

## **Supporting Information**

# **Bismethylene Triphosphate Nucleotides of Uridine-4-Phosphate Analogues – A New Class of Anionic-Pyrimidine Nucleotide Analogues**

*Scott D. Taylor,<sup>1</sup>\* Farzad Mirzaei,<sup>1</sup> and Stephen L. Bearne<sup>2</sup>*

<sup>1</sup>Department of Chemistry, University of Waterloo, 200 University Ave. West, Waterloo, ON Canada.

<sup>2</sup>Department of Biochemistry and Molecular Biology, Dalhousie University, Halifax, Nova Scotia, B3H 1X5, Canada

### **Table of Contents**

General experimental information	<b>S3</b>
Characterization data for compounds <b>13</b> , <b>14</b> , <b>22</b> , <b>24</b> , <b>25</b> , <b>28</b> , <b>34</b> , <b>35</b> , <b>38</b> , <b>45</b> , and <b>49</b> .	<b>S4-S9</b>
References	<b>S9</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>12</b>	<b>S10-S12</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>13</b>	<b>S13-S15</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>14</b>	<b>S16-S18</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>22</b>	<b>S19-S21</b>
<sup>1</sup> H NMR and <sup>13</sup> C NMR spectra for compound <b>24</b>	<b>S22,S23</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR, and <sup>31</sup> P NMR for compound <b>25</b>	<b>S24-S26</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR, <sup>31</sup> P NMR and <sup>19</sup> F NMR spectra for compound <b>26</b>	<b>S27-S32</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>27</b>	<b>S33-S35</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>28</b>	<b>S36-S38</b>

<sup>1</sup> H NMR for compound <b>34</b>	<b>S39</b>
<sup>1</sup> H NMR and <sup>13</sup> C NMR spectra for compound <b>35</b>	<b>S40,S41</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>36</b>	<b>S42-S44</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>37</b>	<b>S45-S47</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>38</b>	<b>S48-S50</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>42</b>	<b>S51-S53</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>43</b>	<b>S54-S56</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>44</b>	<b>S57-S59</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>45</b>	<b>S60-S65</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>46</b>	<b>S66-S71</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>47</b>	<b>S72-S78</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR, <sup>31</sup> P NMR and <sup>19</sup> F NMR spectra for compound <b>48</b>	<b>S79-S85</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR, <sup>31</sup> P NMR spectra and analytical HPLC for compound <b>49</b>	<b>S86-S89</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR, <sup>31</sup> P NMR spectra and analytical HPLC for compound <b>50</b>	<b>S90-S93</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR and <sup>31</sup> P NMR spectra for compound <b>51</b> (with app. 15% <b>52</b> )	<b>S94-S96</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR, <sup>31</sup> P NMR, <sup>19</sup> F NMR spectra and analytical HPLC for compound <b>53</b>	<b>S97-S105</b>
<sup>1</sup> H NMR, <sup>13</sup> C NMR, and <sup>31</sup> P NMR spectra for compound <b>56</b>	<b>S106-S111</b>
<sup>1</sup> H NMR, <sup>31</sup> P NMR spectra and analytical HPLC for compound <b>57</b> (with app. 3% by <sup>1</sup> H NMR of compound <b>58</b> ).	<b>S112-S114</b>

**General.** All starting materials and reagents were obtained commercially and used without further purification with the exception of TMSBr and trimethylphosphite which were distilled under Ar and stored under Ar in a Schlenk tube. Tetrahydrofuran (THF) was distilled from sodium metal in the presence of benzophenone under argon. CH<sub>2</sub>Cl<sub>2</sub> was distilled from calcium hydride under nitrogen. DMF was distilled under aspirator pressure from CaH and stored under Ar. Methanol was dried by refluxing over magnesium metal and distilled under Ar and stored over 4 Å molecular sieves. Silica gel flash chromatography (flash chromatography) was performed using silica gel 60 Å (230-400 mesh). CDCl<sub>3</sub> was used as the solvent for all NMR experiments unless stated otherwise. For <sup>1</sup>H NMR in CDCl<sub>3</sub>, chemical shifts ( $\delta$ ) are reported in ppm relative to tetramethylsilane ( $\delta$  0.0, internal standard). For <sup>1</sup>H NMR in CD<sub>3</sub>OD, chemical shifts are reported in ppm relative to the solvent residual peak ( $\delta$  3.31, central peak). For <sup>1</sup>H NMR in D<sub>2</sub>O, chemical shifts are reported in ppm relative to the solvent residual peak ( $\delta$  4.79). For <sup>1</sup>H NMR in DMSO-*d*<sub>6</sub>, chemical shifts are reported in ppm relative to the solvent residual peak ( $\delta$  2.49). For <sup>13</sup>C NMR in CDCl<sub>3</sub>, chemical shifts are reported in ppm relative to CDCl<sub>3</sub> ( $\delta$  77.0 for the central peak). For <sup>13</sup>C NMR spectra run in CD<sub>3</sub>OD, chemical shifts are reported in ppm relative to the solvent residual peak ( $\delta$  49.00, central peak). For <sup>13</sup>C NMR in DMSO-*d*<sub>6</sub>, chemical shifts are reported in ppm relative to DMSO-*d*<sub>6</sub>, ( $\delta$  39.5 for the central peak). For <sup>13</sup>C NMR spectra run in D<sub>2</sub>O, chemical shifts are reported in ppm relative to CH<sub>3</sub>OH in D<sub>2</sub>O ( $\delta$  49.5, external standard). All <sup>31</sup>P NMR spectra chemicals shifts are reported in ppm relative to 85% H<sub>3</sub>PO<sub>4</sub> ( $\delta$  0.0, external standard). All <sup>19</sup>F spectra chemical shifts are reported in ppm relative to an external trichlorofluoromethane standard ( $\delta$  0.0, Cflash chromatographyl<sub>3</sub>). Melting points are uncorrected. Semi-preparative and analytical HPLC were performed using C-18 columns.

**2',3',5'-Tri-*O*-acetyl-3-deazacytidine 4-*N*-[*O,O*-Bis(2-cyanoethyl)phosphoramidate] (13).** The same procedure as for compound **12** was employed except the following quantities were used: 2',3',5'-tri-*O*-acetyl-3-deazacytidine (**10**<sup>1</sup>, 0.775 g, 2.1 mmol) in dry THF (10 mL), tetrazole (0.297 g, 4.2 mmol), phosphoramidate **15** (1.14 g, 4.2 mmol) in dry THF (5 mL), *tert*-butyl hydroperoxide (1.25 mL, 12.6 mmol). Purification was achieved using flash chromatography (4% MeOH-96% EtOAc then 10% MeOH-90% EtOAc) to give **13** as a white foam (0.785 g, 68%). <sup>1</sup>H NMR (300 MHz): δ 7.83 (d, *J* = 9.9 Hz, 1H), 7.44 (d, *J* = 7.8 Hz, 1H), 6.20 (dd, *J* = 8.0, 1.9, 1H), 6.16 (d, *J* = 3.3 Hz, 1H), 6.04 (d, *J* = 2.0 Hz, 1H), 5.39 (overlapping dd, *J* = 5.3 Hz, 1H), 5.28 (overlapping dd, *J* = 5.9 Hz, 1H), 4.20-4.39 (m, 7H), 2.75-2.81 (m, 4H), 2.08-2.12 (three overlapping singlets, 9H); <sup>13</sup>C NMR (75 MHz): δ 170.4, 169.6, 169.5, 162.7, 150.7, 133.6, 116.7, 102.26 (d, *J* = 7.6 Hz), 101.06 (d, *J* = 8.5 Hz), 88.5, 79.2, 73.8, 69.6, 62.8, 62.01, 61.99 (d, *J* = 4.2 Hz), 20.8, 20.5, 20.4, 19.6 (d, *J* = 6.7 Hz); <sup>31</sup>P NMR (121 MHz): δ 1.7; LR<sup>+</sup>ESIMS *m/z* (relative intensity) 555.1 (M + 1, 100), 297.1 (41), 259.1 (35); HR<sup>+</sup>ESIMS *m/z* calculated for C<sub>22</sub>H<sub>28</sub>N<sub>4</sub>O<sub>11</sub>P (M + 1): 555.1481, found 555.1492.

**2',3',5'-Tri-*O*-benzoyl-3-deazacytidine 4-*N*-[*O,O*-Bis(2-cyanoethyl)phosphoramidate] (14).** Same procedure as for compound **12** except the following quantities were used: 2',3',5'-tri-*O*-benzoyl-3-deazacytidine (**11**,<sup>1</sup> 1.70 g, 3.07 mmol) in dry THF (20 mL), tetrazole (0.430 g, 6.14 mmol), phosphoramidate **15** (1.67 g, 6.14 mmol) in dry THF (10 mL), *tert*-butyl hydroperoxide (3.1 mL, 30.7 mmol). Purification was achieved using flash chromatography (EtOAc then 2% MeOH-98% EtOAc) to give **14** as a white foam (1.82 g, 80%). <sup>1</sup>H NMR (300 MHz): δ 8.06 (d, *J* = 8.0 Hz, 2H), 7.87-7.95 (m, 4H), 7.10-7.60 (m, 10 H), 7.12, (d, *J* = 9.0 Hz, 1H), 6.42 (d, *J* = 3.3 Hz, 1H), 6.05 (s overlapping with d, 2H), 5.79-5.87 (m, 2H), 4.65-4.82 (m, 3H), 4.20-4.369 (m, 4H), 2.71-2.81 (m, 4H); <sup>13</sup>C NMR (75 MHz): δ 166.1, 165.24, 165.17, 162.7, 150.9, 134.1, 133.66, 133.60, 133.4, 129.8, 129.75, 129.71, 129.4, 128.73, 128.70, 128.57, 128.53, 128.44, 116.80, 102.36 (d, *J* = 7.2 Hz), 101.28 (d, *J* = 8.4 Hz), 89.8, 79.7, 74.8, 70.1, 63.7, 62.0

(d,  $J = 3.6$  Hz), 19.6, (d,  $J = 7.8$  Hz);  $^{31}\text{P}$  NMR (121 MHz):  $\delta$  1.72; LR $^+\text{ESIMS}$   $m/z$  (relative intensity) 741.2 (M + 1, 97), 445.2 (100); HR $^+\text{ESIMS}$   $m/z$  calculated for  $\text{C}_{37}\text{H}_{34}\text{N}_4\text{O}_{11}\text{P}$  (M + 1): 741.1962, found 741.1945.

**{1-[3,4-bis-(acetoxyloxy)-5-acetoxyloxymethyltetrahydro-furan-2-yl]-2-oxo-1,2-dihydro-pyridin-4-ylmethyl}phosphonic acid dimethyl ester (22).** (2-Hydroxypyridin-4-ylmethyl)phosphonic acid dimethyl ester<sup>2</sup> (0.330 g, 1.52 mmol) and acetyl 2,3,5-tri-*O*-acetyl- $\beta$ -D-ribofuranose (0.484 g, 1.52 mmol) were suspended in dry  $\text{CH}_3\text{CN}$  (6 mL) under Ar at 0 °C (ice bath). After addition of bis(trimethylsilyl)trifluoroacetamide (0.606 mL, 2.25 mmol) to the suspension, the ice bath was removed and the solution stirred for 1 h. Trimethylsilyltriflate (0.412 mL, 2.25 mmol) was added and the solution stirred for 2 h. The reaction was diluted with EtOAc (200 mL) and ether (100 mL) and washed with sat.  $\text{NaHCO}_3$  and sat. brine. The organic layer was dried and concentrated and the residue purified by flash chromatography (5% MeOH-95% EtOAc then 10% MeOH-90% EtOAc) which gave pure compound **22** as a white foam (0.450 g, 62%).  $^1\text{H}$  NMR (300 MHz):  $\delta$  7.44 (d,  $J = 7.2$  Hz, 1H), 6.37 (s, 1H), 6.24 (s overlapping with d, 2H), 5.35 (overlapping dd,  $J = 4.6$  Hz, 1H), 5.30 (overlapping dd,  $J = 5.1$  Hz, 1H), 4.30-4.40 (m, 3 H),  $\delta$  3.72 (d,  $J = 10.7$  Hz, 3H), 3.72 (d,  $J = 10.9$  Hz, 3H), 2.51 (d,  $J = 36.7$  Hz, 2H), 2.08, (three overlapping singlets, 9H);  $^{13}\text{C}$  NMR (75 MHz):  $\delta$  170.1, 169.4, 169.3, 161.49 (d,  $J = 2.4$  Hz), 145.37 (d,  $J = 8.6$  Hz), 132.0, 120.83 (d,  $J = 8.5$  Hz), 108.34 (d,  $J = 4.5$  Hz), 88.1, 79.3, 73.8, 69.6, 62.7, 52.98 (d,  $J = 6.5$  Hz), 32.21 (d,  $J = 136.2$  Hz), 20.6, 20.3;  $^{31}\text{P}$  NMR (121 MHz):  $\delta$  27.6; LR $^+\text{ESIMS}$   $m/z$  (relative intensity) 476.1 (M + 1, 100); HR $^+\text{ESIMS}$   $m/z$  calculated for  $\text{C}_{19}\text{H}_{27}\text{NO}_{11}\text{P}$  (M + 1): 476.1322, found 476.1321.

**1-(2',3'-Di-*O*-acetyl- $\beta$ -D-ribofuranosyl)-2-oxo-1,2-dihdropyridine (24).** Prepared from compound **21** as a white foam in 75 % yield (90% EtOAc-10% hexane) using the general deacetylation procedure described in the experimental section.  $^1\text{H}$  NMR (300 MHz):  $\delta$  7.76 (d,  $J = 7.4$  Hz, 1H), 7.15-7.21 (m,

1H), 6.32 (d,  $J$  = 9.6 Hz, 1H), 6.07-6.16 (m, 2H), 5.30-5.39 (m, 2H), 4.54 (bs, 1H), 4.10 (s, 1H), 3.80 (d,  $J$  = 11.6 Hz, 1H), 3.65 (d,  $J$  = 12.4 Hz, 1H);  $^{13}\text{C}$  NMR (75 MHz):  $\delta$  169.7, 169.3, 162.5, 140.3, 133.8, 120.1, 106.8, 87.8, 83.1, 73.9, 70.4, 60.8, 20.4, 20.2; LREIMS  $m/z$  (relative intensity) 311.1 (M<sup>+</sup>, 14), 217.1 (100), 162.1 (17), 127.1 (40), 85 (37); HREIMS  $m/z$  calculated for C<sub>14</sub>H<sub>17</sub>NO<sub>7</sub>: 311.1005, found 311.1004

**{1-[3,4-bis-(acetoxyloxy)-5-hydroxymethyltetrahydro-furan-2-yl]-2-oxo-1,2-dihydro-pyridin-4-ylmethyl}phosphonic acid dimethyl ester (25).** Prepared from compound **22** as a white foam in 87% yield (5% MeOH-90% CHCl<sub>3</sub>) using the general deacetylation procedure described in the experimental section.  $^1\text{H}$  NMR (300 MHz):  $\delta$  7.73 (d,  $J$  = 5.9 Hz, 1H), 6.26 (s, 1H), 6.17 (d,  $J$  = 4.8 Hz, 1H), 6.13 (d,  $J$  = 7.4 Hz, 1H), 5.34-5.40 (m, 2H), 4.23 (bs, 1H), 4.11 (s, 1H), 3.80 (d,  $J$  = 13.8 Hz, 1H), 3.67 (d,  $J$  = 13.9 Hz, 1H), 3.61 (d,  $J$  = 11.0 Hz, 3H), 3.60 (d,  $J$  = 11.4 Hz, 3H), 2.87 (d,  $J$  = 22.4 Hz, 2H), 1.97 (s, 3H), 1.93 (s, 3H);  $^{13}\text{C}$  NMR (75 MHz):  $\delta$  170.0, 169.6, 162.13 (d,  $J$  = 2.3 Hz), 145.49 (d,  $J$  = 8.8 Hz), 133.63 (d,  $J$  = 2.0 Hz), 120.53 (d,  $J$  = 9.0 Hz), 108.73 (d,  $J$  = 3.7 Hz), 87.7, 83.5, 74.1, 70.8, 61.2, 53.20 (d,  $J$  = 6.9 Hz), 53.17 (d,  $J$  = 6.1 Hz), 32.2 (d,  $J$  = 136.9 Hz), 20.7, 20.5.  $^{31}\text{P}$  NMR (121 MHz):  $\delta$  28.0; LR<sup>+</sup>ESIMS  $m/z$  (relative intensity) 434.1 (M + 1, 100), 217 (25); HR<sup>+</sup>ESIMS  $m/z$  calculated for C<sub>17</sub>H<sub>25</sub>NO<sub>10</sub>P (M + 1): 434.1216, found 434.1216.

**2',3',5'-Di-O-acetyl-3-deazacytidine 4-N-[O,O-Bis(2-cyanoethyl)phosphoramidate] (28).** Prepared from compound **13** as a white foam in 84% yield (10% MeOH-95% EtOAc) using the general deacetylation procedure described in the experimental section.  $^1\text{H}$  NMR (300 MHz):  $\delta$  7.54 (d,  $J$  = 7.5 Hz, 1H), 6.94 (s, 1H), 6.17 (d,  $J$  = 9.4 Hz, 1H), 6.03 (s, 1H), 5.94 (d,  $J$  = 4.4 Hz, 1H), 5.67 (d,  $J$  = 5.3 Hz, 1H), 5.58 (d,  $J$  = 3.6 Hz, 1H), 5.49 (d,  $J$  = 5.3 Hz, 1H), 4.30 (m, 5H), 3.96 (d,  $J$  = 12.1 Hz, 1H), 3.78 (d,  $J$  = 13.8 Hz, 1H), 3.64 (s, 1H), 2.80 (t,  $J$  = 5.9 Hz, 4H), 2.07, 2.09 (two overlapping singlets, 6H);

<sup>13</sup>C NMR (75 MHz):  $\delta$  170.1, 169.8, 163.2, 151.1, 135.4, 117.0, 102.2 (d,  $J$  = 6.5 Hz), 101.4 (d,  $J$  = 9.6 Hz), 89.2, 83.2, 73.6, 70.4, 62.2 (d,  $J$  = 4.3 Hz), 61.1, 20.6, 20.5, 19.6 (d,  $J$  = 7.8 Hz); <sup>31</sup>P NMR (121 MHz):  $\delta$  1.6; LR<sup>+</sup>ESIMS  $m/z$  (relative intensity) 513.1 (M + 1, 100), HR<sup>+</sup>ESIMS  $m/z$  calculated for C<sub>20</sub>H<sub>26</sub>N<sub>4</sub>O<sub>10</sub>P: 513.1387, found 513.1406.

**2',3'-Di-O-benzoyluridine (34).** Prepared from compound **29** as a white solid in 81% yield (70% EtOAc-30% hexane) using the general debenzylation procedure described in the experimental section. Mp (EtOAc) 195-197 °C (lit.<sup>3</sup> mp 194-196 °C); <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD):  $\delta$  8.15 (d,  $J$  = 8.2 Hz, 1H), 8.03 (d,  $J$  = 6.9 Hz, 2H), 7.88 (d,  $J$  = 8.3, 2H), 7.37-7.67 (m, 6H), 6.42 (d,  $J$  = 6.0 Hz, 1H), 5.79-5.88 (m, 3H), 4.52 (d,  $J$  = 2.4 Hz, 1H), 3.91-4.01 (m, 2H).

**1-(2',3'-Di-O-benzoyl-β-D-ribofuranosyl)-2-oxo-1,2-dihdropyridine (35).** Prepared from compound **30** using the general debenzylation procedure described in the experimental section. Purification was achieved by re-crystallisation from MeOH which gave **35** as a white solid in 84% yield. Mp 203-204 °C; <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>-CDCl<sub>3</sub>):  $\delta$  8.04 (d,  $J$  = 6.1 Hz, 1H), 7.79-7.88 (m, 4H), 7.44-7.50 (m, 2H), 7.28-7.36 (m, 4H), 6.57 (d,  $J$  = 5.3 Hz, 1H), 6.33 (d,  $J$  = 8.9 Hz, 1H), 6.21 (overlapping dd,  $J$  = 6.2 Hz, 1H), 5.66-5.73 (m, 2H), 5.33 (bt,  $J$  = 4.5 Hz, 1H), 4.42 (s, 1H), 3.70-3.90 (m, 2H); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>-CDCl<sub>3</sub>):  $\delta$  165.0, 164.7, 161.9, 140.2, 134.1, 133.6, 129.6, 129.5, 129.1, 128.8, 128.6, 128.5, 120.2, 106.2, 86.7, 83.7, 75.0, 71.7, 60.9; LR<sup>+</sup>ESIMS  $m/z$  (relative intensity) 436.1 (M + 1, 100), 341.1 (75), 183.0 (30); HR<sup>+</sup>ESIMS  $m/z$  calculated for C<sub>24</sub>H<sub>22</sub>NO<sub>7</sub>: 436.1396 (M + 1), found 436.1396.

**2',3',5'-Di-O-acetyl-3-deazacytidine 4-N-[O,O-Bis(2-cyanoethyl)phosphoramidate] (38).** Catalyst **20** (30 mol %) was added to a solution of the nucleoside **14** (200 mg, 0.27 mmol) in dry MeOH-dry THF (8 mL, 1:1) and the resulting solution was stirred at 48 °C (oil bath). After 48 h, an additional 16 mol% catalyst was added and the reaction stirred for an additional 24 h at 48 °C. The solution was concentrated and the resulting residue purified using flash chromatography (4% MeOH-96% EtOAc) to give nucleoside **38**

as a white foam (95.8 mg, 56%).  $^1\text{H}$  NMR (300 MHz):  $\delta$  7.91 (d,  $J = 7.5$  Hz, 4H), 7.64 (d,  $J = 8.0$  Hz, 1H), 7.48-7.55 (m, 2H), 7.29-7.42 (m, 5H), 6.25 (d,  $J = 7.3$  Hz, 1H), 6.20 (d,  $J = 4.4$  Hz, 1H), 6.10 (s, 1H), 5.98 (overlapping dd,  $J = 5.3$  Hz, 1H), 5.88 (overlapping dd,  $J = 5.5$  Hz, 1H), 4.47 (d,  $J = 4.4$  Hz, 1H), 4.31 (t,  $J = 8.1$  Hz, 4H), 4.06 (d,  $J = 12.2$  Hz, 1H), 3.91 (d,  $J = 12.7$  Hz, 1H), 2.78 (t,  $J = 5.9$  Hz, 4H), 1.63 (bs, 1H);  $^{13}\text{C}$  NMR (75 MHz):  $\delta$  165.7, 165.4, 163.3, 151.0, 135.9, 133.7 (d,  $J = 4.6$  Hz), 129.92, 129.86, 129.0, 128.9, 128.6, 102.6 (d,  $J = 7.2$  Hz), 101.4 (d,  $J = 8.1$  Hz), 90.7, 83.7, 74.6, 71.2, 62.1 (d,  $J = 4.8$  Hz), 61.5, 19.7 (d,  $J = 7.2$  Hz);  $^{31}\text{P}$  NMR (121 MHz):  $\delta$  1.67; LR $^+\text{ESIMS}$   $m/z$  (relative intensity): 637.1 (M + 1, 100), 341.1 (23); HR $^+\text{ESIMS}$   $m/z$  calculated for  $\text{C}_{30}\text{H}_{30}\text{N}_4\text{O}_{10}\text{P}$  (M + 1): 637.1700, found 637.1687.

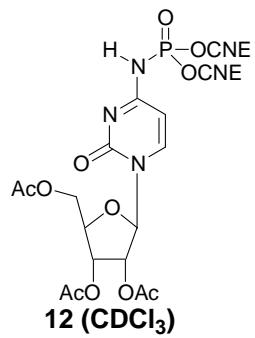
**1-(2',3'-Di-O-acetyl-5'-(bis-[(benzyloxyphosphorylmethyl)benzyloxyphosphorylmethyl]-methoxyphosphoryl)- $\beta$ -D-ribofuranosyl)-2-oxo-1,2-dihdropyridine (45).** Prepared from compound **35** as a white foam in 64% yield (100% EtOAc then 5% MeOH-95% EtOAc) using the general procedure described in the experimental section.  $^1\text{H}$  NMR (300 MHz):  $\delta$  7.94-7.85 (m, 4H), 7.67-7.76 (m, 1H), 7.48-7.53 (m, 2H), 7.20-7.35 (m, 20H), 6.60-6.70 (m, 1H), 6.49 (d,  $J = 8.8$  Hz, 1H), 6.17-6.25 (m, 1H), 5.91 (overlapping dd,  $J = 5.3$  Hz, 0.4 H), 5.79 (overlapping dd,  $J = 5.3$  Hz, 0.6 H), 5.63-5.73 (m, 1H), 4.90-5.20 (m, 6H), 4.37-4.69 (m, 3H), 3.81 (d,  $J = 11.9$  Hz, 1H), 3.73 (d,  $J = 11.7$  Hz, 2H), 2.68-3.12 (m, 4H);  $^{13}\text{C}$  NMR (75 MHz):  $\delta$  165.1, 165.04, 165.00, 164.9, 162.1, 162.03, 162.00, 139.8, 139.72, 139.68, 135.68, 135.66, 135.58, 135.55, 135.49, 133.40, 133.37, 13.31, 133.15, 133.07, 132.95, 132.92, 129.7, 129.6, 128.6, 128.52, 128.48, 128.4, 128.3, 128.2, 128.00, 127.91, 127.87, 127.82, 127.80, 120.86, 120.82, 120.75, 106.5, 106.30, 106.27, 87.5, 87.3, 87.2, 86.9, 81.01, 80.95, 80.91, 80.86, 74.51, 74.47, 74.44, 70.55, 70.49, 70.44, 70.40, 68.04, 67.98, 67.92, 67.86, 67.81, 67.76, 67.08, 67.03, 67.00, 66.93, 66.88, 65.45, 65.41, 65.17, 65.12, 65.07, 65.00, 53.47, 53.43, 53.21, 53.18, 53.16, 53.14, 52.84, 52.80, 26.78-29.49 (m);  $^{31}\text{P}$  NMR (121 MHz):  $\delta$  39.73-39.80 (m), 39.37-39.50 (m), 23.74-23.84 (m), 23.29-23.43 (m), 21.74-21.85 (m); LR $^+\text{ESIMS}$   $m/z$  (relative intensity): 956 (M + 1); HR $^+\text{ESIMS}$   $m/z$  calculated for  $\text{C}_{48}\text{H}_{49}\text{NO}_{14}\text{P}_3$ : 956.2350, found 956.2366.

**{1-[3,4-Bis-(hydroxy)-5-[(dihydroxyphosphorylmethyl)hydroxyphosphorylmethyl]-hydroxyphosphoryl}tetrahydro-furan-2-yl]- 2-oxo-1,2-dihdropyridine tetrasodium salt (49).**

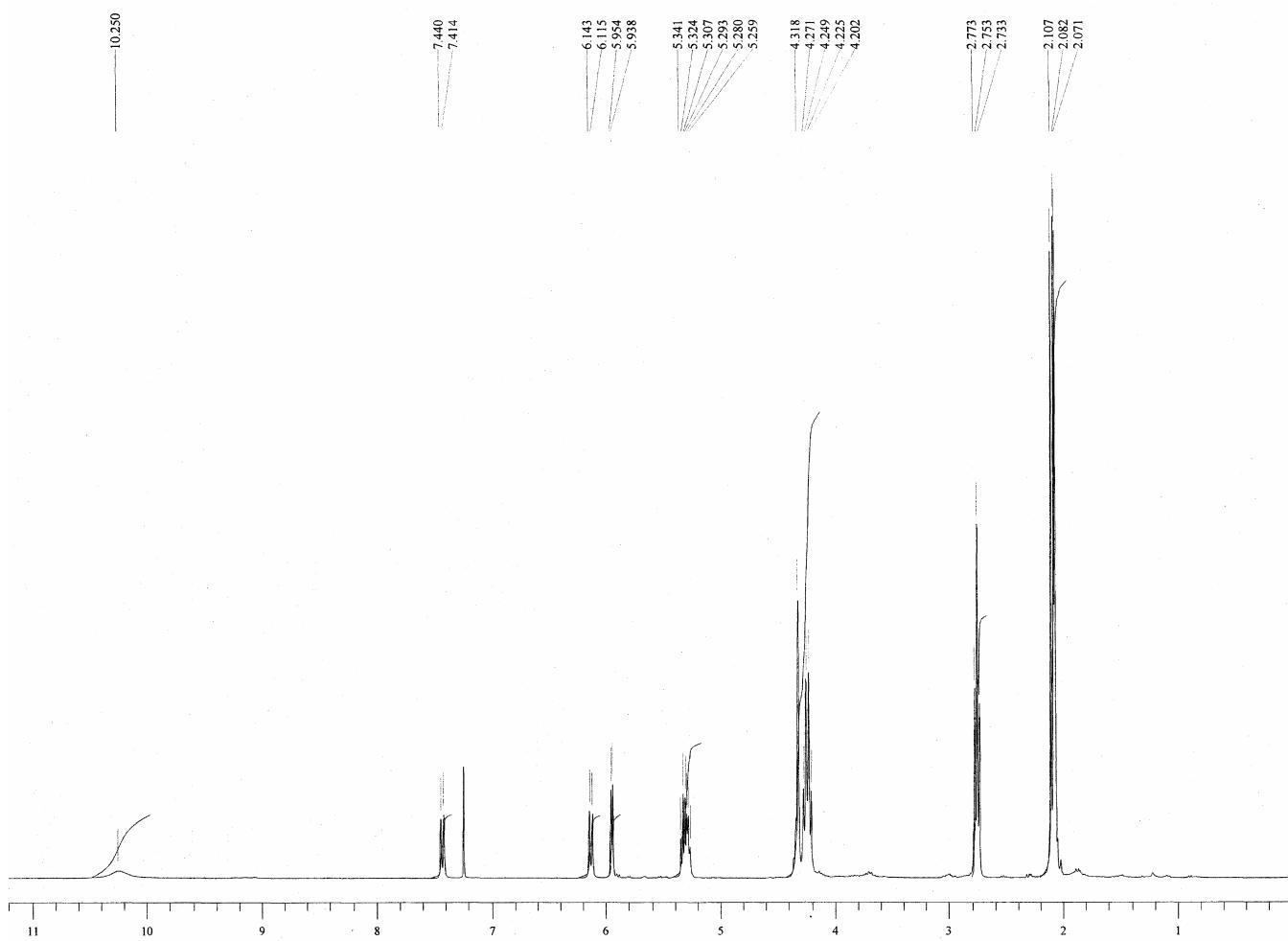
Prepared from compound **45** (0.100 g, 0.105 mmol) using the general procedure described in the experimental section. The following solvent system was used for preparative HPLC: solvent A: 100 mM TEAA, pH 7.0, solvent B: CH<sub>3</sub>CN. The following elution profile was used: 0-13 min: 99% A-1% B; 13-50 min: linear gradient of 99% A-1% B to 93% A-7% B. Flow rate = 8 mL/min. t<sub>r</sub> = 23.36 min. The tetrasodium salt of nucleotide **49**, generated using a Dowex 50Wx8 Na<sup>+</sup> ion exchange column, was a white powder (27.8 mg, 48%). <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): δ 7.92 (d, J = 5.8 Hz, 1H), 7.49 (overlapping dd, J = 7.2 Hz, 1H), 6.51 (overlapping dd, J = 6.8 Hz, 1H), 6.46 (d, J = 8.8 Hz, 1H), 6.60, (d, J = 2.7 Hz, 1H), 3.97-4.28 (m, 5H), 1.93-2.35 (m, 4H); <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O): δ 164.0, 142.4, 133.3, 118.8, 109.4, 89.3, 82.8 (d, J = 7.1 Hz), 75.0, 68.9, 62.7, 30.5-32.6 (m); <sup>31</sup>P NMR (121 MHz, D<sub>2</sub>O): δ 28.4 (P<sub>β</sub>), 18.2 (P<sub>α</sub>), 15.5 (P<sub>γ</sub>); LR-ESIMS m/z (relative intensity) 462 (M - 4Na + 3H<sup>+</sup>, 100). HR-ESIMS m/z calculated for C<sub>12</sub>H<sub>19</sub>NO<sub>12</sub>P<sub>3</sub> (M - 4Na + 3H<sup>+</sup>): 462.0120, found 462.0123. The analytical HPLC chromatogram of **49** (elution profile: linear gradient of 99% A-1% B to 93% A-7% B over 50 minutes) indicated **49** to be greater than 99.8% pure (flow rate 1 mL/min, t<sub>r</sub> = 13.46 min).

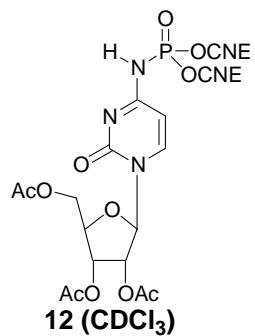
## References

1. Cook, P. D.; Day, R. T.; Robins, R. K. *J. Heterocyclic Chem.* **1997**, *14*, 1295.
2. Taylor, S. D.; Mirzaei, F.; Bearne, S. L. *J Org Chem.* **2006**, *71*, 9420.
3. Lohrmann, R.; Khorana, H. G. *J. Am. Chem. Soc.* **1964**, *86*, 4188.

