

# **C<sub>3</sub> symmetric Ti(IV) amine triphenolate complexes as sulfoxidation catalysts using aqueous hydrogen peroxide**

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**General Remarks:** <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded at 301 K on a Bruker AC-300 and Bruker AC-250 instruments. ESI-MS experiments of complexes **2a-c** were performed in a ESI-TOF Mariner™ Biospectrometry™ Workstation of Applied Biosystems by flow injection analysis using methanol as mobile phase. GC analysis were performed using a Shimadzu GC-2010 gas chromatograph with a FID detector and a capillary column EQUITY™-5 using dodecane as external standard. All chemicals were used as provided without further purifications. Dry solvents were purchased from Fluka, Ti(IV) tetraisopropoxide, thioanisol, dibutylsulfide, benzyl phenyl sulfide, *n*-butyl *p*-tolyl sulfide, *p*-methoxy thioanisol, *p*-nitro thioanisol and 35 % aqueous hydrogen peroxide from Aldrich. Ligands **3a-c** were synthesized as previously reported.<sup>1</sup> Ti(IV) complexes were always handle and stored in glovebox, with exception of complex **2c** which could be handle in open air.

## **Synthesis of Ti(IV) complexes 2a-c.**

Complexes **2a-c** were prepared in glovebox by mixing homogeneous solutions of the corresponding ligands **3a-c** (0.10 M) and Ti(Oi-Pr)<sub>4</sub> (0.18 M) in CHCl<sub>3</sub> or CDCl<sub>3</sub> in a 1:1 ratio, using 1,2-dichloroethane as internal standard, to a final concentration 0.01 M of the complex obtaining a bright yellow solution which was use for kinetic experiments and reactions without further purifications and without removing the three equivalents

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(1) Prins, L. J.; Mba, M.; Kolarović, A.; Licini, G. *Tetrahedron Lett.* **2006**, *47*, 2735-2738.

of *i*-PrOH released from the metal precursor. A quantitative conversion was observed in respect of the internal standard, DCE (3.78 ppm).

In all cases in the NMR spectra resonances relative to free *iso*-propanol released in the reaction were present: <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>): 4.04 (hept, 1H, *J* = 6.1 Hz, CHMe<sub>2</sub>), 1.22 (6H, d, *J* = 6.1 Hz, CH(CH<sub>3</sub>)<sub>2</sub>). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): δ 64.5 (CH), 25.1 (CH<sub>3</sub>).

**Complex 2a.** <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>): δ 7.16 (t, 3H, *J* = 7.8 Hz, ArH), 7.07 (d, 3H, *J* = 7.3 Hz, ArH), 6.82 (t, 3H, *J* = 7.3 Hz, ArH), 6.74 (d, 3H, *J* = 7.8 Hz, ArH), 5.14 (hept, 1H, *J* = 6.1 Hz, CHMe<sub>2</sub>), 3.49 (bs, 6H, NCH<sub>2</sub>), 1.55 (6H, d, *J* = 6.1 Hz, CH(CH<sub>3</sub>)<sub>2</sub>). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): δ 163.0 (C), 129.6 (CH), 129.3 (CH), 124.4 (C), 120.9 (CH), 116.2 (CH), 80.6 (CH, *i*-Pr), 58.7 (CH<sub>2</sub>), 25.1 (CH<sub>3</sub>, *i*-Pr). ESI-MS: 412.1608 (M+H<sup>+</sup>), calc. 412.1208.

**Complex 2b.** <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>): δ 7.08 (d, 3H, *J* = 7.3 Hz, ArH), 6.93 (d, 3H, *J* = 7.3 Hz, ArH), 6.75 (t, 3H, *J* = 7.3 Hz, ArH), 5.24 (hept, 1H, *J* = 6.1 Hz, CHMe<sub>2</sub>), 3.49 (bs, 6H, NCH<sub>2</sub>), 2.30 (9H, s, CH<sub>3</sub>), 1.55 (6H, d, *J* = 6.1 Hz, CH(CH<sub>3</sub>)<sub>2</sub>). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): δ 161.7 (C), 130.4 (CH), 127.2 (CH), 124.9 (C), 124.0 (C), 120.5 (CH), 80.1 (CH, *i*-Pr), 58.7 (CH<sub>2</sub>), 25.8 (CH<sub>3</sub>, *i*-Pr), 16.5 (CH<sub>3</sub>). ESI-MS: 454.1940 (M+H<sup>+</sup>) calc. 454.1498.

