Supporting Information

Selectively Actinide-Catalyzed Tandem Proton-Transfer Esterification of Aldehydes with Alcohols for the Production of Asymmetric Esters

Heng Liu, Moris S. Eisen*

Schulich Faculty of Chemistry, Technion – Israel Institute of Technology, Haifa City, 32000, Israel.

Table of content	S1
Experimental section	S2
General considerations	S2
General procedures for proton transfer esterification process	S2
Characterization data	S4
Kinetic studies	S8
Deuterium labeling studies	S10
Discussion of kinetic rate law	S11
Stoichiometric reactions	S12
References	S14
NMR spectra	S15

Experimental section

General considerations

All manipulations of air-sensitive materials were performed with the rigorous exclusion of oxygen and moisture in flamed Schlenk-type glassware or J-Young Teflon valve-sealed NMR tubes on a dual manifold Schlenk line interfaced to a high vacuum (10⁻⁵ Torr) line, or in a nitrogen-filled Innovative Technologies glovebox with a medium-capacity recirculator $(1 - 2 \text{ ppm of } O_2)$. Argon and nitrogen were purified by passage through MnO oxygen-removal column and a Davison 4Å molecular sieve column. Hydrocarbon solvents benzene- d^6 (Cambridge Isotopes), toluene (Bio-Lab), were distilled under vacuum from Na/K alloy. Liquid aldehydes were distilled over sodium bicarbonate and stored in a glovebox prior to use, solid aldehydes were recrystallized twice and then dried for 12 h on a high vacuum line (10⁻⁵ Torr) and stored in a glovebox prior to use. Methanol, ethanol, isopropanol, *tert*-butanol, benzyl alcohol was dried using sodium (Na) metal (or CaH₂), distilled, and stored over 4 Å molecular sieves. The actinide complexes $[(Me_3Si)_2N]_2An[\kappa^2-(N,C)-CH_2Si(CH_3)_2N(SiMe_3)]$ (An = Th (1), U(2)) and U[N(SiMe_3)_2]_3 (3) were prepared according to published procedures.¹ All the aforementioned reagents were stored in an inert atmosphere glovebox prior to use. O-deuterated methanol was purchased from Sigma Aldrich, and dried according the above procedure, and storing over 4 Å molecular sieves. Deuterated benzylaldehyde, PhCDO, was prepared according to previous reports, storing over 4 Å molecular sieves after being dried.²

NMR spectra were recorded on Bruker Avance 300, Bruker Avance III 400 spectrometers on crude reaction mixtures. Chemical shifts for ¹H and ¹³C NMR are referenced to internal protiosolvent and reported relative to tetramethylsilane.

General procedures for proton transfer esterification process

In a typical experiment, into a J. Young Teflon sealed NMR tube was added aldehyde (1.044 mmol) and alcohol (0.348 mmol) (α , α , α -trifluromethylacetophenone was added if necessary), followed by adding desired amount of catalyst (7µmol) in C₆D₆. Samples were then sealed and placed in an oil bath preheated to 70 °C, and the reaction progress monitored at regular intervals using ¹H NMR spectroscopy for up to 24 hours. The yield was calculated from the ratio of esters and alcohols from the crude ¹H NMR spectra (see examples in Figure S1). After completion of the reaction, the pure product was obtained by flash column chromatography on silica gel (20:1) and compared with previous reports.

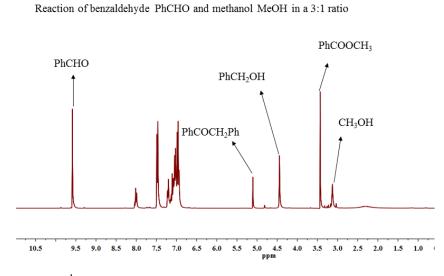
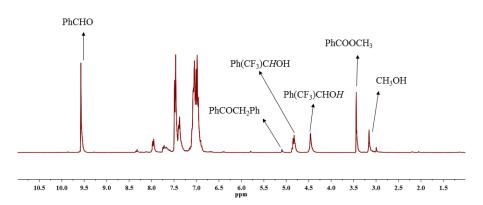


Figure S1. Crude ¹H NMR spectra of the reaction between PhCHO and MeOH (3:1).



Reaction of benzaldehyde PhCHO and methanol MeOH in a 3:1 ratio in the presence of TFMAP

Figure S2. Crude ¹H NMR spectra of the reaction between PhCHO and MeOH (3:1) in the presence of TFMAP

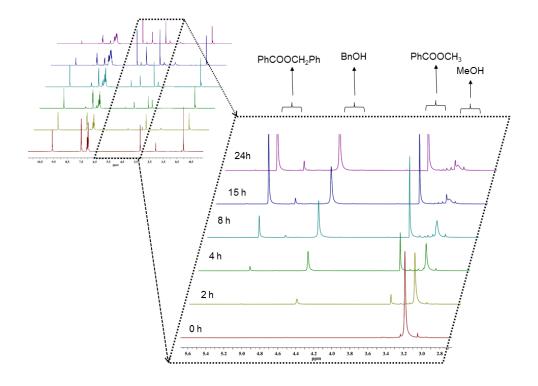
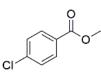


Figure S3. Reaction progress of the reaction between PhCHO and MeOH (3:1).

Characterization data

Methyl benzoate³: reaction of benzaldehyde (1.044mmol, 106.4 μL) with methanol (0.348mmol, 14.1μL) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 8.13 – 7.86 (m, 2H, H_{Ar}), 7.67 – 7.33 (m, 3H, H_{Ar}), 3.94 (s, 3H, OCH₃). ¹³C NMR (75 MHz, CDCl₃) δ 167.13, 132.92, 130.13, 129.57, 128.36, 52.12. MS (APCI): m/z 137.0607(M+H)*.



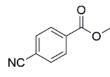
Methyl 4-chlorobenzoate⁴: reaction of 4-chlorobenzaldehyde (1.044mmol, 146.8mg) with methanol (0.348mmol, 14.1µL) was carried out following the general procedure described above.¹H

NMR (300 MHz, CDCl₃) δ 8.05 – 7.84 (m, 2H, *H*_{Ar}), 7.50 – 7.31 (m, 2H, *H*_{Ar}), 3.89 (s, 3H, C*H*₃). ¹³C NMR (75 MHz, CDCl₃) δ 166.15, 139.26, 130.86, 128.61, 128.45, 51.92. MS (APCI): m/z 171.0204 (M+H)*.

 O_2N O_2N

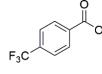
CDCl₃) δ 8.41 – 8.22 (m, 2H, *H*_{Ar}), 8.17 (m, 2H, *H*_{Ar}), 3.98 – 3.86 (m, 3H, C*H*₃). ¹³C NMR (75 MHz, CDCl₃) δ 165.07, 150.35, 135.35, 130.60, 123.44, 52.74. MS (APCI): m/z 182.0476(M+H)*.

Methyl 3-nitrobenzoate⁶: reaction of 3-nitrobenzaldehyde O_2N (1.044mmol, 157.8mg) with methanol (0.348mmol, 14.1µL) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 8.91 – 8.76 (m, 1H, *H*_{Ar}), 8.50 – 8.25 (m, 2H, *H*_{Ar}), 7.74 – 7.50 (m, 1H, *H*_{Ar}), 3.98 (s, 3H, C*H*₃). ¹³C NMR (75 MHz, CDCl₃) δ 164.86, 148.14, 135.18, 131.73, 129.54, 127.30, 124.51, 52.72. MS (APCI): m/z 182.0411(M+H)*.



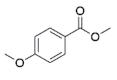
Methyl 4-cyanobenzoate⁵: reaction of 4-cyanobenzaldehyde (1.044mmol, 136.9mg) with methanol (0.348mmol, 14.1µL) was carried out following the general procedure described above. ¹H

NMR (300 MHz, CDCl₃) δ 8.15 – 8.00 (m, 2H, *H*_{Ar}), 7.80 – 7.55 (m, 2H, *H*_{Ar}), 3.93 (s, 3H, C*H*₃). ¹³C NMR (75 MHz, CDCl₃) δ 165.33, 133.79, 132.12, 129.99, 117.87, 116.27, 52.64. MS (APCI): m/z 162.0563 (M+H)*.



Methyl 4-trifluoromethylbenzoate⁷: reaction of 4-trifluorobenzadehyde (1.044mmol, 140.0 μ L) with methanol (0.348mmol, 14.1 μ L) was carried out following the general

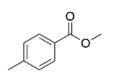
procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 8.14 (d, *J* = 8.1 Hz, 1H, *H*_{Ar}), 7.69 (d, *J* = 8.1 Hz, 1H, *H*_{Ar}), 3.94 (s, 1H, *CH*₃). MS (APCI): m/z 205.0433(M+H)*.