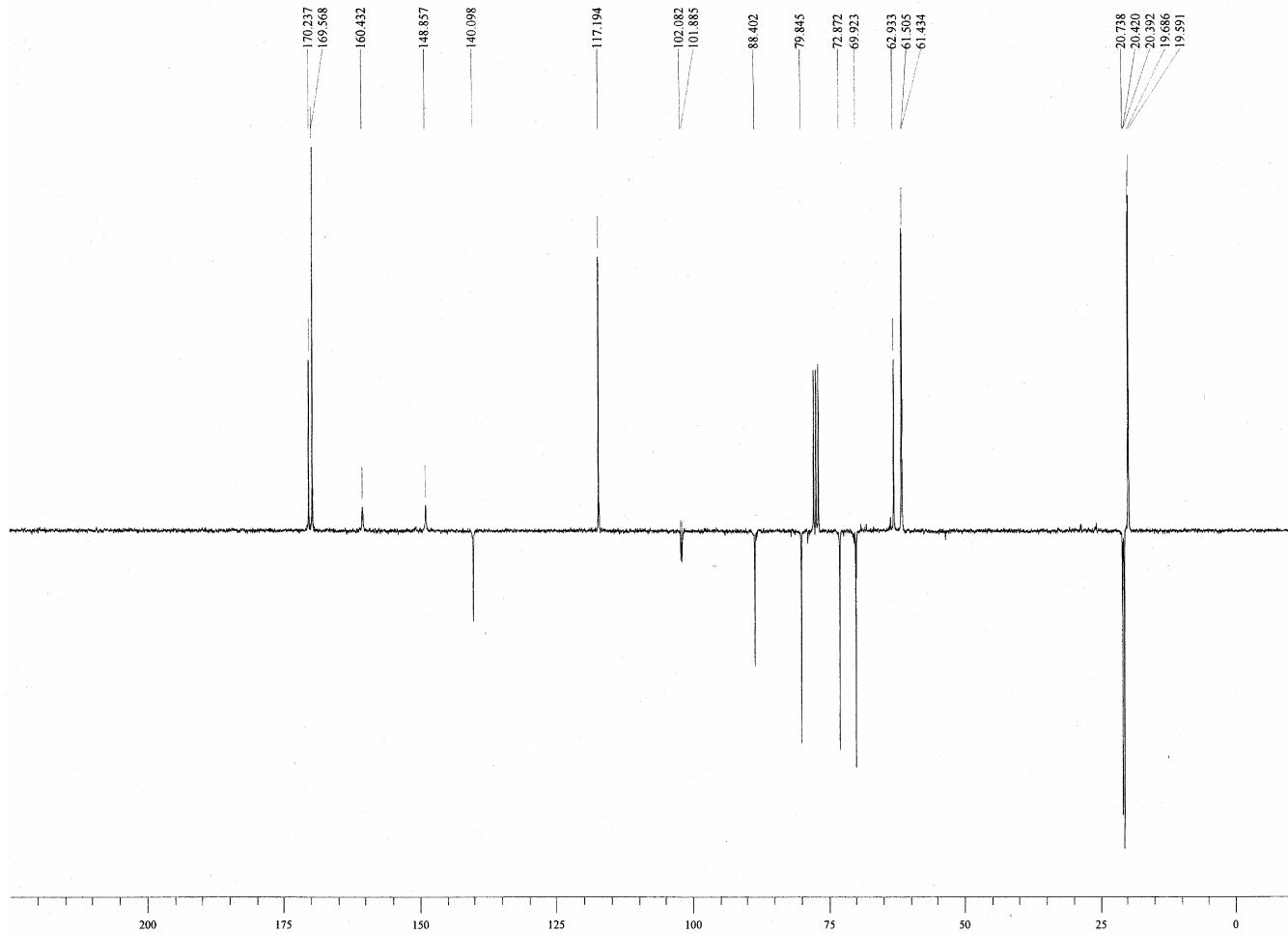


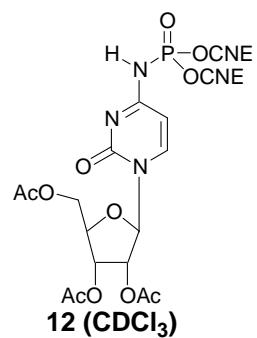
<sup>1</sup>H



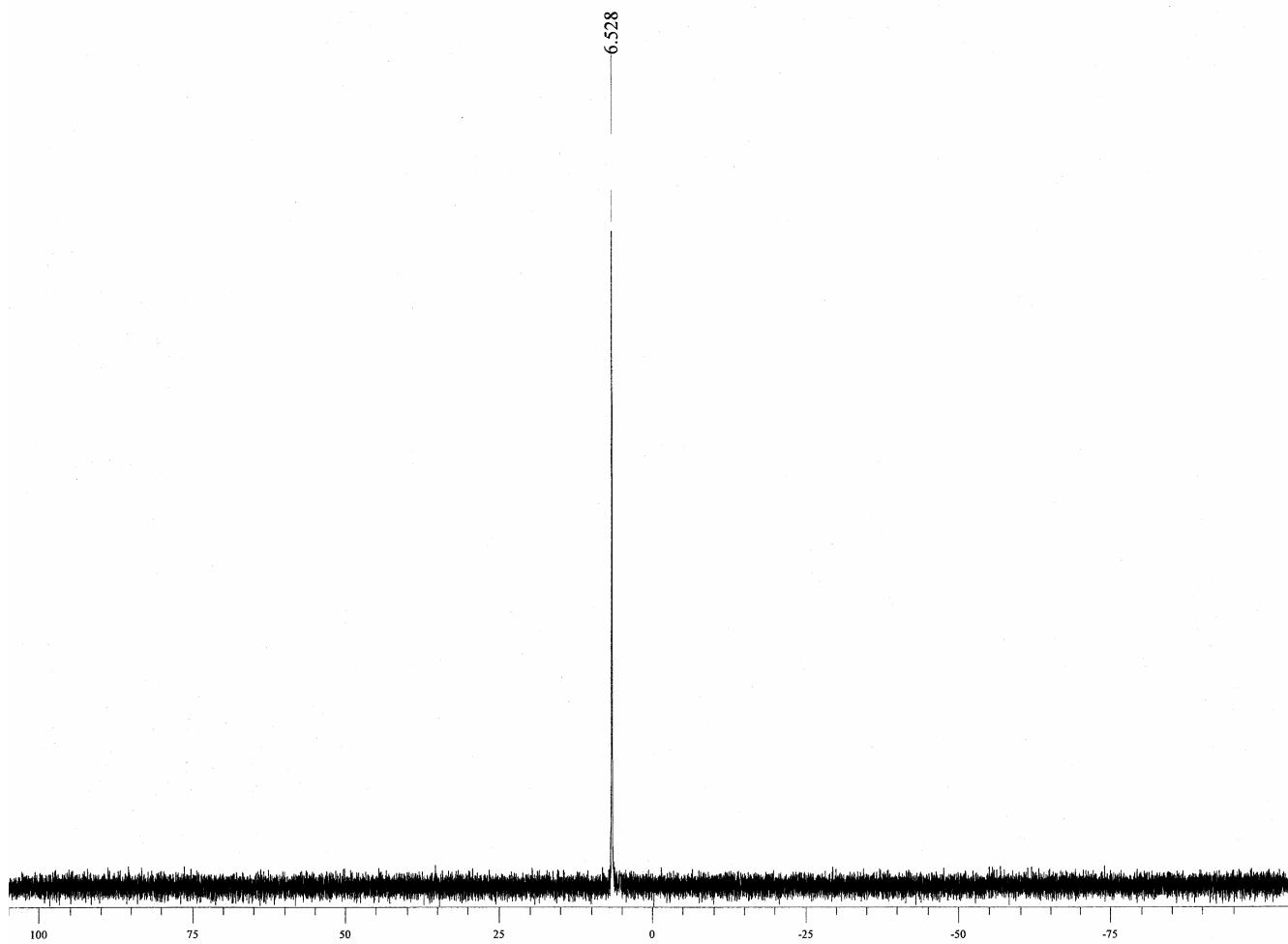


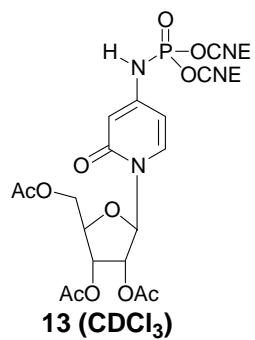
<sup>13</sup>C



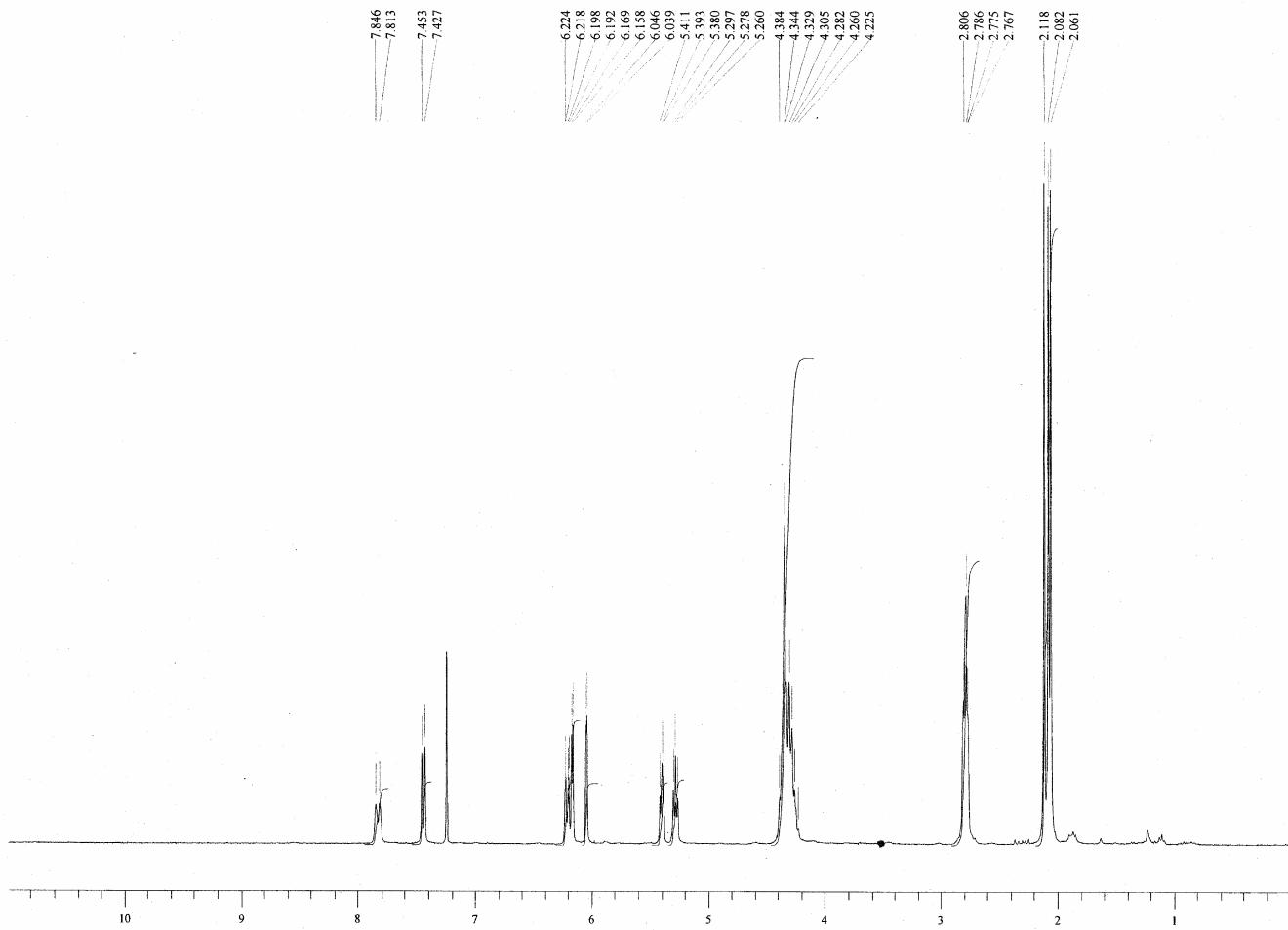


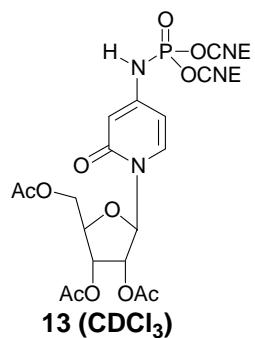
$^{31}\text{P}$



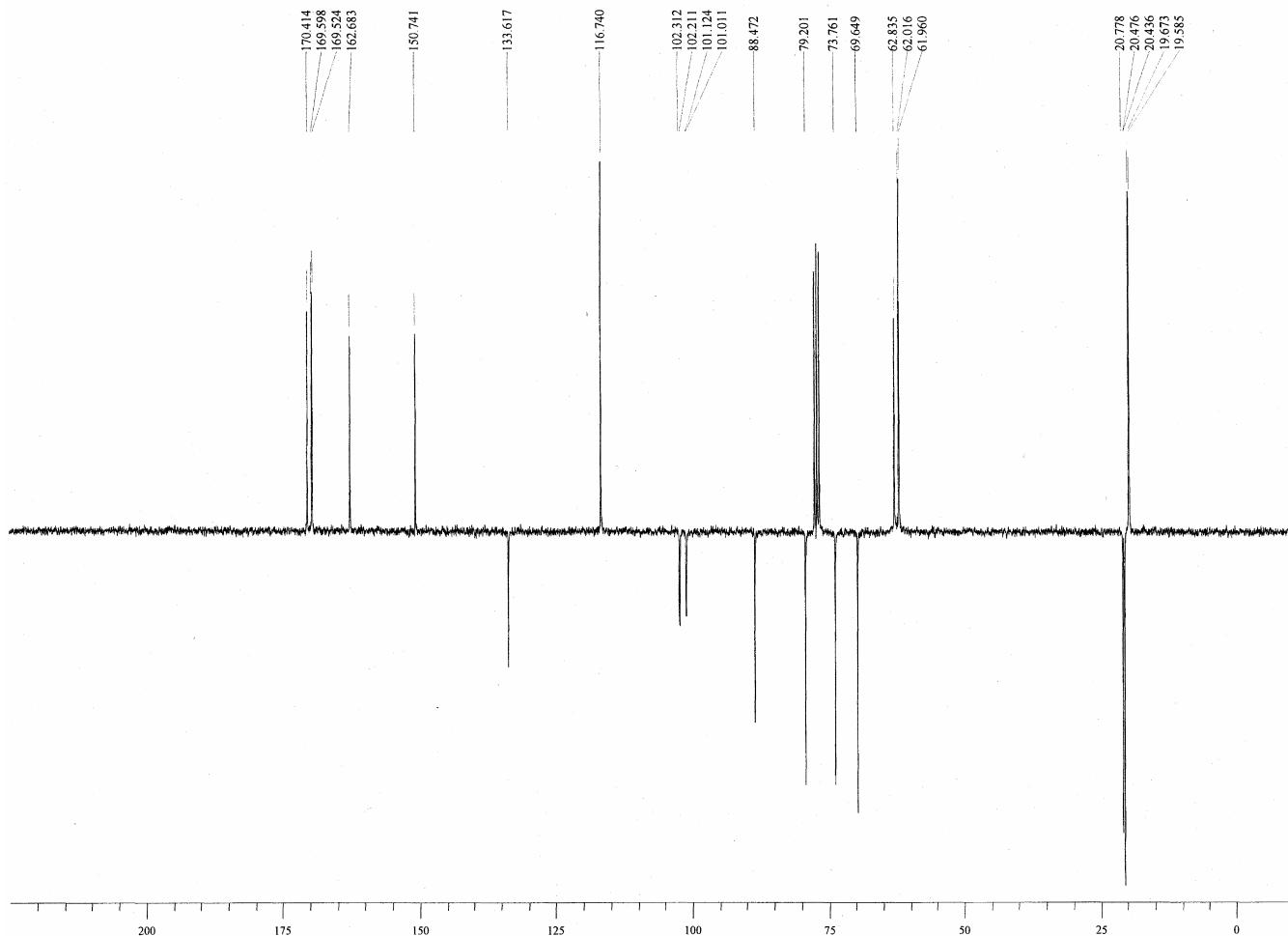


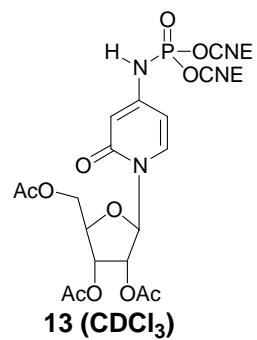
<sup>1</sup>H



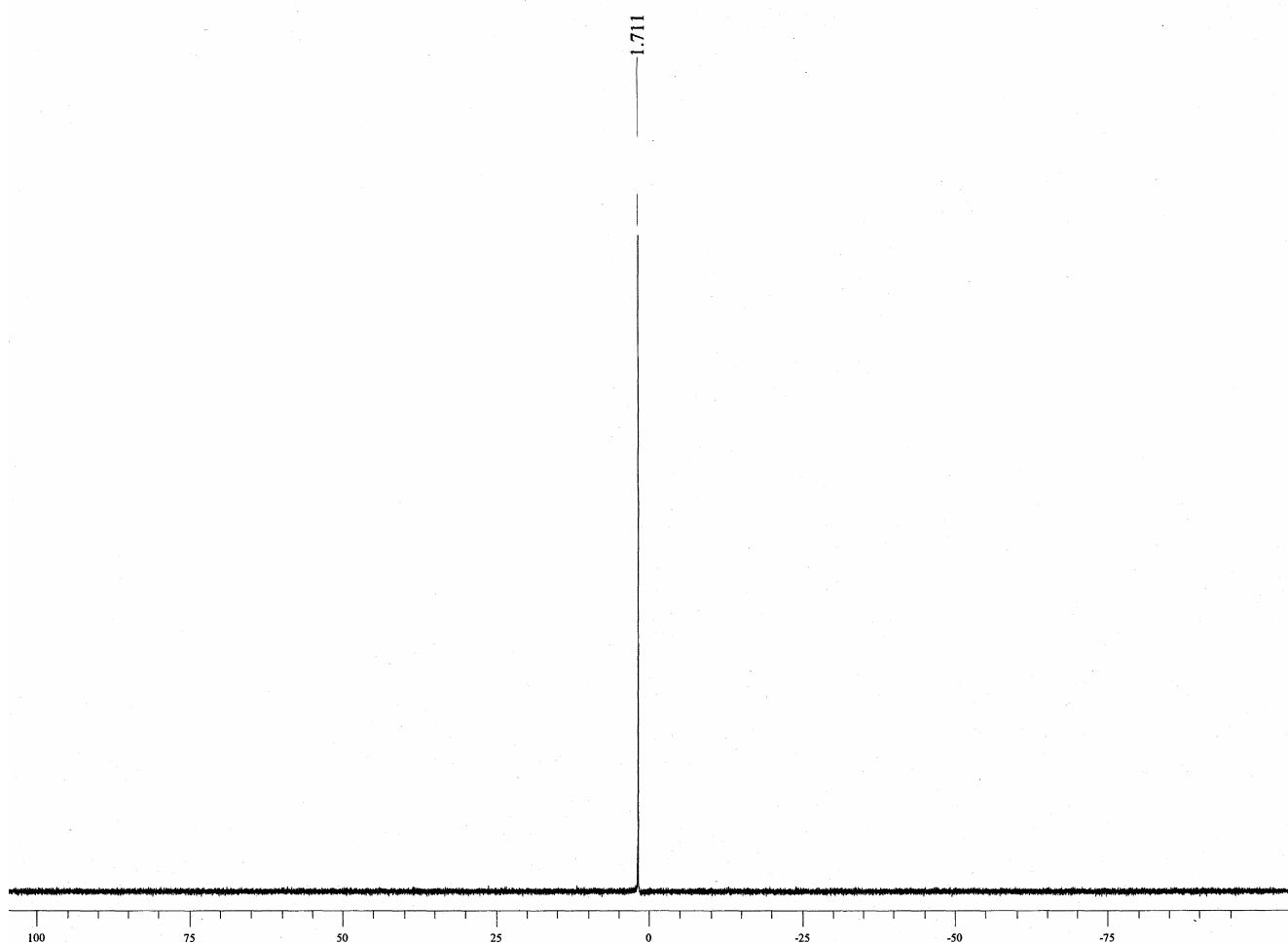


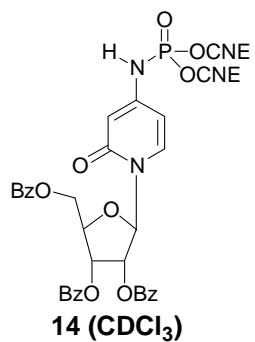
<sup>13</sup>C



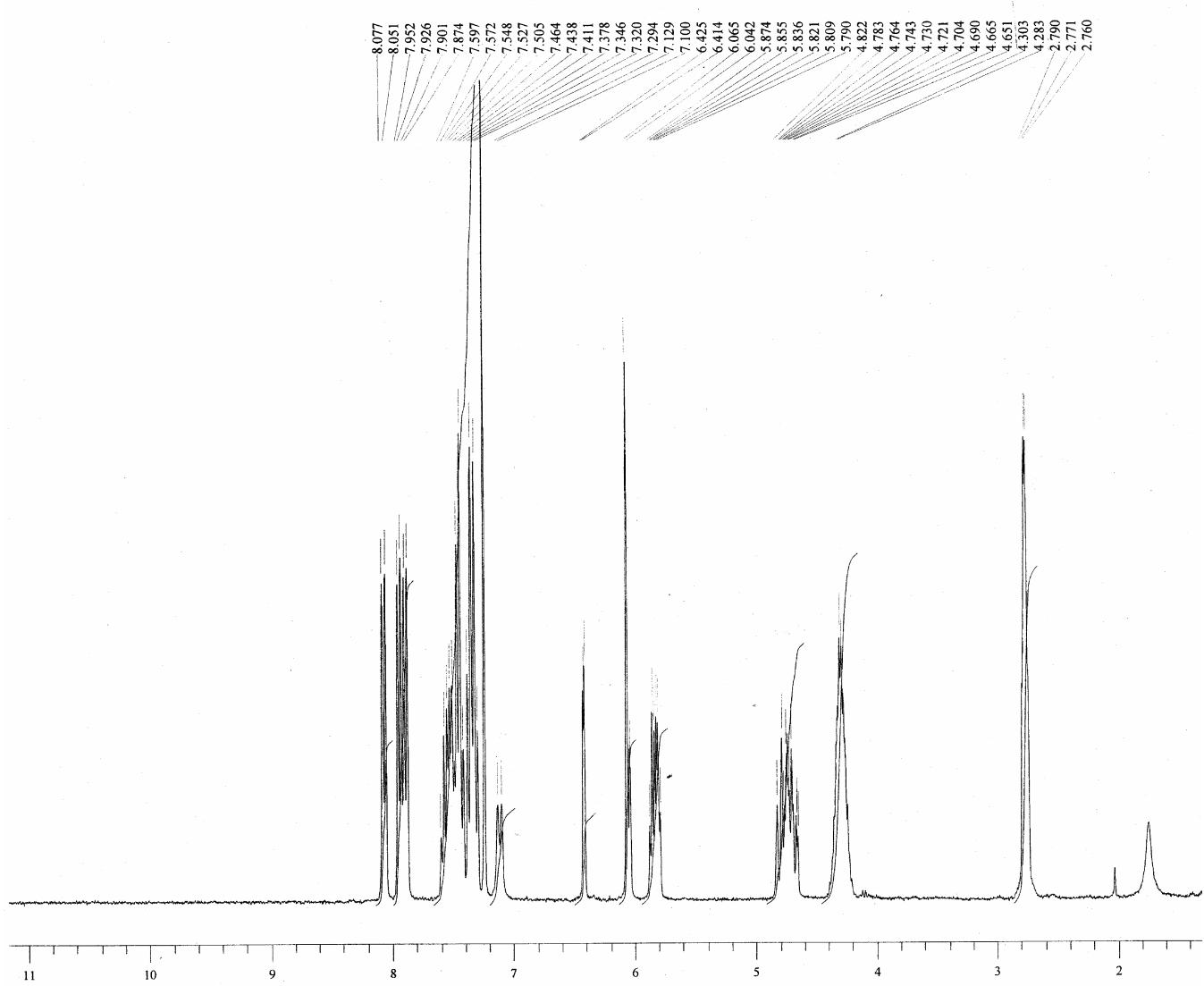


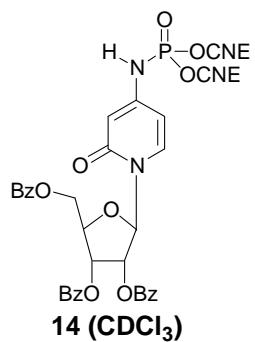
$^{31}\text{P}$



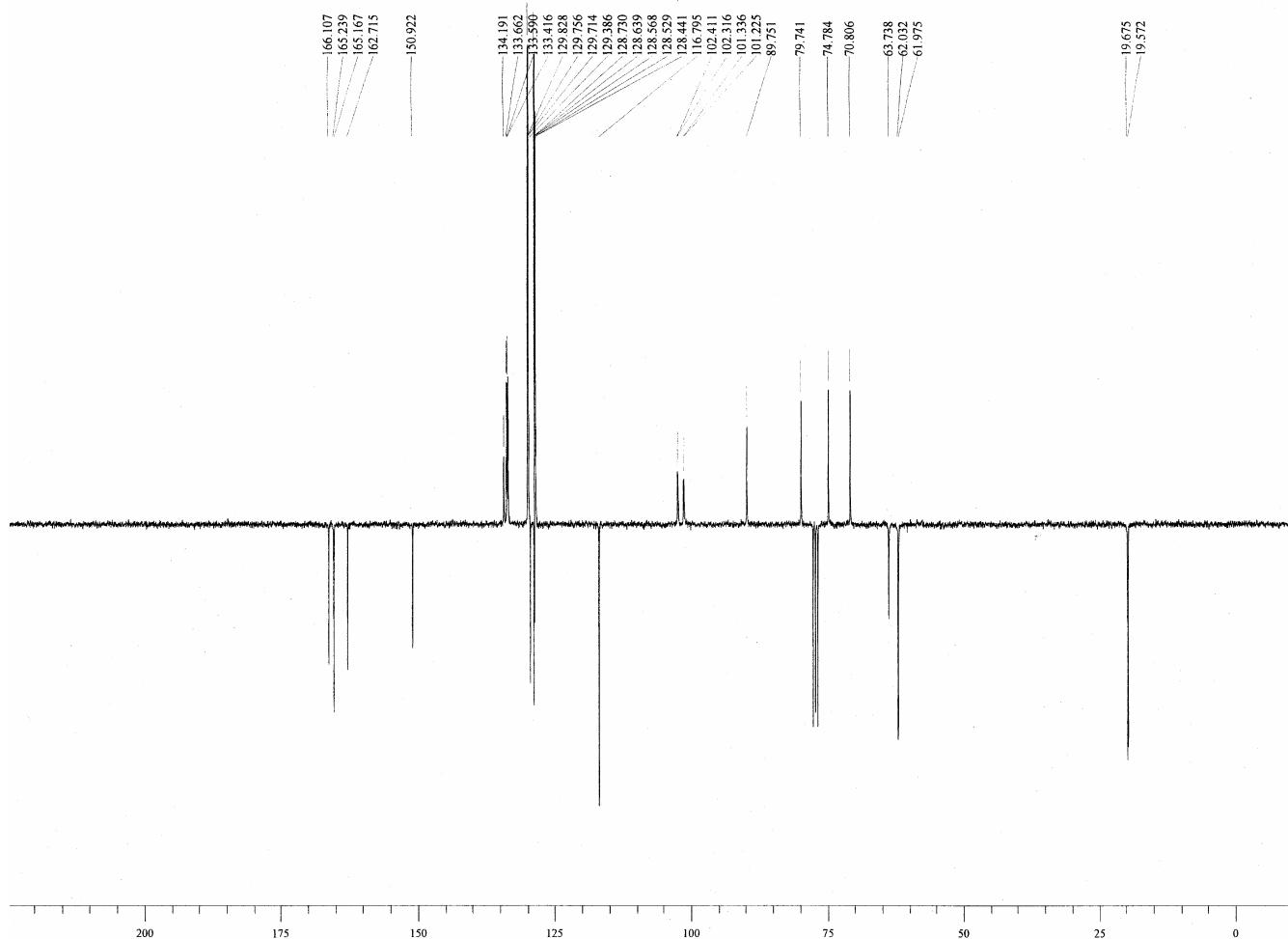


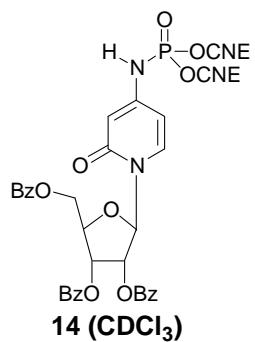
<sup>1</sup>H





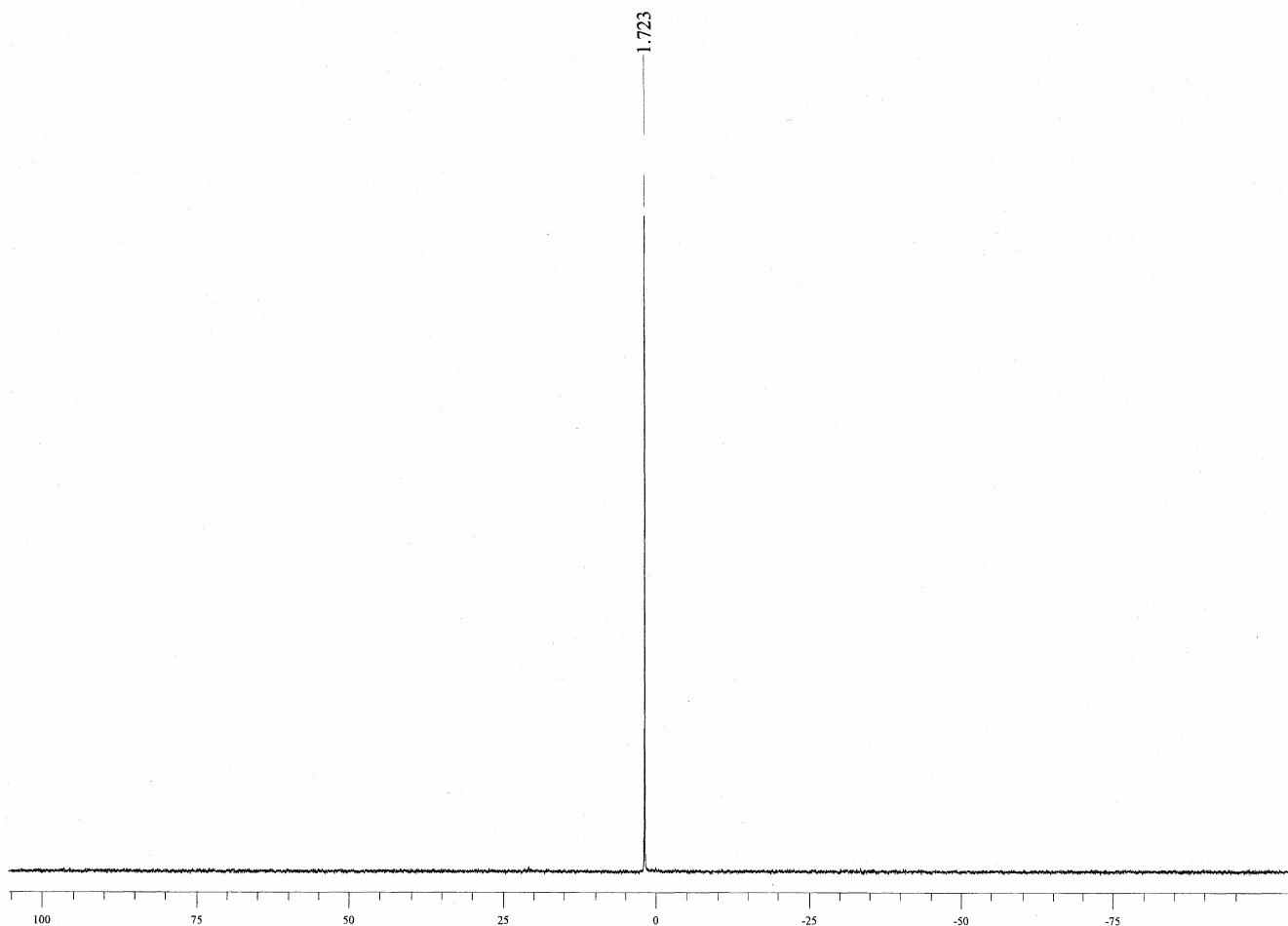
<sup>13</sup>C

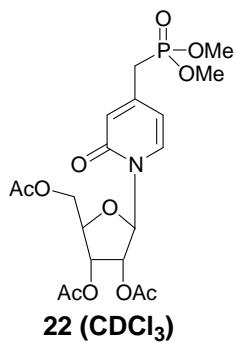




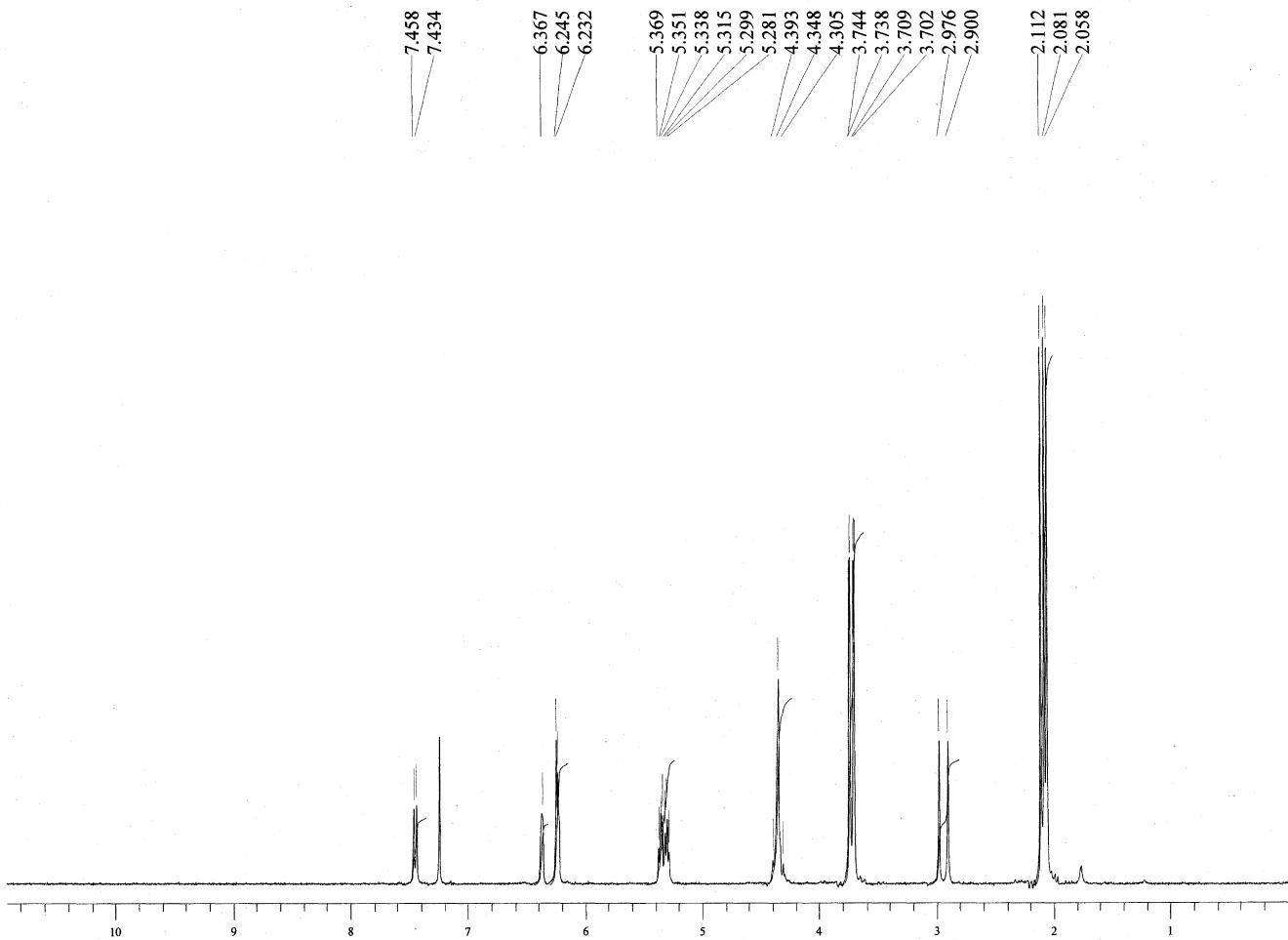
**14 (CDCl<sub>3</sub>)**

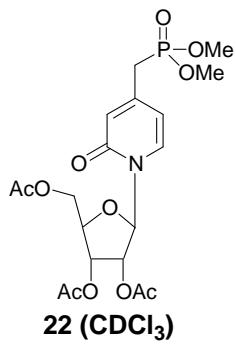
<sup>31</sup>P



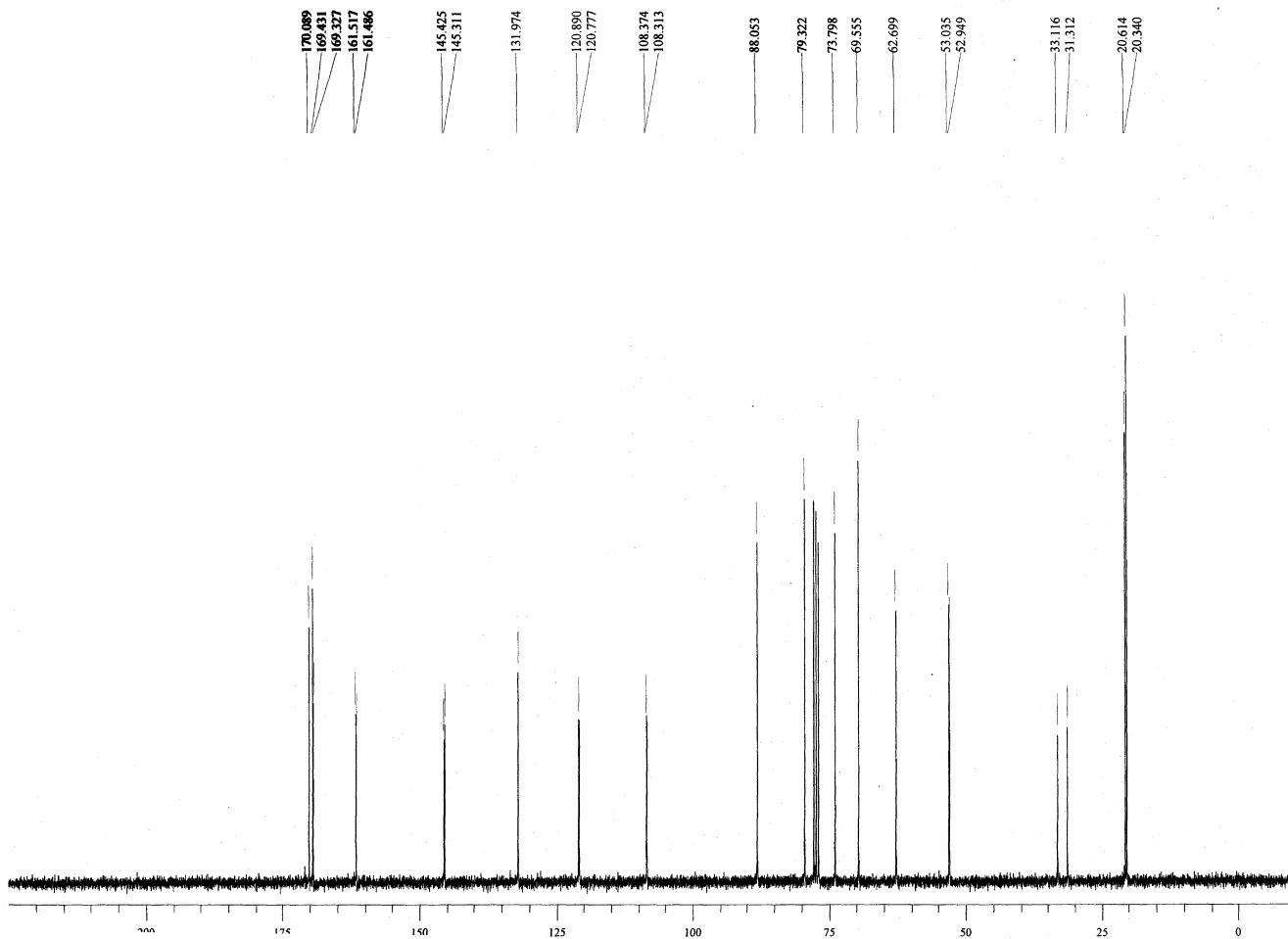


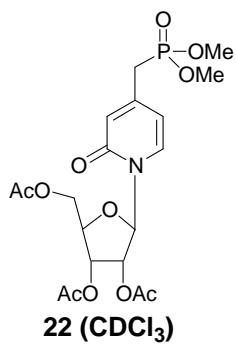
<sup>1</sup>H



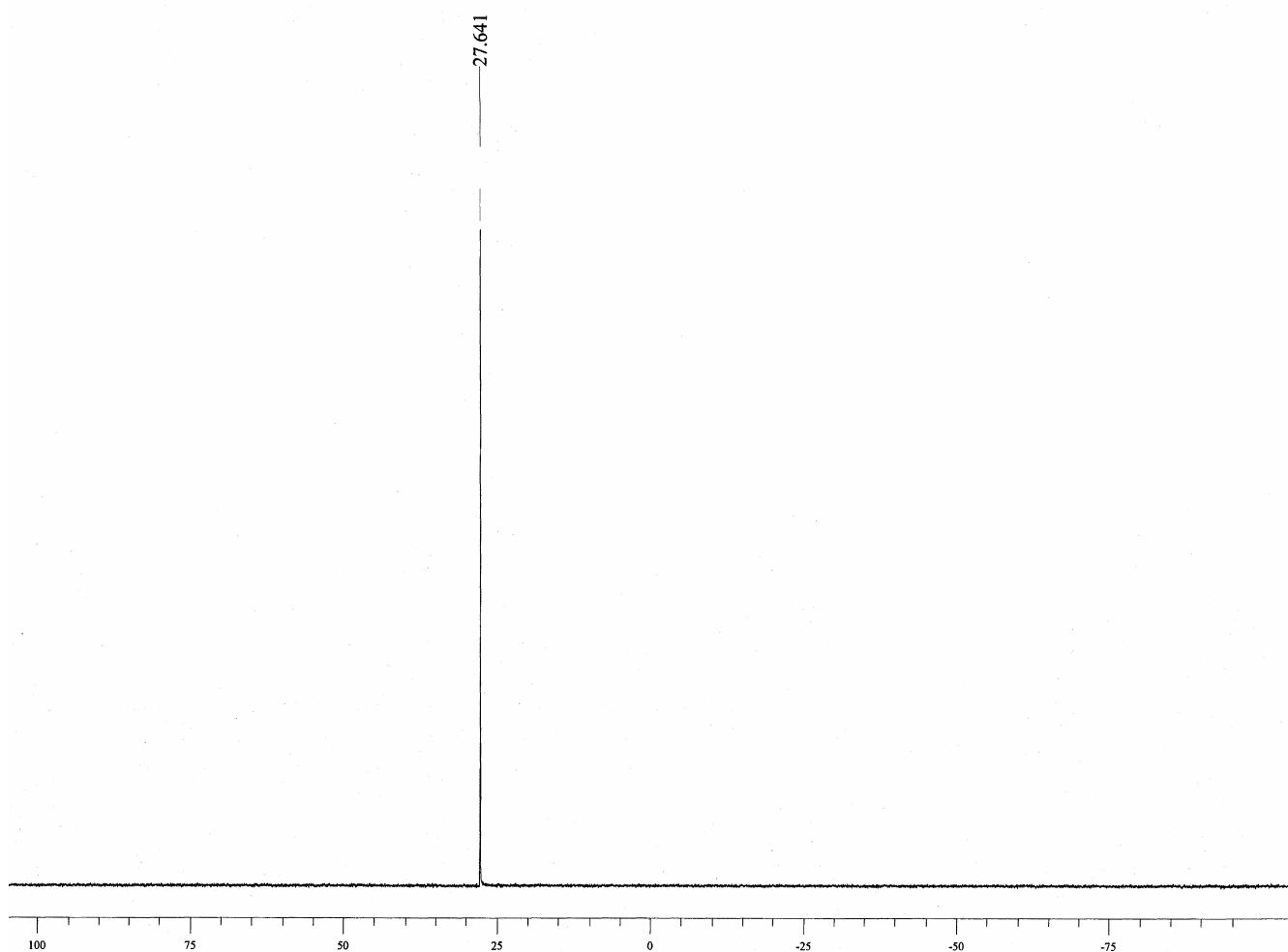