**Complex 2c:** <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>): δ 7.19 (d, 3H, *J* = 7.8 Hz, ArH), 6.96 (d, 3H, *J* = 6.5 Hz, ArH), 6.77 (t, 3H, *J* = 7.5 Hz, ArH), 5.24 (hept, 1H, *J* = 6.1 Hz, CHMe<sub>2</sub>), 3.94 (d, 3H, *J* = 13.0 Hz, NCH<sub>2</sub>), 2.89 (d, 3H, *J* = 13.0 Hz, NCH<sub>2</sub>), 1.51 (6H, d, *J* = 6.1 Hz, CH(CH<sub>3</sub>)<sub>2</sub>), 1.45 (s, 27H, *t*-Bu). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): δ 162.7 (C), 136.4 (C), 127.8 (CH), 126.4 (CH), 125.2 (C), 120.3 (CH), 80.2 (CH, *i*-Pr), 58.6 (CH<sub>2</sub>), 35.1 (C), 29.7 (CH<sub>3</sub>), 26.7 (CH<sub>3</sub>, *i*-Pr). ESI-MS: 580.3605 (M+H<sup>+</sup>), calc. 580.2906.

### **General procedure for monitoring the sulfoxidation reactions catalyzed by 2a-c using aqueous H<sub>2</sub>O<sub>2</sub> as oxidant (Table 1).**

A screw-cap NMR tube was charged with a solution of the corresponding *in situ* formed complex in CDCl<sub>3</sub> (0.003 mmol), solvent was removed under vacuum and then CD<sub>3</sub>OD, the internal standard (1,2-dichloroethane, DCE), 35% aqueous H<sub>2</sub>O<sub>2</sub> (0.3 mmol) and thioanisole (0.3 mmol) were added with a final volume of 0.6 ml. Concentrations of sulfide, sulfoxide and sulfone were determined by integration of the methyl group

signals: Ph-S-*Me* (2.4 ppm), Ph-SO-*Me* (2.8 ppm) and Ph-SO<sub>2</sub>-*Me* (3.1 ppm) in respect of the internal standard, DCE (3.78 ppm).

**General procedure for monitoring the sulfoxidation reactions catalyzed by 2c using aqueous H<sub>2</sub>O<sub>2</sub> as oxidant (Table 2).**

A screw-cap NMR tube was charged with a solution of the *in situ* formed complex 2c in CDCl<sub>3</sub>, solvent was removed under vacuum and then CD<sub>3</sub>OD followed by the internal standard (1,2-dichloroethane), 35% aqueous H<sub>2</sub>O<sub>2</sub> and thioanisole (4a) were added with final concentrations as reported in Table 2 to a final volume of 0.6 ml. The monitoring of the concentration of sulfide, sulfoxide and sulfone was made by integration of the methyl group signals: Ph-S-*Me* (2.4 ppm), Ph-SO-*Me* (2.8 ppm) and Ph-SO<sub>2</sub>-*Me* (3.1 ppm). Final yields were determined by quantitative GC analysis after complete H<sub>2</sub>O<sub>2</sub> consumption (iodometric test) in respect of the internal standard 1,2-dichloroethane (3.78 ppm).

**General procedure for sulfoxidation reactions catalyzed by 2c using aqueous H<sub>2</sub>O<sub>2</sub> as oxidant (Table 3).**

To a 1 ml solution of the corresponding thioethers 4a-f (0.5 mmol) and catalyst 2c (0.005 mmol) in MeOH, was added 35% aqueous H<sub>2</sub>O<sub>2</sub> (0.5 mmol). The mixture was stirred at rt until all the oxidant has been consumed (iodometric test), and CHCl<sub>3</sub> was added. The mixture was washed with 5% sodium metabisulfite aqueous solution, the layers were separated and the aqueous one extracted twice with chloroform. The organic layers were washed with brine, dried over MgSO<sub>4</sub> and the solvent was removed under reduce pressure. Ratios sulfoxide:sulfone were determined by quantitative GC analysis and by <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz). Yields were determined by quantitative GC analysis. The sulfoxides 5a-f and sulfones 6a-f <sup>1</sup>H NMR spectra match those already reported in the literature.<sup>2</sup>

