Crown Ethers as Building Blocks for Carbohydrate Receptors

Monika Mazik,* Matthias Kuschel, and Willi Sicking

		Page
1.	Syntheses of the receptors 1 and 2.	S2
1.1	Synthesis of the receptor 1 .	S2
1.2	Synthesis of the receptor 2 .	S 3
2.	¹ H NMR titration of 1 with β-glucopyranoside 3a (chemical shifts of the CH_3 and the pyridine CH protons).	S4
3.	¹ H NMR titration of 1 with α-glucopyranoside 4a (chemical shifts of the CH_3 and the pyridine CH protons).	S4
4.	¹ H NMR titration of 1 with β- and α-glucopyranoside, 3a and 4a (typical titration	S 5
	curves).	S 6
5.	Extraction of β - and α -methyl-glucopyranoside (3b and 4b) from the solid state	
	into a CDCl ₃ -solution of receptor 1 .	S 6
6.	¹ H NMR Titration of receptor 2 with α-glucopyranoside 4a (Chemical shifts of the	
	CH ₃ and CH resonances, protons C, E, H, and I).	S 7
7.	¹ H NMR titration of 2 with α-glucopyranoside 4a (typical titration curve and a	
	representative mole ratio plot).	

1 Syntheses of the receptors 1 and 2.

1.1 Synthesis of the receptor 1.

The synthesis of **1** started from 1,3,5-tris(bromomethyl)-2,4,6-trimethyl-benzene (**5**), which was converted into the compound **7** *via* reaction with two equivalents of 2-amino-4,6-dimethyl-pyridine (**6**), followed by the reaction with one equivalent of 2-aminomethyl-15-crown-5 (**8**), as shown in Scheme S1.

SCHEME S1

$$\begin{array}{c} \text{CH}_3 \\ \text{Br} \\ \text{Br} \\ \text{Br} \\ \text{Br} \\ \text{Br} \\ \text{Br} \\ \text{H}_2\text{N} \\ \text{N} \\ \text{CH}_3 \\ \text{CH}_3\text{CN/THF} \\ \text{K}_2\text{CO}_3 \\ \text{N} \\ \text{N} \\ \text{CH}_3\text{CN/THF} \\ \text{K}_2\text{CO}_3 \\ \text{N} \\ \text{N} \\ \text{CH}_3 \\ \text{N} \\ \text{CH}_3 \\ \text{CH}_$$

Compound 7. To a mixture of 1,3,5-tris(bromomethyl)-2,4,6-trimethyl-benzene (3.00 g, 6.80 mmol) and K₂CO₃ (1.88 g, 13.60 mmol) in CH₃CN/THF (1:1 v/v; 40 mL) was added dropwise a CH₃CN (10 mL) solution of 2-amino-4,6-dimethyl-pyridine (1.66 g, 13.60 mmol). The mixture was stirred at room temperature for 72 h. After filtration and evaporation of solvents, the crude product was purified by column chromatography (ethyl acetate/toluene, 1:3 v/v). Yield 30 %. M.p. 77-78 °C. ¹H-NMR (400 MHz, CDCl₃) δ = 1.22 (t, 3H, J = 7.5 Hz), 1.29 (t, 6H, J = 7.5 Hz), 2.24 (s, 6H), 2.36 (s, 6 H), 2.73 (q, 2H, J = 7.5 Hz), 2.85 (q, 4H, J = 7.5 Hz), 4.23 (t, 2H, J = 4.2 Hz), 4.37 (d, 4H, J = 4.2 Hz), 4.62 (s, 2 H), 6.10 (s, 2 H), 6.35 (s, 2 H). ¹³C-NMR (100 MHz, CDCl₃) δ = 16.4, 16.7, 21.1, 22.8, 23.0, 24.1, 29.6, 40.5, 103.6, 113.9, 131.9, 133.4, 143.8, 144.9, 148.9, 156.5, 158.0. HR-MS calcd for C₂₉H₃₉BrN₄ 5232.2353; found: 522.2360. R_f = 0.31 (ethyl acetate/toluene, 1:3 v/v).

