

Supporting information

**Palladium-Catalyzed Asymmetric Allylic Alkylation of Alkyl
Substituted Allyl Reagents with Acyclic Amides**

Yang-Jie Jiang,[†] Gao-Peng Zhang,[†] Jian-Qiang Huang,[†] Di Chen,[†] Chang-Hua Ding,^{*,†} and Xue-Long Hou^{*,†,‡}

[†]State Key Laboratory of Organometallic Chemistry, [‡]Shanghai–Hong Kong Joint Laboratory in Chemical Synthesis, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, 345 Lingling Road, Shanghai 200032, China.

xlhou@sioc.ac.cn; dingch@sioc.ac.cn

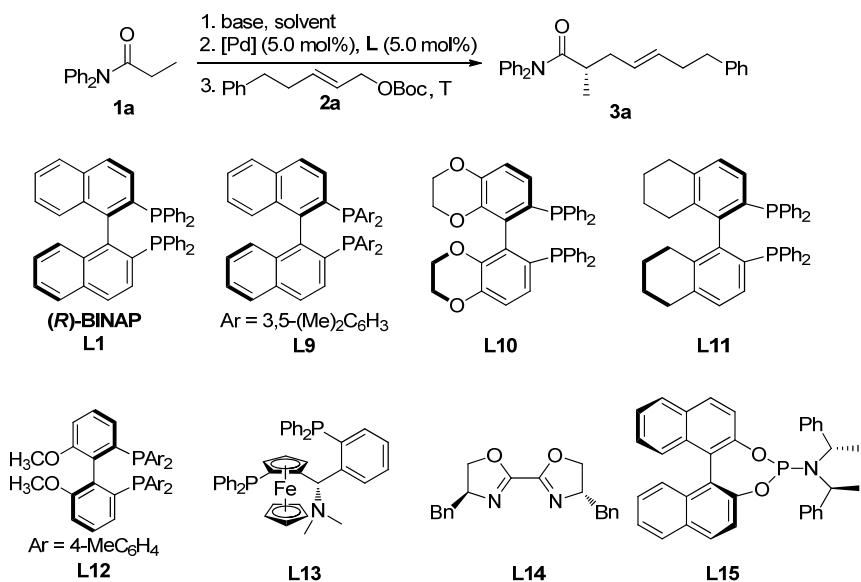
Table of Contents

| | |
|--|------------|
| 1. General methods..... | S2 |
| 2. Screening of reaction conditions for Pd-catalyzed asymmetric allylic alkylation of amide 1a with allyl reagent 2a..... | S2 |
| 3. Effect of amide on the Pd-catalyzed asymmetric allylic alkylation..... | S4 |
| 4. Synthesis of allylic reagents 2..... | S4 |
| 5. General experimental procedure and characterization of products..... | S8 |
| 6. Pd-Catalyzed Asymmetric Allylic Alkylation of Amide 1a with Allyl Reagents 2a on 1.0 mmol Scale | S18 |
| 7. Transformation of the allylated product 3d and its determination of absolute configuration..... | S18 |
| 8. Enantioselective formal synthesis of enantiomer of Dubiusamine A..... | S21 |
| 9. Preliminary studies of the beta-hydride elimination for Pd-catalyzed allylic alkylation of amide 1a with allyl reagent 2j^a..... | S23 |
| 10. References..... | S26 |
| 11. Spectra..... | S27 |

1. General methods

Commercially available reagents were used without further purification. Solvents were purified prior to use according to the standard methods. Column chromatography was performed on silica gel (300–400 mesh) using a forced flow of eluent. NMR spectra were recorded at room temperature on an NMR instrument operating at 400 MHz. Chemical shifts for ¹H NMR are reported in parts per million with the solvent resonance as the internal standard (7.26 ppm for CHCl₃). The data were reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, sep = septet, bs = broad singlet, m = multiplet), coupling constants (Hz), and integration. ¹³C NMR spectra were recorded on an NMR instrument operating at 100 MHz with complete proton decoupling. Chemical shifts were reported in parts per million with the solvent resonance as the internal standard (77.1 ppm for CDCl₃). MS and HRMS were measured in EI, ESI, or DART (direct analysis in real-time) mode and the mass analysis mode of the HRMS was TOF. Infrared spectra were recorded from thin films of pure samples. Melting points were measured on an XT-4 micromelting point apparatus. Thin layer chromatography was performed on precoated glass-backed plates and visualized with UV light at 254 nm. Enantiomeric ratios were determined by chiral HPLC analysis in comparison with authentic racemic materials.

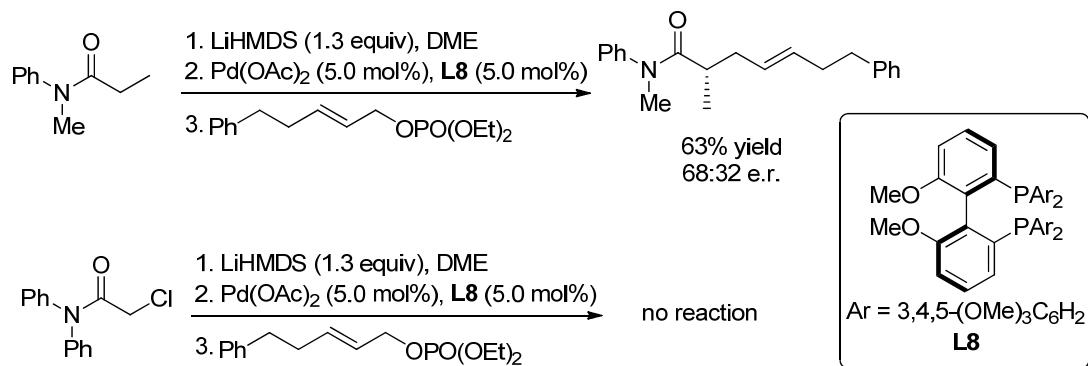
2. Screening of reaction conditions for Pd-catalyzed asymmetric allylic alkylation of amide 1a with allyl reagent 2a^a



| entry | base | solvent | [Pd] | L | yield % ^[b] | ee% ^[c] |
|-------|---------------------------------|-------------------|---|------------|------------------------|--------------------|
| 1 | LDA | THF | [Pd(C ₃ H ₅)Cl] ₂ | L1 | 50 | 32 |
| 2 | LDA | THF | Pd ₂ (dba) ₃ | L1 | 90 | 34 |
| 3 | LDA | Et ₂ O | Pd(OAc) ₂ | L1 | 90 | 32 |
| 4 | LDA | toluene | Pd(OAc) ₂ | L1 | 79 | 37 |
| 5 | LDA | DME | Pd(OAc) ₂ | L1 | 91 | 42 |
| 6 | LDA | DCM | Pd(OAc) ₂ | L1 | NR | — |
| 7 | LiHMDS | DME | Pd(OAc) ₂ | L1 | 94 | 46 |
| 8 | NaHMDS | DME | Pd(OAc) ₂ | L1 | 90 | 29 |
| 9 | KHMDS | DME | Pd(OAc) ₂ | L1 | trace | — |
| 10 | ^t BuOK | DME | Pd(OAc) ₂ | L1 | trace | — |
| 11 | Cs ₂ CO ₃ | DME | Pd(OAc) ₂ | L1 | NR | — |
| 12 | LiHMDS | DME | Pd(OAc) ₂ | L9 | 94 | 48 |
| 13 | LiHMDS | DME | Pd(OAc) ₂ | L10 | 98 | 41 |
| 14 | LiHMDS | DME | Pd(OAc) ₂ | L11 | 99 | 51 |
| 15 | LiHMDS | DME | Pd(OAc) ₂ | L12 | 72 | 23 |
| 16 | LiHMDS | DME | Pd(OAc) ₂ | L13 | 75 | 12 |
| 17 | LiHMDS | DME | Pd(OAc) ₂ | L14 | NR | — |
| 18 | LiHMDS | DME | Pd(OAc) ₂ | L15 | NR | — |

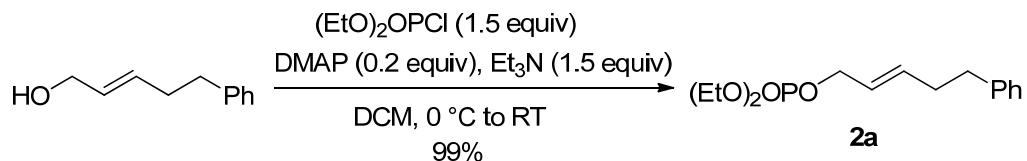
^aMolar ratio of **1a**/**2a**/base/[Pd]/ligand/additive = 130/100/130/5/5/100. ^bThe yield determined by ¹H NMR using mesitylene as internal standard. ^cDetermined by chiral HPLC.

3. Effect of amide on the Pd-catalyzed asymmetric allylic alkylation



4. Synthesis of allylic reagents 2

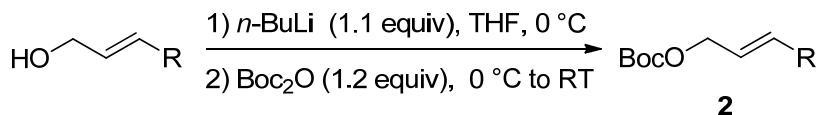
Experimental procedure for the preparation of allylic reagent **2a**¹



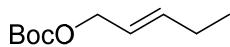
4-Dimethylaminopyridine (257 mg, 2.1 mmol) and triethylamine (2.2 mL, 15.7 mmol) were added to a solution of (*E*)-5-phenylpent-2-en-1-ol (1.7 g, 10.5 mmol) in CH₂Cl₂ (35 mL). The resulting solution was cooled to 0 °C. Diethyl chlorophosphate (2.3 mL, 15.7 mmol) was added dropwise, and the reaction mixture was stirred overnight at room temperature. The reaction was quenched with saturated aqueous NH₄Cl and extracted with EtOAc. The organic layer was dried over anhydrous MgSO₄, filtered, and concentrated under vacuum. The residue was purified by chromatography on silica gel with hexane/EtOAc = 1/1 to afford **2a**² as a colorless oil (3.12g, 99% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.28 (dd, *J* = 8.1, 6.3 Hz, 2H), 7.21–7.14 (m, 3H), 5.89–5.79 (m, 1H), 5.69–5.59 (m, 1H), 4.52–4.43 (m, 2H), 4.16–4.03 (m, 4H), 2.77–2.65 (m, 2H), 2.39 (dd, *J* = 15.0, 7.1 Hz, 2H), 1.33 (td, *J* = 7.1, 0.9 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 141.40, 135.26, 128.37, 128.33, 125.92, 125.05 (d, *J* = 6.9 Hz), 67.97 (d, *J* = 5.4 Hz), 63.73 (d, *J* = 5.6 Hz), 35.21, 33.90, 16.17 (d, *J* = 6.9 Hz).

General experimental procedure for the preparation of allylic *tert*-butyl carbonates³

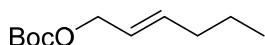


To a solution of allylic alcohol (10 mmol) in THF (45 mL) under a nitrogen atmosphere was added *n*-BuLi (2.0 M in hexane, 11 mmol) at 0 °C and the mixture was stirred for 30 min. Boc₂O (12 mmol) was added at 0 °C and the reaction mixture was stirred overnight at room temperature. The reaction was quenched with H₂O and the solvent was removed in vacuo. The residue was diluted with ethanol (15 mL) and imidazole (3 mmol) was added to remove the unreacted Boc₂O. After stirring for 1 h at room temperature, the solvent was removed and the residue was purified by flash column chromatography (petroleum ether: ethyl acetate = 100:1) to give the corresponding allylic *tert*-butyl carbonate.



2b

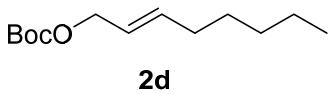
(E)-*tert*-butyl pent-2-en-1-yl carbonate: General procedure was followed on a 10.0 mmol scale, a colorless oil (1.7 g, 95%). ¹H NMR (400 MHz, CDCl₃) δ 5.91–5.77 (m, 1H), 5.65–5.51 (m, 1H), 4.50 (d, *J* = 6.6 Hz, 2H), 2.14–2.01 (m, 2H), 1.49 (s, 9H), 1.00 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 153.30, 138.28, 122.52, 81.69, 67.60, 27.67, 25.14, 12.94; IR: 2967, 1738, 1458, 1369, 1272, 1251, 1159, 1102, 857, 793; MS (EI): m/z (%) 69 (58), 57 (100), 55 (5), 42 (4); HRMS (DART) m/z Calcd for C₁₀H₁₉O₃ [M+H]⁺: 187.1329, found 187.1329.



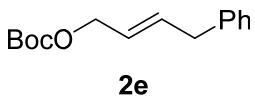
2c

(E)-*tert*-butyl hex-2-en-1-yl carbonate: General procedure was followed on a 8.5 mmol scale, a colorless oil (1.6 g, 95%). ¹H NMR (400 MHz, CDCl₃) δ 5.84–5.74 (m, 1H), 5.58 (ddd, *J* = 13.9, 7.2, 5.9 Hz, 1H), 4.50 (d, *J* = 6.6 Hz, 2H), 2.03 (q, *J* = 7.1 Hz, 2H), 1.48 (s, 9H), 1.46–1.35 (m, 2H), 0.90 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (101

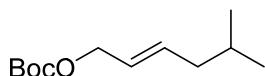
MHz, CDCl₃) δ 153.36, 136.86, 123.60, 81.87, 67.71, 34.26, 27.74, 21.96, 13.60; IR: 2962, 1738, 1458, 1369, 1273, 1251, 1159, 1104, 971, 928, 858, 793, 761; MS (EI): m/z (%) 200 (M⁺, 0.1), 144 (13), 83 (53), 57 (100), 56 (21), 43 (7); HRMS (DART) m/z Calcd for C₁₁H₂₄O₃N [M+NH₄]⁺: 218.1751, found 218.1751.



(E)-tert-butyl oct-2-en-1-yl carbonate: General procedure was followed on a 13.3 mmol scale, a colorless oil (2.8 g, 92%). ¹H NMR (400 MHz, CDCl₃) δ 5.85–5.74 (m, 1H), 5.63–5.53 (m, 1H), 4.50 (d, J = 6.6 Hz, 2H), 2.04 (q, J = 7.0 Hz, 2H), 1.49 (s, 9H), 1.43–1.33 (m, 2H), 1.33–1.22 (m, 4H), 0.88 (t, J = 6.9 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 153.37, 137.17, 123.38, 81.89, 67.75, 32.18, 31.30, 28.47, 27.75, 22.47, 14.00; IR: 2858, 1739, 1458, 1369, 1272, 1251, 1159, 1102, 1036, 972, 928, 859, 793; MS (EI): m/z (%) 228 (M⁺, 0.03), 144 (2), 117 (1), 84 (2), 71 (3), 57 (100), 41 (10); HRMS (DART) m/z Calcd for C₁₃H₂₈O₃N [M+NH₄]⁺: 246.2064, found 246.2064.

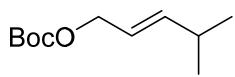


(E)-tert-butyl (4-phenylbut-2-en-1-yl) carbonate: General procedure was followed on a 2.4 mmol scale, a colorless oil (0.45 g, 77%). ¹H NMR (400 MHz, CDCl₃) δ 7.28 (dd, J = 13.8, 6.1 Hz, 2H), 7.24–7.14 (m, 3H), 5.95 (dt, J = 13.9, 6.7 Hz, 1H), 5.65 (dt, J = 13.6, 6.5 Hz, 1H), 4.53 (d, J = 6.4 Hz, 2H), 3.40 (d, J = 6.7 Hz, 2H), 1.48 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 153.33, 139.45, 135.02, 128.60, 128.45, 126.21, 124.97, 82.08, 67.34, 38.61, 27.76; IR: 2979, 1737, 1454, 1369, 1273, 1251, 1157, 1090, 971, 857, 793, 747, 698, 461; MS (EI): m/z (%) 248 (M⁺, 0.02), 131 (45), 104 (4), 91 (23), 77 (4), 57 (30); HRMS (DART) m/z Calcd for C₁₅H₂₁O₃ [M+H]⁺: 249.1485, found 249.1485.



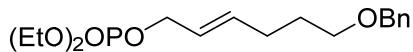
2f

(E)-tert-butyl (5-methylhex-2-en-1-yl) carbonate: General procedure was followed on a 3.7 mmol scale, a colorless oil (0.73 g, 92%). ^1H NMR (400 MHz, CDCl_3) δ 5.77 (dt, $J = 14.4, 7.2$ Hz, 1H), 5.62–5.52 (m, 1H), 4.50 (d, $J = 6.5$ Hz, 2H), 1.94 (t, $J = 6.9$ Hz, 2H), 1.68–1.60 (m, 1H), 1.49 (s, 9H), 0.88 (d, $J = 6.6$ Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 153.32, 135.63, 124.62, 81.75, 67.58, 41.51, 28.00, 27.70, 22.19; IR: 2957, 1738, 1462, 1368, 1274, 1251, 1159, 1100, 972, 858, 793, 758; MS (DART): m/z (%) 214 (M^+ , 0.2), 97 (60), 57 (100), 43 (24); HRMS (EI) m/z Calcd for $\text{C}_{12}\text{H}_{26}\text{O}_3\text{N} [\text{M}+\text{NH}_4]^+$: 232.1907, found 232.1907.



2g

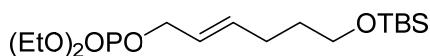
(E)-tert-butyl (4-methylpent-2-en-1-yl) carbonate: General procedure was followed on a 2.6 mmol scale, a colorless oil (0.42 g, 80%). ^1H NMR (400 MHz, CDCl_3) δ 5.76 (ddt, $J = 15.4, 6.4, 1.1$ Hz, 1H), 5.53 (dtd, $J = 15.4, 6.6, 1.4$ Hz, 1H), 4.52–4.48 (m, 2H), 2.31 (dq, $J = 13.6, 6.8$ Hz, 1H), 1.49 (s, 9H), 0.99 (d, $J = 6.8$ Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 153.36, 143.73, 120.55, 81.96, 67.89, 30.75, 27.77, 21.93; IR: 2961, 1738, 1462, 1368, 1273, 1251, 1159, 1090, 971, 927, 860, 793, 763; MS (EI): m/z (%) 200 (M^+ , 1.1), 83 (14), 69 (7), 57 (100), 56 (25), 43 (28); HRMS (DART) m/z Calcd for $\text{C}_{11}\text{H}_{24}\text{O}_3\text{N} [\text{M}+\text{NH}_4]^+$: 218.1751, found 218.1751.



2h

(E)-6-(benzyloxy)hex-2-en-1-yl diethyl phosphate: Procedure was followed as the preparation of **2a** on a 4.85 mmol scale, a colorless oil (1.68 g, 99%). ^1H NMR (400 MHz, CDCl_3) δ 7.42–7.25 (m, 5H), 5.85–5.75 (m, 1H), 5.66–5.57 (m, 1H), 4.53–4.42 (m, 4H), 4.10 (p, $J = 7.2$ Hz, 4H), 3.47 (t, $J = 6.4$ Hz, 2H), 2.17 (q, $J = 7.1$ Hz, 2H),

1.78–1.64 (m, 2H), 1.33 (t, J = 7.1 Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 138.47, 135.58, 128.34, 127.59, 127.52, 124.88 (d, J = 6.7 Hz), 72.88, 69.48, 67.99 (d, J = 5.1 Hz), 63.67 (d, J = 6.6 Hz), 28.89, 28.79, 16.16 (d, J = 6.7 Hz); ^{31}P NMR (162 MHz, CDCl_3) δ -0.83; IR: 2982, 2936, 2855, 1453, 1392, 1367, 1263, 1166, 1099, 1024, 967, 845, 800, 736, 698, 610, 515; MS (ESI) m/z: 365.1 ($\text{M}+\text{Na}^+$); HRMS (ESI) m/z Calcd for $\text{C}_{17}\text{H}_{31}\text{NO}_5\text{P}$ ($\text{M}+\text{NH}_4^+$): 360.1934, found 360.1932.

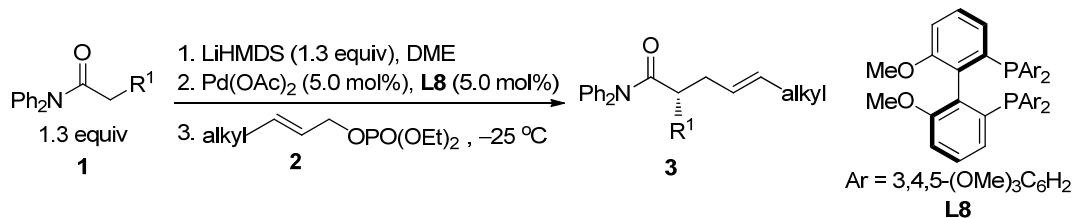


2i

(E)-6-((tert-butyldimethylsilyl)oxy)hex-2-en-1-yl diethyl phosphate: Procedure was followed as the preparation of **2a** on a 2.6 mmol scale, a colorless oil (0.95g, 99%). ^1H NMR (400 MHz, CDCl_3) δ 5.87–5.76 (m, 1H), 5.68–5.57 (m, 1H), 4.48 (t, J = 7.2 Hz, 2H), 4.11 (q, J = 7.3 Hz, 4H), 3.61 (t, J = 6.3 Hz, 2H), 2.13 (dd, J = 14.7, 7.2 Hz, 2H), 1.66–1.56 (m, 2H), 1.34 (t, J = 7.1 Hz, 6H), 0.89 (s, 9H), 0.04 (s, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 136.01, 124.62 (d, J = 7.0 Hz), 68.10 (d, J = 4.7 Hz), 63.70 (d, J = 5.8 Hz), 62.36, 31.87, 28.49, 25.93, 16.18 (d, J = 6.5 Hz), -5.30; ^{31}P NMR (162 MHz, CDCl_3) δ -0.83; IR: 2930, 1470, 1254, 1098, 967, 834, 774, 662, 526; MS (ESI) m/z: 389.1 ($\text{M}+\text{Na}^+$); HRMS (ESI) m/z Calcd for $\text{C}_{16}\text{H}_{39}\text{NO}_5\text{PSi}$ ($\text{M}+\text{NH}_4^+$): 384.233, found 384.2327.

5 General experimental procedure and characterization of products

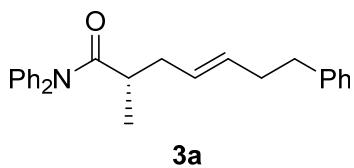
5.1 General experimental procedure



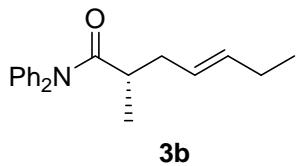
A dry Schlenk tube was flame dried and flushed with Argon. LiCl (8.4 mg), amide **1** (0.26 mmol) and DME (1.8 mL) were added into the dry Schlenk tube. LiHMDS (1.0 M in THF, 0.26 mL, 0.26 mmol) were added at 0 °C, and stirred at room temperature for 30 min. In a separated flushed flask, Pd(OAc)₂ (2.25 mg, 0.01 mmol), **L8** (9.43, 0.01 mmol) and DME (1.0 mL) were added. The resulting mixture was stirred at room temperature for 30 min. Then the in-situ generated catalyst was added to the enolate solution. The allylic substrates **2** (0.2 mmol) was then added and the mixture was stirred at -25 °C. After the reaction was complete (monitored by TLC), the reaction mixture was quenched by H₂O (0.3 mL). The solution was dried (anhydrous Na₂SO₄) and then filtered through a 0.5 inch plug of silica gel (eluting with EtOAc) to remove the solid. The crude reaction mixture was concentrated under reduced pressure. The resulting crude product was purified by flash column chromatography on silica gel (eluting with petroleum ether/ethyl acetate 10/1) to yield the product.

Special Note: ¹³C NMR spectra of products **3** show broad peaks around 125–130 ppm, which was assigned as peaks of phenyl groups on nitrogen of the amide. To better understand these phenomena, the ¹³C NMR spectrum of *N,N*-diphenylpropionamide **1a** was measured at 50 °C and -30 °C respectively. It revealed that the broad peaks around 125–130 ppm were observed for ¹³C NMR spectrum measured at 50 °C, while six single peaks around 125–130 ppm were observed for ¹³C NMR spectrum measured at -30 °C (these spectra was included in the spectra section). These phenomena may attributed to rapid rotation of two phenyl groups on nitrogen of the amide at room temperature and 50 °C, while the rotation was significantly slowed down at -30 °C.

5.2 characterization of products

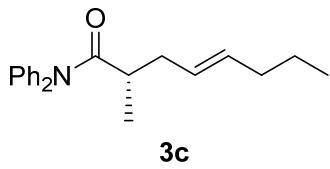


colorless oil, 69.5 mg, yield: 95%, 95:5 er. $[\alpha]_D^{31} = 63.6$ (1.0, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.40–7.14 (m, 15H), 5.58–5.47 (m, 1H), 5.40–5.29 (m, 1H), 2.71–2.64 (m, 2H), 2.59 (dt, $J = 13.1, 6.6$ Hz, 1H), 2.43 (dt, $J = 14.7, 7.5$ Hz, 1H), 2.33 (dd, $J = 14.9, 7.2$ Hz, 2H), 2.09–1.98 (m, 1H), 1.10 (d, $J = 6.7$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3): δ 176.61, 143.00, 141.95, 131.81, 129.68, 128.80, 128.42, 128.31, 127.97, 126.49, 125.81, 37.81, 37.72, 35.98, 34.52, 17.68; IR: 3062, 3026, 2969, 2930, 2849, 1669, 1591, 1490, 1452, 1379, 1254, 968, 755, 693, 620 cm^{-1} ; MS (ESI) m/z: 370.1 ($\text{M}+\text{H}$) $^+$, 392.1 ($\text{M}+\text{Na}$) $^+$; HRMS (ESI) m/z Calcd for $\text{C}_{26}\text{H}_{28}\text{NO}$ ($\text{M}+\text{H}$) $^+$: 370.2165, found 370.2164; HPLC (Chiralpak OD-H, Hexane:*i*-Propanol = 99:1, 1 mL/min, 214 nm): $t_{\text{major}} = 21.22$ min, $t_{\text{minor}} = 24.70$ min.

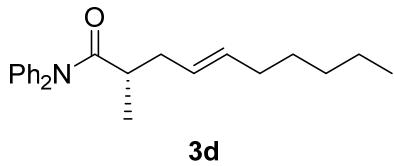


colorless oil, 48.9 mg, yield: 84%, 95.7:4.3 er. $[\alpha]_D^{31} = 97.5$ (1.0, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.26 (dd, $J = 28.0, 20.2$ Hz, 10H), 5.59–5.49 (m, 1H), 5.37–5.27 (m, 1H), 2.75–2.55 (m, 1H), 2.52–2.37 (m, 1H), 2.11–1.96 (m, 3H), 1.14 (d, $J = 6.7$ Hz, 3H), 1.00 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 176.68, 143.05, 134.33, 128.83, 126.19, 37.88, 37.84, 25.59, 17.73, 13.80; IR: 3063, 3029, 2962, 2931, 2873, 2850, 1669, 1591, 1490, 1452, 1379, 1256, 1162, 1074, 1029, 967, 755, 691, 665, 649, 620 cm^{-1} ; MS (ESI) m/z: 294.1 ($\text{M}+\text{H}$) $^+$, 316.1 ($\text{M}+\text{Na}$) $^+$; HRMS (ESI) m/z

Calcd for C₂₀H₂₄NO (M+H)⁺: 294.1852, found 294.1852; HPLC (Chiralpak OD-H, Hexane:*i*-Propanol = 99:1, 1 mL/min, 214 nm): t_{major} = 9.88 min, t_{minor} = 11.17 min.

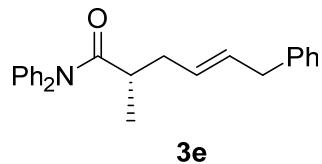


colorless oil, 56.9 mg, yield: 92%, 96:4 er. [α]_D³¹ = 79.5 (1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.34–7.22 (m, 10H), 5.54–5.44 (m, 1H), 5.38–5.28 (m, 1H), 2.75–2.55 (m, 1H), 2.52–2.30 (m, 1H), 2.15–1.95 (m, 3H), 1.48–1.34 (m, 2H), 1.14 (d, *J* = 6.7 Hz, 3H), 0.91 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 176.68, 143.03, 132.67, 129.59, 128.77, 127.58, 127.35, 126.48, 126.23, 125.89, 125.48, 37.86, 37.85, 34.74, 22.61, 17.74, 13.78; IR: 3063, 3037, 2959, 2929, 2872, 2841, 1671, 1591, 1490, 1452, 1379, 1254, 967, 755, 691, 620 cm⁻¹; MS (ESI) m/z: 308.1 (M+H)⁺, 330.1 (M+Na)⁺; HRMS (ESI) m/z Calcd for C₂₁H₂₆NO (M+H)⁺: 308.2009, found 308.2009; HPLC (Chiralpak OD-H, Hexane:*i*-Propanol = 99:1, 1 mL/min, 214 nm): t_{major} = 9.58 min, t_{minor} = 10.58 min.

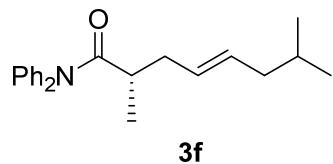


colorless oil, 55.0 mg, yield: 82%, 94.6:5.4 er. [α]_D³¹ = 76.3 (1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.21 (m, 10H), 5.54–5.43 (m, 1H), 5.37–5.26 (m, 1H), 2.75–2.52 (m, 1H), 2.53–2.35 (m, 1H), 2.11–1.93 (m, 3H), 1.47–1.21 (m, 6H), 1.12 (d, *J* = 6.7 Hz, 3H), 0.88 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 176.68, 143.04, 132.92, 129.60, 128.77, 127.69, 127.12, 126.52, 125.95, 37.87, 32.62, 31.45, 29.20, 22.58, 17.75, 14.08; IR: 3063, 3037, 2957, 2925, 2872, 2854, 1672, 1591, 1490,

1452, 1378, 1253, 1160, 1074, 968, 902, 755, 699, 691, 665, 649, 620 cm^{-1} ; MS (ESI) m/z: 336.2 ($\text{M}+\text{H}$)⁺, 358.1 ($\text{M}+\text{Na}$)⁺; HRMS(ESI) m/z Calcd for $\text{C}_{23}\text{H}_{30}\text{NO}$ ($\text{M}+\text{H}$)⁺: 336.2322, found 336.2322; HPLC (Chiraldak IG, Hexane:*i*-Propanol = 90:10, 0.7 mL/min, 214 nm): $t_{\text{major}} = 13.12$ min, $t_{\text{minor}} = 14.82$ min.

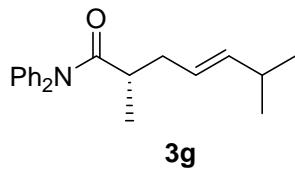


colorless oil, 59.5 mg, yield: 83%, 95.5:4.5 er. $[\alpha]_D^{31} = 59.1$ (1.0, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39–7.12 (m, 15H), 5.73–5.62 (m, 1H), 5.46–5.35 (m, 1H), 3.34 (d, $J = 6.6$ Hz, 2H), 2.69–2.55 (m, 1H), 2.55–2.42 (m, 1H), 2.13–1.97 (m, 1H), 1.13 (d, $J = 6.7$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 176.44, 142.97, 140.59, 131.69, 131.22, 128.84, 128.60, 128.43, 125.99, 39.01, 37.79, 37.71, 17.97; IR: 3055, 3028, 2975, 2960, 2926, 2868, 2834, 1658, 1590, 1490, 1452, 1427, 1386, 1277, 1180, 1154, 1072, 1028, 969, 907, 773, 747, 715, 692, 648, 621 cm^{-1} ; MS (ESI) m/z: 356.1 ($\text{M}+\text{H}$)⁺, 378.1 ($\text{M}+\text{Na}$)⁺; HRMS (ESI) m/z Calcd for $\text{C}_{25}\text{H}_{26}\text{NO}$ ($\text{M}+\text{H}$)⁺: 356.2009, found 356.2007; HPLC (Chiraldak OD-H, Hexane:*i*-Propanol = 99:1, 1 mL/min, 214 nm): $t_{\text{major}} = 18.60$ min, $t_{\text{minor}} = 22.79$ min.

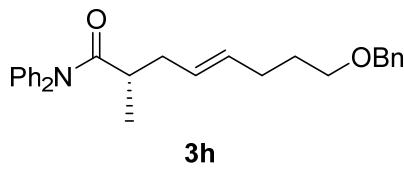


colorless oil, 57.8 mg, yield: 89%, 96.4:3.6 er. $[\alpha]_D^{31} = 68.5$ (0.8, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.33–7.22 (m, 10H), 5.46 (dt, $J = 14.3, 7.1$ Hz, 1H), 5.35–5.25 (m, 1H), 2.73–2.54 (m, 1H), 2.45 (dt, $J = 14.7, 7.6$ Hz, 1H), 2.04 (dt, $J = 13.2, 6.6$ Hz, 1H), 1.99–1.80 (m, 2H), 1.70–1.50 (m, 1H), 1.12 (d, $J = 6.7$ Hz, 3H), 0.87 (d, $J = 6.7$

Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 176.68, 142.99, 131.66, 131.58, 128.78, 128.31, 126.49, 42.06, 37.88, 37.82, 28.43, 22.42, 22.32, 17.75; IR: 3063, 3038, 2955, 2930, 2899, 2869, 2837, 1671, 1591, 1490, 1452, 1379, 1252, 1163, 969, 755, 700, 691, 665, 649, 620 cm^{-1} ; MS (ESI) m/z: 322.1 ($\text{M}+\text{H}$) $^+$, 344.1 ($\text{M}+\text{Na}$) $^+$; HRMS (ESI) m/z Calcd for $\text{C}_{22}\text{H}_{28}\text{NO}$ ($\text{M}+\text{H}$) $^+$: 322.2165, found 322.2164; HPLC (Chiralpak OD-H, Hexane:*i*-Propanol = 99:1, 1mL/min, 214nm): $t_{\text{major}} = 9.51$ min, $t_{\text{minor}} = 10.63$ min.

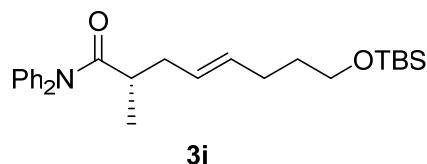


colorless oil, 55.2 mg, yield: 90%, 96:4 er. $[\alpha]_D^{25} = 94.9$ (1.0, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.50–7.09 (m, 10H), 5.47 (dd, $J = 15.3, 6.5$ Hz, 1H), 5.33–5.22 (m, 1H), 2.68–2.55 (m, 1H), 2.50–2.38 (m, 1H), 2.26 (dq, $J = 13.4, 6.7$ Hz, 1H), 2.08–1.95 (m, 1H), 1.13 (d, $J = 6.7$ Hz, 3H), 0.98 (d, $J = 6.8$ Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 176.73, 142.99, 139.84, 134.86, 128.73, 126.49, 124.14, 119.82, 37.92, 31.03, 22.62, 22.55, 17.87; IR: 2958, 2930, 1671, 1591, 1490, 1452, 1379, 1257, 970, 756, 692, 620, 522; MS (ESI) m/z: 308.2 ($\text{M}+\text{H}$) $^+$, 330.1 ($\text{M}+\text{Na}$) $^+$; HRMS (ESI) m/z Calcd for $\text{C}_{21}\text{H}_{26}\text{NO}$ ($\text{M}+\text{H}$) $^+$: 308.2009, found 308.2011; HPLC (Chiralpak OD-H, Hexane:*i*-Propanol = 99:1, 1 mL/min, 214 nm): $t_{\text{major}} = 9.25$ min, $t_{\text{minor}} = 10.79$ min.

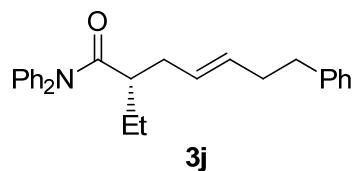


colorless oil, 74.9 mg, yield: 91%, 96:4 er. $[\alpha]_D^{27} = 60.80$ (1.0, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.52–7.06 (m, 15H), 5.53–5.42 (m, 1H), 5.38–5.27 (m, 1H), 4.48 (s, 2H), 3.46 (t, $J = 6.5$ Hz, 2H), 2.73–2.52 (m, 1H), 2.51–2.35 (m, 1H), 2.10 (dd, $J =$

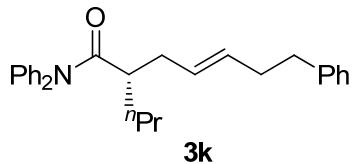
14.4, 7.1 Hz, 2H), 2.07–1.98 (m, 1H), 1.76–1.60 (m, 2H), 1.11 (d, J = 6.7 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 176.61, 143.01, 138.61, 132.02, 128.82, 128.37, 127.80, 127.64, 127.52, 126.49, 72.86, 69.75, 37.81, 37.79, 29.52, 29.22, 17.72; IR: 2932, 2850, 1670, 1590, 1490, 1452, 1378, 1258, 1101, 968, 756, 697, 620, 522 cm^{-1} ; MS (ESI) m/z: 414.2 ($\text{M}+\text{H}$) $^+$, 436.2 ($\text{M}+\text{Na}$) $^+$; HRMS (ESI) m/z Calcd for $\text{C}_{28}\text{H}_{32}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 414.2428, found 414.2425; HPLC (Chiralpak OD-H, Hexane:*i*-Propanol = 95:5, 0.7 mL/min, 214 nm): $t_{\text{major}} = 17.53$ min, $t_{\text{minor}} = 19.26$ min.



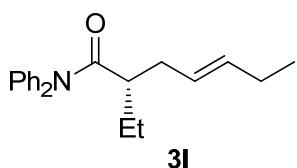
colorless oil, 76.2 mg, yield: 88%, 92.8:7.2 er. $[\alpha]_D^{27} = 55.3$ (1.0, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.33–7.21 (m, 10H), 5.54–5.44 (m, 1H), 5.39–5.27 (m, 1H), 3.60 (t, J = 6.4 Hz, 2H), 2.70–2.55 (m, 1H), 2.51–2.35 (m, 1H), 2.13–1.97 (m, 3H), 1.67–1.51 (m, 2H), 1.12 (d, J = 6.7 Hz, 3H), 0.89 (s, 9H), 0.04 (s, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 176.63, 143.00, 132.23, 129.63, 128.77, 127.52, 126.51, 62.59, 37.81, 32.59, 28.86, 25.97, 18.34, 17.68, -5.24; IR: 2929, 2855, 1673, 1592, 1491, 1380, 1251, 1098, 968, 834, 774, 755, 700, 620, 521 cm^{-1} ; MS (ESI) m/z: 438.3 ($\text{M}+\text{H}$) $^+$, 460.2 ($\text{M}+\text{Na}$) $^+$; HRMS (ESI) m/z Calcd for $\text{C}_{27}\text{H}_{40}\text{NO}_2\text{Si}$ ($\text{M}+\text{H}$) $^+$: 438.2823, found 438.2819; HPLC (Chiralpak OD-H, Hexane:*i*-Propanol=99:1, 1.0 mL/min, 214 nm): $t_{\text{major}} = 9.63$ min, $t_{\text{minor}} = 11.16$ min.



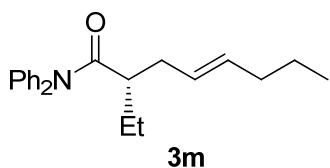
colorless oil, 68.6 mg, yield: 91%, 94.7:5.3 er. $[\alpha]_D^{31} = -22.2$ (1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.50–7.14 (m, 15H), 5.62–5.51 (m, 1H), 5.46–5.34 (m, 1H), 2.72 (t, *J* = 7.8 Hz, 2H), 2.51 (dq, *J* = 8.3, 5.6 Hz, 1H), 2.45–2.32 (m, 3H), 2.25–2.08 (m, 1H), 1.75 (tt, *J* = 15.4, 7.5 Hz, 1H), 1.57–1.37 (m, 1H), 0.95 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 175.93, 143.03, 141.94, 131.72, 129.61, 129.26, 128.82, 128.43, 128.32, 128.10, 127.74, 126.55, 125.82, 44.65, 36.30, 35.96, 34.56, 25.97, 12.12; IR: 3026, 2961, 2927, 2854, 1667, 1591, 1490, 1453, 1385, 1275, 1239, 969, 754, 693, 619 cm⁻¹; MS (ESI) m/z: 384.2 (M+H)⁺, 406.1 (M+Na)⁺; HRMS (ESI) m/z Calcd for C₂₇H₃₀NO (M+H)⁺: 384.2322, found 384.2321; HPLC (Chiralpak IG, Hexane:*i*-Propanol = 90:10, 0.7 mL/min, 214 nm): t_{major} = 19.09 min, t_{minor} = 25.35 min.



colorless oil, 76.2 mg, yield: 97%, 95:5 er. $[\alpha]_D^{31} = -27.6$ (1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.53–7.11 (m, 15H), 5.61–5.50 (m, 1H), 5.45–5.34 (m, 1H), 2.71 (t, *J* = 7.8 Hz, 2H), 2.66–2.50 (m, 1H), 2.49–2.31 (m, 3H), 2.13 (dt, *J* = 13.2, 6.4 Hz, 1H), 1.82–1.65 (m, 1H), 1.47–1.24 (m, 3H), 0.86 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 176.07, 143.01, 141.94, 134.77, 131.72, 129.59, 129.22, 128.81, 128.43, 128.32, 128.11, 127.74, 126.54, 125.82, 43.00, 36.51, 35.96, 35.21, 34.54, 20.79, 14.31; IR: 3027, 2956, 2927, 2856, 1668, 1591, 1490, 1452, 1383, 1259, 970, 754, 697, 662, 620 cm⁻¹; MS (ESI) m/z: 398.1 (M+H)⁺, 420.1 (M+Na)⁺; HRMS (ESI) m/z Calcd for C₂₈H₃₂NO (M+H)⁺: 398.2478, found 398.2477; HPLC (Chiralpak IC, Hexane:*i*-Propanol = 99:1, 1 mL/min, 214 nm): t_{minor} = 30.59 min, t_{major} = 33.32 min.

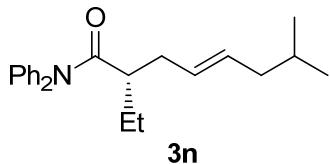


colorless oil, 45.8 mg, yield: 74%, 92.4:7.6 er. $[\alpha]_D^{28} = -7.7$ (1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.49–7.11 (m, 10H), 5.60–5.48 (m, 1H), 5.39–5.28 (m, 1H), 2.58–2.44 (m, 1H), 2.44–2.34 (m, 1H), 2.17–2.00 (m, 3H), 1.74 (td, *J* = 14.9, 7.7 Hz, 1H), 1.54–1.39 (m, 1H), 1.01 (t, *J* = 7.4 Hz, 3H), 0.94 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 175.99, 134.22, 129.54, 128.27, 126.55, 126.31, 44.71, 36.38, 26.09, 25.61, 13.77, 12.10; IR: 2961, 2930, 2873, 1667, 1591, 1490, 1454, 1383, 1275, 1239, 1161, 1074, 1029, 968, 754, 693, 662, 619, 521, 464 cm⁻¹; MS (ESI) m/z: 308.2 (M+H)⁺, 330.1 (M+Na)⁺; HRMS (ESI) m/z Calcd for C₂₁H₂₆NO (M+H)⁺: 308.2009, found 308.2003; HPLC (Chiralpak IG, Hexane:*i*-Propanol = 90:10, 0.7 mL/min, 214 nm): t_{major} = 15.69 min, t_{minor} = 20.15 min.

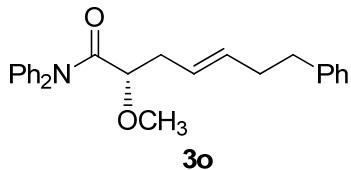


colorless oil, 50.1 mg, yield: 80%, 91.3:8.7 er. $[\alpha]_D^{28} = -18.8$ (1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.48–7.06 (m, 10H), 5.52–5.42 (m, 1H), 5.37–5.27 (m, 1H), 2.53–2.44 (m, 1H), 2.43–2.33 (m, 1H), 2.15–2.05 (m, 1H), 1.99 (dd, *J* = 14.0, 7.1 Hz, 2H), 1.84–1.66 (m, 1H), 1.52–1.32 (m, 3H), 0.91 (q, *J* = 7.7 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 175.97, 143.05, 132.56, 129.26, 128.76, 127.45, 126.56, 44.69, 36.37, 34.76, 26.06, 22.60, 13.79, 12.08; IR: 2959, 2927, 2872, 1668, 1591, 1490, 1453, 1382, 1274, 1239, 1161, 1074, 1029, 968, 754, 693, 662, 618, 520 cm⁻¹; MS (ESI) m/z: 322.2 (M+H)⁺, 344.2 (M+Na)⁺; HRMS (ESI) m/z Calcd for C₂₂H₂₈NO (M+H)⁺:

322.2165, found 322.2159; HPLC (Chiralpak IE, Hexane:*i*-Propanol = 95:5, 0.7 mL/min, 214 nm): $t_{\text{major}} = 24.67 \text{ min}$, $t_{\text{minor}} = 26.57 \text{ min}$.



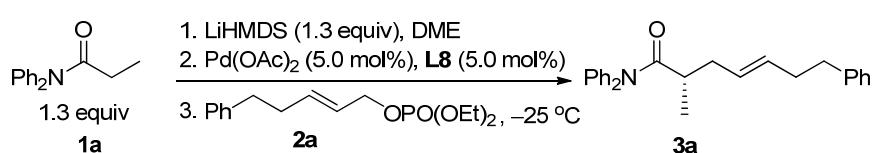
colorless oil, 47.8 mg, yield: 72%, 92.9:7.1 er. $[\alpha]_D^{28} = -34.4$ (1.0, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.55–6.98 (m, 10H), 5.46 (dt, $J = 14.3, 6.9 \text{ Hz}$, 1H), 5.37–5.27 (m, 1H), 2.54–2.45 (m, 1H), 2.40 (dt, $J = 14.6, 7.4 \text{ Hz}$, 1H), 2.22–2.07 (m, 1H), 1.98–1.83 (m, 2H), 1.74 (tt, $J = 15.0, 7.5 \text{ Hz}$, 1H), 1.61 (tt, $J = 13.2, 6.7 \text{ Hz}$, 1H), 1.53–1.41 (m, 1H), 0.99–0.78 (m, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 175.93, 143.05, 131.46, 129.23, 128.77, 128.40, 126.54, 44.68, 42.06, 36.32, 28.44, 26.04, 22.40, 22.33, 12.06; IR: 2957, 2926, 2870, 2837, 1669, 1592, 1490, 1454, 1383, 1273, 1239, 1162, 1074, 1029, 970, 754, 693, 662, 619, 520 cm^{-1} ; MS (ESI) m/z: 336.2 ($\text{M}+\text{H}$) $^+$, 358.2 ($\text{M}+\text{Na}$) $^+$; HRMS (ESI) m/z Calcd for $\text{C}_{23}\text{H}_{30}\text{NO}$ ($\text{M}+\text{H}$) $^+$: 336.2322, found 336.2315; HPLC (Chiralpak IE, Hexane:*i*-Propanol = 95:5, 0.7 mL/min, 214 nm): $t_{\text{major}} = 21.89 \text{ min}$, $t_{\text{minor}} = 23.85 \text{ min}$.



colorless oil, 59.7 mg, yield: 79%, 94.4:5.6 er. $[\alpha]_D^{26} = 54.4$ (1.0, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.46–7.11 (m, 15H), 5.58–5.47 (m, 1H), 5.45–5.35 (m, 1H), 3.92 (t, $J = 6.4 \text{ Hz}$, 1H), 3.30 (s, 3H), 2.65 (dd, $J = 8.5, 6.8 \text{ Hz}$, 2H), 2.53–2.35 (m, 2H), 2.32 (dd, $J = 14.9, 7.1 \text{ Hz}$, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.52, 141.86, 132.95, 128.96, 128.44, 128.30, 125.81, 125.36, 78.31, 57.06, 35.78, 35.65, 34.47; IR:

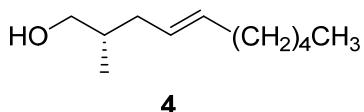
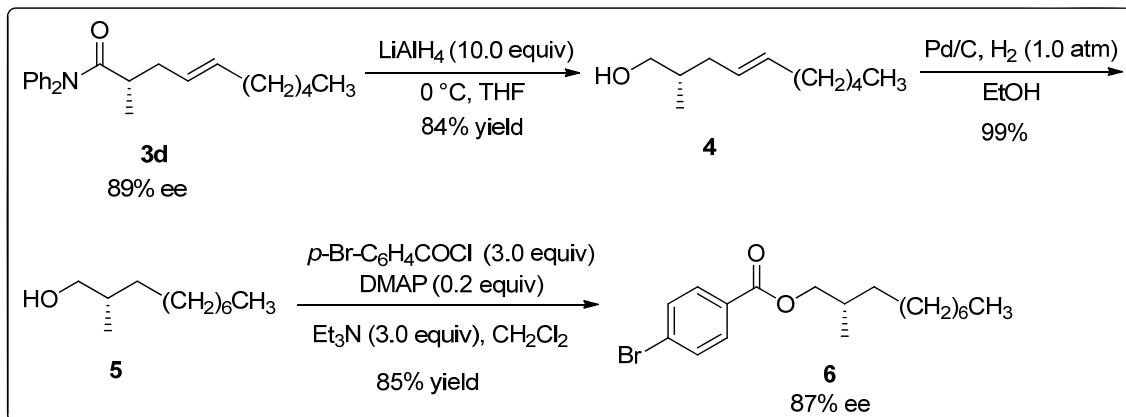
3061, 2923, 1679, 1591, 1490, 1379, 1270, 1114, 969, 753, 698, 618, 568 cm⁻¹; MS (ESI) m/z: 386.2 (M+H)⁺, 408.2 (M+Na)⁺; HRMS (ESI) m/z Calcd for C₂₆H₂₈NO₂ (M+H)⁺: 386.2115, found 386.2125; HPLC (Chiralpak OD-H, Hexane:*i*-Propanol = 90:10, 1.0 mL/min, 214 nm): t_{major} = 13.04 min, t_{minor} = 17.16 min.