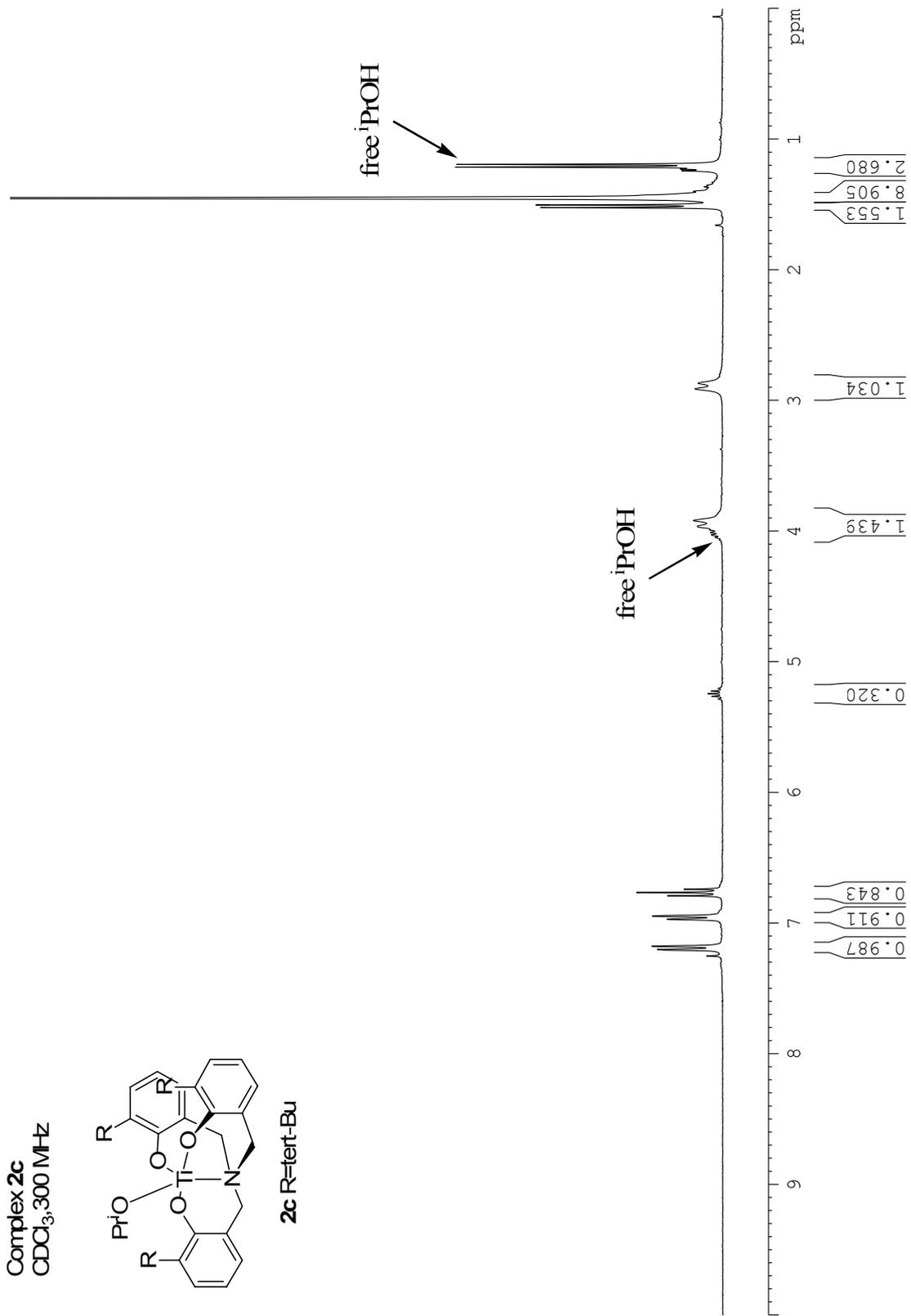
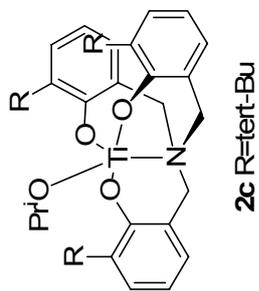
**Oxidation of thioanisole on gram-scale.**