Receptor 1. To a mixture of compound **7** (262.5 mg, 0.50 mmol) and K_2CO_3 (69.3 mg, 0.50 mmol) in CH₃CN/THF (1:1 v/v; 20 mL) was added dropwise a CH₃CN (10 mL) solution of 2-aminomethyl-15-crown-5 (125 mg, 0.50 mmol). The mixture was stirred at room temperature for 48 h. After filtration and evaporation of solvents, the crude product was purified by column chromatography (chloroform/methanol 7:1 v/v). Yield 62 %. M.p. 57-58 °C. ¹H-NMR (500 MHz, CDCl₃, [**1**] = 0.9 mM) δ = 1.23 (t, 9H, J = 7.6 Hz), 2.23 (s, 6 H), 2.35 (s, 6 H), 2.77 (m, 7 H), 3.70 (m, 23 H), 4.25 (br. s, 2 H) 4.36 (d, 4H, J = 4.0 Hz), 6.08 (s, 2 H), 6.33 (s, 2 H). ¹³C-NMR (100 MHz, CDCl₃) δ = 16.8, 16.9, 21.1, 22.8, 24.1, 40.6, 47.8, 52.1, 70.1, 70.4, 70.5, 70.6, 70.8, 70.9, 71.0, 72.9, 78.7, 103.5, 113.8, 132.7, 142.9, 143.2, 148.8, 156.5, 158.2, 162.7. HR-MS calcd for $C_{40}H_{61}N_5O_5$ 691.4667; found: 691.4671. R_f = 0.10 (chloroform/methanol, 7:1 v/v).

1.2 Synthesis of the receptor 2.

To a solution of benzene-1,3,5-tricarbonyl chloride (187.4 mg, 0.70 mmol) in dry CH₂Cl₂ (10 ml) was added a CH₂Cl₂ (10 mL) solution of 4'-aminobenzo-15-crown-5 (200 mg, 0.70 mmol) and triethylamine (0.1 mL). The reaction mixture was stirred at room temperature for 45 min and then a CH₂Cl₂ (10 mL) solution of 2-amino-4,6-dimethylpyridine (172.5 mg, 1.41 mmol) was added. The mixture was stirred at room temperature for 72 h. The organic phase was washed three times with water (3 x 10 mL), dried, and the solvent was evaporated. The crude product was was purified by column chromatography (chloroform/methanol 7:1 v/v). Yield 41 %. M.p. 200 °C (decomp.). 1 H-NMR (500 MHz, CDCl₃; [2] = 0.9 mM) δ = 2.38 (s, 6H), 2.45 (s, 6H), 3.77 (br.s, 8H), 3.92 (m, 4H), 4.16 (m, 2H), 4.21 (m, 2H), 6.82 (s, 2H), 6.90 (d, 1H, J = 8.5 Hz), 7.10 (dd, 1H, J = 8.5/2.2 Hz), 7.47 (d, 1H, J = 2.2 Hz), 7.99 (s, 1H), 8.02 (s, 2H), 8.63 (s, 2H), 8.65 (s, 1H), 8.67 (s, 2H). 13 C-NMR (50 MHz, CDCl₃) δ = 21.3, 23.7, 68.7, 69.4, 69.5, 69.6, 70.4, 70.5, 70.9, 107.3, 112.0, 113.1, 114.6, 121.0, 128.9, 129.1, 131.9, 135.4, 136.2, 149.2, 150.3, 150.4, 156.6, 163.6, 163.9. HR-MS calcd for C_{37} H₄₁N₅O₈ 683.2950; found: 683.2958. R_f = 0.66 (chloroform/methanol 7:1 v/v).

¹H NMR titration of **1** with β -glucopyranoside **3a** (chemical shifts of the CH₃ and the pyridine CH protons).

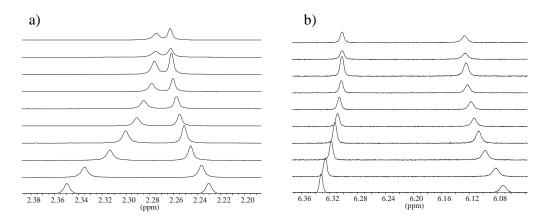


FIGURE S1. Partial ¹H NMR spectra (500 MHz, CDCl₃, 25 °C) of **1** after addition of (from bottom to top) 0, 0.27, 0.55, 0.83, 1.11, 1.38, 2.08, 2.77, 3.47 and 4.16 equiv of **3a** ([**1**] = 0.87 mM). (a) Chemical shifts of the CH₃ resonances. (b) Chemical shifts of the pyridine CH resonances.

3. 1 H NMR titration of 1 with α-glucopyranoside 4a (chemical shifts of the CH₃ and the pyridine CH protons).

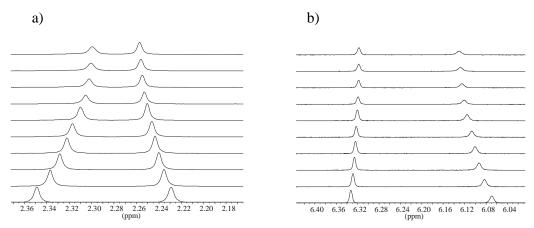


FIGURE S2. Partial ¹H NMR spectra (500 MHz, CDCl₃, 25 °C) of **1** after addition of (from bottom to top) 0, 0.44, 0.88, 1.33, 1.77, 2.66, 3.55, 4.44, 5.33 and 6.22 equiv of **4a** ([**1**] = 0.85 mM). (a) Chemical shifts of the CH₃ resonances. (b) Chemical shifts of the pyridine CH resonances.

