

## Multiplexed Detection of Epigenetic Markers Using Quantum Dot (QD)-Encoded Hydrogel Microparticles

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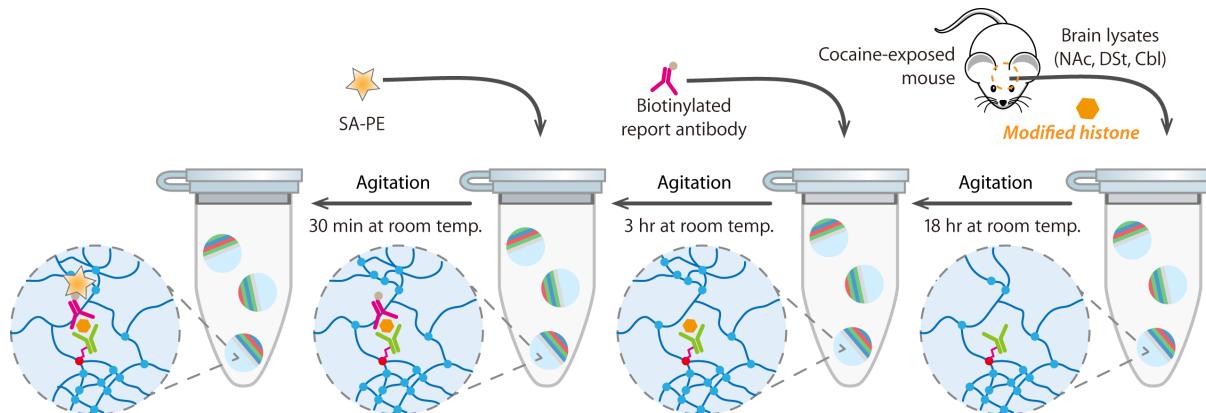
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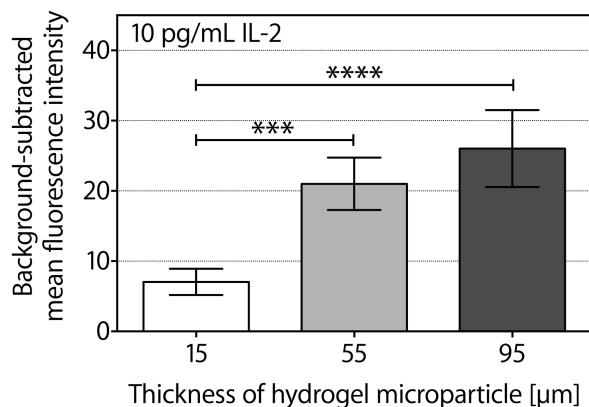
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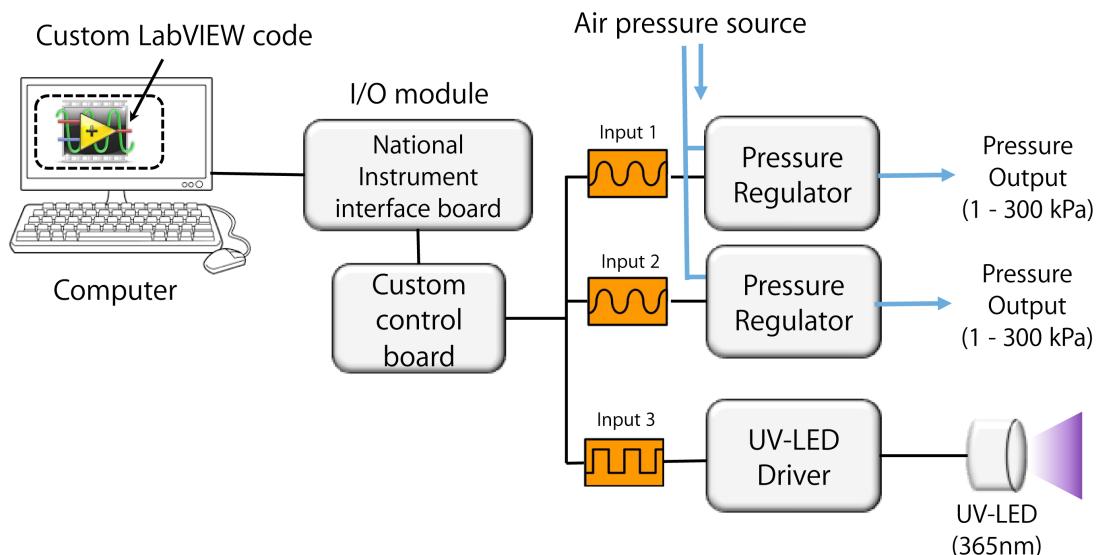
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**Figure S1.** Detailed assay scheme for multiplexed detection of modified histones from mouse brain lysates. 1) incubation with a brain lysate per mouse containing 3 modified histones (NAc, DSt, or Cbl), 2) incubation with a biotinylated report antibody, and 3) incubation with a reporter fluorophore of SA-PE.



