwise of NaClO₂ (0.43 g), the resulting mixture was vigorously stirred at RT for 10 min. 5 mL of NaCl saturated aqueous solution was added, the mixture was extracted with ethyl acetate (5 mL × 3), and combined organic layers were washed with brine (5 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The residue was purified by chromatography on silica gel (petroleum ether/ethyl acetate 1:1 – 1:2, v/v) to afford compound **10** (0.18 g, 88%) as white foam: $[\alpha]^{17}_{D}$ +22 (*c* 2.5, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃) δ 7.21 (dd, *J* = 16.0 Hz, 1H), 5.37 (d, *J* = 9.4 Hz, 1H), 5.17 (dd, *J* = 16.5 Hz, 1H), 5.77 (s, 1H), 5.61 (d, *J* = 16.0 Hz, 1H), 5.37 (d, *J* = 9.4 Hz, 1H), 5.17 (dd, *J* = 16.2, 9.0 Hz, 1H), 5.10 (t, *J* = 6.0 Hz, 1H), 2.19 (s, 3H), 1.93 (s, 3H), 1.67 (s, 3H), 1.58 (s, 3H), 1.37 (s, 3H), 1.06 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 211.0, 196.8, 167.4, 159.9, 156.3, 145.1, 144.5, 134.9, 132.0, 131.6, 124.1, 123.9, 115.1, 82.2, 74.3, 53.7, 44.9, 41.2, 39.1, 27.6, 25.6, 23.1, 20.5, 18.4, 17.6; HRMS (ESI): [M+CI]⁻ calcd for C₂₅H₃₄O₆Cl, 465.2049; found, 465.2067.

General Procedure for Synthesis of 18-*N*,*N*-dialkyl formamide vibsanin B (24b – 24f). To a solution of compound 10 (20 mg), corresponding amine (2 equiv.), DMAP (3 mg) in DCM (2 mL) was added EDCI (9 mg), the resulting mixture was stirred at RT for 12 - 24 h. After completion, water (2 mL) and DCM (10 mL) were added, and the organic layer was washed with brine (5 mL × 3), dried over Na₂SO₄, and concentrated under vacuum. The residue was purified by chromatography on silica gel.

18-N,N-dimethyl formamide vibsanin B (24a) (71%), slightly yellow oil: $[α]^{17}_{D}$ +8 (*c* 3.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.94 (d, *J* = 16.3 Hz, 1H), 6.06 (d, *J* = 16.3 Hz, 1H), 6.01 (dd, *J* = 12.9, 3.9 Hz, 1H), 5.76 (s, 1H), 5.66 (d, *J* = 16.1 Hz, 1H), 5.43 (d, *J* = 9.1 Hz, 1H), 5.17 (dd, *J* = 16.1, 9.2 Hz, 1H), 5.05 (t, *J* = 7.1 Hz, 1H), 3.11 (s, 3H), 2.96 (s, 3H), 2.18 (s, 3H), 2.11 (dd, *J* = 13.8, 4.0 Hz, 1H), 1.92 (s, 3H), 1.65 (s, 3H), 1.56 (s, 3H), 1.37 (s, 3H), 1.09 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 197.5, 167.3, 166.9, 158.9, 158.0, 145.0, 139.7, 131.9, 131.8, 128.2, 124.2, 124.0, 115.6, 81.3, 74.2, 44.5, 40.8, 39.3, 39.1, 35.2, 27.7, 25.8, 23.2, 23.0, 20.6, 18.3, 17.8; HRMS (ESI): [M+H]⁺ calcd for C₂₇H₄₀NO₅, 458.2901; found, 458.2901.