Methyl anisate⁵: reaction of 4-anisaldehyde (1.044mmol, 126.8 μ L) with methanol (0.348mmol, 14.1 μ L) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ

8.03 – 7.87 (m, 2H, *H*_{Ar}), 6.95 – 6.73 (m, 2H, *H*_{Ar}), 3.87 – 3.86 (m, 3H, *CH*₃), 3.85 – 3.83 (m, 2H, *CH*₃). ¹³C NMR (75 MHz, CDCl₃) δ 166.78, 163.22, 131.48, 122.53, 113.32, 55.21, 51.71. MS (APCI): m/z 166.01 (M+H)*.

Methyl 4-methylbenzoate⁵: reaction of 4-methylbenzaldehyde (1.044mmol, 123.1 μ L) with methanol (0.348mmol, 14.1 μ L) was carried out following the general procedure



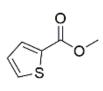
described above. ¹H NMR (300 MHz, CDCl₃) δ 7.92 – 7.84 (m, 2H, HAr), 7.19 – 7.14 (m, 2H, HAr), 3.87 (s, 3H, CH₃), 2.38 (s, 3H, CH₃). ¹³C NMR (75 MHz, CDCl₃) δ 167.10, 143.46, 129.48, 128.97,

127.28, 51.39, 21.29. MS (APCI): m/z 151.0758 (M+H)*.



Methyl picolinate⁸: reaction of 2-pyridinecarboxaldehyde (1.044mmol, 99.3 µL) with methanol (0.348mmol, 14.1µL) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 8.81 - 8.63 (m, 1H, H_{Ar}), 8.28 - 7.96 (m, 1H, H_{Ar}), 7.88 - 7.64 (m, 1H, H_{Ar}), 7.54 - 647.36 (m, 1H, H_{Ar}), 3.99 (s, 3H, CH₃). ¹³C NMR (75 MHz, CDCl₃) δ 165.59, 149.69,

147.82, 136.92, 126.83, 125.01, 52.78. MS (APCI): m/z 138.0572 (M+H)*.



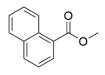
Methyl thenoate⁹: reaction of 2-thenaldehyde (1.044mmol, 97.6 µL) with methanol (0.348mmol, 14.1µL) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 7.80 - 7.77 (m, 1H, H_{Ar}), 7.55 - 7.52 (m, 1H, H_{Ar}), 7.11 - 7.06 (m, 1H,

*H*_{Ar}), 3.87 (s, 3H, C*H*₃). ¹³C NMR (75 MHz, CDCl₃) δ 162.83, 133.71, 133.38, 132.22, 127.65, 51.96. MS (APCI): m/z 143.0174 (M+H)*



Methyl 2-furoate⁹: reaction of 2-furaldehyde (1.044mmol, 87µL) with methanol (0.348mmol, 14.1µL) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 7.59 – 7.53 $(m, 1H, H_{Ar}), 7.19 - 7.13 (m, 1H, H_{Ar}), 6.54 - 6.46 (m, 1H, H_{Ar}), 3.88 (s, 3H, CH_3).$

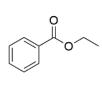
NMR (75 MHz, CDCl₃) δ 158.95, 146.15, 143.41, 117.80, 111.71, 51.79. MS (APCI): m/z 127.0411 (M+H)*.



Methyl naphthalene-1-carboxylate⁵: reaction of 1-naphthaldehyde (1.044mmol, 141.8µL) with methanol (0.348mmol, 14.1µL) was carried out following the general procedure described above. ¹H NMR

 $(300 \text{ MHz}, \text{CDCl}_3) \delta 8.92 - 8.83 \text{ (m, 1H, } H_{\text{Ar}}\text{)}, 8.20 - 8.11 \text{ (m, 1H, } H_{\text{Ar}}\text{)}, 8.01 \text{ (d, } J =$ 8.2 Hz, 1H, H_{Ar}), 7.91 – 7.80 (m, 1H, H_{Ar}), 7.64 – 7.56 (m, 1H, H_{Ar}), 7.56 – 7.43 (m, 2H, H_{Ar}), 3.99 (s, 3H, CH₃). ¹³C NMR (75 MHz, CDCl₃) δ 167.95, 133.71, 133.27, 131.20, 130.11, 128.43, 127.66, 126.95, 126.10, 125.68, 124.39, 51.71. MS (APCI): m/z 187.0838 (M+H)*.

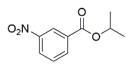
Methyl naphthalene-2-carboxylate⁵: reaction of 2-naphthaldehyde (1.044mmol, 163.5mg) with methanol (0.348mmol, 14.1μL) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 8.60 (s, 1H, H_{Ar}), 8.05 (d, J = 8.6 Hz, 1H, H_{Ar}), 7.98 – 7.91 (m, 1H, H_{Ar}), 7.90 – 7.82 (m, 2H, H_{Ar}), 7.61 – 7.48 (m, 2H, H_{Ar}), 3.97 (s, 3H, CH₃). ¹³C NMR (75 MHz, CDCl₃) δ 167.17, 135.39, 132.36, 130.96, 129.25, 128.13, 128.05, 127.65, 127.26, 126.53, 125.11, 52.03. MS (APCI): m/z 187.0762 (M+H)*.



Methyl benzoate⁹: reaction of benzaldehyde (1.044mmol, 106.4 μ L) with ethanol (0.348mmol, 20.3 μ L) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 8.07

- 7.95 (m, 2H, *H*_{Ar}), 7.57 - 7.45 (m, 1H, *H*_{Ar}), 7.45 - 7.34 (m, 2H, *H*_{Ar}), 4.51 - 4.19 (q, *J* = 7.2 Hz, 2H, CH₂), 1.36 (t, *J* = 7.2 Hz, 3H, CH₃). ¹³C NMR (75 MHz, CDCl₃) δ 166.43, 132.68, 130.37, 129.22, 128.07, 60.57, 14.19. MS (APCI): m/z 151.0748 (M+H)*.

 O_2N **Ethyl 3-nitrobenzoate**¹⁰: reaction of 3-nitrobenzaldehyde (1.044mmol, 157.8mg) with ethanol (0.348mmol, 20.3μL) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 8.84 – 8.70 (m, 1H, *H*_{Ar}), 8.39 – 8.17 (m, 2H, *H*_{Ar}), 7.64 – 7.46 (m, 1H, *H*_{Ar}), 4.35 (q, *J* = 7.1 Hz, 2H, *CH*₂), 1.34 (d, *J* = 7.1 Hz, 3H, *CH*₃). ¹³C NMR (75 MHz, C₆D₆) δ 161.77, 148.12, 135.17, 131.97, 129.49, 127.12, 124.23, 61.46, 13.78. MS (APCI): m/z 195.0291.



Isopropyl 3-nitrobenzoate¹⁰: reaction of 3-nitrobenzaldehyde (1.044mmol, 157.8mg) with isopropanol (0.348mmol, 26.6μL) was carried out following the general procedure described above.

¹H NMR (300 MHz, CDCl₃) δ 8.90 – 8.78 (m, 1H, *H*_{Ar}), 8.45 – 8.30 (m, 2H, *H*_{Ar}), 7.67 – 7.56 (m, 1H, *H*_{Ar}), 5.28 (hept, *J* = 6.3 Hz, 1H, *CH*), 1.39 (d, *J* = 6.3 Hz, 6H, *CH*₃). ¹³C

NMR (75 MHz, CDCl₃) δ 191.69, 162.21, 149.45, 135.23, 132.55, 129.38, 126.98, 124.35, 69.38, 21.76. MS (APCI): m/z 209.0431.

Phenylmethyl 3-nitrobenzote¹⁰ : reaction of 3-nitro benzaldehyde (1.044mmol, 157.8mg) with benzyl alcohol (0.348mmol, 36.05µL) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 8.91 – 8.82 (m, 1H), 8.54 – 8.26 (m, 2H), 7.63 (t, J = 7.7 Hz, 1H), 7.48 – 7.34 (m, 5H), 5.40 (s, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 164.21, 148.20, 135.26, 135.14, 131.81, 129.52, 128.63, 128.54, 128.37, 127.37, 124.57, 67.51. MS (APCI): m/z 257.0742.

Benzyl alcohol- d_2^{11} : reaction of benzaldehyde-d (1.044mmol, 110.2mg) With methanol (0.348mmol, 14.1µL) was carried out following the general procedure described above. ¹H NMR (300 MHz, CDCl₃) δ 7.35

(d, 4H, *H*_{Ar}), 7.32 – 7.25 (m, 1H, *H*_{Ar}), 1.65 (s, 1H, O*H*). ¹³C NMR (75 MHz, CDCl₃) δ 140.68, 128.52, 127.51, 126.87, 64.95.