6. Pd-Catalyzed Asymmetric Allylic Alkylation of Amide **1a** with Allyl Reagents **2a** on 1.0 mmol Scale.

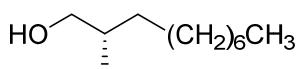


A dry Schlenk tube was flame dried and flushed with Argon. LiCl (42.9 mg), amide **1a** (292.9 mg, 1.3 mmol) and DME (9.0 mL) were added into the dry Schlenk tube. LiHMDS (1.0 M in THF, 1.3 mL, 1.3 mmol) were added at 0 °C, and stirred at room temperature for 30 min. In a separated flushed flask, Pd(OAc)₂ (11.2 mg, 0.05 mmol), **L8** (47.2 mg, 0.05 mmol) and DME (5.0 mL) were added. The resulting mixture was stirred at room temperature for 30 min. Then the in-situ generated catalyst was added to the enolate solution. Allylic substrate **2a** (298.3 mg, 1.0 mmol) was then added and the mixture was stirred at -25 °C. After the reaction was complete (monitored by TLC), the reaction mixture was quenched by H₂O (1.0 mL). The solution was dried (anhydrous Na₂SO₄) and then filtered through a 2.0 inch plug of silica gel (eluting with EtOAc) to remove the solid. The crude reaction mixture was concentrated under reduced pressure. The resulting crude product was purified by flash column chromatography on silica gel (eluting with petroleum ether/ethyl acetate 10/1) to yield the product **3a** (0.35 g, 93%, 93.5:6.5 er).

7. Transformation of the allylated product **3d** and its determination of absolute configuration

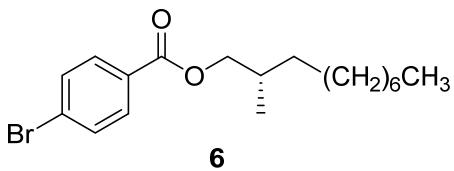


To a solution of LiAlH₄ (62.2 mg, 1.64 mmol) in THF (12 mL) at 0 °C, was added dropwise a solution of **3d** (50 mg, 0.164 mmol, 89% ee) in THF (4 mL). After stirring for 1 h, the reaction was quenched by the addition of water (3 mL). The mixture was filtered through Celite, which was washed with THF. The filtrate was concentrated, and the residue was purified by flash column chromatography on silica gel (eluting with petroleum ether/ethyl acetate 10/1) to furnished the product **4**⁴ (colorless oil, 21.4 mg, 84%). $[\alpha]_D^{21} = -0.62$ (1.0, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃) δ 5.60–5.26 (m, 2H), 3.50 (dd, *J* = 10.5, 6.1 Hz, 1H), 3.42 (dd, *J* = 10.6, 6.2 Hz, 1H), 2.16–2.03 (m, 1H), 1.99 (dd, *J* = 13.3, 6.5 Hz, 2H), 1.94–1.83 (m, 2H), 1.68 (dq, *J* = 13.1, 6.6 Hz, 1H), 1.42–1.20 (m, 6H), 0.89 (dd, *J* = 10.3, 6.9 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 132.36, 127.95, 67.91, 36.56, 35.96, 32.56, 31.37, 29.24, 22.51, 16.41, 14.06.



Palladium-charcoal (2.0 mg, 10%) was added to a solution of **4** (18 mg, 0.106 mmol) in EtOH (1mL). The suspension was stirred under H₂ (balloon) for 24 h at room temperature. The catalyst was filtered off through Celite, and the solid was washed

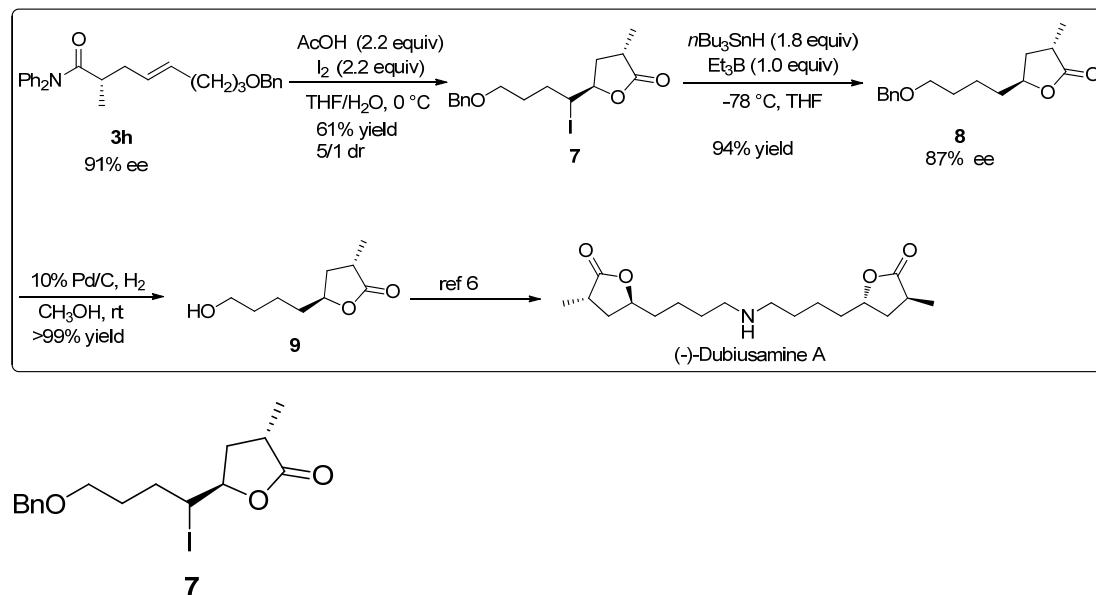
with EtOH. The combined solution was concentrated in vacuo to provide **5**⁵ (colorless oil, 18 mg, 99%). $[\alpha]_D^{26} = -7.2$ (1.0, CH₂Cl₂) $\{(S)\text{-}\mathbf{5}\}$: $[\alpha]_D = -9.4$ (1.0, CH₂Cl₂) $\}^{[5]}$; ¹H NMR (400 MHz, CDCl₃) δ 3.51 (dd, *J* = 10.5, 5.8 Hz, 1H), 3.41 (dd, *J* = 10.4, 6.6 Hz, 1H), 1.60 (dt, *J* = 12.6, 6.1 Hz, 1H), 1.47 (s, 1H), 1.40–1.05 (m, 14H), 0.97–0.83 (m, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 68.40, 35.74, 33.13, 31.90, 29.94, 29.61, 29.33, 26.98, 22.68, 16.58, 14.12.



The alcohol **5** (15 mg, 0.087 mmol) and DCM (1.5 mL) was added to a dry Schlenk tube, and then Et₃N (36 μL, 0.26mmol), DMAP (2.1 mg, 0.0174 mmol), and 4-bromobenzoyl chloride (57.3 mg, 0.26 mmol) were added. The mixture was stirred at room temperature until **5** was fully consumed (monitored by TLC). After the reaction mixture was diluted with DCM (3 mL), the organic phase was washed with 1M HCl (3 mL), sat. aq. Na₂CO₃ solution (3 mL), and brine (3 mL). After the organic phase was dried over anhydrous MgSO₄, the solvent was removed under reduced pressure and the residue was purified by flash column chromatography on silica gel (eluting with petroleum ether/ethyl acetate 10/1) to give the product **6** (colorless oil, 36.7 mg, 85%, 93.7:6.3 er). $[\alpha]_D^{26} = 1.5$ (1.0, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃) δ 7.92 (d, *J* = 8.6 Hz, 2H), 7.60 (d, *J* = 8.5 Hz, 2H), 4.22 (dd, *J* = 10.7, 5.8 Hz, 1H), 4.12 (dd, *J* = 10.7, 6.8 Hz, 1H), 2.11–1.78 (m, 1H), 1.48–1.25 (m, 14H), 1.03 (d, *J* = 6.7 Hz, 3H), 0.90 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 165.93, 131.66, 131.05, 129.40, 127.88, 70.14, 33.42, 32.65, 31.89, 29.82, 29.55, 29.30, 26.84, 22.67, 17.03, 14.12; IR: 2923, 2853, 1721, 1591, 1266, 1172, 1012, 847, 756, 722, 469; MS (EI): m/z (%) 354 (M⁺, 0.4), 200 (11), 185 (71), 183 (73), 155 (29), 141 (1), 113 (3), 99 (5),

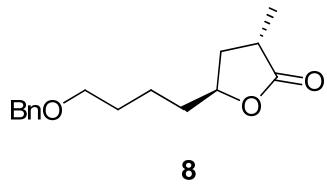
85 (12), 79 (3), 71 (17), 57 (66), 43 (40); HRMS (EI) Calcd for $C_{18}H_{27}O_2Br$ [M]⁺: 354.1194, found 354.1187; HPLC (Chiralpak OJ-H, Hexane: *i*-Propanol=98:2, 0.7 mL/min, 214nm): $t_{\text{minor}} = 6.87$ min, $t_{\text{minor}} = 7.52$ min.

8. Enantioselective formal synthesis of enantiomer of Dubiusamine A

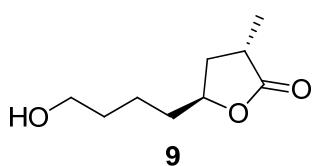


To a solution of **3h** (70 mg, 0.17mmol) and AcOH (21.3 μ L, 0.37mmol) in THF/H₂O (2:1, 2.4 mL) was added iodine (93.9 mg, 0.37 mmol) in THF (1.6 mL) at 0 °C. After being stirred at 0 °C for 48 h, the reaction mixture was poured into ice-cold 40% NaHSO₃ and extracted with Et₂O. The combined organic layers were washed with 1 N HCl, saturated NaHCO₃, and saturated NaCl. The organic phase was dried over anhydrous Na₂SO₄, filtered, and concentrated under vacuum. The diastereoselectivity was determined to be 5/1 by ¹H NMR spectroscopy. Then, the resulting crude product was purified by flash column chromatography on silica gel (eluting with petroleum ether/ethyl acetate 10/1) to yield the pure major isomer of **7** (colorless oil, 40.2 mg, 61%) and the minor isomer was completely removed. $[\alpha]_D^{28} = -11.6$ (1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 7.39–7.25 (m, 5H), 4.53–4.46 (m, 2H), 4.41 (td, $J = 7.7, 5.6$ Hz, 1H), 4.13 (ddd, $J = 10.1, 7.9, 3.5$ Hz, 1H), 3.56–3.46 (m, 2H), 2.76 (dp, $J =$

9.5, 7.3 Hz, 1H), 2.35 (ddd, J = 13.5, 9.6, 5.5 Hz, 1H), 2.17–2.08 (m, 1H), 2.08–2.00 (m, 1H), 1.94 (dtd, J = 19.9, 10.1, 6.0 Hz, 1H), 1.90–1.82 (m, 1H), 1.70 (ddt, J = 15.6, 11.9, 5.8 Hz, 1H), 1.30 (d, J = 7.4 Hz, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 179.29, 138.29, 128.39, 127.65, 127.61, 80.19, 72.96, 69.03, 39.07, 35.80, 34.48, 32.76, 29.41, 16.41; IR: 2935, 2855, 1772, 1495, 1453, 1360, 1174, 1096, 1022, 919, 736, 697, 597, 459 cm^{-1} ; MS (ESI) m/z: 389.0 ($\text{M}+\text{H}$) $^+$, 406.1 ($\text{M}+\text{NH}_4$) $^+$; HRMS (ESI) m/z Calcd for $\text{C}_{16}\text{H}_{25}\text{INO}_3$ ($\text{M}+\text{NH}_4$) $^+$: 406.0874, found 406.0872.



To compound **7** (40 mg, 0.10 mmol) in THF (3 mL) was added $n\text{Bu}_3\text{SnH}$ (51.3 μL , 0.19 mmol) and Et_3B (1.0 M in THF, 0.1 mL, 0.1 mmol) at -78°C , and stirred at -78°C for 2 hours. After quenching the reaction with saturated aqueous KF (3 mL), work-up with Et_2O (5 mL \times 3) and chromatographic separation ($\text{Et}_2\text{O}/\text{hexane} = 1/20$) provided **8** (colorless oil, 25.4 mg, 94 %, 93.8:6.2 er). $[\alpha]_D^{26} = -22.2$ (1.0, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.43–7.23 (m, 5H), 4.55–4.45 (m, 3H), 3.47 (t, J = 6.3 Hz, 2H), 2.80–2.57 (m, 1H), 2.10 (ddd, J = 13.5, 8.9, 4.9 Hz, 1H), 1.98 (dt, J = 12.9, 7.5 Hz, 1H), 1.73–1.57 (m, 4H), 1.51 (ddd, J = 19.3, 9.6, 4.9 Hz, 2H), 1.27 (d, J = 7.3 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 180.09, 138.44, 128.37, 127.65, 127.57, 78.33, 72.96, 69.93, 35.39, 35.20, 34.00, 29.41, 22.20, 15.90; IR: 2936, 2861, 1765, 1454, 1357, 1186, 1098, 996, 925, 736, 698, 610, 528 cm^{-1} ; MS (ESI) m/z: 263.1 ($\text{M}+\text{H}$) $^+$, 285.1 ($\text{M}+\text{Na}$) $^+$; HRMS (ESI) m/z Calcd for $\text{C}_{16}\text{H}_{26}\text{NO}_3$ ($\text{M}+\text{NH}_4$) $^+$: 280.1907, found 280.1907; HPLC (Chiraldak OD-H, Hexane:*i*-Propanol = 95:5, 0.7 mL/min, 214 nm): $t_{\text{major}} = 27.66$ min, $t_{\text{minor}} = 29.62$ min.



Palladium-charcoal (4 mg, 10%) was added to a solution of compound **8** (25 mg, 0.095 mmol) in EtOH (2 mL). The suspension was stirred under H₂ (balloon) for 24 h at room temperature. The catalyst was filtered off through Celite, and the solid was washed with EtOH. The combined solution was concentrated in vacuo to provide **9**. (colorless oil, 16.4 mg, quant). [α]_D³⁰ = -29.9 (1.0, CHCl₃) {(3*R*,5*R*)-**9**: [α]_D²² = +40 (0.01, CHCl₃)}⁶; ¹H NMR (400 MHz, CDCl₃) δ 4.53 (tt, *J* = 7.7, 5.0 Hz, 1H), 3.67 (dd, *J* = 12.8, 6.6 Hz, 2H), 2.83–2.61 (m, 1H), 2.18–2.08 (m, 1H), 2.02 (dt, *J* = 12.8, 7.5 Hz, 1H), 1.82–1.69 (m, 2H), 1.61 (dd, *J* = 13.3, 5.9 Hz, 2H), 1.57–1.45 (m, 2H), 1.28 (d, *J* = 7.3 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 180.17, 78.37, 62.48, 35.39, 35.17, 34.02, 32.23, 21.73, 15.89; HRMS (ESI) m/z Calcd for C₉H₂₀NO₃ (M+NH₄)⁺: 190.1438, found 190.1437.

9. Preliminary studies of the beta-hydride elimination for Pd-catalyzed allylic alkylation of amide **1a** with allyl reagent **2j**^a

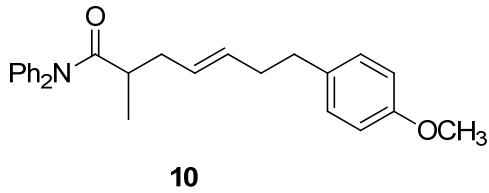
| | | | | | | | |
|-----------|--|---------------|---|------------------------------------|------------------------------------|-----------------------------------|--|
| 1a | 1) base (1.5 equiv), THF 2) Pd(OAc) ₂ (5.0 mol %), L1 (5.0 mol %) | 2j | 10 : Ph ₂ N-C(=O)-CH ₂ -CH=CH-CH ₂ -C ₆ H ₄ -OCH ₃ 11 : C ₆ H ₅ -CH=CH-CH ₂ -C ₆ H ₄ -OCH ₃ 12 : Ph ₂ N-CH ₂ -CH=CH-CH ₂ -C ₆ H ₄ -OCH ₃ | | | | |
| L1 | | | 12 | 10 | 11 | yield(2g,%)^c | |
| entry | base | <i>t</i> (°C) | yield (3p ,%) ^b | yield (10 ,%) ^b | yield (11 ,%) ^b | yield(2g ,%) ^c | |
| 1 | LDA | 0 | 76 | trace | 6 | 10 | |

| | | | | | | |
|----------------|--------|----|----|----|---|----|
| 2 | LDA | 25 | 79 | 9 | 8 | 0 |
| 3 | LDA | 50 | 63 | 28 | 8 | 0 |
| 4 ^d | LDA | 25 | 78 | 0 | 4 | 18 |
| 5 ^d | LDA | 50 | 81 | 0 | 4 | 15 |
| 6 ^e | LDA | 50 | 74 | 14 | 8 | 4 |
| 7 ^f | LDA | 25 | 76 | 14 | 6 | 0 |
| 8 | LiHMDS | 25 | 97 | 0 | 0 | 0 |

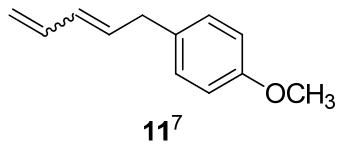
^aMolar ratio of **1a/2g/base/Pd(OAc)₂/L1** = 150/100/150/5/5. ^bIsolated yield.

^cIsolated yield of **2g** recovered. ^dLiCl (1.0 equiv) added as additive. ^e**L8** used as ligand. ^fDME used as solvent.

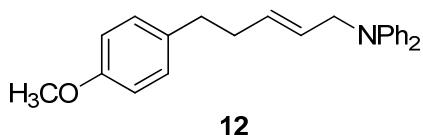
General experimental procedure for the Pd-catalyzed allylic alkylation of amide **1a** with allyl reagent **2j** under reaction condition as shown in above table: a dry Schlenk tube was flame dried and flushed with argon. Amide **1a** (67.59 mg, 0.3 mmol) and solvent (1.7 mL) were added into the dry Schlenk tube. Base (0.3 mmol) were added at 0 °C, and stirred at room temperature for 30 min. In a separated flushed flask, Pd(OAc)₂ (2.25 mg, 0.01 mmol), ligand (0.01 mmol) and solvent (1.0 mL) were added. The resulting mixture was stirred at room temperature for 30 min. Then the in-situ generated catalyst was added to the enolate solution. The allylic substrate **2j** (58.48 mg, 0.2 mmol) was then added and the mixture was stirred at the corresponding temperature. After the reaction was complete (monitored by TLC), the reaction mixture was quenched by H₂O (0.3 mL). The solution was dried (anhydrous Na₂SO₄) and then filtered through a 0.5 inch plug of silica gel (eluting with EtOAc) to remove the solid. The crude reaction mixture was concentrated under reduced pressure. The resulting crude product was purified by flash column chromatography on silica gel (eluting with petroleum ether/ethyl acetate 10/1) to yield the product.



¹H NMR (400 MHz, CDCl₃) δ 7.38–7.18 (m, 10H), 7.08 (d, *J* = 8.4 Hz, 2H), 6.81 (d, *J* = 8.4 Hz, 2H), 5.57–5.46 (m, 1H), 5.39–5.29 (m, 1H), 3.77 (s, 3H), 2.60 (q, *J* = 8.0 Hz, 3H), 2.49–2.37 (m, 1H), 2.35–2.22 (m, 2H), 2.11–1.98 (m, 1H), 1.10 (d, *J* = 6.7 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 176.60, 157.71, 142.99, 134.03, 131.87, 129.26, 127.87, 113.69, 55.25, 37.80, 37.71, 35.03, 34.73, 17.64; IR: 3033, 2930, 1670, 1592, 1511, 1491, 1379, 1246, 1033, 804, 758, 698, 522 cm⁻¹; MS(ESI) m/z: 400.2(M+H⁺), 422.2(M+Na⁺); HRMS(ESI) m/z Calcd for C₂₇H₃₀NO₂(M+H)⁺: 400.2271, found 400.2267.



E/Z = 62:38. ¹H NMR (400 MHz, CDCl₃) δ 7.14–7.07 (m, 2.0H), 6.88–6.80 (m, 2.0H), 6.33 (dt, *J* = 17.0, 10.3 Hz, 0.52H), 6.17–6.03 (m, 0.84H), 5.89–5.76 (m, 0.61H), 5.14 (ddd, *J* = 65.9, 45.6, 13.7 Hz, 1.84H), 3.79 (d, *J* = 2.0 Hz, 3.0H), 3.48 (d, *J* = 8.0 Hz, 0.70H), 3.36 (d, *J* = 6.9 Hz, 1.12H). ¹³C NMR (101 MHz, CDCl₃) δ 157.98, 136.96, 133.94, 132.07, 131.73, 131.03, 129.67, 129.52, 129.28, 117.87, 115.58, 113.89, 113.84, 55.27, 38.01, 33.00. HRMS (EI⁺) Calcd for C₁₂H₁₄O: 174.1045, found 174.1046.

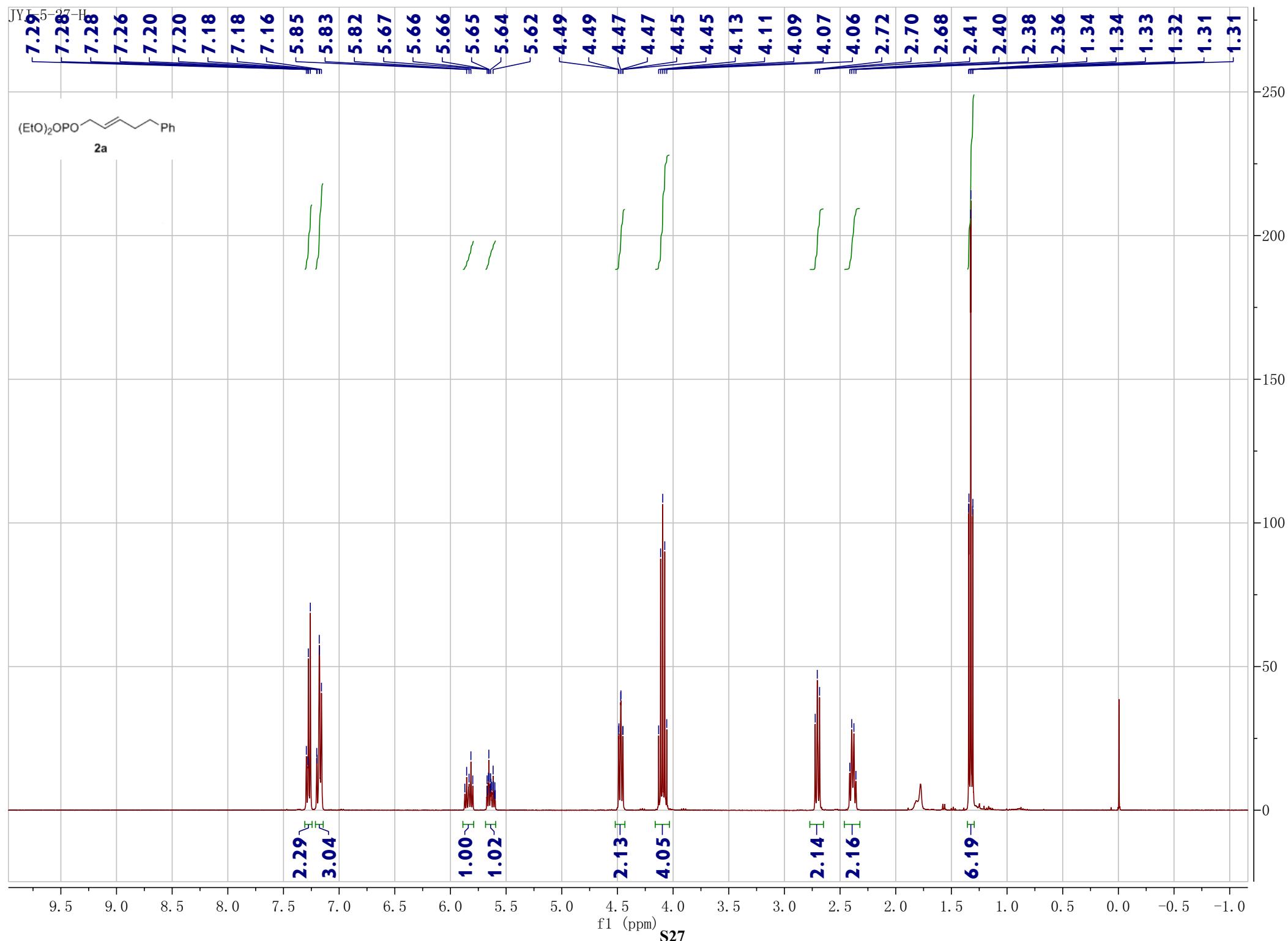


¹H NMR (400 MHz, CDCl₃) δ 7.24 (dd, *J* = 10.6, 5.0 Hz, 4H), 6.99 (d, *J* = 8.5 Hz, 6H), 6.93 (t, *J* = 7.3 Hz, 2H), 6.78 (d, *J* = 8.5 Hz, 2H), 5.71–5.59 (m, 1H), 5.53 (dt, *J*

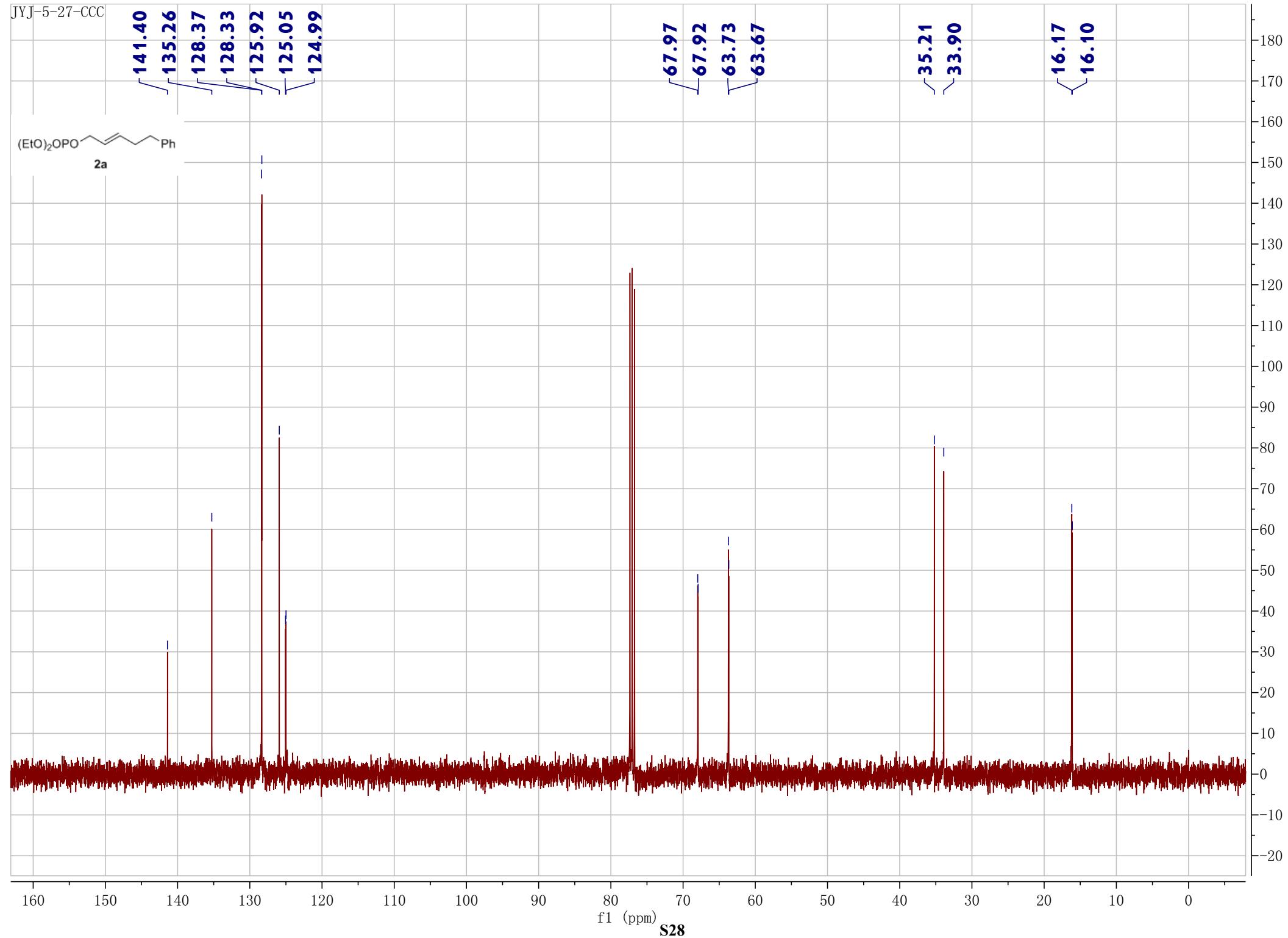
= 15.5, 5.0 Hz, 1H), 4.27 (d, J = 4.9 Hz, 2H), 3.77 (s, 3H), 2.56 (t, J = 7.6 Hz, 2H), 2.28 (q, J = 7.1 Hz, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 157.68, 147.88, 133.82, 131.87, 129.40, 129.15, 129.07, 126.31, 122.88, 121.70, 121.10, 120.77, 113.79, 113.63, 55.24, 53.99, 34.85, 34.42; HRMS(ESI) m/z Calcd for $\text{C}_{24}\text{H}_{26}\text{NO}$ ($\text{M}+\text{H}$) $^+$: 344.2009, found 344.2002.

10. References

- (1) Shintani, R.; Takatsu, K.; Takeda, M.; Hagashi, T. *Angew. Chem. Int. Ed.* **2011**, *50*, 8656.
- (2) Delvos, L. B.; Vyas, D. J.; Oestreich, M. *Angew. Chem. Int. Ed.* **2013**, *52*, 4650.
- (3) Yuan, Q.-J.; Yao, K.; Zhang, W.-B. *Chem. Commun.* **2015**, *51*, 11834.
- (4) Huang, Z.-H.; Tan, Z.; Novak, T.; Zhu, G.-G.; Negishi, E. I. *Adv. Synth. Catal.* **2007**, *349*, 539.
- (5) Cardillo, G.; D'Amico, A.; Orena, M.; Sandri, S. *J. Org. Chem.* **1988**, *53*, 2354.
- (6) Tan, M. A.; Kitajima, M.; Kogure, N.; Nonata, M. G.; Takayama, H. *Tetrahedron* **2010**, *66*, 3353.
- (7) Gao, D.-X.; Aplin, T.; Bauld, N. L. *J. Phys. Org. Chem.* **1999**, *12*, 808.



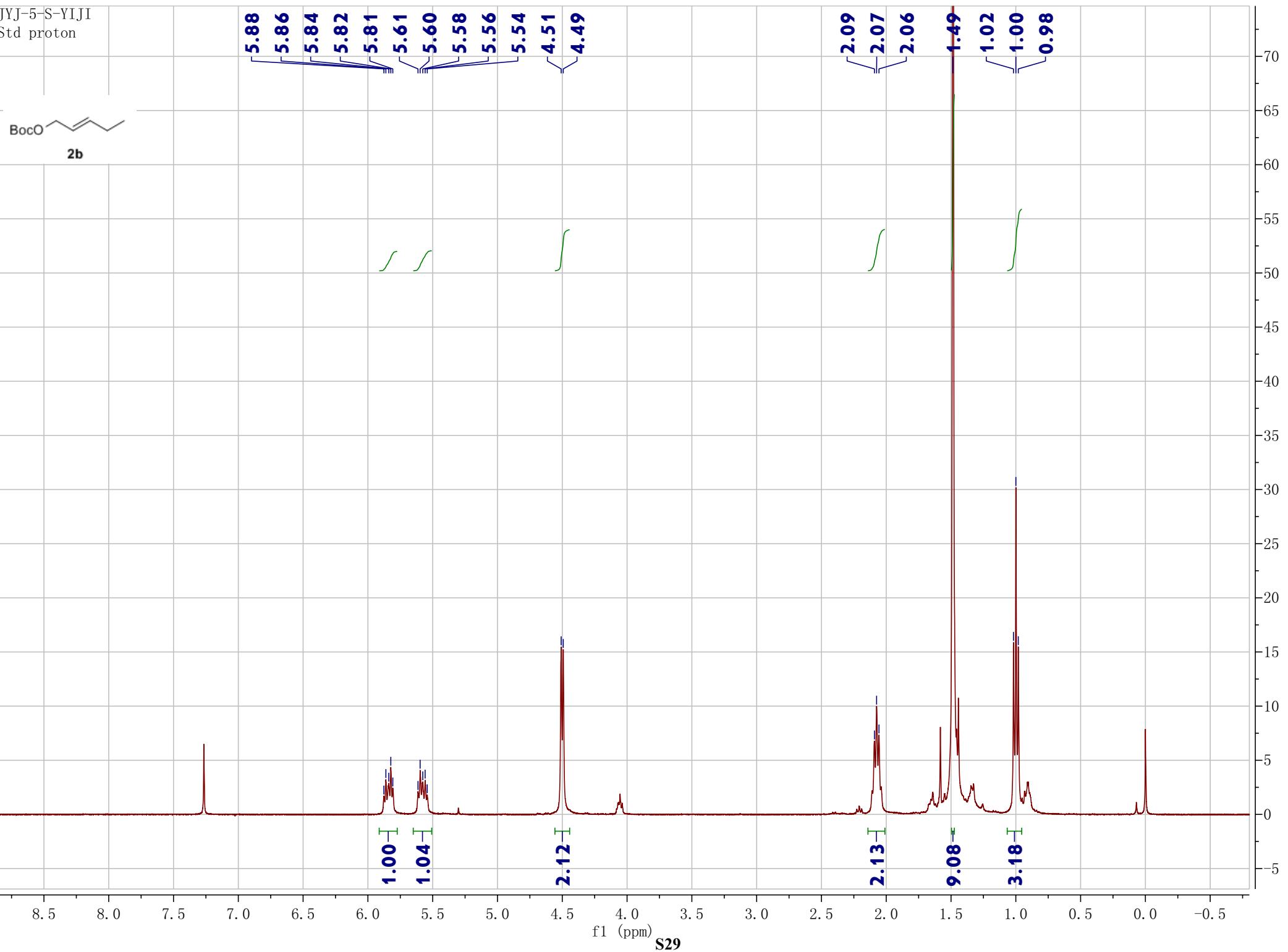
JYJ-5-27-CCC



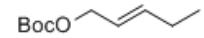
JYJ-5-S-YIJI
Std proton



2b

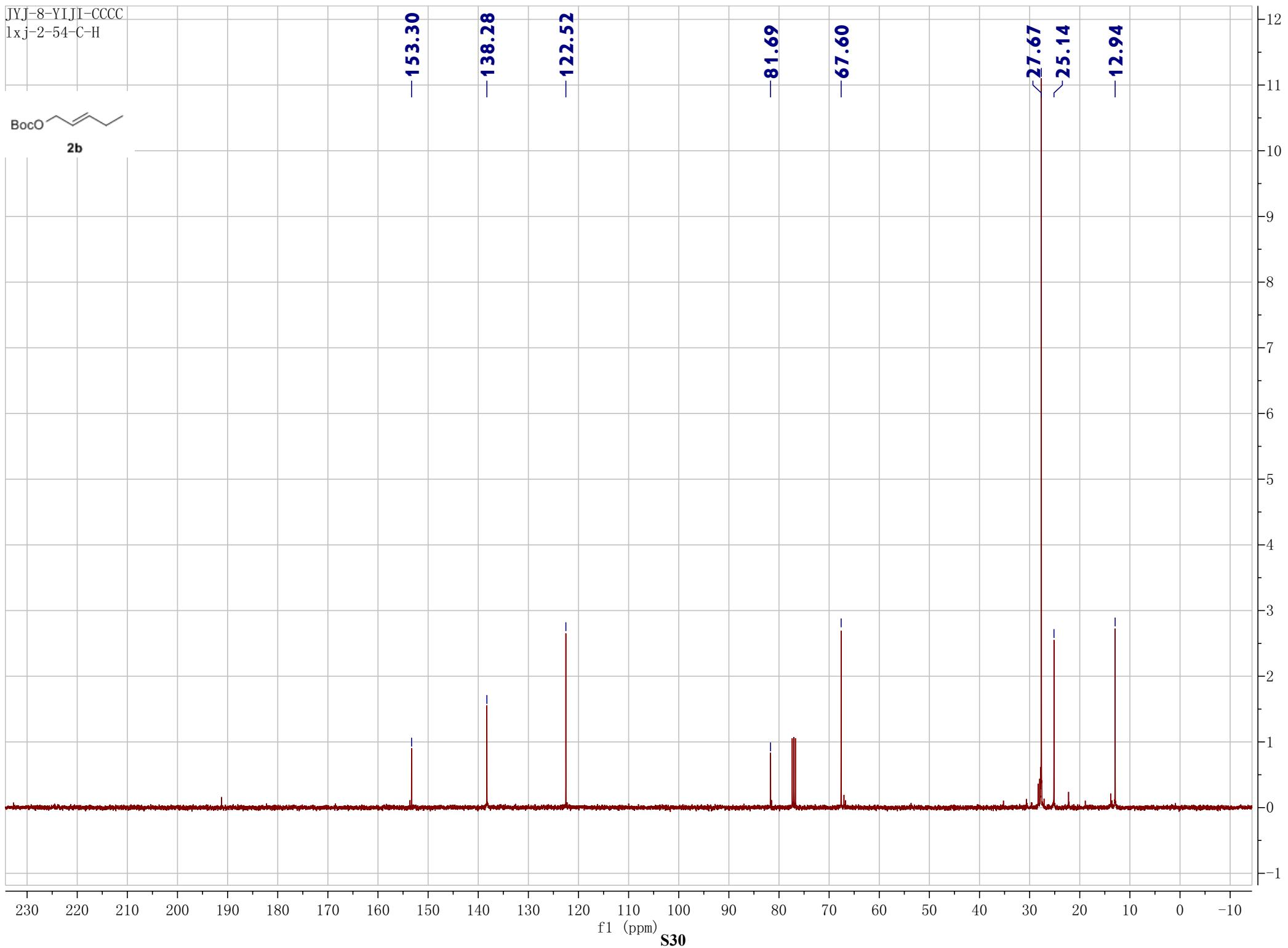


JYJ-8-YIJI-CCCC
1xj-2-54-C-H

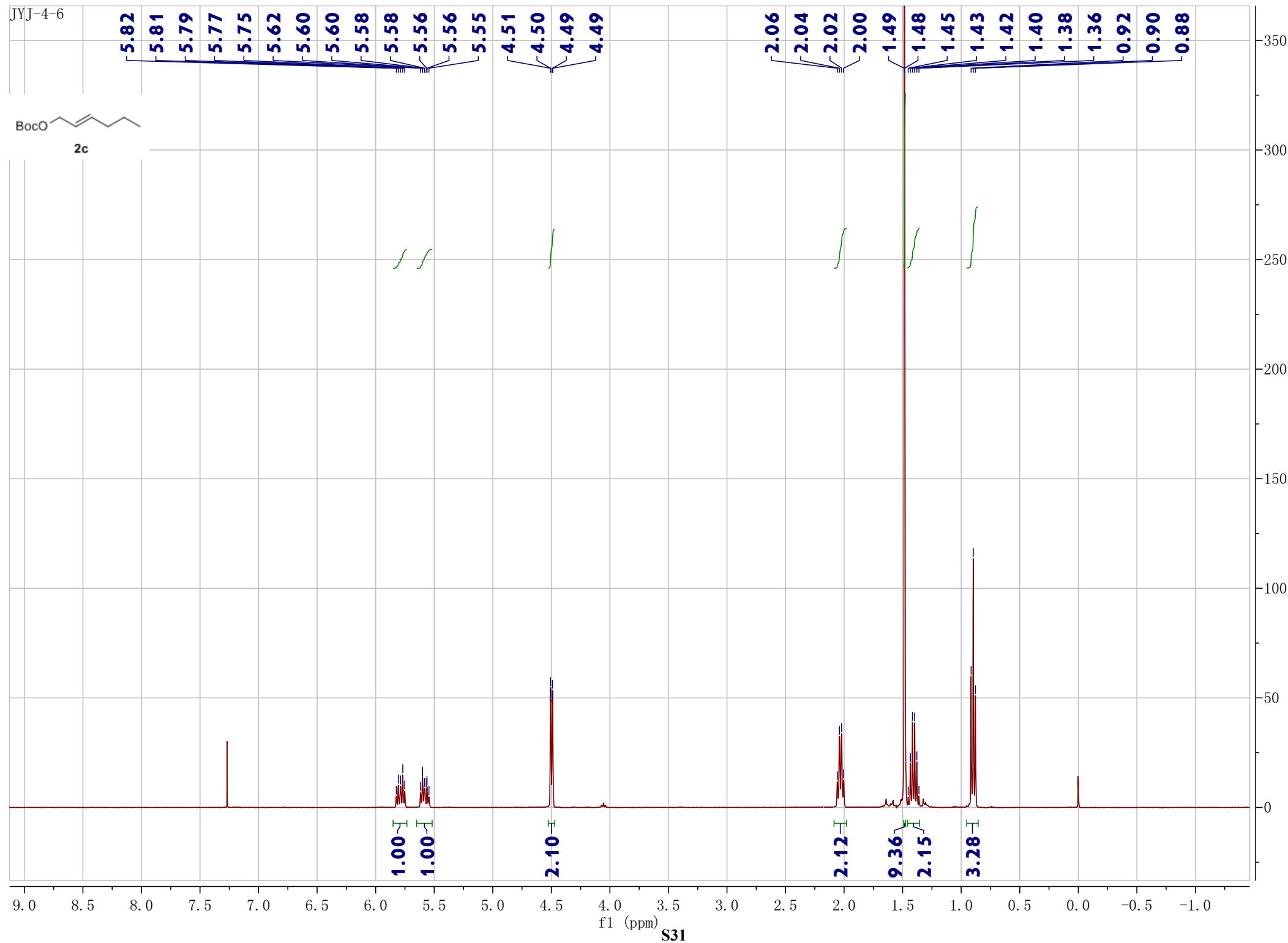


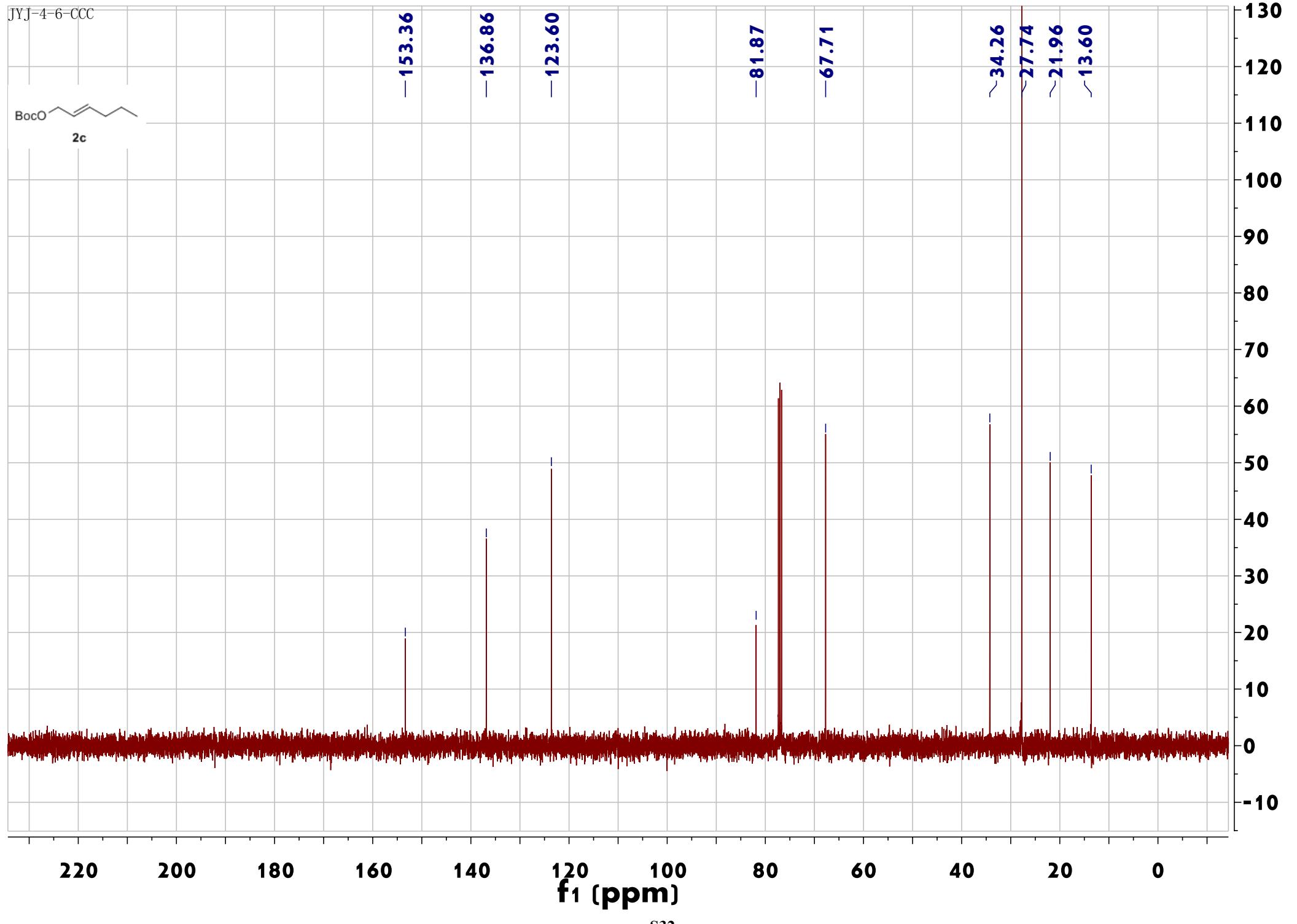
2b

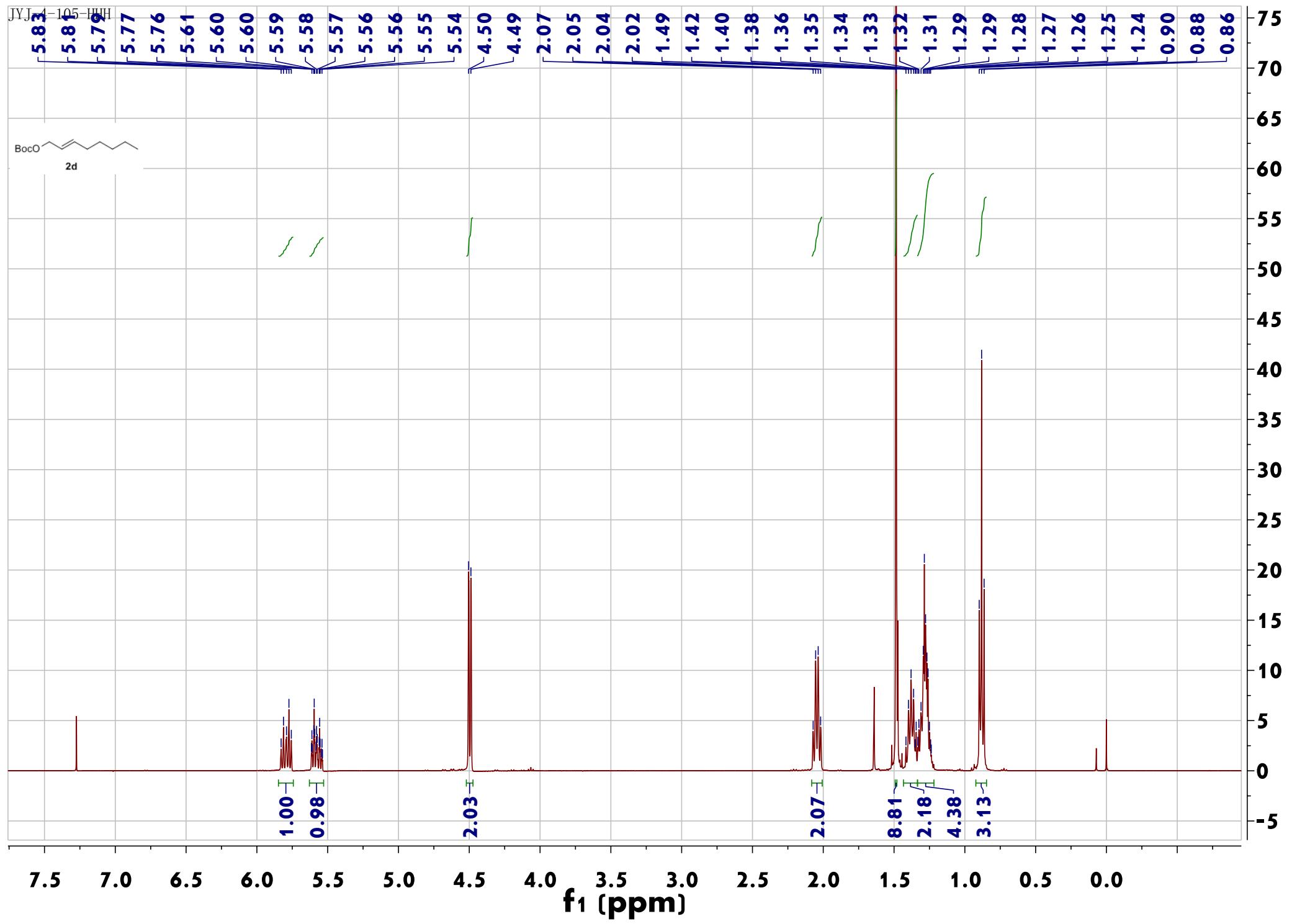
-153.30
-138.28
-122.52
-81.69
-67.60
27.67
25.14
-12.94