18-N,N-diethyl formamide vibsanin B (**24b**) (78%), slightly yellow oil: $[α]^{17}_{D}$ +30 (*c* 3.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 7.02 (d, *J* = 16.3 Hz, 1H), 6.06 (d, *J* = 16.3 Hz, 1H), 5.94 (dd, *J* = 12.9, 3.8 Hz, 1H), 5.76 (s, 1H), 5.69 (d, *J* = 16.1 Hz, 1H), 5.44 (d, *J* = 9.1 Hz, 1H), 5.17 (dd, *J* = 16.1, 9.1 Hz, 1H), 5.07 (t, *J* = 7.0 Hz, 1H), 3.67 (m, 1H), 3.44 (m, 1H), 3.37 (m, 2H), 2.19 (s, 3H), 1.93 (s, 3H), 1.66 (s, 3H), 1.56 (s, 3H), 1.37 (s, 3H), 1.12 (m, 6H), 1.03 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 197.4, 167.0, 166.7, 159.0, 158.4, 145.1, 140.5, 131.9, 130.1, 128.3, 124.2, 124.0, 115.6, 81.5, 74.3, 44.5, 43.2, 40.8, 39.4, 39.1, 27.7, 25.8, 23.3, 23.0, 20.6, 18.3, 17.8, 14.3, 12.8; HRMS (ESI): [M-H]⁻ calcd for C₂₉H₄₂NO₅, 484.3068; found, 484.3100.

18-N,N-diisopropyl formamide vibsanin B (24c) (70%), slightly yellow oil: $[\alpha]^{17}_{D}$ +12 (c 1.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.49 (d, J = 16.2 Hz, 1H), 6.26 (d, J = 16.2 Hz, 1H), 5.78 (d, J = 15.8 Hz, 1H), 5.76 (s, 1H), 5.37 (d, J = 9.8 Hz, 1H), 5.15 (dd, J = 16.4, 8.9 Hz, 1H), 5.05 (t, J = 7.0 Hz, 1H), 4.07 (dt, J = 13.2, 6.7 Hz, 1H), 3.44 (dt, J = 13.6, 6.7 Hz, 1H), 2.32 (dd, J = 14.4, 10.1 Hz, 1H), 2.17 (s, 3H), 2.01 (dd, J = 14.4, 3.8 Hz, 1H), 1.92 (s, 3H), 1.68 (s, 3H), 1.56 (s, 3H), 1.47 (d, J = 6.8 Hz, 6H), 1.38 (s, 3H), 1.12 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 197.9, 167.0, 165.3, 159.1, 155.5, 145.1, 144.1, 141.0, 138.3, 131.9, 124.2, 123.8, 115.6, 83.2, 76.5, 51.4, 45.9, 42.8, 41.7, 40.0, 37.1, 35.4, 27.7, 25.8, 23.4, 21.1, 21.0, 20.7, 20.6, 20.4, 17.7; HRMS (ESI): [M+H]⁺ calcd for C₃₁H₄₈NO₅, 514.3527; found, 514.3529.

18-pyrrolidinyl formamide vibsanin B (24d) (68%), slightly yellow oil: $[\alpha]_{D}^{16} + 15$ (*c* 2.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.82 (d, *J* = 16.3 Hz, 1H), 6.37 (dd, *J* = 13.0, 3.8 Hz, 1H), 6.19 (dd, *J* = 12.5, 4.6 Hz, 1H), 6.10 (t, *J* = 12.7 Hz, 1H), 5.77 (s, 1H), 5.64 (d, *J* = 16.2 Hz, 1H), 5.40 (d, *J* = 9.1 Hz, 1H), 5.18 (dd, J = 16.2, 9.2 Hz, 1H), 5.07 (t, J = 7.1 Hz, 1H), 3.64 (dd, J = 10.5, 5.2 Hz, 1H), 3.49 (m, 1H), 3.42 (dd, J = 16.9, 6.9 Hz, 1H), 3.36 (m, 1H), 2.19 (s, 3H), 1.94 (s, 3H), 1.66 (s, 3H), 1.57 (s, 3H), 1.37 (s, 3H), 1.04 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 198.2, 167.1, 166.5, 159.3, 157.4, 145.1, 140.3, 134.6, 132.0, 128.7, 124.2, 124.0, 115.5, 81.6, 74.3, 48.7, 46.5, 44.7, 41.0, 39.1, 27.8, 26.3, 25.8, 24.3, 23.3, 23.1, 20.6, 18.5, 17.8; HRMS (ESI): [M+H]⁺ calcd for C₂₉H₄₂NO₅, 484.3057; found, 484.3058.