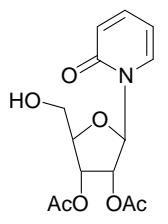
<sup>13</sup>C





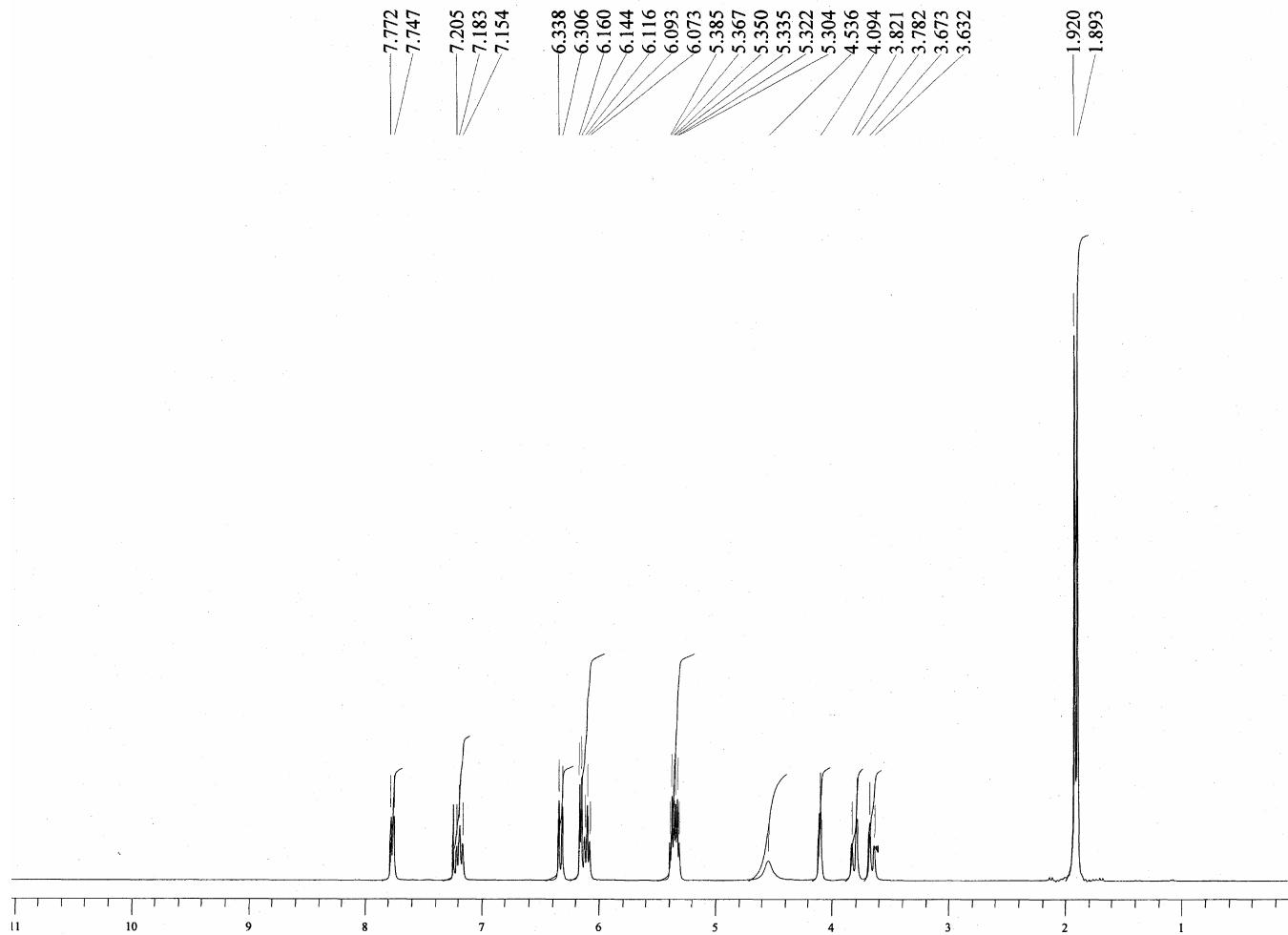
<sup>31</sup>P

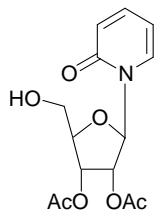




**24 ( $\text{CDCl}_3$ )**

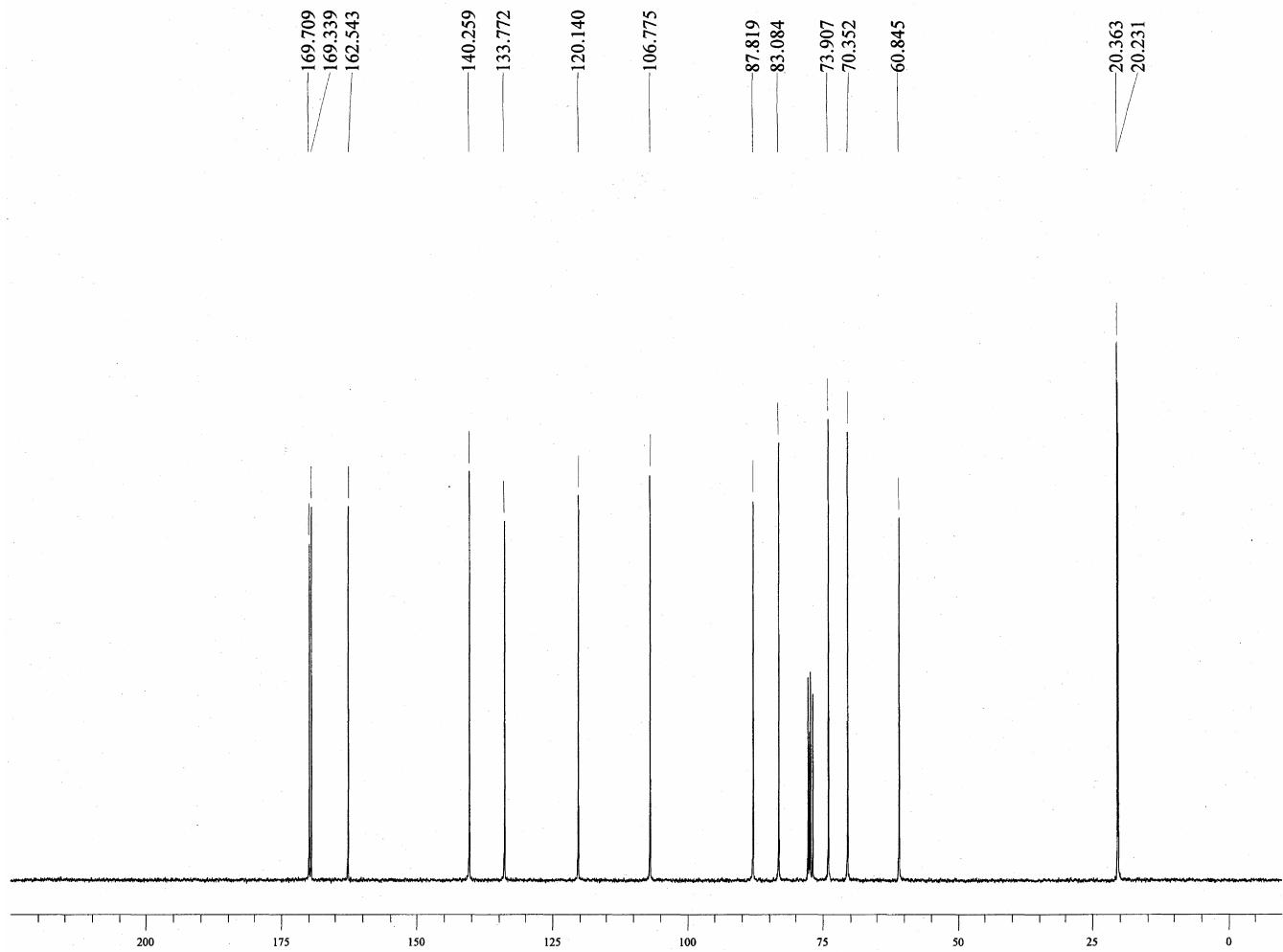
$^1\text{H}$

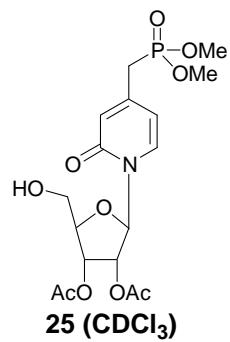




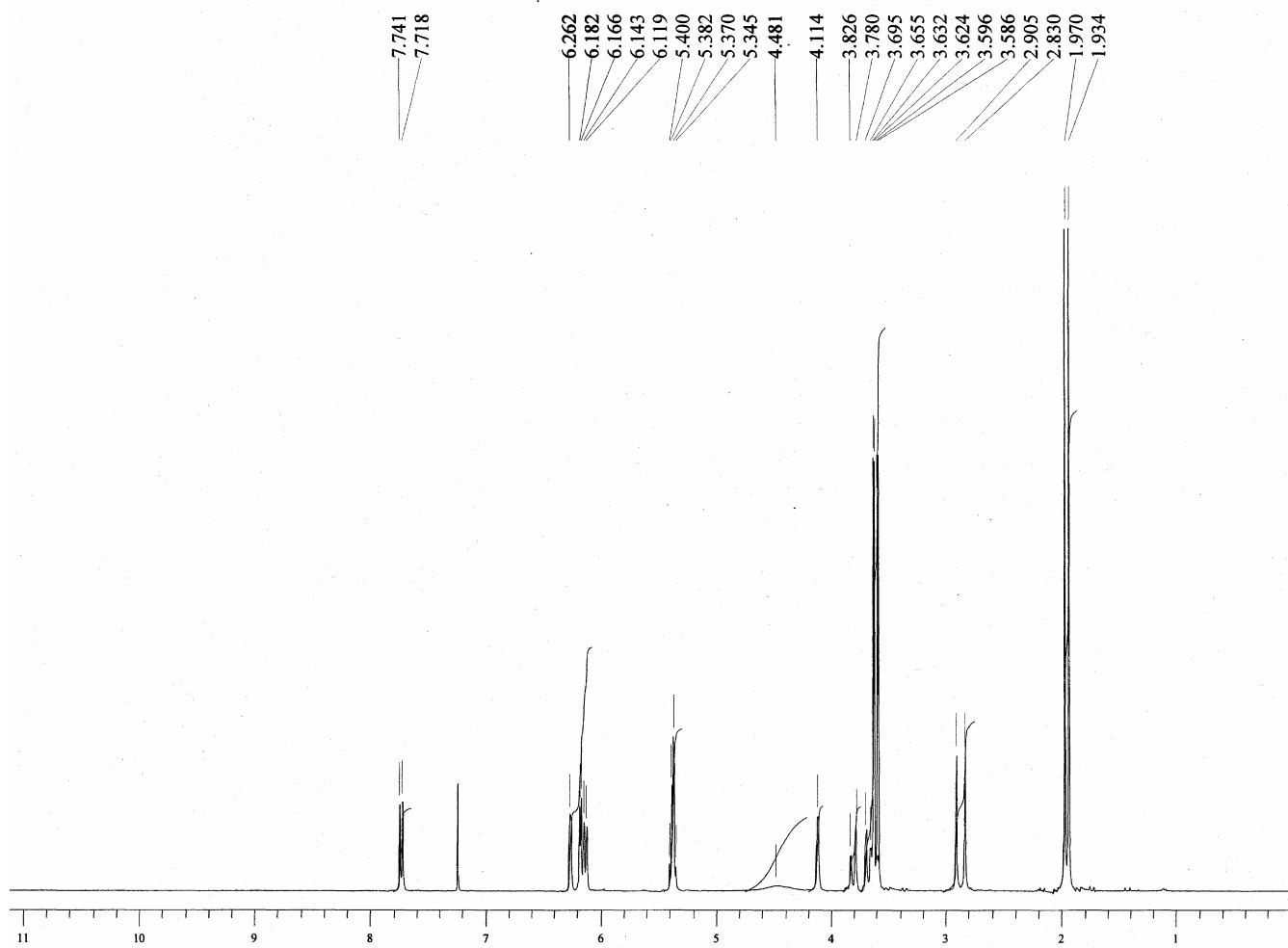
**24 (CDCl<sub>3</sub>)**

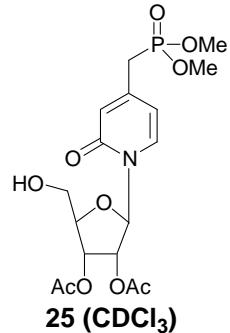
<sup>13</sup>C



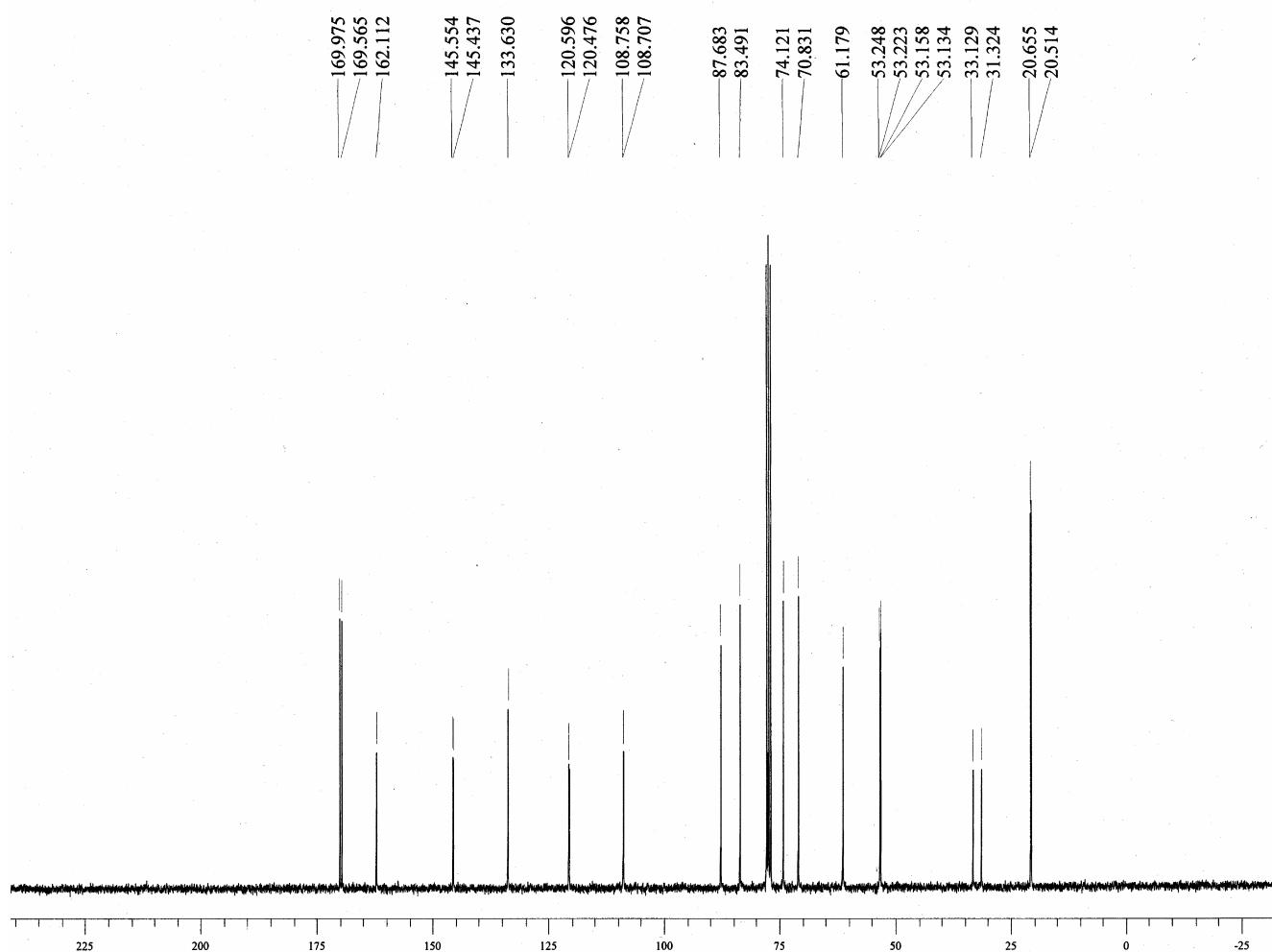


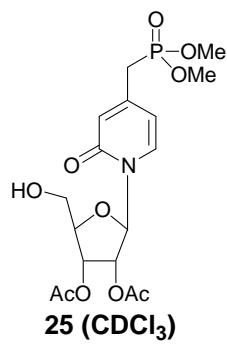
<sup>1</sup>H





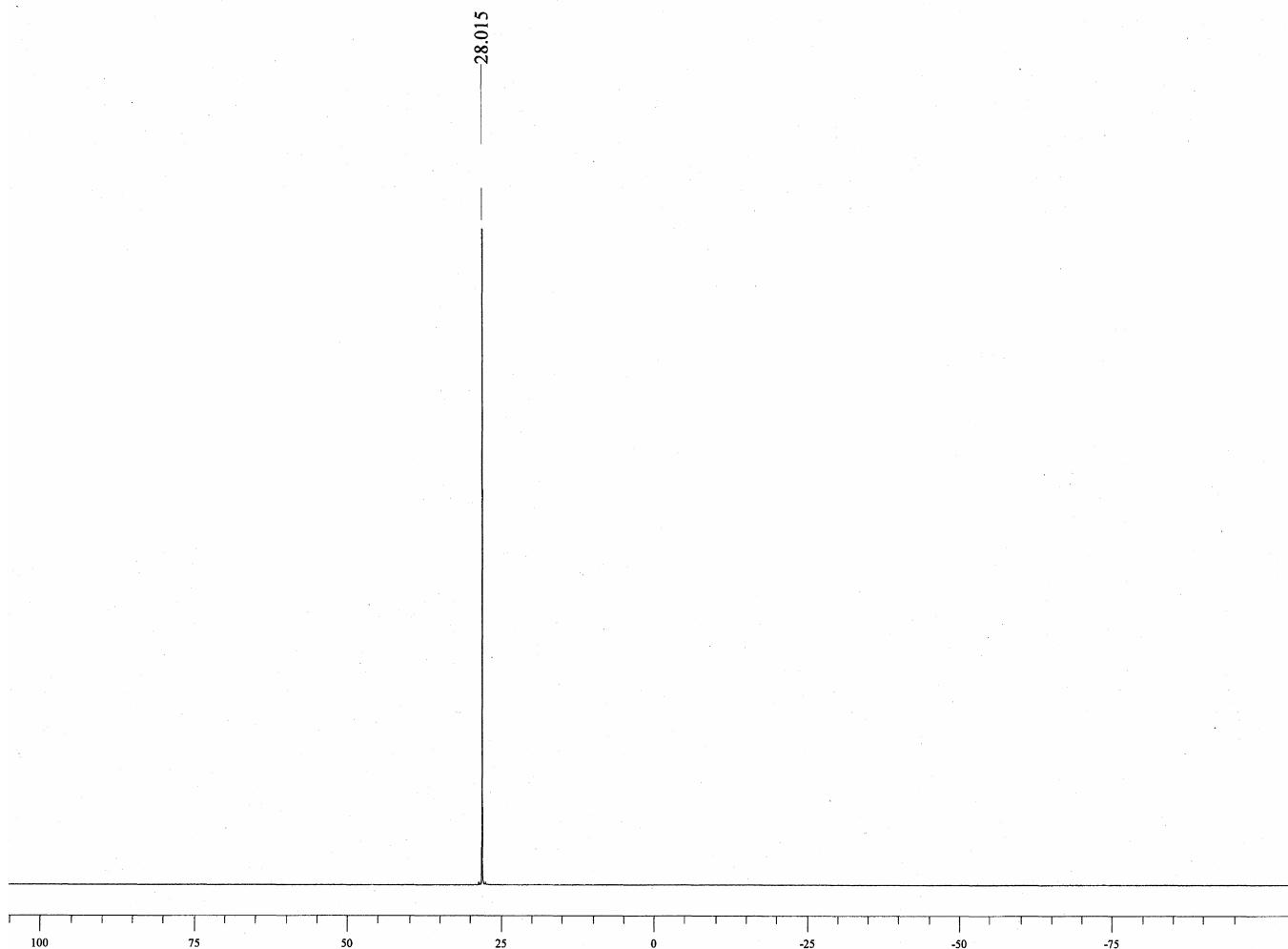
$^{13}\text{C}$

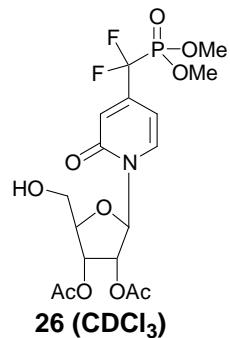




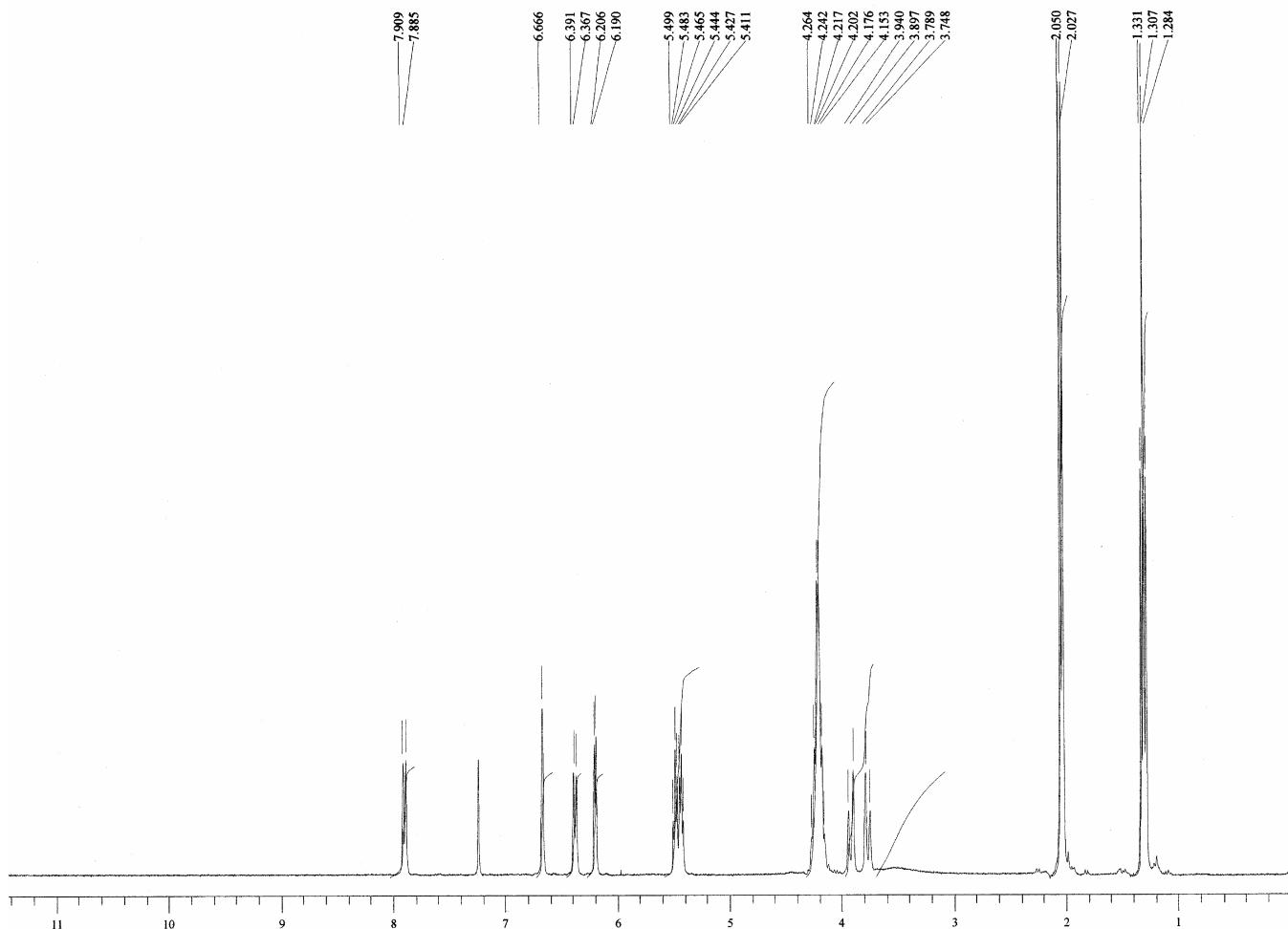
**25 (CDCl<sub>3</sub>)**

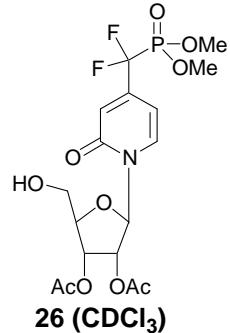
<sup>31</sup>P



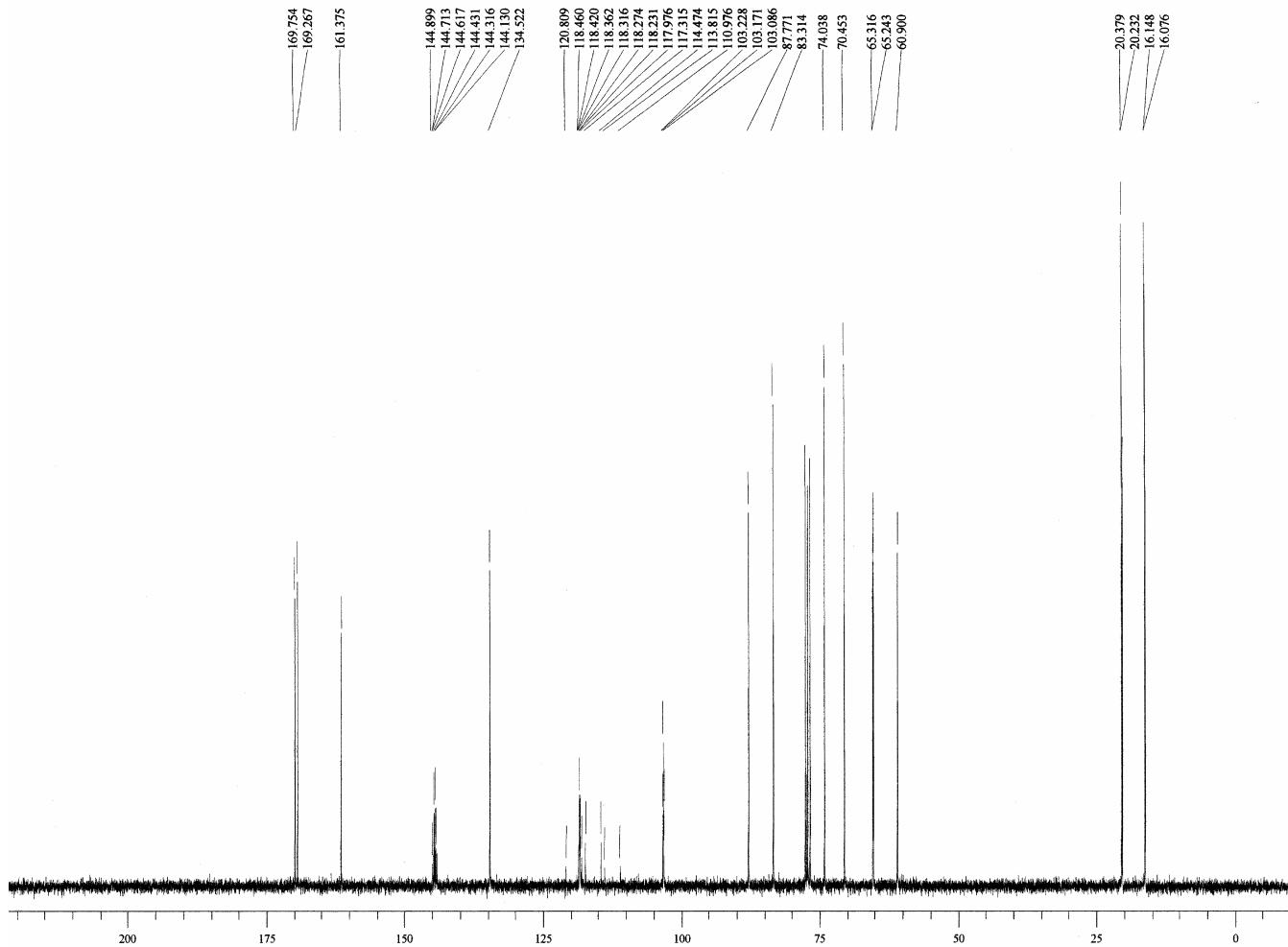


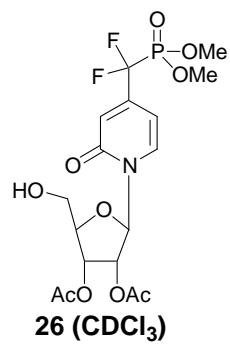
$^1\text{H}$



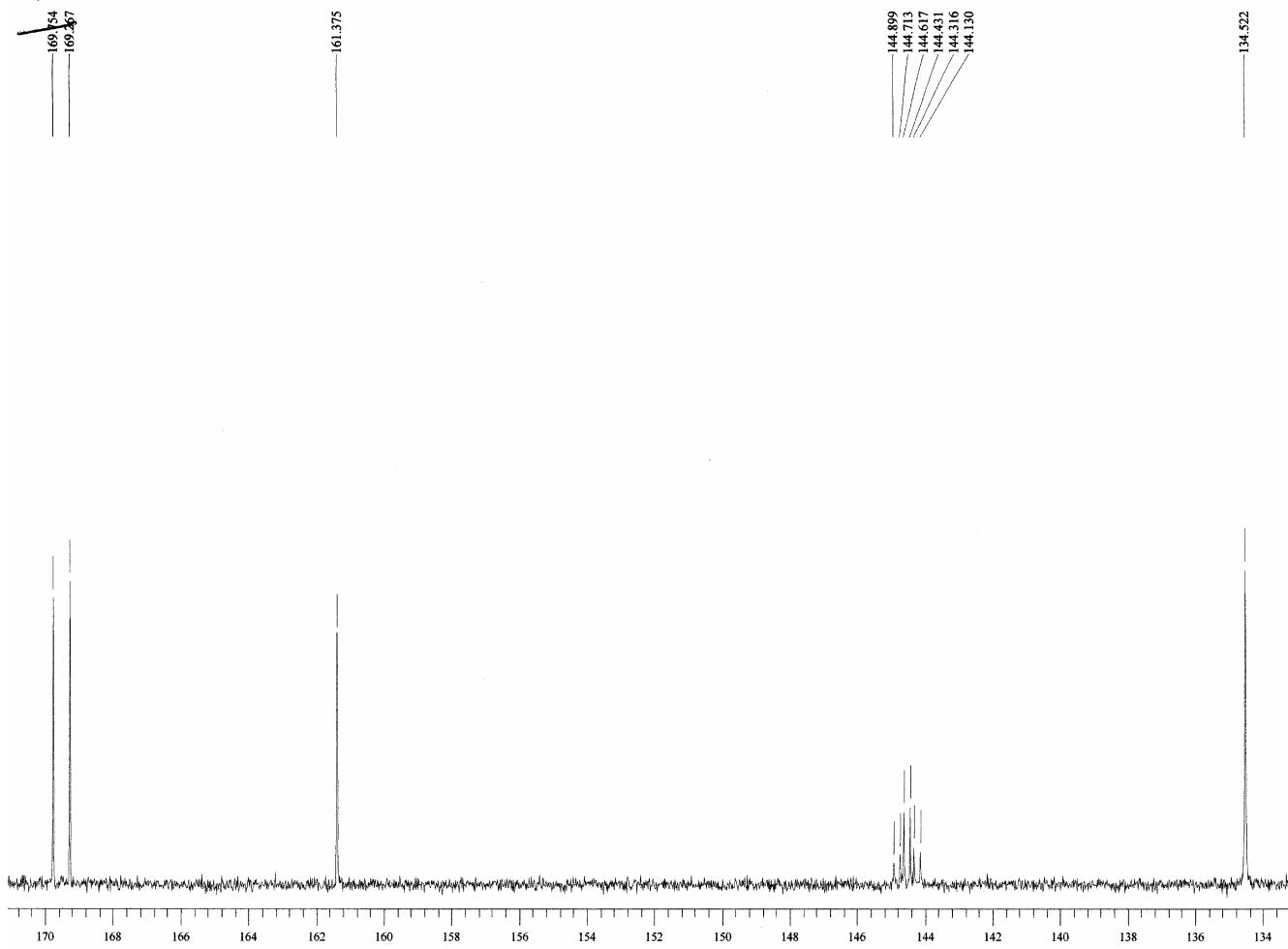


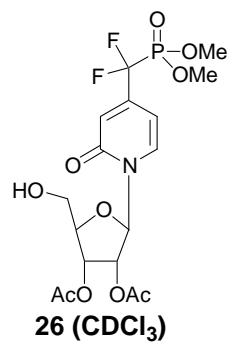
<sup>13</sup>C



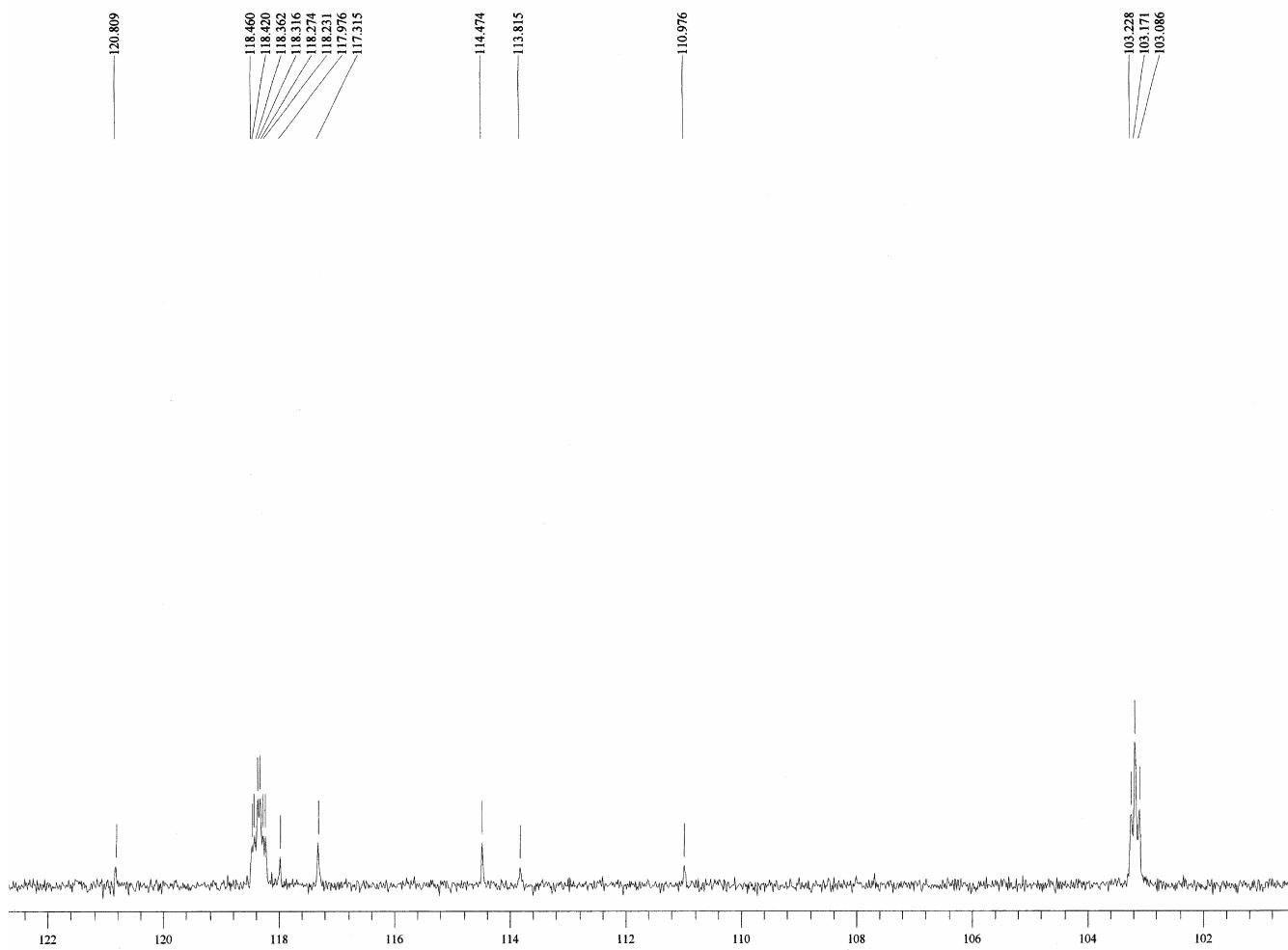


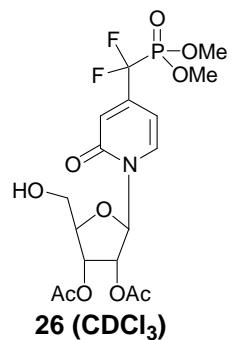
<sup>13</sup>C (expanded)





$^{13}\text{C}$  (expanded)