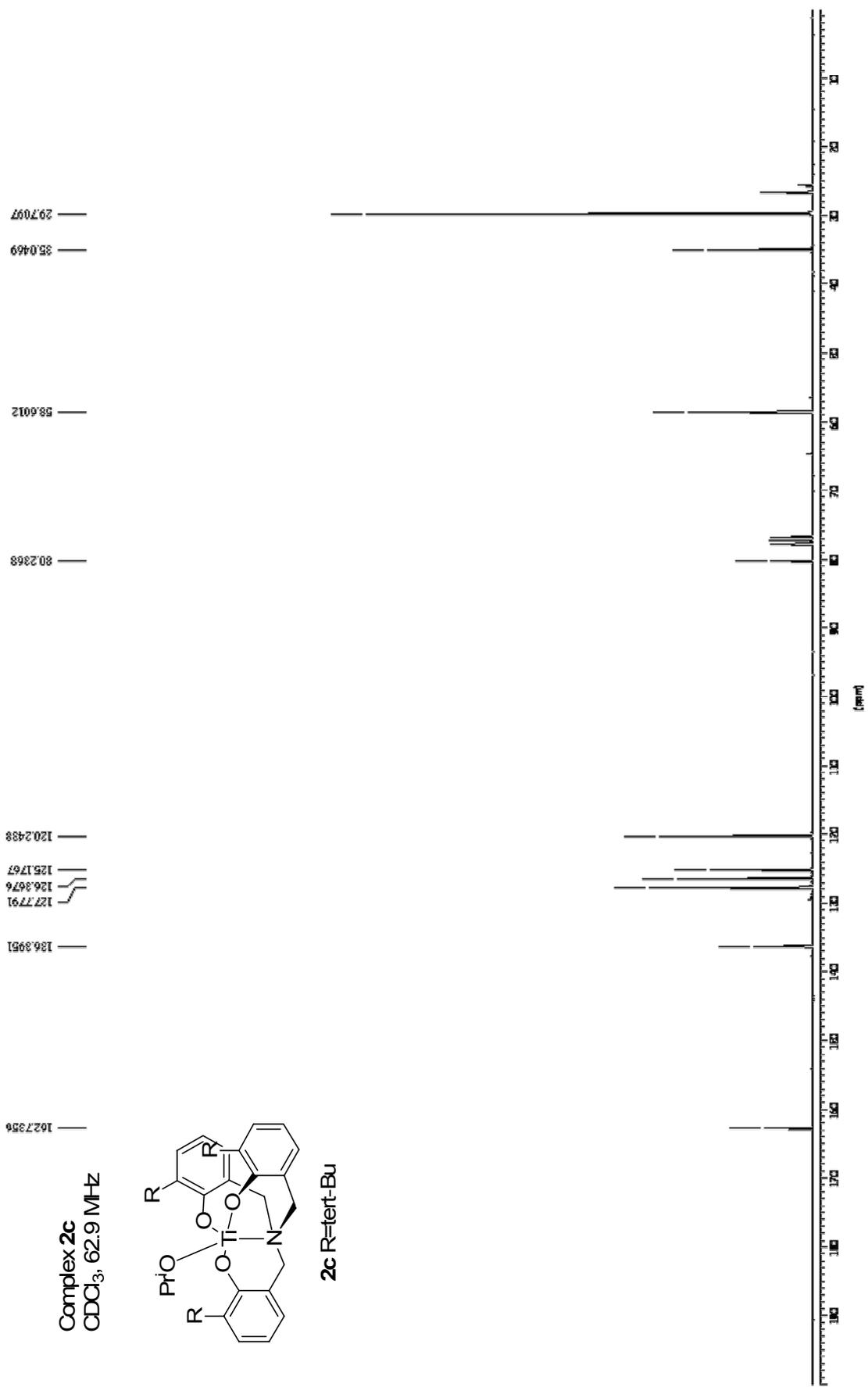
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(2) a) Brunel, J. M.; Diter, P.; Duetsch, M.; Kagan, H. B. *J. Org. Chem.* **1995**, *60*, 8086. b) Rebiere, F.; Samuel, O.; Ricard, L.; Kagan, H. B. *J. Org. Chem.* **1991**, *56*, 5991. c) Pitchen, P.; Dunach, E.; Dshemukh, M. N.; Kagan H. B. *J. Am. Chem. Soc.* **1984**, *106*, 8188.

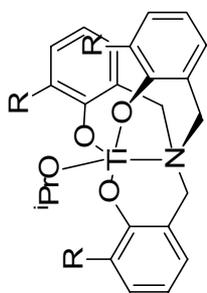
To a solution of thioanisole (**4a**) (859 mg, 6.9 mmol) and catalyst **2c** (42 mg, 0.069 mmol) in MeOH (13.8 ml), was added 35% aqueous H<sub>2</sub>O<sub>2</sub> (0.63 ml, 6.9 mmol). The mixture was stirred at rt for 9h, then concentrated to half volume and chloroform was added. The mixture was washed with 5% sodium metabisulfite aqueous solution, the layers were separated and the aqueous one extracted with chloroform. The organic layers were washed with brine, dried over MgSO<sub>4</sub> and the solvent was removed under reduce pressure. A conversion of 97% with sulfoxide:sulfone ratio 98:2 was obtained (quantitative GC analysis and <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz). The crude was purified by column chromatography on silica gel (petroleum ether/ ethyl acetate 1:2) obtaining 897mg (93%) of methyl phenyl sulfoxide (**5a**) and 31 mg (3%) of methyl phenyl sulfone (**6a**) corresponding to a ratio sulfoxide:sulfone 97:3 and 96% conversion. The sulfoxide and sulfone <sup>1</sup>H NMR spectra match those already reported in the literarture.<sup>2</sup>