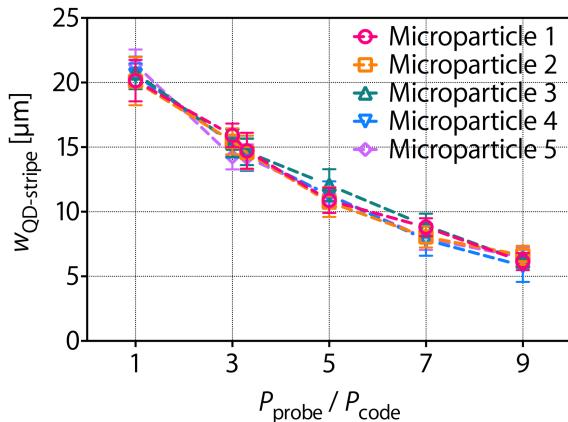
**Figure S2.** Optimization of thickness of hydrogel microparticles. Bar graphs presenting background-subtracted mean fluorescence intensity of IL-2 (10 pg/mL) from hydrogel microparticles with thickness of 15 (white), 55 (gray), and 95 (dark gray)  $\mu\text{m}$ . 5 hydrogel microparticles were analyzed for each data point. Error bars represent standard deviation. \*\*\*\* and \*\*\* denote significant differences of  $p < 0.0001$  and  $p < 0.001$ , respectively.