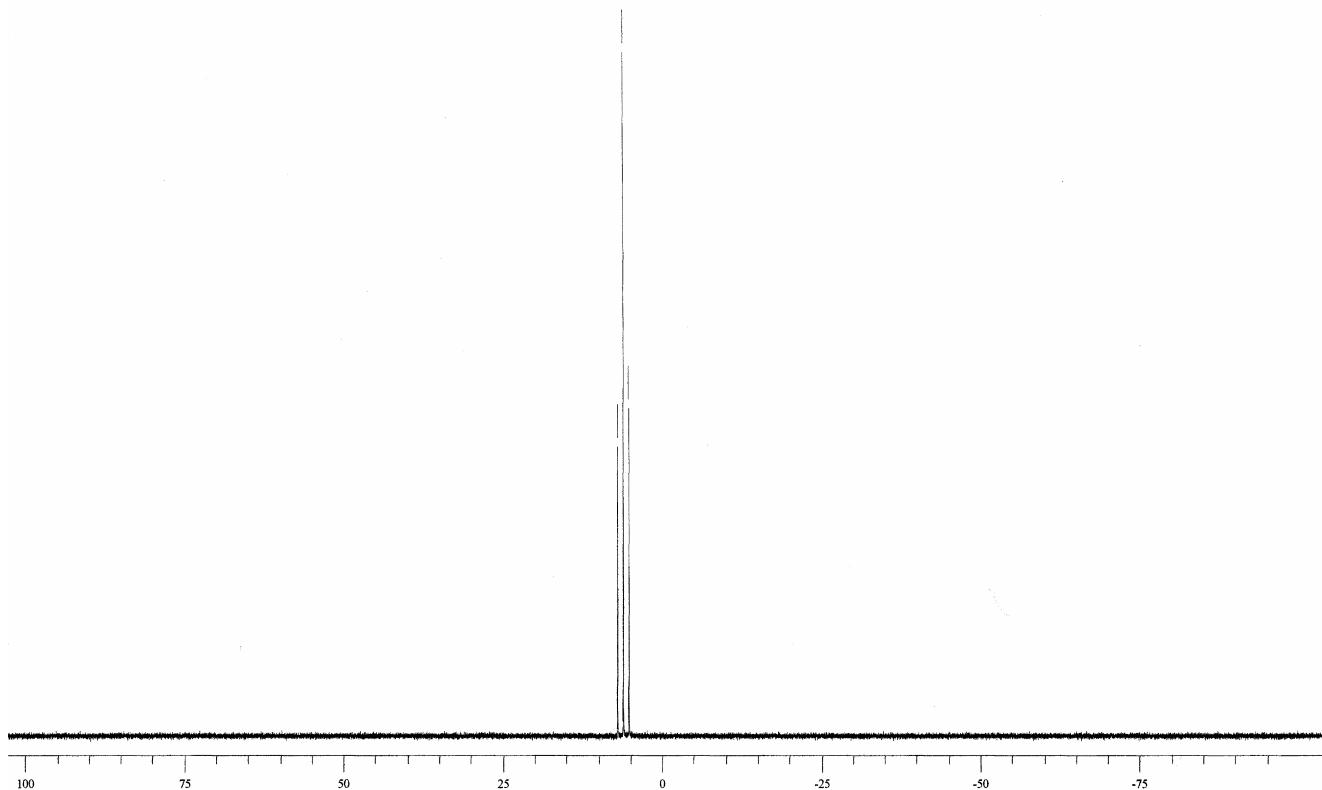


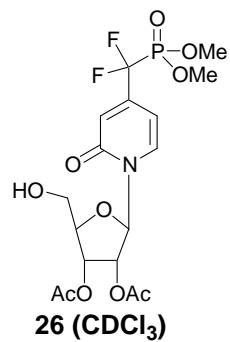


**26 (CDCl<sub>3</sub>)**

<sup>31</sup>P

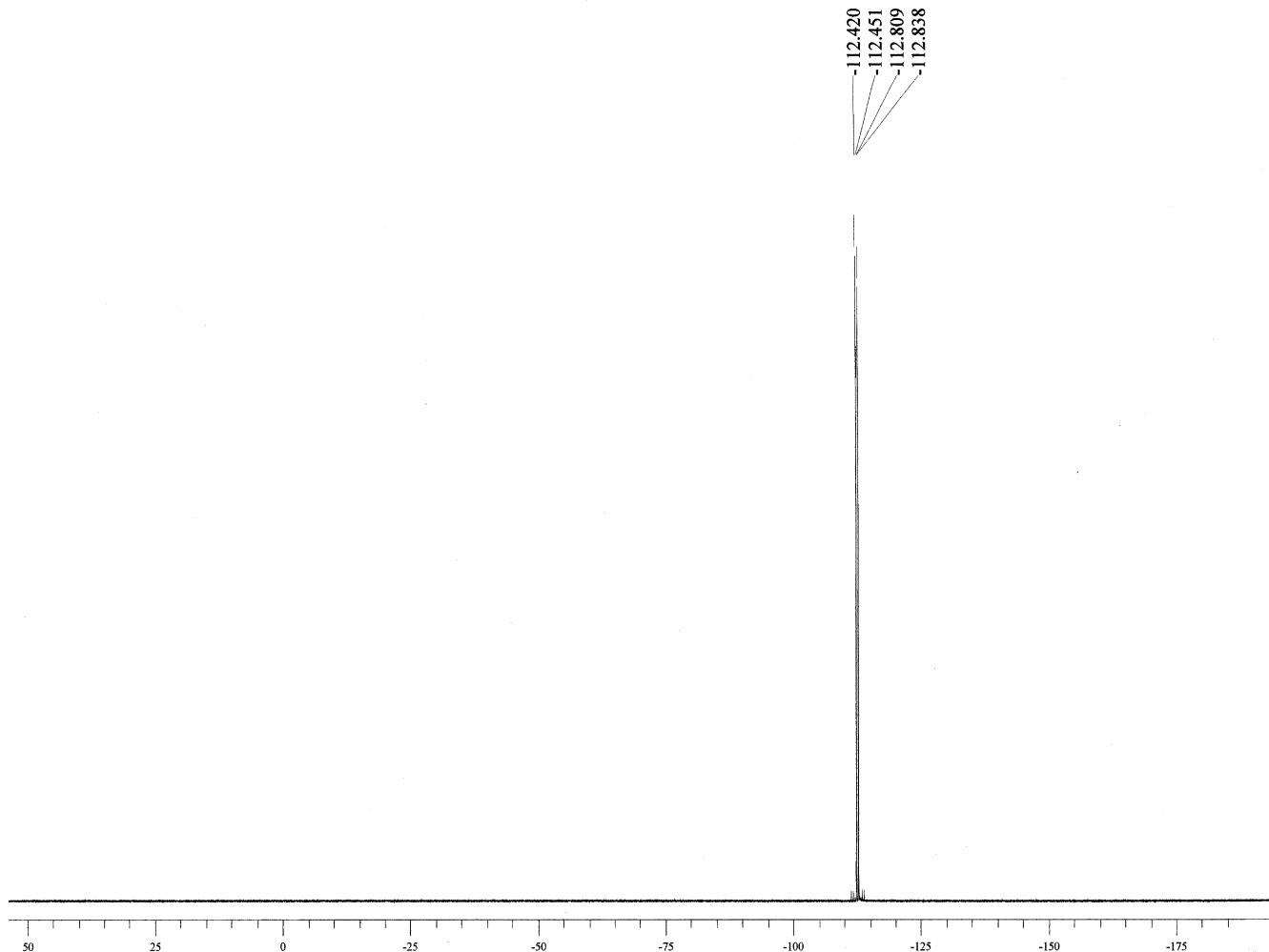
6.964  
6.068  
5.171

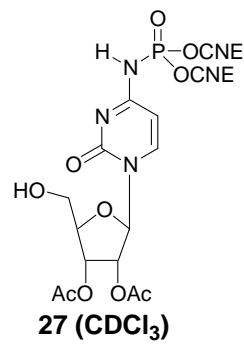




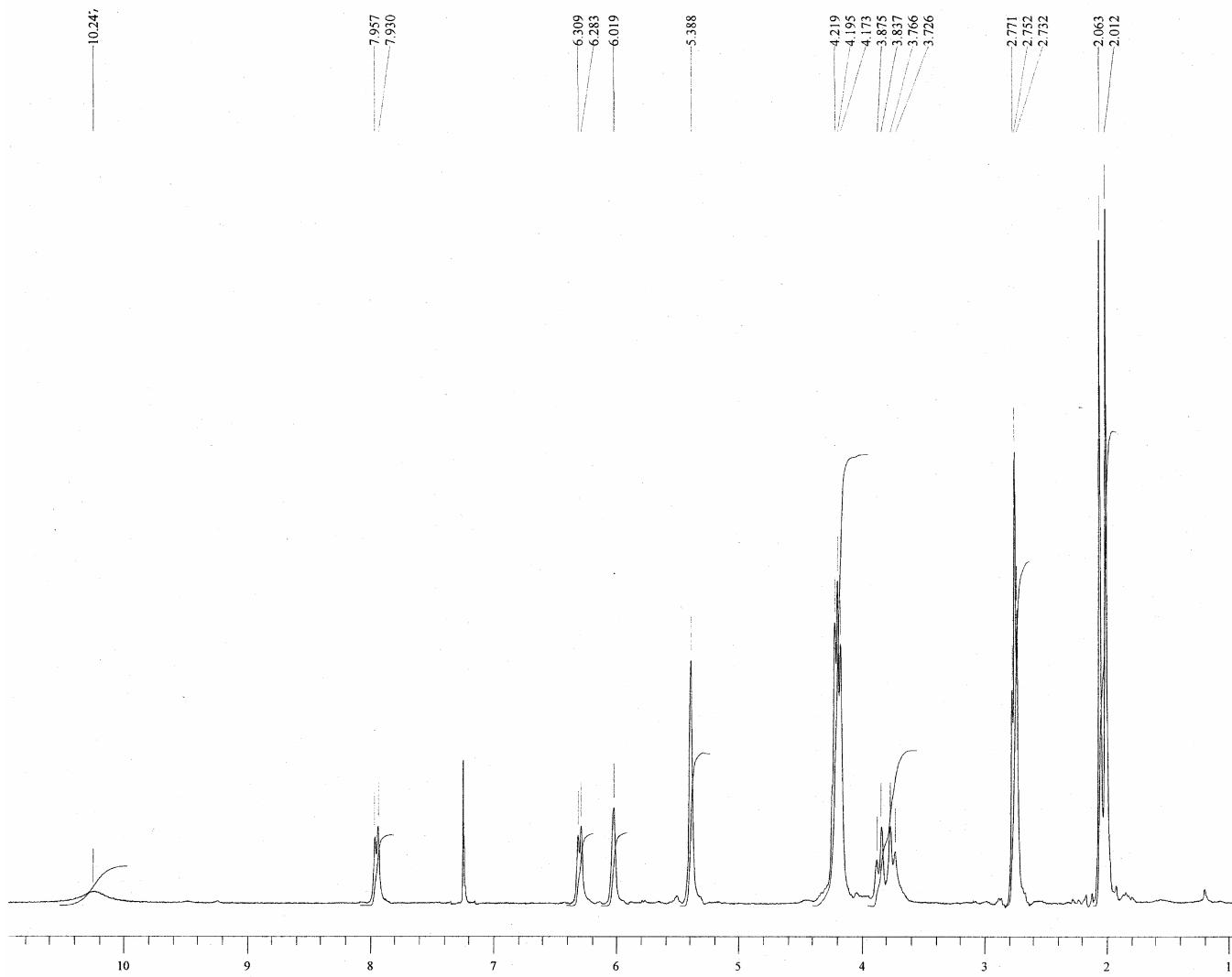
**26 (CDCl<sub>3</sub>)**

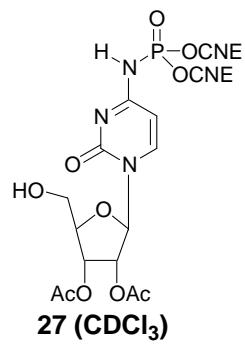
<sup>19</sup>F



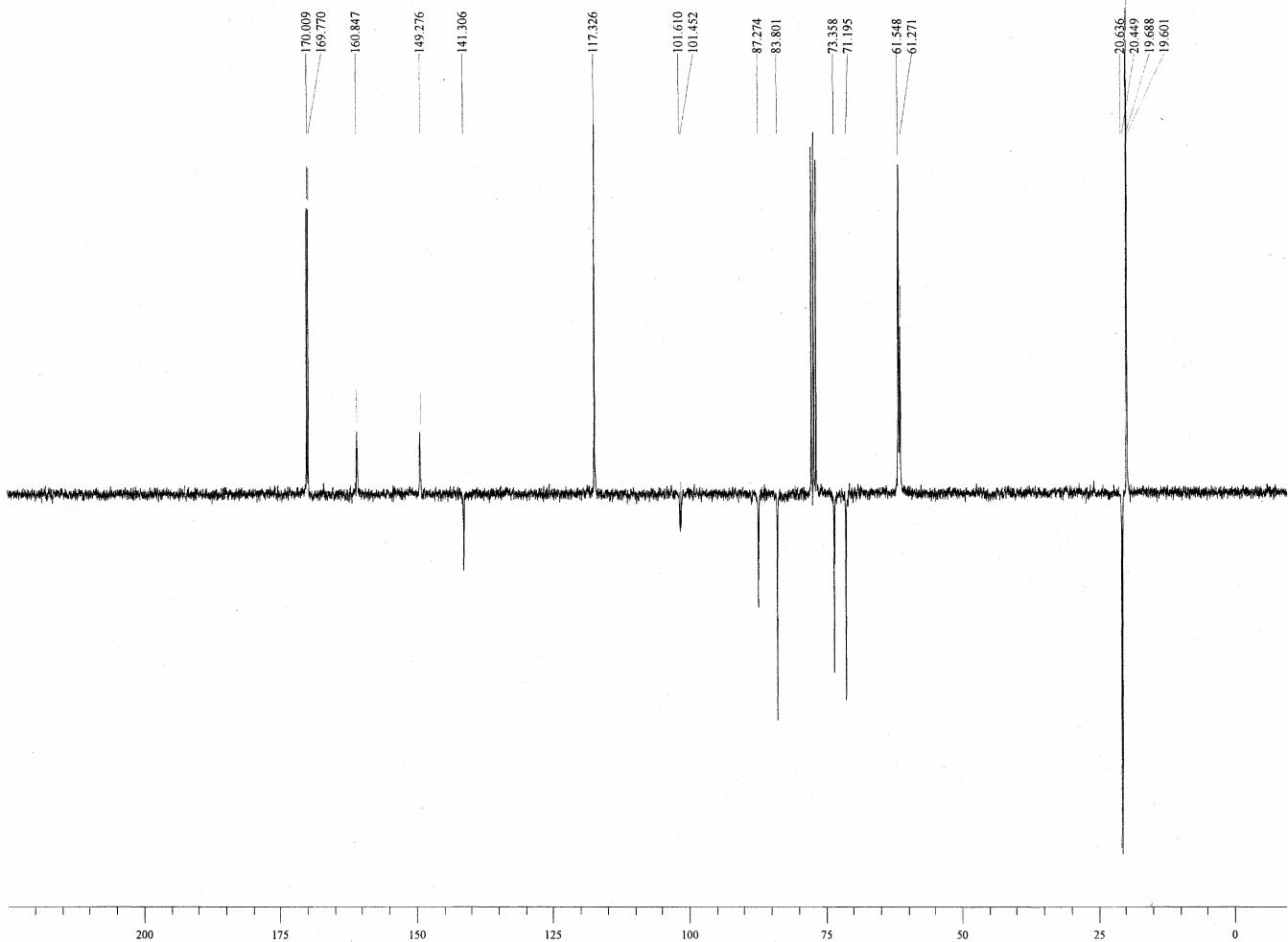


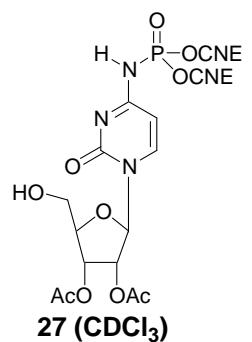
$^1\text{H}$



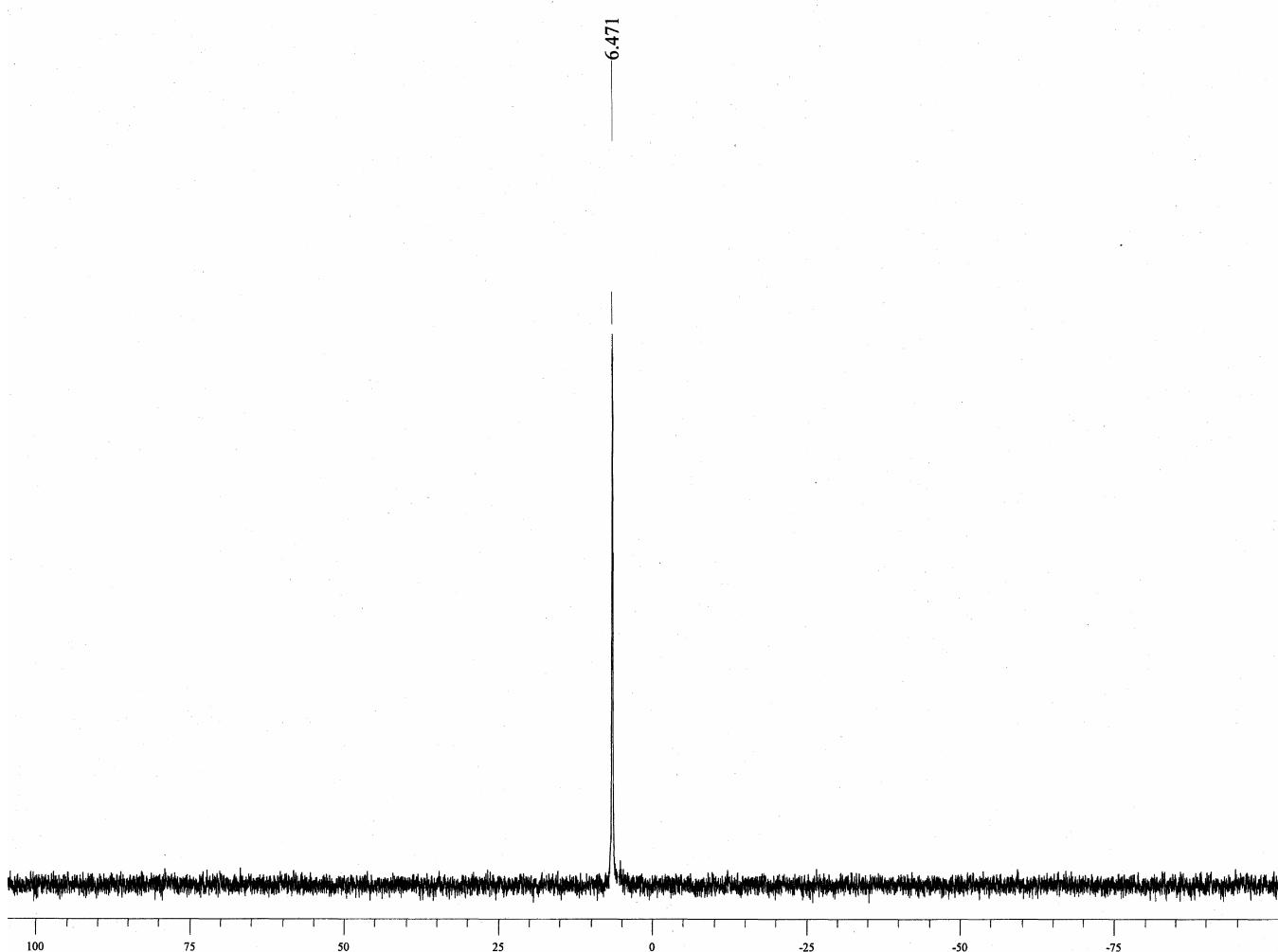


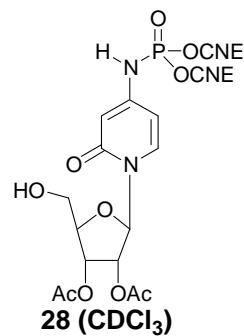
<sup>13</sup>C



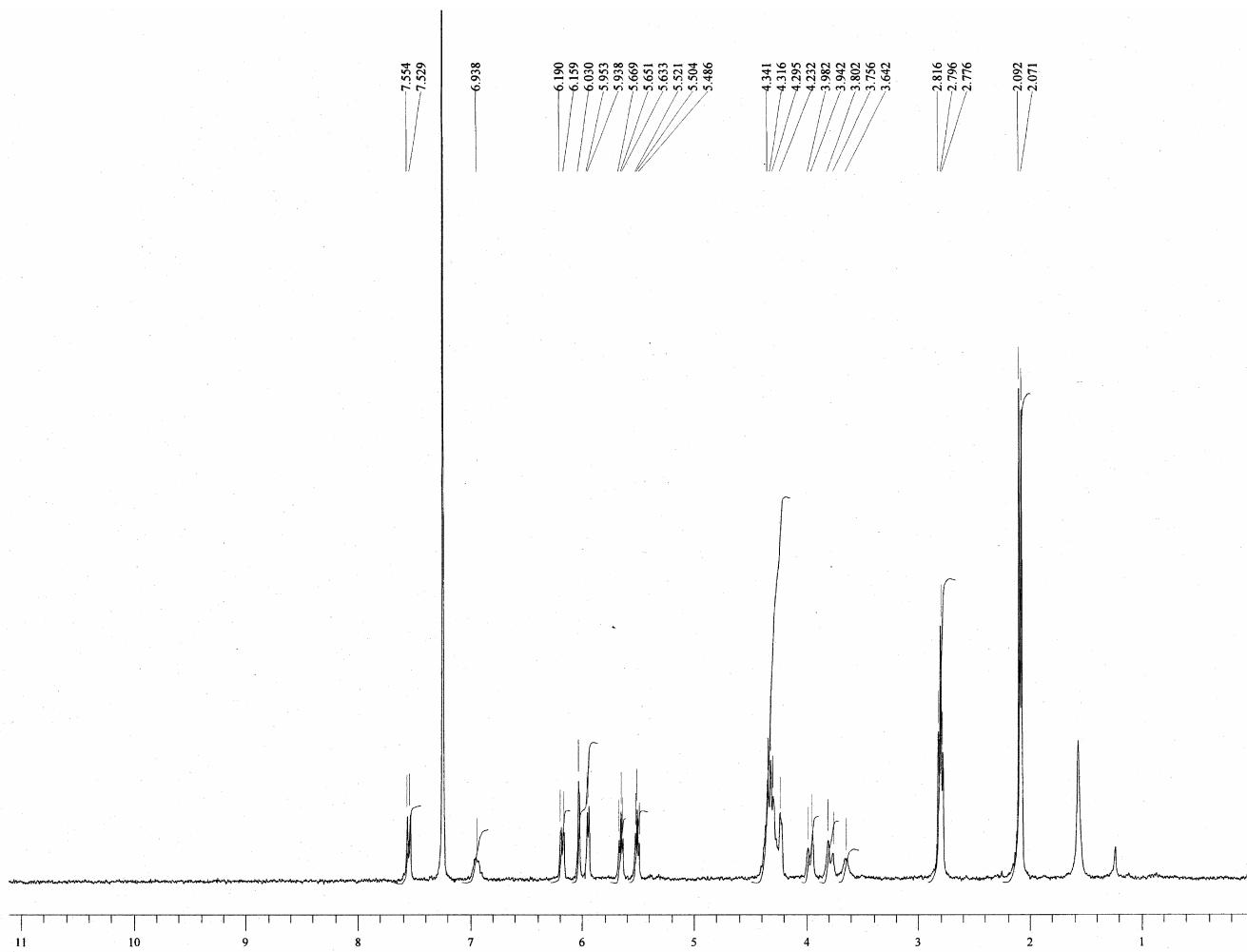


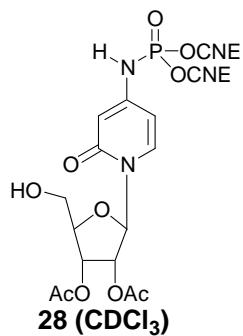
<sup>31</sup>P



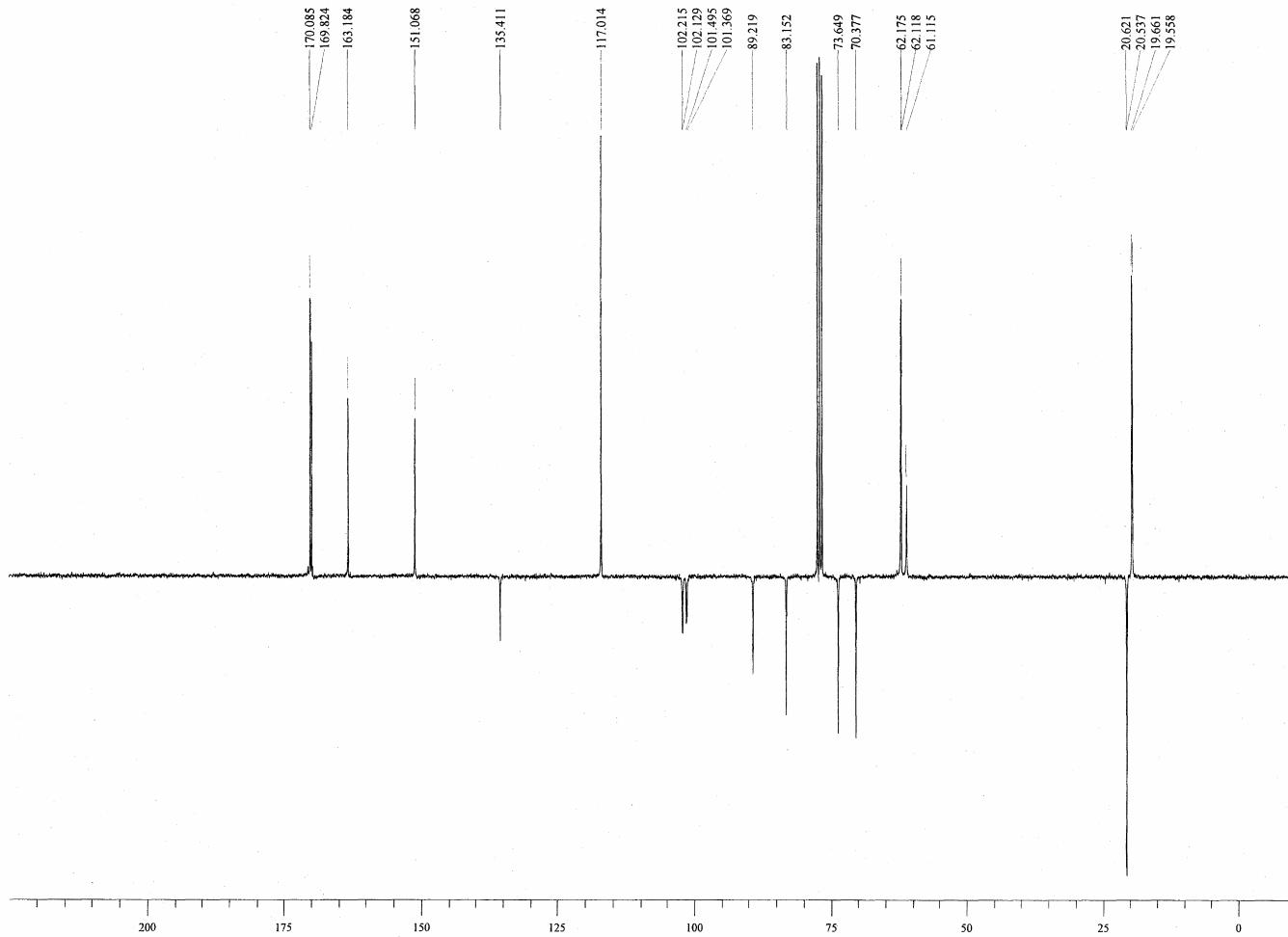


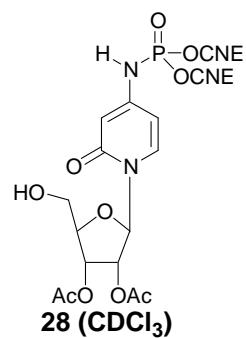
<sup>1</sup>H



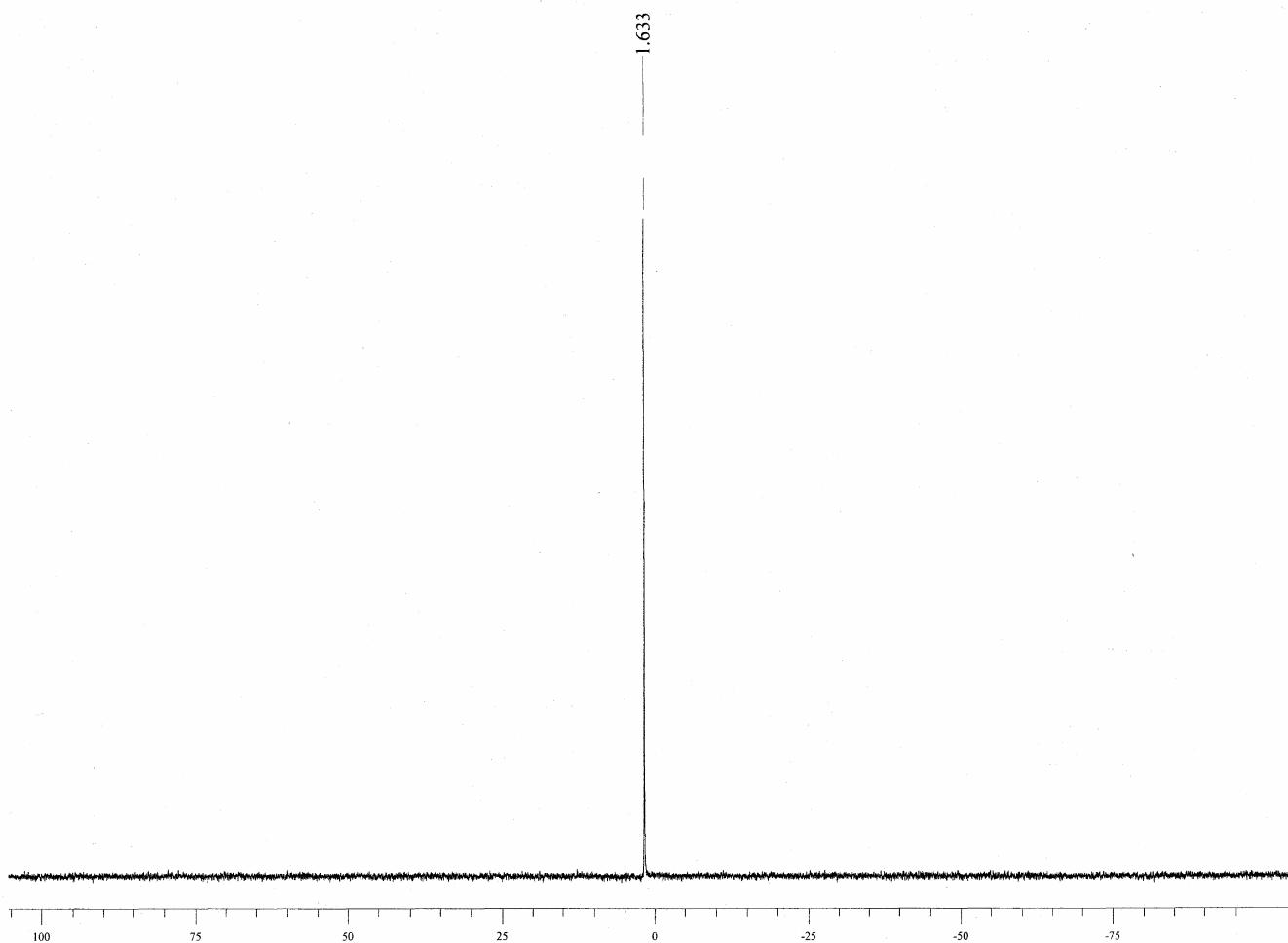


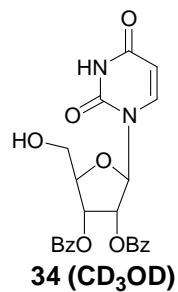