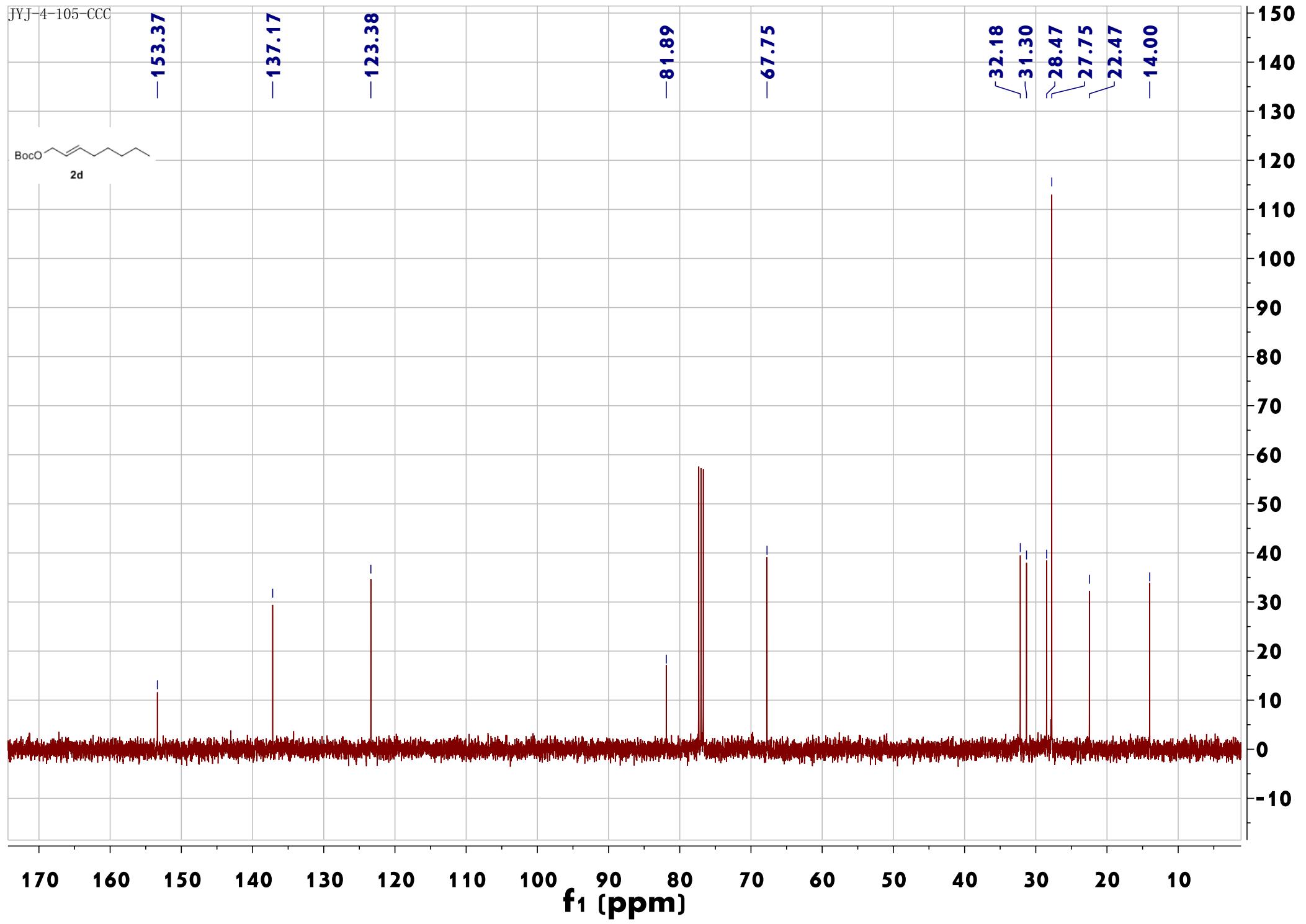


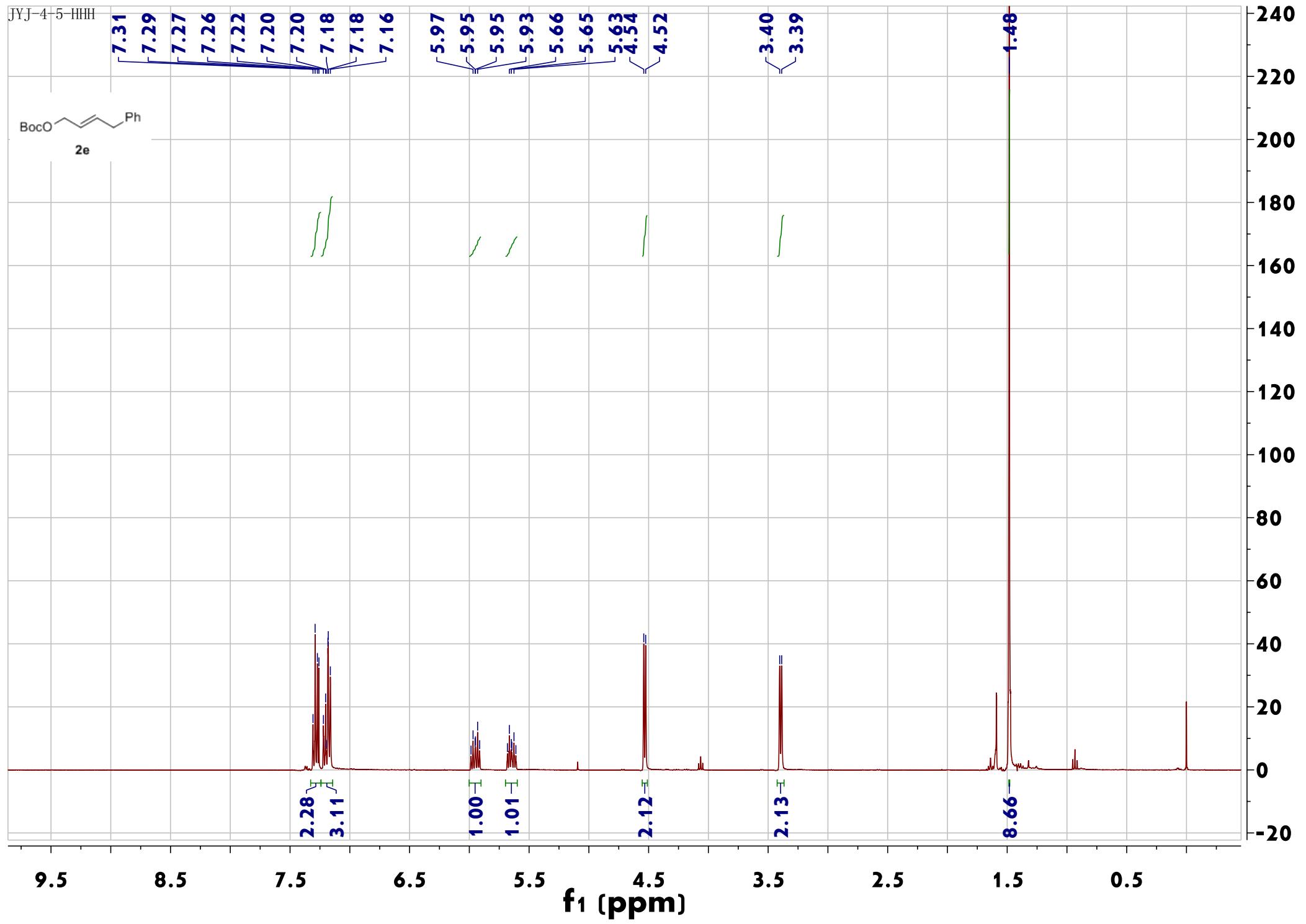
JYJ-4-6

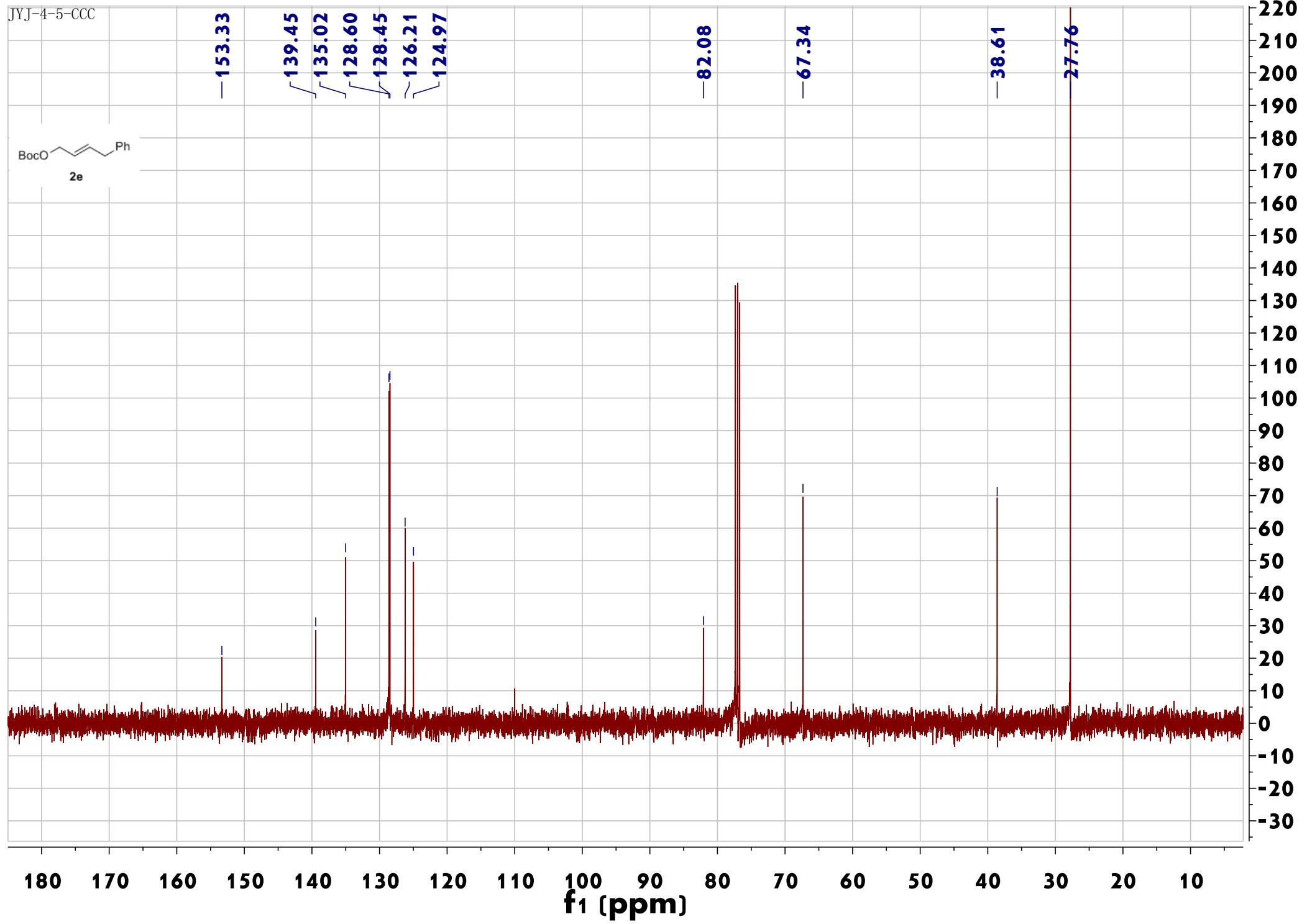
**2c**

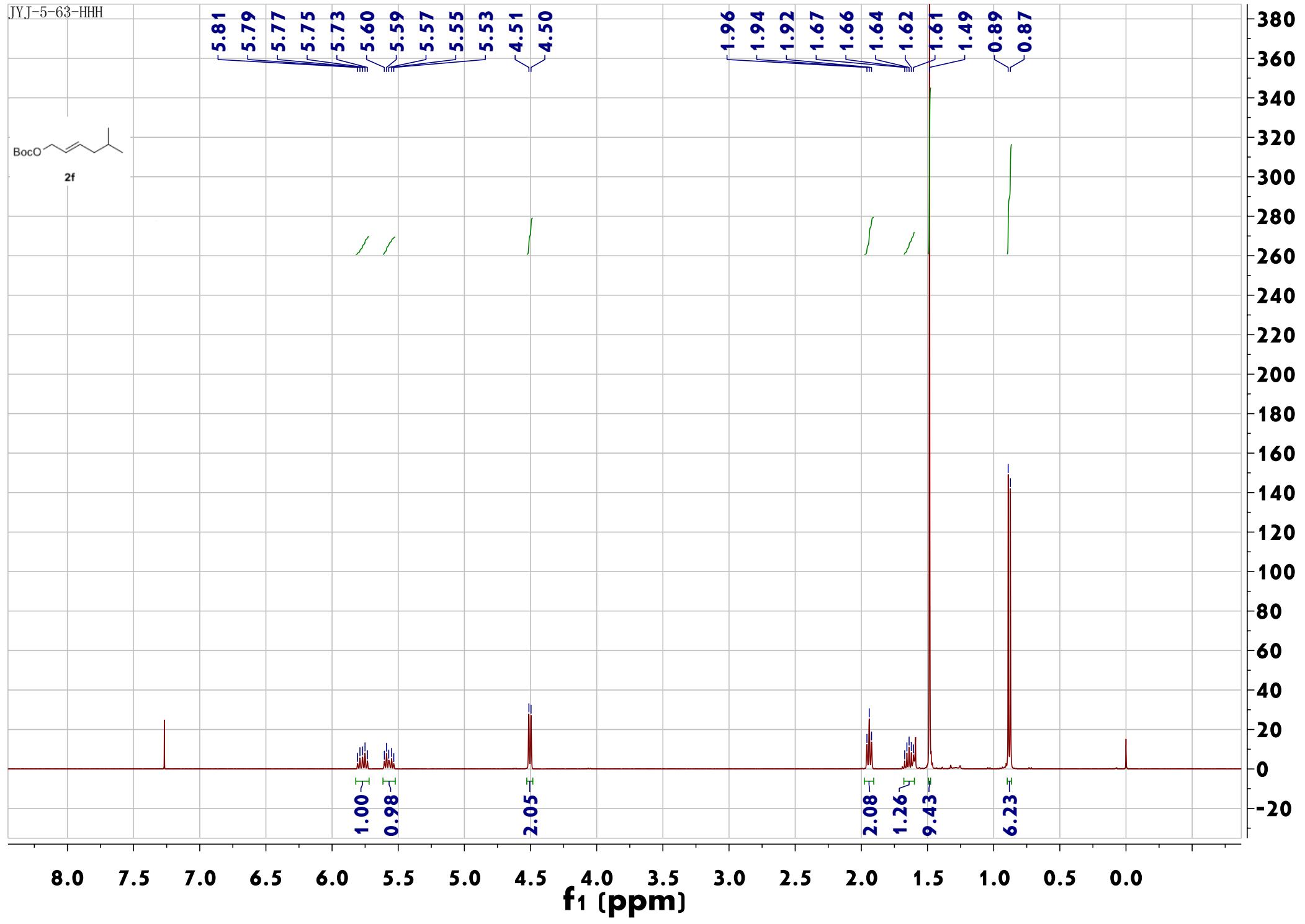




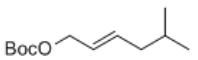








JYJ-5-63-CCC
1xj=2-54-C-H



2f

-153.32

-135.63

-124.62

-81.75

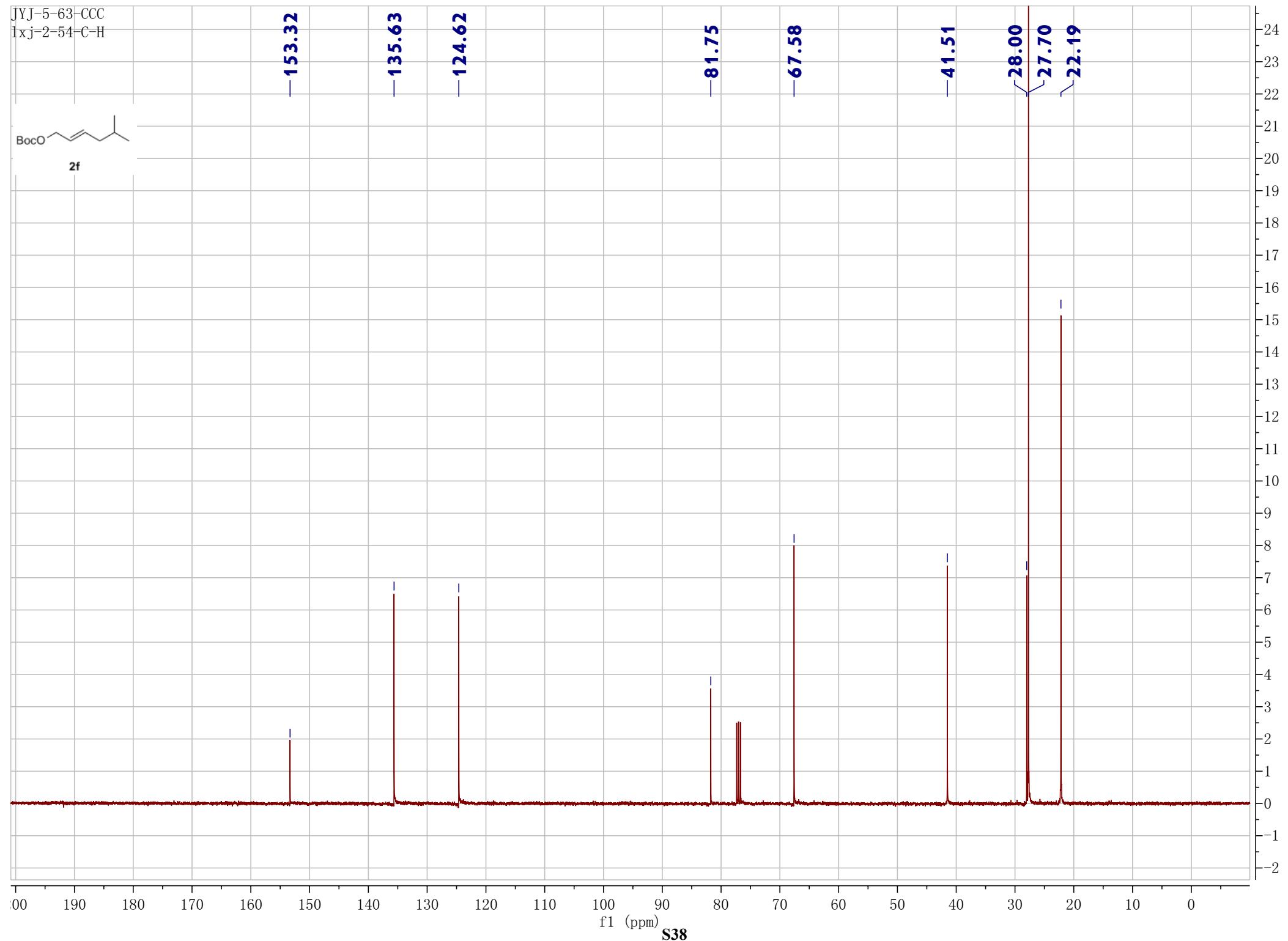
-67.58

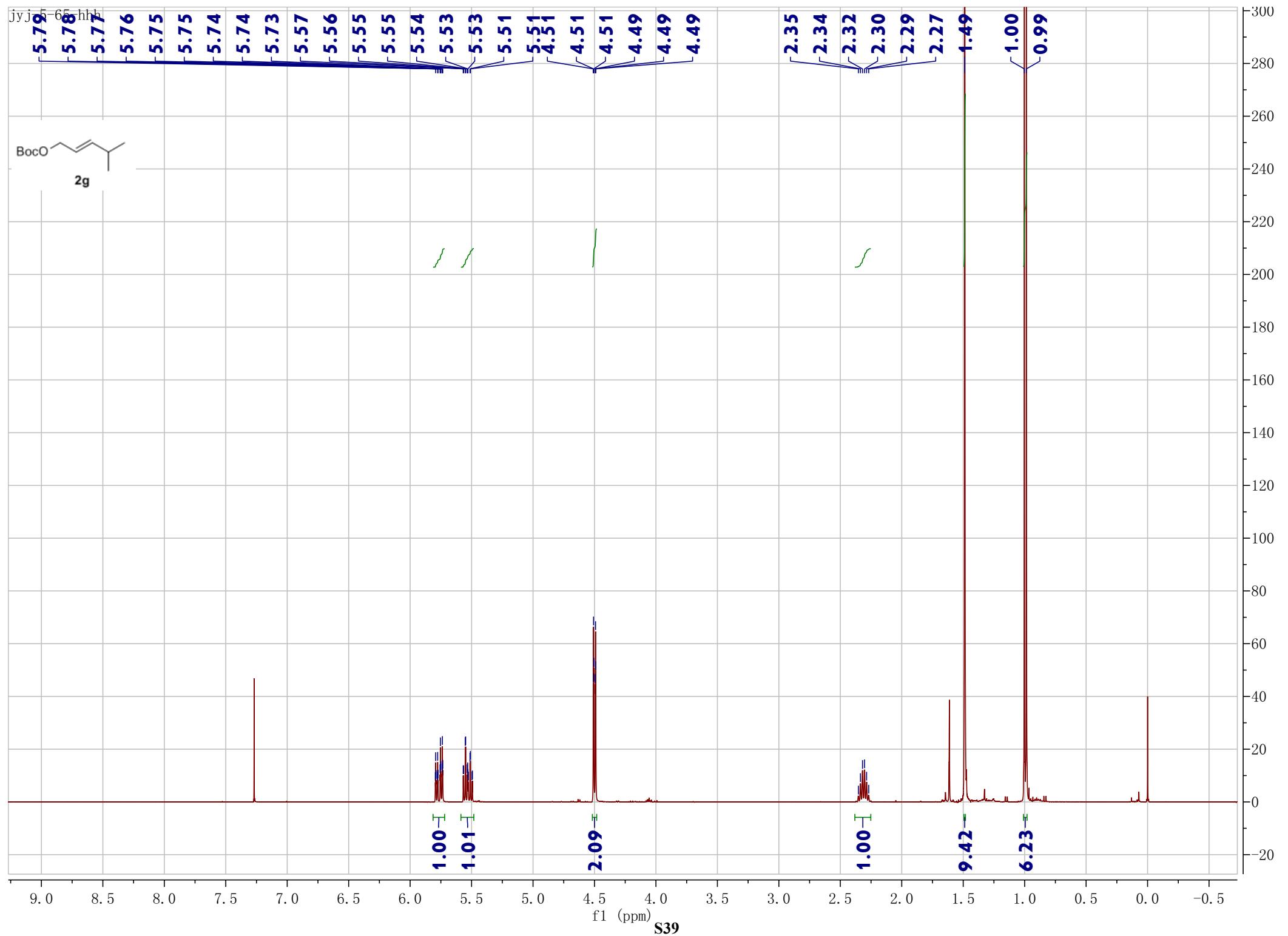
-41.51

28.00

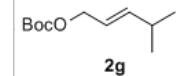
27.70

22.19





JYJ-5-65-CCC



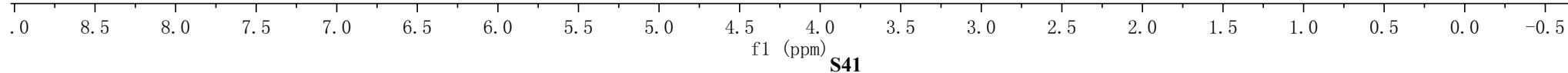
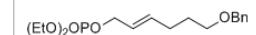
-153.36
-143.73
-120.55
-81.96
-67.89
~30.75
~27.77
~21.93

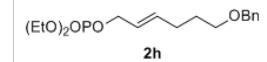
230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm) S40

150
140
130
120
110
100
90
80
70
60
50
40
30
20
10
0
-10
-20

JYJ-6-73-HHH

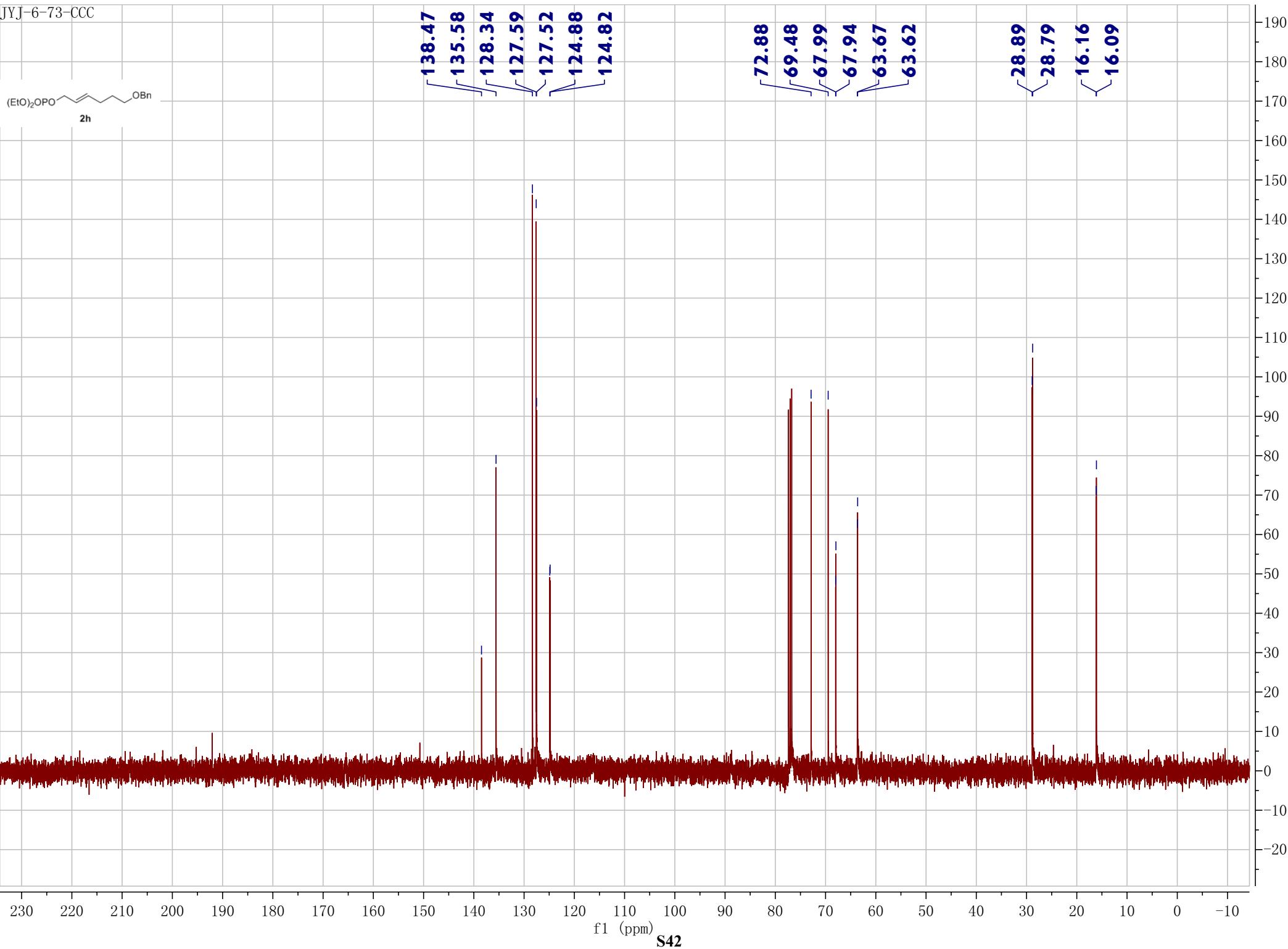




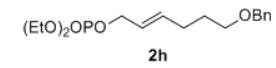
138.47
135.58
128.34
127.59
127.52
124.88
124.82

