## Monomer/Oligomer Quasi-racemic Protein Crystallography

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## 1. general information

## A. Materials

2-Chlorotrityl Chloride Resin and Rink Amide-AM Resin were purchased from Tianjin Nankai Hecheng Science \& Technology Co., Ltd (China). Fmoc-amino acids, Fmoc-D-amino acid and 1-hydroxy-7-azabenzotriazole (HOAt) were purchased from CSBio (Shanghai) Ltd, GL Biochem (Shanghai) Ltd. And BO MAI JIETECHNOLOGY CO, LTD. 4-mercaptophenylacetic acid (MPAA) was purchased from Alfa Aesar. Triisopropyldilsne (TIPS), N,N-Dimethylformamide (DMF), acetic acid (HPLC grade), thioanisole, trifluoroacetic acid (TFA, HPLC grade) and phenylsilane were purchased from J\&K Chemical Ltd. (Shanghai). Ethyl cyanoglyoxylate-2-oxime, N,N-diisopropyl-carbodiimide (DIC), 1,2-ethanedithiol and N,N-diisopropylethylamine (DIEPA) were purchased from adamas-beta. Dithiothreitol (DTT) was purchased from Aladdin (Shanghai, China). Acetonitrile (HPLC grade) was purchase from J. T. Baker. $\mathrm{Na}_{2} \mathrm{HPO}_{4} \cdot 12 \mathrm{H}_{2} \mathrm{O}$, guanidine hydrochloride $(\mathrm{Gn} \cdot \mathrm{HCl})$, and ether was purchase from Sinopharm Chemical Reagent Co., Ltd. Dichloromethane (DCM) and sodium nitrite $\left(\mathrm{NaNO}_{2}\right)$ were purchased from Beijing Chemical Industry Group CO., LTD. Glycyl 1-(2,4-dimethoxyphenyl)-2-mercaptoethyl auxiliary and Fmoc-N-(2,4-dimethoxybenzyl) glycine were purchased from Nantong peptide Biotech LTD (China). Liberty blue (automated microwave peptide synthesizer) was purchased from CEM corporation.

## B. HPLC, Mass spectrometry and FPLC

Analytical RP-HPLC (SHIMADZU) was used to monitor the purity of crude peptide, chemical reaction progress and the purity of product with an analytical column (welch C18, 30 min , flow rate $1 \mathrm{ml} / \mathrm{min}$ ) at 214 nm and 254 nm . Semi-preparative HPLC (SHIMADZU) was used to separate and purify the crude peptide and reaction product with a semi preparative column (Grace Vydac C18, flow rate $5 \mathrm{ml} / \mathrm{min}$ and welch $\mathrm{XB}-\mathrm{C} 18$, flow rate $7 \mathrm{ml} / \mathrm{min}$ ) at 214 nm and 254 nm . Mobile
phase buffer A is acetonitrile ( $0.1 \%$ TFA) and Mobile phase buffer B deionized distilled water ( $0.1 \% \mathrm{TFA}$ ). Both buffers were sonicated for 30 min .

Crude peptide and reaction product were characterized by normal ESI mass spectra on LC/MS 2020 (SHIMADZU). And ultimate product was characterized by high-resolution ESI mass spectra on SYNAPTTM G2-Si HDMS.

Lyophilized polyubiquitin chains were dissolved in water contained $6 \mathrm{~mol} / \mathrm{L} \mathrm{Gn}$ $\mathrm{HCl}, 0.1 \mathrm{mM} \mathrm{Na} 2 \mathrm{HPO}_{4}, \mathrm{pH} 3.0$. Polyubiquitin chains were purified by AKTA (GE Healthcare Life Science) with Mono S cation exchange chromatography column and Superdex 75 column. Every injection was monitored at 280 nm and 214 nm .

## 2. Experimental figure

## A. HPLC Chromatography and mass spectra



Figure S1. Analytical HPLC chromatograms of purified di-Ubs and D-Ub. Inset Electrospray ionization mass spectrum (di-Ub-K6: obs. 17110.3 Da, calc. 17110.6 Da; di-Ub-K11: obs. 17110.1 Da calc. 17110.6 Da ; di-Ub-K27: obs. 17110.2 Da, calc. 17110.6 Da; di-Ub-K29: obs. 17110.0 Da , calc. 17110.6 Da ; di-Ub-K33: obs. 17110.0 Da, calc. 17110.6 Da; di-Ub-K48: obs. 17109.5 Da, calc. 17110.6 Da; di-Ub-K63: obs. 17109.4 Da, calc. 17110.6 Da ; D-Ub: obs. 8545.0 Da, calc. 8545.8 Da).