18-morpholinyl formamide vibsanin B (24e) (82%), slightly yellow oil: $[\alpha]^{16}_{D}$ +11 (*c* 5.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.96 (d, *J* = 16.3 Hz, 1H), 6.06 (d, *J* = 16.3 Hz, 1H), 5.99 (dt, *J* = 13.2, 6.6 Hz, 1H), 5.76 (s, 1H), 5.67 (d, *J* = 16.1 Hz, 1H), 5.44 (d, *J* = 9.1 Hz, 1H), 5.18 (dd, *J* = 16.1, 9.2 Hz, 1H), 5.05 (t, *J* = 7.1 Hz, 1H), 3.95 – 3.34 (m, 8H), 2.19 (s, 3H), 2.10 (dd, *J* = 13.8, 3.9 Hz, 1H), 1.93 (s, 3H), 1.65 (s, 3H), 1.56 (s, 3H), 1.37 (s, 3H), 1.10 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 197.1, 167.0, 165.9, 159.2, 158.3, 145.0, 139.5, 139.2, 132.0, 131.9, 128.1, 124.1, 115.5, 81.4, 74.2, 67.2, 66.7, 48.0, 44.5, 42.5, 40.9, 39.1, 27.7, 25.8, 23.3, 23.0, 20.6, 18.3, 17.8; HRMS (ESI): [M+H]⁺ calcd for C₂₉H₄₂NO₆, 484.3057; found, 484.3058.

18-(*N*-methylpiperazinyl) formamide vibsanin *B* (24*f*) (75%), yellow oil: $[α]^{23}_{D}$ +35 (*c* 13.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.98 (d, *J* = 16.3 Hz, 1H), 6.06 (d, *J* = 16.3 Hz, 1H), 5.97 (dd, *J* = 12.9, 3.9 Hz, 1H), 5.77 (s, 1H), 5.67 (d, *J* = 16.2 Hz, 1H), 5.44 (d, *J* = 9.1 Hz, 1H), 5.18 (dd, *J* = 16.1, 9.2 Hz, 1H), 5.06 (t, *J* = 7.1 Hz, 1H), 3.84 (m, 4H), 3.52 (brs, 2H), 3.42 (brs, 2H), 2.30 (s, 3H), 2.19 (s, 3H), 2.10 (dd, *J* = 13.8, 3.9 Hz, 1H), 1.93 (s, 3H), 1.66 (s, 3H), 1.56 (s, 3H), 1.37 (s, 3H), 1.10 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 197.2, 167.1, 165.7, 159.2, 158.4, 145.0, 139.8, 132.0, 131.4, 128.2, 124.2, 124.0, 115.5, 81.5, 74.3, 55.4, 54.6, 47.5, 46.1, 44.5, 42.0, 40.9, 39.1, 27.7, 25.8, 23.3, 23.0, 20.6, 18.4, 17.8; HRMS (ESI): $[M+H]^+$ calcd for C₃₀H₄₅N₂O₅, 513.3328; found, 513.3329.

General Procedure for Synthesis of Compounds 26b and 26c. To a solution of compound 13b (0.5 g), $CeCl_3 \cdot 7H_2O$ (35 mg) or compound 13c (0.5 g), $CeCl_3 \cdot 7H_2O$ (28 mg) in MeOH (10 mL) at -78 °C, was added portionwise of NaBH₄ (0.11 g) or (86 mg), the resulting mixture was stirred at -78 °C for 2 h. After completion, NH₄Cl saturated aqueous solution (1 mL) and ethyl acetate (20 mL) were added, the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and

concentrated under vacuum to yield crude 25b or 25c which was used in next step without further purification.