72.88
69.48
67.99
67.94
63.67
63.62

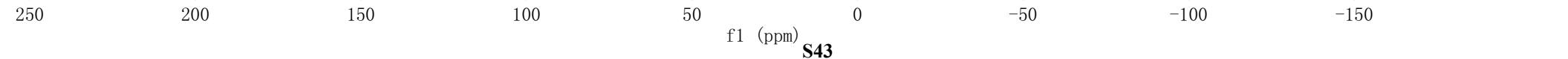
28.89
28.79
16.16
16.09



JYJ-6-73-PPP
STANDARD CARBON PARAMETERS



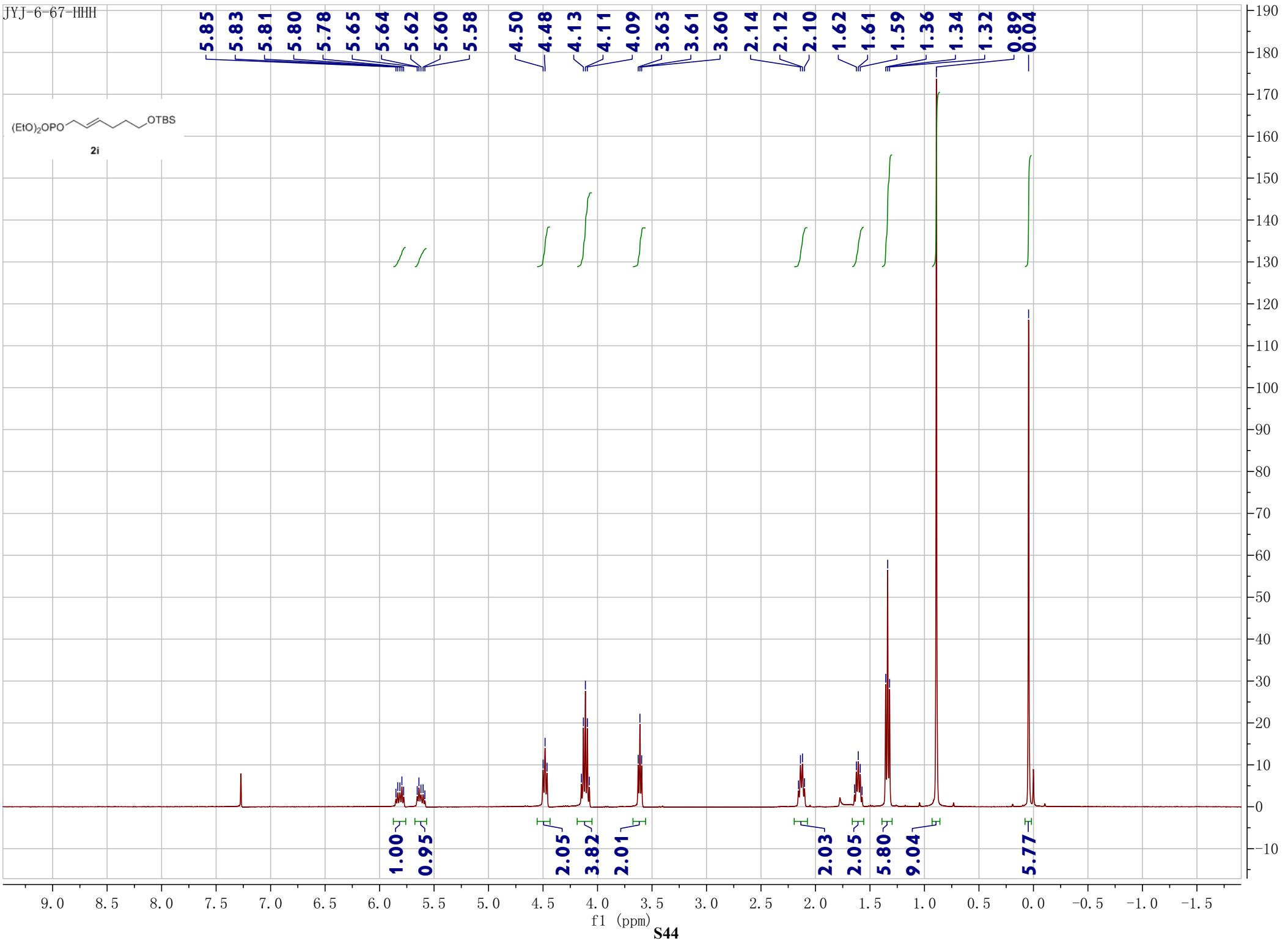
-0.83



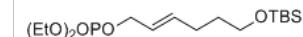
JYJ-6-67-HHH



2i



JYJ-6-67-CCC



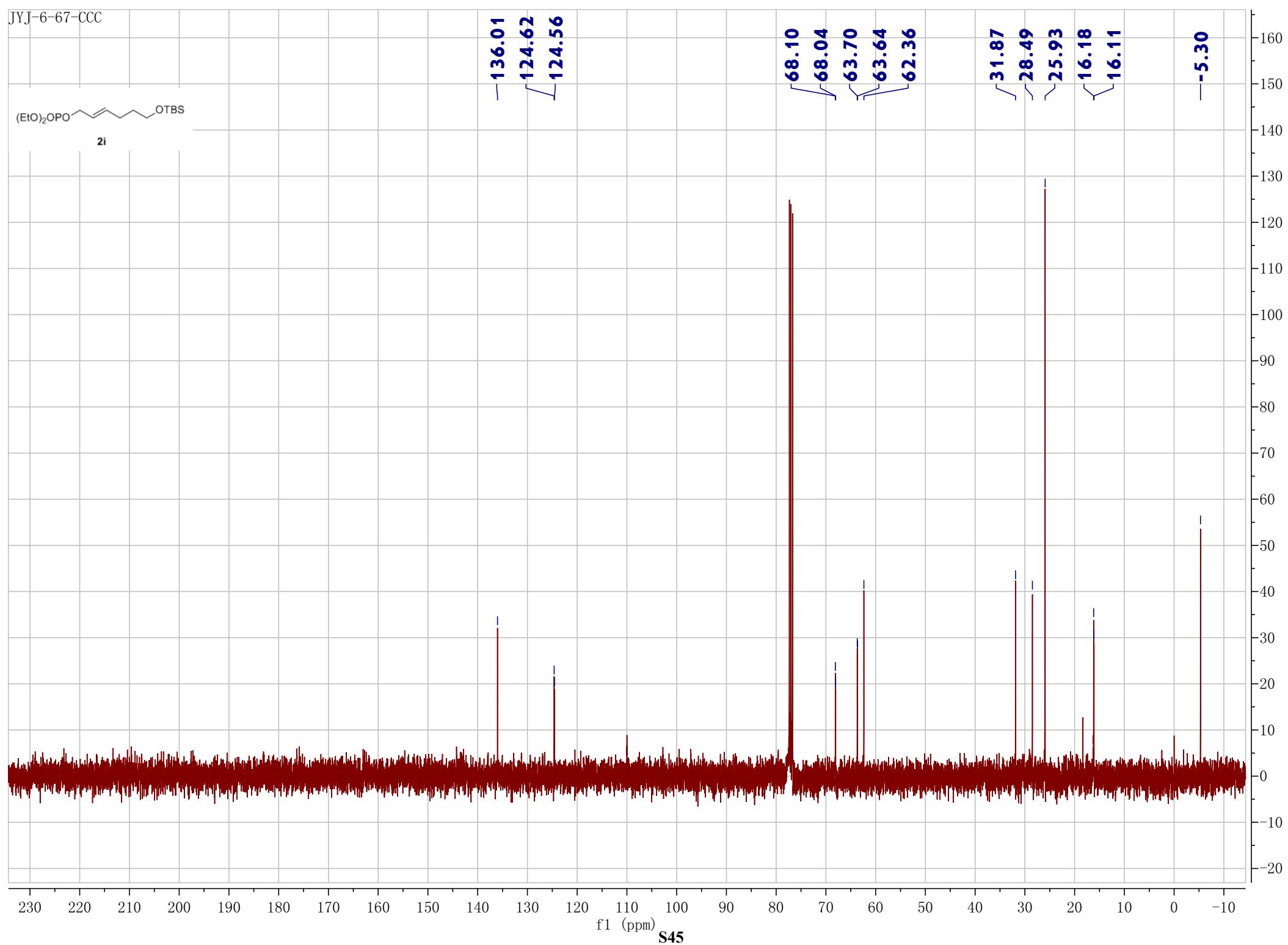
2i

— 136.01
— 124.62
— 124.56

— 68.10
— 68.04
— 63.70
— 63.64
— 62.36

— 31.87
— 28.49
— 25.93
— 16.18
— 16.11

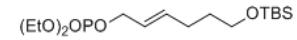
— -5.30



230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm) S45

JYJ-6-67-PPP
STANDARD CARBON PARAMETERS



2i

-0.83

250

200

150

100

50

0

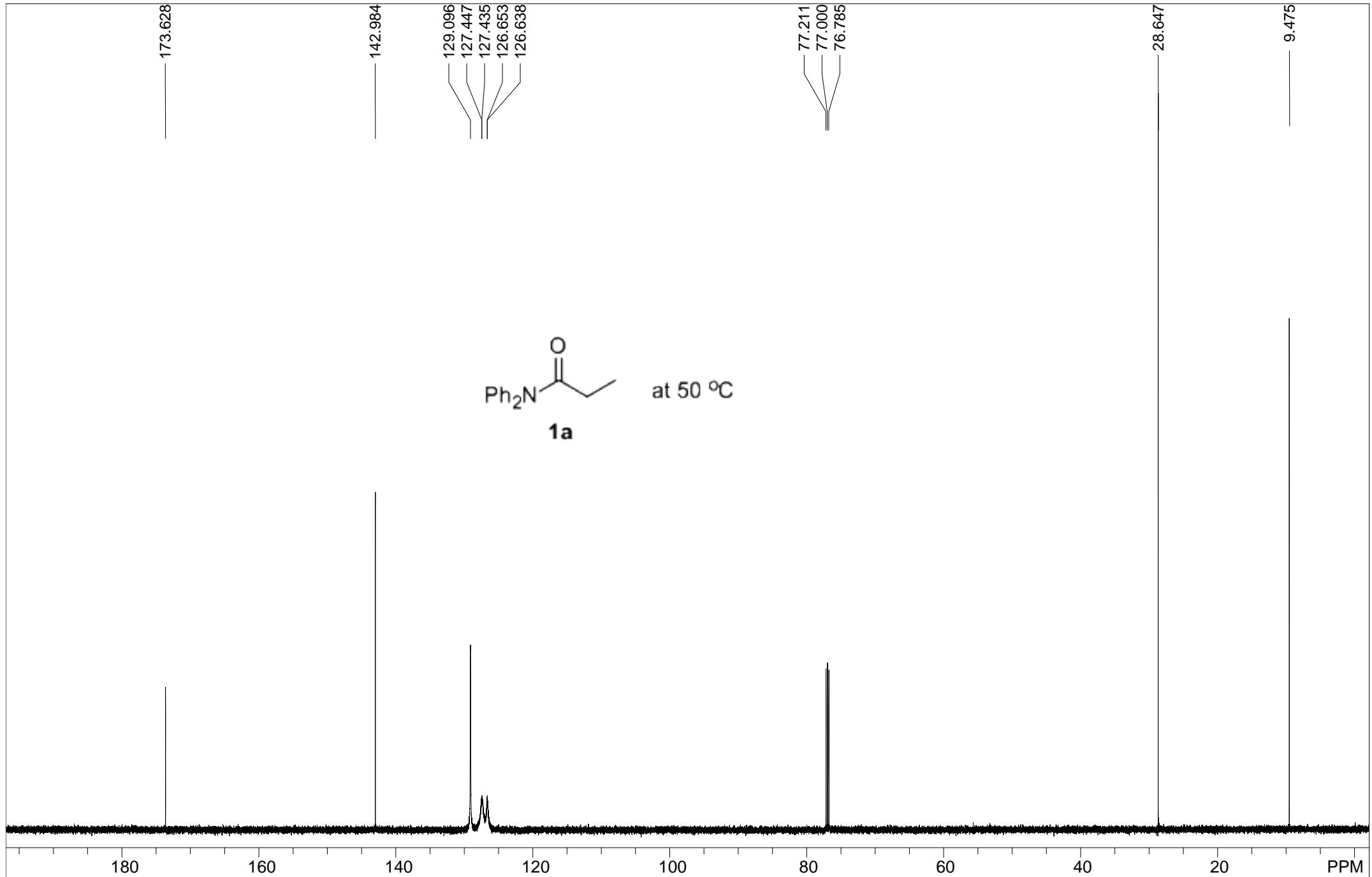
-50

-100

-150

f1 (ppm) S46

75
70
65
60
55
50
45
40
35
30
25
20
15
10
5
0
-5



:blank line

F1: 150.829

F2: 599.778

SW1: 35714

OF1: 15396.1

PTS1d: 65536

EX: s2pul

PW: 5.2 usec

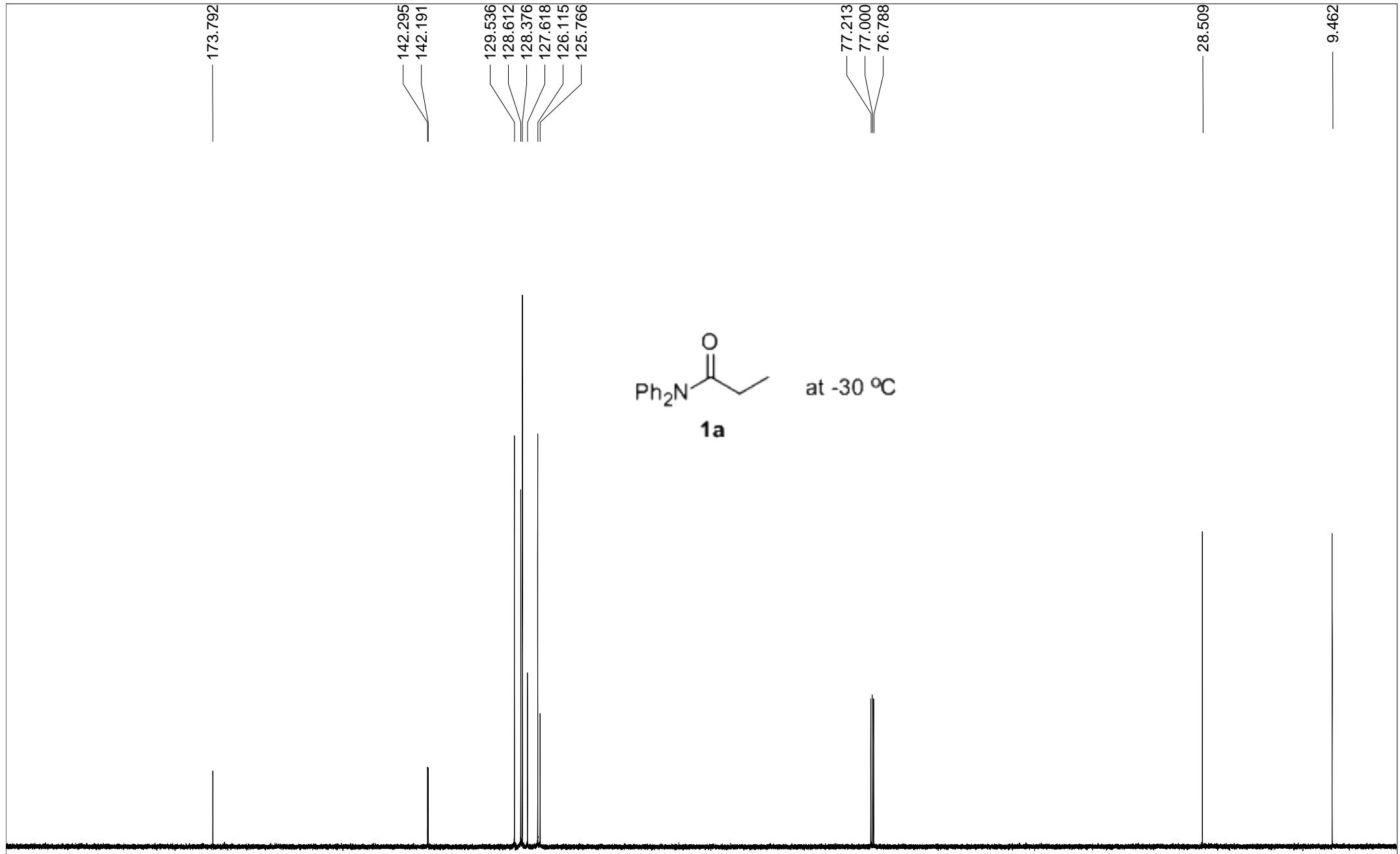
PD: 1.0 sec

NA: 96

LB: 0.0

USER: -- DATE: Sep 23 2014

Nuts - \$50oC-C.fid



200

150

100

50

PPM

:blank line

USER: -- DATE: Sep 23 2014

F1: 150.829

F2: 599.778

SW1: 35714

OF1: 15348.4

PTS1d: 65536

EX: s2pul

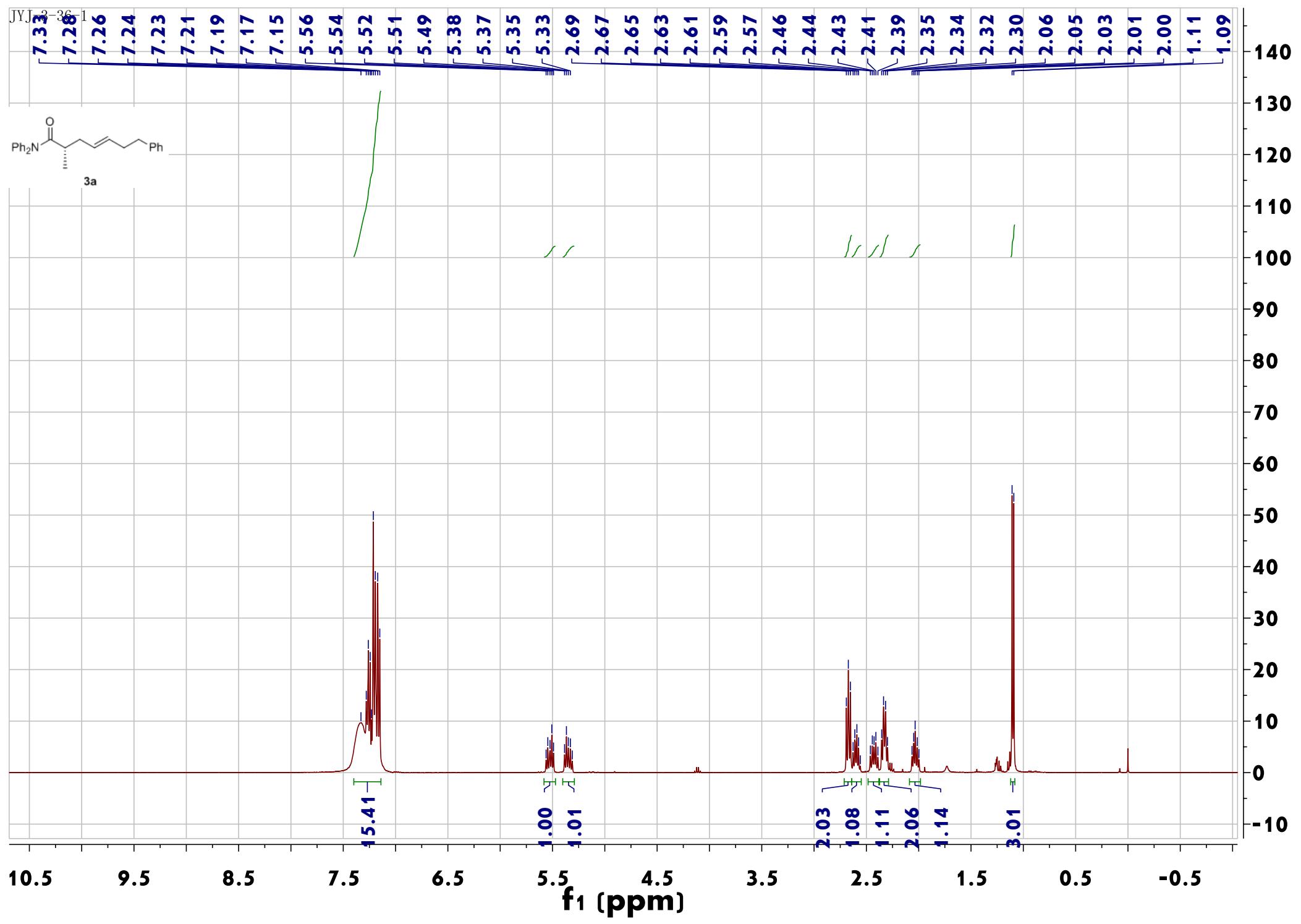
PW: 5.2 usec

PD: 1.0 sec

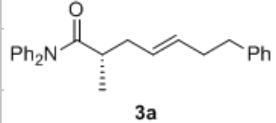
NA: 32

LB: 0.0

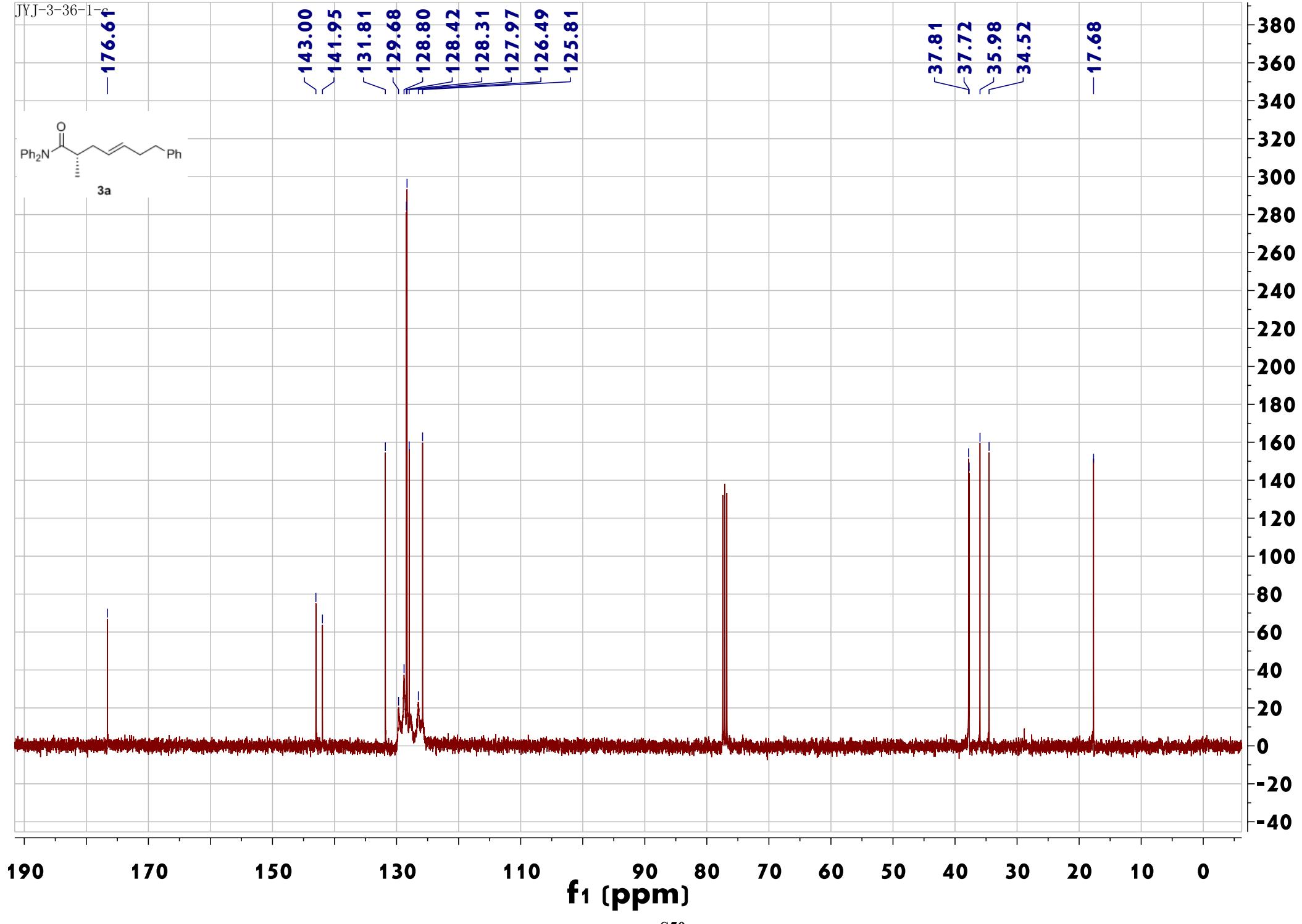
Nuts - \$-30oC-C.fid



JYJ-3-36-1-e
—176.61

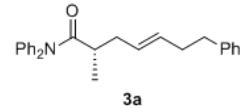


3a

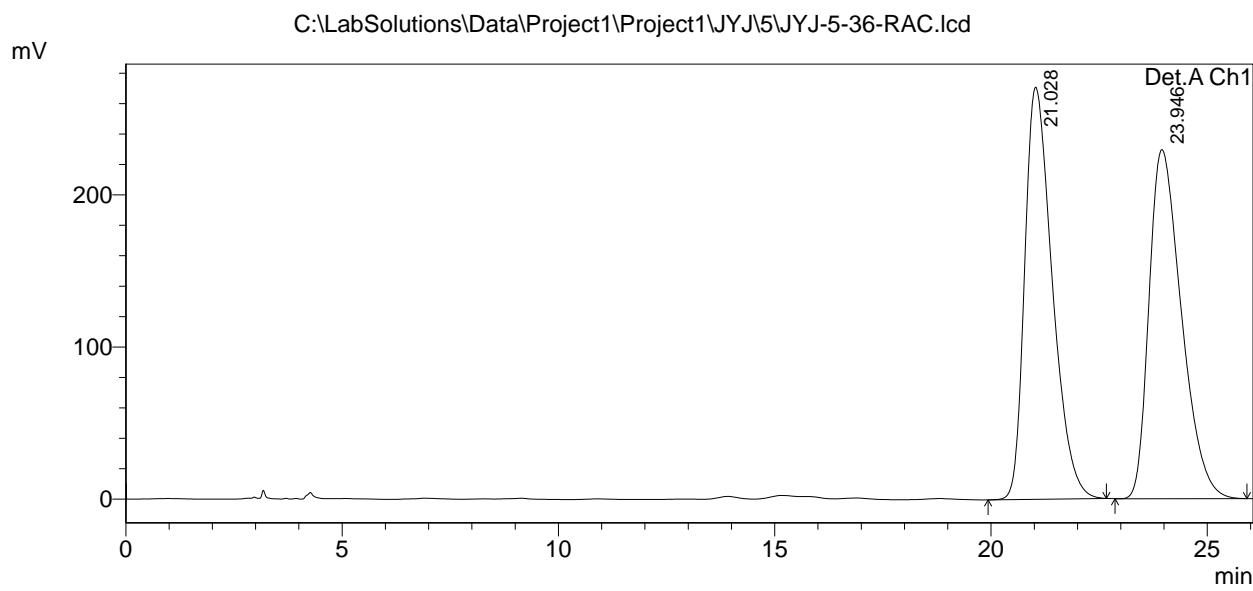


==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : JYJ-5-36-RAC
 Sample ID : OD-H,99/1,1,214
 Vial # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-5-36-RAC.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-10-13 9:51:10
 Data Processed : 2016-10-13 10:46:36



<Chromatogram>



PeakTable

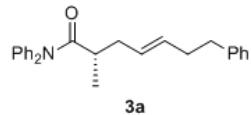
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 21.028 | 12015408 | 271024 | 50.173 | 54.143 |
| 2 | 23.946 | 11932609 | 229549 | 49.827 | 45.857 |
| Total | | 23948017 | 500573 | 100.000 | 100.000 |

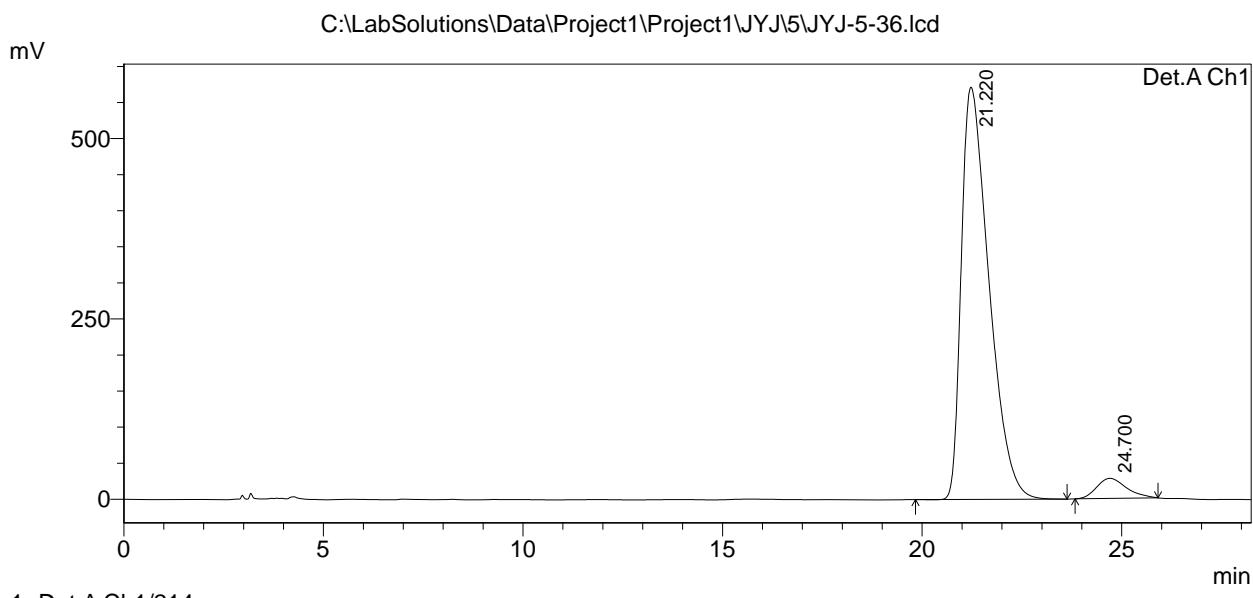
==== Shimadzu LCsolution Analysis Report ====

C:\LabSolutions\Data\Project1\Project1\JYJ\5\JYJ-5-36.lcd

Acquired by : Admin
 Sample Name : JYJ-5-36-1
 Sample ID : OD-H,99/1,1,214
 Vial # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-5-36.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-10-13 10:18:14
 Data Processed : 2016-10-13 10:52:12



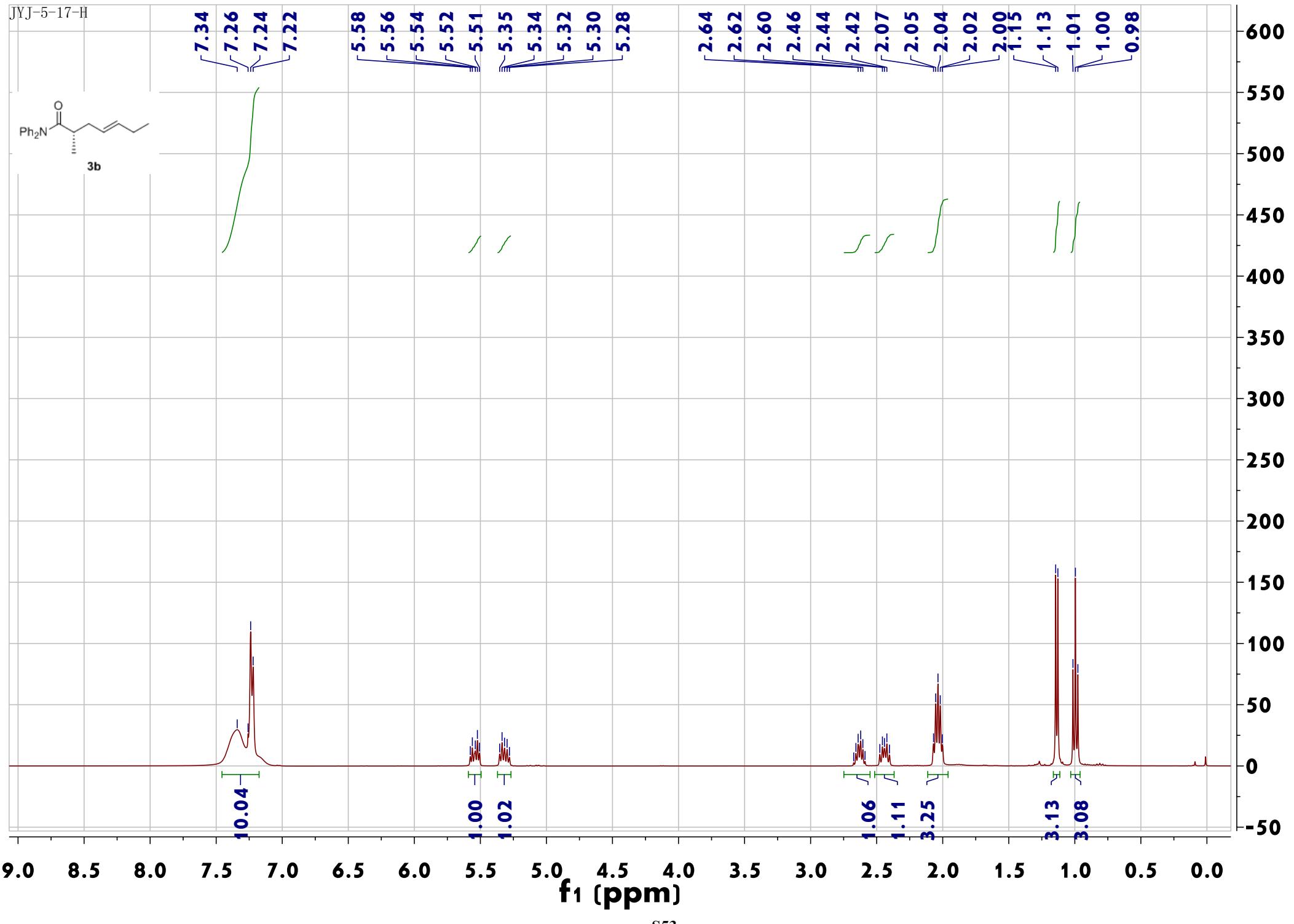
<Chromatogram>



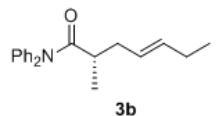
PeakTable

Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 21.220 | 27462370 | 571498 | 94.985 | 95.345 |
| 2 | 24.700 | 1449818 | 27899 | 5.015 | 4.655 |
| Total | | 28912188 | 599397 | 100.000 | 100.000 |



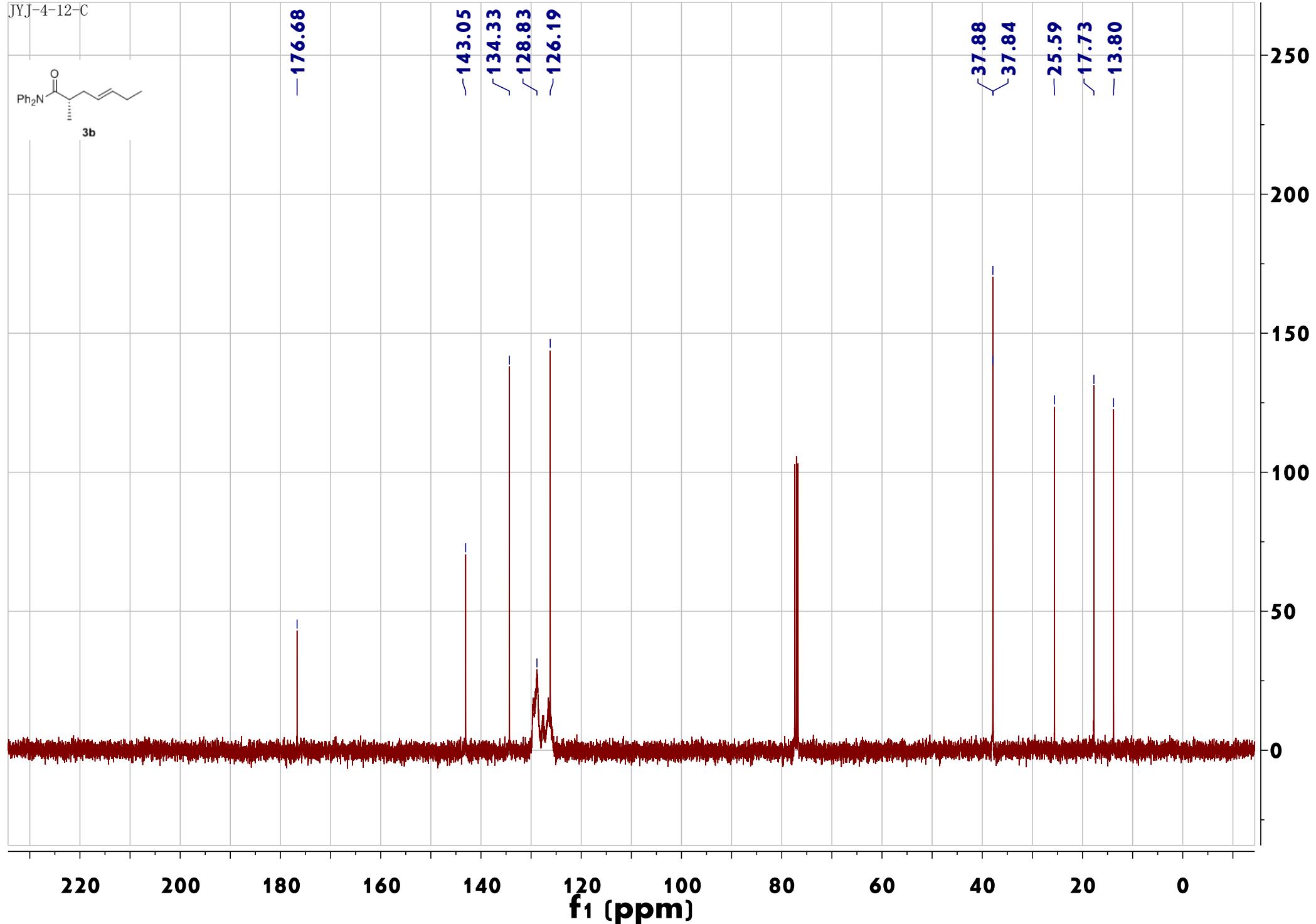
JYJ-4-12-C



-176.68

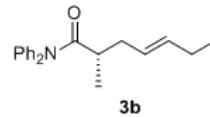
~~143.05~~ ~~134.33~~ ~~128.83~~ ~~126.19~~

$$\begin{array}{r} \boxed{37.88} \\[-1ex] \boxed{37.84} \\[-1ex] - 25.59 \\[-1ex] \hline 17.73 \\[-1ex] - 13.80 \\[-1ex] \hline \end{array}$$

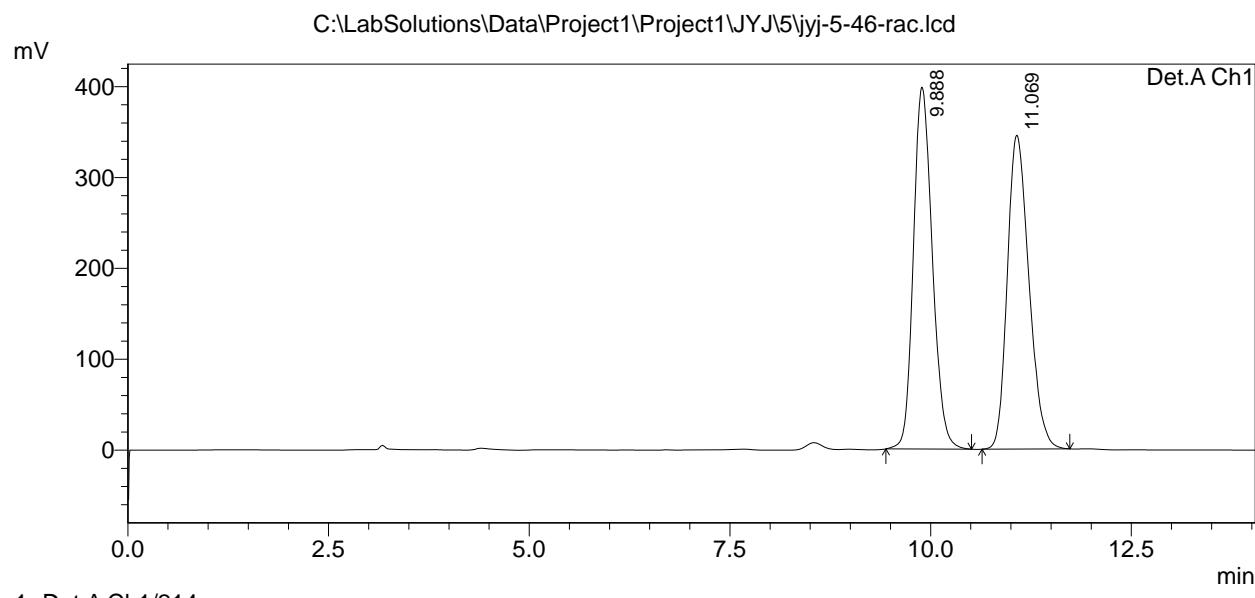


==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : jyj-5-46-rac
 Sample ID : OD-H,99/1,1,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : jyj-5-46-rac.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-9-11 9:34:21
 Data Processed : 2016-9-11 10:08:41



<Chromatogram>



PeakTable

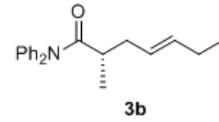
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 9.888 | 6525634 | 398221 | 50.195 | 53.560 |
| 2 | 11.069 | 6474938 | 345290 | 49.805 | 46.440 |
| Total | | 13000572 | 743511 | 100.000 | 100.000 |

==== Shimadzu LCsolution Analysis Report ====

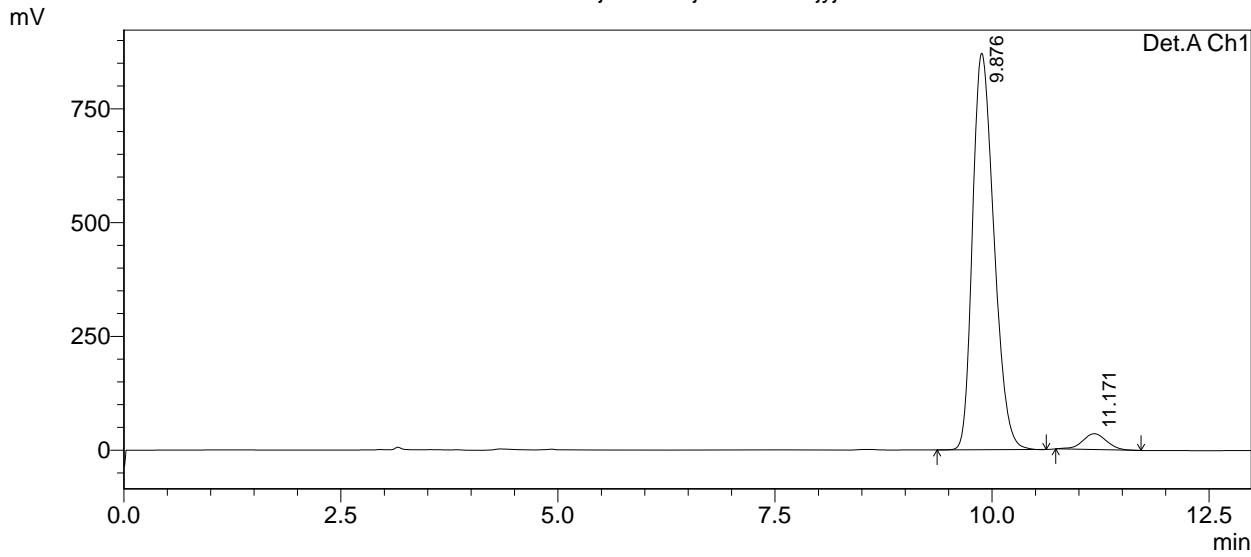
C:\LabSolutions\Data\Project1\Project1\JYJ\5\jyj-5-60.lcd

Acquired by : Admin
 Sample Name : jyj-5-60
 Sample ID : OD-H,99/1,1,214
 Vail # :
 Injection Volume : 2 uL
 Data File Name : jyj-5-60.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-9-24 22:18:14
 Data Processed : 2016-9-24 22:44:39



<Chromatogram>

C:\LabSolutions\Data\Project1\Project1\JYJ\5\jyj-5-60.lcd

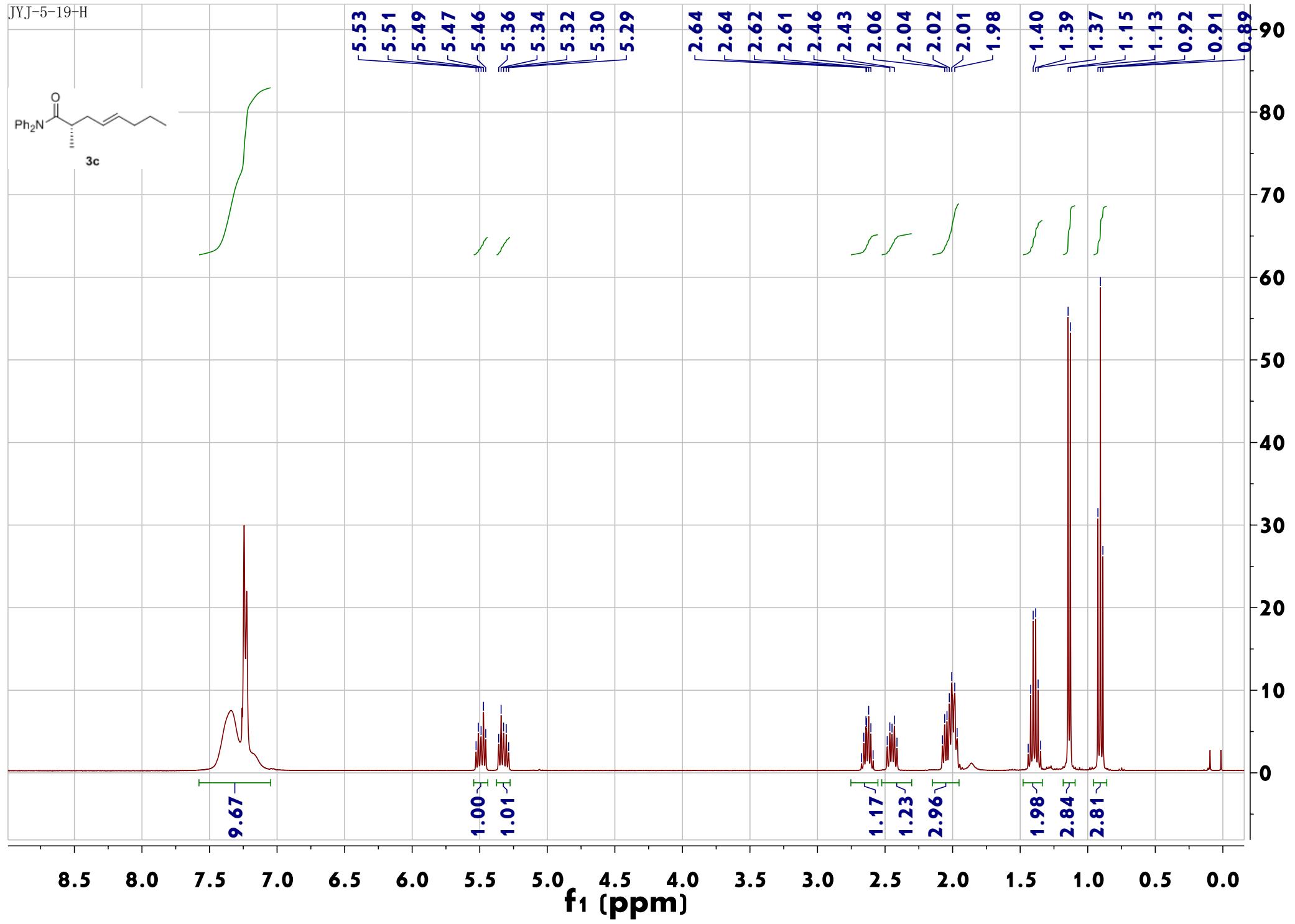


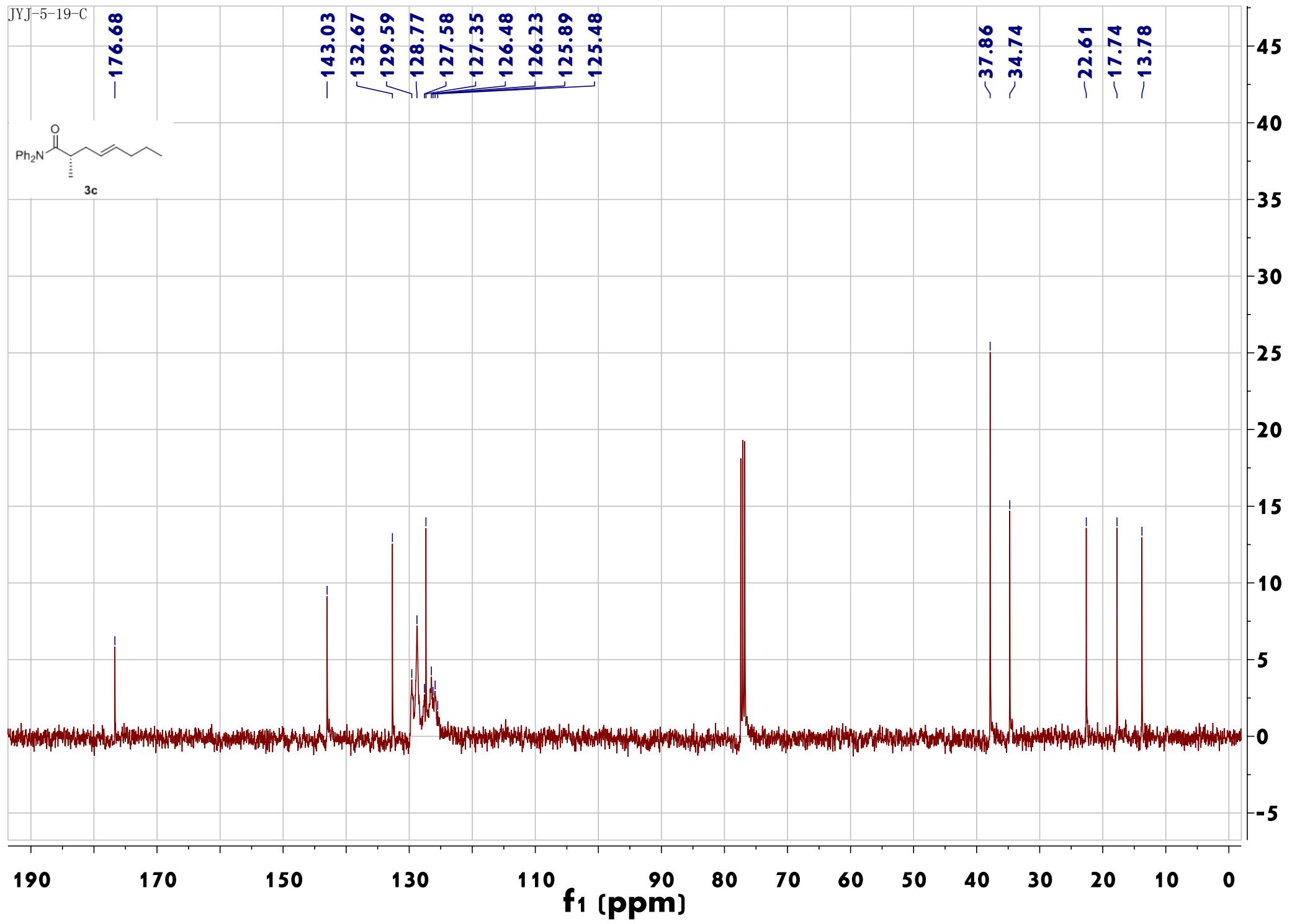
1 Det.A Ch1/214nm

PeakTable

Detector A Ch1 214nm

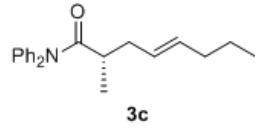
| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 9.876 | 14917162 | 871371 | 95.676 | 96.136 |
| 2 | 11.171 | 674226 | 35023 | 4.324 | 3.864 |
| Total | | 15591388 | 906394 | 100.000 | 100.000 |



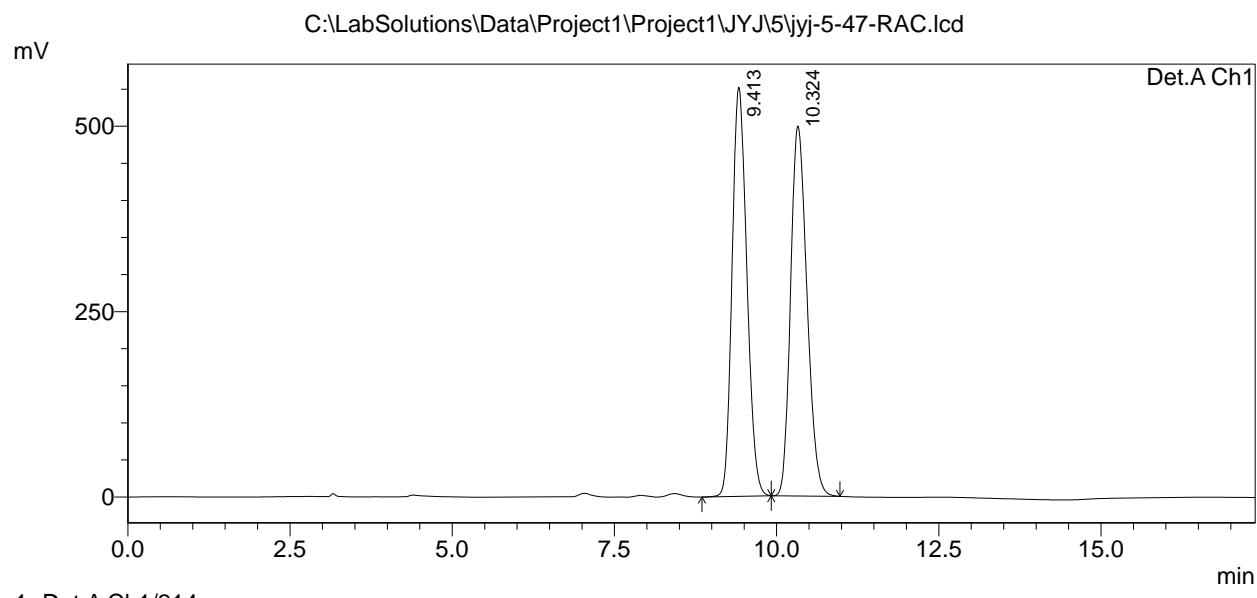


==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : jyj-5-47-RAC
 Sample ID : OD-H,99/1,1,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : jyj-5-47-RAC.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-9-11 10:13:03
 Data Processed : 2016-9-11 10:44:03



<Chromatogram>



1 Det.A Ch1/214nm

PeakTable

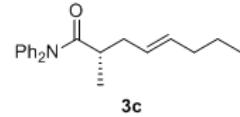
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|---------|---------|----------|
| 1 | 9.413 | 8753559 | 551879 | 49.937 | 52.524 |
| 2 | 10.324 | 8775757 | 498831 | 50.063 | 47.476 |
| Total | | 17529316 | 1050710 | 100.000 | 100.000 |

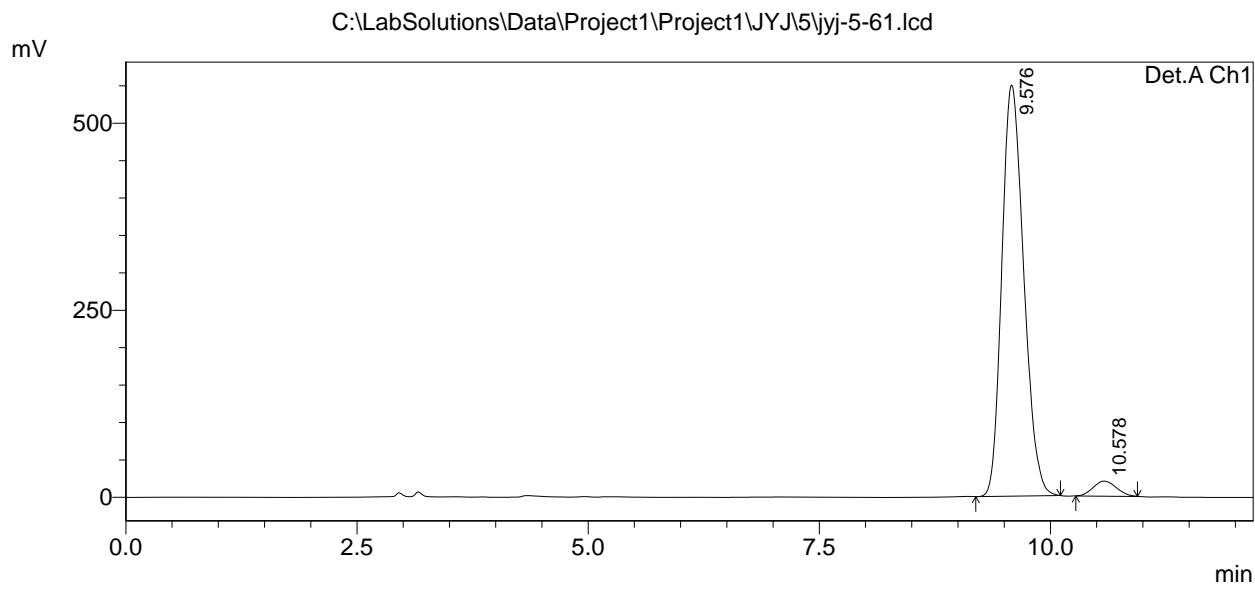
==== Shimadzu LCsolution Analysis Report ====

C:\LabSolutions\Data\Project1\Project1\JYJ\5\jyj-5-61.lcd

Acquired by : Admin
 Sample Name : jyj-5-61
 Sample ID : OD-H,99/1,1,214
 Vail # :
 Injection Volume : 2 uL
 Data File Name : jyj-5-61.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-9-24 22:32:18
 Data Processed : 2016-9-25 18:41:11



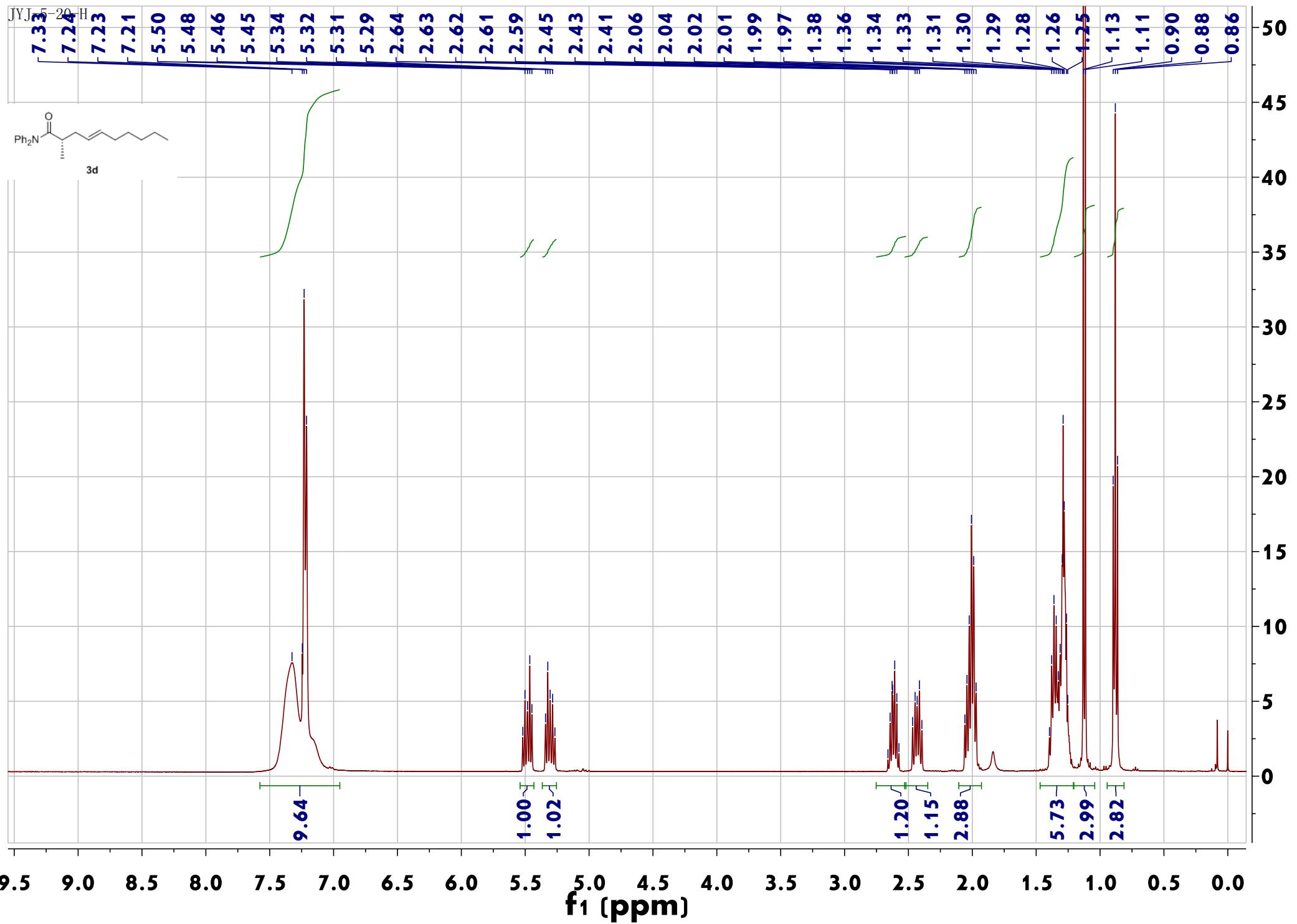
<Chromatogram>

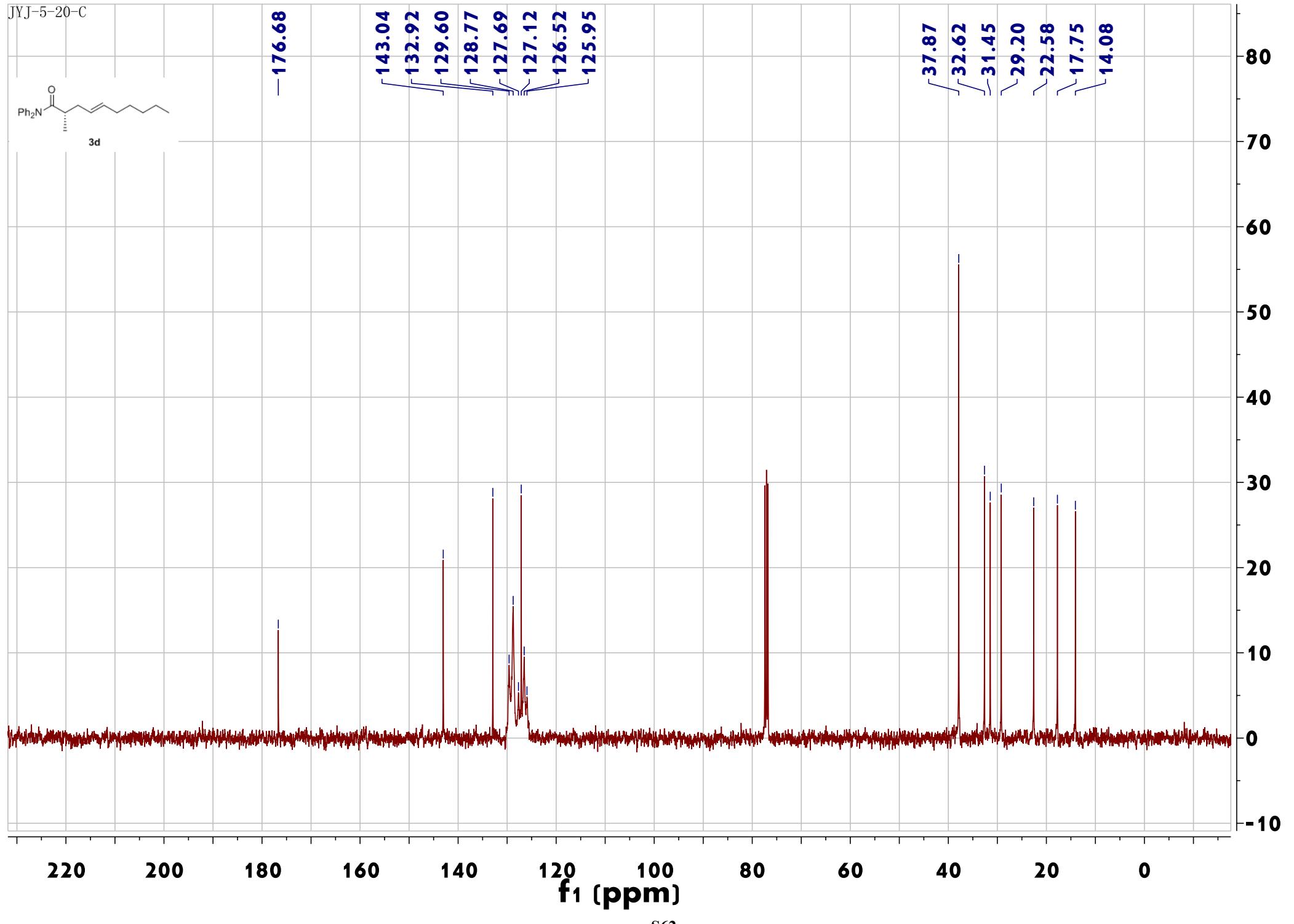


PeakTable

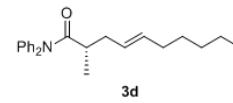
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|---------|--------|---------|----------|
| 1 | 9.576 | 9049392 | 549273 | 96.230 | 96.440 |
| 2 | 10.578 | 354551 | 20275 | 3.770 | 3.560 |
| Total | | 9403943 | 569548 | 100.000 | 100.000 |