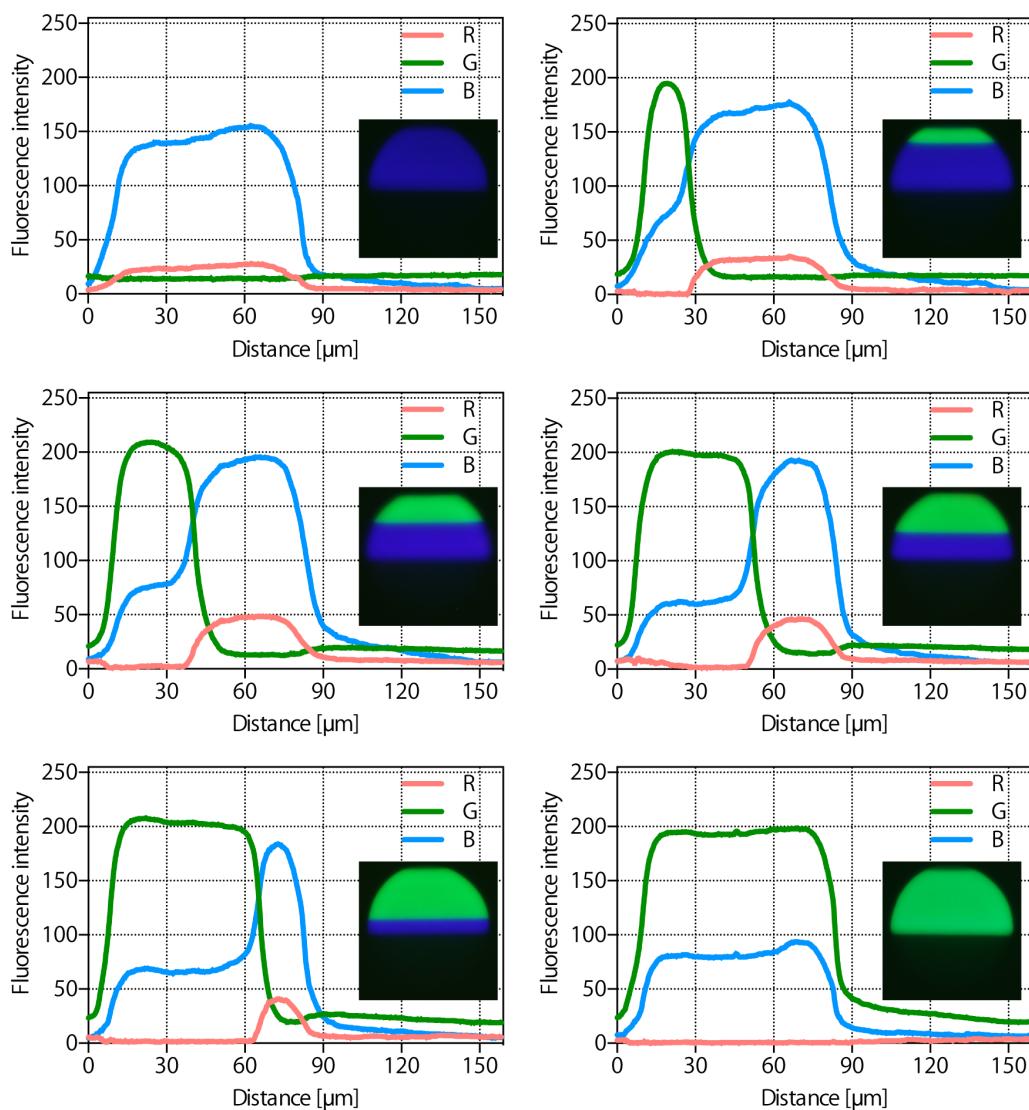
**Figure S3.** Block diagram of circuit design to control both pressure regulators and UV LED. Computer sends commands to a custom control board through the interface board to generate signals. Two analog signals and one digital signal are generated to control pressure regulators and UV-LED driver, respectively.

**Table S1.** Width of QD stripes by controlling pressure to probe and code regions

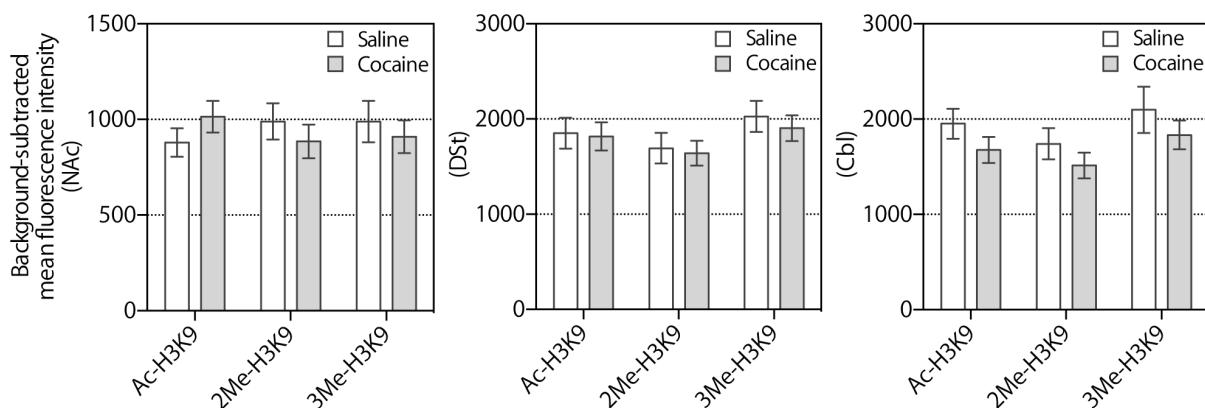
$P_{\text{probe}}$	$P_{\text{code}}$	$P_{\text{probe}}/P_{\text{code}}$	$w_{\text{QD-stripe,mean}}$ [ $\mu\text{m}$ ]	$w_{\text{QD-stripe,SD}}$ [ $\mu\text{m}$ ]	Coefficient of variation (CV) [%]
30	30	1	20.6	1.37	6.64
45	15	3	15.2	1.08	7.09
100	30	3.3	14.5	1.12	7.75
50	10	5	11.3	1.05	9.31
50.25	7.5	7	8.31	0.97	11.7
104	6	9	6.25	0.81	12.9