The crude **25b** or **25c** was dissolved in DCM (10 mL), to this solution was added TEA (390 μ L) or (320 μ L) and TESCl (315 μ L) or (255 μ L), the resulting mixture was stirred at RT for 8 h. After completion, water (1 mL) and DCM (20 mL) were added, the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum to yield crude disilane intermediate. This crude product was dissolved in THF/Et₂O (10 mL, 1/1, v/v) at 0 °C, to this solution was added LiBH₄ (41 mg) or (33 mg), the reaction was allowed to warm to RT and stirred for 12 h. After completion, NH₄Cl saturated aqueous solution (1 mL) and ethyl acetate (20 mL) were added, the organic layer was washed with brine (10 mL × 3), dried over Na₂SO₄, and concentrated under vacuum, the residue was purified by chromatography on silica gel (petroleum ether/ethyl acetate 6:1, v/v) to afford compound **26b** (0.395 g, 75% over 3 steps) as colorless oil and compound **26c** (0.365 g, 70% over 3 steps). **26b**: MS ESI (*m/z*): 586 [M+Na]⁺; **26c**: MS ESI (*m/z*): 711 [M+Na]⁺.

General Procedure for Synthesis of compounds 28b - 28e. To a solution of compound 26b (50 mg), corresponding acid (0.18 mmol), DMAP (5mg) in DCM (5 mL) at 0 °C was added DCC (37 mg), the reaction was allowed to warm to RT and stirred for 12 - 24 h. After completion, water (1 mL) and ethyl acetate (10 mL) were added, and the organic layer was washed with brine (5 mL × 3), dried over Na₂SO₄, and concentrated under vacuum to give crude ester. This crude product was dissolved in AcOH/THF/H₂O (6.5 mL, 8/2/1, v/v) and stirred for 2 - 4 h. After completion, ethyl acetate (10 mL) was added, the organic layer was washed with NaHCO₃ saturated aqueous solution (10 mL × 2), brine (5 mL × 3), dried over Na₂SO₄, and concentrated under vacuum to give the crude C4-OTES removed intermediate. This crude product was dissolved in DCM/DMSO (6 mL, 5/1, v/v), to this solution was added IBX (75 mg), the resulting mixture was stirred at RT for 2 - 6 h. After completion, the mixture was passed a short pad of celite, the filter cake was washed with ethyl acetate (5 mL × 3), and the combined organic layers were washed with brine (5 mL × 3), dried over Na₂SO₄, and concentrated under vacuum to give crude ketones 27b - 27e. This crude product was dissolved in THF/AcOH (2.1 mL, 20/1, v/v), to this solution was added TBAF (90 μ L, 2 M in THF), the resulting mixture was stirred 1 - 4 h. After completion, water (1 mL) and ethyl acetate (10 mL)

were added, and the organic layer was washed with brine (5 mL \times 3), dried over Na₂SO₄, and concentrated under vacuum, and the residue was purified by chromatography on silica gel.

28b, (32% over 4 steps), colorless oil: $[\alpha]^{17}_{D}$ +9 (*c* 4.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.55 (d, *J* = 16.3 Hz, 1H), 6.08 (d, *J* = 16.3 Hz, 1H), 5.97 (dd, *J* = 12.9, 3.8 Hz, 1H), 5.70 (d, *J* = 16.2 Hz, 1H), 5.33 (d, *J* = 9.1 Hz, 1H), 5.14 (dd, *J* = 16.2, 9.2 Hz, 1H), 5.10 (t, *J* = 7.0 Hz, 1H), 4.42 (d, *J* = 12.6 Hz, 1H), 4.12 (d, *J* = 12.7 Hz, 1H), 2.15 (s, 3H), 2.04 (brd, *J* = 10.9 Hz, 1H), 1.68 (s, 3H), 1.58 (s, 3H), 1.38 (s, 3H), 1.02 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 202.9, 171.8, 154.8, 146.1, 142.9, 131.9, 129.8, 129.0, 124.3, 123.0, 82.0, 74.0, 65.3, 44.8, 41.0, 40.8, 38.9, 25.8, 22.9, 21.4, 18.3, 17.8; HRMS (ESI): [M+Na]⁺ calcd for C₂₂H₃₂O₅Na, 399.2147; found, 399.2141.

