Stable Bio-oil Production from Proteinaceous Cyanobacteria: Tail Gas Reactive Pyrolysis of Spirulina Supporting Information

Bruna M. E. Chagas^{1,2}, Charles A. Mullen², Christina Dorado², Yassen Elkasabi², Akwasi A. Boateng*², Marcus A.F. Melo¹, Carlos H.Ataíde³

¹Universidade Federal do Rio Grande do Norte, CT, Departamento de Engenharia Química, 59078-970, Natal/RN, Brazil m

²USDA-ARS, Eastern Regional Research Center, 600 E. Mermaid Lane, Wyndmoor, PA 19038, United States

³Universidade Federal de Uberlândia (UFU), Faculdade de Engenharia Quimica, Campus Santa Mônica, Bloco 1K, 38408-144 Uberlândia, MG, Brazil

*Corresponding author. E-mail: akwasi.boateng@ars.usda.gov

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Table S1

Ultimate analysis (wt.%, daf)		Proximate analysis (wt.	Proximate analysis (wt.%)		
С	48.05	Moisture	5.60		
Н	7.04	Ash	7.94		
Ν	10.74	Volatile matter	79.38		
S	0.72	Fixed Carbon	5.28		
O^a	33.45	HHV (MJ/kg)	22.56		

Characterization of Spirulina used in this study.

Compounds	% Area		Functional group/structure	
	Regular ^e	TGRP ^f	group/structure	
Nitriles ^a	5.8	9.5 - 11.4	R−C≡N	
Pyrroles ^b	4.0	4.2 - 4.3	NH	
Amides ^c	13.0	9.7 – 11.0	R-C ^O NH ₂	
Indoles ^d	8.2	10.3- 11.9		
2,5-dimethylpiperazine	8.5	Nd		
2-Aminopyridine	0.4	1.0	NH ₂	
1,4-dimethylpyrazole	0.7	nd	N-N	
2,2,6,6 - Tetramethyl-4-piperidone	3.9	nd	↓ ↓ ↓ ↓ ↓	
2,5-Pyrrolidinedione	2.3	nd	o=	
5-(2-methylpropyl)-2,4- imidazoledinedione	0.8	nd		
5-oxoproline	2.5	nd		

Identified nitrogen containing compounds in regular and TGRP pyrolysis of spirulina (% area)

^a 4-methyl-pentanitrile, 3-methyl-butanitrile, benzonitrile, benzyl nitrile, benzenepropanenitrile, hexadecanenitrile ^b pyrrole, 3-methyl-1H-Pyrrole, hexahydropyrrolo[1,2-α]pyrazine-1,4-dione ^c acetamide, propanamide, hexanamide, hexadecanamide, 3-methyl-butanamide ^d Indole, 3-methyl-1H-indole

Table S2

^e Results from a single experiment ^fExperiments conducted in duplicate

NMR Analysis of ESP Oils and Condensers' Organic Fraction

Carbon 13 (¹³C) Nuclear Magnetic Resonance Spectroscopy (NMR) and Proton (¹H) NMR were conducted on the ESP oils from the regular and tail-gas reactive pyrolysis of Spirulina as well as the condensers' organic fraction from the TGRP of Spirulina. Solution-state NMR spectra were recorded at 9.4 T on a Varian Inova NMR spectrometer (Palo Alto, CA) using a 5 mm dual broad-band probe equipped with z-axis pulsed field gradients or on a 14 T Agilent VNMRS NMR spectrometer (Santa Clara, CA) using a 5 mm One NMR probe with z-axis pulsed field gradients. All spectra were acquired at 40 °C in CD₃OD. The ¹H NMR spectra, at 400 MHz, had a spectral width of 13 ppm and were acquired with a 45° pulse angle with a 6 s relaxation delay and were referenced to the sodium salt of 3-(trimethylsilyl)propionic acid-d₄ (TSP). All ¹³C NMR spectra had a sprectral width of 250 ppm, were acquired using a 45° pulse angle, inverse-gating, and were referenced to the solvent ¹³C peak. Reasonable signal-to-noise was achieved with 15,000–70,000 transients, utilizing a 4 or 6 s relaxation delay to provide adequate recovery of the signal for integration purposes. All data processing was performed using Spinworks4 (Version 4.1.0.0) (Copyright 2015, Kirk Marat, University of Manitoba (ftp://davinci.chem.umanitoba.ca/pub/marat/SpinWorks/).



Figure S1. ¹³C NMR of ESP oil from the Regular Pyrolysis of Spirulina recorded at a frequency of 150 MHz in Deuterated Methanol



Figure S2. ¹³C NMR of ESP oil from the TGRP of Spirulina recorded at a frequency of 150 MHz in Deuterated Methanol



Figure S3. ¹³C NMR of the Condensers' Organic Fraction from the TGRP of Spirulina recorded at a frequency of 150 Mz in Deuterated Methanol



S4. ¹H NMR of the ESP oil from the Regular Pyrolysis of Spirulina recorded at a frequency of 600 MHz in Deuterated Methanol



Figure S5. ¹H NMR of the ESP oil from the TGRP of Spirulina recorded at a frequency of 600 MHz in Deuterated Methanol



Figure S6. ¹H NMR of the Condensers' Organic Fraction from the TGRP of Spirulina recorded at a frequency of 600 MHz in Deuterated Methanol



Α



Figure S7. GC/MS Chromatograms of distillate fractions from Spirulina TGRP bio-oil. a. propenitrile b. benzene c. C4-nitriles d. toluene e. *p*-xylene f. pyrrole g. styrene h. pentenitrile i. dimethyl pyridines j. C3-alkyl benzenes k. indene l. benzonitrile m. phenol n. cresols o. indole p. hexadecane q. hexadecane nitrile r. hexdecanamide (A)Retention time 6-40 min (B) Retention time 40-85 min



Figure S8 - (a) Spirulina TGRP bio-oil distillation fractions, with fraction 3 (b) exhibiting triphasic behavior. The top organic phase consisted primarily of hexadecane