Benzyl benzoate- d_2 : reaction of benzaldehyde-d (1.044mmol, 110.2mg) with methanol (0.348mmol, 14.1µL) was carried out following the general procedure described above. ¹H NMR (300

MHz, CDCl₃) δ 8.07 (d, *J* = 7.7 Hz, 2H), 7.54 (t, *J* = 7.1 Hz, 1H), 7.49 – 7.28 (m, 7H). ¹³C NMR (75 MHz, CDCl₃) δ 166.31, 135.82, 132.90, 130.12, 129.59, 128.48, 128.25, 128.14, 128.09, 65.83. MS (APCI): m/z 214.1290.

Kinetic studies

All the kinetic experiments were done in a similar method. In a J. Young NMR tube, typical amount of precatalyst **1** (3 – 14 μ mol), PhCHO (0.348 - 2.1mmol), CH₃OH (0.174 – 1.044) and C₆D₆ was added in the glove box and then the tube was sealed. Take the tube out of the glove box and freeze it in ice bath until the ¹H NMR experiment began. All the experiments were done by changing one substrate or catalyst while keeping the other reagents constant, and the data was collected every five minutes up to six hours. The product concentrations were measured by the area ratio of *methyl* group at 3.08 ppm and 3.42 ppm, which were assigned to the starting material MeOH

and ester product PhCOOMe respectively. Reaction rates were determined by leastsquare fit of product concentration versus time, and the plots were shown in Figures S4-6.

Activation parameters including enthalpy (ΔH^{\neq}) , entropy (ΔS^{\neq}) and activation energy (E_a) were calculated from kinetic data using Eyring and Arrhenius plots. In a typical sample, the J. Young tube was loaded with desired amount of precatalyst **1**, PhCHO, CH₃OH, C₆D₆ and sealed. Then the sample was inserted into Bruker Avance 300 spectrometer which had been previously set to the desired temperature. The data was collected every one minute up to six hours. Reaction rates were determined by the least square fit of product concentration versus time, and Eyring and Arrhenius plots were shown in Figures S7-8. Enthalpy (ΔH^{\neq}), entropy (ΔS^{\neq}) and activation energy (E_a) were calculated from the slope and intercept of the least-square fit.

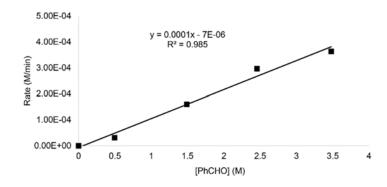


Figure S4. Plot of reaction rate versus concentration of PhCHO by complex 1.

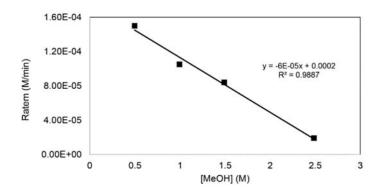


Figure S5. Plot of reaction rate versus concentration of MeOH by complex 1.

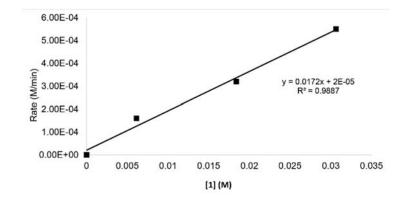


Figure S6. Plot of reaction rate versus concentration of complex 1.

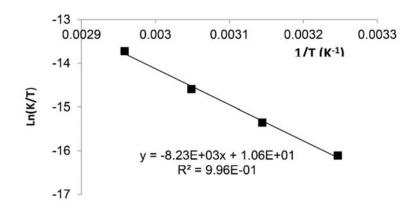


Figure S7. Eyring plot for the reaction of PHCO and MeOH using complex 1.

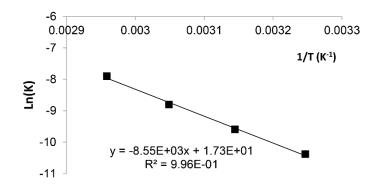


Figure S8. Arrhenius plot for the reaction of PHCHO and MeOH using complex 1.

Deuterium labeling studies.

In a J. Young NMR tube, typical amount of precatalyst **1** (7 μ mol), PhCHO (or PhCDO, 1.044mmol), CH₃OH (MeOD, 1.044 mmol) and C₆D₆ was added in the glove box and then the tube was sealed. Take the tube out of the glove box and freeze it in ice bath until the ¹H NMR experiment began. All the experiments were done by changing

one substrate or catalyst while keeping the other reagents constant, and the data was collected every five minutes up to six hours. The reaction progresses were shown in Figure S9.

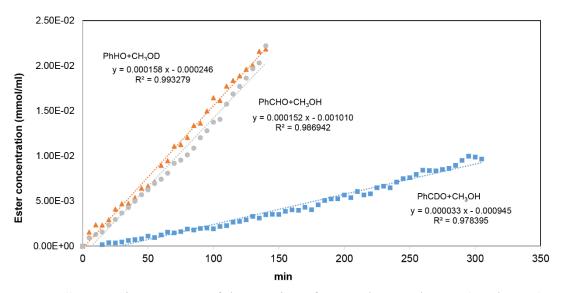
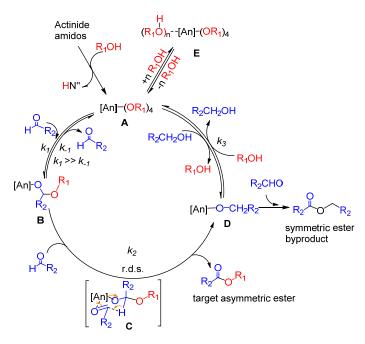


Figure S9. Reaction progress of the reaction of precatalyst **1**, PhCHO (or PhCDO), CH₃OH (MeOD).

Discussion of kinetic rate law

According to the kinetic and thermodynamic data, the following mechanism is proposed:



S11

Deuterium labeling studies showed that the hydride transfer through the sixmember ring insertion is the turnover limiting step, and the remaining steps of the cycle are rapid (k_1 , k_3), so the following rate equation can be obtained based on the assumption of a steady-state:

$$\frac{\partial p}{\partial t} = k_2 [\mathbf{B}] [R_2 CHO]$$
$$k_1 [\mathbf{A}] [R_2 CHO] - k_{-1} [\mathbf{B}] - k_2 [\mathbf{B}] [R_2 CHO] = 0$$

Substitution **B** with **A** gives the following equation:

$$\frac{\partial p}{\partial t} = k_2 \frac{k_1 [\mathbf{A}] [R_2 CHO]^2}{k_{-1} + k_2 [R_2 CHO]}$$

In the formation of species **A**, alcoholysis of actinide amido compounds is rapid and irreversible, however, the equilibrium of alcohol coordination/decoordination to actinide center must be considered,¹² giving the following expression:

$$K_{eq} = \frac{[E]}{[R_1 OH][A]}$$

From the reaction cycle, it can be found that the sum of active catalysts (**A**) and alcohol-saturated complex (**E**) is the total amount of catalyst loading [An] used in the reaction: [A] + [E] = [An]. After substitution of [A] from equilibrium expression into rate equation, the following kinetic rate law is obtained:

$$\frac{\partial p}{\partial t} = k_2 \frac{k_1 [An] [R_2 CHO]^2}{(1 + K_{eq} [R_1 OH])(k_{-1} + k_2 [R_2 CHO])}$$

From this equation, inverse-first order behavior on R₁OH and first order dependence on $[R_2CHO]$ can be concluded if $K_{eq}[R_1OH]$ is very large and $k_2[R_2CHO] \gg k_{-1}$, giving rise to the equation as following:

$$\frac{\partial p}{\partial t} = \frac{k_1}{K_{eq}} \frac{[An][R_2CHO]}{[R_1OH]} = k'[An][R_2CHO][R_1OH]^{-1}$$

Stoichiometric reactions

A J. Young NMR tube was loaded with 0.056 mmol complex **1**, 0.56 mmol of MeOH, and 500 μ L C₆D₆, and then the tube was put into the oil bath which had been previously set to desired temperature (70°C), and monitored by ¹H NMR spectroscopy.

After 12h, another 0.56 mmol of PhCHO was added, and the reaction was monitored by ¹H NMR spectroscopy.

A J. Young NMR tube was loaded with 0.056 mmol complex **1**, 0.56 mmol of PhCHO and 500 μ L C₆D₆, and then the tube was put into the oil bath which had been previously set to desired temperature (70°C), and monitored by ¹H NMR spectroscopy. After 12h, another 0.56 mmol of MeOH was added, and the reaction was monitored by ¹H NMR spectroscopy.