<sup>13</sup>C



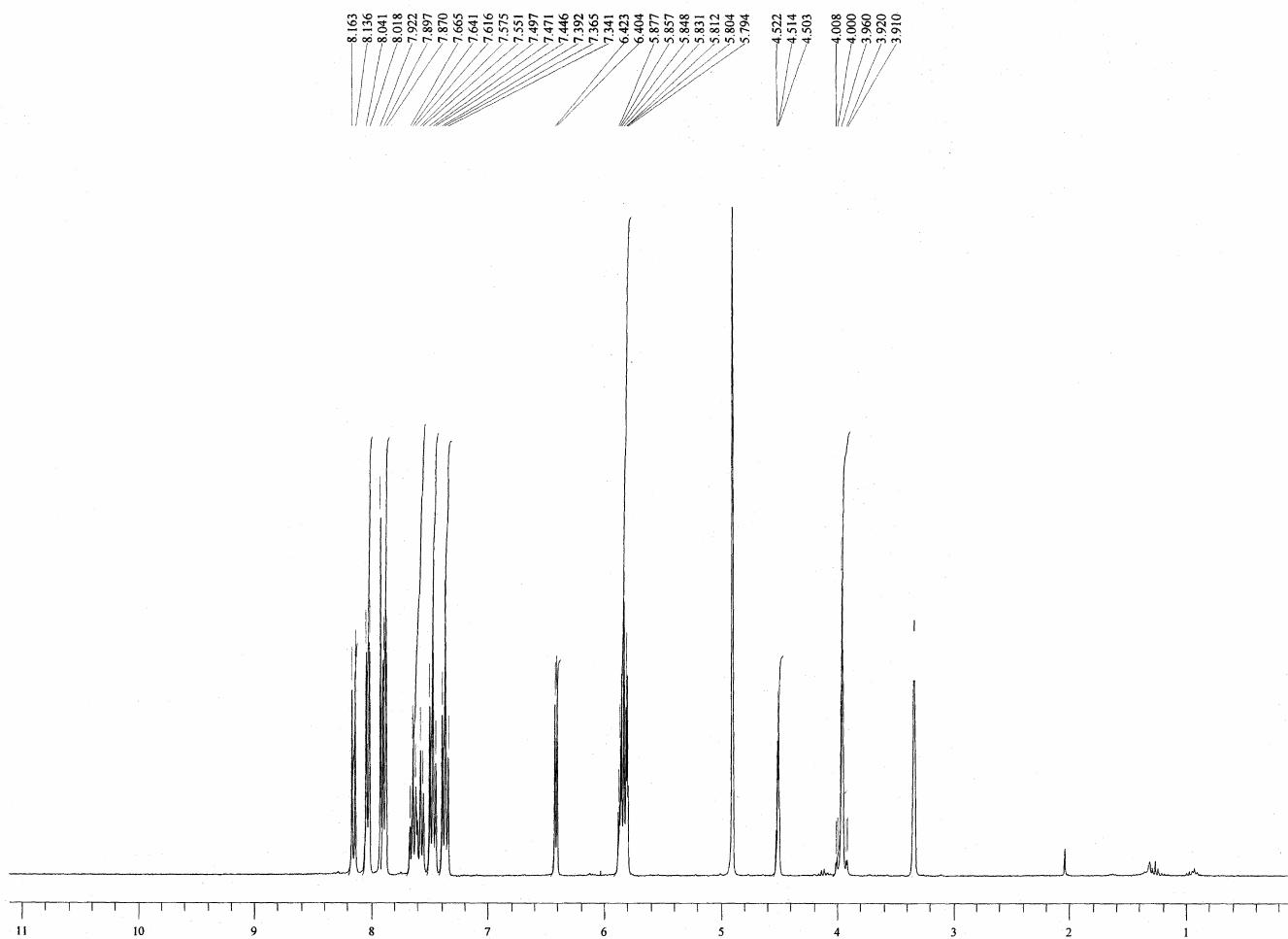


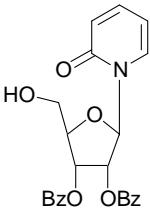
$^{31}\text{P}$





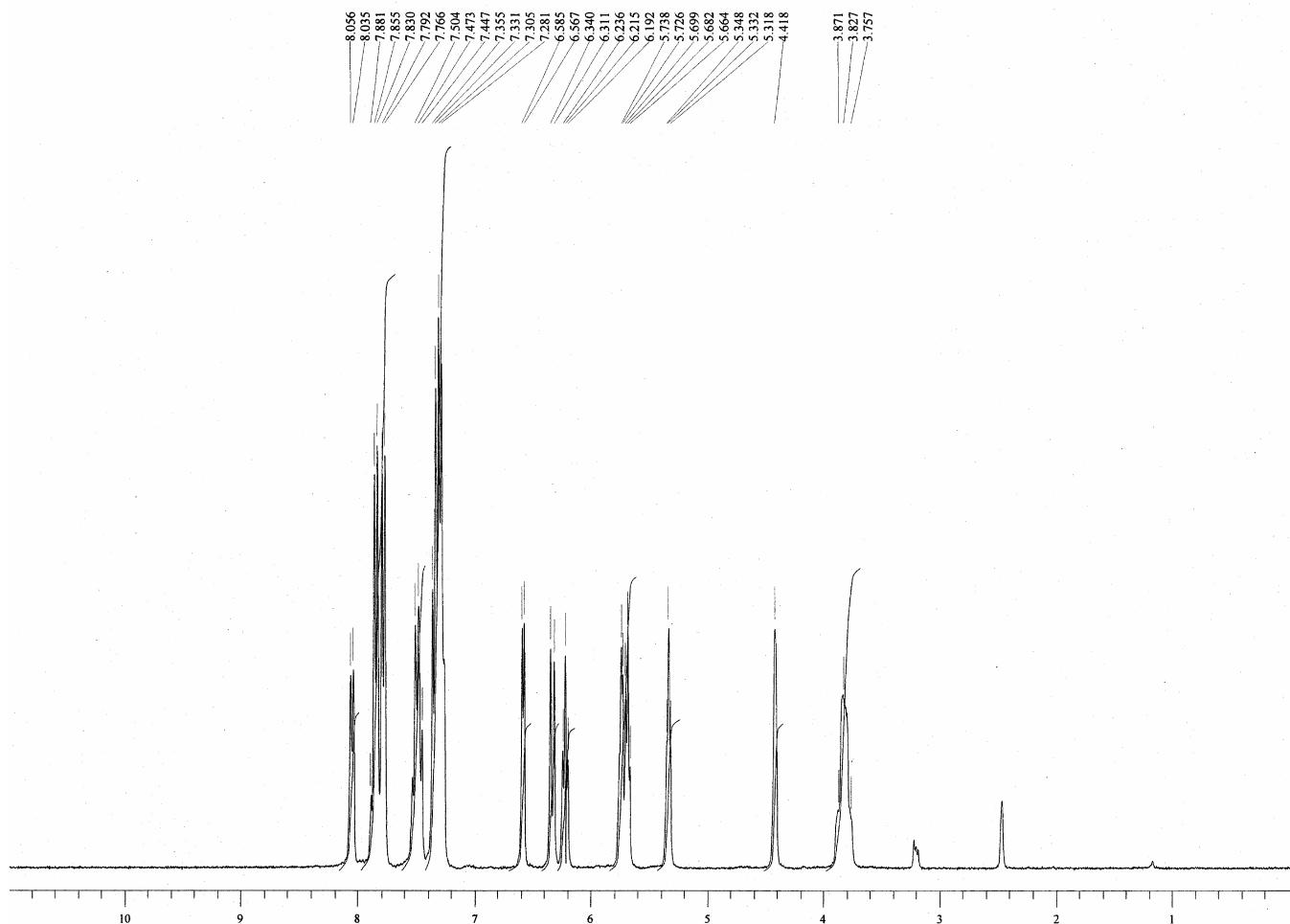
<sup>1</sup>H

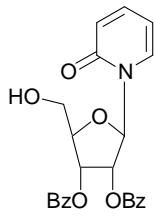




**35 (DMSO-*d*<sub>6</sub>-CDCl<sub>3</sub>)**

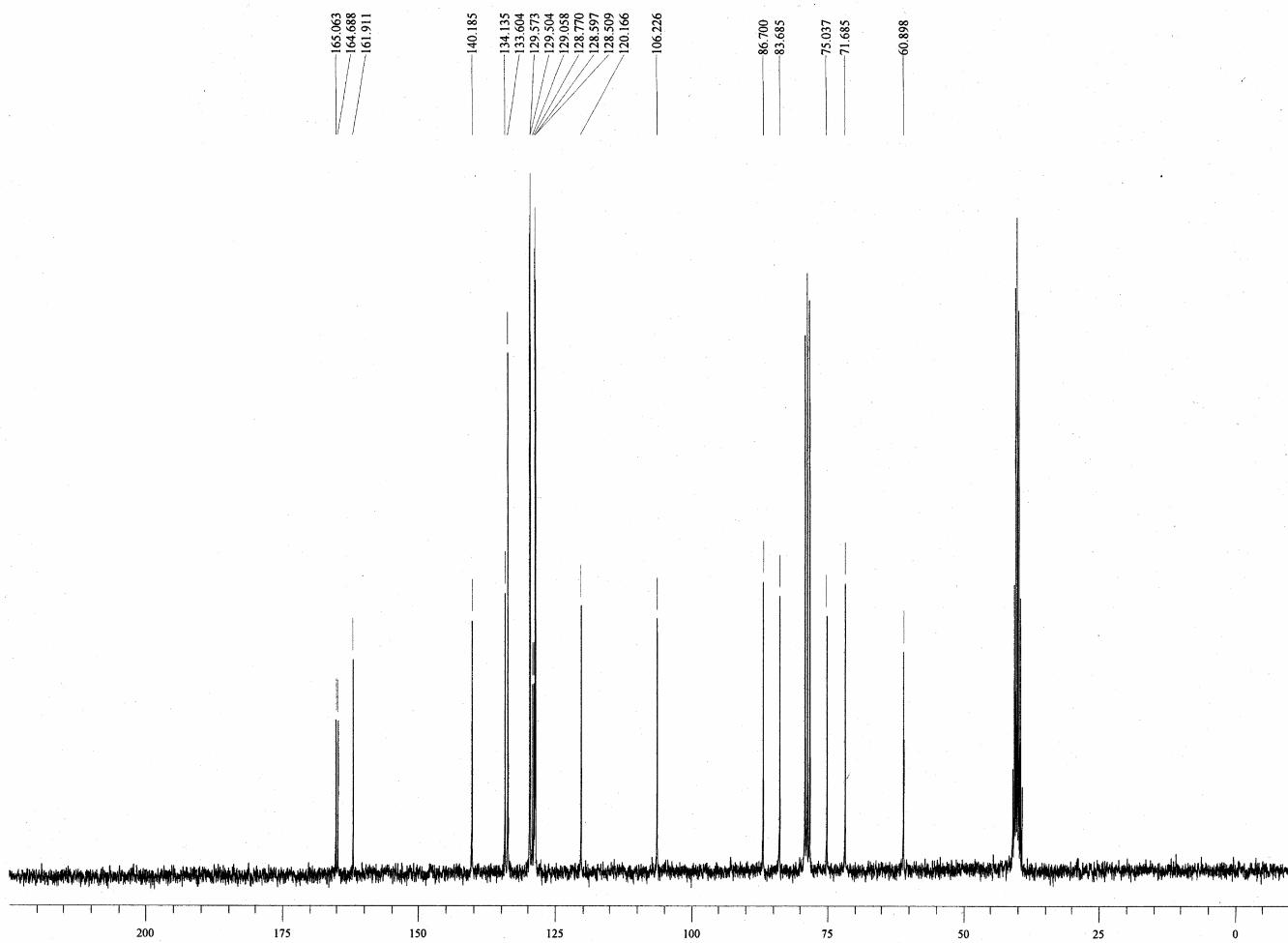
<sup>1</sup>H

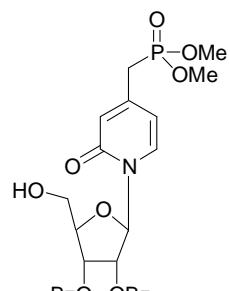




**35 (DMSO-*d*<sub>6</sub>-CDCl<sub>3</sub>)**

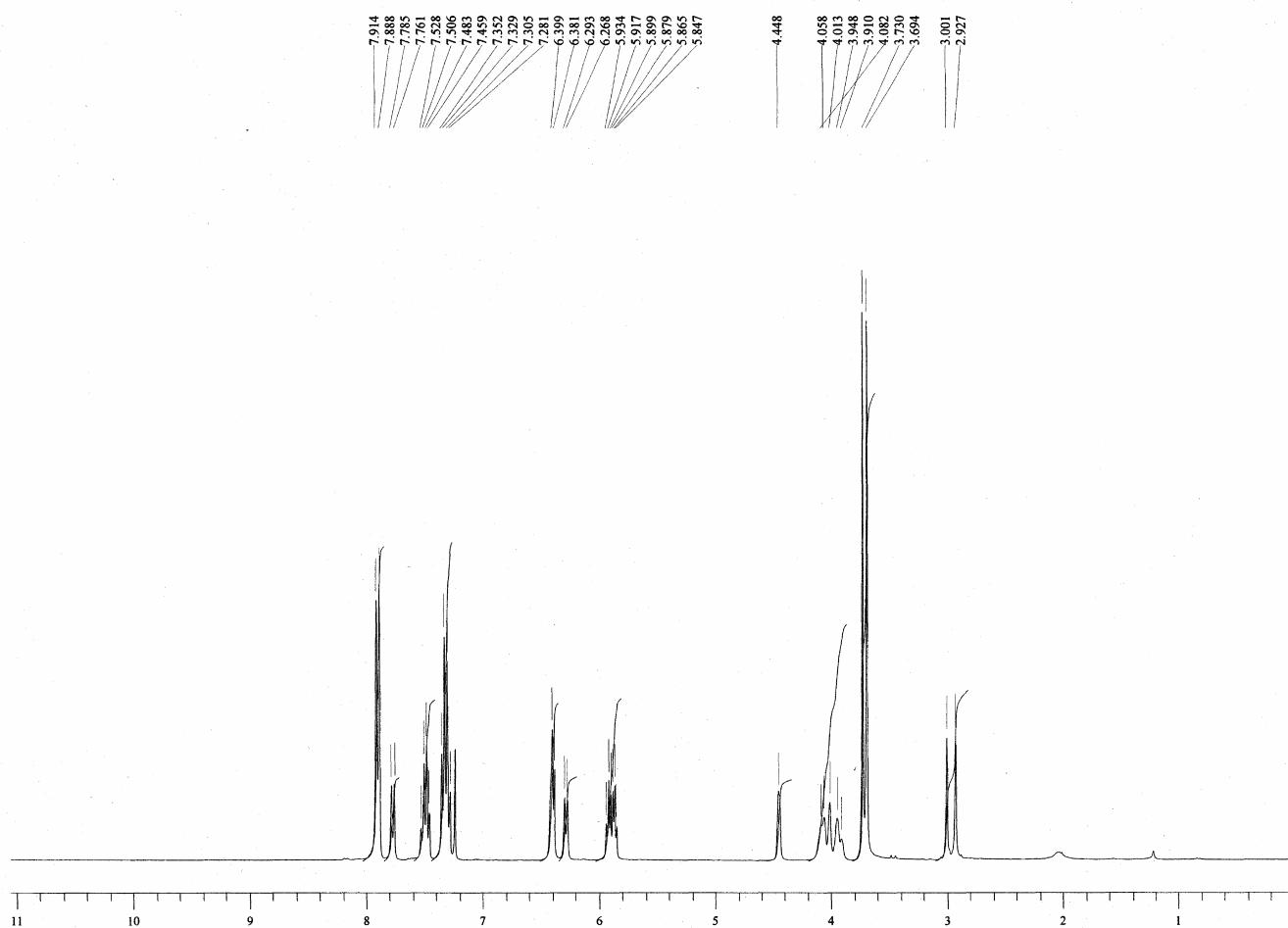
<sup>13</sup>C

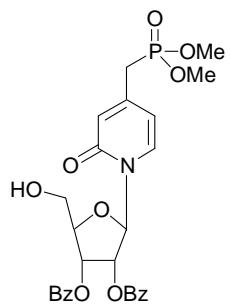




**36 (CDCl<sub>3</sub>)**

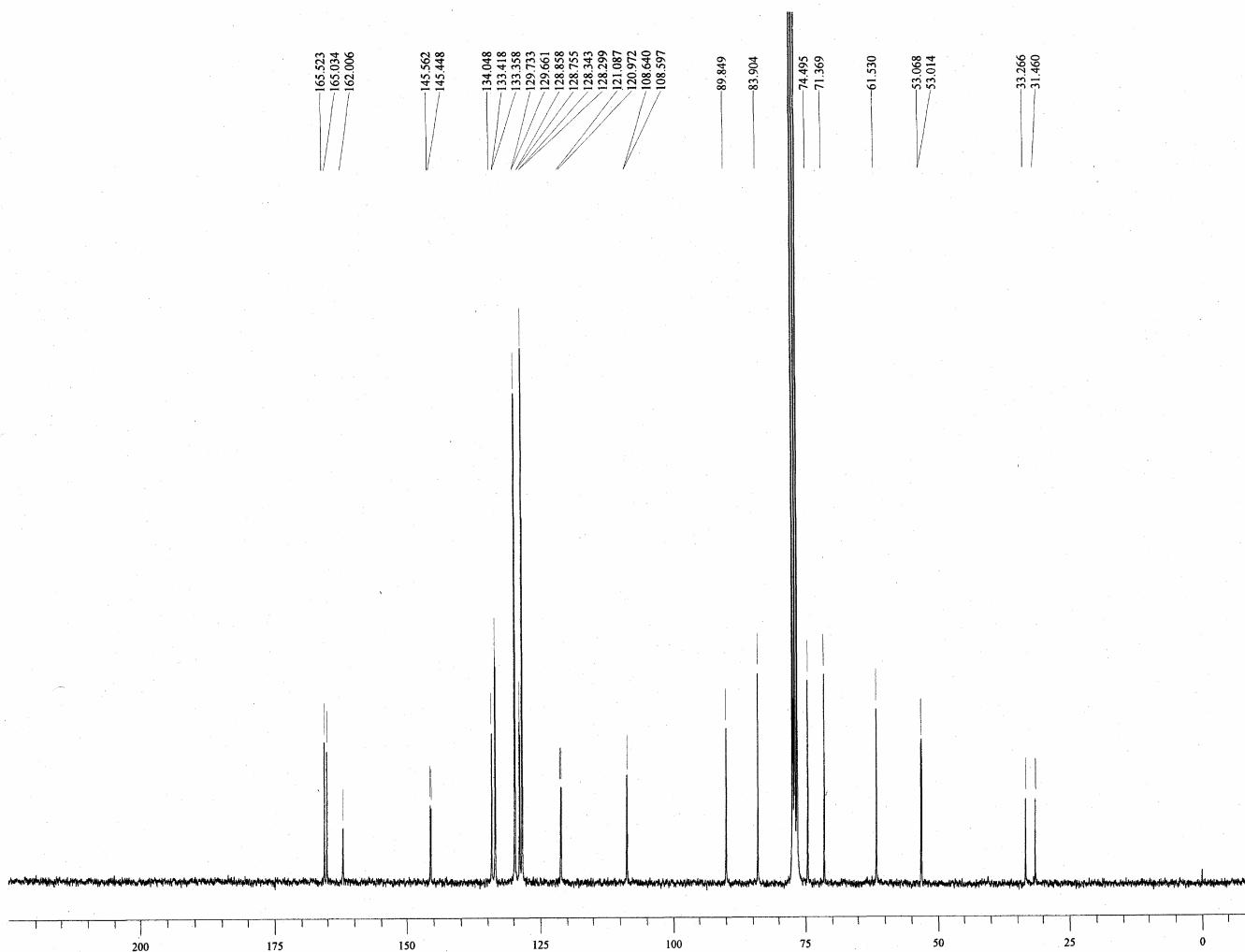
<sup>1</sup>H

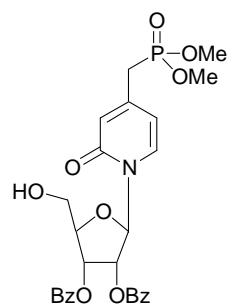




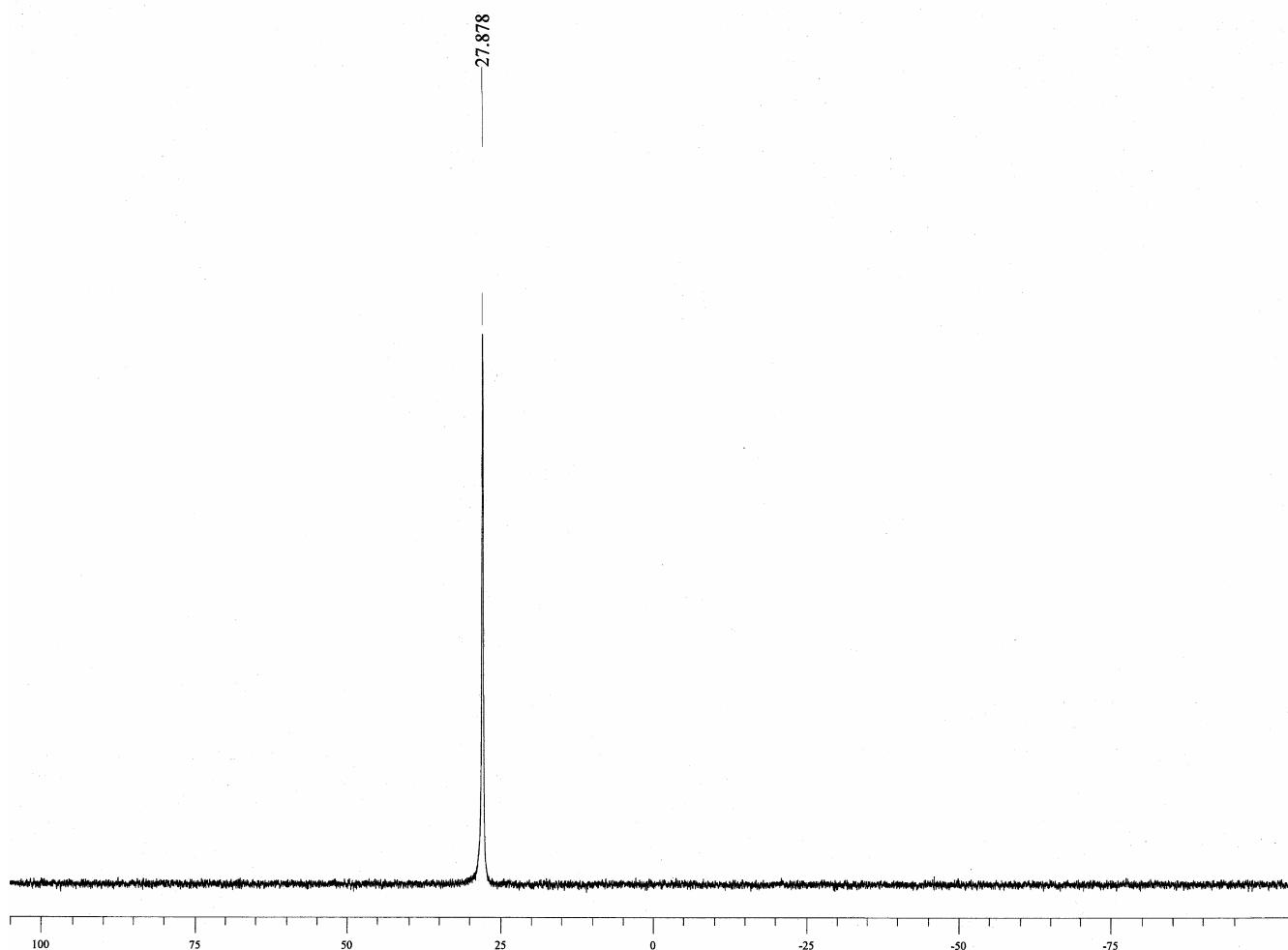
**36 (CDCl<sub>3</sub>)**

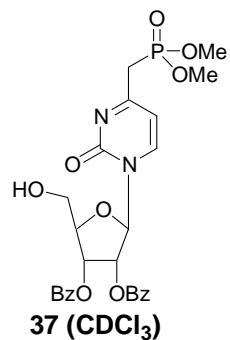
<sup>13</sup>C



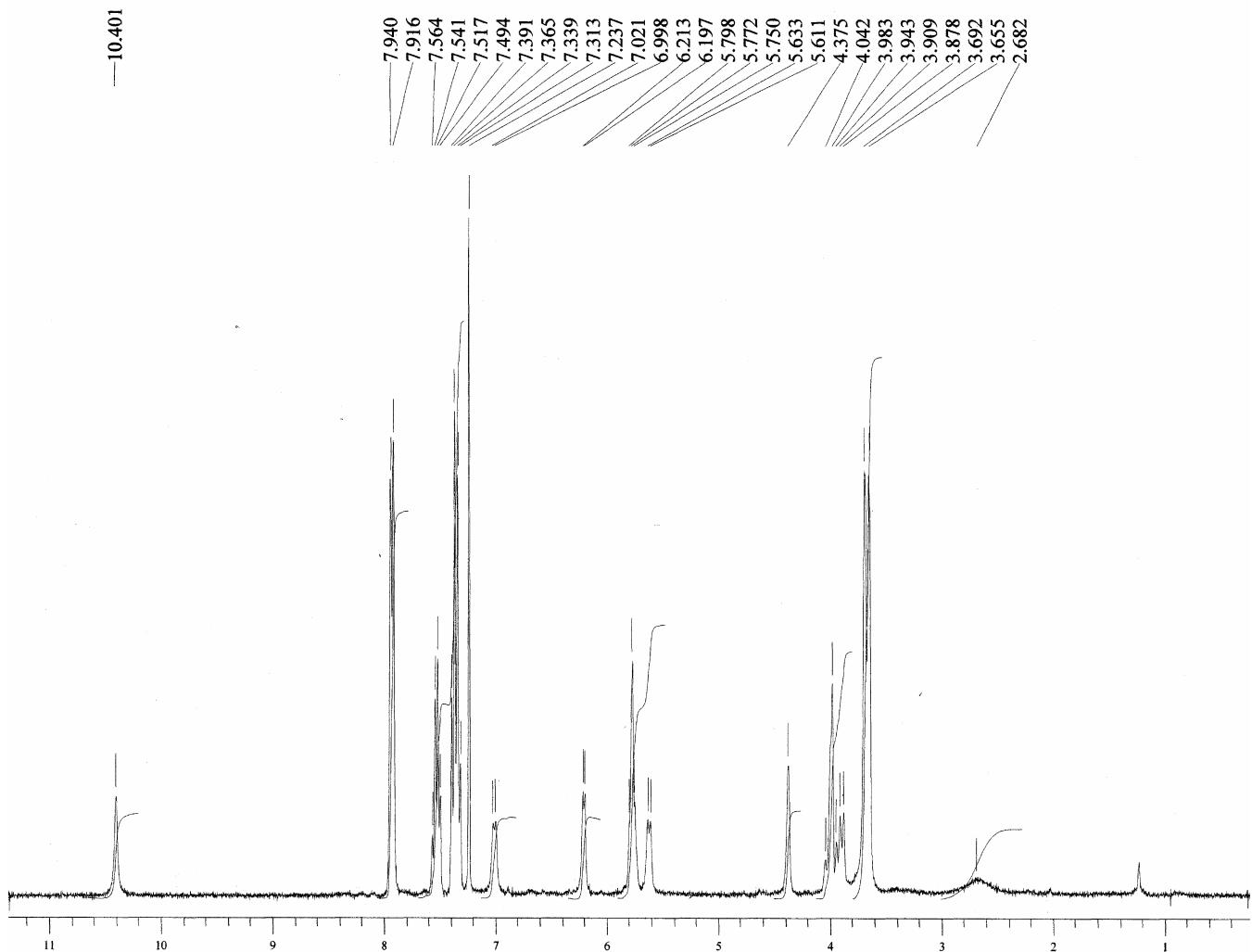


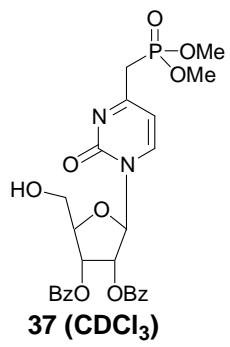