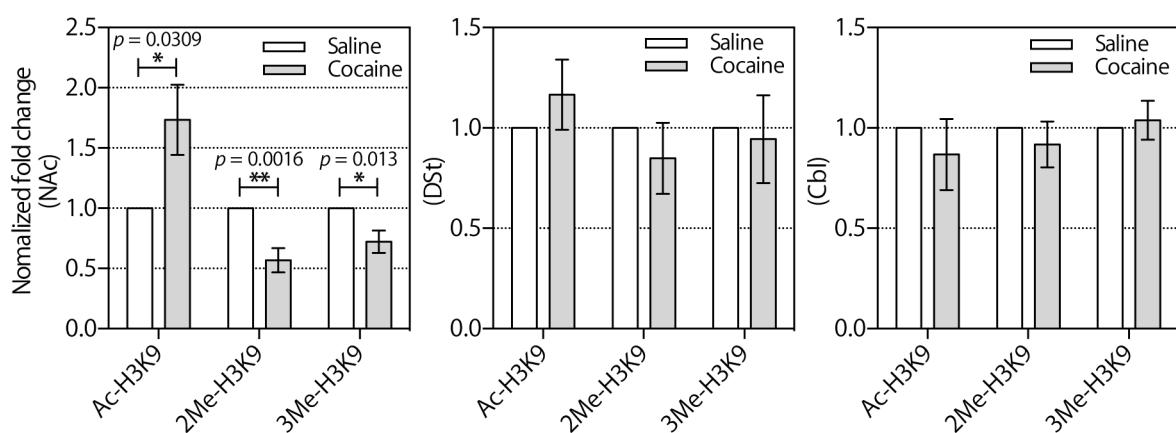
**Figure S4.** Inter- and intra-particle variations of width of QD-stripes by controlling ratio of pressure to probe and code regions. Red circle, orange square, green triangle, blue inverse triangle, and purple lozenge indicate mean of 5 QD-stripes per representative microparticle 1, 2, 3, 4, and 5, respectively. Error bars represent standard deviation.