28c, (26% over 4 steps), colorless oil: $[\alpha]^{20}_{D}$ +28 (*c* 2.1, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.56 (d, *J* = 16.3 Hz, 1H), 6.07 (d, *J* = 16.3 Hz, 1H), 5.97 (dd, *J* = 12.8, 3.8 Hz, 1H), 5.69 (d, *J* = 16.2 Hz, 1H), 5.33 (d, *J* = 9.1 Hz, 1H), 5.15 (dd, *J* = 16.2, 9.2 Hz, 1H), 5.09 (t, *J* = 7.0 Hz, 1H), 4.41 (d, *J* = 12.5 Hz, 1H), 4.14 (d, *J* = 12.7 Hz, 1H), 2.64 (m, 1H), 1.67 (s, 3H), 1.58 (s, 3H), 1.37 (s, 3H), 1.23 (d, *J* = 7.0 Hz, 3H), 1.20 (d, *J* = 6.9 Hz, 3H), 1.03 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 202.8, 178.0, 154.9, 146.0, 142.9, 131.9, 129.7, 128.8, 124.3, 123.0, 81.9, 74.2, 65.2, 44.7, 40.8, 38.9, 34.3, 25.8, 23.3, 23.0, 19.2, 18.9, 18.3, 17.8; HRMS (ESI): [M+Na]⁺ calcd for C₂₄H₃₆O₅Na, 427.2460; found, 427.2462.

28d, (35% over 4 steps), colorless oil: $[\alpha]^{17}_{D}$ +27 (*c* 2.1, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.53 (d, *J* = 16.3 Hz, 1H), 6.08 (d, *J* = 16.4 Hz, 1H), 5.98 (dd, *J* = 12.8, 3.9 Hz, 1H), 5.68 (d, *J* = 16.2 Hz, 1H), 5.35 (d, *J* = 9.2 Hz, 1H), 5.16 (dd, *J* = 16.2, 9.2 Hz, 1H), 5.09 (t, *J* = 7.0 Hz, 1H), 4.41 (d, *J* = 12.7 Hz, 1H), 4.16 (d, *J* = 12.7 Hz, 1H), 2.40 (m, 2H), 1.68 (s, 3H), 1.58 (s, 3H), 1.37 (s, 3H), 1.03 (s, 3H), 0.98 (td, *J* = 7.4, 2.3 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 202.6, 174.7, 154.5, 146.0, 142.8, 132.0, 129.8, 128.9, 124.3, 123.1, 82.1, 74.2, 65.3, 44.8, 40.8, 38.9, 36.5, 25.9, 23.3, 23.0, 18.6, 18.5, 17.8, 13.8; HRMS (EI): [M]⁺ calcd for C₂₄H₃₆O₅, 404.2563; found, 404.2562.

28e, (42% over 4 steps), colorless oil: $[\alpha]^{18}{}_{D}$ +50 (*c* 2.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 8.09 (m, 2H), 7.60 (t, *J* = 7.5 Hz, 1H), 7.47 (t, *J* = 7.8 Hz, 2H), 6.61 (d, *J* = 16.4 Hz, 1H), 6.13 (d, *J* = 16.4 Hz, 1H), 6.01 (dd, *J* = 12.8, 4.0 Hz, 1H), 5.78 (d, *J* = 16.2 Hz, 1H), 5.61 (d, *J* = 9.1 Hz, 1H), 5.31 (dd, *J* = 16.2, 9.2 Hz, 1H), 5.10 (t, *J* = 7.0 Hz, 1H), 4.43 (d, *J* = 12.6 Hz, 1H), 4.19 (d, *J* = 12.6 Hz, 1H), 1.68 (s, 3H), 1.58 (s, 3H), 1.50 (s, 3H), 1.06 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 202.5, 167.2, 154.3, 146.3, 142.8, 133.7, 132.0, 130.0, 129.8, 129.7, 129.0, 128.7, 124.3, 123.1, 82.8, 74.5, 65.3, 44.9, 40.9, 39.0, 25.8, 23.3, 23.0, 18.7, 17.9; HRMS (EI): [M]⁺ calcd for C₂₇H₃₄O₅, 438.2406; found, 438.2399.