Reference

- 1 Dormond, A.; El Bouadili, A.; Aaliti, A.; Moise, C., J. Organomet. Chem. 1985, 288 (1), C1-C5.
- 2 Gajewski, J. J.; Bocian, W.; Harris, N. J.; Olson, L. P.; Gajewski, J. P., *J. Am. Chem. Soc.* 1999, 121
 (2), 326-334.
- 3 Kim, J.-J.; Park, Y.-D.; Kweon, D.-H.; Kang, Y.-J.; Kim, H.-K.; Lee, S.-G.; Cho, S.-D.; Lee, W.-S.; Yoon, Y.-J., *Bull. Korean Chem. Soc.* **2004**, *25* (4), 501-505.
- 4 Yamamoto, Y., Adv. Synth. Catal. 2010, 352 (2-3), 478-492.
- 5 Lerebours, R.; Wolf, C., J. Am. Chem. Soc. 2006, 128 (40), 13052-13053.
- 6 Aridoss, G.; Laali, K. K., J. Org. Chem. 2011, 76 (19), 8088-8094.
- 7 Knauber, T.; Arikan, F.; Röschenthaler, G.-V.; Gooßen, L. J., Chem. Eur. J. 2011, 17 (9), 2689-2697.
- 8 Mamane, V.; Aubert, E.; Fort, Y., J. Org. Chem. 2007, 72 (19), 7294-7300.
- 9 Gowrisankar, S.; Neumann, H.; Beller, M., Angew. Chem. Int. Ed. 2011, 50 (22), 5139-5143.
- 10 Wang, Z.; Kang, J.; Yu, M., J. Chem. Res 2007, 2007 (6), 323-324.
- 11 Golisz, S. R.; Labinger, J. A.; Bercaw, J. E., Organometallics 2010, 29 (21), 5026-5032.
- 12 Van der Sluys, W. G.; Sattelberger, A. P., Chem. Rev. 1990, 90 (6), 1027-1040.

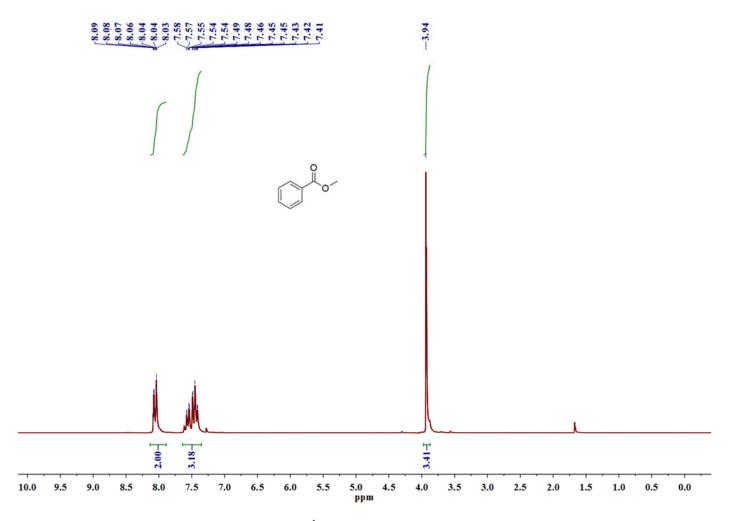
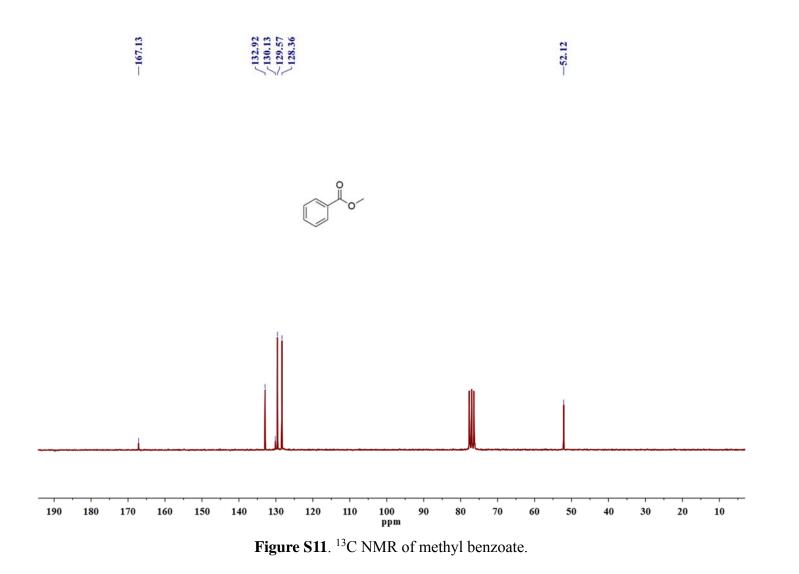


Figure S10. ¹H NMR of methyl benzoate.



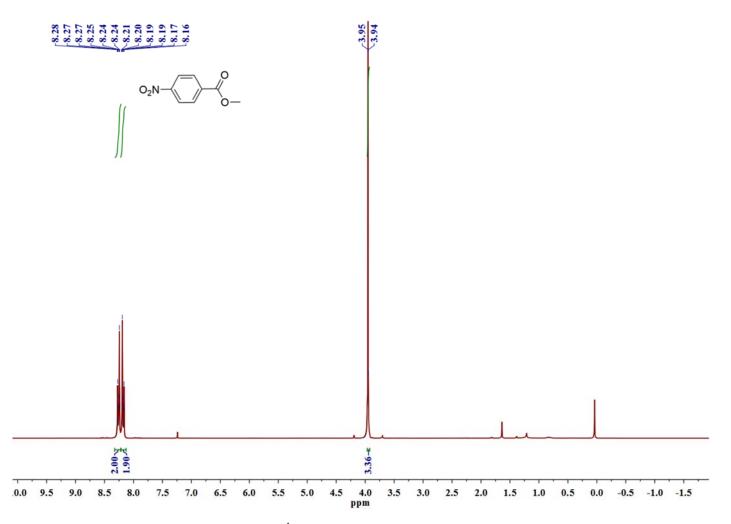


Figure S12. ¹H NMR of methyl 4-nitrobenzoate.

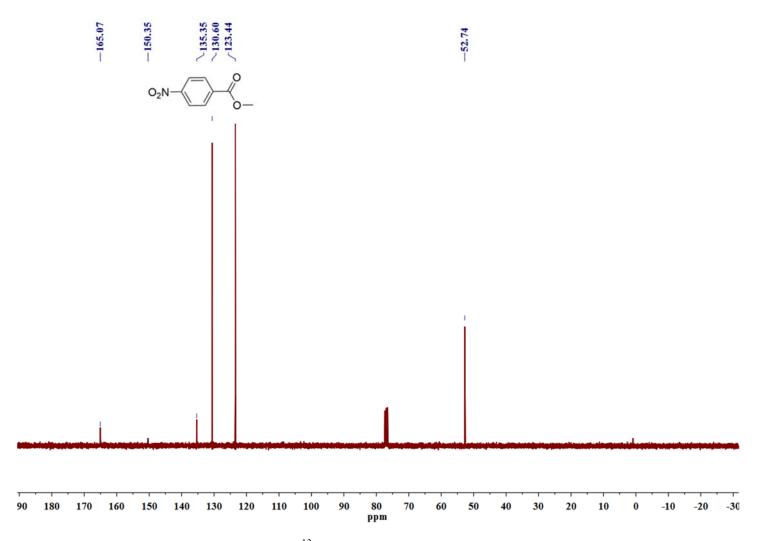


Figure S13. ¹³C NMR of methyl 4-nitrobenzoate.

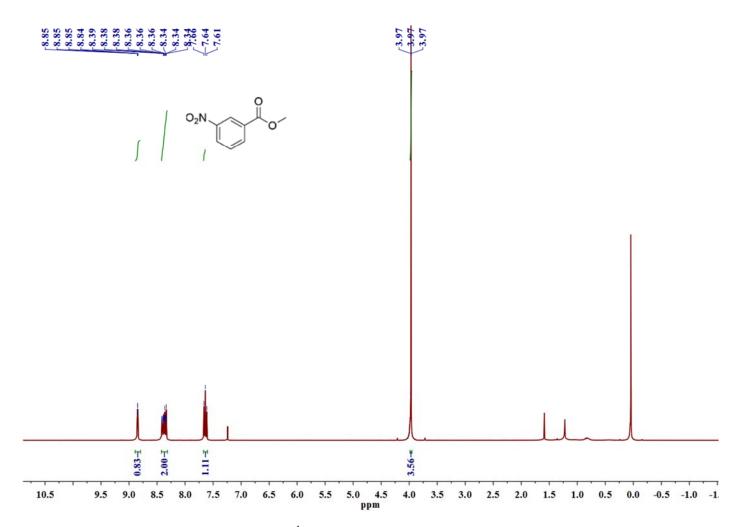


Figure S14. ¹H NMR of methyl 3-nitrobenzoate.