Complex **2c**  
CDCl<sub>3</sub>, 300 MHz

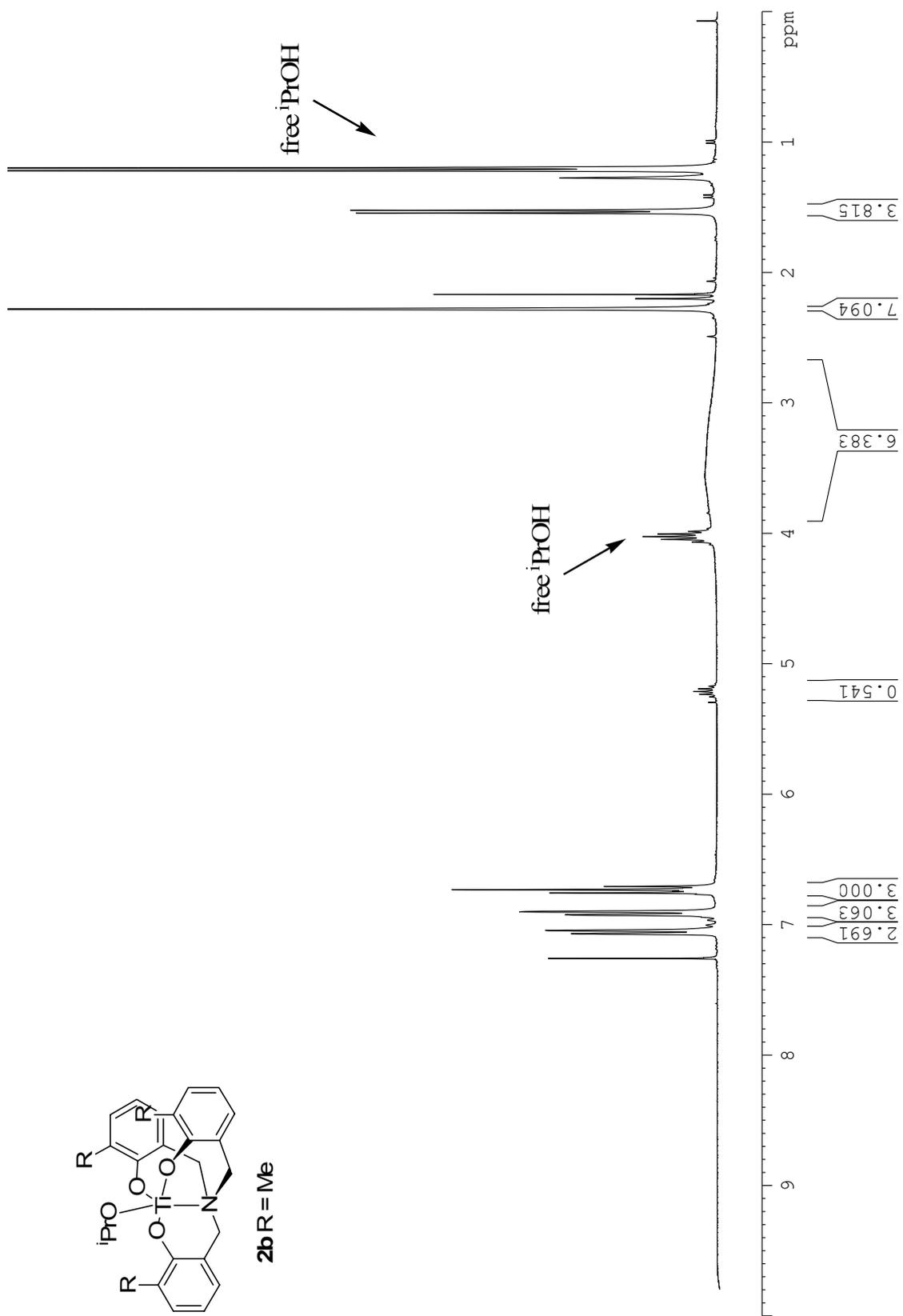


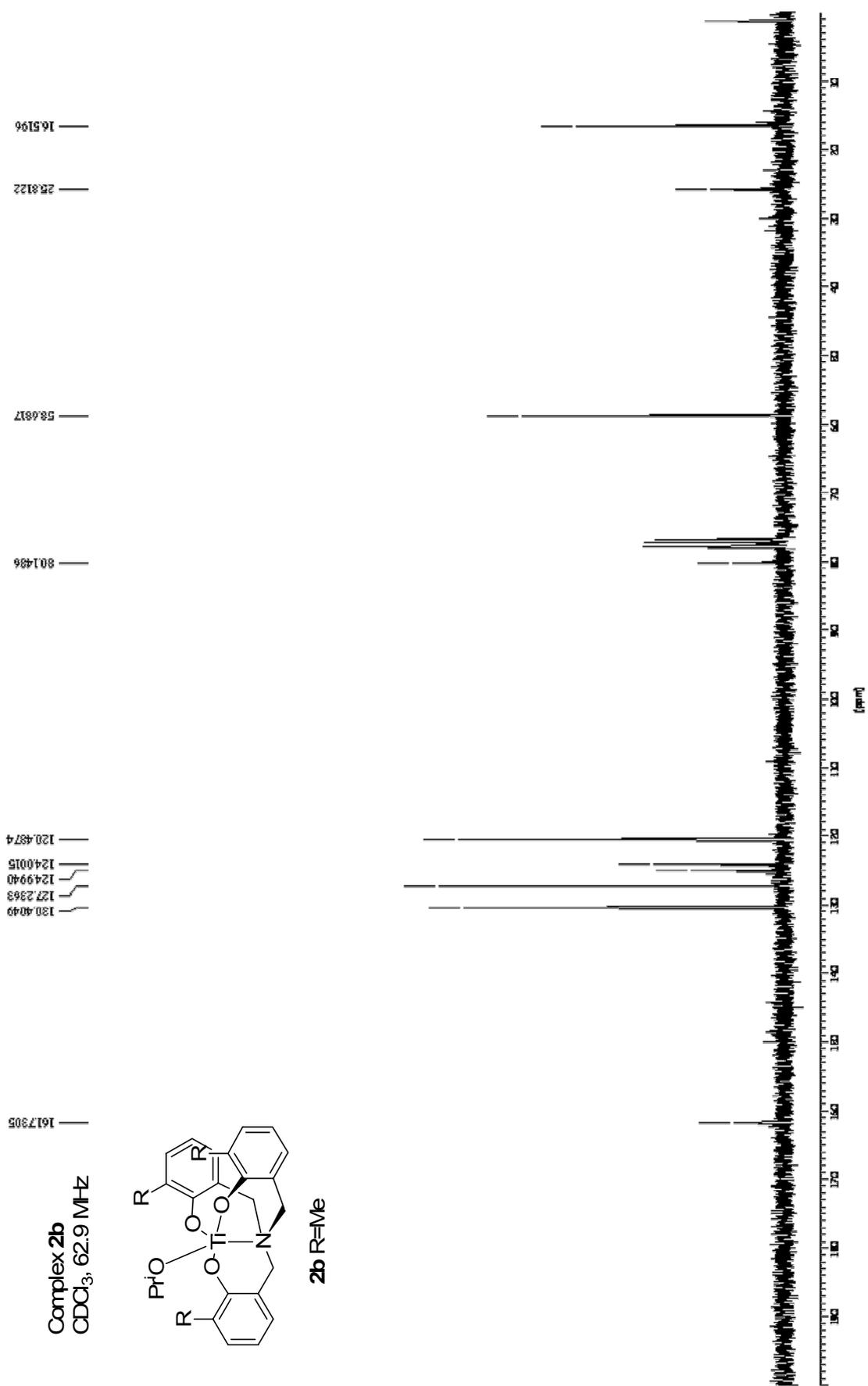