Synthesis of Compounds 28*f*. Prepared by using 26*c* (50 mg), diisopropylethylamine (38 μ L) and MOMCl (11 μ L) following the general procedure for synthesis of compounds 28*b* – 28*e* to afford 28*f* (28% over 4 steps) as colorless oil: [α]¹⁶_D +11 (*c* 5.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 6.46 (d, *J* = 16.4 Hz, 1H), 6.14 (d, *J* = 16.4 Hz, 1H), 5.99 (dd, *J* = 12.8, 3.8 Hz, 1H), 5.61 (d, *J* = 16.1 Hz, 1H), 5.21 (dd, *J* = 16.1, 9.0 Hz, 1H), 5.09 (t, *J* = 6.9 Hz, 1H), 4.71 (d, *J* = 7.3 Hz, 1H), 4.62 (d, *J* = 7.4 Hz, 1H), 4.43 (dd, *J* = 12.2, 3.9 Hz, 1H), 4.28 (d, *J* = 9.0 Hz, 1H), 4.17 (d, *J* = 12.3 Hz, 1H), 3.38 (s, 3H), 1.68 (s, 3H), 1.59 (s, 3H), 1.39 (s, 3H), 1.04 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 202.2, 153.6, 144.9, 142.7, 131.9, 130.4, 130.1, 125.1, 124.4, 92.2, 80.6, 80.5, 65.5, 55.8, 45.1, 40.8, 39.0, 25.8, 23.4, 23.0, 17.8, 14.4; HRMS (ESI): [M+Na]⁺ calcd for C₂₂H₃₄O₅Na, 401.2298; found, 401.2301.

4*R*-methoxyl vibsanin *B* (**29a**) and 4*S*-methoxyl vibsanin *B* (**29b**). Prepared by using crude **25b** (0.1 g), silver oxide (220 mg) and MeI (60 μ L) following the general procedure for synthesis of compounds **28b** – **28e** to afford **29a** (42% over 2 steps) and **29b** (41% over 2 steps) as colorless oil.

29a: $[\alpha]^{15}_{D}$ +26 (*c* 1.3, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 5.75 (s, 1H), 5.63 (m, 2H), 5.50 (d, J = 15.8 Hz, 1H), 5.39 (d, J = 15.7 Hz, 1H), 5.16 (dd, J = 15.8, 9.4 Hz, 1H), 5.06 (d, J = 9.3 Hz, 2H), 4.69 (d, J = 9.6 Hz, 1H), 4.31 (t, J = 9.6 Hz, 1H), 3.98 (d, J = 12.0 Hz, 1H), 3.27 (s, 3H), 2.18 (s, 3H), 1.99 (dd, J = 13.0, 4.2 Hz, 1H), 1.92 (s, 3H), 1.65 (s, 3H), 1.55 (s, 3H), 1.28 (s, 3H), 1.05 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 166.8, 158.1, 146.5, 141.3, 139.2, 130.5, 125.2, 124.6, 123.4, 120.0, 114.4, 86.6, 81.9, 74.9, 65.5, 55.0, 40.8, 39.6, 37.1, 30.0, 26.6, 24.7, 22.3, 22.0, 19.4, 16.6; HRMS (ESI): $[M+Na]^+$ calcd for C₂₆H₄₀O₅Na, 455.2768; found, 455.2770.

29b: $[\alpha]_{D}^{15} + 22$ (*c* 1.0, CH₂Cl₂). ¹H NMR (600 MHz, CDCl₃) δ 5.75 (s, 1H), 5.73 (d, *J* = 4.7 Hz, 1H), 5.63 (d, *J* = 5.2 Hz, 1H), 5.50 (d, *J* = 0.9 Hz, 1H), 5.35 (m, 1H), 5.21 (d, *J* = 7.3 Hz, 1H), 5.07 (s, 1H), 4.35 (d, *J* = 4.6 Hz, 1H), 4.10 (s, 1H), 3.28 (s, 2H), 2.31 (t, *J* = 13.0 Hz, 1H), 2.17 (s, 3H), 1.91 (s, 3H), 1.66 (s, 3H), 1.56 (s, 3H), 1.32 (s, 3H), 1.04 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ

166.9, 158.3, 147.3, 141.6, 135.1, 134.0, 131.5, 127.2, 124.7, 120.7, 115.9, 79.5, 76.8, 69.3, 66.8, 56.7, 43.1, 40.7, 38.4, 31.1, 27.7, 25.8, 23.4, 23.0, 20.6, 17.8; HRMS (ESI): $[M+Na]^+$ calcd for $C_{26}H_{40}O_5Na$, 455.2768; found, 455.2769.