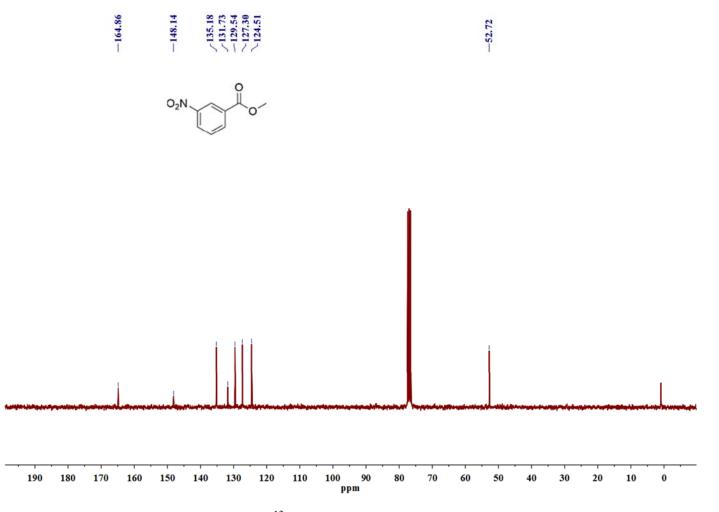


Figure S15. ¹³C NMR of methyl 3-nitrobenzoate.

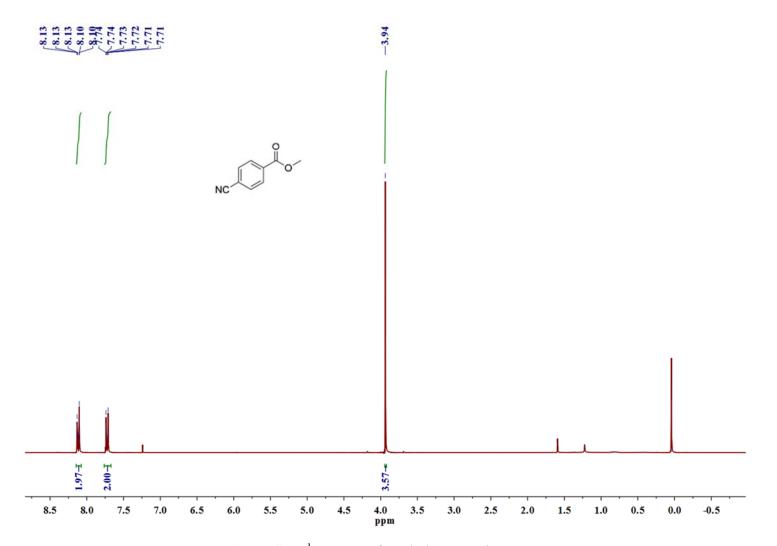


Figure S16. ¹H NMR of methyl 4-cyanobenzoate.

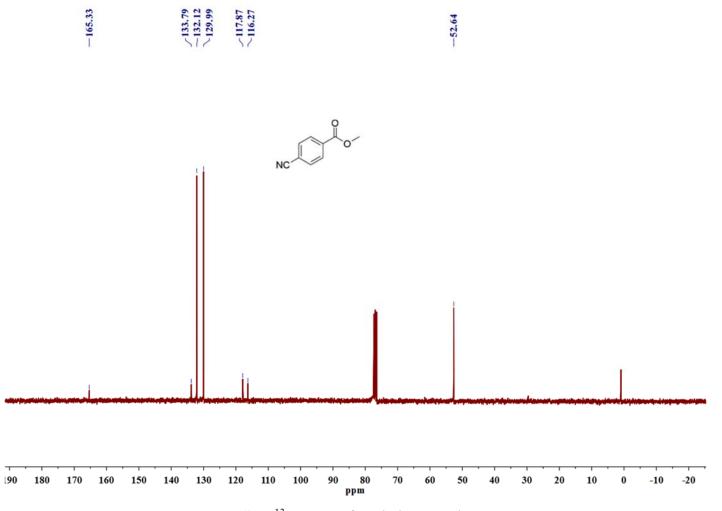


Figure S17. ¹³C NMR of methyl 4-cyanobenzoate.

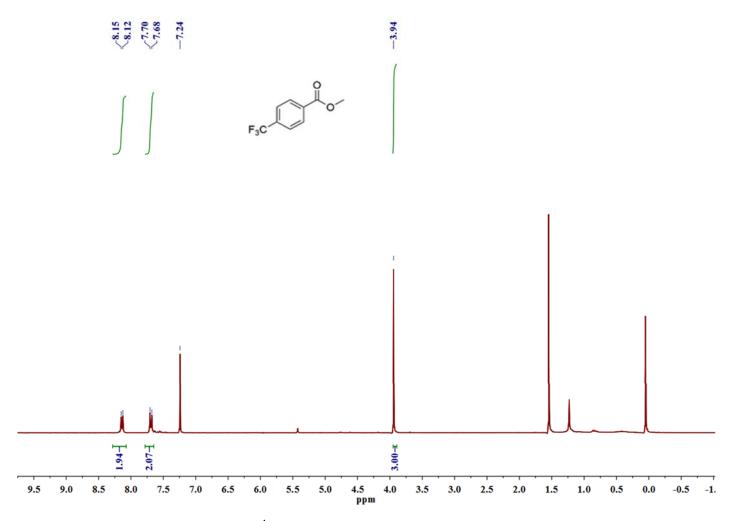
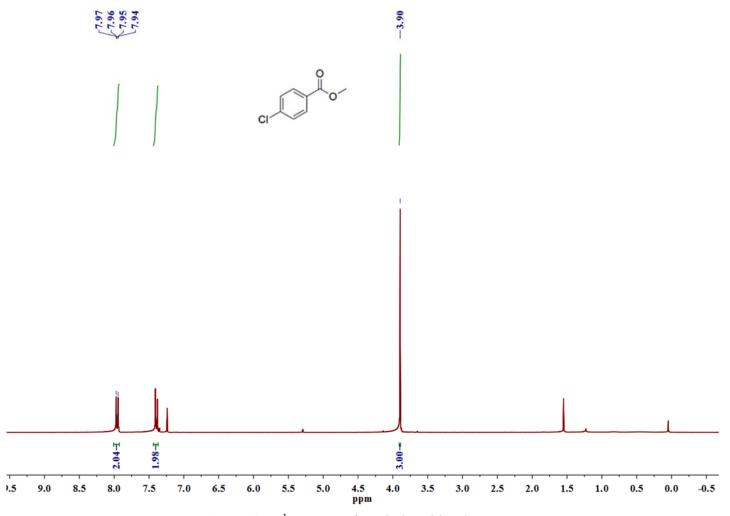
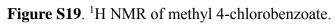


Figure S18. ¹H NMR of methyl 4-trifluromethylbenzoate.





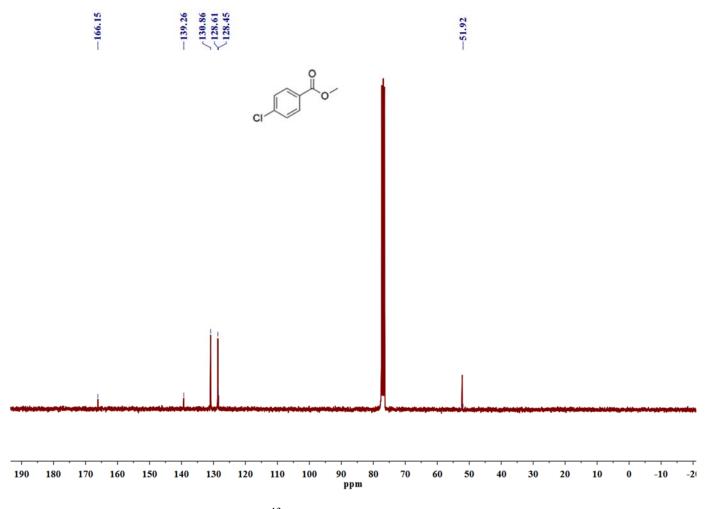


Figure S20. ¹³C NMR of methyl 4-chlorobenzoate.