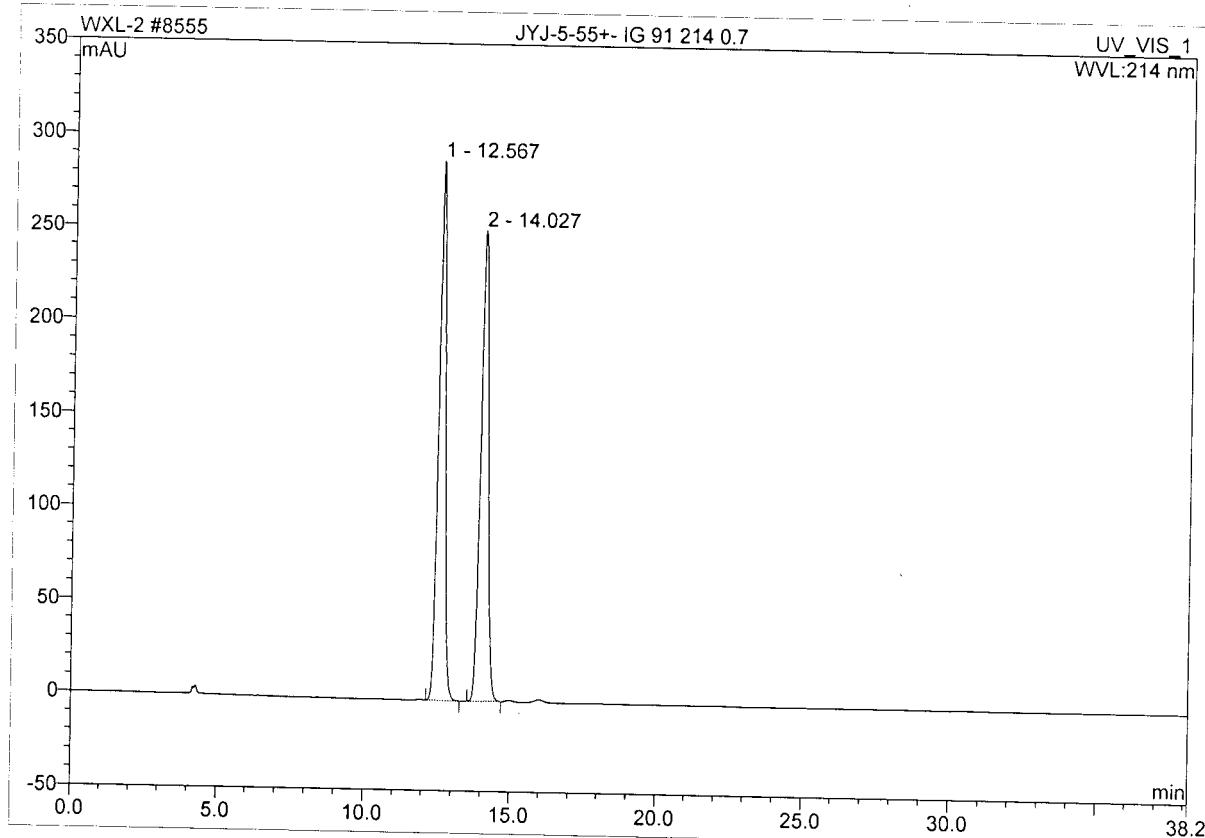




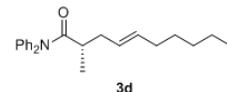
8555 JYJ-5-55+- IG 91 214 0.7



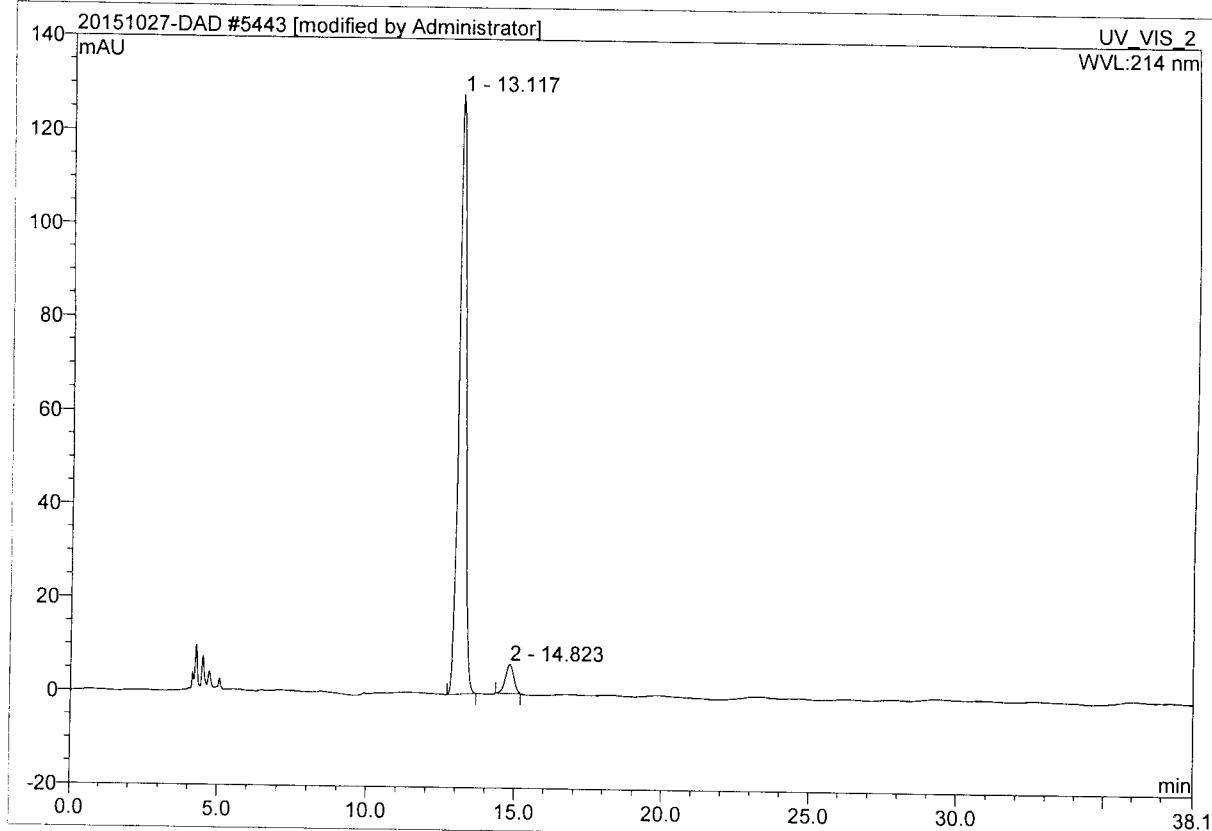
| | | | |
|-------------------------|--------------------------|--------------------------|----------|
| Sample Name: | JYJ-5-55+- IG 91 214 0.7 | <i>Injection Volume:</i> | 3.0 |
| Vial Number: | RC6 | <i>Channel:</i> | UV_VIS_1 |
| Sample Type: | unknown | <i>Wavelength:</i> | 214 |
| Control Program: | WXL-2014-4 | <i>Bandwidth:</i> | n.a. |
| Quantif. Method: | WXL | <i>Dilution Factor:</i> | 1.0000 |
| Recording Time: | 2016/9/21 13:09 | <i>Sample Weight:</i> | 1.0000 |
| Run Time (min): | 38.15 | <i>Sample Amount:</i> | 1.0000 |



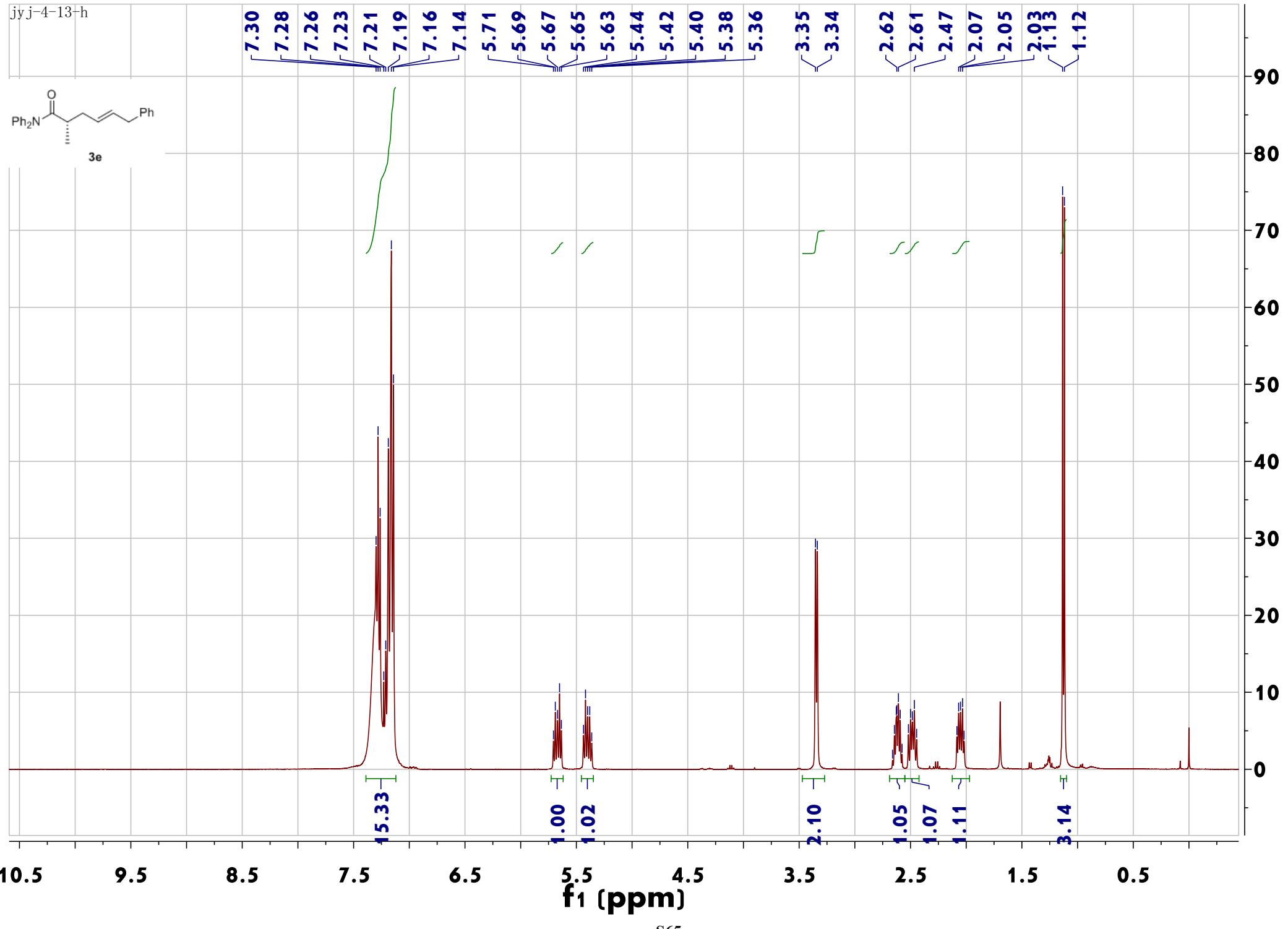
| No. | Ret.Time min | Peak Name | Height mAU | Area mAU*min | Rel.Area % | Amount | Type |
|---------------|-----------------|-----------|---------------|-----------------|---------------|--------|------|
| 1 | 12.57 | n.a. | 289.356 | 78.989 | 50.38 | n.a. | BMB |
| 2 | 14.03 | n.a. | 252.287 | 77.808 | 49.62 | n.a. | BMB |
| Total: | | | 541.642 | 156.797 | 100.00 | 0.000 | |

5443 JYJ-7-8 IG 91 214 0.7

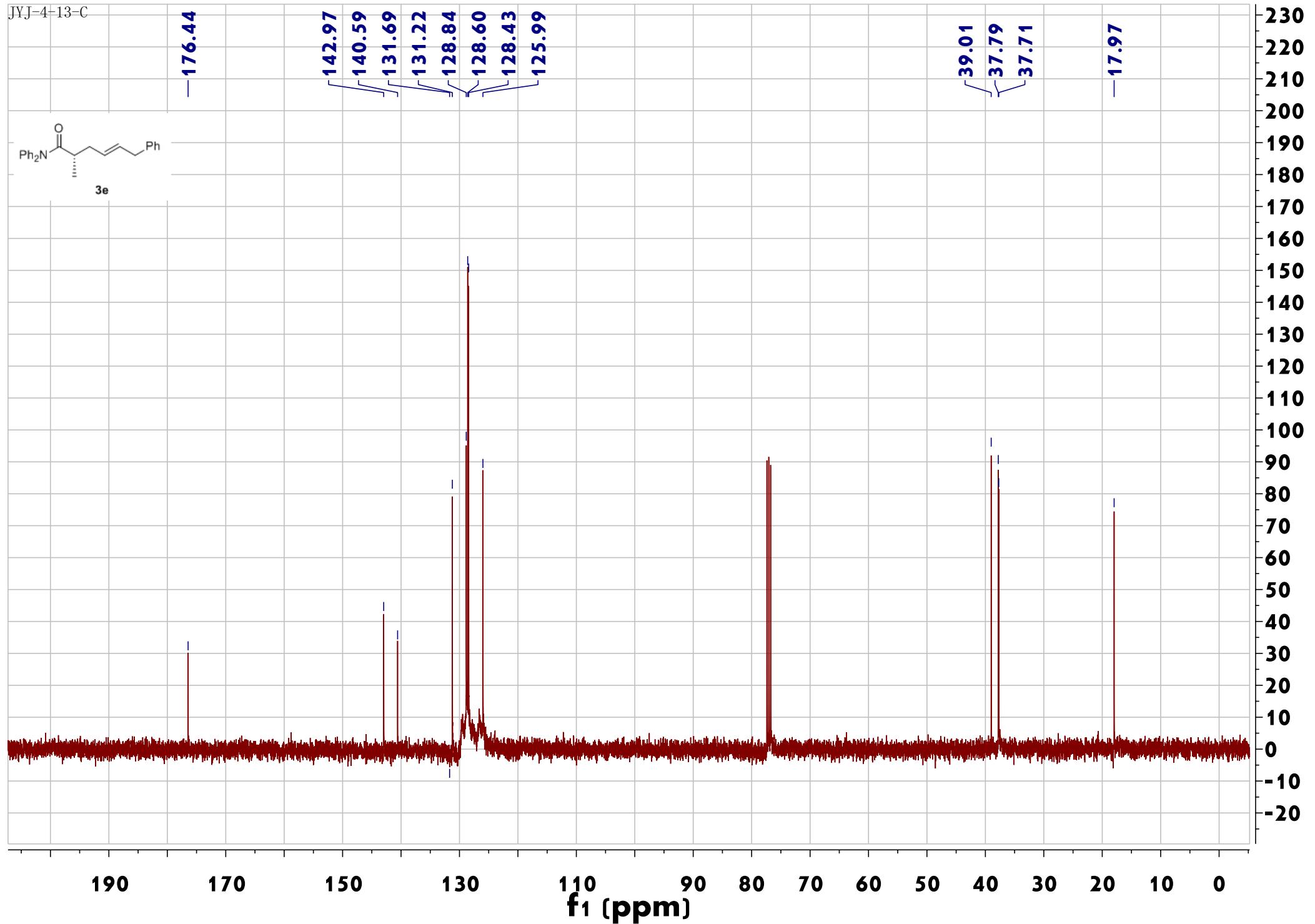
| | | | |
|-------------------------|------------------------------|--------------------------|-----------------|
| Sample Name: | JYJ-7-8 IG 91 214 0.7 | Injection Volume: | 2.0 |
| Vial Number: | RD4 | Channel: | UV_VIS_2 |
| Sample Type: | unknown | Wavelength: | 214.0 |
| Control Program: | test-dad5 | Bandwidth: | 4 |
| Quantif. Method: | WXL | Dilution Factor: | 1.0000 |
| Recording Time: | 2017-2-24 10:35 | Sample Weight: | 1.0000 |
| Run Time (min): | 38.09 | Sample Amount: | 1.0000 |



| No. | Ret.Time min | Peak Name | Height mAU | Area mAU*min | Rel.Area % | Amount | Type |
|---------------|-----------------|-----------|---------------|-----------------|---------------|--------|------|
| 1 | 13.12 | n.a. | 128.146 | 36.392 | 94.61 | n.a. | BMB |
| 2 | 14.82 | n.a. | 6.244 | 2.071 | 5.39 | n.a. | BMB |
| Total: | | | 134.390 | 38.463 | 100.00 | 0.000 | |

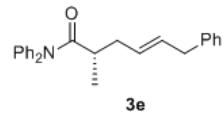


JYJ-4-13-C

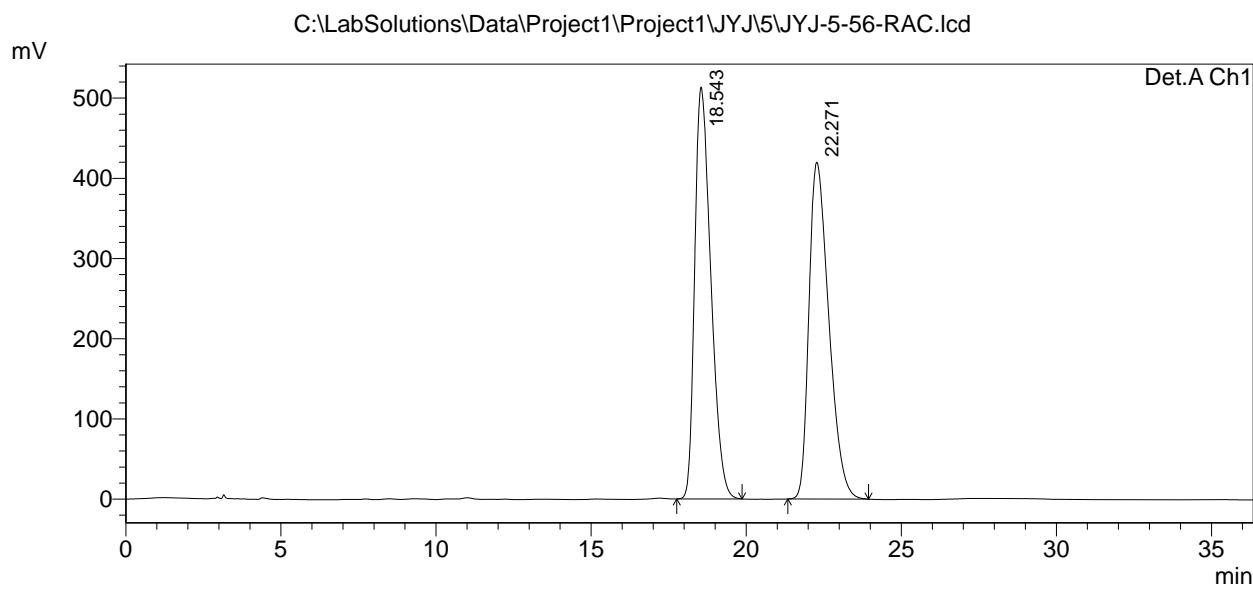


==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : JYJ-5-56-RAC
 Sample ID : OD-H,99/1,1,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-5-56-RAC.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-9-19 10:02:38
 Data Processed : 2016-9-19 11:07:55



<Chromatogram>



1 Det.A Ch1/214nm

PeakTable

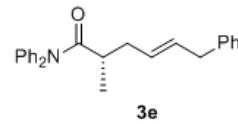
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 18.543 | 18578616 | 513453 | 49.820 | 55.007 |
| 2 | 22.271 | 18712804 | 419981 | 50.180 | 44.993 |
| Total | | 37291420 | 933434 | 100.000 | 100.000 |

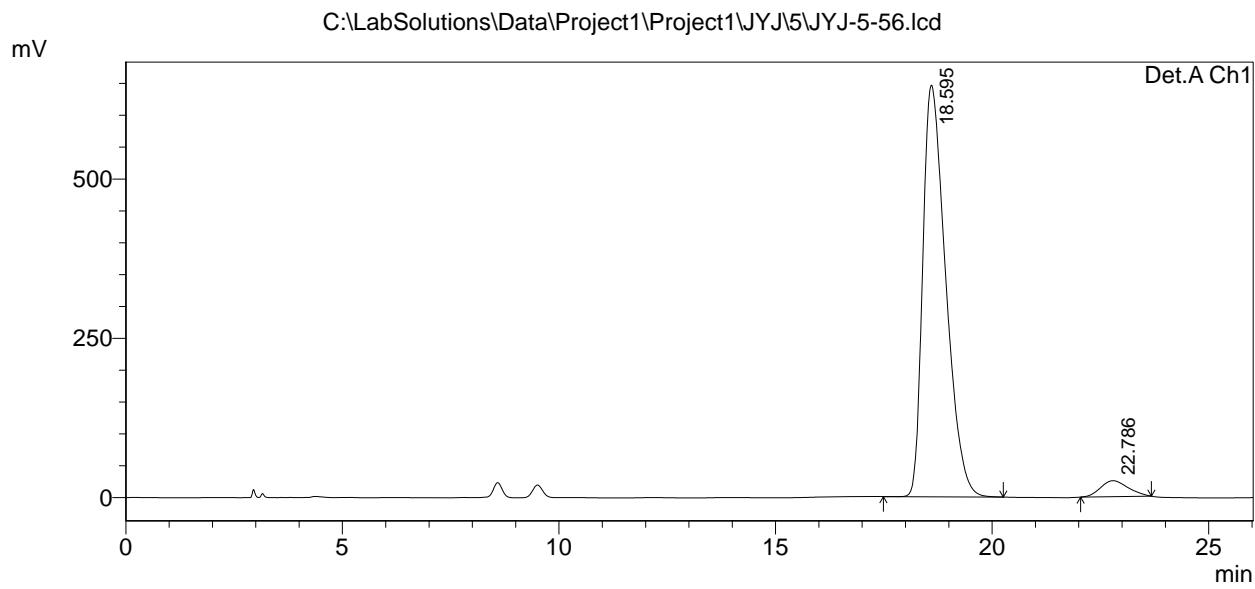
==== Shimadzu LCsolution Analysis Report ====

C:\LabSolutions\Data\Project1\Project1\JYJ\5\JYJ-5-56.lcd

Acquired by : Admin
 Sample Name : JYJ-5-56
 Sample ID : OD-H,99/1,1,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-5-56.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-9-19 10:41:42
 Data Processed : 2016-9-19 11:11:00



<Chromatogram>

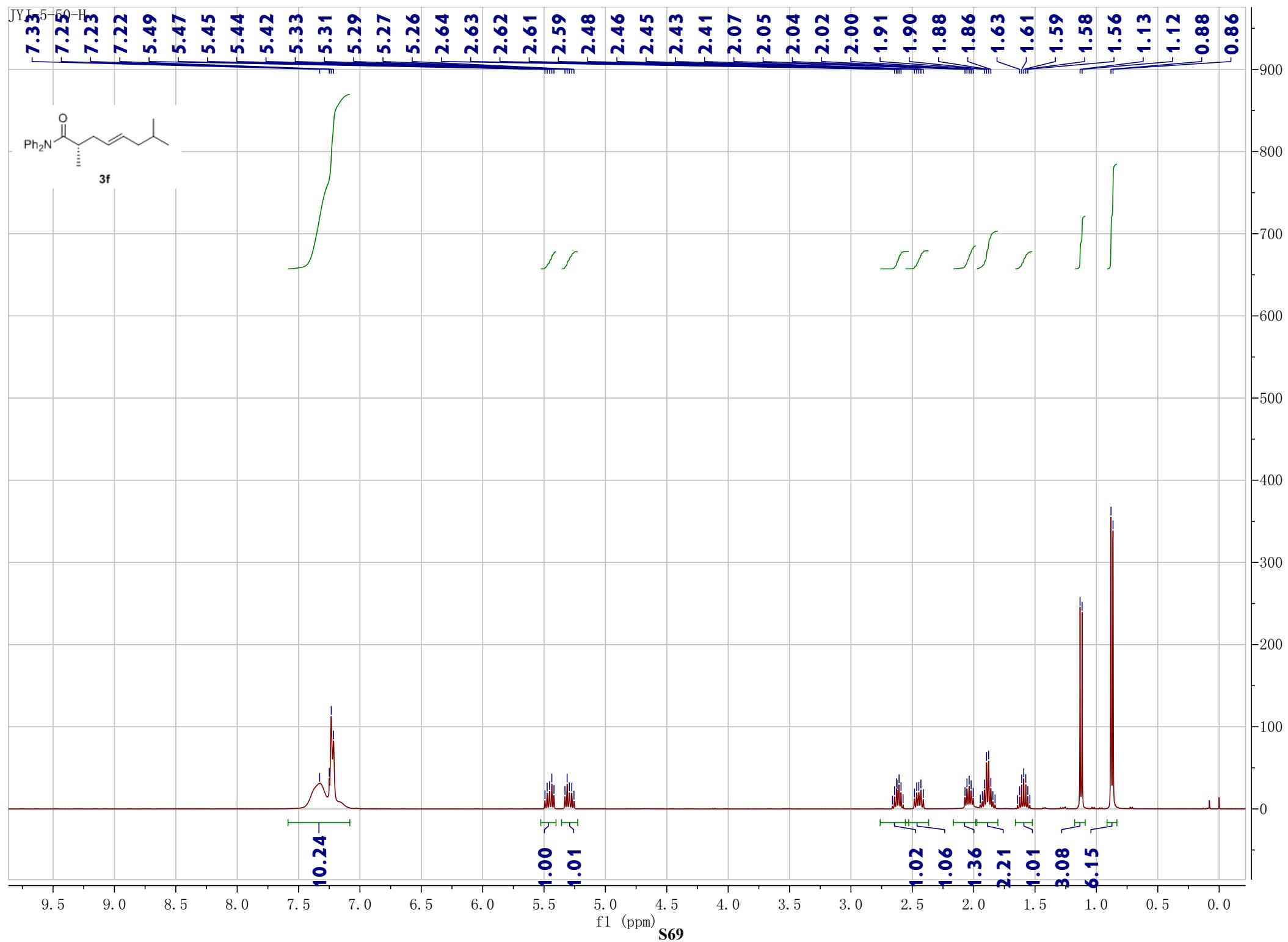


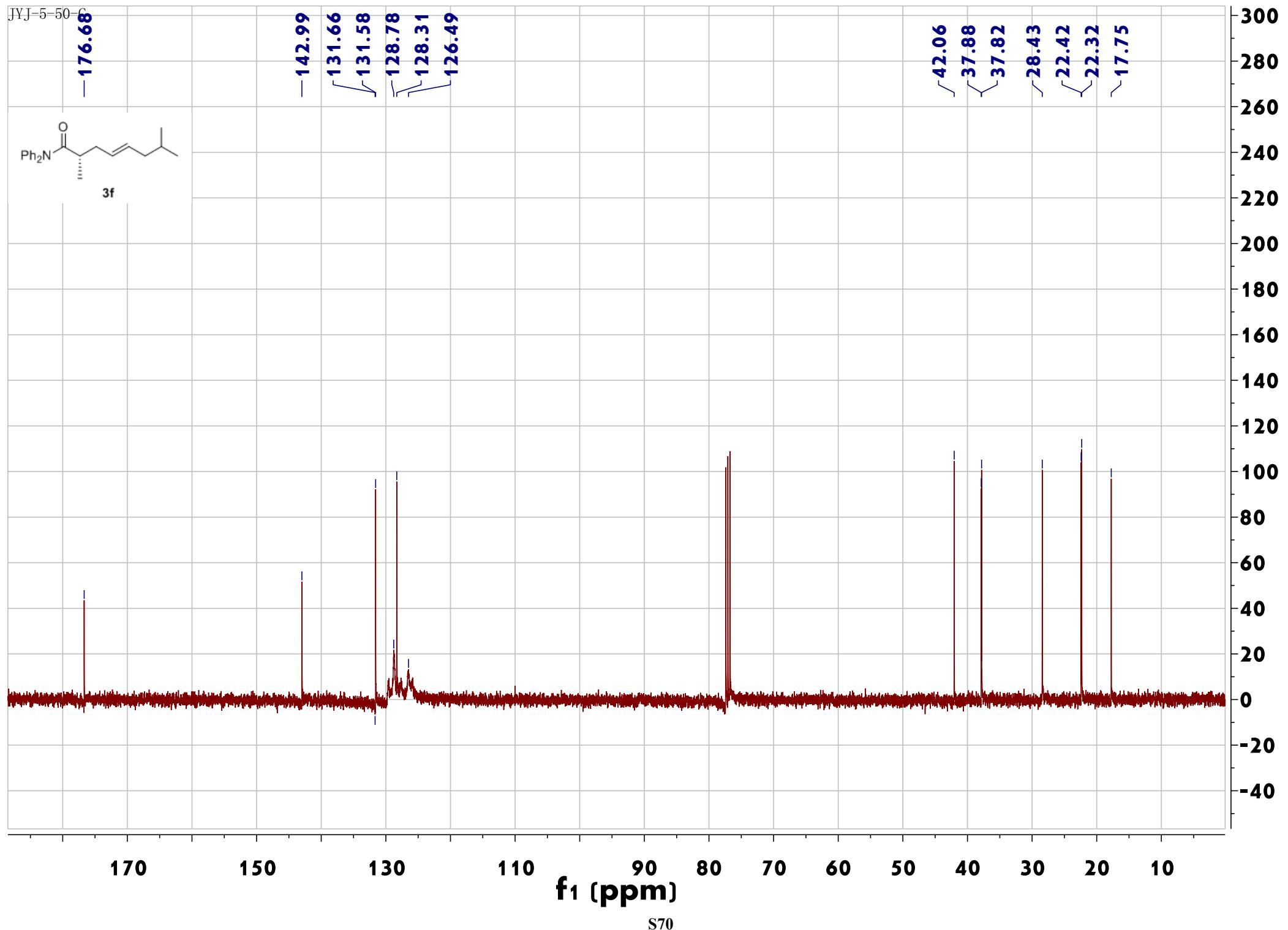
1 Det.A Ch1/214nm

PeakTable

Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 18.595 | 23764689 | 646177 | 95.501 | 96.255 |
| 2 | 22.786 | 1119647 | 25143 | 4.499 | 3.745 |
| Total | | 24884337 | 671320 | 100.000 | 100.000 |

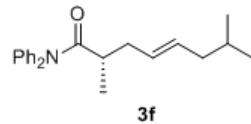




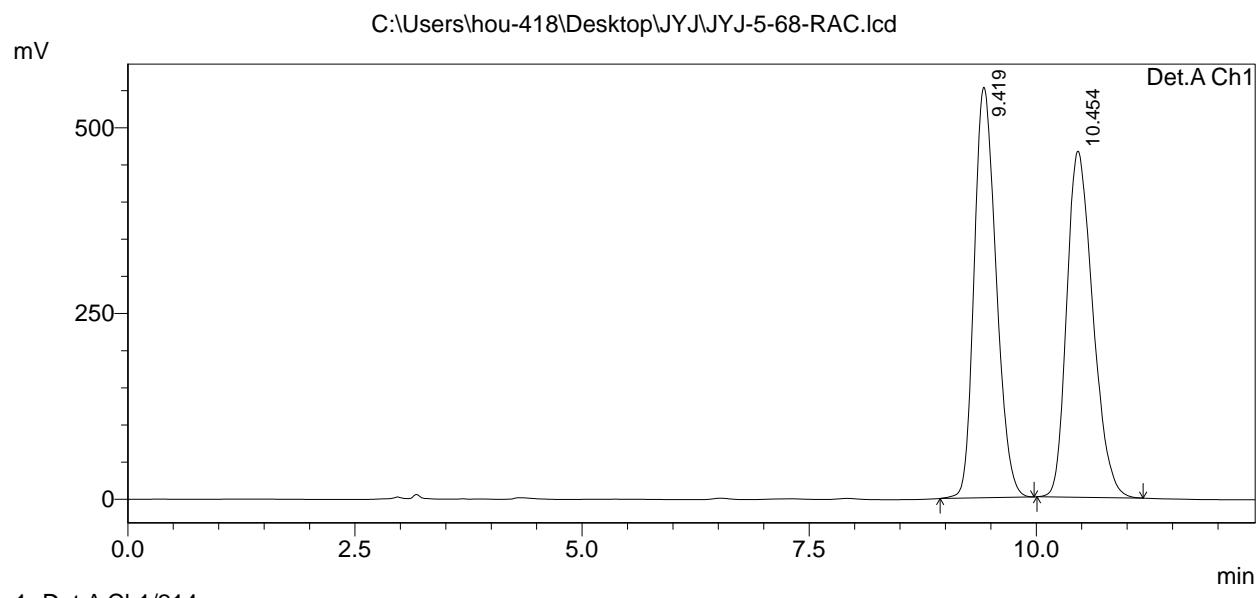
==== Shimadzu LCsolution Analysis Report ====

C:\Users\hou-418\Desktop\JYJ\JYJ-5-68-RAC.lcd

Acquired by : Admin
 Sample Name : JYJ-5-68-RAC
 Sample ID : OD-H,99/1,1,214
 Vial # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-5-68-RAC.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-10-11 21:03:39
 Data Processed : 2016-10-11 21:33:29



<Chromatogram>



PeakTable

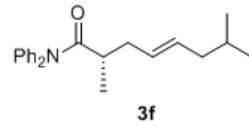
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|---------|---------|----------|
| 1 | 9.419 | 9308541 | 552707 | 49.943 | 54.257 |
| 2 | 10.454 | 9329847 | 465970 | 50.057 | 45.743 |
| Total | | 18638389 | 1018677 | 100.000 | 100.000 |

==== Shimadzu LCsolution Analysis Report ====

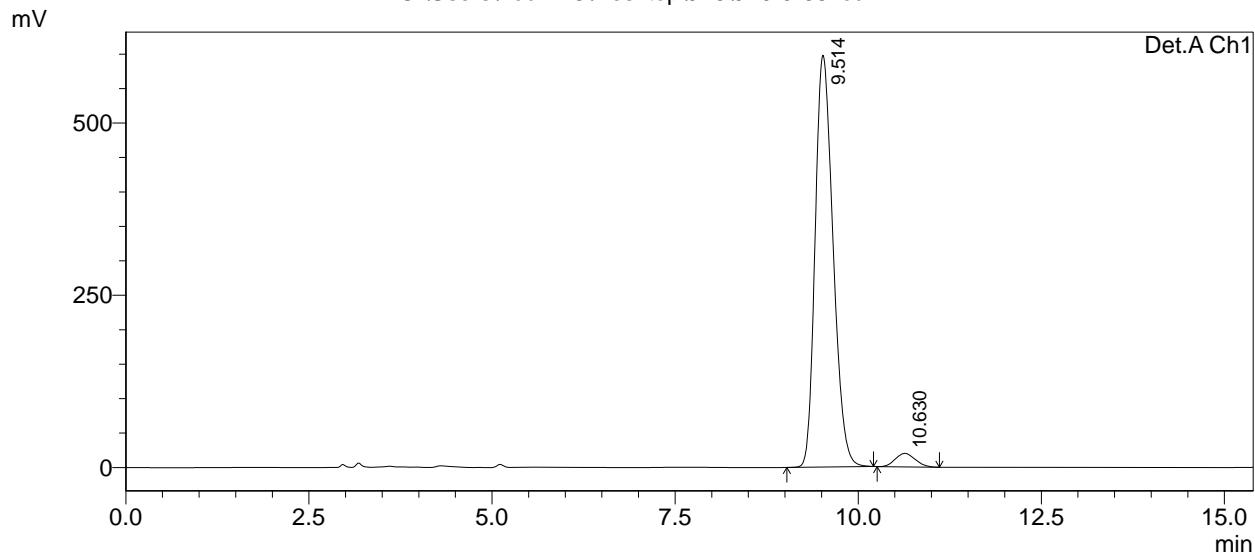
C:\Users\hou-418\Desktop\JYJ\JYJ-5-68.lcd

Acquired by : Admin
 Sample Name : JYJ-5-68
 Sample ID : OD-H,99/1,1,214
 Vial # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-5-68.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-10-11 21:17:15
 Data Processed : 2016-10-12 8:43:59



<Chromatogram>

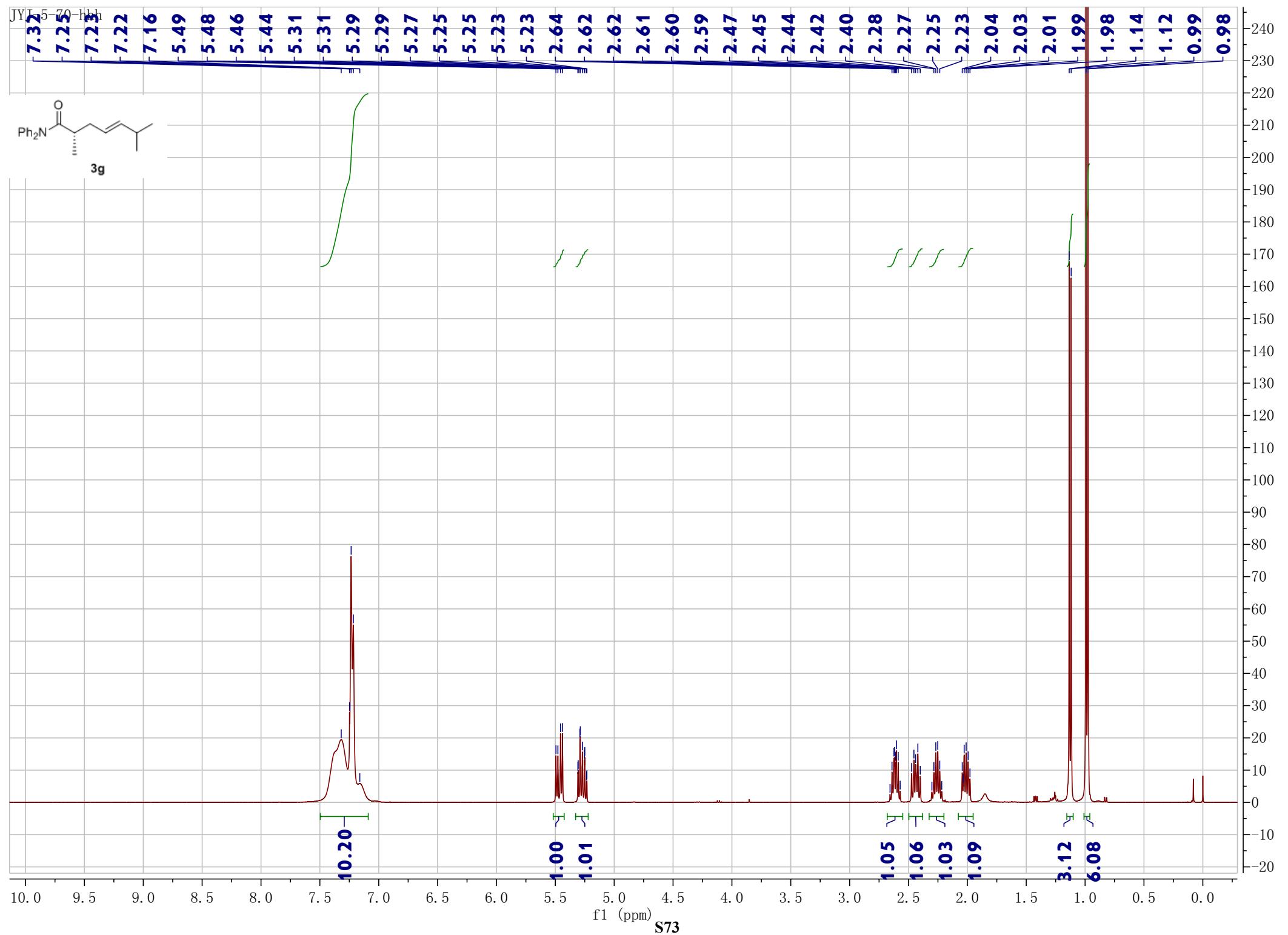
C:\Users\hou-418\Desktop\JYJ\JYJ-5-68.lcd



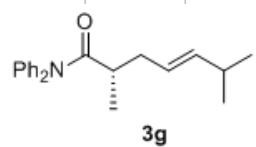
PeakTable

Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 9.514 | 10170178 | 597868 | 96.445 | 96.822 |
| 2 | 10.630 | 374859 | 19624 | 3.555 | 3.178 |
| Total | | 10545036 | 617492 | 100.000 | 100.000 |



JYJ-5-70-CCC

-176.73**3g**

✓ **142.99**
✓ **139.84**
✓ **134.86**
✓ **128.73**
✓ **126.49**
✓ **124.14**
✓ **119.82**

✓ **~37.92**
✓ **31.03**
✓ **22.62**
✓ **22.55**
✓ **-17.87**

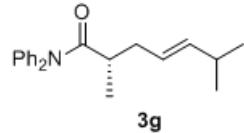
190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

f1 (ppm) **S74**

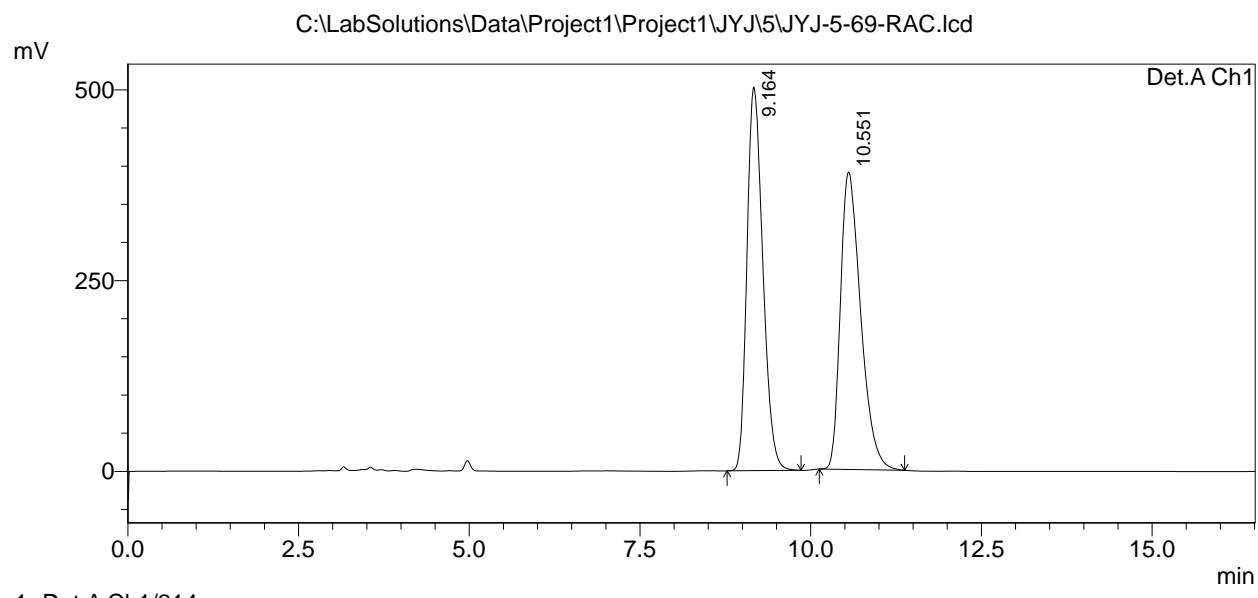
130
120
110
100
90
80
70
60
50
40
30
20
10
0
-10

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : JYJ-5-69-RAC
 Sample ID : OD-H,99/1,1,214
 Vail # :
 Injection Volume : 2 uL
 Data File Name : JYJ-5-69-RAC.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-10-11 20:12:05
 Data Processed : 2016-10-11 20:46:58



<Chromatogram>



PeakTable

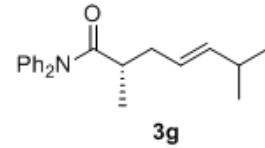
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 9.164 | 8130042 | 502691 | 50.219 | 56.330 |
| 2 | 10.551 | 8059217 | 389718 | 49.781 | 43.670 |
| Total | | 16189258 | 892409 | 100.000 | 100.000 |

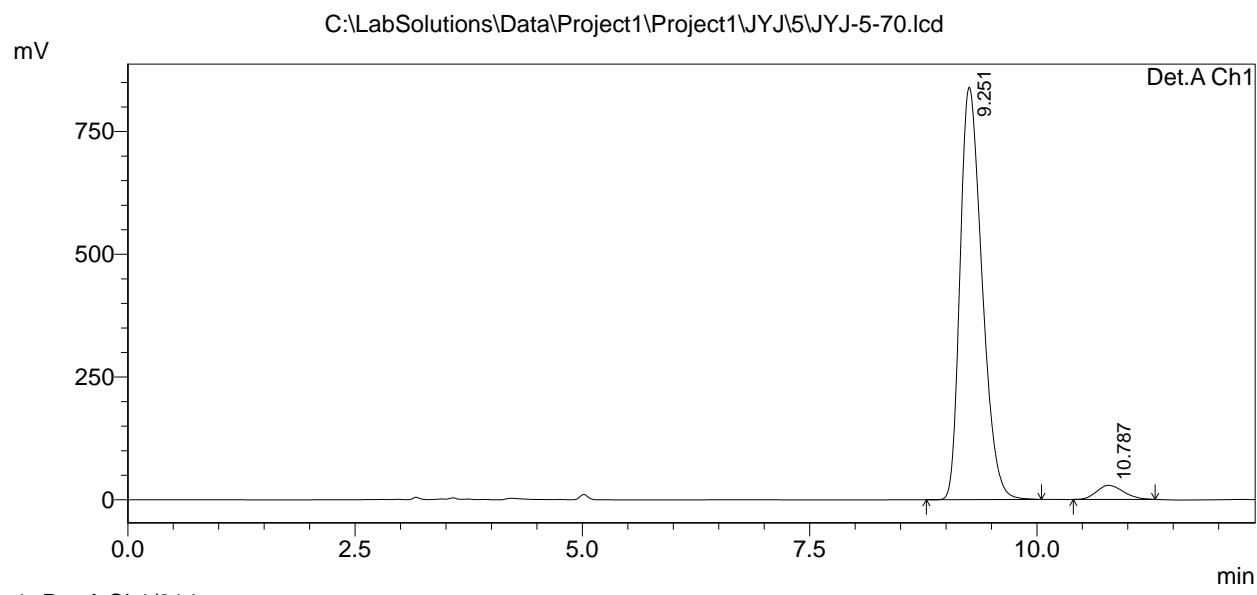
==== Shimadzu LCsolution Analysis Report ====

C:\LabSolutions\Data\Project1\Project1\JYJ\5\JYJ-5-70.lcd

Acquired by : Admin
 Sample Name : JYJ-5-70
 Sample ID : OD-H,99/1,1,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-5-70.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-10-11 20:34:17
 Data Processed : 2016-10-11 20:47:42



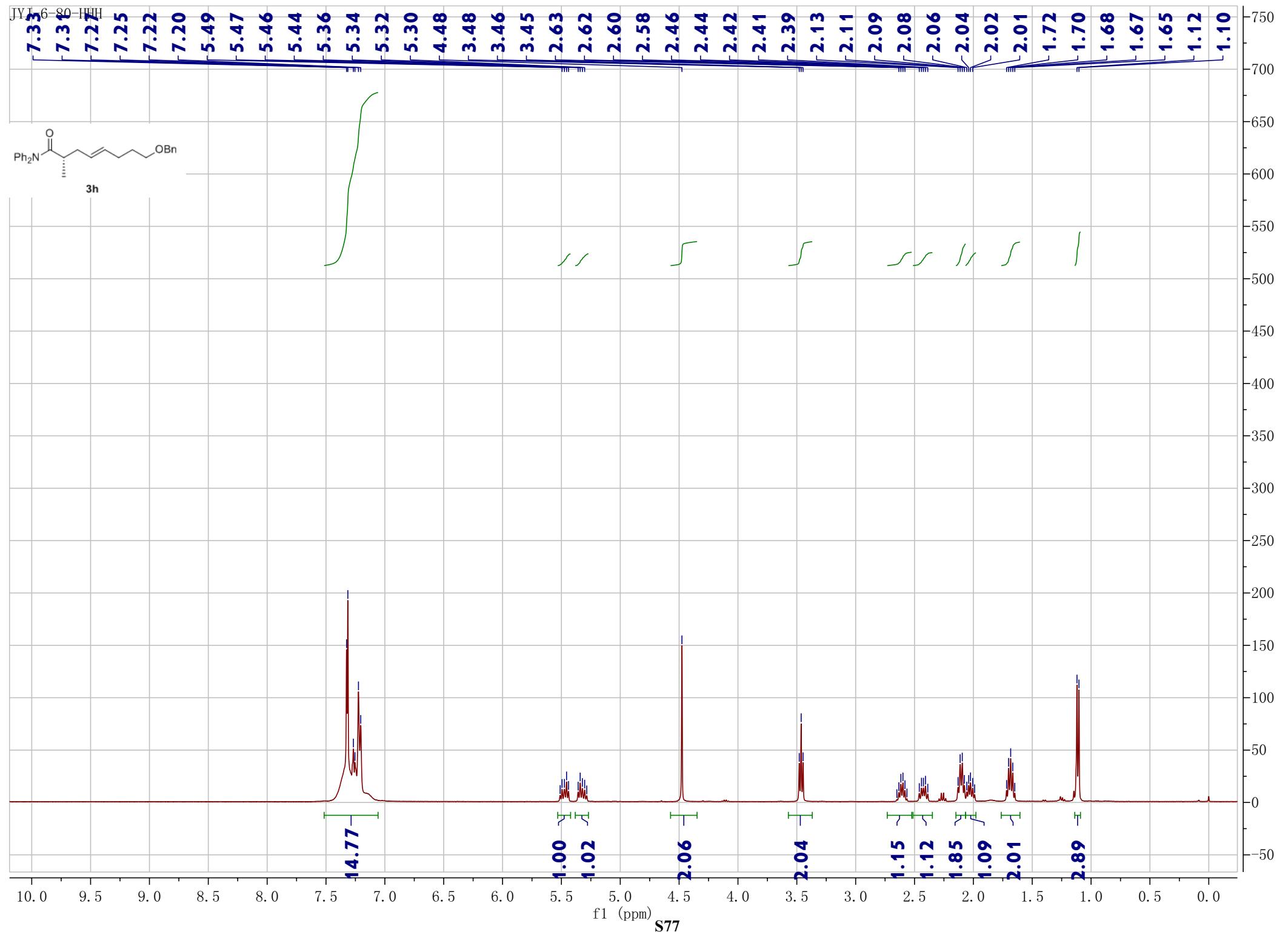
<Chromatogram>

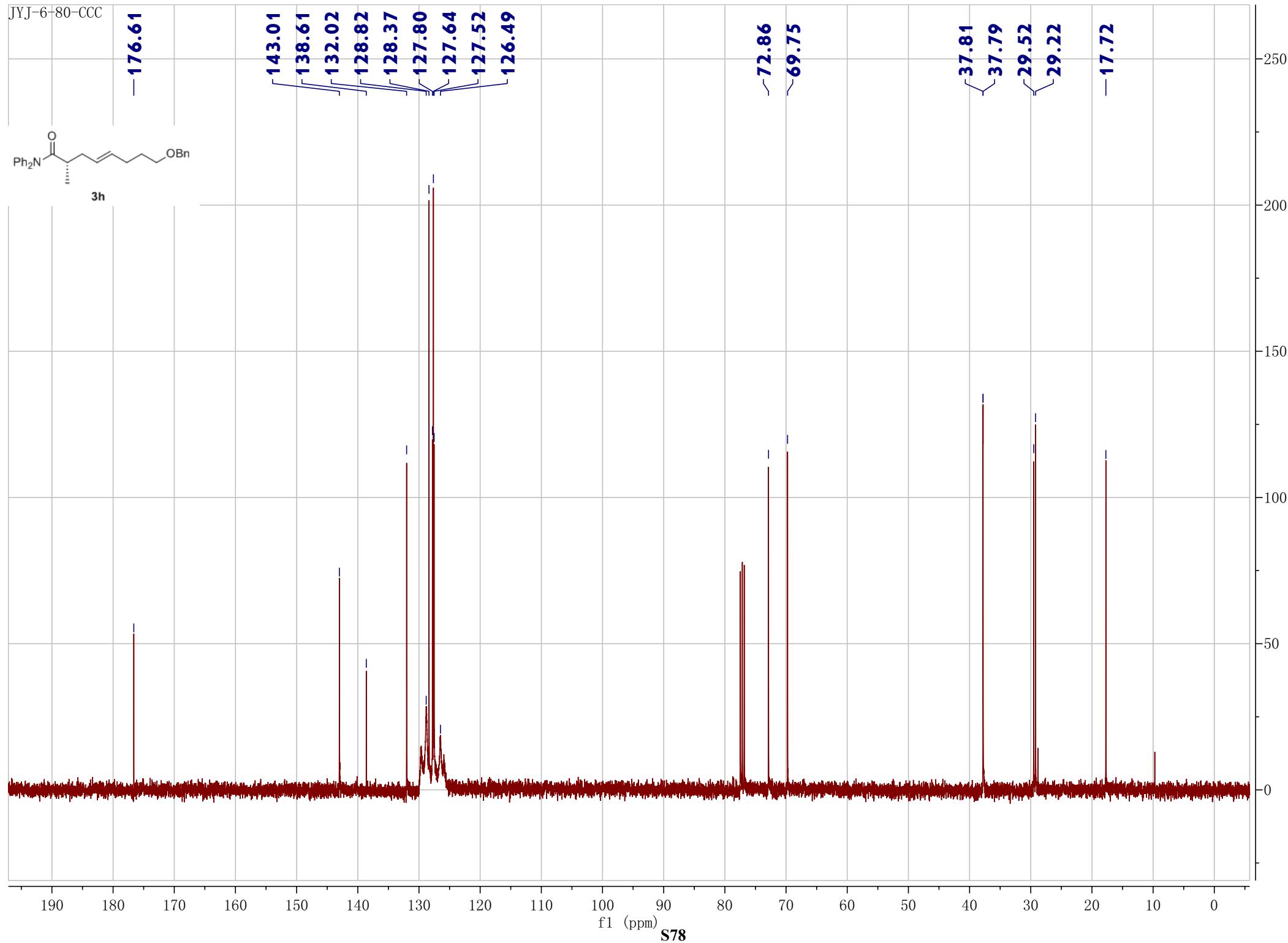


PeakTable

Detector A Ch1 214nm

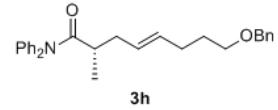
| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 9.251 | 14105938 | 840023 | 95.943 | 96.670 |
| 2 | 10.787 | 596475 | 28933 | 4.057 | 3.330 |
| Total | | 14702413 | 868956 | 100.000 | 100.000 |