300 MHz, CDCl<sub>3</sub>

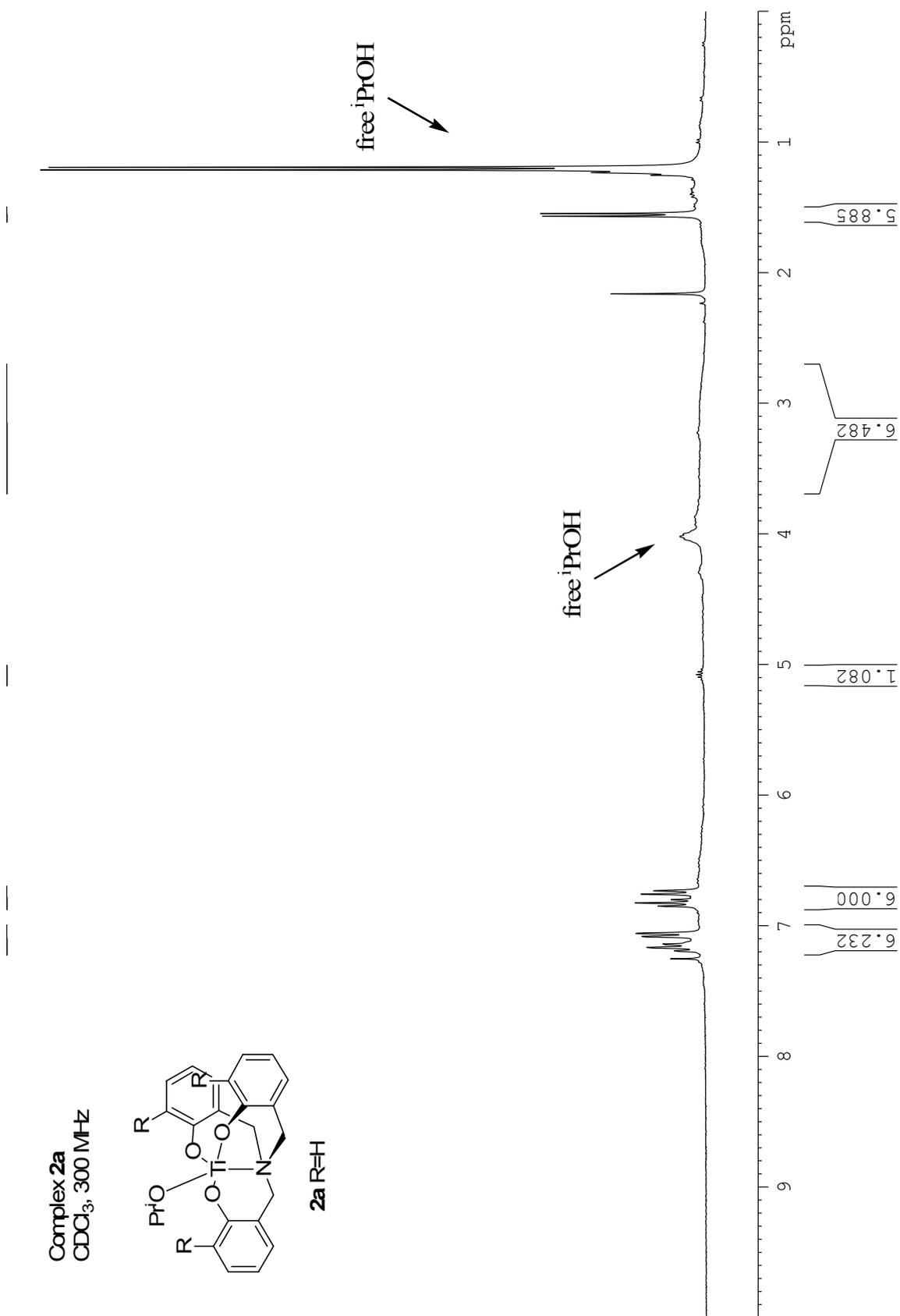
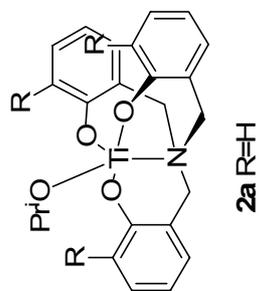