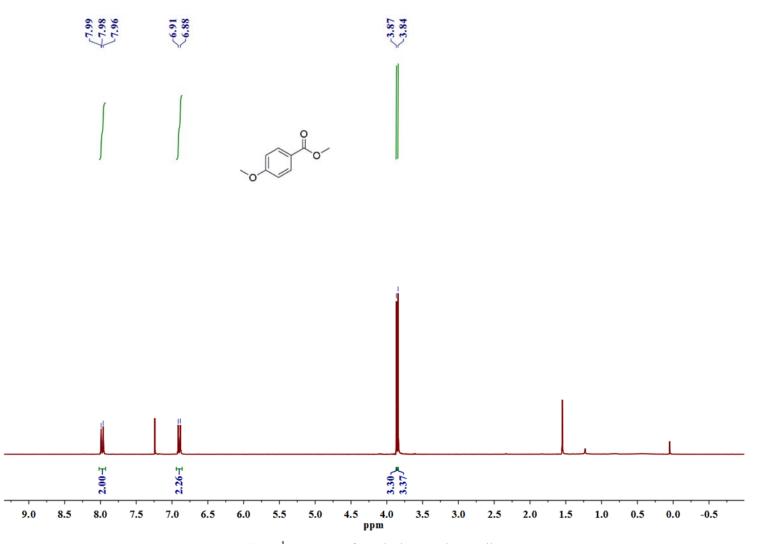


Figure S21. ¹H NMR of methyl 4-methyoxylbenzoate.

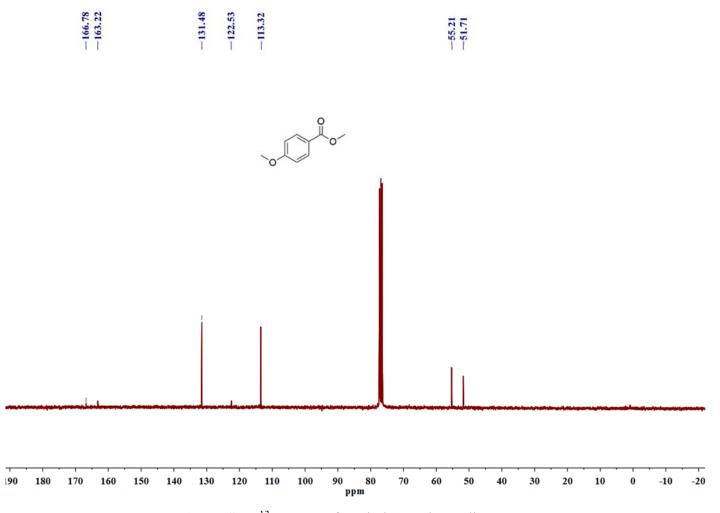


Figure S22. ¹³C NMR of methyl 4-methyoxylbenzoate.

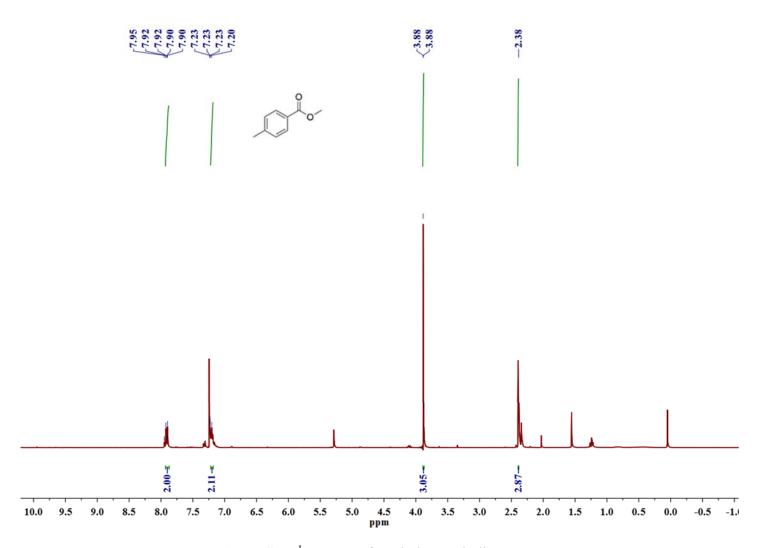


Figure S23. ¹H NMR of methyl 4-methylbenzoate.

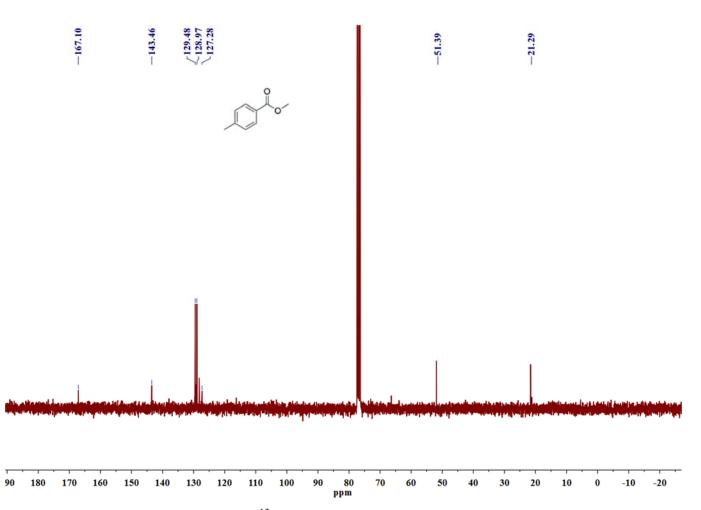


Figure S24. ¹³C NMR of methyl 4-methylbenzoate.

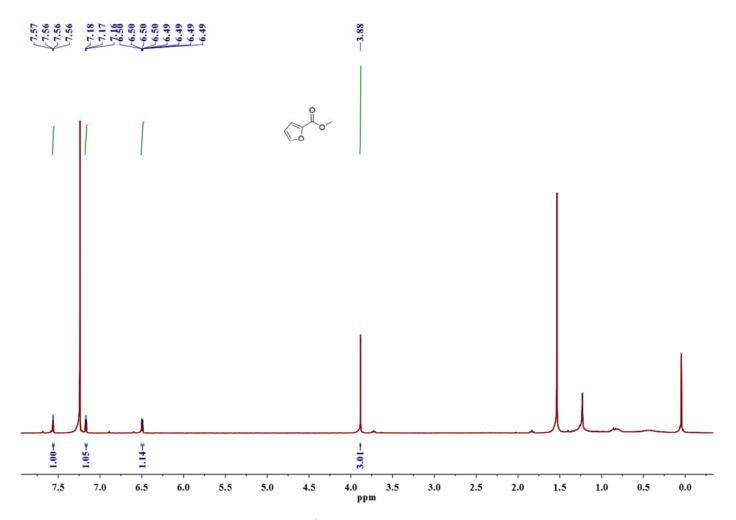
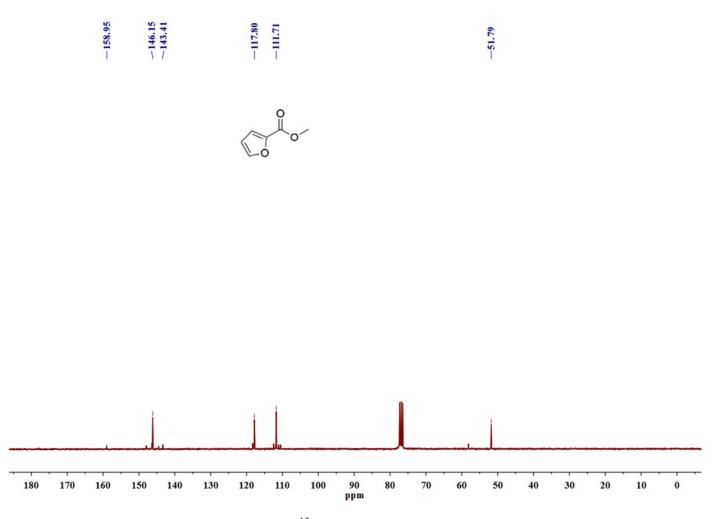
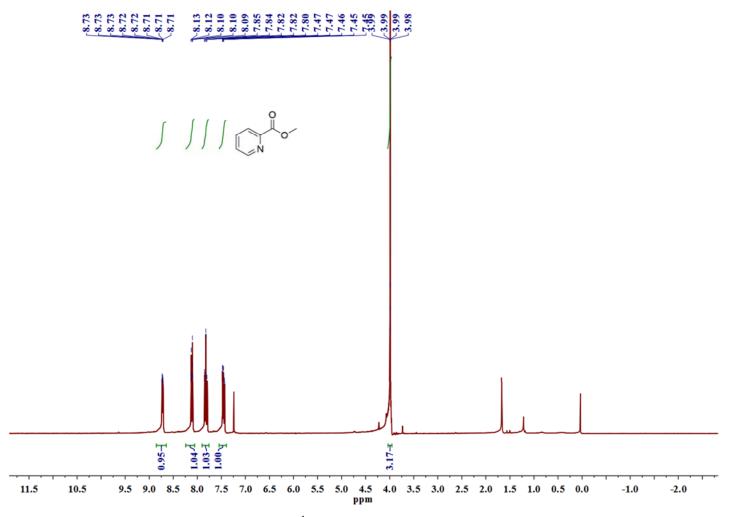
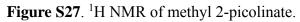