## B Characterization of synthetic peptide segments of K11/K63-branched tri-Ub



Figure S2. Analytical HPLC chromatograms of peptide segments of 11/63-branched tri-Ub. Inset Electrospray ionization mass spectrum (1-45-NHNH2: obs. 5127.1 Da , calc. $5127.6 \mathrm{Da} ; \mathbf{1 - 4 5 ( K 1 1 a g ) - \mathbf { N H N H } _ { 2 }}$ : obs. 5381.4 Da calc. 5381.0 Da ; Acm-46-75-NHNH $\mathbf{2}^{2}$ : obs. 3528.3 Da , calc. 3528.9 Da ; Acm-46-76-K63-76-46-NH2 obs. 7052.8 Da, calc. 7053.9 Da; Acm-46-76-K11-1-45-NHNH $\mathbf{2}_{2}$ : obs. 8680.5 Da, calc. 8680.9 Da ; 46-76-K11-1-76-K63-76-46-NH2 $\mathbf{2}$ : obs. 15560.9 Da , calc. 15561.8 Da ).

## C. Crystals of di-ubiquitins



Figure S3. Di-ubiquitins crystals.

## D. Number of crystallization conditions



Figure S4. Number of crystallization conditions.

## E. Static light scattering (SLS) and analytical ultracentrifugation (AUC) experiments



Figure S5. No interaction was found between linear tri-Ub and D-mono-Ub in the solution through static light scattering and analytical ultracentrifugation.

Supplementary Table 1 Data collection and refinement statistics

| Crystal | K6-linked di-Ub | K11-linked di-Ub |
| :--- | :--- | :--- |
| Data collection |  |  |
| Space group | P 1 | P 1 |
| Unit cell | $24.730,26.380,45.120 ;$ | $26.523,26.733,43.768 ;$ |
|  | $98.07,93.85,106.54$ | $75.17,76.00,80.96$ |
| Resolution $(\AA)$ | $44 \sim 1.16(1.20 \sim 1.16)$ | $26 \sim 1.73(1.8 \sim 1.73)$ |
| $\mathrm{R}_{\text {merge }}$ | $0.066(0.749)$ | $0.061(0.176)$ |
| $\mathrm{I} / \sigma \mathrm{I}$ | $11.25(1.74)$ | $8.43(2.41)$ |
| Completeness $(\%)$ | $62.8(8.6)$ | $68.2(5.4)$ |
| Redundancy | $3.54(3.09)$ | $1.8(1.3)$ |
| No. reflections | 23459 | 7976 |
| Wilson B-factor $\left(\AA^{2}\right)$ | 10.0 | 13.8 |
| Refinement |  |  |


| $\mathrm{R}_{\text {work }} / \mathrm{R}_{\text {free }}$ | 0.1912/0.2285 | 0.1868/0.2355 |
| :---: | :---: | :---: |
| No. atoms |  |  |
| Protein | 1194 | 1164 |
| Water | 191 | 165 |
| R.m.s. deviations |  |  |
| Bond lengths $(\AA)$ | 0.005 | 0.007 |
| Bond angles ( ${ }^{\circ}$ ) | 1.014 | 1.084 |
| Ramachandran plot statistics (\%) |  |  |
| Most favourable | 100\% | 100\% |
| Disallowed | 0\% | 0\% |
| Crystallization condition | 0.2M Magnesium chloride hexahydrate, 20\%PEG3350 pH5.9 | 0.2M Li $2 \mathrm{SO} 4,0.1 \mathrm{M}$ tris $8.5,30 \% \mathrm{PEG} 4000$ |