<sup>31</sup>P



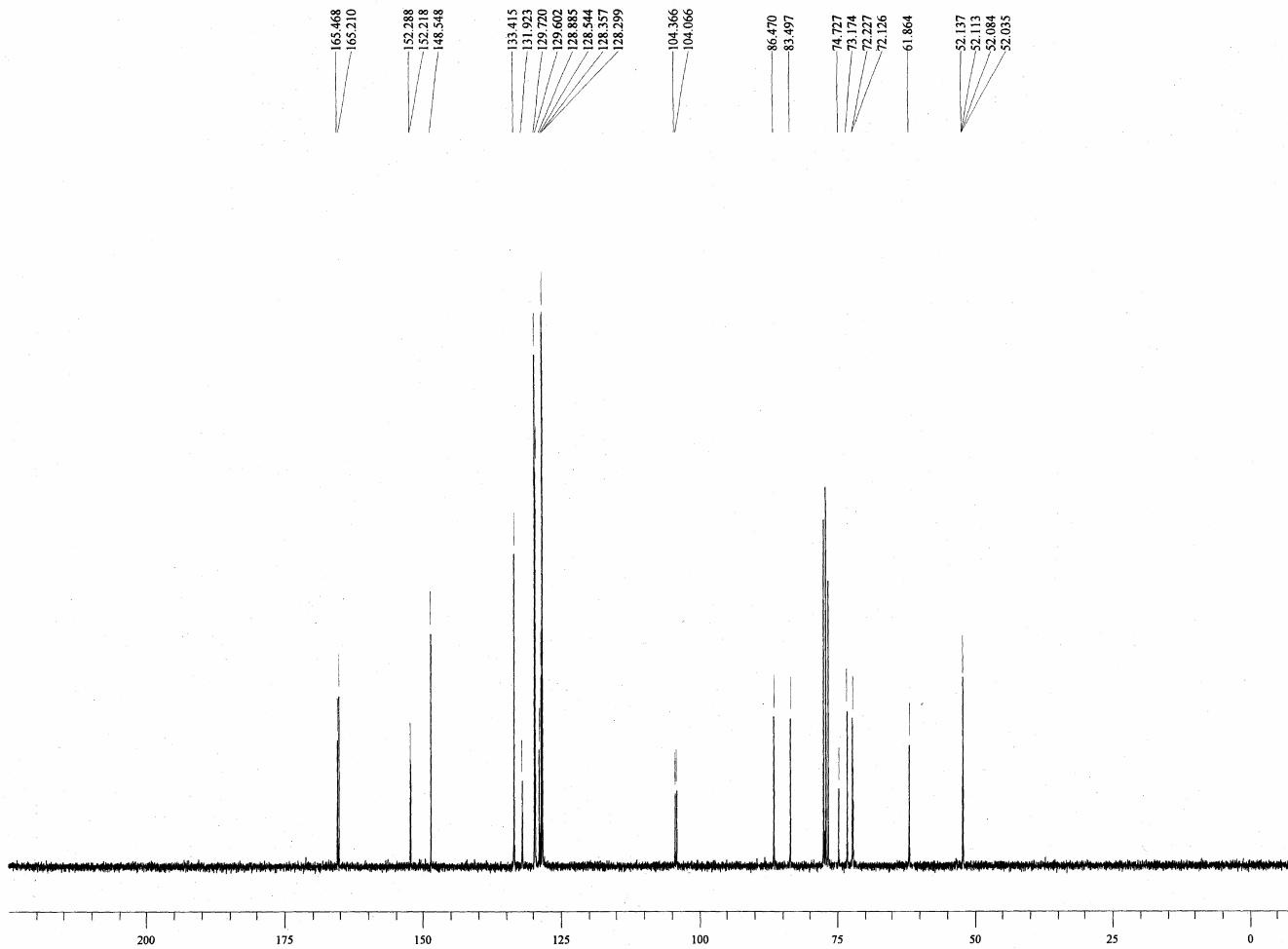


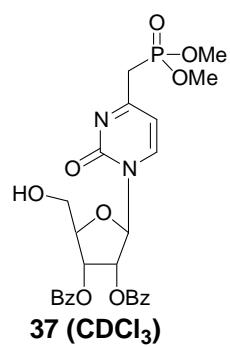
$^1\text{H}$



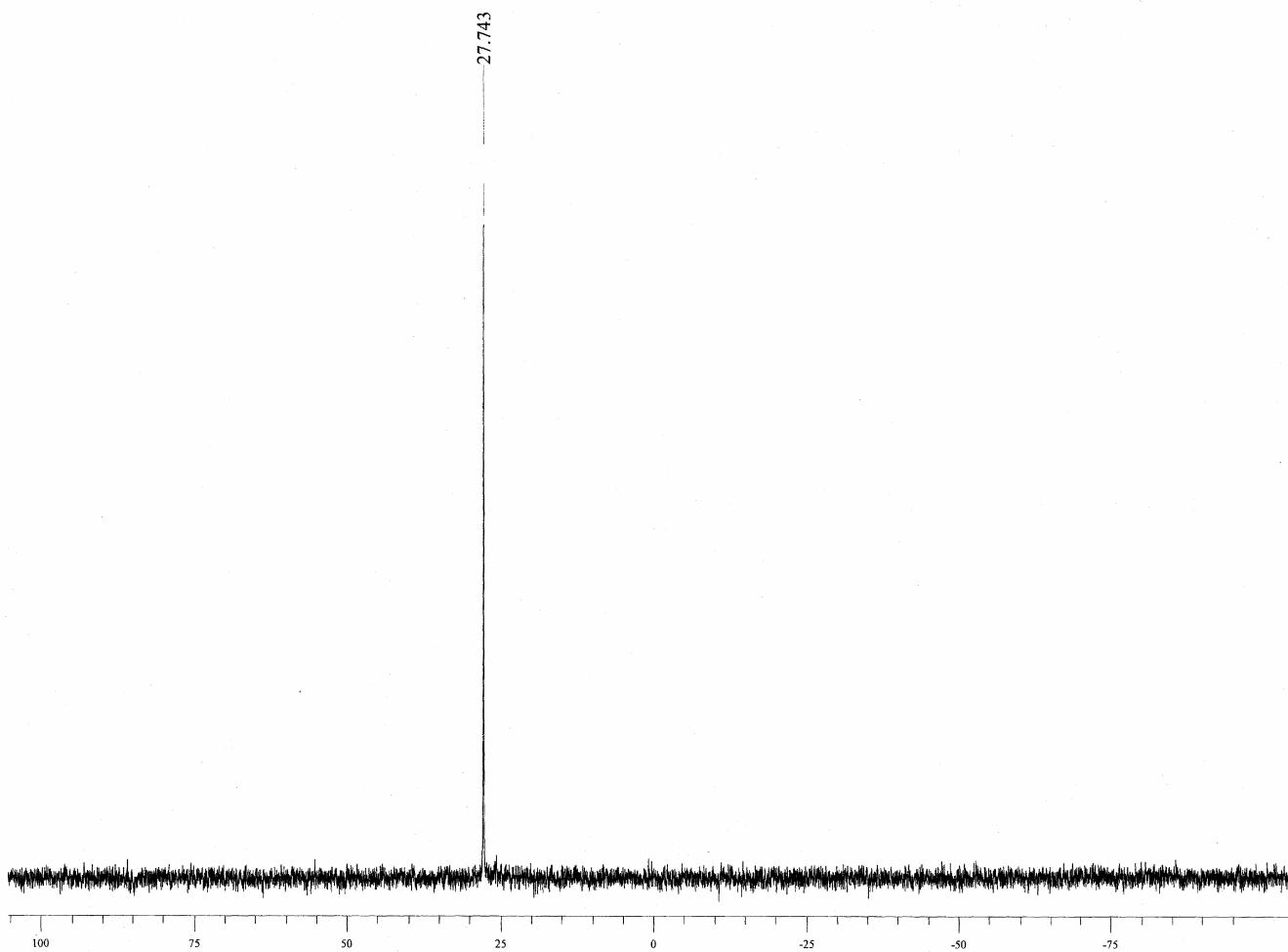


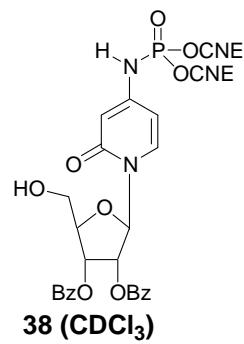
<sup>13</sup>C



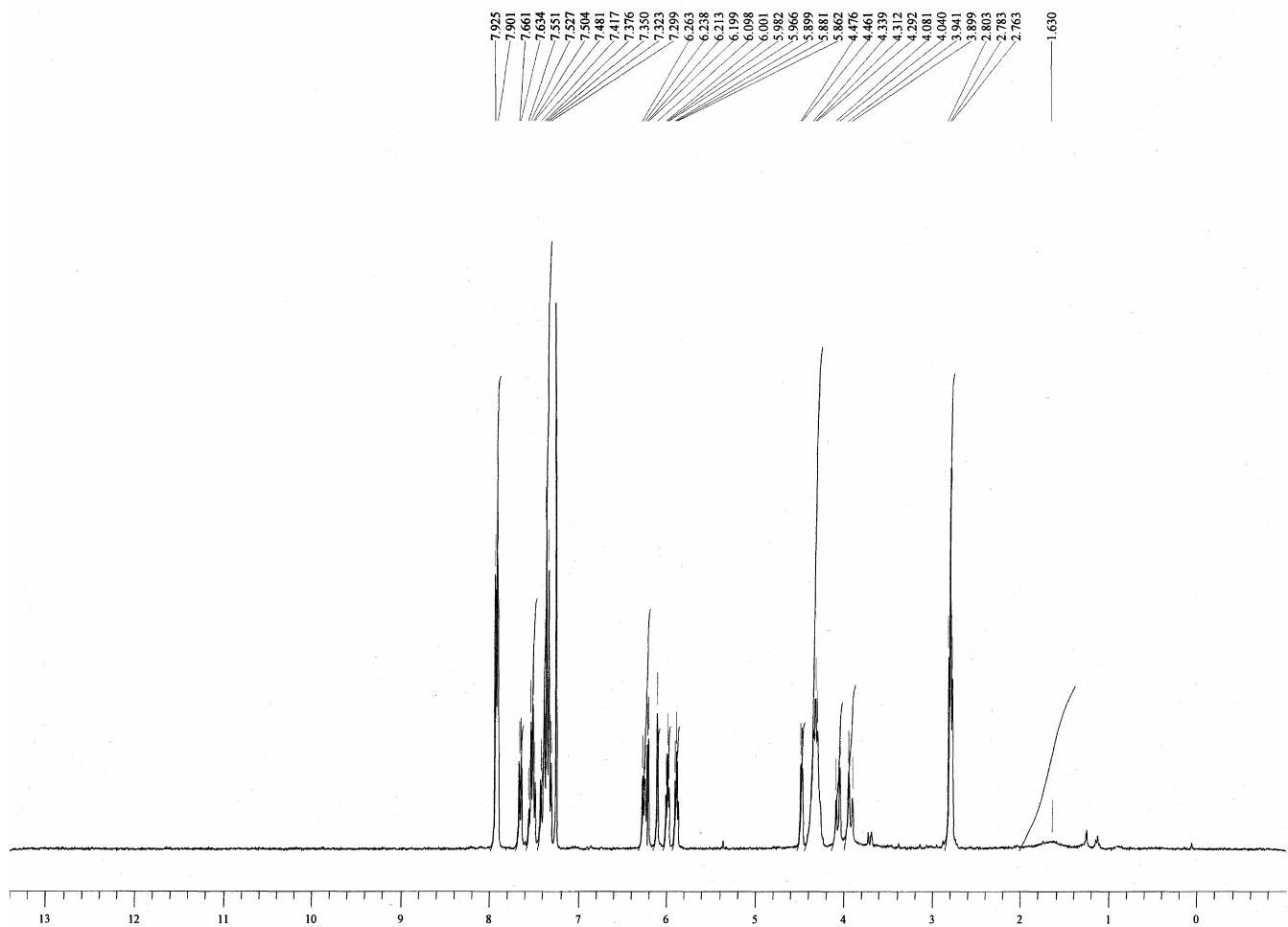


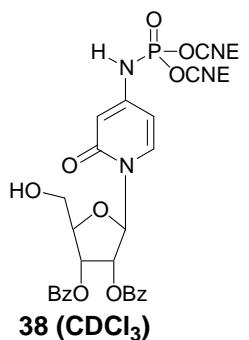
<sup>31</sup>P



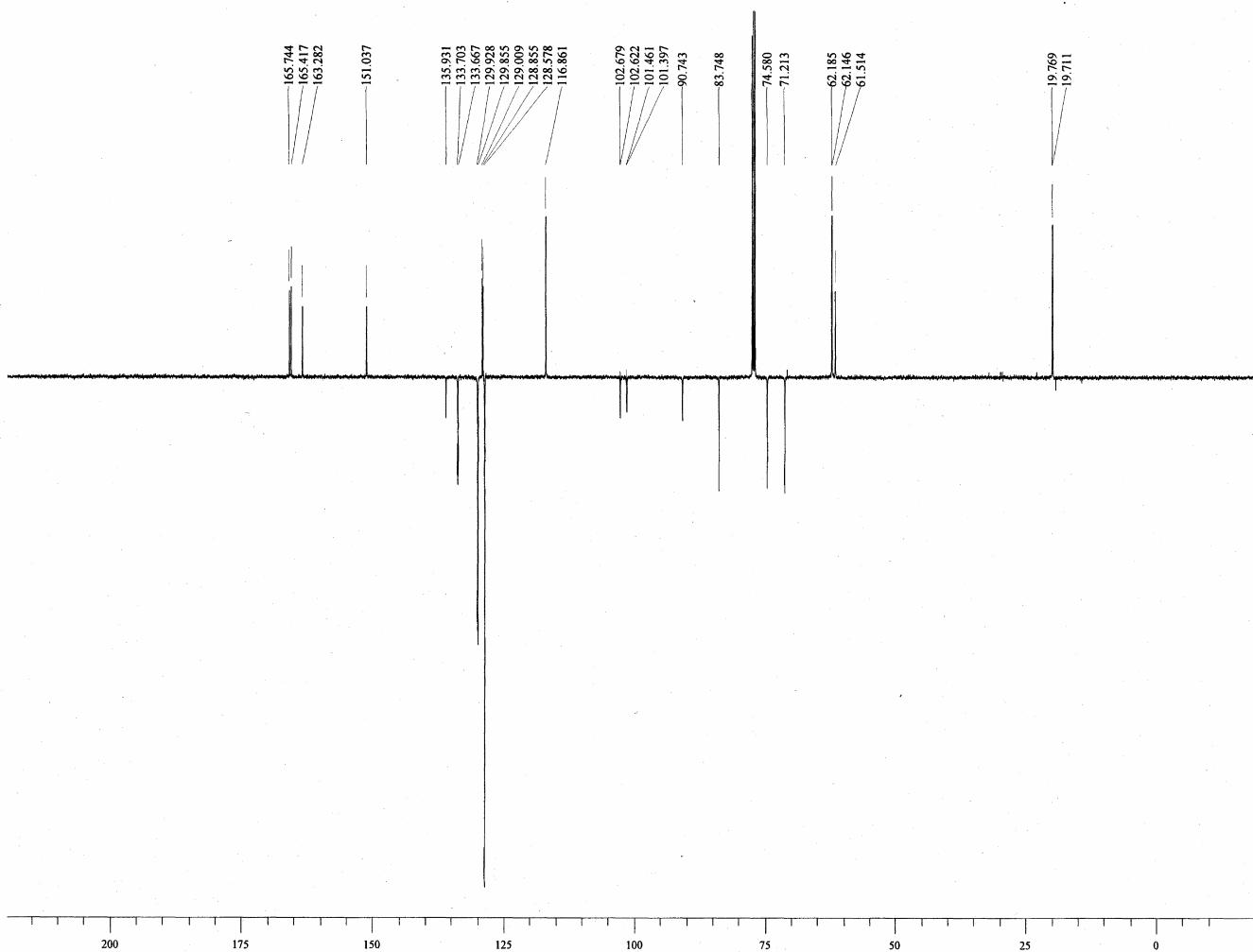


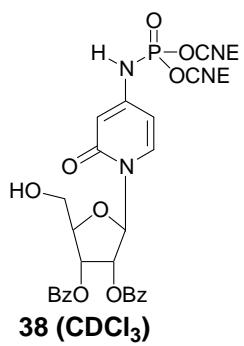
<sup>1</sup>H



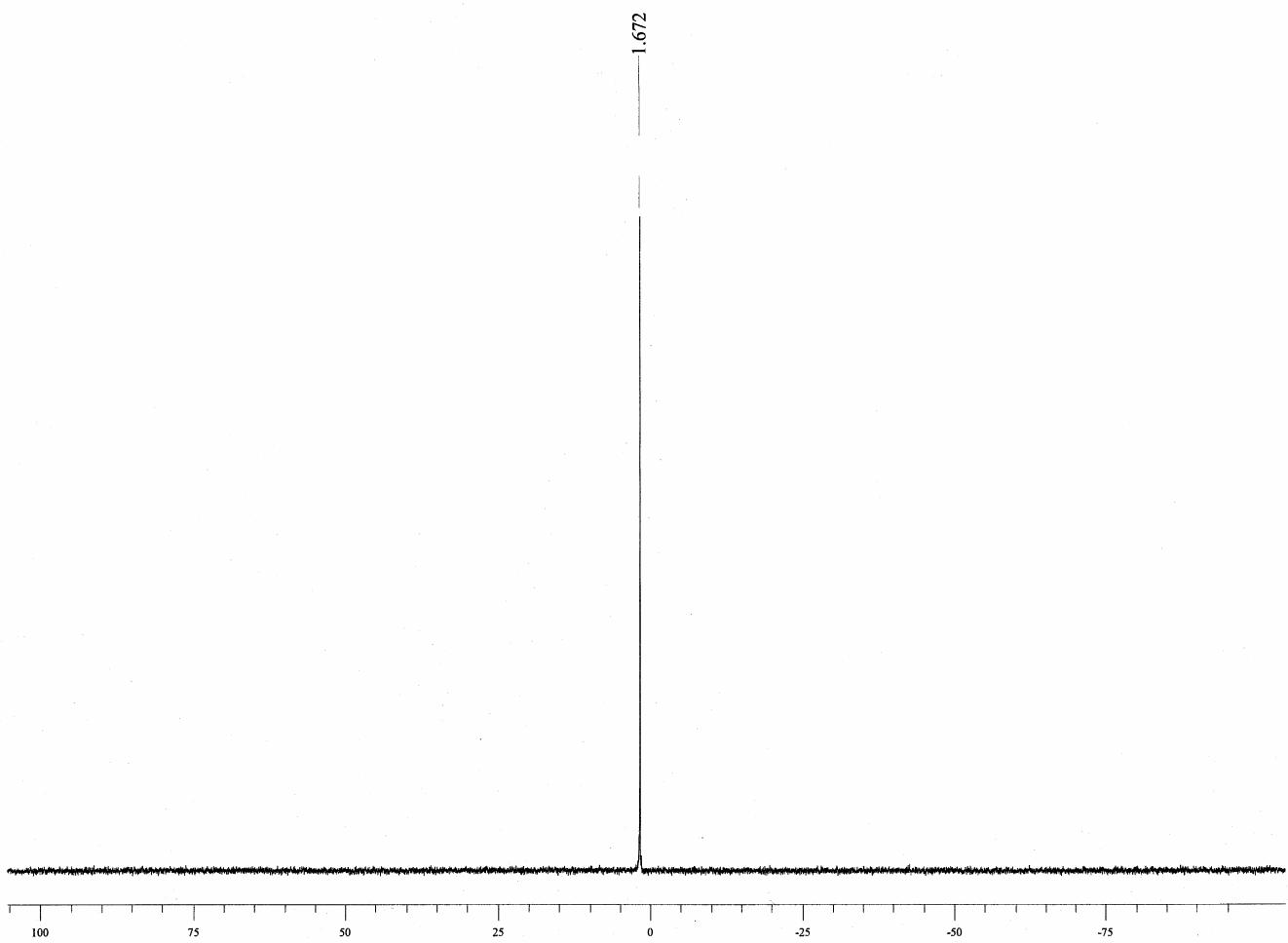


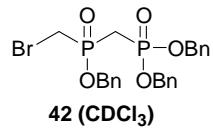
<sup>13</sup>C



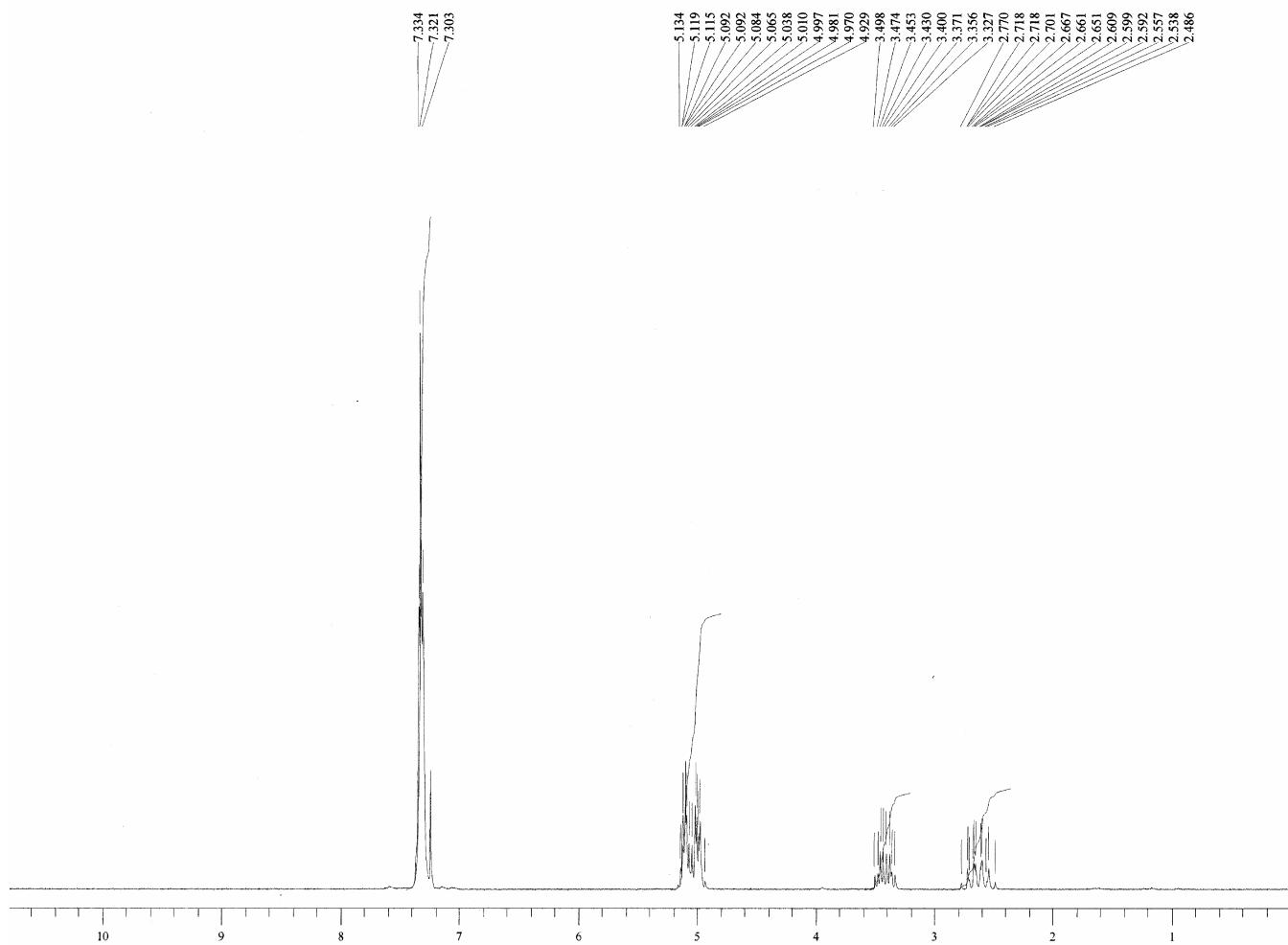


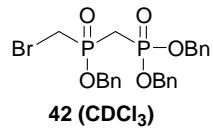
<sup>31</sup>P



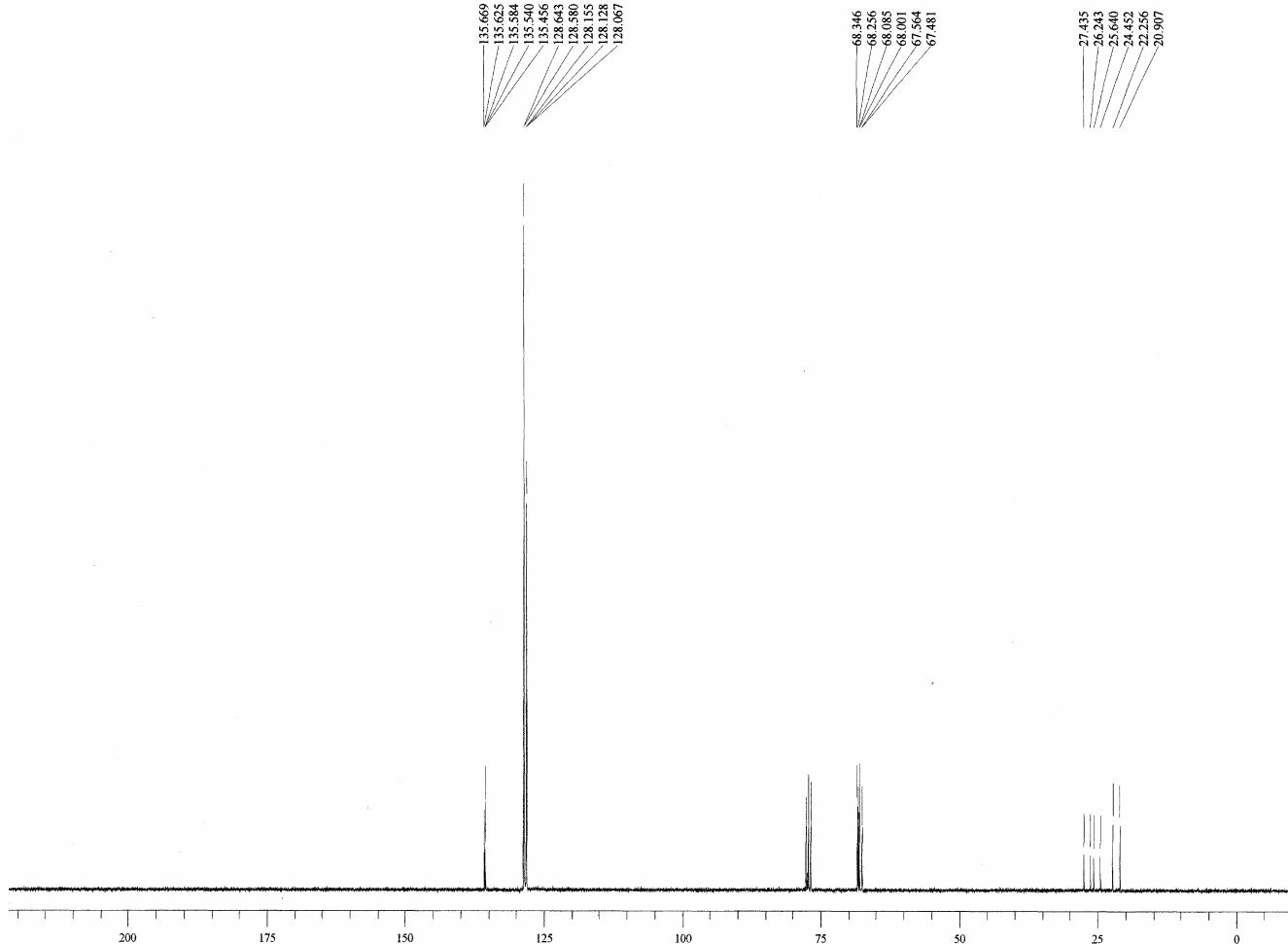


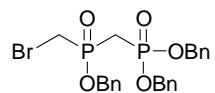
<sup>1</sup>H





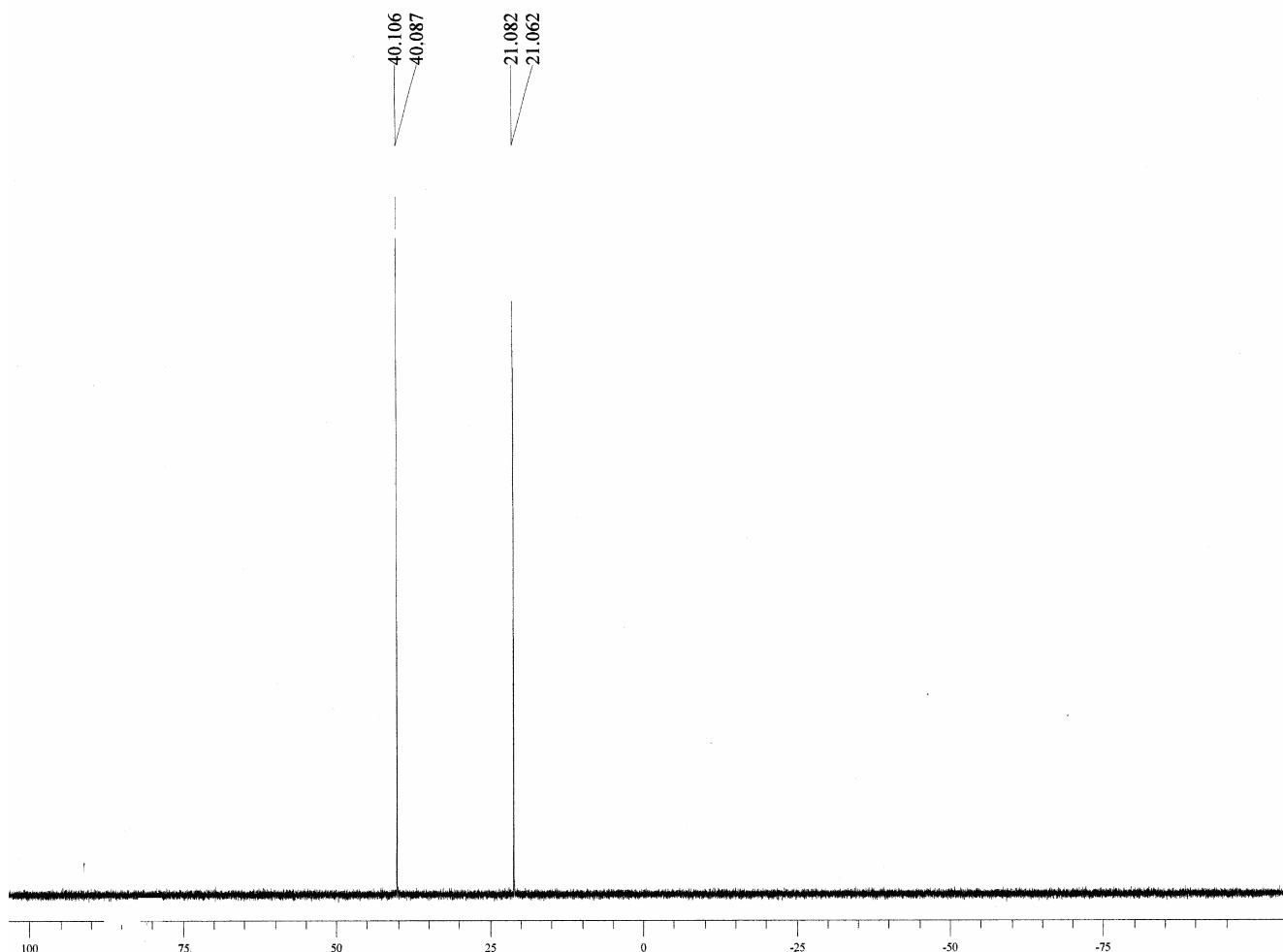
<sup>13</sup>C

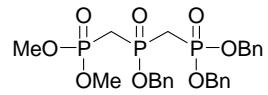