4. ¹H NMR titration of **1** with β- and α-glucopyranoside, **3a** and **4a** (typical titration curves).

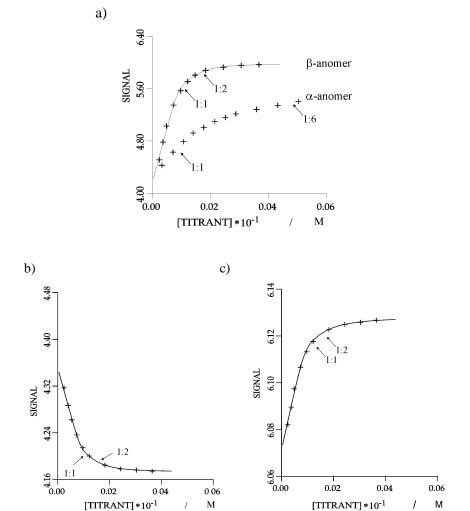


FIGURE S3. Plot of the downfield chemical shifts of the NH^A, resonances of **1** as a function of added β- and α-glucopyranoside, **3a** and **4a** (a) ([**1**] = 0.87 mM; Equiv of **3a** = 0.00-3.47; Equiv of **4a** = 0.00-6.22). Plot of the observed upfield chemical shifts of the CH_2^C (b) and downfield chemical shifts of the pyr-CH resonances (c) of **1** as a function of added β-glucopyranoside **3a** ([**1**] = 0.87 mM; Equiv of **3a** = 0.00-4.18). The [receptor]:[sugar] ratio is marked.

5. Extraction of β - and α -methyl-glucopyranoside (3b and 4b) from the solid state into a CDCl₃-solution of receptor 1.

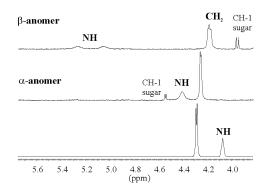


FIGURE S4. ¹H NMR spectra showing the NH and the CH_2 protons of receptor **1** before and after the extraction of solid methyl-α-glucopyranoside **4b** or methyl-β-glucopyranoside **3b** by a CDCl₃-solution of receptor **1** (0.9 mM).

6 ¹H NMR Titration of receptor **2** with α-glucopyranoside **4a** (chemical shifts of the CH₃ and CH resonances, protons C, E, H, and I).

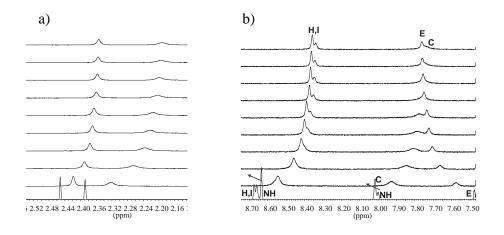


FIGURE S5. Partial ¹H NMR spectra (500 MHz, CDCl₃, 25 °C) of **2** after addition of (from bottom to top) 0, 0.20, 0.40, 0.60, 0.80, 1.00, 1.50, 2.00, 2.50 and 3.01 equiv of **4a** ([1] = 0.87 mM). (a) Chemical shifts of the CH₃ resonances. (b) Chemical shifts of the CH resonances, protons C, E, H, and I.

7. 1 H NMR titration of 2 with α -glucopyranoside 4a in CDCl₃ (typical titration curve and a representative mole ratio plot).

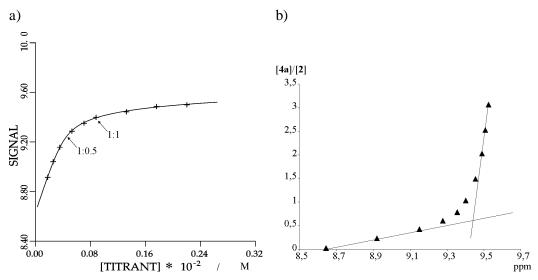


FIGURE S6. (a) Plot of the observed (\mathbf{x}) and calculated (—) downfield chemical shifts of the NH^A resonances of $\mathbf{2}$ as a function of added α -glucopyranoside $\mathbf{4a}$; [$\mathbf{2}$] = 0.88 mM; Equiv of $\mathbf{4a}$ = 0, 0.20, 0.30, 0.40, 0.60, 0.80, 1.00, 1.50, 2.00, 2.50. The [receptor]:[sugar] ratio is marked. (b) Mole ratio plot (analysis of the shifts of the NH^A of $\mathbf{2}$).