| Crystal | K27-linked di-Ub | K29-linked di-Ub |
| :--- | :--- | :--- |
| Data collection |  |  |
| Space group | P1 | P2 |
| Unit cell | $38.240,38.260,48.000 ;$ | $28.027,46.497,43.157 ;$ |
|  | $98.15,98.15,109.67$ | $90.00,97.74,90.00$ |
| Resolution $(\AA)$ | $35 \sim 1.15(1.19 \sim 1.15)$ | $46 \sim 1.98(2.05 \sim 1.98)$ |
| $\mathrm{R}_{\text {merge }}$ | $0.034(0.604)$ | $0.117(0.382)$ |
| I / $\sigma$ | $7.31(1.40)$ |  |
| Completeness $(\%)$ | $78.6(47.0)$ | $78.8(44.6)$ |
| Redundancy | $1.93(1.90)$ | $2.5(1.5)$ |
| No. reflections | 67778 | 5292 |
| Wilson B-factor $\left(\AA^{2}\right)$ | 11.6 | 28.0 |
| Refinement |  |  |

$\mathrm{R}_{\text {work }} / \mathrm{R}_{\text {free }} \quad 0.2021 / 0.2215 \quad 0.2451 / 0.3297$

| No. atoms |  |  |
| :--- | :--- | :--- |
| Protein | 2404 | 1153 |
| Water | 340 | 54 |

R.m.s. deviations

Bond lengths $0.006 \quad 0.019$
( $\AA$ )
Bond angles $\left(^{\circ}\right) \quad 0.980 \quad 1.847$

| Ramachandran plot |  |  |
| :--- | :--- | :--- |
| statistics (\%) |  | $96 \%$ |
| Most favourable | $98 \%$ | $0 \%$ |
| Disallowed | $1 \%$ | 0.2 MPotassium sulfate 20\%PEG3350 |
| Crystallization | 0.2 M Magnesium acetate tetrahydrate, |  |
| condition | 0.1 M Sodium cacodylate trihydrate |  |
|  | $\mathrm{pH} 6.5,20 \%$ PEG 8000 |  |


| Crystal | K33-linked di-Ub | K48-linked di-Ub |
| :--- | :--- | :--- |
| Data collection |  |  |
| Space group | P1 | P1 |
| Unit cell | $26.436,29.200,44.232 ;$ | $27.969,40.834,52.395 ;$ |
|  | $83.14,86.82,71.18$ | $98.469,101.190,105.937$ |
| Resolution $(\AA)$ | $27 \sim 1.95(2.02 \sim 1.95)$ | $38 \sim 1.59(1.65 \sim 1.59)$ |
| R $_{\text {merge }}$ | $0.076(0.291)$ | $0.089(1.086)$ |
| I/бI | $16.00(3.61)$ | $4.00(0.64)$ |
| Completeness $(\%)$ | $93.2(70.5)$ | $79.9(67.3)$ |
| Redundancy | $3.4(2.9)$ | $1.69(1.62)$ |
| No. reflections | 8490 | 23422 |
| Wilson B-factor $\left(\AA^{2}\right)$ | 22.1 | 17.6 |
| Refinement |  |  |

$\mathrm{R}_{\text {work }} / \mathrm{R}_{\text {free }} \quad 0.2359 / 0.2713 \quad 0.2642 / 0.3106$

| No. atoms |  |  |
| :--- | :--- | :--- |
| Protein | 1183 | 2407 |
| Water | 139 | 221 |

R.m.s. deviations

Bond lengths $0.009 \quad 0.004$
( $\AA$ )
Bond angles $\left(^{\circ}\right) \quad 1.095 \quad 0.783$

| Ramachandran plot |  |  |
| :--- | :--- | :--- |
| statistics (\%) |  | $99 \%$ |
| Most favourable | $96 \%$ | $0 \%$ |
| Disallowed | $1 \%$ | 0.1 M Sodium citrate tribasic dehydrate pH5.6, |
| Crystallization | 0.2 M Magnesium sulfate heptahydrate, | $20 \%$ 2-Propanol, 20\%PEG 4000 |
| condition | $20 \%$ PEG 3350 |  |