**42 (CDCl<sub>3</sub>)**

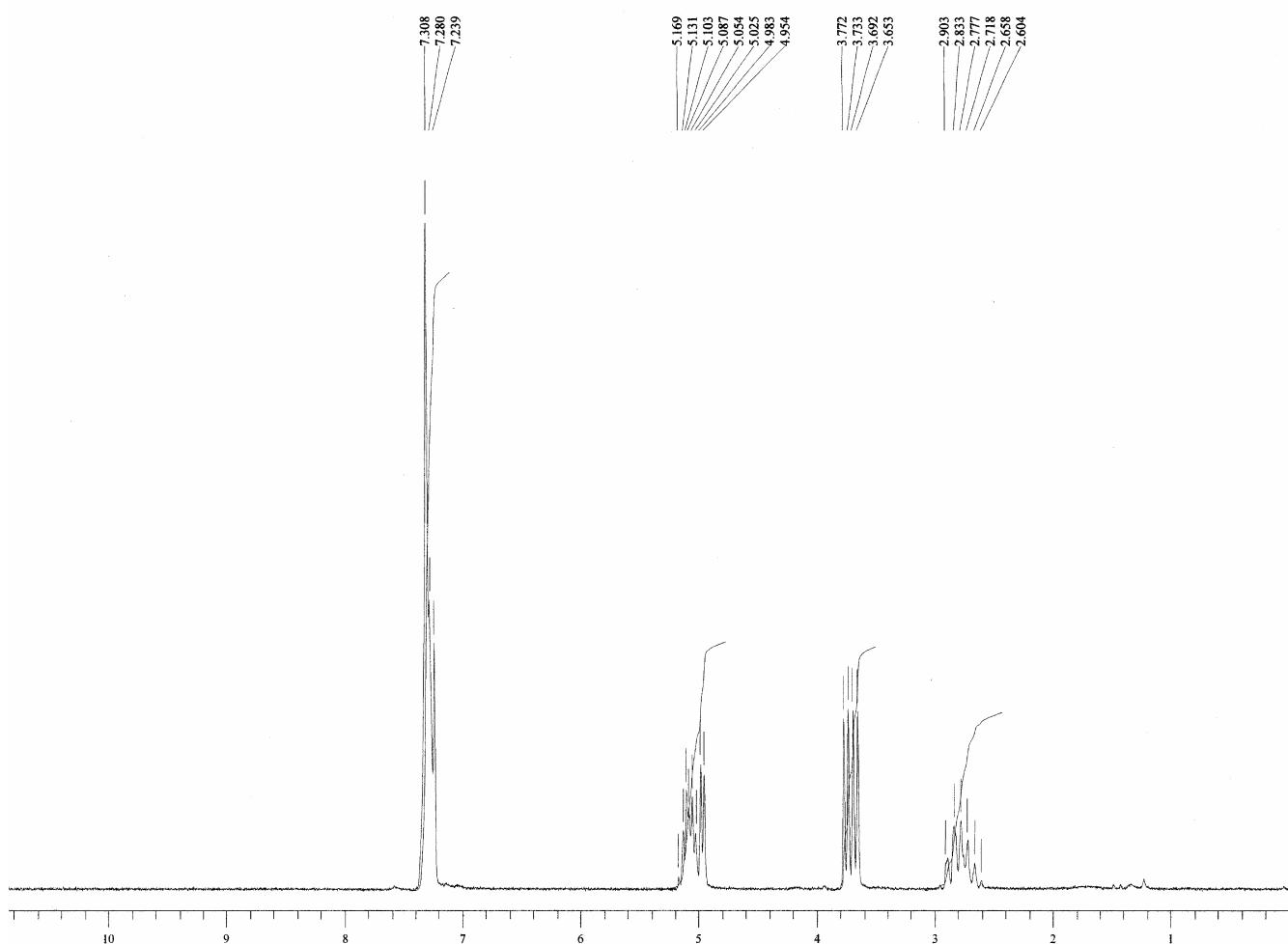
<sup>31</sup>P

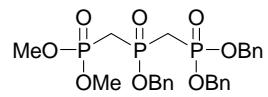




**43 (CDCl<sub>3</sub>)**

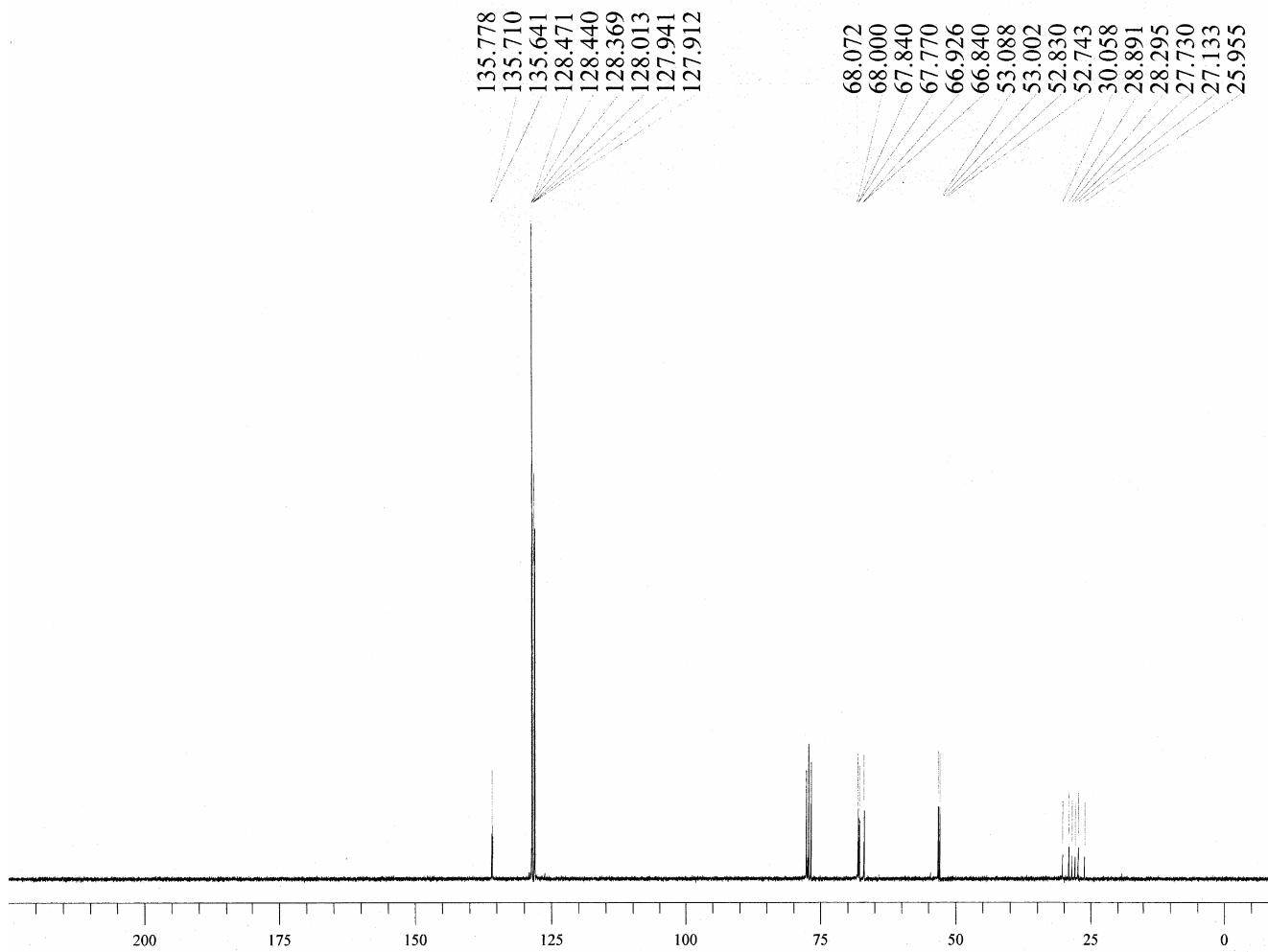
<sup>1</sup>H

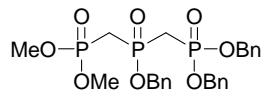




**43 ( $\text{CDCl}_3$ )**

$^{13}\text{C}$



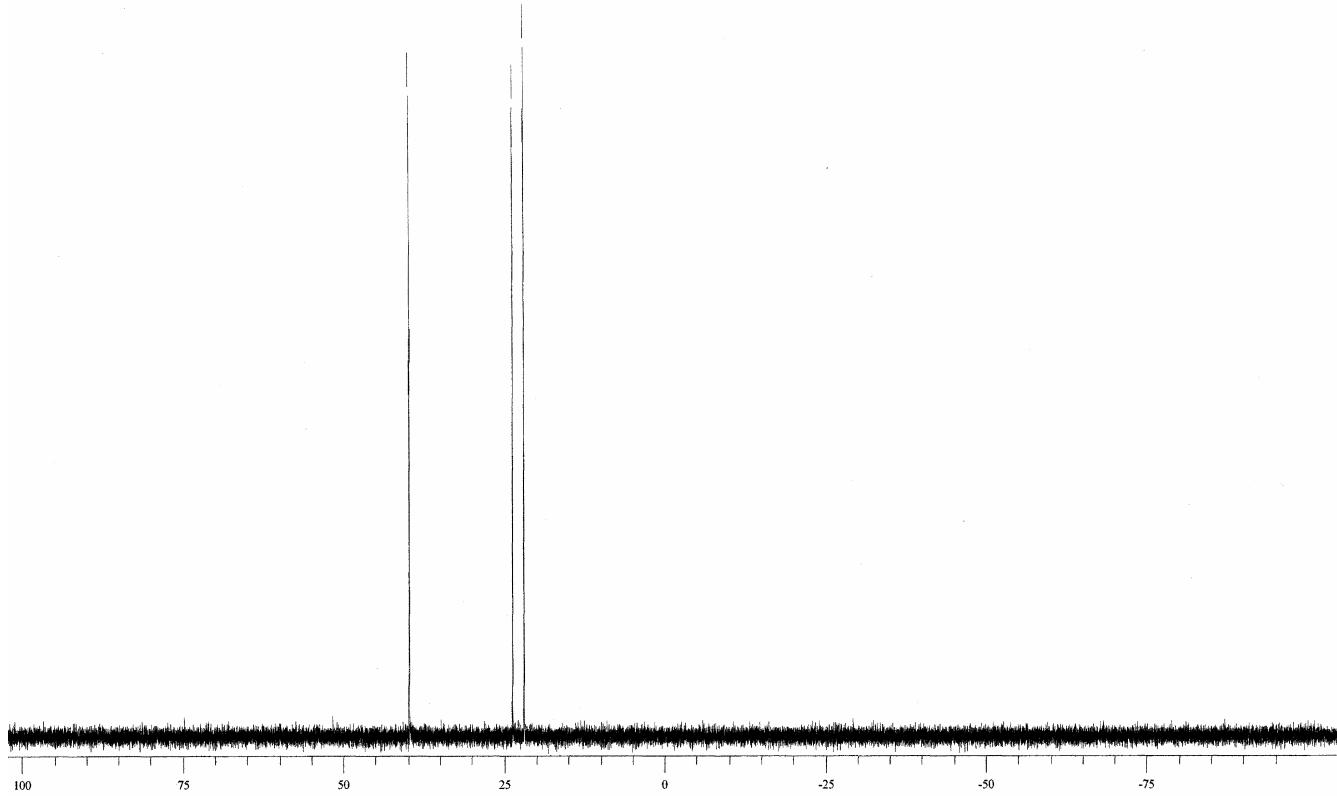


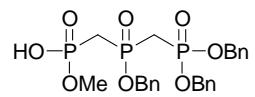
43 ( $\text{CDCl}_3$ )

$^{31}\text{P}$

39.809  
39.770  
39.745

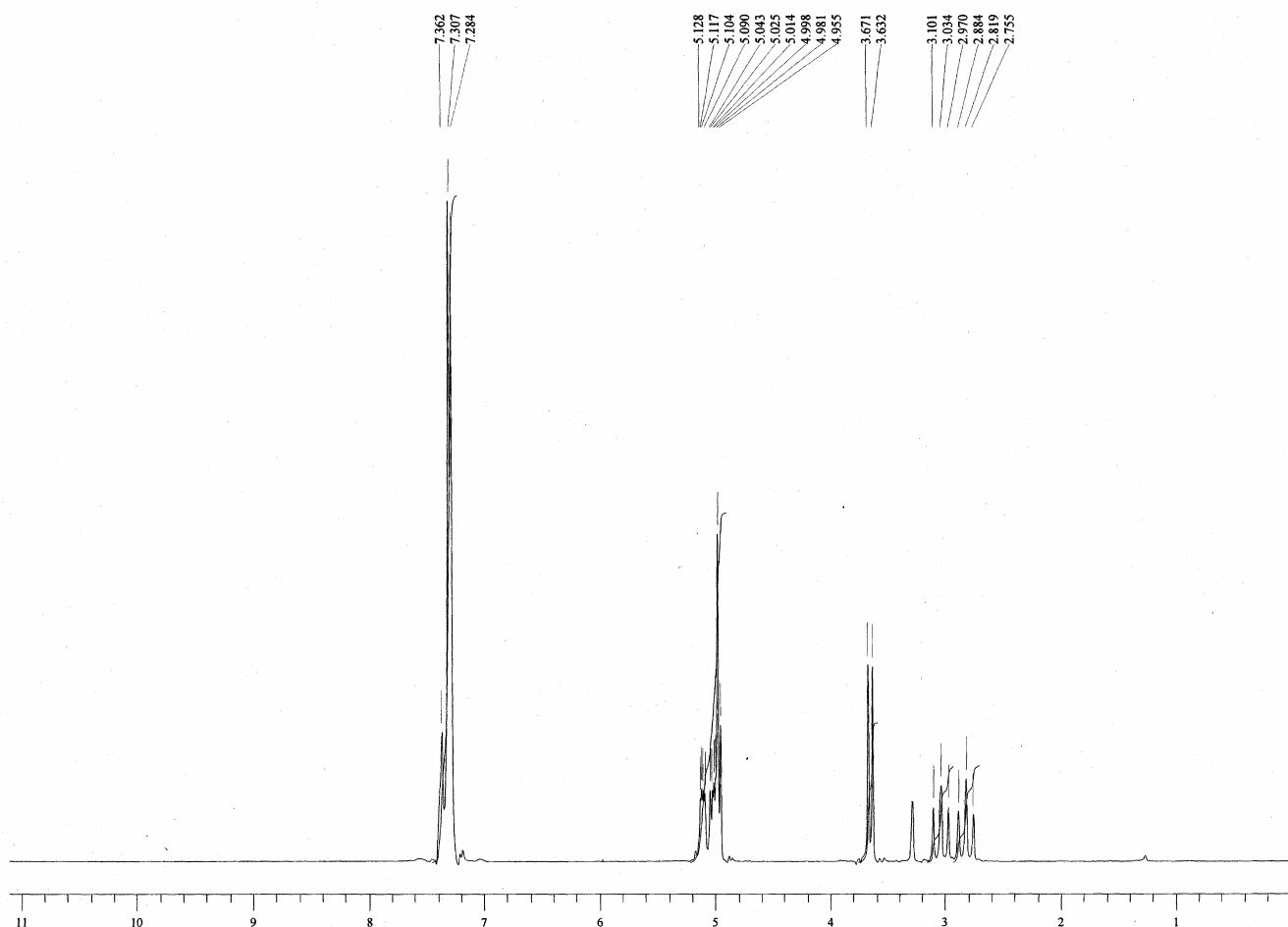
23.657  
23.624  
21.899  
21.870

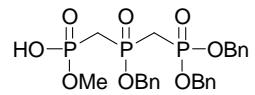




**44 (CD<sub>3</sub>OD)**

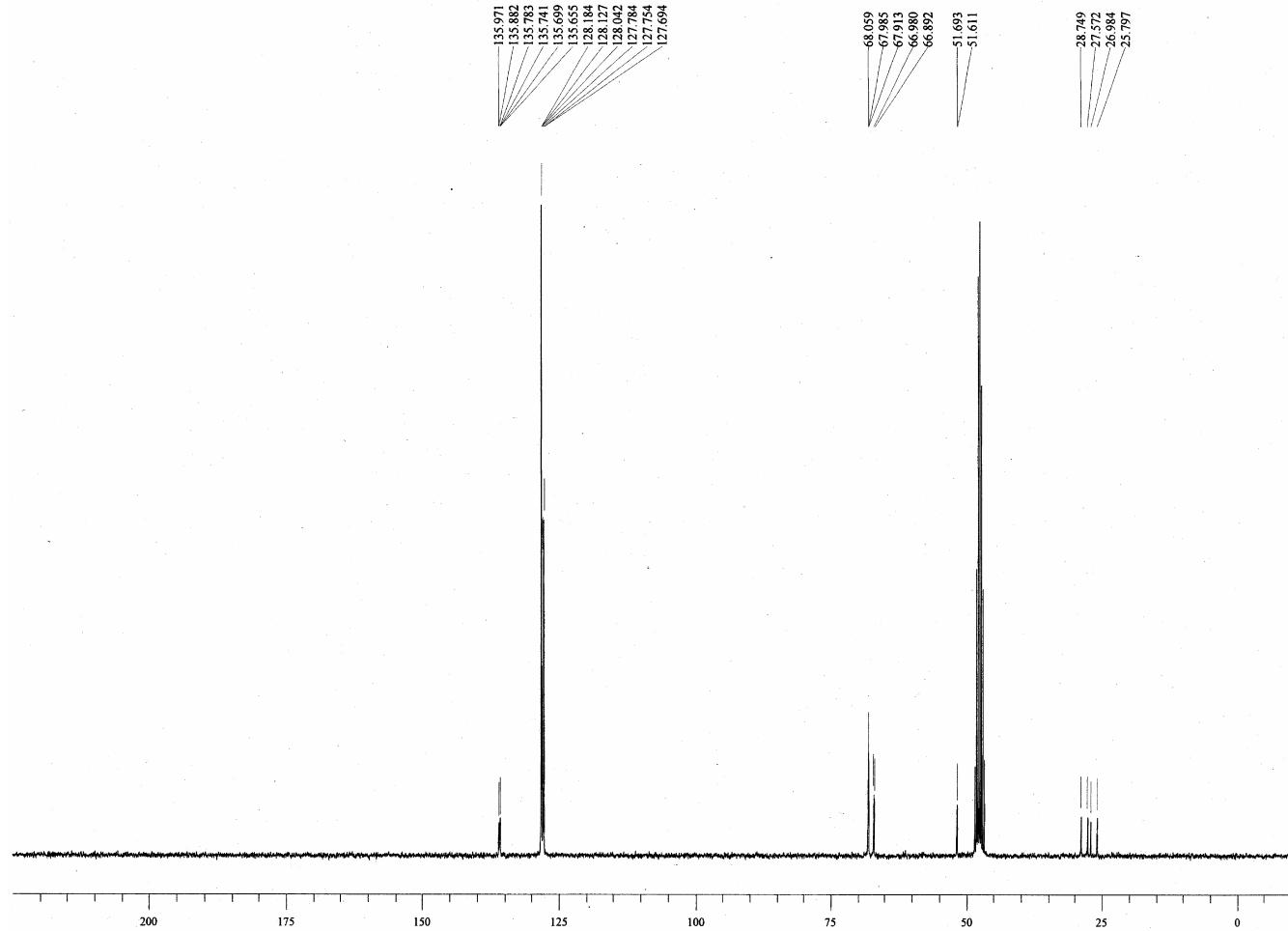
<sup>1</sup>H

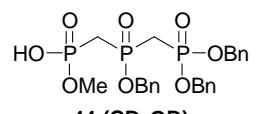




**44 (CD<sub>3</sub>OD)**

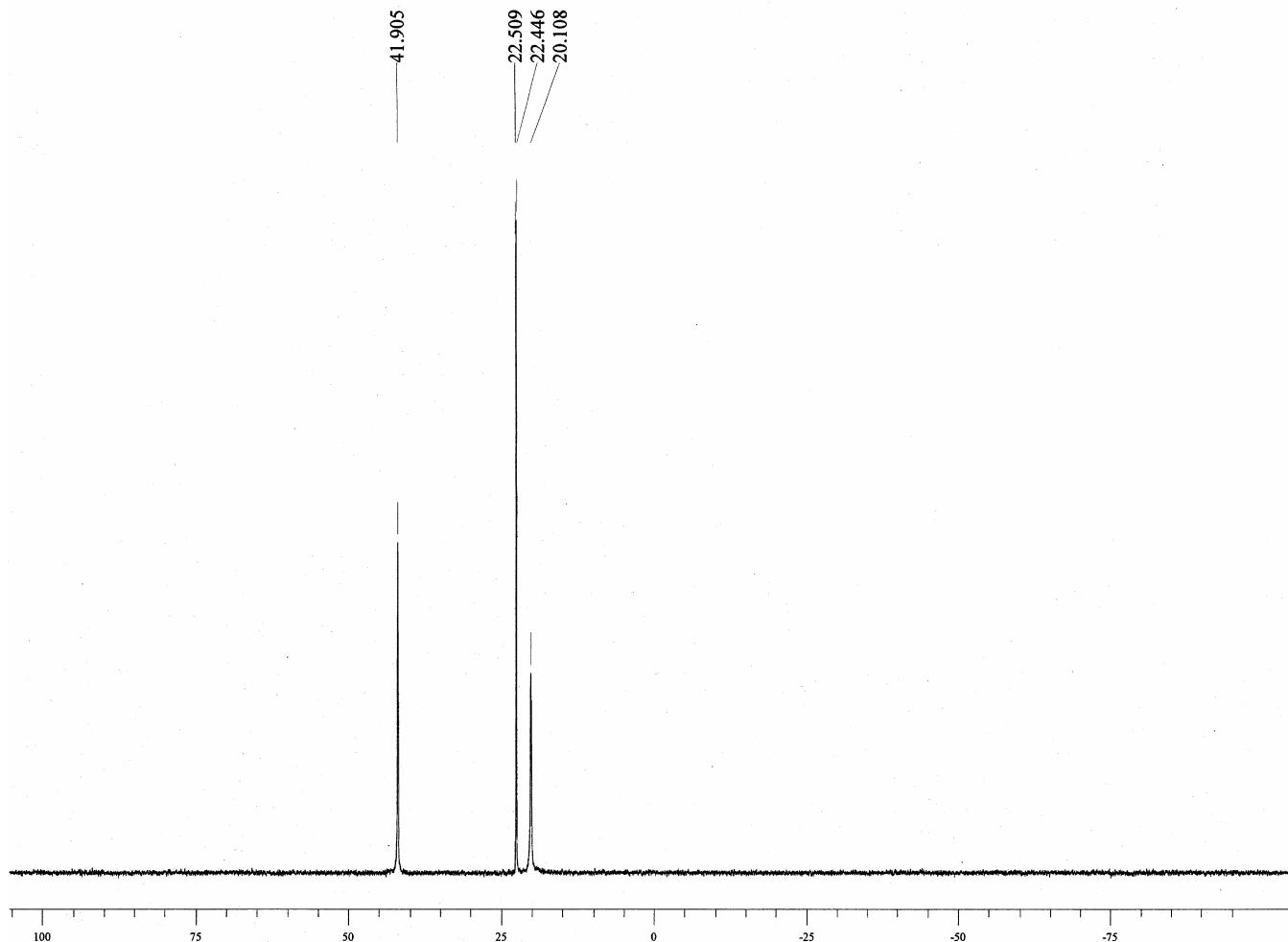
<sup>13</sup>C

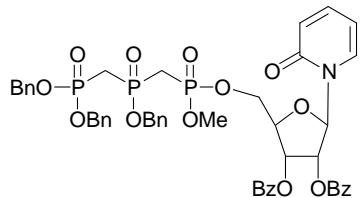




44 ( $\text{CD}_3\text{OD}$ )