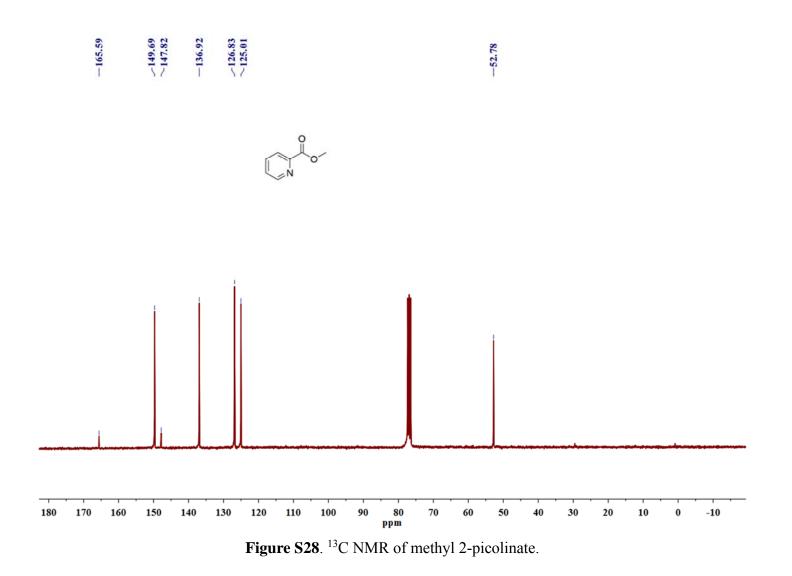
Figure S25. ¹H NMR of methyl 2-furoate.











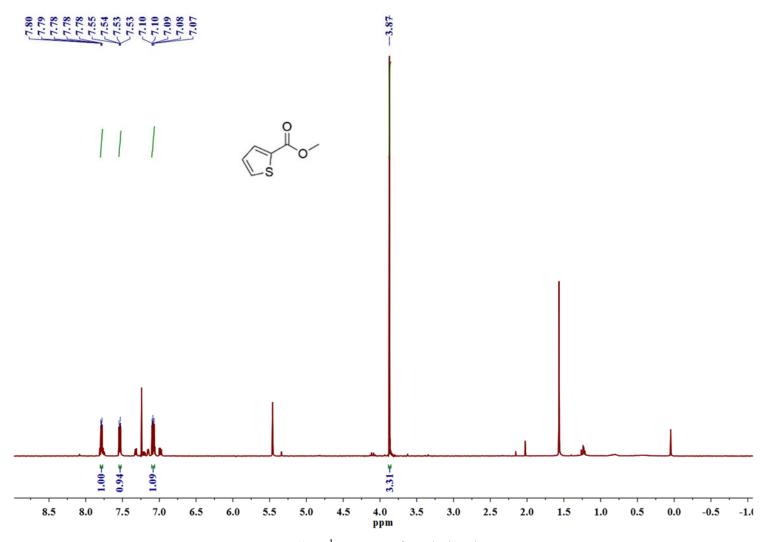


Figure S29. ¹H NMR of methyl 2-thenote.

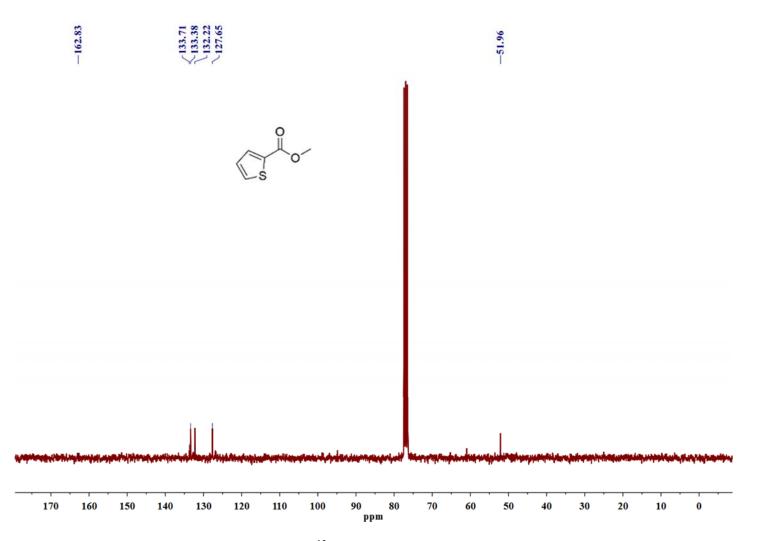


Figure S30. ¹³C NMR of methyl 2-thenote.

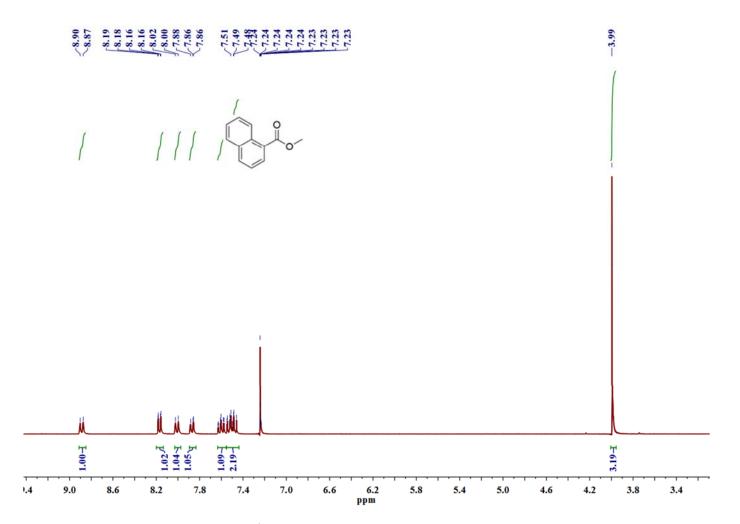


Figure S31. ¹H NMR of methyl naphthalene-1-carboxylate.

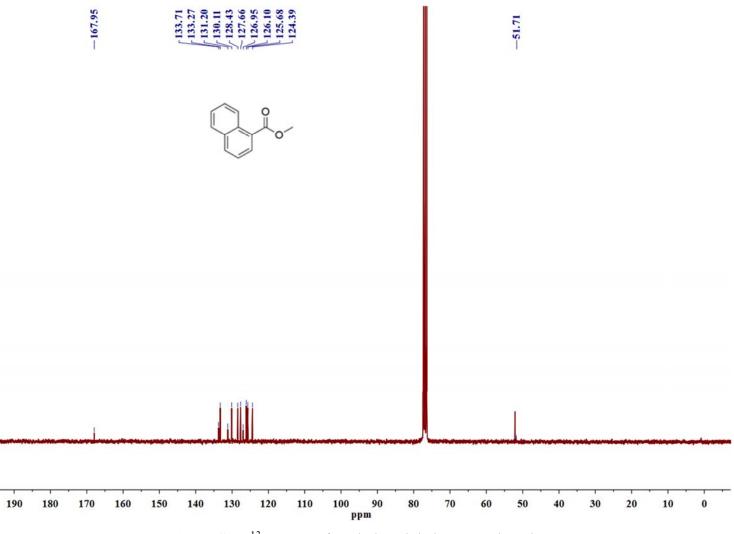


Figure S32. ¹³C NMR of methyl naphthalene-1-carboxylate.

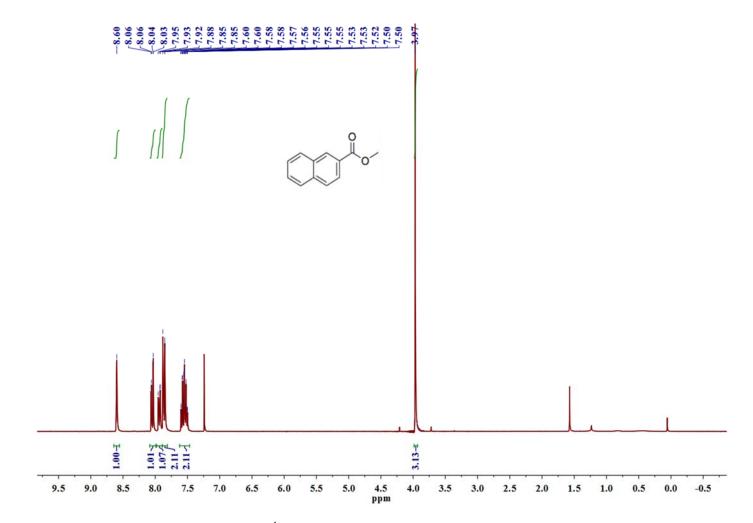


Figure S33. ¹H NMR of methyl naphthalene-2-carboxylate.