**2b** R = Me



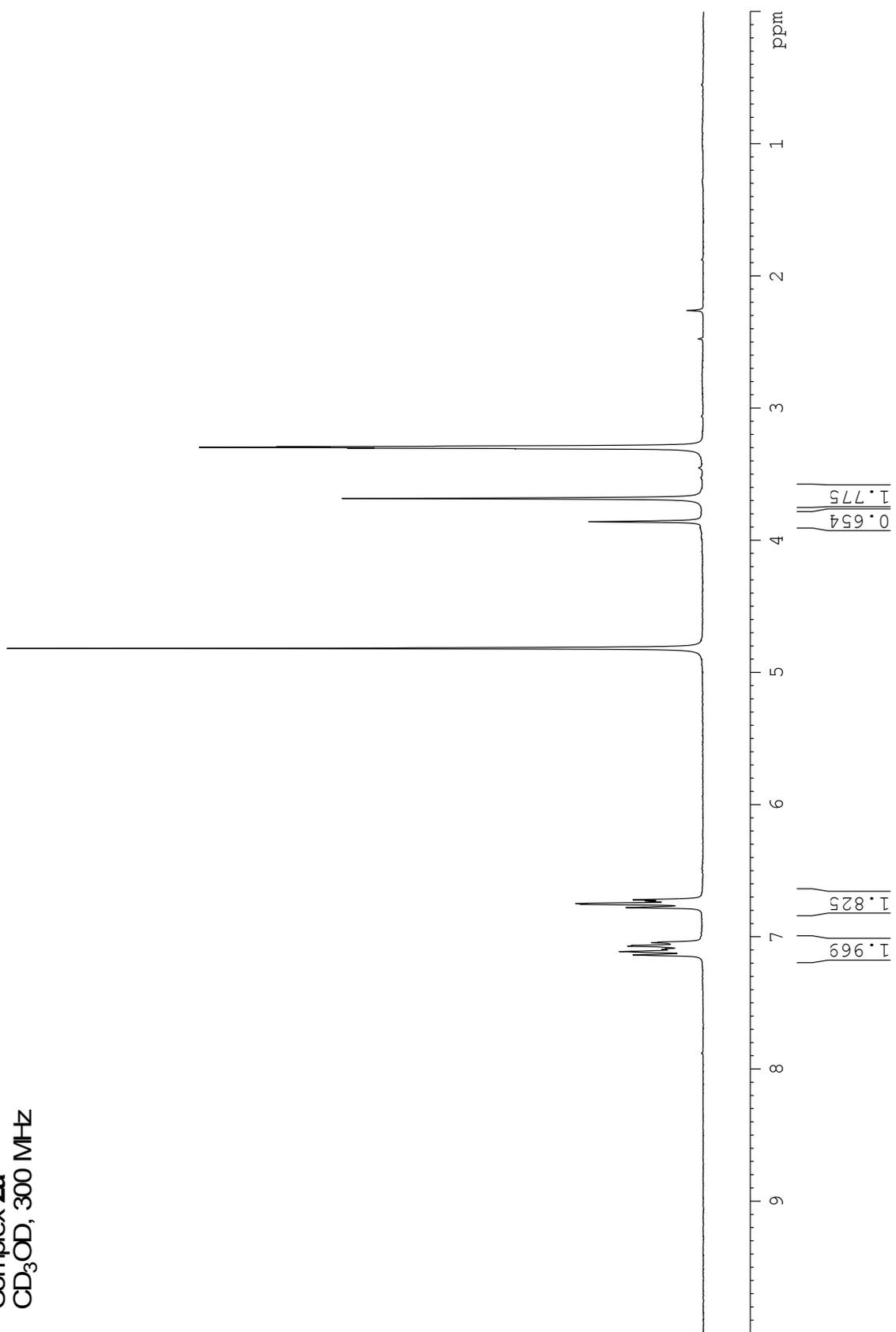


Complex **2a**  
CDCl<sub>3</sub>, 300 MHz

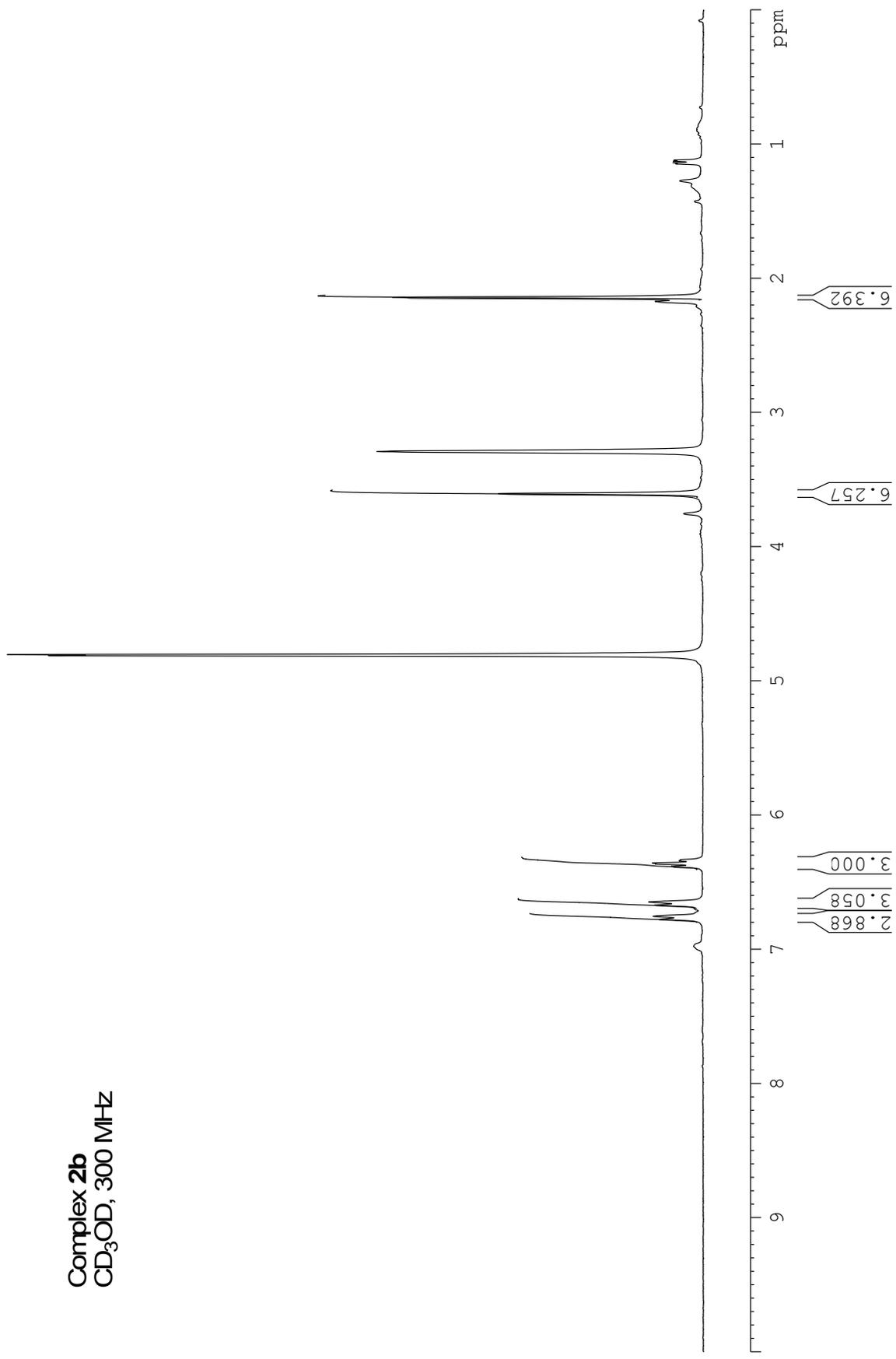




Complex **2a**  
CD<sub>3</sub>OD, 300 MHz



Complex **2b**  
CD<sub>3</sub>OD, 300 MHz



Complex **2c**  
MeOD, 250 MHz