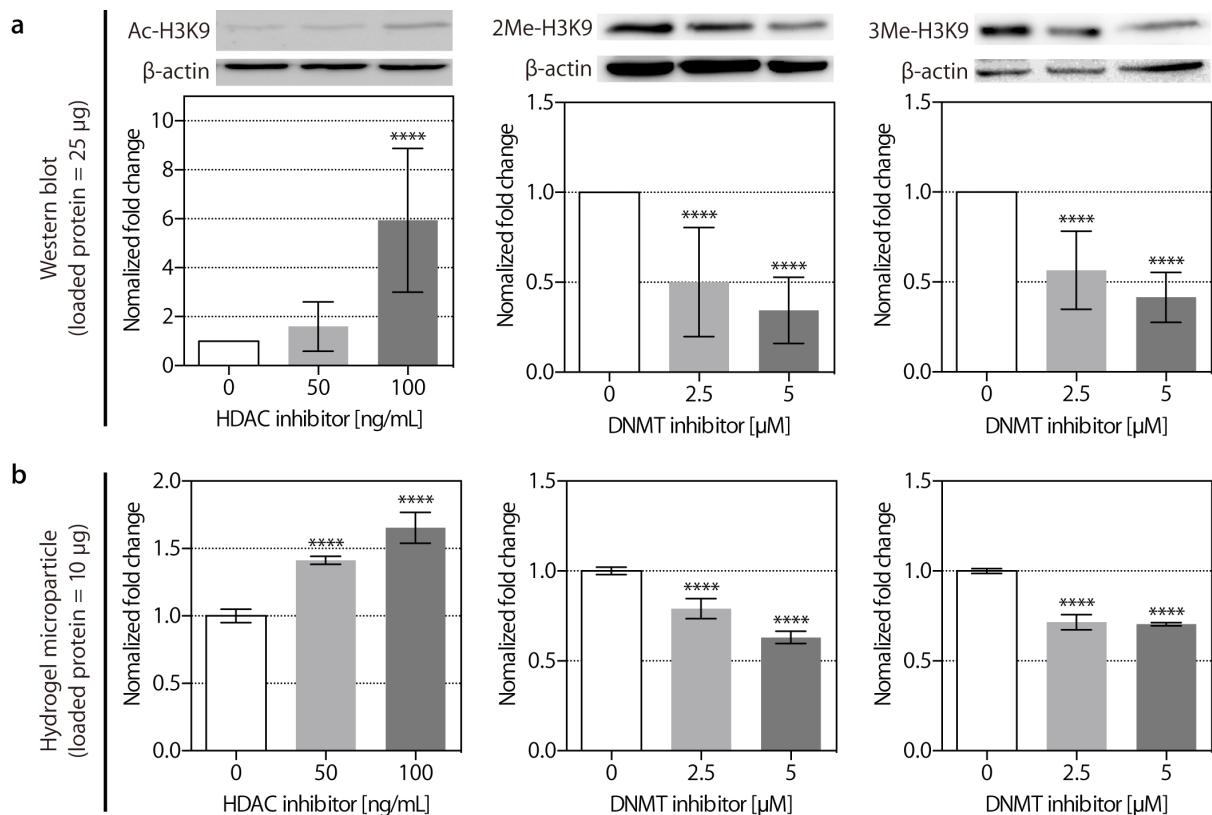
**Figure S5.** Spatial intensity profiles of red, green, and blue channels from representative microparticles (inset fluorescence micrographs) encoded with identical green and blue stripes consecutively located in the code region.



**Figure S6.** Representative example displaying detection of histone H3K9 modifications from mixture of multiple brain lysates. Bar graphs presenting background-subtracted mean fluorescence intensity of Ac-H3K9, 2Me-H3K9, and 3Me-H3K9 simultaneously detected from NAc (left), DSt (middle), and Cbl (right) of mice exposed to saline or cocaine for 7 days. White and gray bars are background-subtracted mean fluorescence intensity from saline- and cocaine-exposed groups, respectively. Error bars represent standard error of mean.



**Figure S7.** Western blot analysis from 50 µg of total proteins in the brain lysates of cocaine-exposed mice. Normalized fold changes in H3K9 modifications from NAc (left), DSt (middle), and Cbl (right). Cocaine-exposed mice display significant elevation in Ac-H3K9 and reduction in Me2- & Me3-H3K9 in response to cocaine. In both DSt and Cbl, no significant changes were observed. n ≥ 4 mice per group.



**Figure S8.** Quantification of histone H3K9 modifications from lysates of HEK cells treated with HDAC inhibitor or DNMT inhibitor. **a.** Western blot analysis with 25 µg of total proteins. HDAC inhibitor dose-dependently increased Ac-H3K9 status, while DNMT inhibitor also dose-dependently reduced 2Me-H3K9 and 3Me-H3K9. \*\*\* denotes  $p < 0.0001$ . n = 3 per group. **b.** Plots of normalized fold changes for hydrogel microparticle-based assay with the identical lysates (10 µg of total proteins). \*\*\* denotes  $p < 0.0001$ .