| Crystal | K63-linked di-Ub | K11/K63-branched tri-Ub |
| :--- | :--- | :--- |
| Data collection |  |  |
| Space group | $\mathrm{P} 2_{1} 2_{1} 2$ | $\mathrm{P} 2_{1} 2_{1} 2_{1}$ |
| Unit cell | $40.456,79.268,35.819 ;$ | $50.073,52.911,58.230 ;$ |
|  | $90.000,90.000,90.000$ | $90.00,90.00,90.00$ |
| Resolution $(\AA)$ | $36 \sim 1.55(1.61 \sim 1.55)$ | $31 \sim 1.84(1.91 \sim 1.84)$ |
| $\mathrm{R}_{\text {merge }}$ | $0.076(2.126)$ | $0.124(0.490)$ |
| $\mathrm{I} / \sigma \mathrm{I}$ | $11.07(0.57)$ | $8.36(0.65)$ |
| Completeness $(\%)$ | $98.9(98.4)$ | $77.1(10.6)$ |
| Redundancy | $6.20(5.58)$ | $3.2(1.7)$ |
| No. reflections | 17087 | 10779 |
| Wilson B-factor $\left(\AA^{2}\right)$ | 25.6 | 24.8 |
| Refinement |  |  |

$\mathrm{R}_{\text {work }} / \mathrm{R}_{\text {free }} \quad 0.2490 / 0.2528 \quad 0.2208 / 0.2452$

No. atoms

| Protein | 1197 | 1179 |
| :--- | :--- | :--- |
| Water | 99 | 131 |

R.m.s. deviations

Bond lengths $0.006 \quad 0.003$
(Å)
Bond angles $\left({ }^{\circ}\right) \quad 0.999 \quad 0.753$

| Ramachandran plot statistics (\%) |  |  |
| :---: | :---: | :---: |
| Most favourable | 99\% | 99\% |
| Disallowed | 1\% | 0\% |
| Crystallization condition | 0.1M TRIS hydrochloride $\mathrm{pH} 8.5,2.0 \mathrm{M}$ Ammonium phosphate monobasic | 0.2M $\mathrm{MgSO}_{4}, 20 \% \mathrm{PEG} 3350,4 \mathrm{mM} \mathrm{CdCl} 2$, |
|  |  | pH6.0 |


| Crystal | linear tri-Ub | linear tetra-Ub |
| :--- | :--- | :--- |
| Data collection |  |  |
| Space group | P 1 | $\mathrm{P} 2_{1}$ |
| Unit cell | $30.955,31.123,32.180 ;$ | $29.436,56.462,38.532 ;$ |
|  | $71.54,77.42,89.43$ | $90.000,90.915,90.000$ |
| Resolution $(\AA)$ | $30 \sim 1.80(1.87 \sim 1.80)$ | $24 \sim 2.21(2.29 \sim 2.21)$ |
| $\mathrm{R}_{\text {merge }}$ | $0.207(0.426)$ | $0.150(0.288)$ |
| $\mathrm{I} / \sigma \mathrm{I}$ | $4.2(2.4)$ | $7.73(1.36)$ |
| Completeness $(\%)$ | $74.7(14.5)$ | $56.2(7.1)$ |
| Redundancy | $1.7(1.3)$ | $3.5(1.4)$ |
| No. reflections | 6648 | 3575 |
| Wilson B-factor $\left(\AA^{2}\right)$ | 6.5 | 29.2 |
| Refinement |  |  |

$\mathrm{R}_{\text {work }} / \mathrm{R}_{\text {free }} \quad 0.2258 / 0.3148 \quad 0.2220 / 0.3222$

| No. atoms |  |  |
| ---: | :--- | :--- |
| Protein | 1175 | 1160 |
| Water | 105 | 35 |

R.m.s. deviations
Bond lengths $0.008 \quad 0.012$
(Å)
Bond angles ( ${ }^{\circ}$ ) 1.116
1.337

| Ramachandran plot |  |  |
| :--- | :--- | :--- |
| statistics (\%) |  | $96 \%$ |
| $\quad$ Most favourable | $99 \%$ | $1 \%$ |
| Disallowed | $0 \%$ | 0.2 M Sodium acetate trihydrate, |
| Crystallization | $0.2 \mathrm{M} \mathrm{MgSO} 4,20 \%$ PEG3350, 4 mM | $20 \% \mathrm{PEG} 3350, \mathrm{pH} 8.0$ |
| condition | $\mathrm{CdCl} 2, \mathrm{pH} 6.0$ |  |