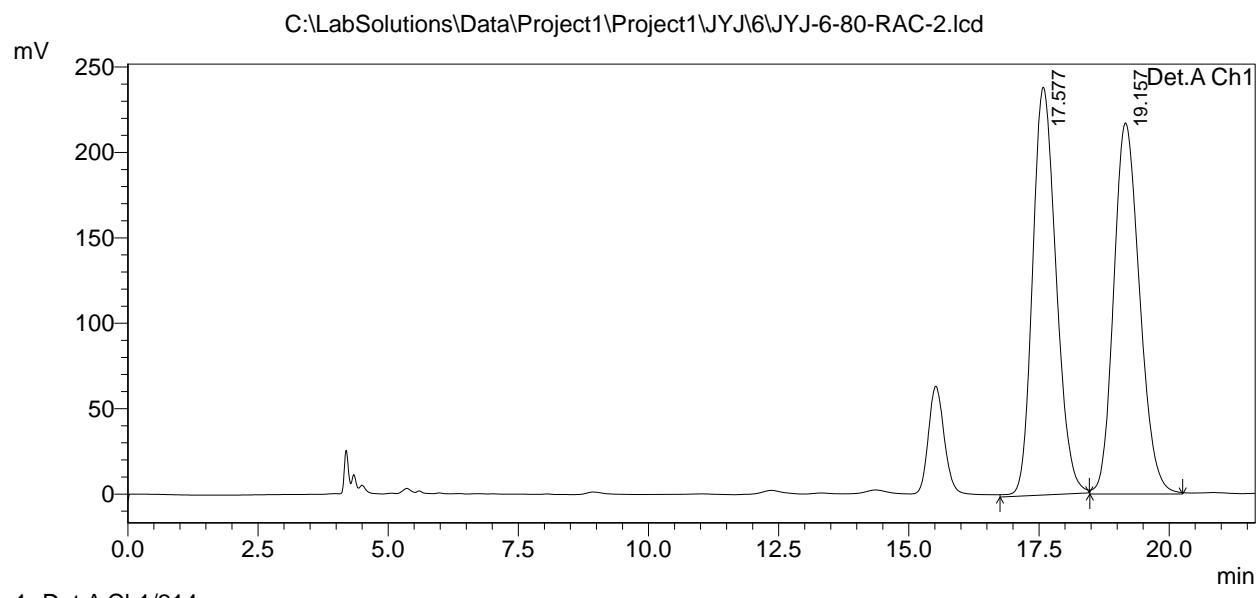


==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : JYJ-6-80-RAC-2
 Sample ID : OD-H,95/5,0.7,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-6-80-RAC-2.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2017-1-7 19:06:03
 Data Processed : 2017-1-7 19:52:37



<Chromatogram>



1 Det.A Ch1/214nm

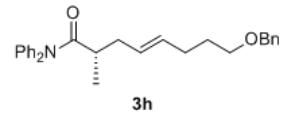
PeakTable

Detector A Ch1 214nm

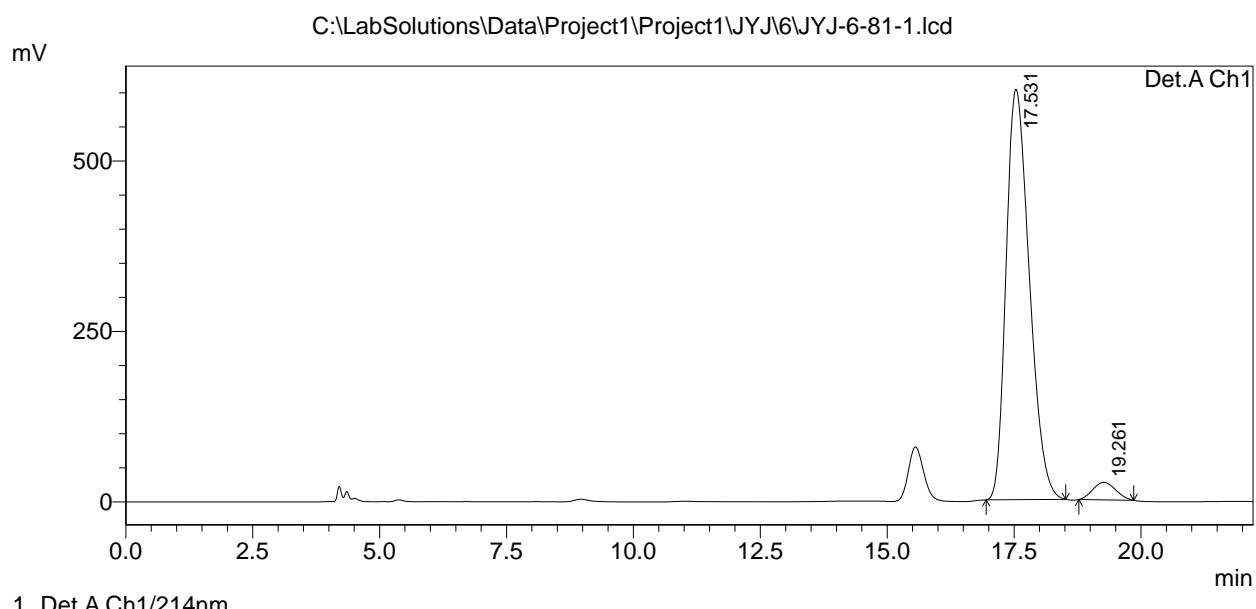
| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 17.577 | 7520350 | 238806 | 50.013 | 52.368 |
| 2 | 19.157 | 7516554 | 217205 | 49.987 | 47.632 |
| Total | | 15036903 | 456011 | 100.000 | 100.000 |

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : JYJ-6-81-1
 Sample ID : OD-H,95/5,0.7,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-6-81-1.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2017-1-7 19:28:46
 Data Processed : 2017-7-3 11:13:04



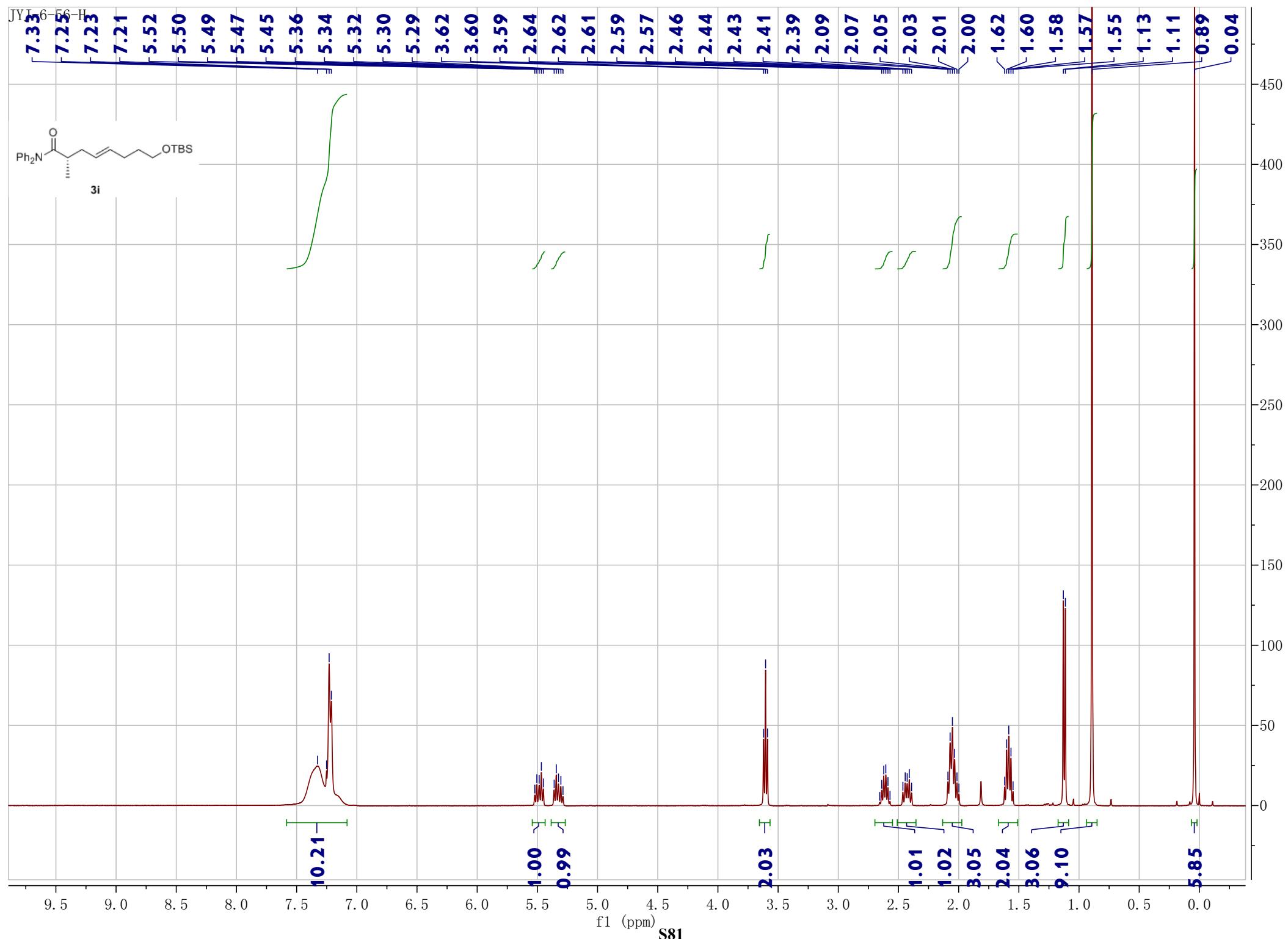
<Chromatogram>



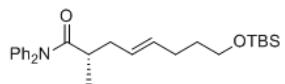
PeakTable

Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 17.531 | 19196460 | 602139 | 95.964 | 95.902 |
| 2 | 19.261 | 807257 | 25728 | 4.036 | 4.098 |
| Total | | 20003716 | 627867 | 100.000 | 100.000 |



JYJ-6-56-C

-176.63**3i****-143.00**
132.23
129.63
128.77
127.52
126.51**-62.59****37.81**
32.59
28.86
25.97
18.34
17.68**-5.24**

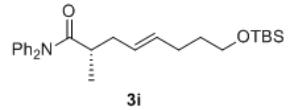
190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm)

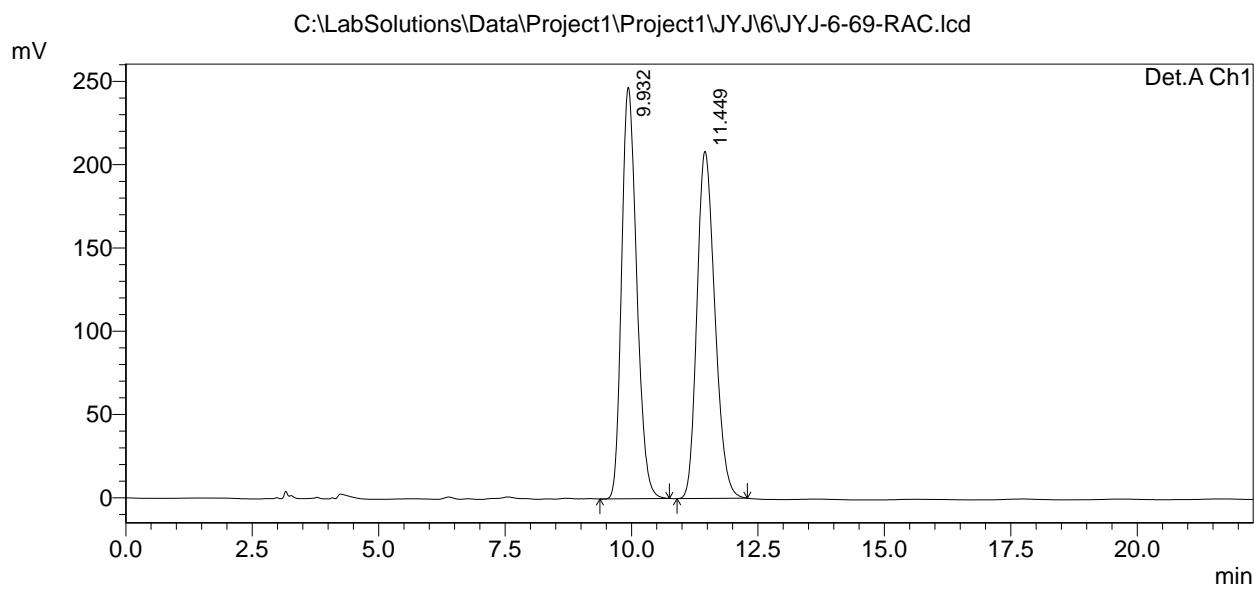
S82

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : JYJ-6-69-RAC
 Sample ID : OD-H,99/1,1,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-6-69-RAC.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-12-30 9:42:02
 Data Processed : 2016-12-30 10:05:05



<Chromatogram>



1 Det.A Ch1/214nm

PeakTable

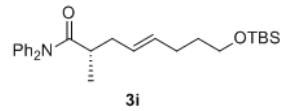
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 9.932 | 5134543 | 247163 | 49.991 | 54.248 |
| 2 | 11.449 | 5136435 | 208457 | 50.009 | 45.752 |
| Total | | 10270978 | 455621 | 100.000 | 100.000 |

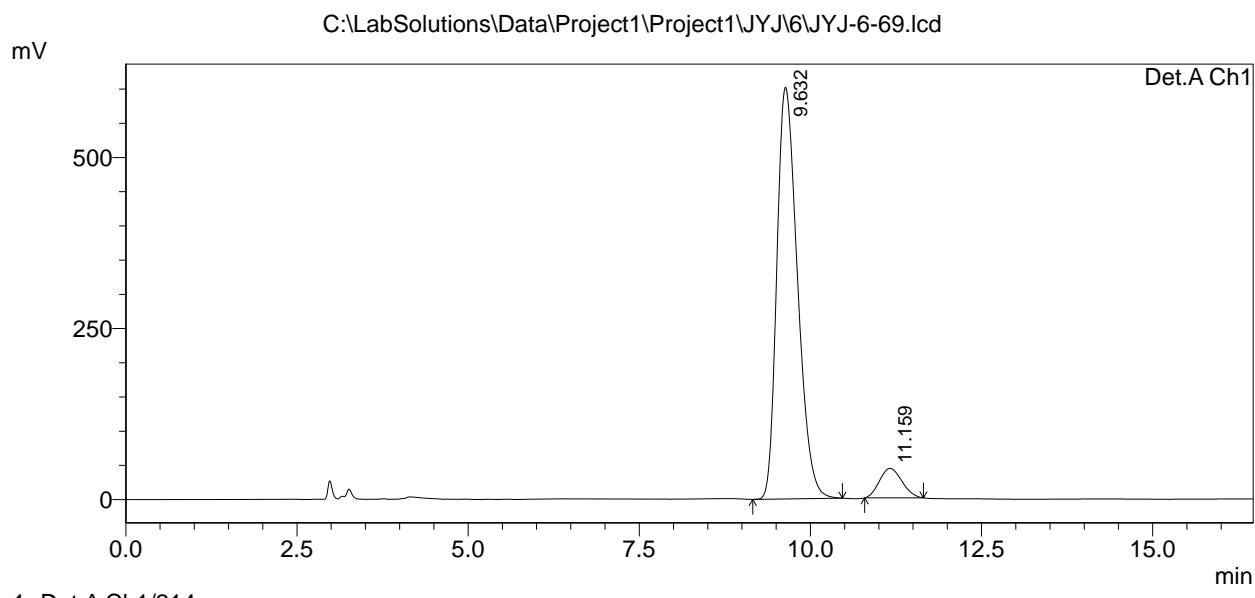
==== Shimadzu LCsolution Analysis Report ====

C:\LabSolutions\Data\Project1\Project1\JYJ\6\JYJ-6-69.lcd

Acquired by : Admin
 Sample Name : JYJ-6-69-1
 Sample ID : OD-H,99/1,1,214
 Vial # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-6-69.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-12-30 10:07:18
 Data Processed : 2016-12-30 10:24:45



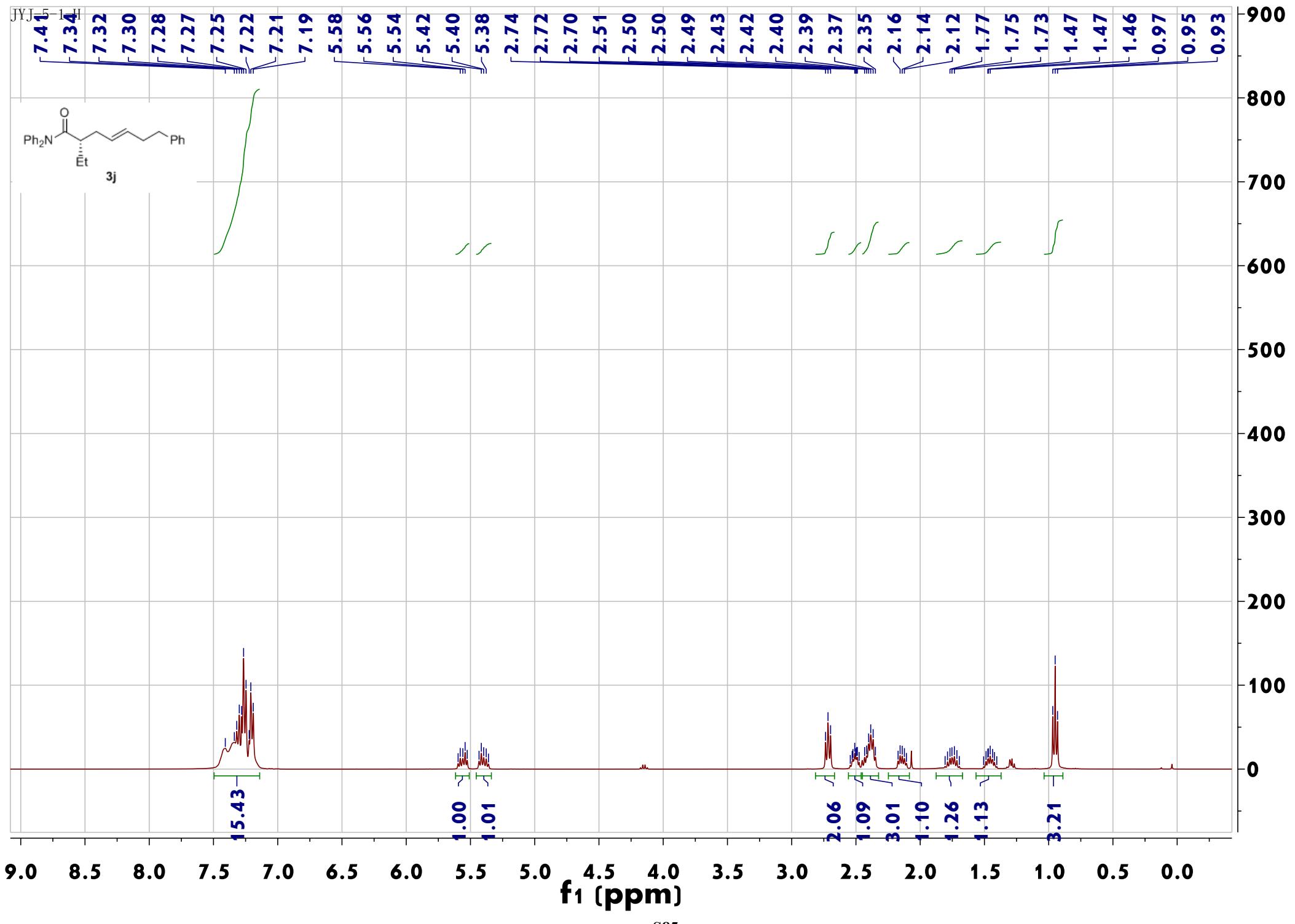
<Chromatogram>

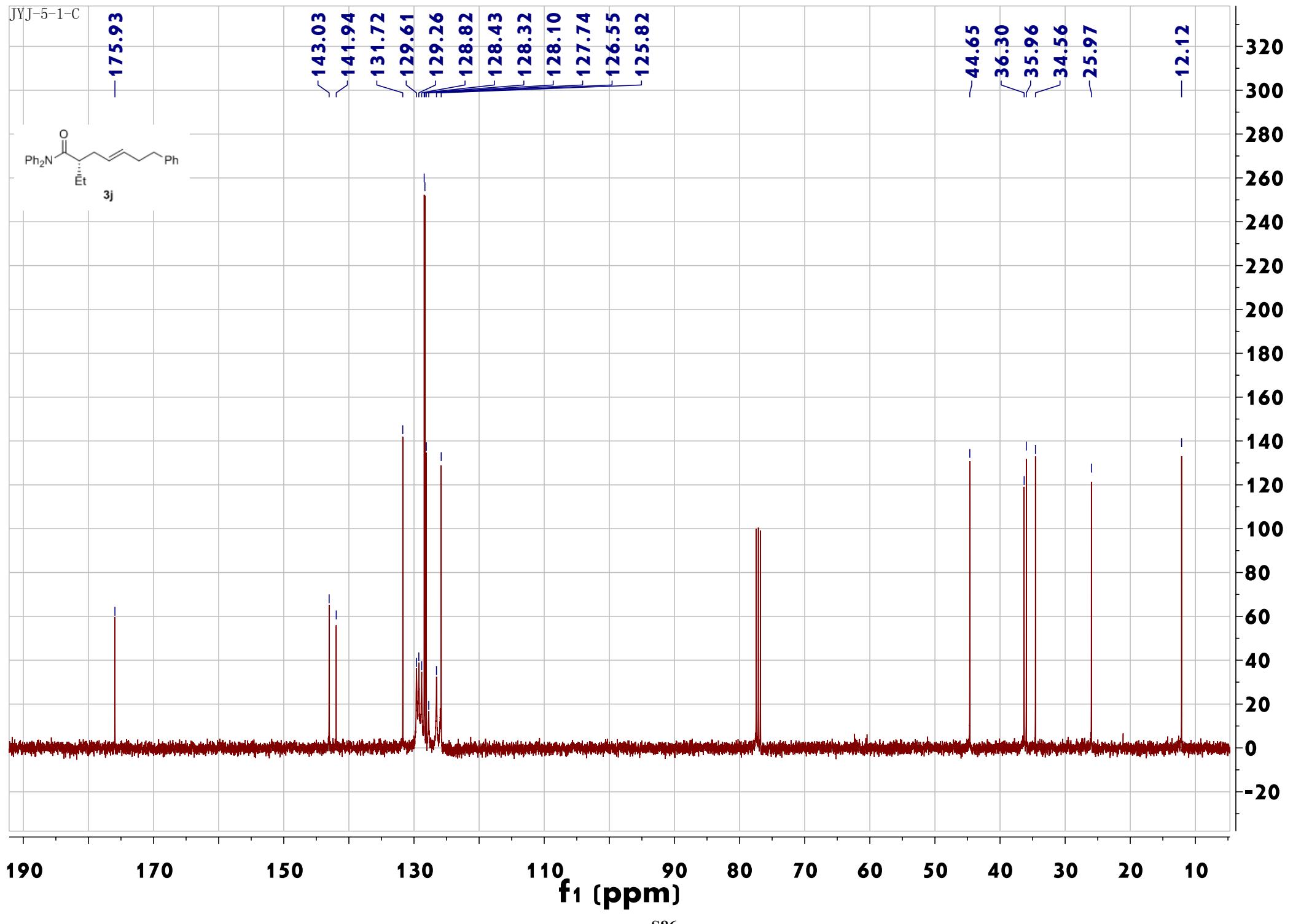


PeakTable

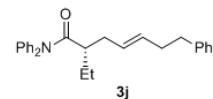
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 9.632 | 12604649 | 601958 | 92.758 | 93.317 |
| 2 | 11.159 | 984139 | 43109 | 7.242 | 6.683 |
| Total | | 13588787 | 645067 | 100.000 | 100.000 |

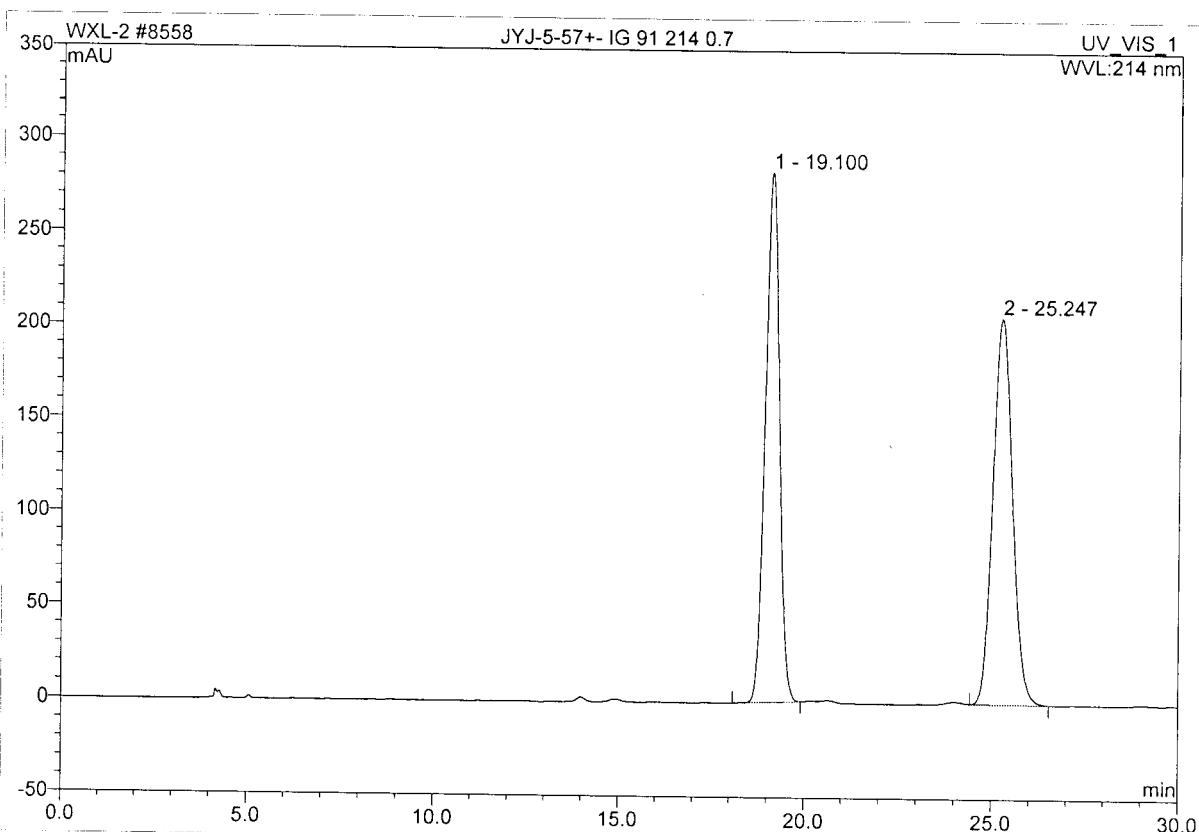




8558 JYJ-5-57+- IG 91 214 0.7

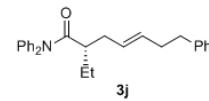


| | | | |
|-------------------------|--------------------------|--------------------------|----------|
| <i>Sample Name:</i> | JYJ-5-57+- IG 91 214 0.7 | <i>Injection Volume:</i> | 3.0 |
| <i>Vial Number:</i> | RC5 | <i>Channel:</i> | UV_VIS_1 |
| <i>Sample Type:</i> | unknown | <i>Wavelength:</i> | 214 |
| <i>Control Program:</i> | WXL-2014 | <i>Bandwidth:</i> | n.a. |
| <i>Quantif. Method:</i> | WXL | <i>Dilution Factor:</i> | 1.0000 |
| <i>Recording Time:</i> | 2016/9/21 14:42 | <i>Sample Weight:</i> | 1.0000 |
| <i>Run Time (min):</i> | 30.00 | <i>Sample Amount:</i> | 1.0000 |

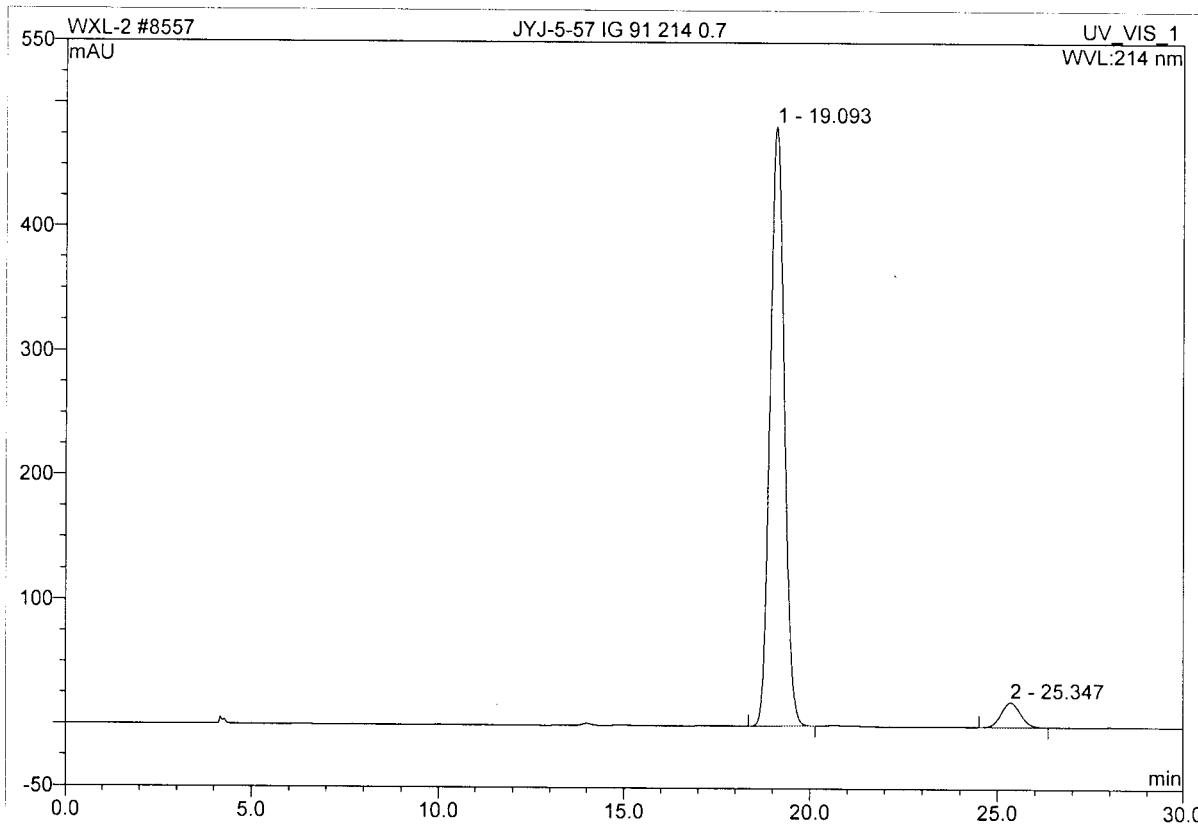


| No. | Ret.Time min | Peak Name | Height mAU | Area mAU*min | Rel.Area % | Amount | Type |
|---------------|-----------------|-----------|---------------|-----------------|---------------|--------|------|
| 1 | 19.10 | n.a. | 282.777 | 123.700 | 49.80 | n.a. | BMB |
| 2 | 25.25 | n.a. | 206.131 | 124.710 | 50.20 | n.a. | BMB |
| Total: | | | 488.908 | 248.410 | 100.00 | 0.000 | |

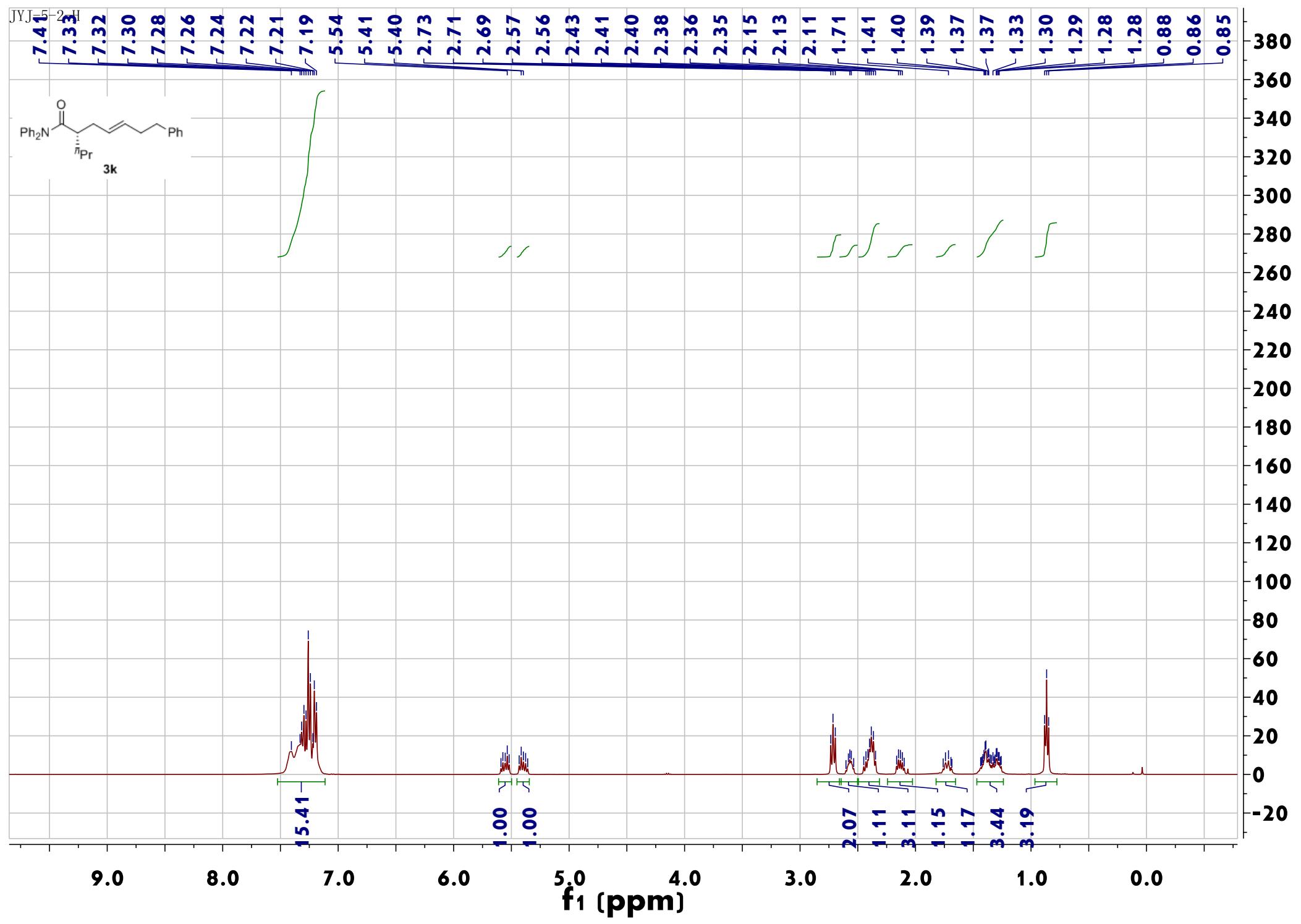
8557 JYJ-5-57 IG 91 214 0.7



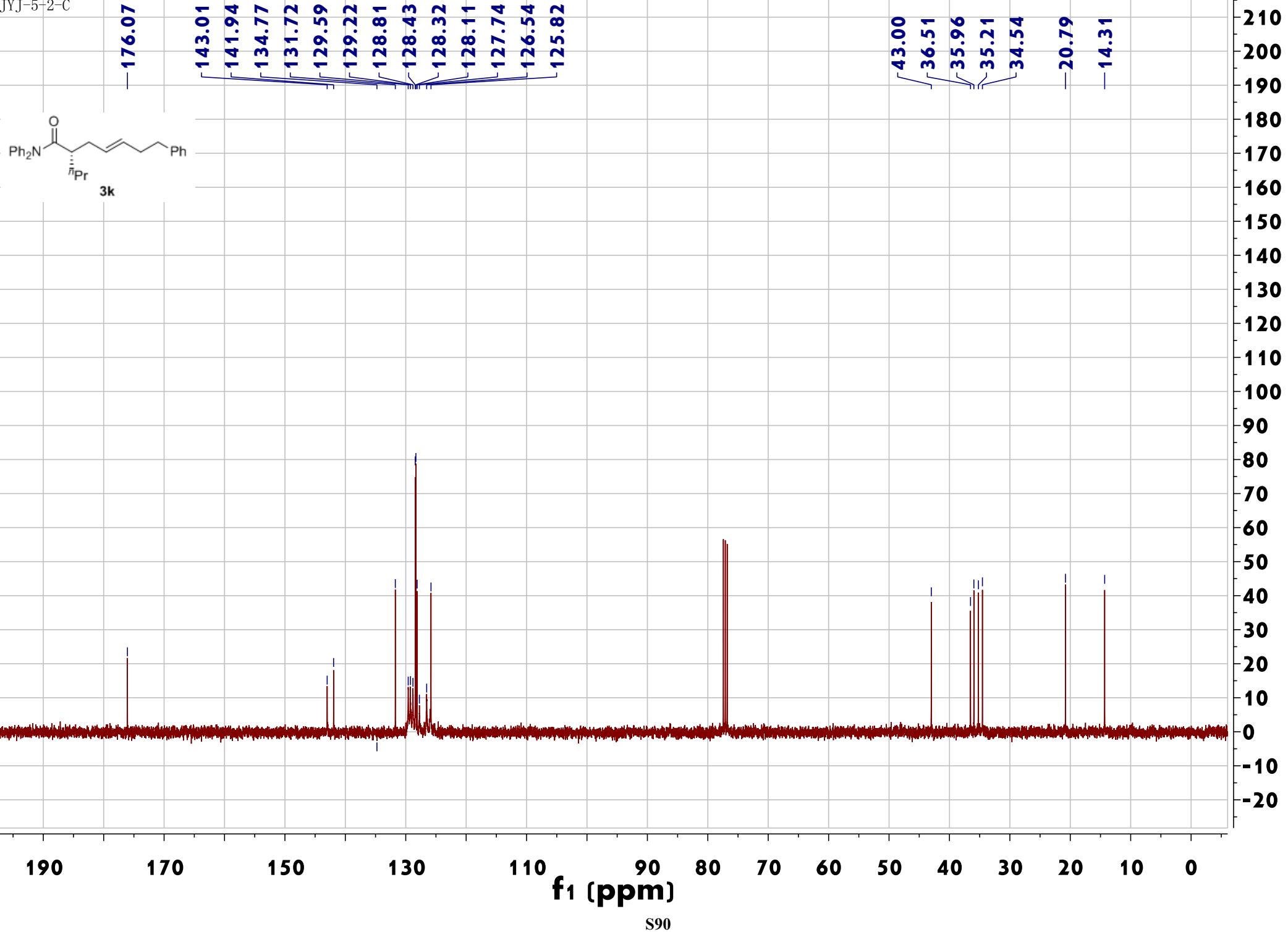
| | | | |
|------------------|------------------------|-------------------|----------|
| Sample Name: | JYJ-5-57 IG 91 214 0.7 | Injection Volume: | 3.0 |
| Vial Number: | RD7 | Channel: | UV_VIS_1 |
| Sample Type: | unknown | Wavelength: | 214 |
| Control Program: | WXL-2014 | Bandwidth: | n.a. |
| Quantif. Method: | WXL | Dilution Factor: | 1.0000 |
| Recording Time: | 2016/9/21 14:10 | Sample Weight: | 1.0000 |
| Run Time (min): | 29.96 | Sample Amount: | 1.0000 |



| No. | Ret.Time min | Peak Name | Height mAU | Area mAU*min | Rel.Area % | Amount | Type |
|---------------|-----------------|-----------|---------------|-----------------|---------------|--------|------|
| 1 | 19.09 | n.a. | 482.582 | 214.439 | 94.71 | n.a. | BMB |
| 2 | 25.35 | n.a. | 20.162 | 11.975 | 5.29 | n.a. | BMB |
| Total: | | | 502.744 | 226.414 | 100.00 | 0.000 | |

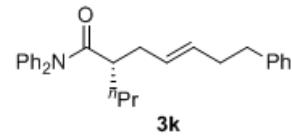


JYJ-5-2-C

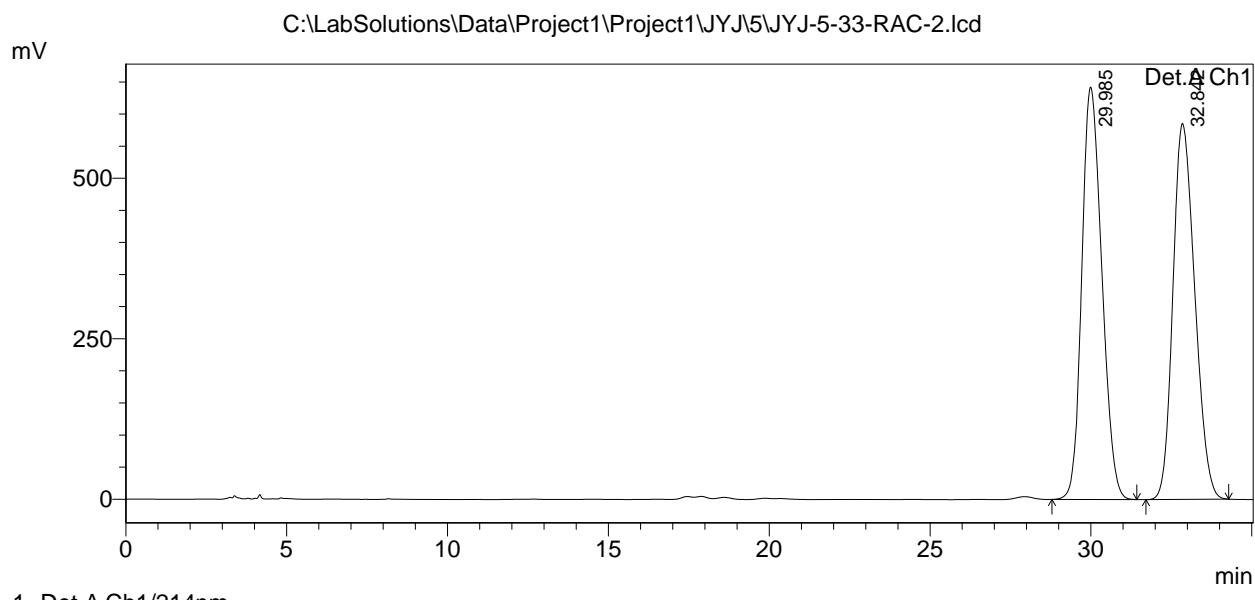


==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : JYJ-5-33-RAC-2
 Sample ID : IC,99/1,1,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-5-33-RAC-2.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-9-7 17:34:29
 Data Processed : 2016-9-7 18:10:16



<Chromatogram>

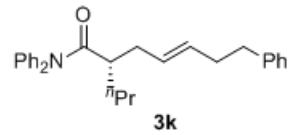


PeakTable

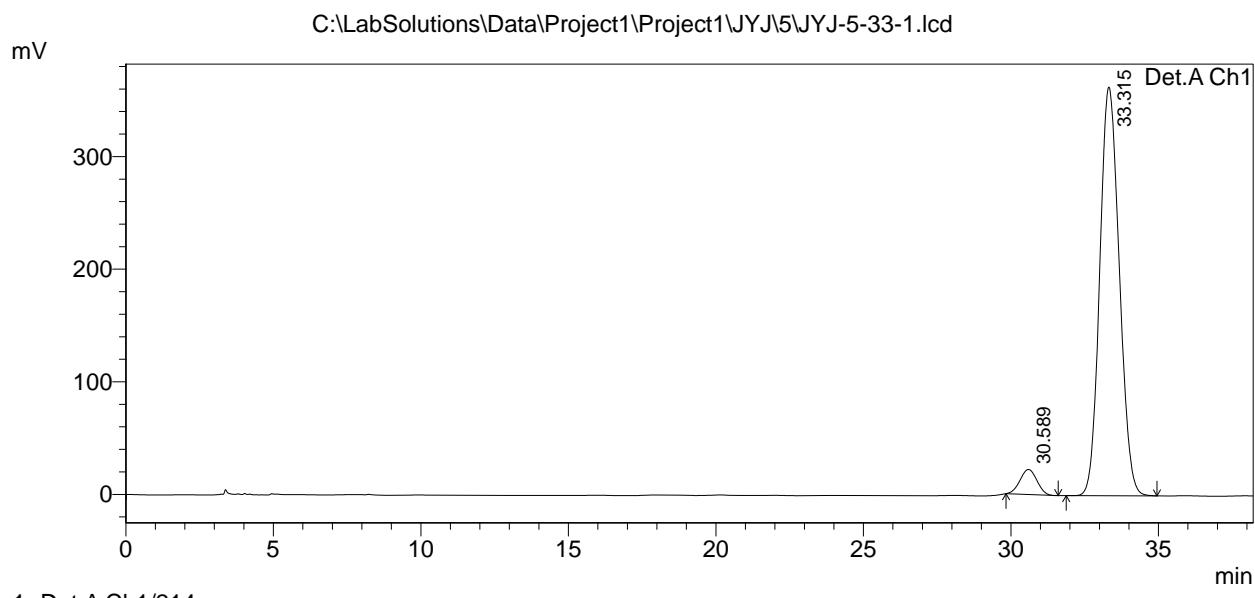
| Detector A Ch1 214nm | | | | | |
|----------------------|-----------|----------|---------|---------|----------|
| Peak# | Ret. Time | Area | Height | Area % | Height % |
| 1 | 29.985 | 27837361 | 642449 | 50.148 | 52.312 |
| 2 | 32.842 | 27673266 | 585658 | 49.852 | 47.688 |
| Total | | 55510626 | 1228107 | 100.000 | 100.000 |