Precedure for preparation of plasimids and recobinant HSP90β-CTD protein

Plasimids. Human HSP90β gene coding sequnece is cloned from the plasmid vector pcDNA3.1-Flag-HSP90β in Chen's lab,⁵ and subcloned into pET28a(+) vector to generate a 6×His-tagged fusion protein. The HSP90β CTD coding sequence(E547-D724) is also cloned from vector pcDNA3.1-Flag-HSP90β and then subcloned into the pET28a(+) vector, producing the vector called pET-28a(+)-HSP90β-CTD. HSP90β NTD (amino acid: P2-K234), MD (amino acid: S261-K552) and CTD (amino acid: E547-D724) sequences were also cloned from pcDNA3.1-Flag-HSP90β vector and subcloned into pcDA3.1-Flag empity vector, generating pcDNA3.1-Flag-HSP90β-NTD, pcDNA3.1-Flag-HSP90β-MD and pcDNA3.1-Flag-HSP90β-CTD vectors, respectively. The specific primers used to amplify the coding sequence for the specific vectors are listed in the TableS4 and TableS5. All of the coding genes in above vectors were sequenced and convinced a 100% identies match.

Preparation of recombinant HSP90β-CTD protein. The *E. coli* strain BL21(DE3) was transformed with pET28a-HSP90β-C, and cultured in LB medium containing 100µg/mL kanamycin at 37°C. When the absorbance of culturing medium is from 0.6 to 1.0 at 600 nm, the BL21 *E. coli* with transformed vectors was induced with 0.5 mM isopropyl-D-1-thiogalactopyranoside (IPTG) for 4 hours before being harvested by centrifugation. Then the cell pellets were suspended in equilibration buffer (20 mM TrisHCl, 100 mM NaCl, pH 8.0, 5mM β-Mercaptoethanol (β-ME), 1mM PMSF) and homonized by sonication. After centrifugation(25000rpm, 45min), the supernatant was loaded to a HisTrap column (GE Healthcare) and washed with washing buffer (20mM Tris, 300mM NaCl, 25mM imidazole, 5mM β-ME, pH 8.0). Then the column was eluted with elution buffer (20 mM TrisHCl, 100 mM NaCl, 150mM imidazole, 1mM PMSF, 5mM β-ME, pH 8.0) and the expressed proteins were collected according to the absorbance peak at 280 nm on the FPLC (GE Healthcare). Then the protein within elution buffer was subject to dialysis in the dialysis buffer (20mM TrisHCl, 20mM NaCl, EDTA 5mM, pH 8.0) at 4°C overnight. After dialysis, the protein exchanged with dialysis buffer was loaded to ion exchange column Q (GE Healthcare) and washed

with low salt buffer (20 mM Tris, 20 mM NaCl, 5mM EDTA, 5mM β -ME, pH 8.0). The HSP90 β -CTD protein was eluted with a NaCl gradient (20 mM Tris, pH 8.0, 0 - 1 M NaCl).

Western blot analysis. Cell lysates were subjected to be electrophoresed and seperated in the 10% SDS-PAGE, and then the proteins in the gel transferred were to enhanced chemiluminescence-nitrocellulose membranes (Bio-Rad, catlog: 162-0112). After blocking with 5% nonfat milk in Tris-buffered saline, the membranes were incubated with the antibody against His or Flag tags (Cell signaling Technology) overnight at 4 °C, followed by incubation of horseradish peroxidase (HRP)-linked secondary antibody (Cell Signaling Technology) for 1 hour at room temperature. Detection was performed by Immobilon Western Chemiluminescent HRP substrate Kit (Millipore, cat. No. WBKLS0100) according to the chemoiluminal manufacturer's instruction (LAS-4000, FUJIFILM). Signal intensity of protein was normalized against ubiquitously expressed proteins GAPDH or actin using Quantity One (Bio-Rad, Hercules, CA, USA) software.






Supporting information







































































s71






















HPLC chromatograms of tested vibsanin B derivatives





































Supporting information









Supporting information

















s97













s100













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