**Video S1.** Video clip showing microfluidic capture of hydrogel microparticles.

### **MATLAB code to rotate and crop acquired images**

```
clc; %Clear Command Window
clear all; %Remove items from workspace, freeing up system memory
close all;

%Open standard dialog box for selecting directory
dname_fl = uigetdir('any desired location on a computer to find images' );

%List all tif files
cd(dname_fl);
img_path_fl=strcat(dname_fl,'*.TIF');
file_fl = dir(img_path_fl);
file_num_fl = length(file_fl);
file_name_fl = {file_fl.name};
file_date_fl = {file_fl.date};

file_num_fl

clims_fl=[100 700];
rot_degree= [ ];
upperleftcorner = [ ];

img_w=620;
img_h=620;

probe_w=550;
probe_h=250;

for i = 1:file_num_fl
    %rotate image=====
    figure(1)
    subplot(2, 1, 1)
    cd(dname_fl);
    img_fl=imread(file_name_fl{i});
    imagesc(img_fl, clims_fl)
    colormap(summer)
    axis equal
    grid on
    grid minor
    title(char(file_name_fl{i}))

    rot_degree_index=0;

    while rot_degree_index == 0
        prompt_fl={'Enter rotation degree:'};
        name_fl='Image rotation degree';

```

```

numlines_fl=1;
defaultanswer_fl={'90'};

options.Resize='on';
options.WindowStyle='normal';
options.Interpreter='tex';

rot_degree_dialog=inputdlg(prompt_fl,name_fl,numlines_fl,defaultanswer_fl,options);

img_rot_fl=imrotate(img_fl,str2num(rot_degree_dialog{1}));
%figure(2)
subplot(2, 1, 2)
imagesc(img_rot_fl,clims_fl)
colormap(summer)
axis equal
grid on
grid minor
title(char(file_name_fl{i}))

selectedButton = uigetpref(
    'mygraphics',...
    % Group
    'savebeforeclosing',...
    % Preference
    'Closing Rotation of Particle',...
    % Window title
    {'Are you satisfied?'},...
    % Values and button strings
    {'Yes','No','Pass'},...
    % Default choice
    'DefaultButton','No');
switch selectedButton
    case 'no'
        rot_degree_index=0;
    case 'yes'
        rot_degree_index=1;
    case 'pass'
        rot_degree_index=2;
end

if rot_degree_index==0
    continue
elseif rot_degree_index ==1
    rot_degree = [rot_degree; str2num(rot_degree_dialog{1})];
    break
elseif rot_degree_index ==2
    break
else
end
end

%crop rotated image=====

```

```

figure(2)
%subplot(2, 1, 1)
imagesc(img_rot_fl, clims_fl)
colormap(summer)
axis equal
grid on
grid minor
title(char(file_name_fl{i}))

crop_point_index=0;

while crop_point_index == 0
    figure(2)
    %subplot(2, 1, 1)
    p1=impoint
    temp_upperleftcorner=getPosition(p1);

    img_crop_fl=imcrop(img_rot_fl, [temp_upperleftcorner(1) temp_upperleftcorner(2) img_w
img_h]);
    figure(3)
    %subplot(2, 1, 2)
    imagesc(img_crop_fl, clims_fl)
    colormap(summer)
    axis equal
    grid on
    grid minor
    title(char(file_name_fl{i}))

selectedButton = uigetpref...
'mygraphics',...
'Group'
'savebeforeclosing',...
'Preference'
'Upper left corner of particle',...
'Window title'
{'Are you satisfied?'},...
{'Yes','No','Pass'},...
'Values and button strings'
'DefaultButton','No'); ...
'Default choice'
switch selectedButton
    case 'no'
        crop_point_index=0;
    case 'yes'
        crop_point_index=1;
    case 'pass'
        crop_point_index=2;
end

if crop_point_index==0
    delete(p1)
    continue
elseif crop_point_index ==1