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : JYJ-5-33-1
 Sample ID : IC,99/1,1,214
 Vial # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-5-33-1.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-9-7 16:53:01
 Data Processed : 2016-9-7 17:32:40



<Chromatogram>

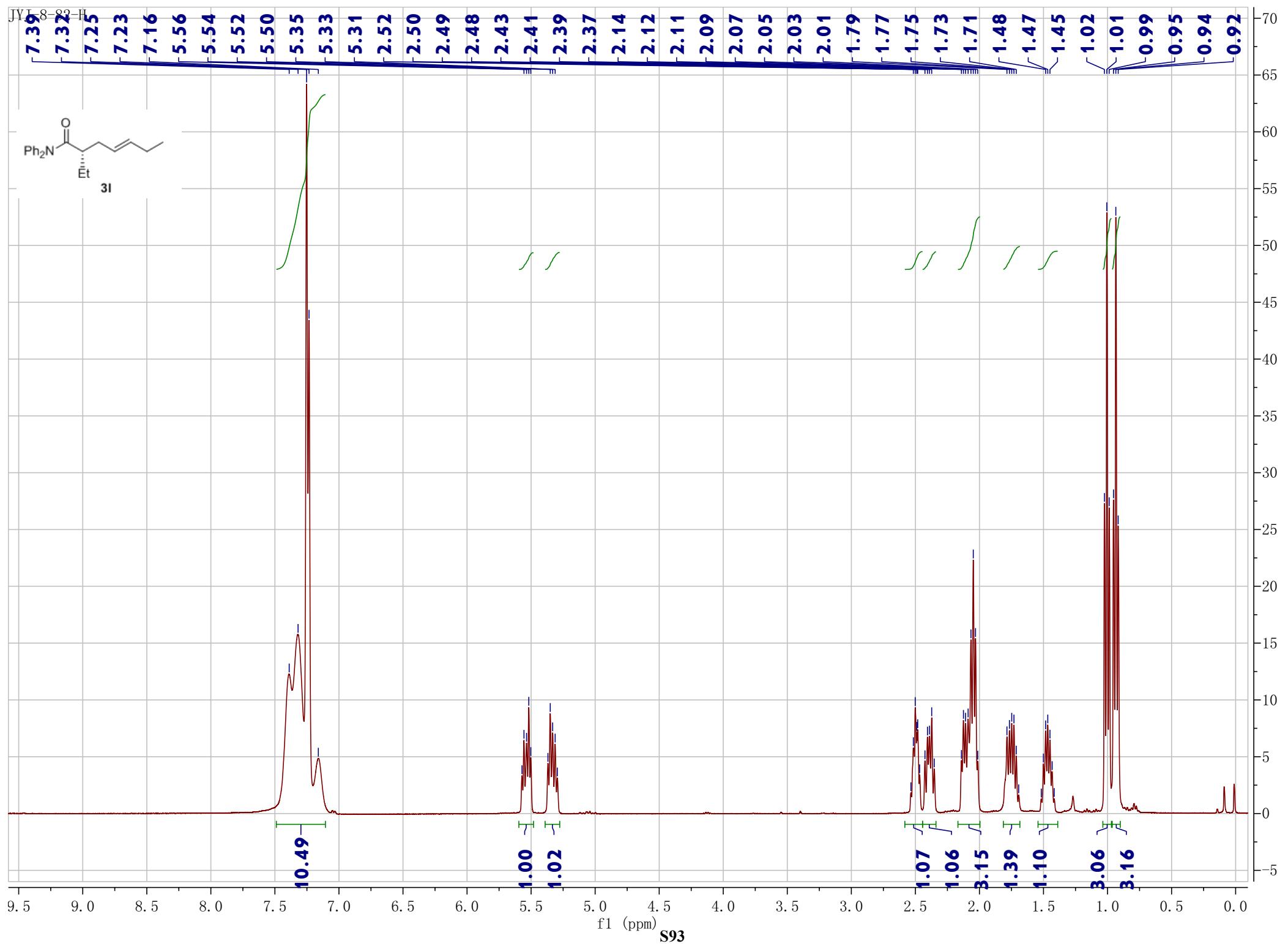


1 Det.A Ch1/214nm

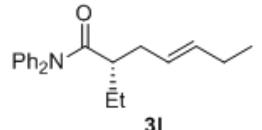
PeakTable

Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 30.589 | 885298 | 22249 | 5.042 | 5.777 |
| 2 | 33.315 | 16671612 | 362855 | 94.958 | 94.223 |
| Total | | 17556910 | 385103 | 100.000 | 100.000 |



JYJ-8-82-C
xcc



3l

-175.99

134.22
129.54
128.27
126.55
126.31

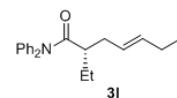
~44.71
~36.38
~26.09
~25.61

13.77
12.10

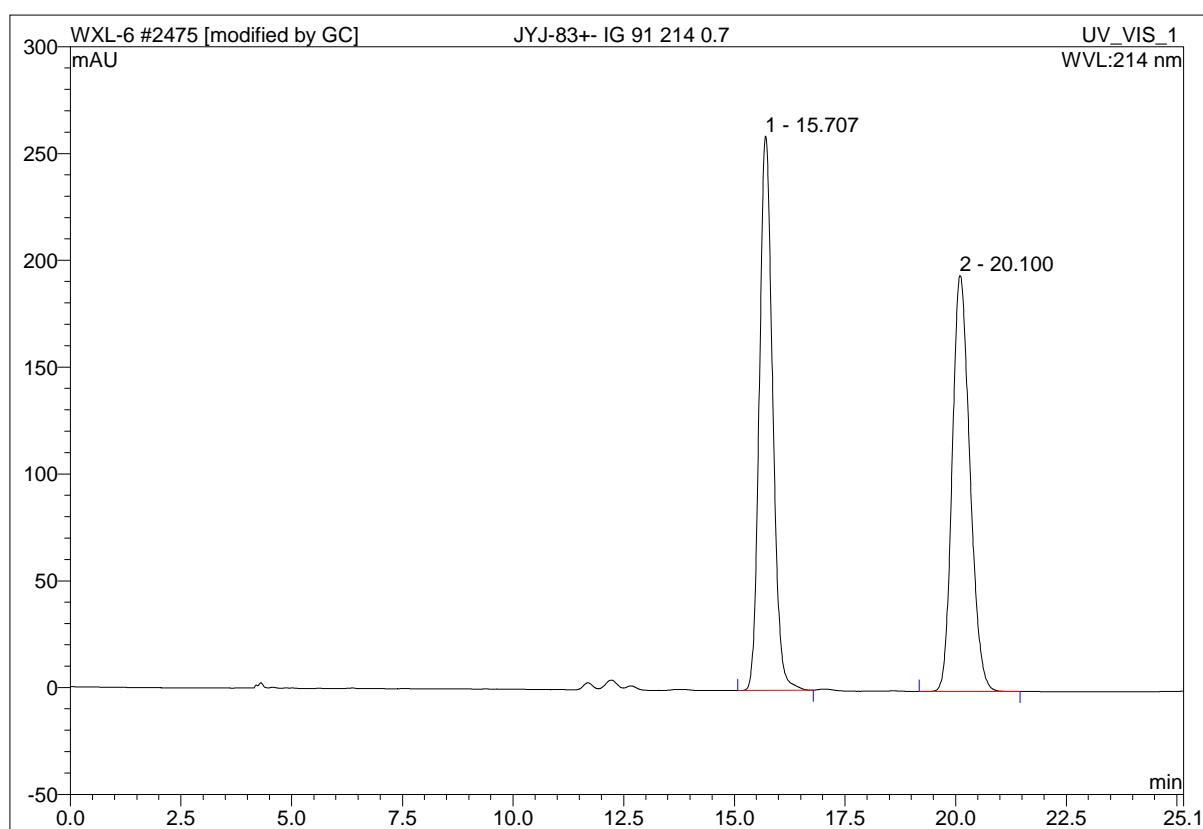
230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm) **S94**

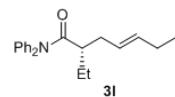
400
350
300
250
200
150
100
50
0
-50

2475 JYJ-83+- IG 91 214 0.7

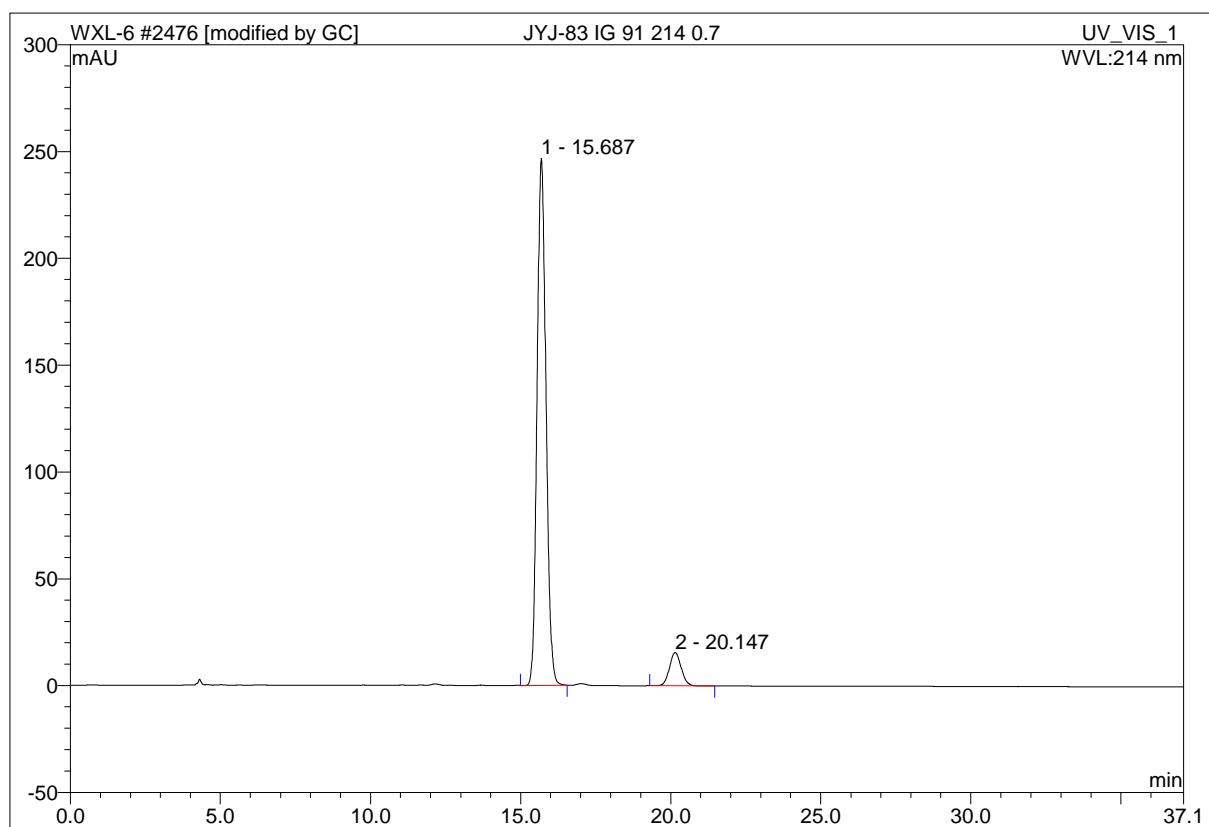
| | | | |
|-------------------------|-------------------------------|--------------------------|-----------------|
| Sample Name: | JYJ-83+- IG 91 214 0.7 | Injection Volume: | 2.0 |
| Vial Number: | RC1 | Channel: | UV_VIS_1 |
| Sample Type: | unknown | Wavelength: | 214 |
| Control Program: | 201701-4 | Bandwidth: | n.a. |
| Quantif. Method: | 201701 | Dilution Factor: | 1.0000 |
| Recording Time: | 2017/7/14 11:04 | Sample Weight: | 1.0000 |
| Run Time (min): | 25.15 | Sample Amount: | 1.0000 |



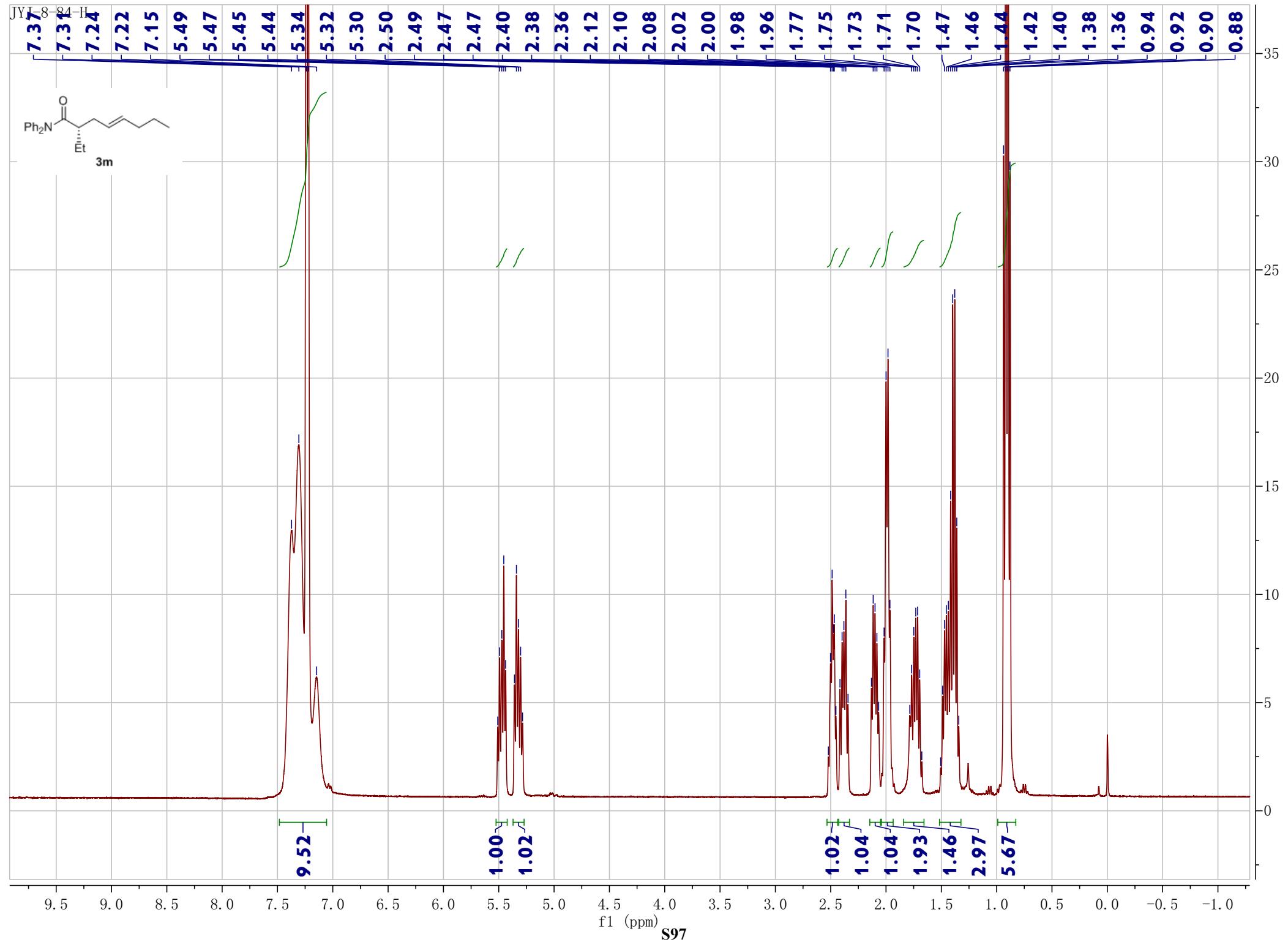
| No. | Ret.Time min | Peak Name | Height mAU | Area mAU*min | Rel.Area % | Amount | Type |
|---------------|-----------------|-----------|---------------|-----------------|---------------|--------|------|
| 1 | 15.71 | n.a. | 259.490 | 90.611 | 50.15 | n.a. | BMB* |
| 2 | 20.10 | n.a. | 194.522 | 90.082 | 49.85 | n.a. | BMB* |
| Total: | | | 454.011 | 180.693 | 100.00 | 0.000 | |

2476 JYJ-83 IG 91 214 0.7

| | | | |
|-------------------------|-----------------------------|--------------------------|-----------------|
| Sample Name: | JYJ-83 IG 91 214 0.7 | Injection Volume: | 2.0 |
| Vial Number: | RC2 | Channel: | UV_VIS_1 |
| Sample Type: | unknown | Wavelength: | 214 |
| Control Program: | 201701-4 | Bandwidth: | n.a. |
| Quantif. Method: | 201701 | Dilution Factor: | 1.0000 |
| Recording Time: | 2017/7/14 11:32 | Sample Weight: | 1.0000 |
| Run Time (min): | 37.08 | Sample Amount: | 1.0000 |

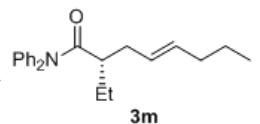


| No. | Ret.Time min | Peak Name | Height mAU | Area mAU*min | Rel.Area % | Amount | Type |
|---------------|-----------------|-----------|---------------|-----------------|---------------|--------|------|
| 1 | 15.69 | n.a. | 246.675 | 85.171 | 92.41 | n.a. | BMB* |
| 2 | 20.15 | n.a. | 15.478 | 6.993 | 7.59 | n.a. | BMB* |
| Total: | | | 262.153 | 92.164 | 100.00 | 0.000 | |



JYJ-8-84-C

xcc



3m

-175.97**143.05**
132.56
129.26
128.76
127.45
126.56**-44.69**
36.37
-34.76
26.06
-22.60
13.79
12.08

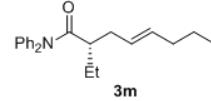
230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm) S98

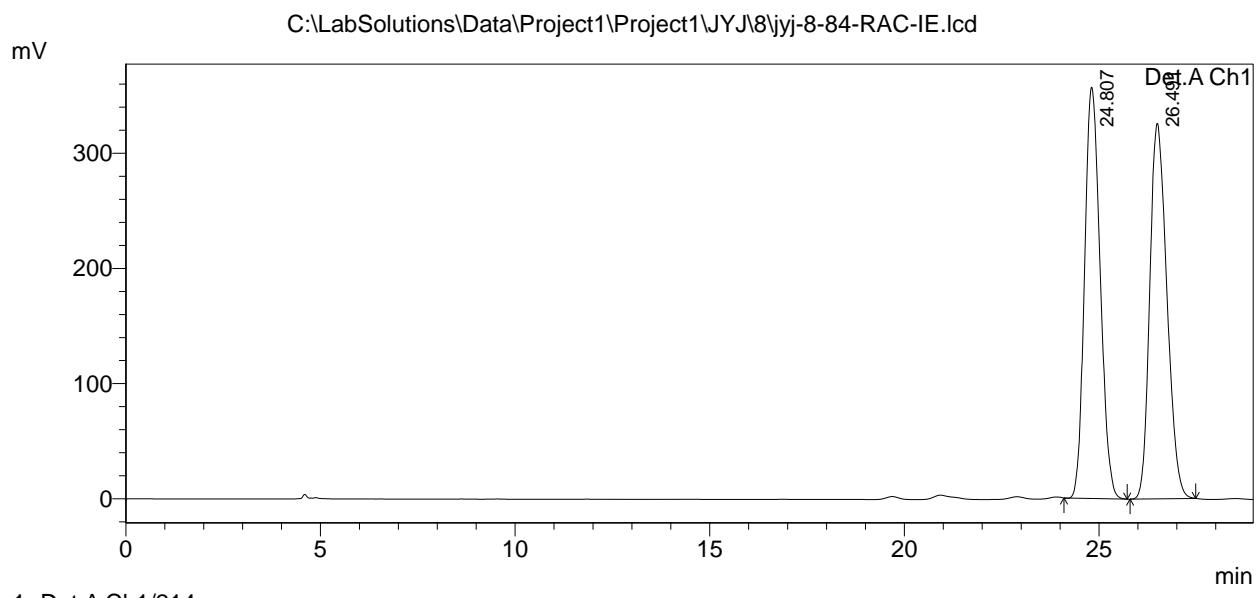
700
650
600
550
500
450
400
350
300
250
200
150
100
50
0
-50
-100

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : jyj-8-84-RAC-IE
 Sample ID : IE,95/5,0.7,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : jyj-8-84-RAC-IE.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2017-7-14 11:21:08
 Data Processed : 2017-7-14 12:20:30



<Chromatogram>



1 Det.A Ch1/214nm

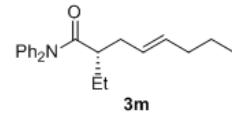
PeakTable

| Detector A Ch1 214nm | | | | | |
|----------------------|-----------|----------|--------|---------|----------|
| Peak# | Ret. Time | Area | Height | Area % | Height % |
| 1 | 24.807 | 9947744 | 357162 | 49.903 | 52.276 |
| 2 | 26.491 | 9986598 | 326064 | 50.097 | 47.724 |
| Total | | 19934342 | 683226 | 100.000 | 100.000 |

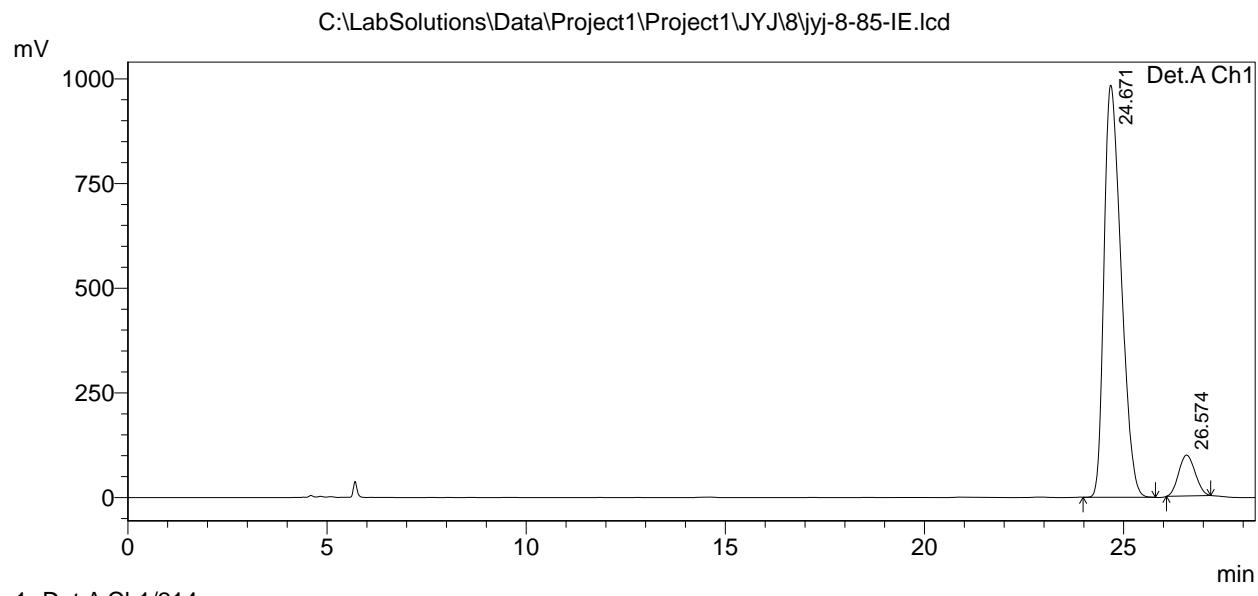
==== Shimadzu LCsolution Analysis Report ====

C:\LabSolutions\Data\Project1\Project1\JYJ\8\jyj-8-85-IE.lcd

Acquired by : Admin
 Sample Name : jyj-8-85-IE
 Sample ID : IE,95/5,0.7,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : jyj-8-85-IE.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2017-7-14 11:50:42
 Data Processed : 2017-7-14 13:14:20



<Chromatogram>

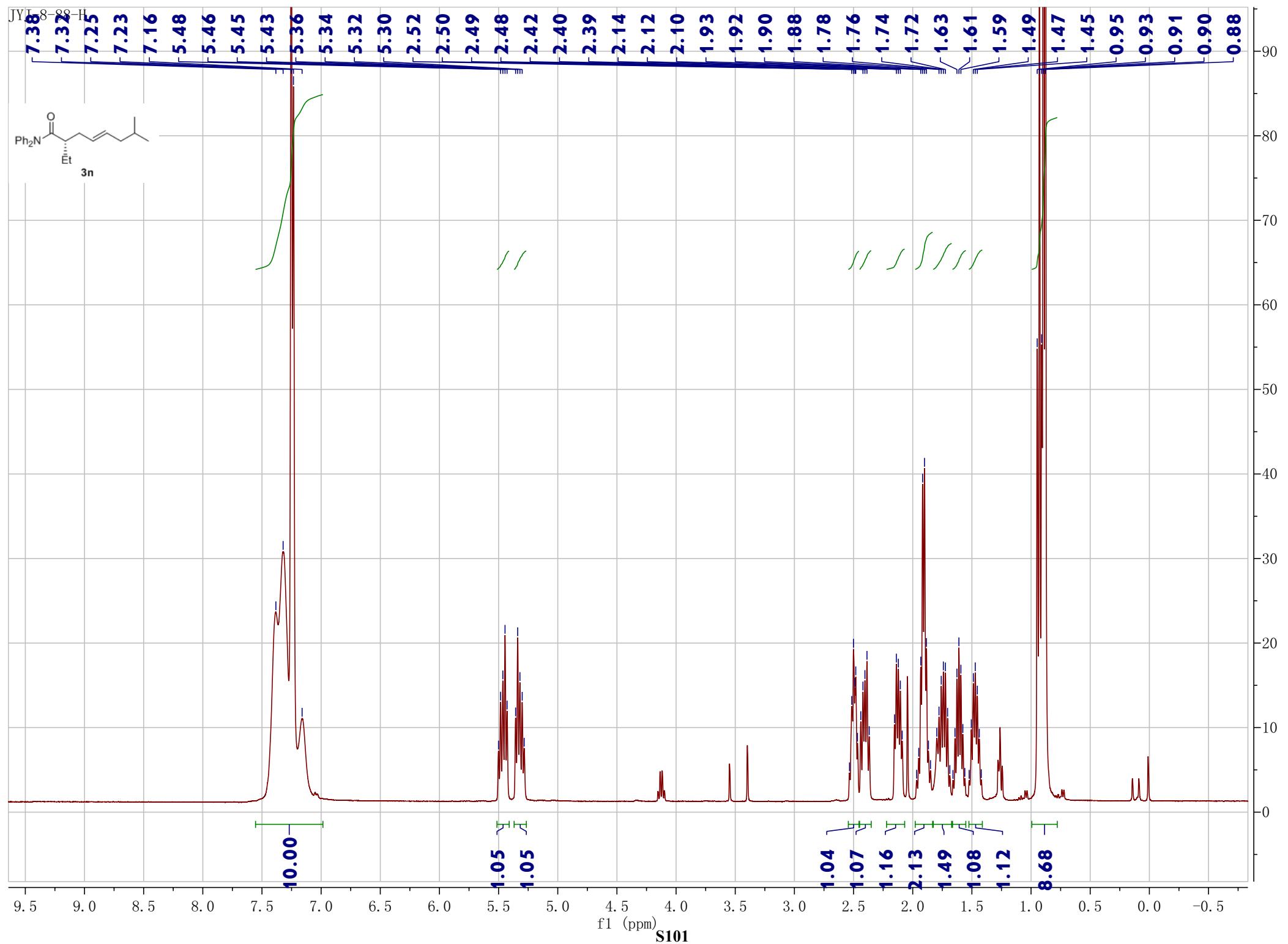


Detector A Ch1 214nm

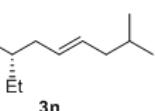
PeakTable

Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|---------|---------|----------|
| 1 | 24.671 | 29416798 | 984227 | 91.269 | 90.974 |
| 2 | 26.574 | 2813971 | 97653 | 8.731 | 9.026 |
| Total | | 32230769 | 1081880 | 100.000 | 100.000 |



JYJ-8-88-C
xcc



3n

-175.93

143.05
131.46
129.23
128.77
128.40
126.54

44.68
42.06
36.32
28.44
26.04
22.40
22.33
12.06

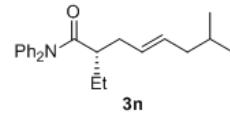
230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm) **S102**

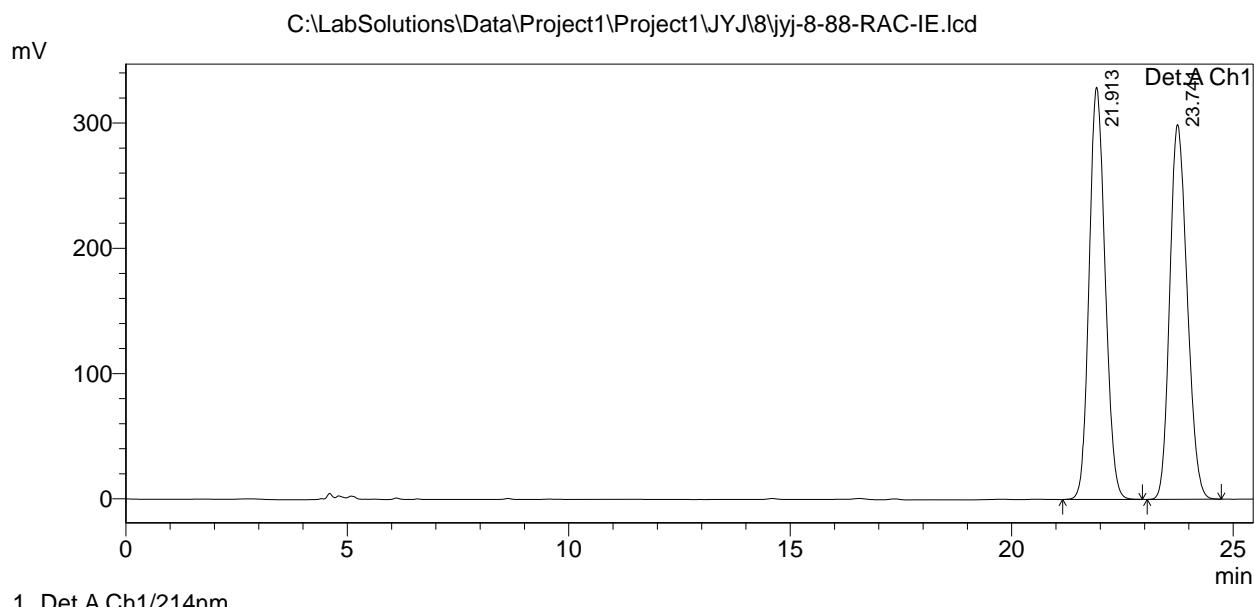
1000
900
800
700
600
500
400
300
200
100
0
-100

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : jyj-8-88-RAC-IE
 Sample ID : IE,95/5,0.7,214
 Vail # :
 Injection Volume : 2 uL
 Data File Name : jyj-8-88-RAC-IE.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2017-7-14 12:50:52
 Data Processed : 2017-7-14 13:49:40



<Chromatogram>



PeakTable

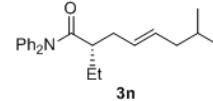
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 21.913 | 8276698 | 329138 | 50.249 | 52.387 |
| 2 | 23.741 | 8194664 | 299145 | 49.751 | 47.613 |
| Total | | 16471362 | 628284 | 100.000 | 100.000 |

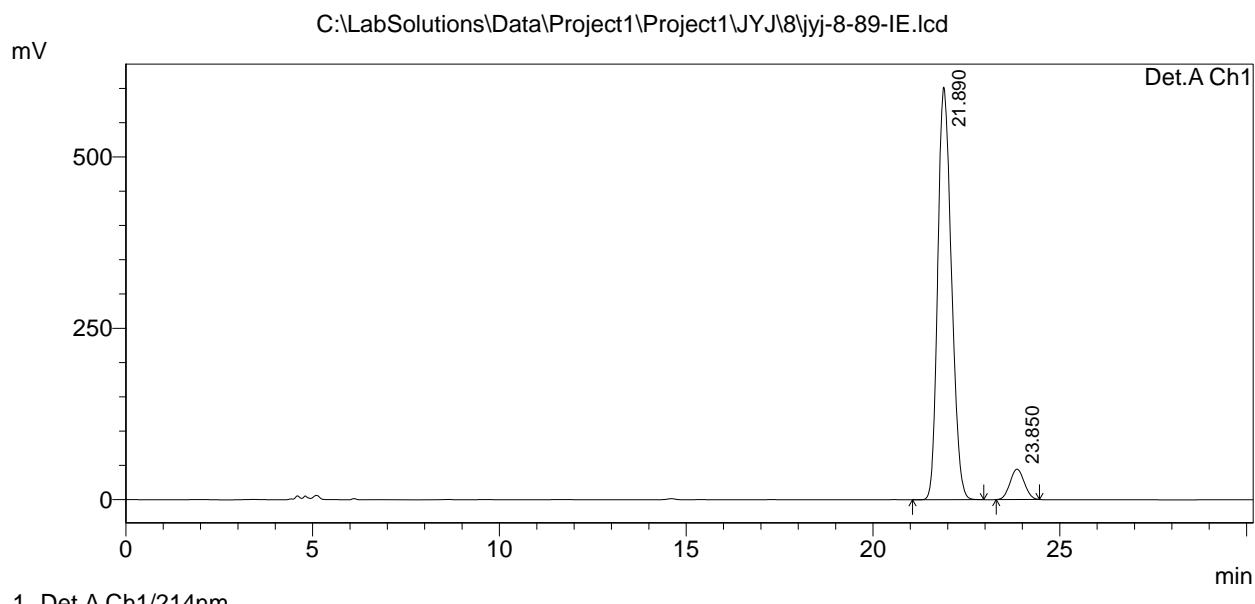
==== Shimadzu LCsolution Analysis Report ====

C:\LabSolutions\Data\Project1\Project1\JYJ\8\jyj-8-89-IE.lcd

Acquired by : Admin
 Sample Name : jyj-8-89-IE
 Sample ID : IE,95/5,0.7,214
 Vail # :
 Injection Volume : 2 uL
 Data File Name : jyj-8-89-IE.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2017-7-14 13:17:31
 Data Processed : 2017-7-14 13:48:40



<Chromatogram>

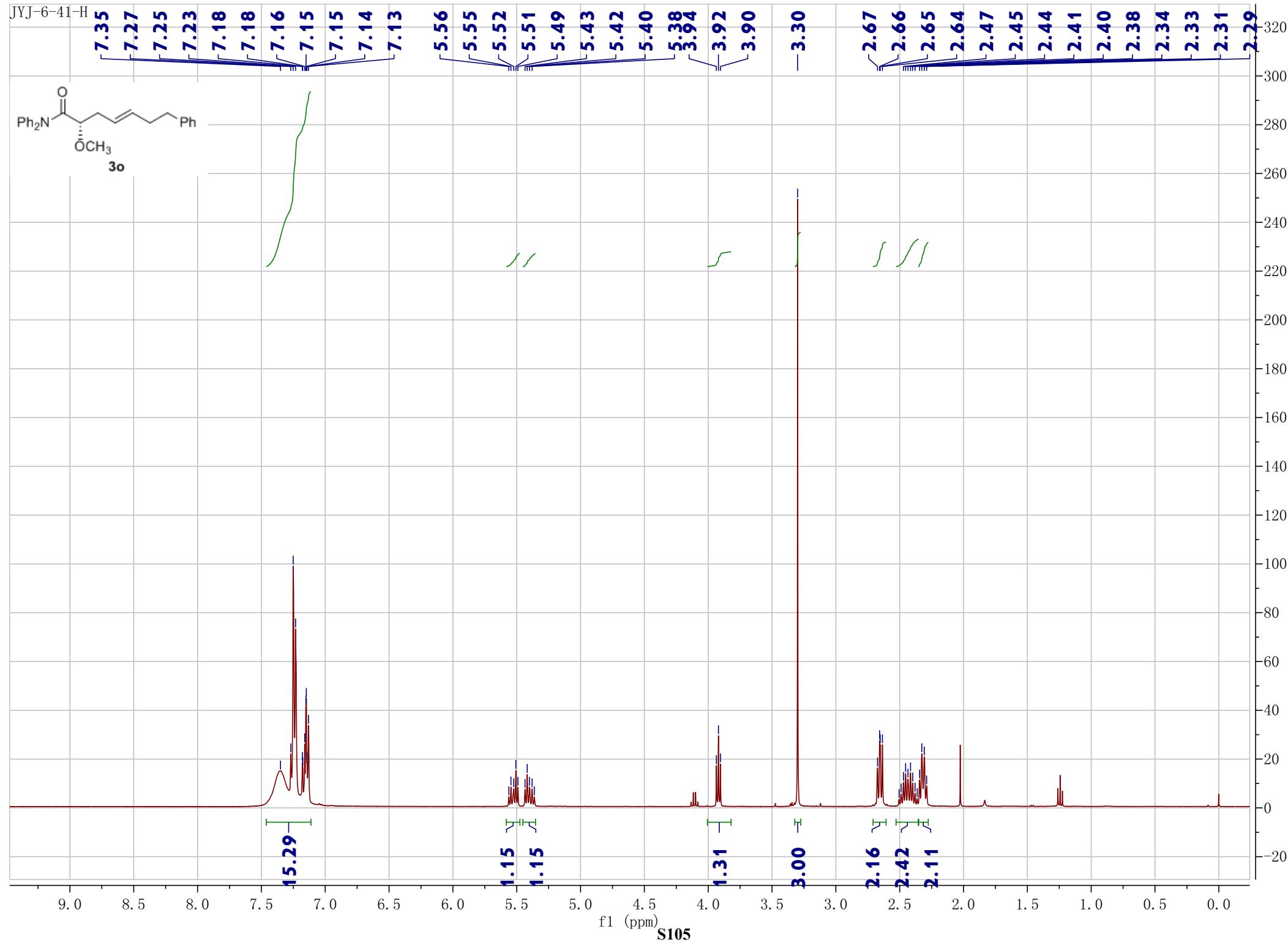


PeakTable

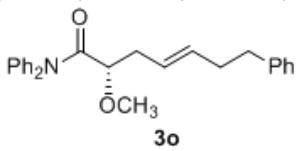
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 21.890 | 15393868 | 602031 | 92.876 | 93.141 |
| 2 | 23.850 | 1180796 | 44331 | 7.124 | 6.859 |
| Total | | 16574664 | 646362 | 100.000 | 100.000 |

JYJ-6-41-H



JYJ-6-41-C

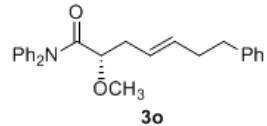
-171.52**141.86**
132.95
128.96
128.44
128.30
125.81
125.36**-78.31****-57.06****35.78**
35.65
34.47

190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

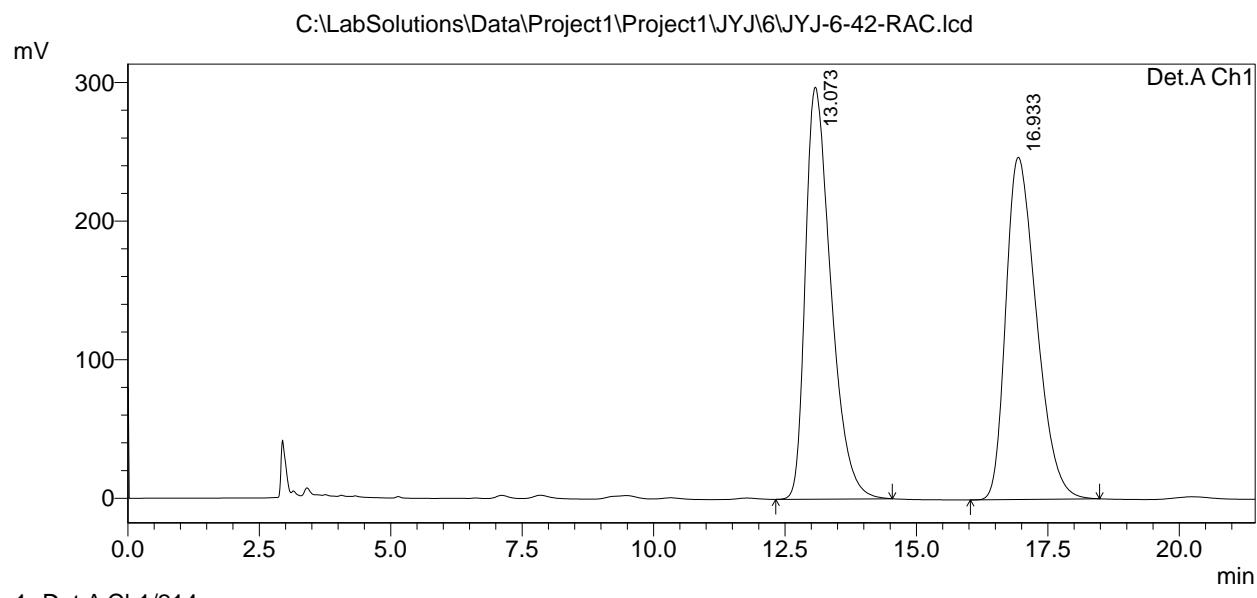
f1 (ppm) **S106**260
240
220
200
180
160
140
120
100
80
60
40
20
0
-20

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : JYJ-6-42-RAC
 Sample ID : OD-H,99/1,1,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-6-42-RAC.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-12-10 16:25:58
 Data Processed : 2016-12-10 17:09:00



<Chromatogram>



PeakTable

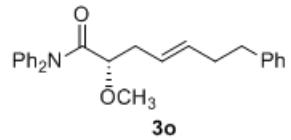
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 13.073 | 10036249 | 297346 | 50.048 | 54.625 |
| 2 | 16.933 | 10017083 | 246999 | 49.952 | 45.375 |
| Total | | 20053332 | 544345 | 100.000 | 100.000 |

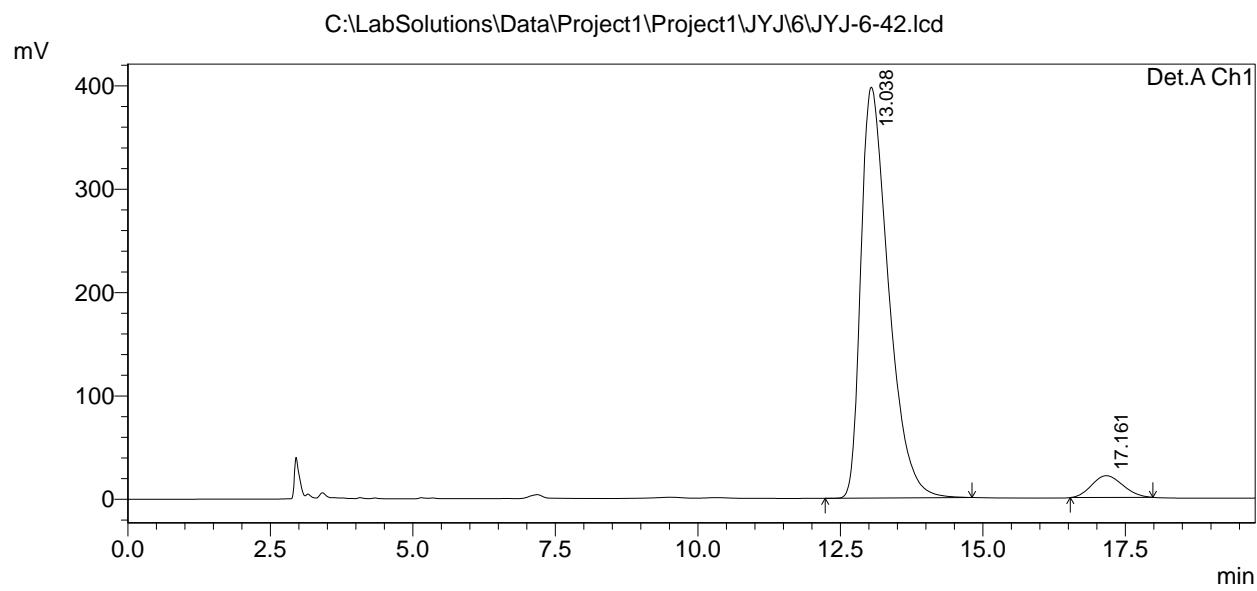
==== Shimadzu LCsolution Analysis Report ====

C:\LabSolutions\Data\Project1\Project1\JYJ\6\JYJ-6-42.lcd

Acquired by : Admin
 Sample Name : JYJ-6-42
 Sample ID : OD-H,90/10,1,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-6-42.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2016-12-10 16:49:03
 Data Processed : 2016-12-10 17:11:26



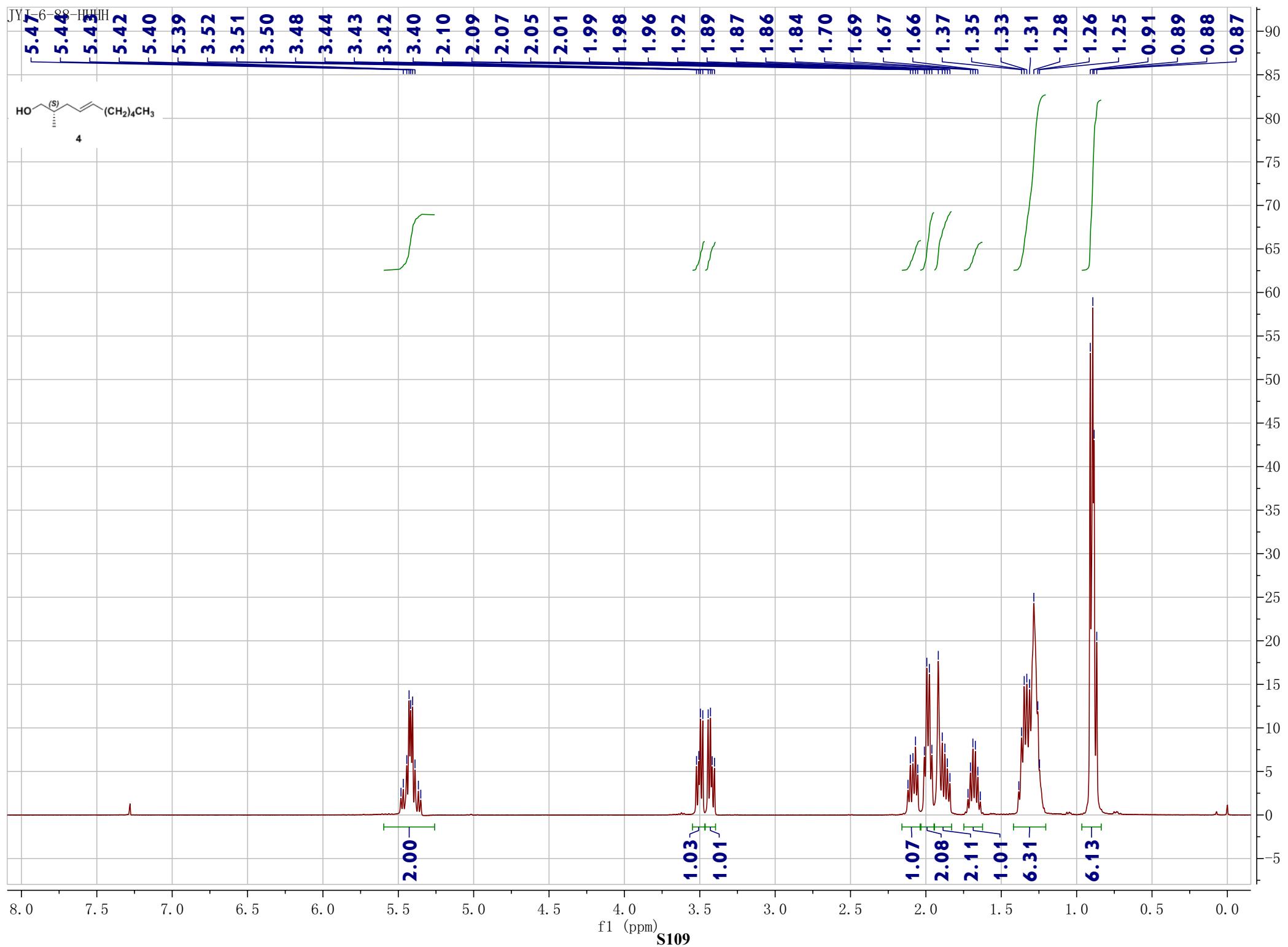
<Chromatogram>



PeakTable

Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 13.038 | 13422479 | 397594 | 94.367 | 94.958 |
| 2 | 17.161 | 801170 | 21111 | 5.633 | 5.042 |
| Total | | 14223649 | 418705 | 100.000 | 100.000 |