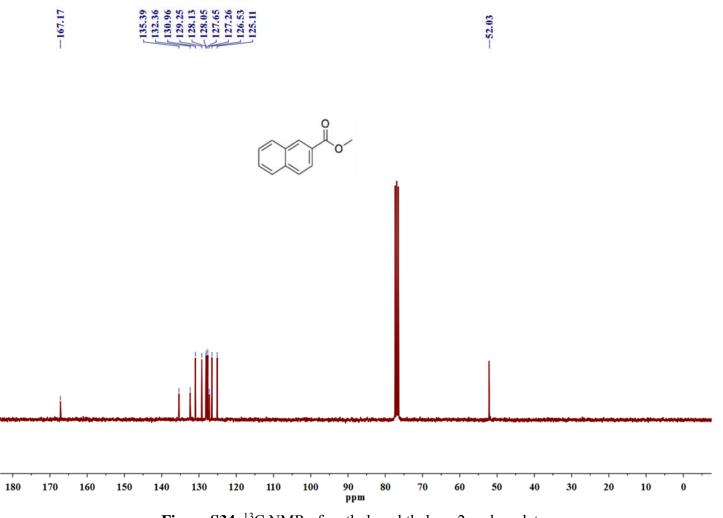
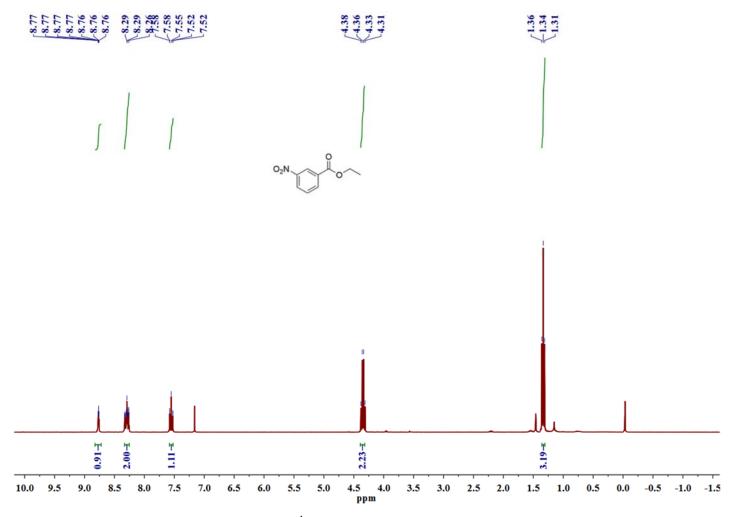
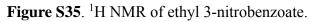
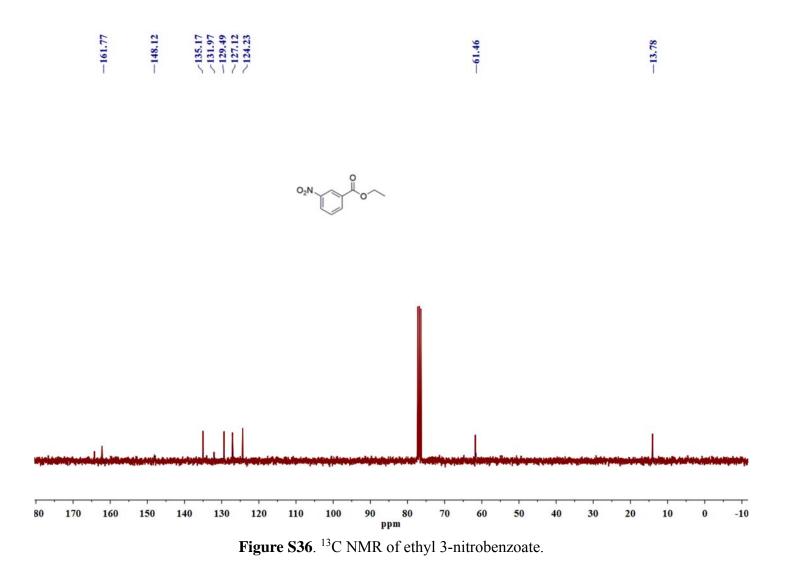


Figure S34. ¹³C NMR of methyl naphthalene-2-carboxylate.







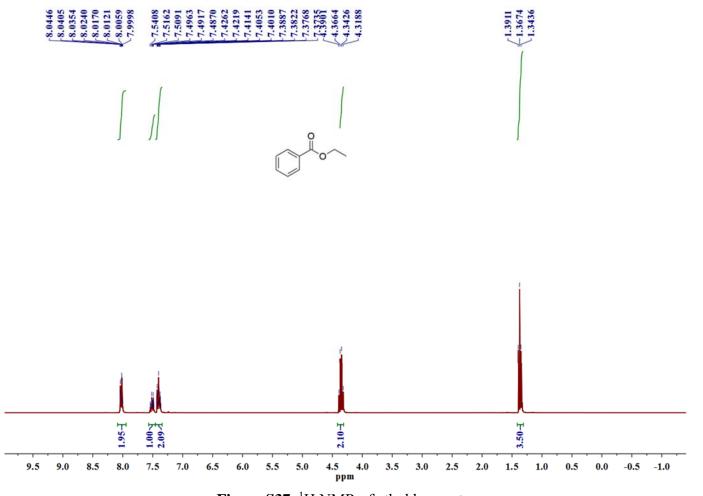
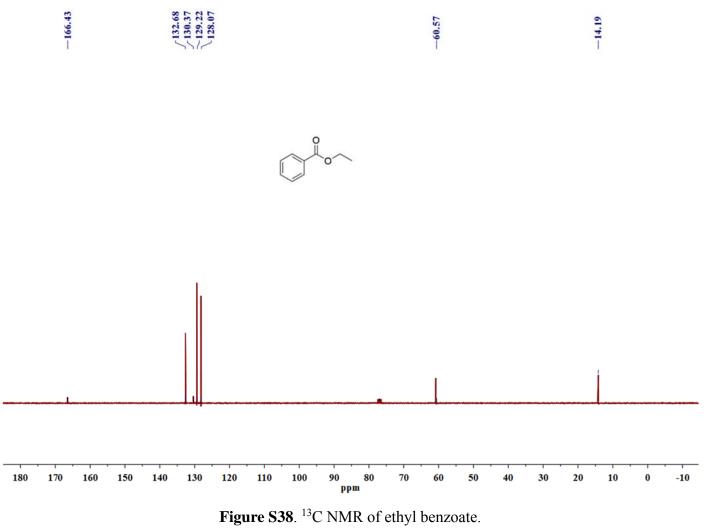


Figure S37. ¹H NMR of ethyl benzoate.





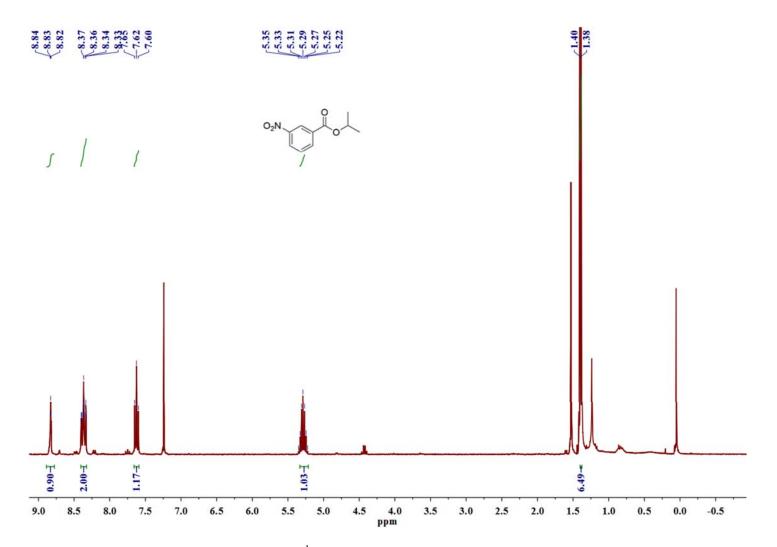


Figure S39. ¹H NMR of isopropyl 3-nitrobenzoate.