```

```

    upperleftcorner = [upperleftcorner; temp_upperleftcorner];
    break
elseif crop_point_index == 2
    break
else
end

end

figure(3)
imsave

%crop rotated image=====
figure(3)

imagesc(img_crop_fl, clim_s_fl)
colormap(summer)
axis equal
grid on
grid minor
title(char(file_name_fl{i}))

crop_point_index=0;

while crop_point_index == 0
    figure(3)

    p1=impoint
    temp_upperleftcorner=getPosition(p1);

    img_crop_fl_probe=imcrop(img_crop_fl, [temp_upperleftcorner(1) temp_upperleftcorner(2)
probe_w probe_h]);
    figure(4)

    imagesc(img_crop_fl_probe, clim_s_fl)
    colormap(summer)
    axis equal
    grid on
    grid minor
    title(char(file_name_fl{i}))

selectedButton = uigetpref...
'mygraphics',...
    % Group
'savebeforeclosing',...
    % Preference
'Upper left corner of particle',...
    % Window title
{'Are you satisfied?'},...

```

```

{'Yes','No','Pass'},...    % Values and button strings
'DefaultButton','No');      % Default choice
switch selectedButton
  case 'no'
    crop_point_index=0;
  case 'yes'
    crop_point_index=1;
  case 'pass'
    crop_point_index=2;
end

if crop_point_index==0
  delete(p1)
  continue
elseif crop_point_index ==1
  upperleftcorner = [upperleftcorner; temp_upperleftcorner];
  break
elseif crop_point_index ==2
  break
else
end
end

figure(4)
imsave

end

close all

```

## MATLAB code to analyze fluorescence intensity from (rotated and cropped) images

```
clc; %Clear Command Window
clear all; %Remove items from workspace, freeing up system memory
close all;

%Open standard dialog box for selecting directory
dname = uigetdir('any desired location on a computer to find images');

%List all tif files
cd(dname);
img_path=strcat(dname,'*.tif');
file = dir(img_path);
file_num = length(file);
file_name = {file.name};
file_date = {file.date};

file_num

%Determine row & column numbers for subplot
prompt1={'Enter row number:', 'Enter column number:'};
name1='Input for Limits of Fluorescence Intensity';
numlines1=1;
defaultanswer1={'5','1'};

options.Resize='on';
options.WindowStyle='normal';
options.Interpreter='tex';

rowcolumn_dialog=inputdlg(prompt1,name1,numlines1,defaultanswer1,options);

subplotsize = [str2num(rowcolumn_dialog{1,1}) str2num(rowcolumn_dialog{2,1})];

%Determine limits of fluorescence intensity
prompt2={'Enter the lower limit of fluorescence intensity:', 'Enter the upper limit of fluorescence
intensity:'};
name2='Input for Limits of Fluorescence Intensity';
numlines2=1;
defaultanswer2={'200','13000'};

options.Resize='on';
options.WindowStyle='normal';
options.Interpreter='tex';

clims_dialog=inputdlg(prompt2,name2,numlines2,defaultanswer2,options);

clims = [str2num(clims_dialog{1,1}) str2num(clims_dialog{2,1})];
```

```

probe_sig=[];

%Generate color-mapped images with the fixed limits for all images
figure()
for i = 1:file_num
    img=imread(file_name{i});
    colormap(jet);
    subplot(subplotsize(1),subplotsize(2),i);
    imagesc(img,clims)
    axis equal
    axis off
    %colorbar
    probe_sig=[probe_sig; mean(mean(img))];
end

mean_probe_sig=mean(probe_sig)

stdev_probe_sig=std(probe_sig)

CV_probe_sig=stdev_probe_sig/mean_probe_sig*100

```