JYJ-6-88-CCC



4

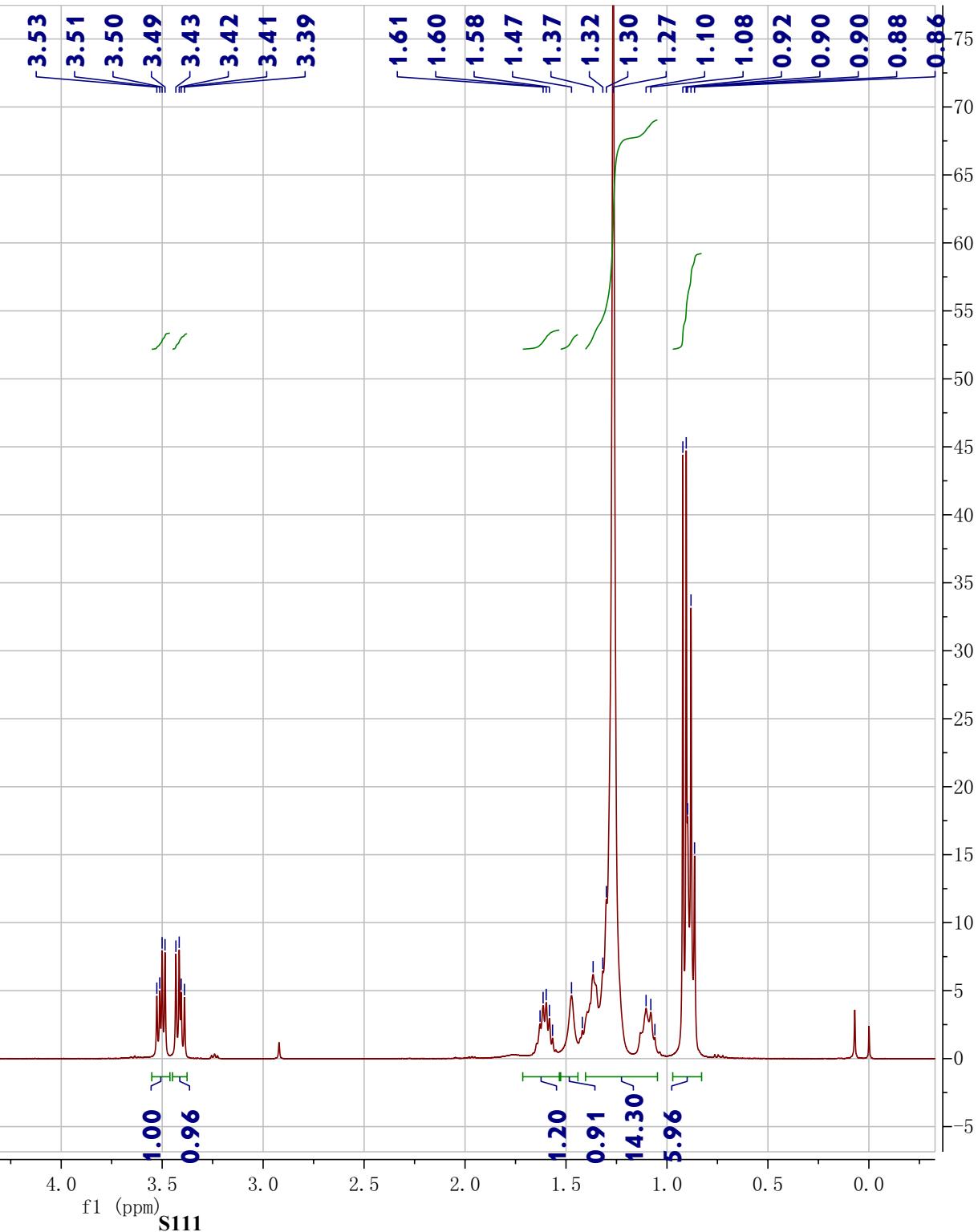
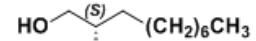
-132.36
-127.95**-67.91****36.56**
35.96
32.56
31.37
29.24
22.51
16.41
14.06

160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

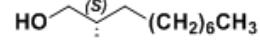
f1 (ppm)

S110

JYJ-6-95-HHH



JYJ-6-95-CCC



5

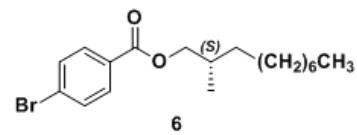
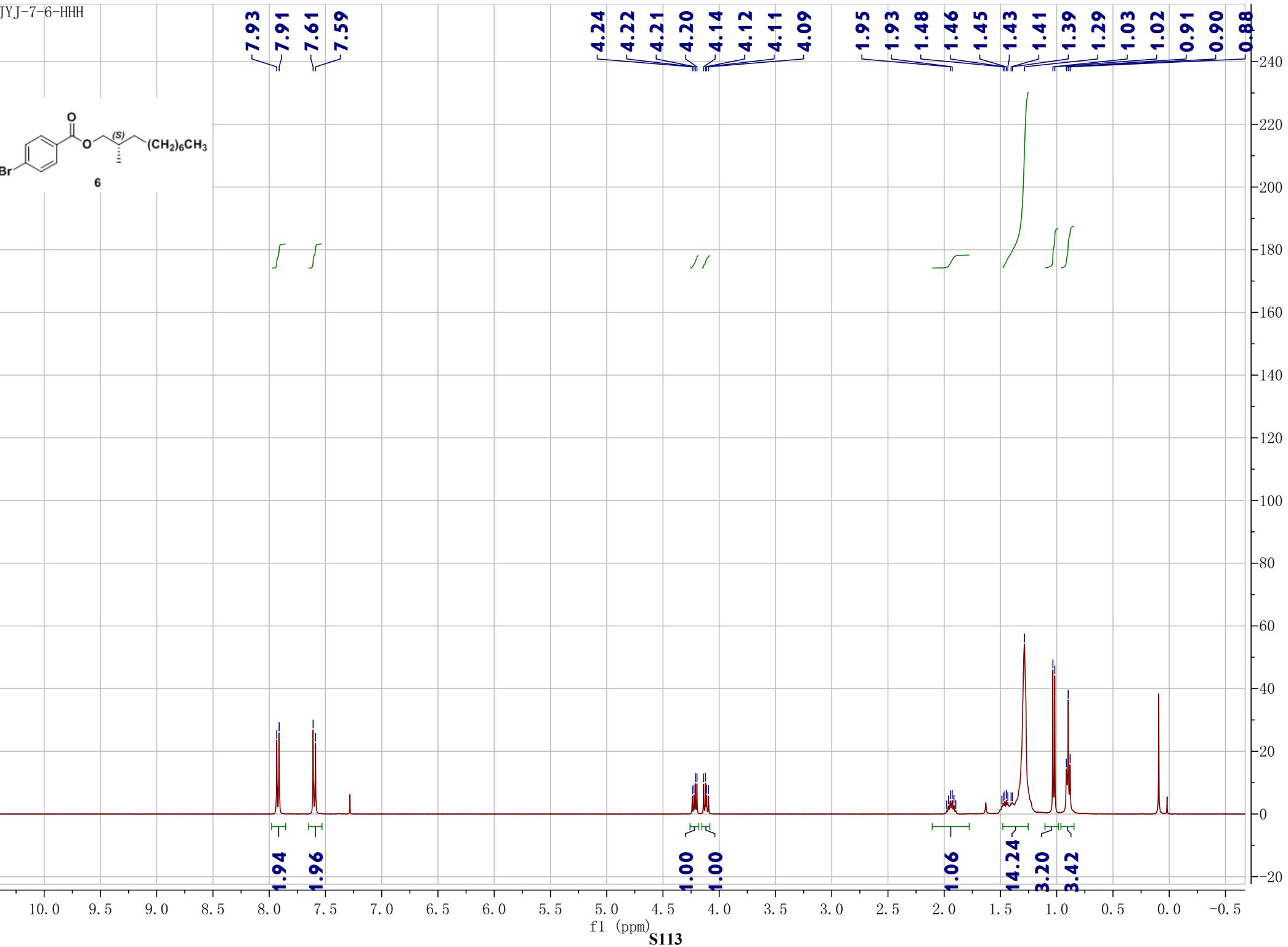
-68.40

35.74
33.13
31.90
29.94
29.61
29.33
26.98
22.68
16.58
14.12

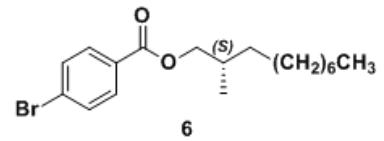
200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm) S112

JYJ-7-6-HHH

**6****S113**

JYJ-7-6-CCC

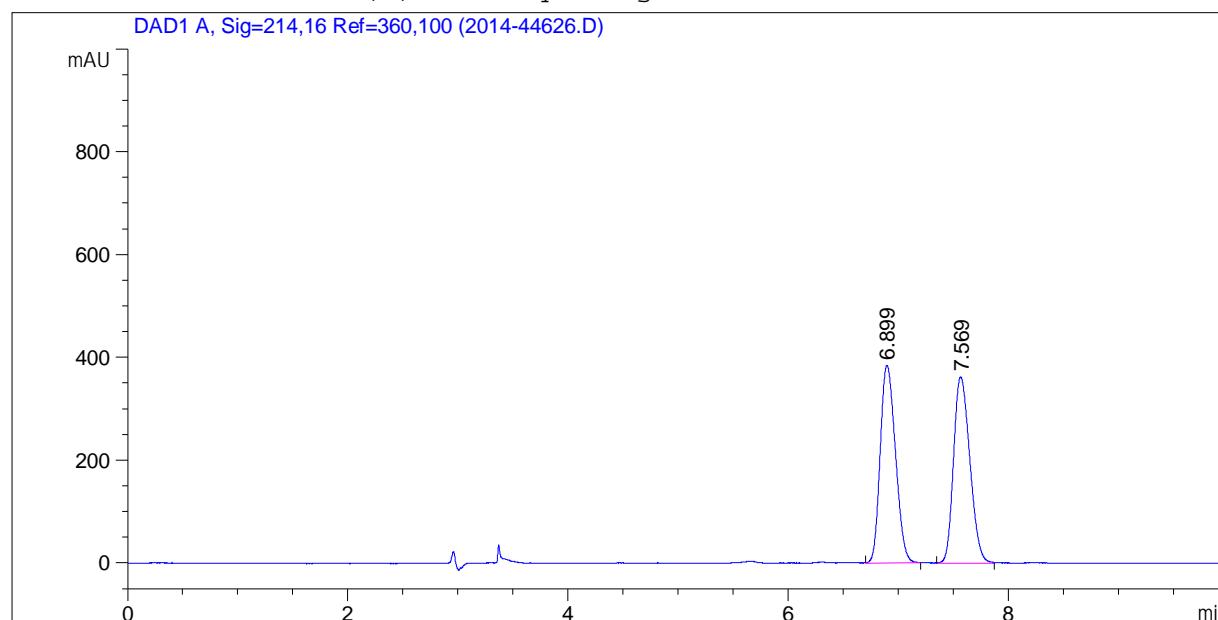
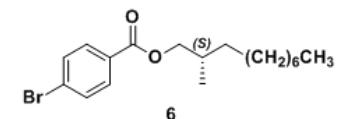
-165.93131.66
131.05
129.40
127.88**-70.14**33.42
32.65
31.89
29.82
29.55
29.30
26.84
22.67
17.03
14.12

230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm)

S114

=====
 Acq. Operator : 系统
 Sample Operator : 系统
 Acq. Instrument : SFC Location : Vial 2
 Injection Date : 07/03/2017 15:59:36 Inj Volume : 3.000 μ l
 Acq. Method : C:\CHEM32\1\METHODS\DEF_LC-TEST-2016.M
 Last changed : 07/03/2017 15:51:55 by 系统
 (modified after loading)
 Analysis Method : C:\CHEM32\1\METHODS\DEF_LC-TEST-2016.M
 Last changed : 13/03/2017 16:49:53 by 系统
 (modified after loading)
 Additional Info : Peak(s) manually integrated



=====
 Area Percent Report
 =====

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=214,16 Ref=360,100

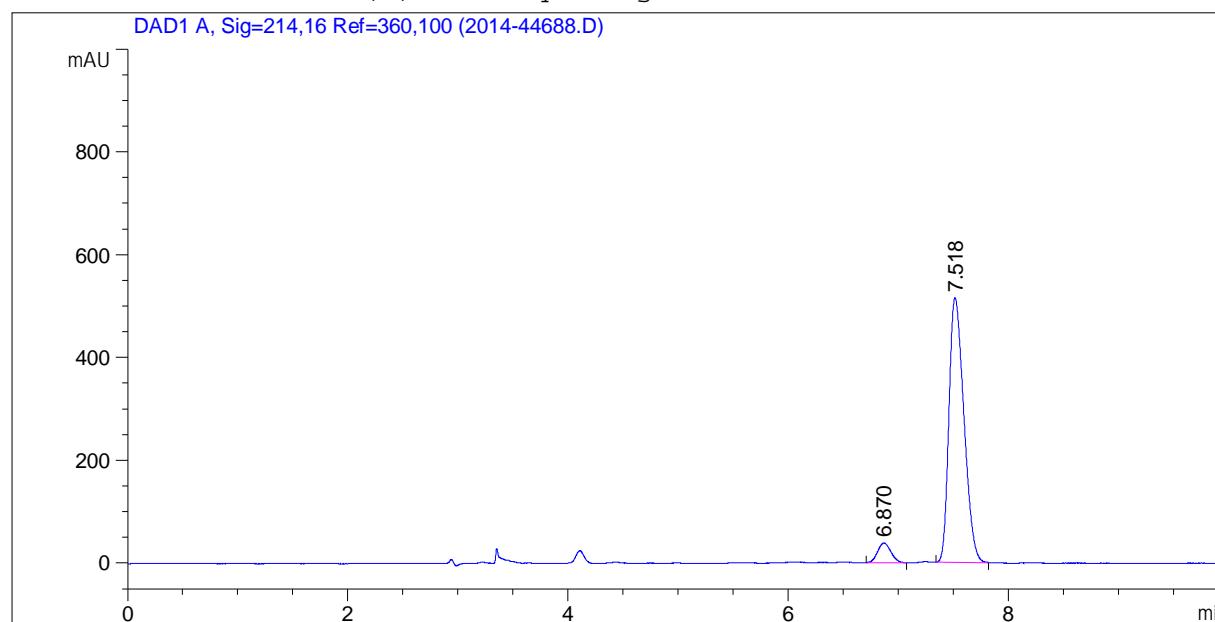
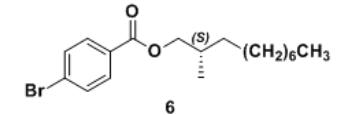
| Peak # | RetTime [min] | Type | Width [min] | Area [mAU*s] | Height [mAU] | Area % |
|--------|---------------|------|-------------|--------------|--------------|---------|
| 1 | 6.899 | VV | 0.1528 | 3700.52661 | 384.53714 | 49.9081 |
| 2 | 7.569 | VV | 0.1593 | 3714.14746 | 361.81332 | 50.0919 |

Totals : 7414.67407 746.35046

=====
 *** End of Report ***
 =====

Sample Name: JYJ-7-22

```
=====
Acq. Operator   : 系统
Sample Operator : 系统
Acq. Instrument : SFC
Injection Date   : 16/03/2017 14:31:05
Location       : Vial 11
Inj Volume    : 1.000 µl
Acq. Method     : C:\CHEM32\1\METHODS\DEF_LC-TEST-2016.M
Last changed    : 16/03/2017 14:19:22 by 系统
                  (modified after loading)
Analysis Method : C:\CHEM32\1\METHODS\DEF_LC-TEST-2016.M
Last changed    : 27/02/2017 16:26:37 by 系统
Additional Info : Peak(s) manually integrated
```



```
=====
Area Percent Report
=====
```

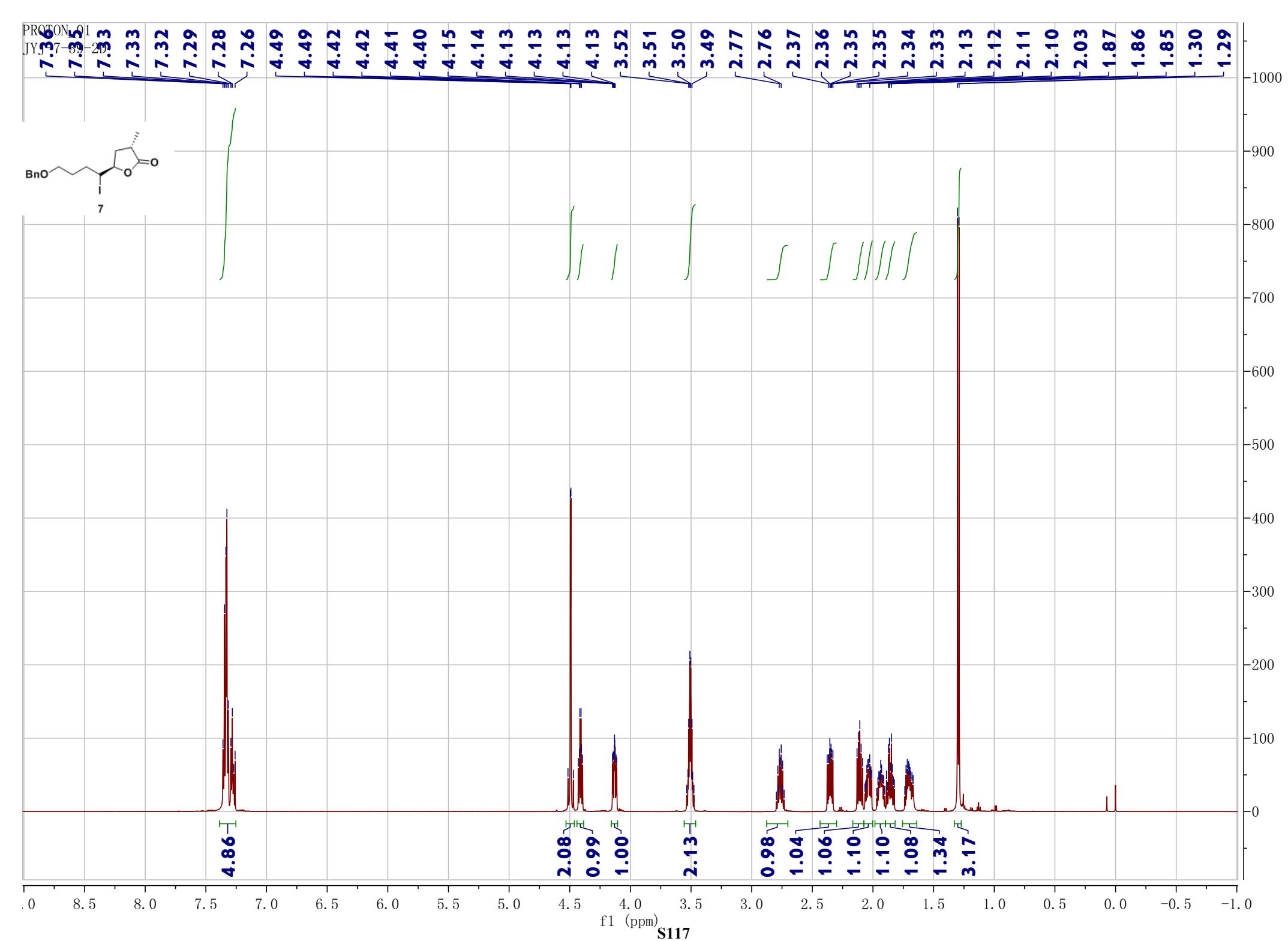
```
Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Do not use Multiplier & Dilution Factor with ISTDs
```

Signal 1: DAD1 A, Sig=214,16 Ref=360,100

| Peak # | RetTime [min] | Type | Width [min] | Area [mAU*s] | Height [mAU] | Area % |
|--------|---------------|------|-------------|--------------|--------------|---------|
| 1 | 6.870 | VV | 0.1209 | 329.39648 | 38.67027 | 6.3413 |
| 2 | 7.518 | VV | 0.1426 | 4865.03174 | 515.31256 | 93.6587 |

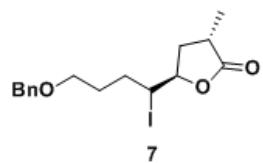
Totals : 5194.42822 553.98283

===== *** End of Report ***



CARBON_01
JYJ-7-39-2D

-179.29



138.29
128.39
127.65
127.61

-80.19
-72.96
-69.03

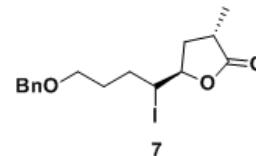
39.07
35.80
34.48
32.76
29.41
-16.41

210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

f1 (ppm)

S118

JYJ-7-39-2D



Sample Name:
JYJ-7-39-2D

Data Collected on:
OMC-NMR600-vnmrs600

Archive directory:
/home/omc/vnmr600/data
Sample directory:
JYJ-7-39-2D_20170320_01
FidFile: NOESY_01

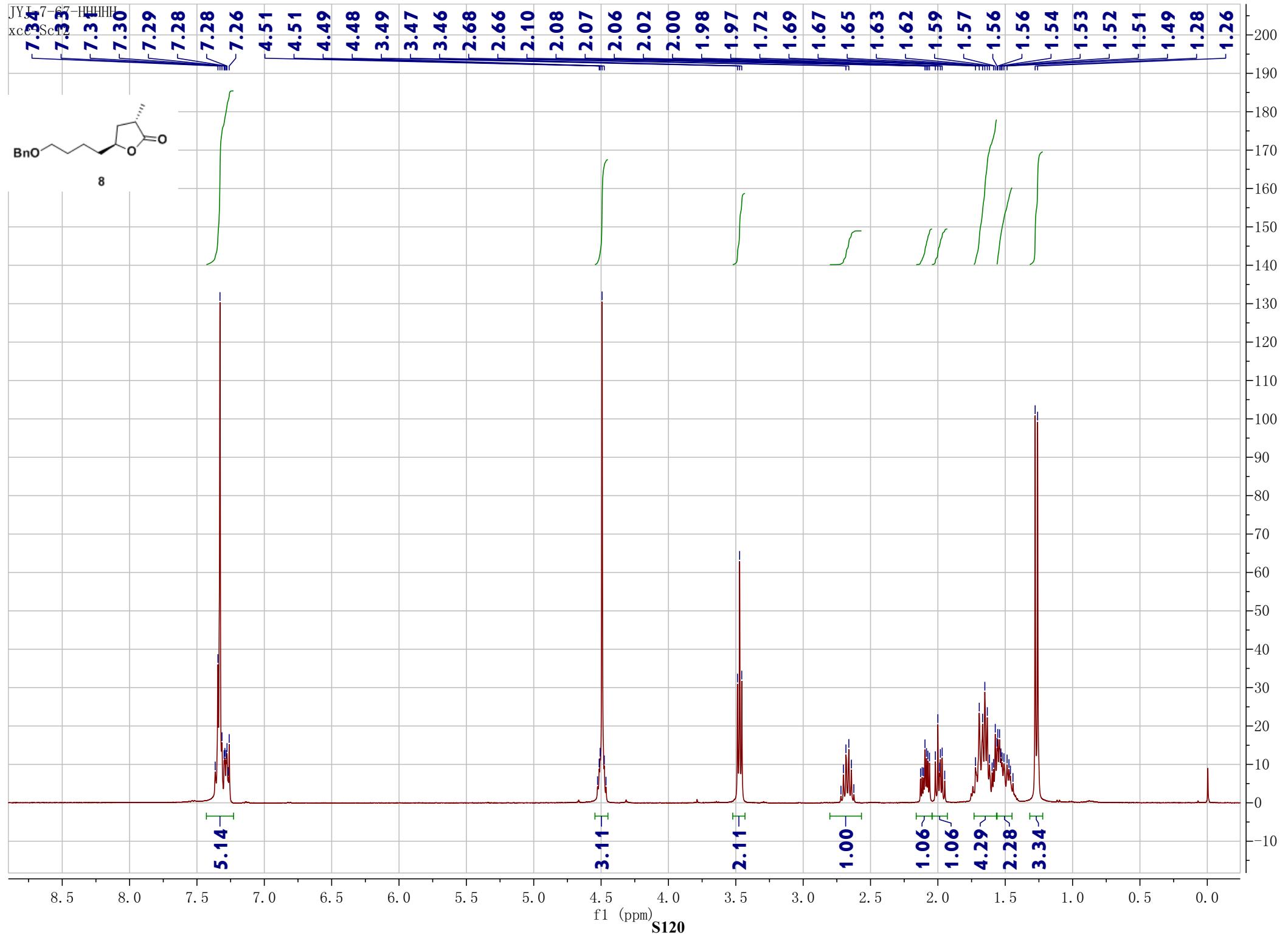
Pulse Sequence: NOESY
Solvent: cdcl₃
Data collected on: Mar 20 2017

Temp. 25.0 C / 298.1 K
Operator: omc

Relax. delay 1.500 sec
Acq. time 0.382 sec
Width 5787.0 Hz
2D Width 5787.0 Hz
8 repetitions
2 x 128 increments
OBSERVE H1, 599.7751422 MHz
DATA PROCESSING
Line broadening 3.0 Hz
Gauss apodization 0.056 sec
F1 DATA PROCESSING
Gauss apodization 0.012 sec
FT size 4096 x 2048
Total time 1 hr, 20 min

Plotname: --Not assigned--

JYI-7-67-HHHH
xxc.Scpt



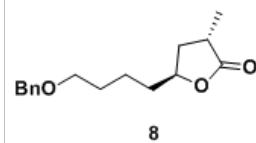
JYJ-7-67-CCCCCC
1xj-2-54-C-H

-180.09

138.44
128.37
127.65
127.57

78.33
72.96
69.93

35.39
35.20
34.00
29.41
22.20
15.90



8

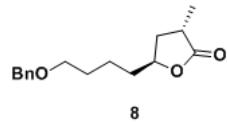
210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

f1 (ppm) **S121**

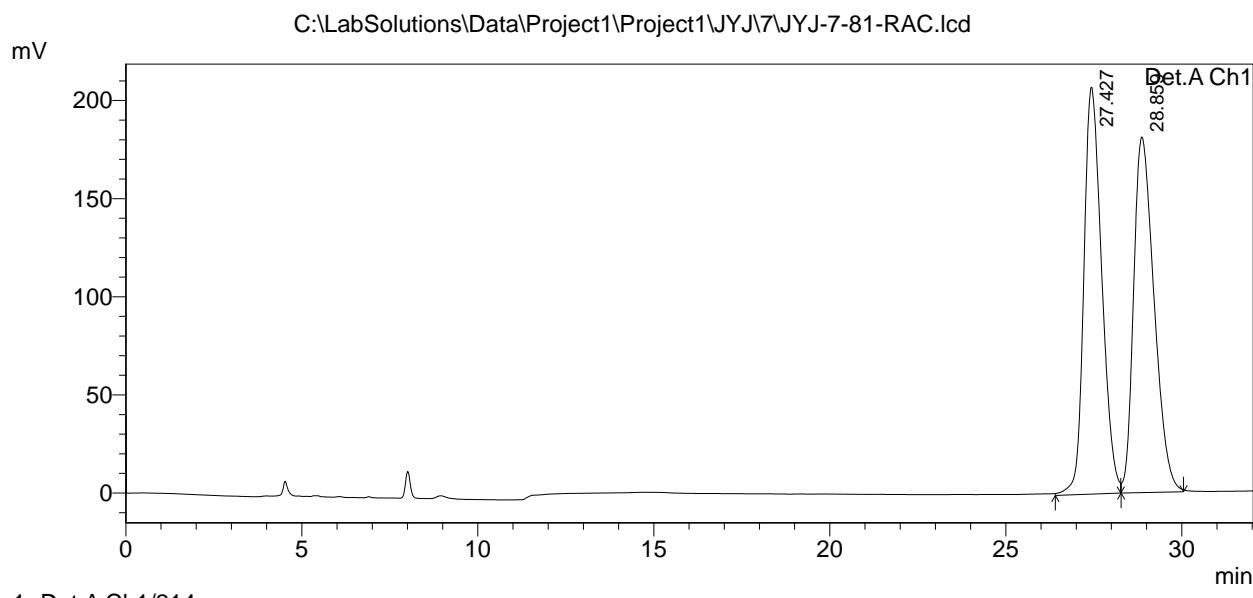
9
8
7
6
5
4
3
2
1
0
-1

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : JYJ-7-81-RAC
 Sample ID : OD-H,95/5,0.7,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-7-81-RAC.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2017-4-18 21:39:40
 Data Processed : 2017-4-18 22:51:23



<Chromatogram>



PeakTable

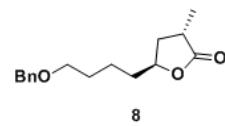
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 27.427 | 7344401 | 207373 | 50.039 | 53.380 |
| 2 | 28.859 | 7333018 | 181108 | 49.961 | 46.620 |
| Total | | 14677420 | 388481 | 100.000 | 100.000 |

==== Shimadzu LCsolution Analysis Report ====

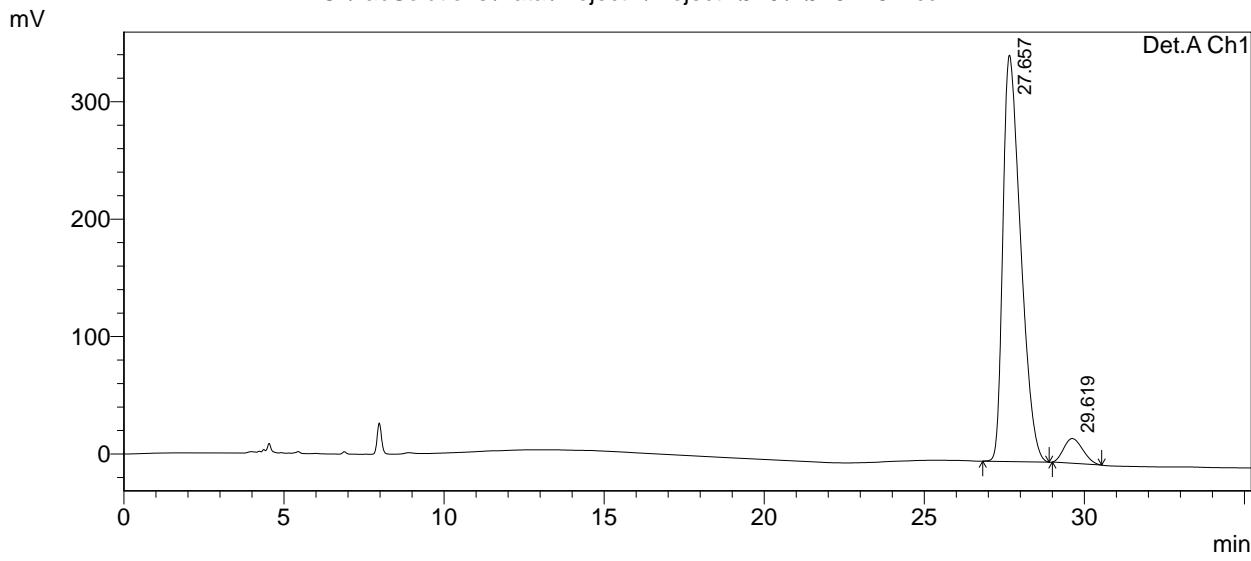
C:\LabSolutions\Data\Project1\Project1\JYJ\7\JYJ-7-82.lcd

Acquired by : Admin
 Sample Name : JYJ-7-82
 Sample ID : OD-H,95/5,0.7,214
 Vail # :
 Injection Volume : 2 μ L
 Data File Name : JYJ-7-82.lcd
 Method File Name : 123.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Data Acquired : 2017-4-18 22:15:55
 Data Processed : 2017-4-20 10:57:47



<Chromatogram>

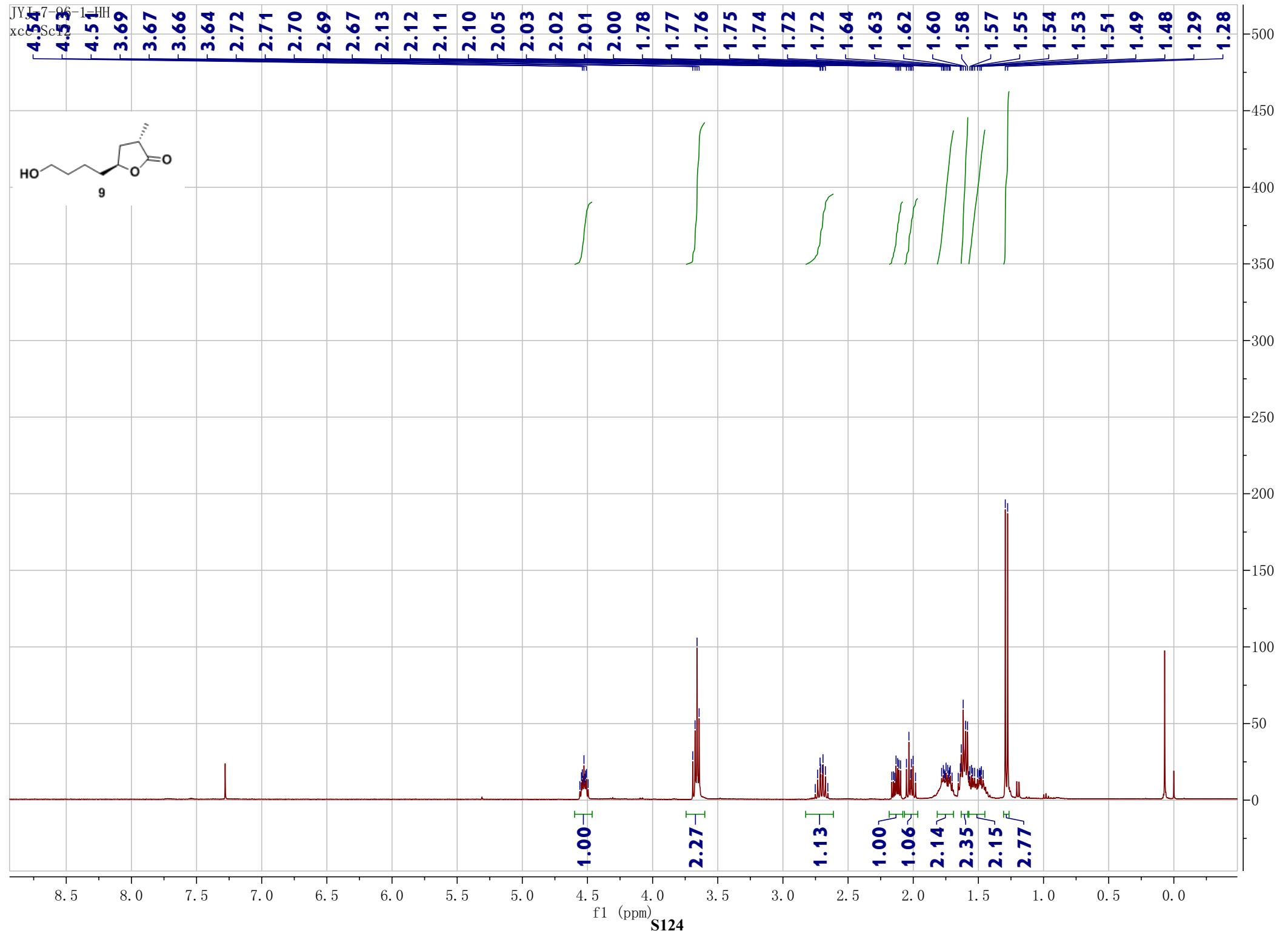
C:\LabSolutions\Data\Project1\Project1\JYJ\7\JYJ-7-82.lcd



PeakTable

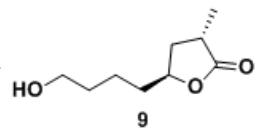
Detector A Ch1 214nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 27.657 | 13357004 | 346109 | 93.821 | 94.258 |
| 2 | 29.619 | 879748 | 21083 | 6.179 | 5.742 |
| Total | | 14236752 | 367192 | 100.000 | 100.000 |



JYJ-7-96-1-CC
1xj-2-54-C-H

-180.17

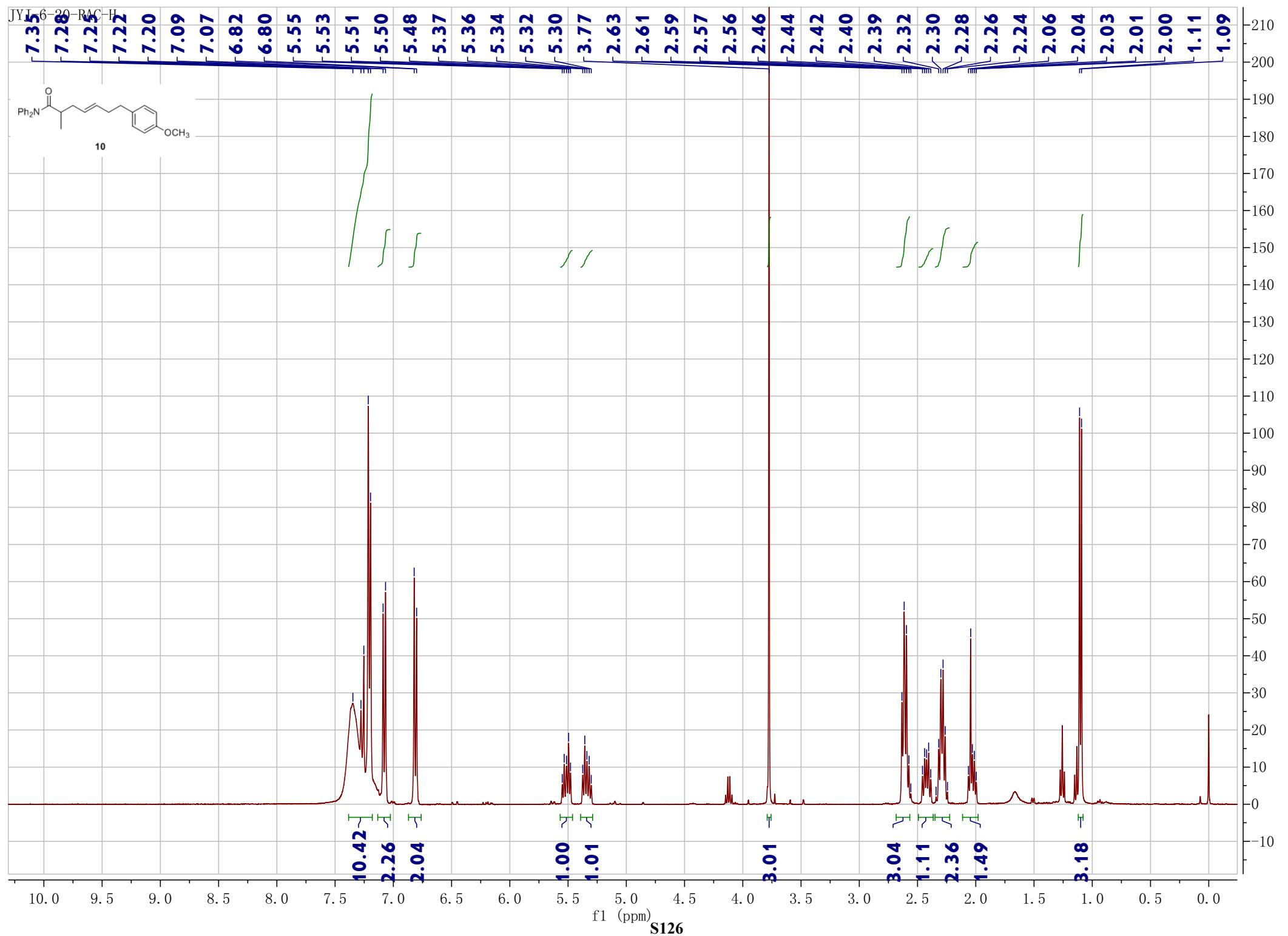


-78.37

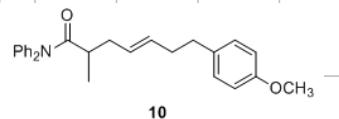
-62.48

35.39
35.17
34.02
32.23
21.73
15.89

220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 f1 (ppm) S125



JYJ-6-20-RAC-C



10

-176.60**-157.71****142.99**
134.03
131.87
129.26
127.87**-113.69****-55.25****37.80**
37.71
35.03
34.73**-17.64**

230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm) **S127**

350

300

250

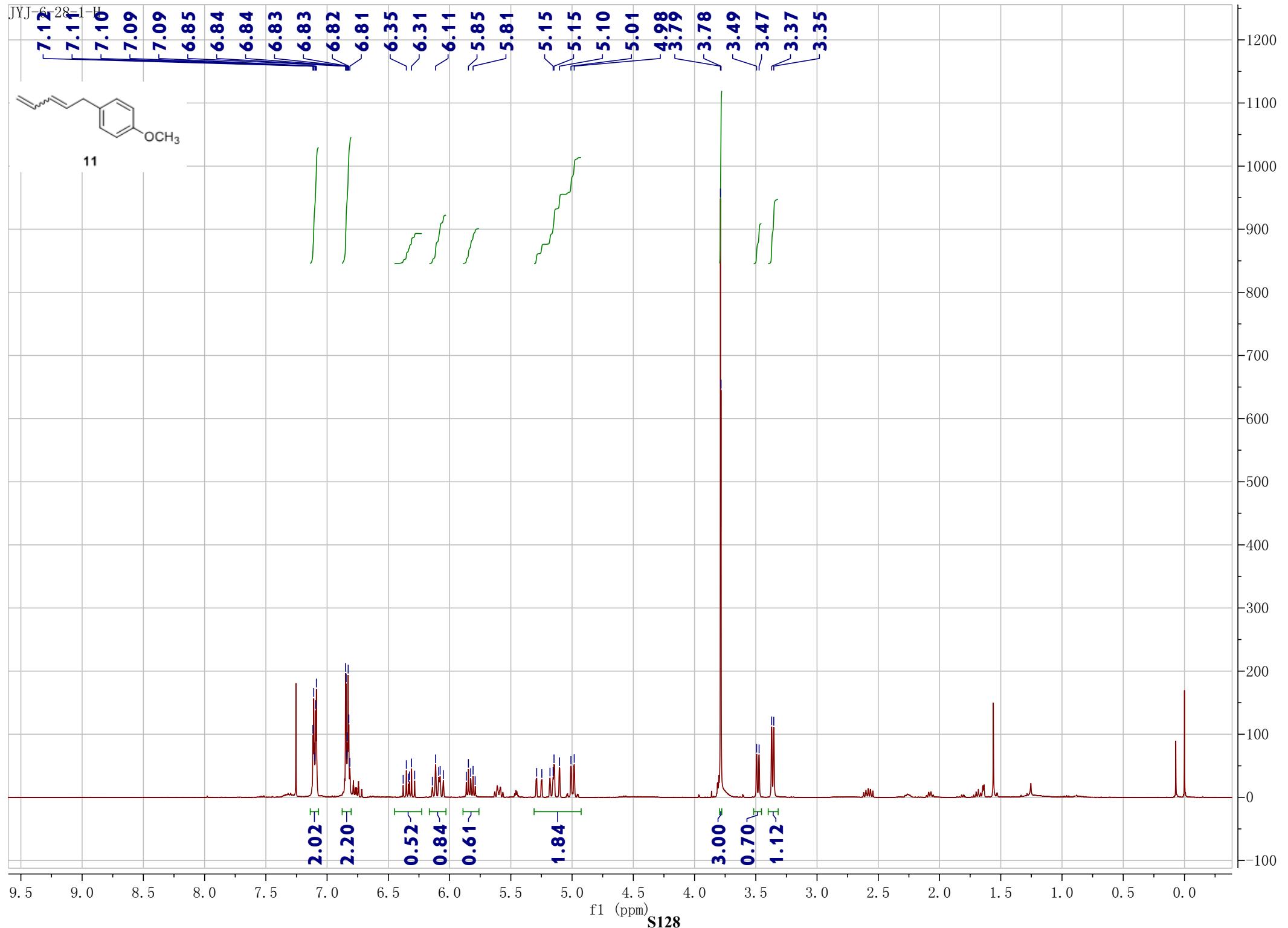
200

150

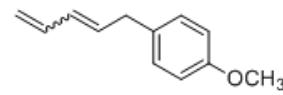
100

50

0



JYJ-6-28-1-C



11

-157.98

136.96
133.94
131.73
131.03
129.67
129.52
129.28
117.87
115.58
113.89
113.84

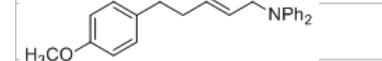
-55.27

38.01
33.00

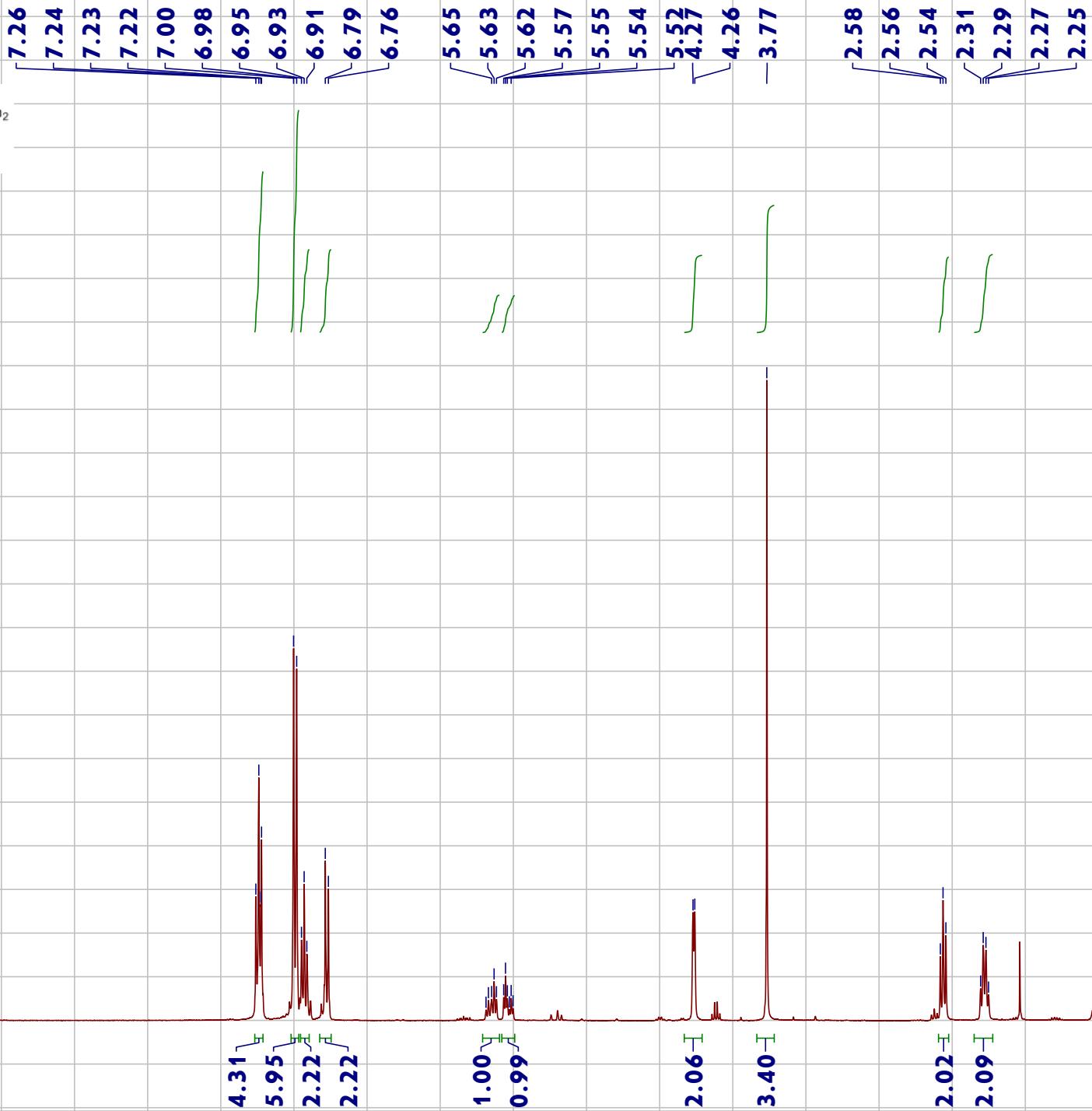
f1 (ppm)
S129

320
300
280
260
240
220
200
180
160
140
120
100
80
60
40
20
0
-20
-40

JYJ-6-29-2-HHH



12



S130

JYJ-6-29

2-C