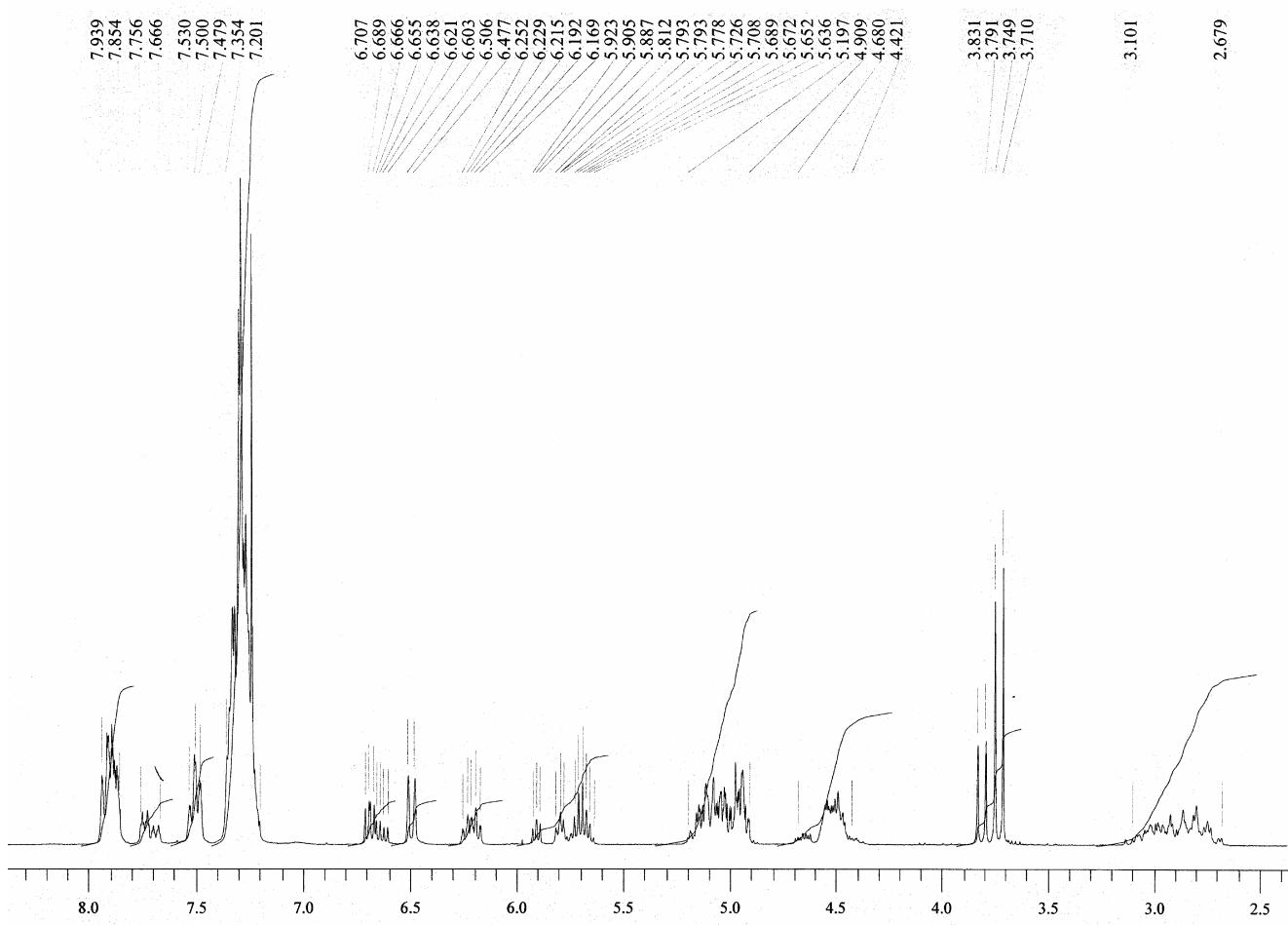
$^{31}\text{P}$

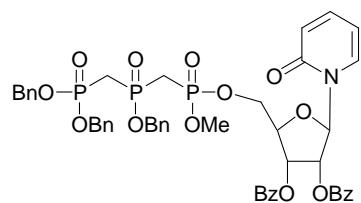




**45 ( $\text{CDCl}_3$ )**

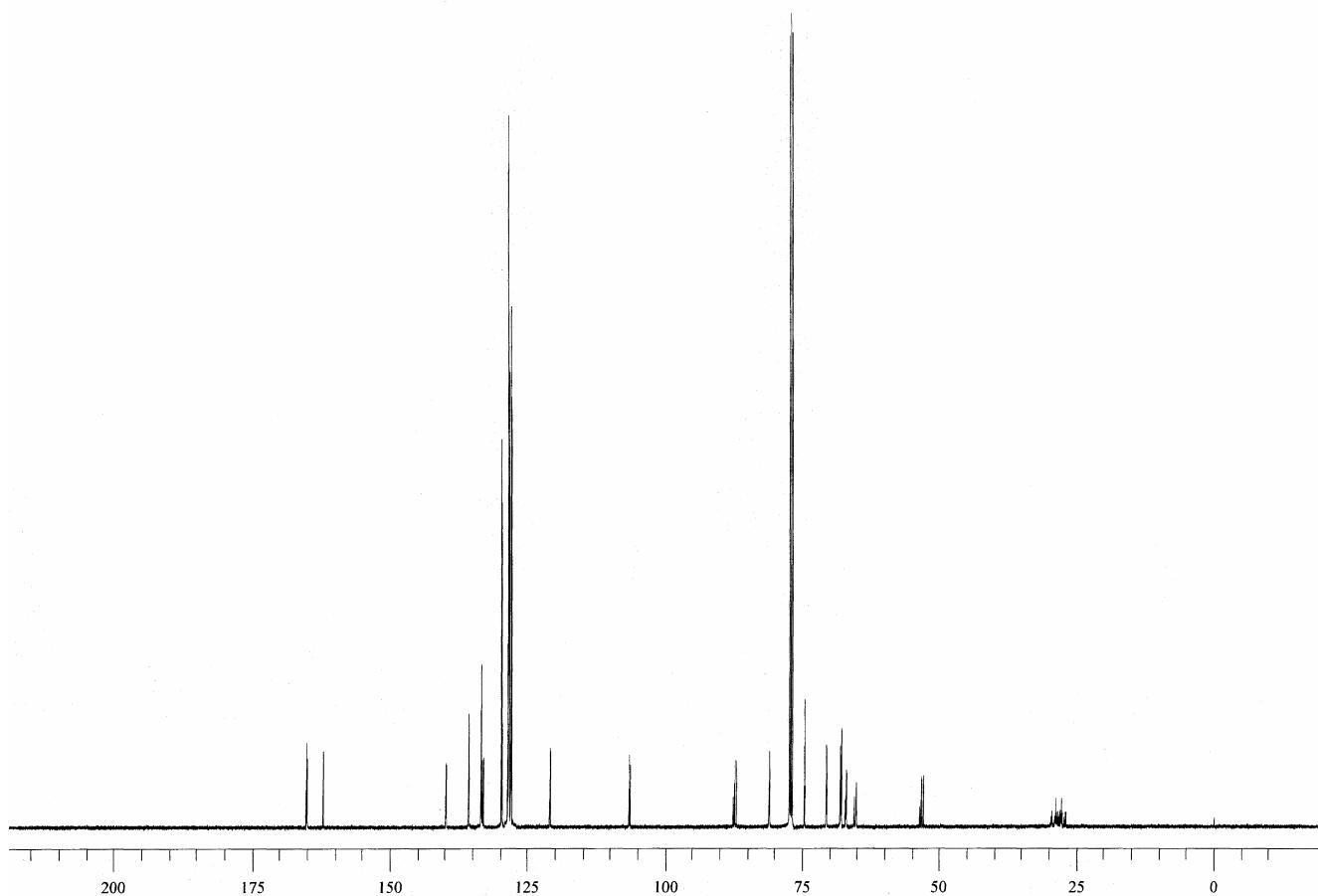
$^1\text{H}$

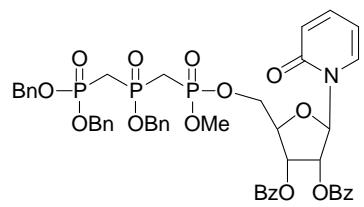




**45 (CDCl<sub>3</sub>)**

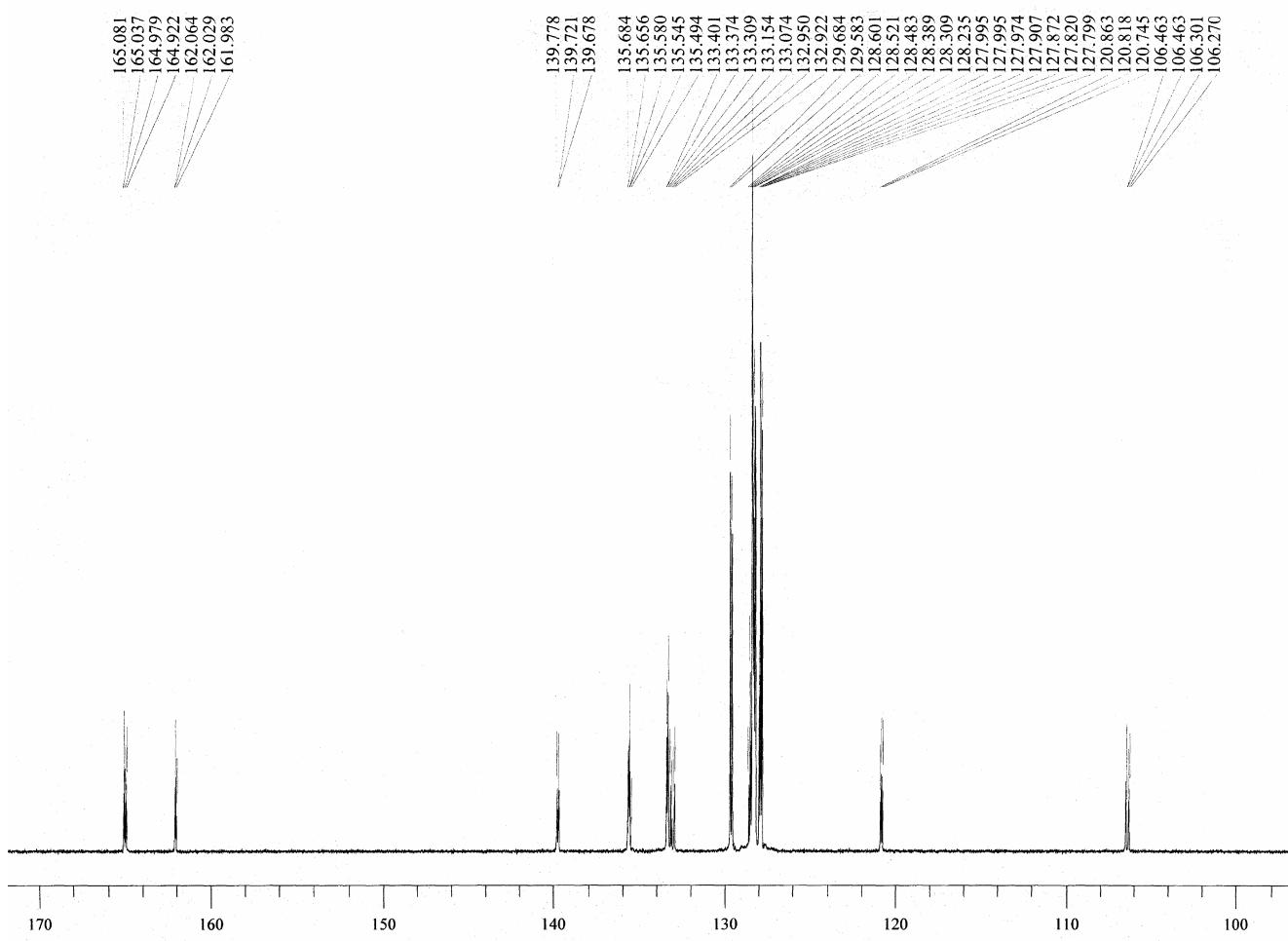
<sup>13</sup>C

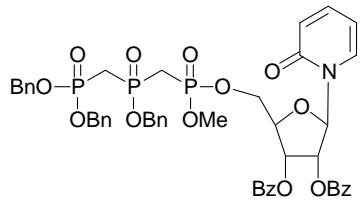




**45 ( $\text{CDCl}_3$ )**

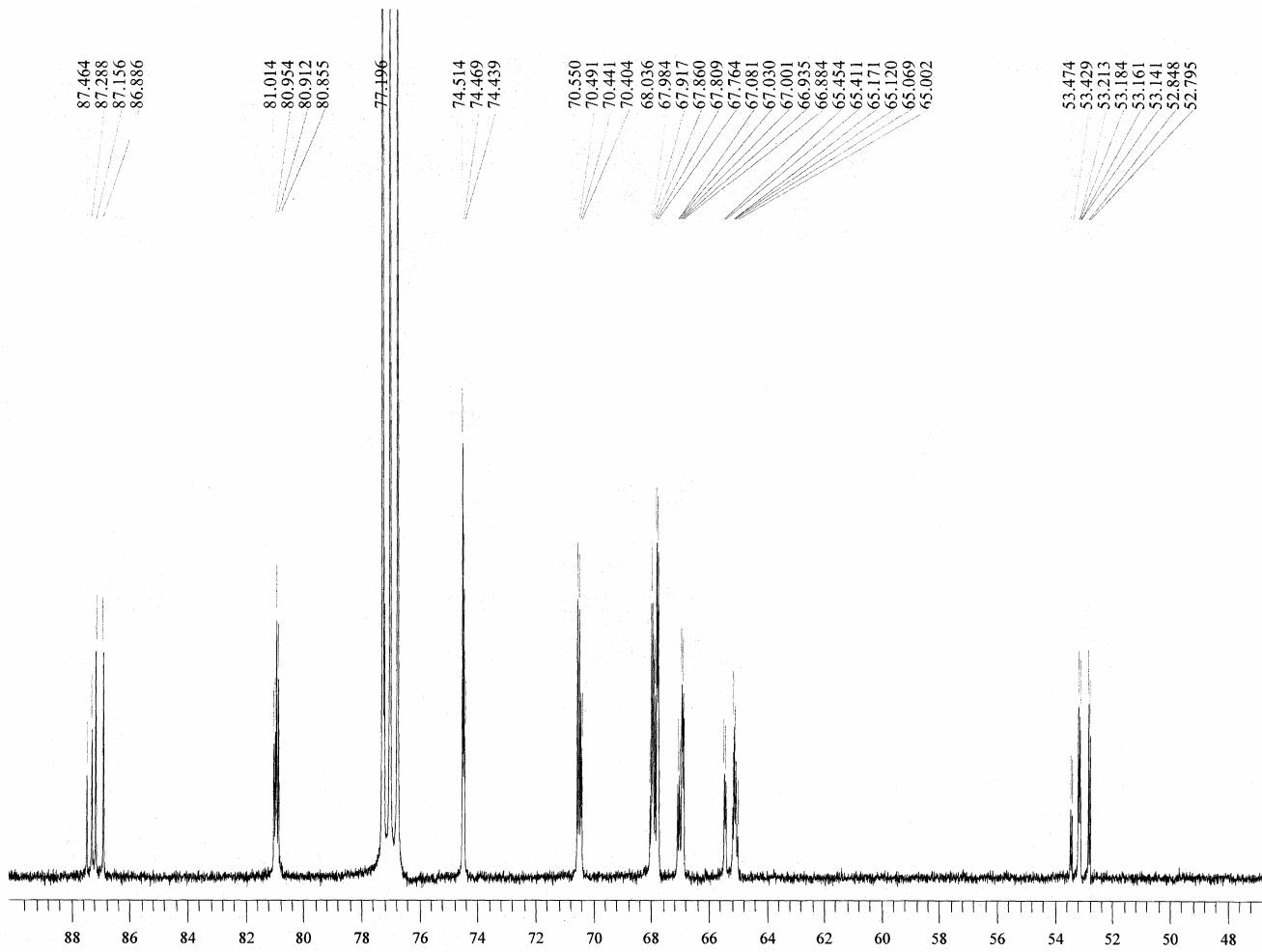
$^{13}\text{C}$  (expanded)

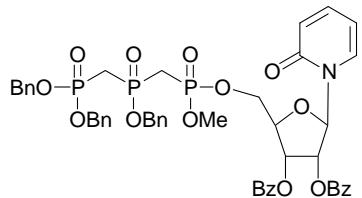




**45 (CDCl<sub>3</sub>)**

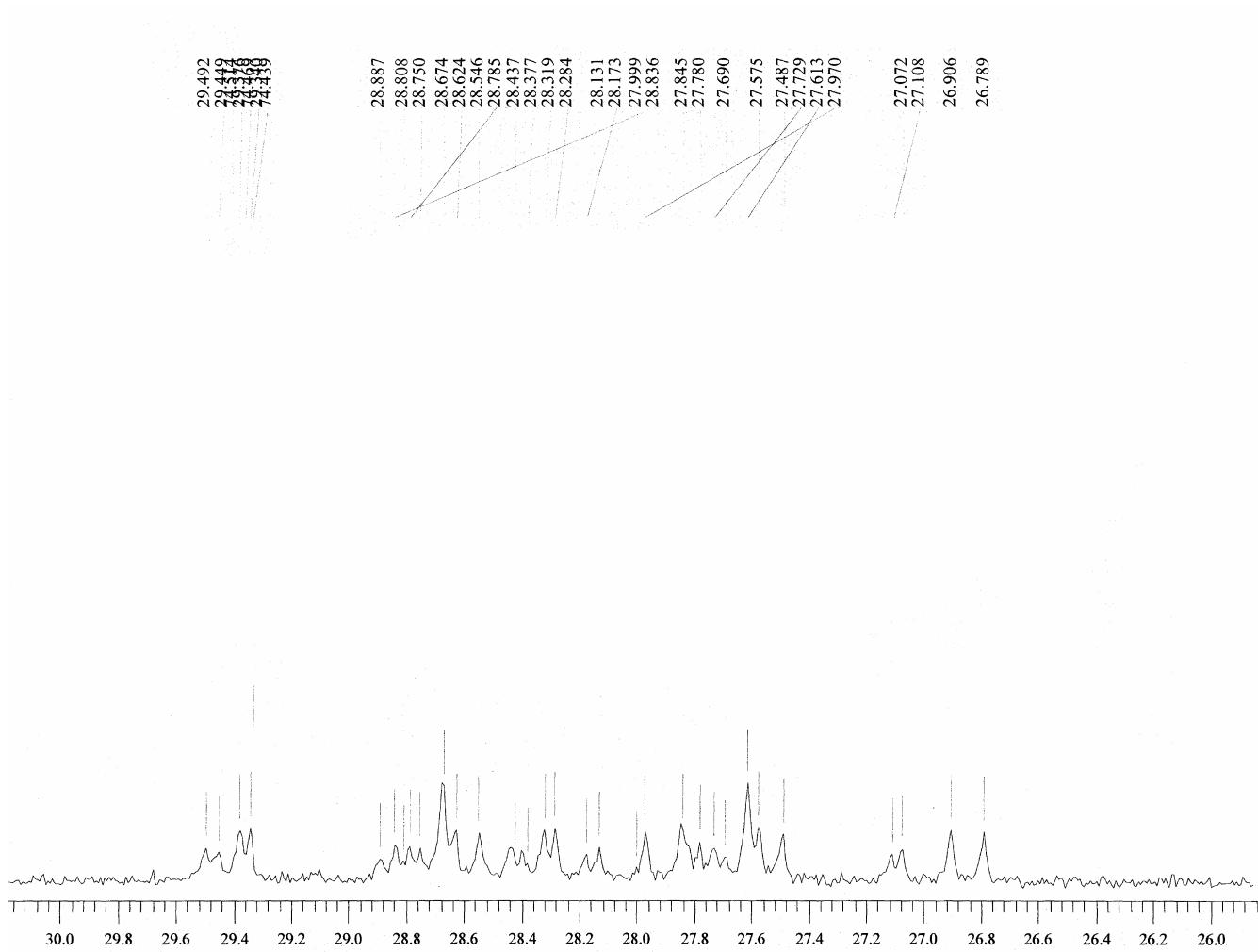
<sup>13</sup>C (expanded)

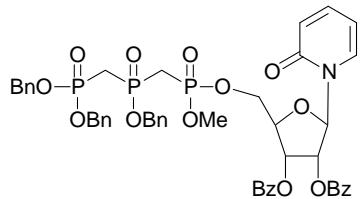




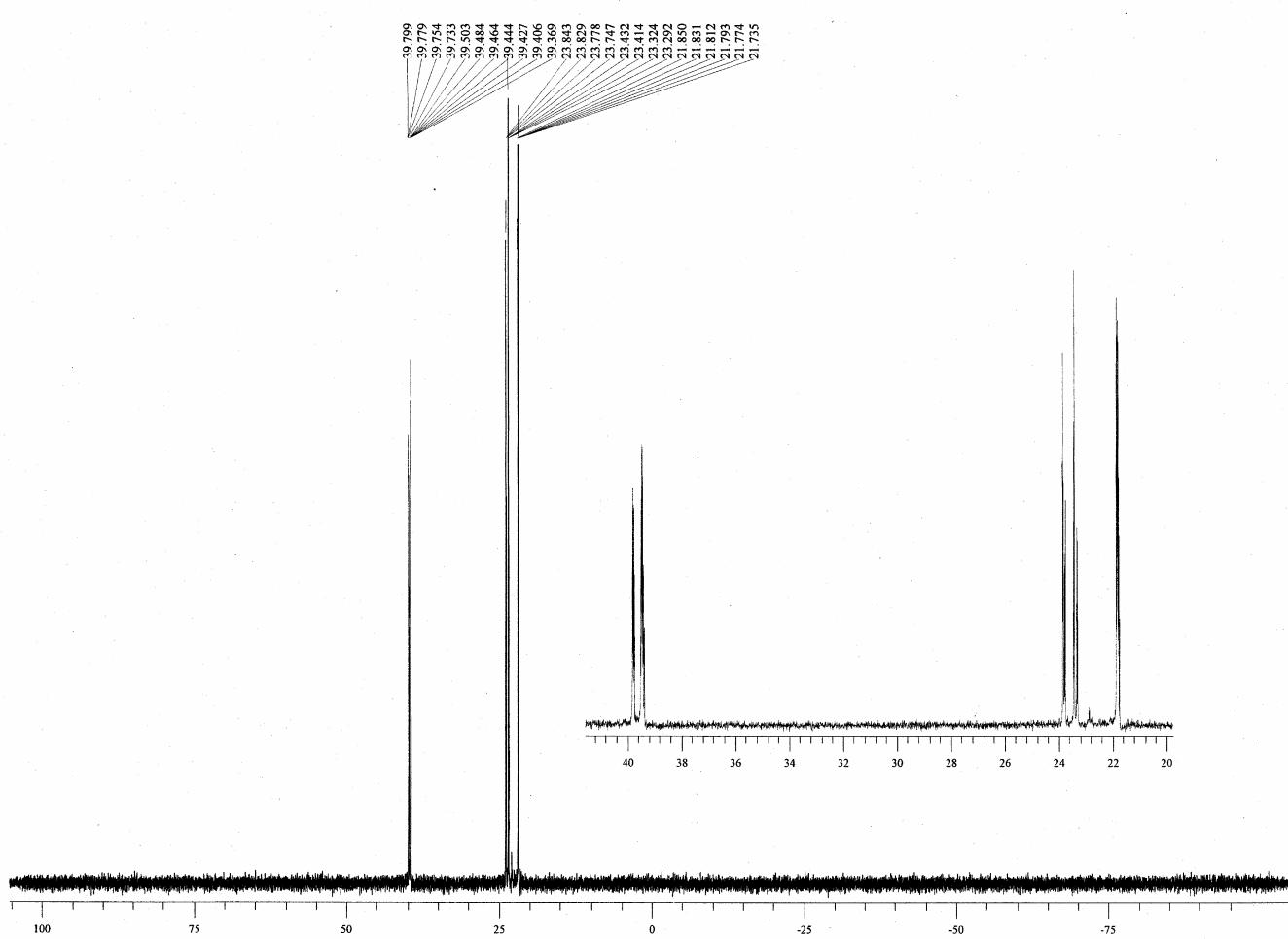
**45 ( $\text{CDCl}_3$ )**

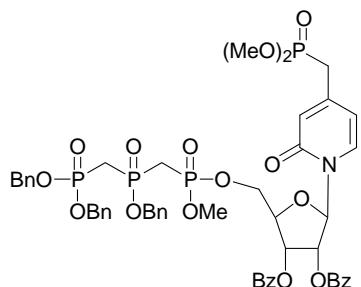
$^{13}\text{C}$  (expanded)





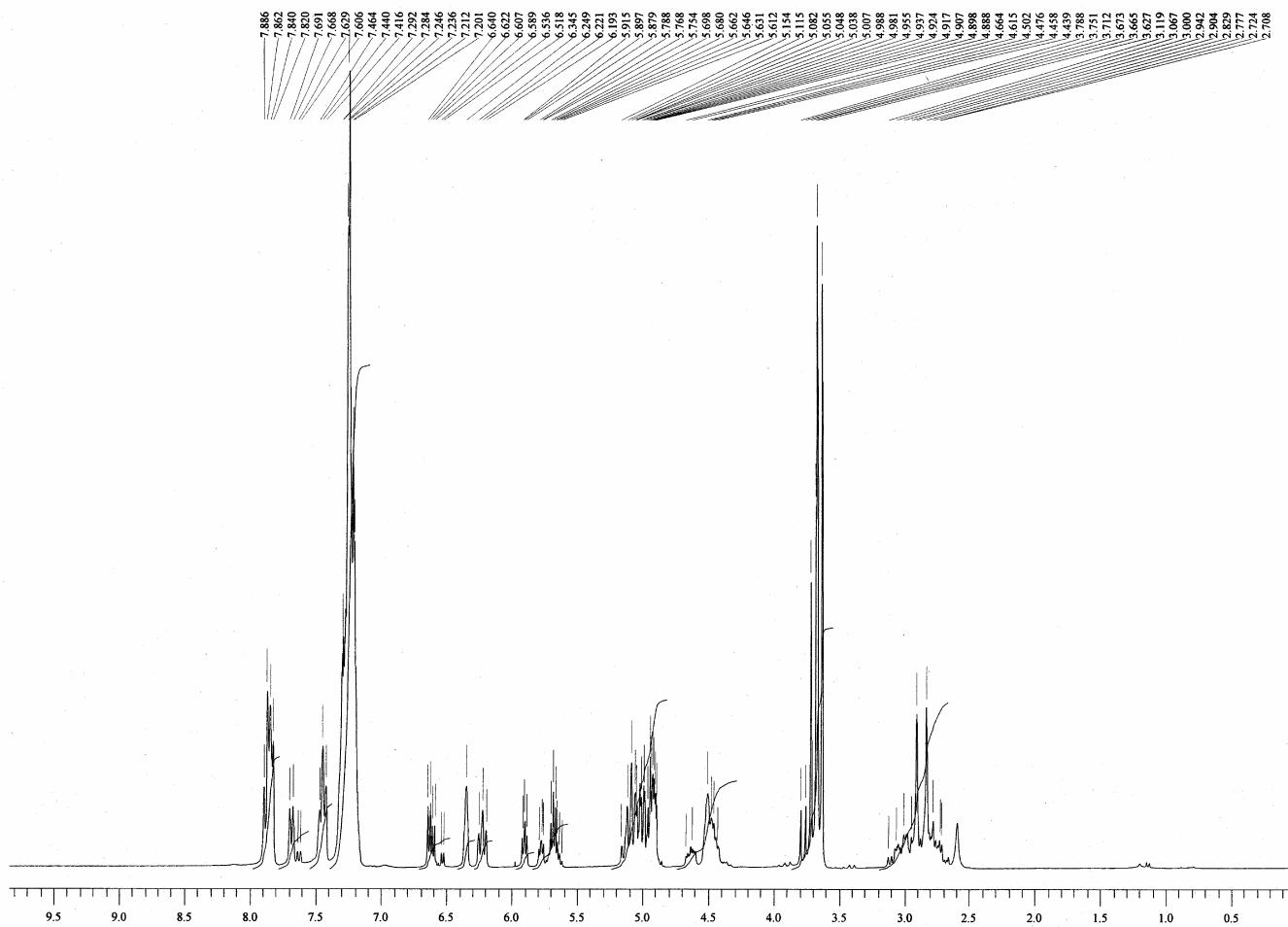
$^{31}\text{P}$

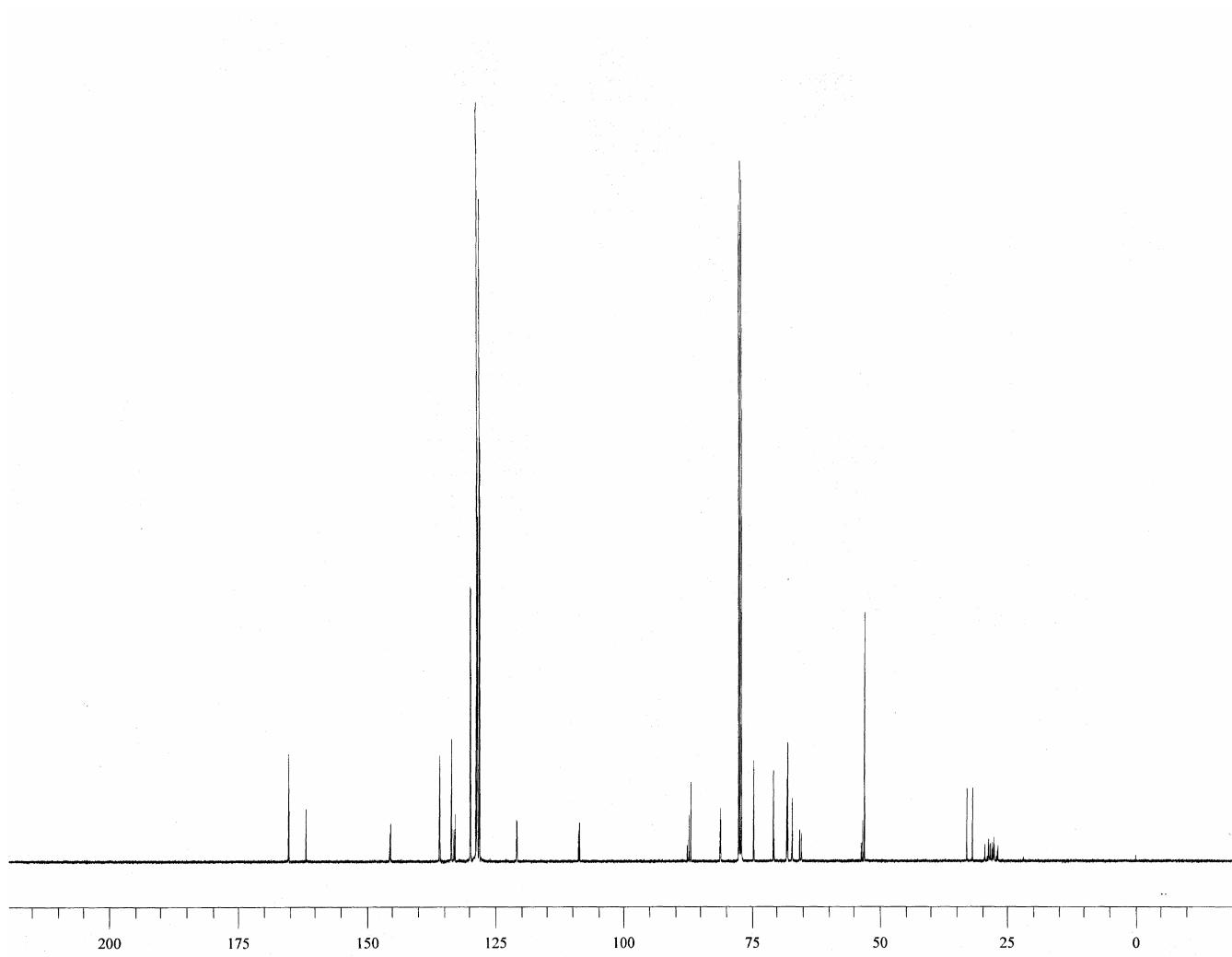
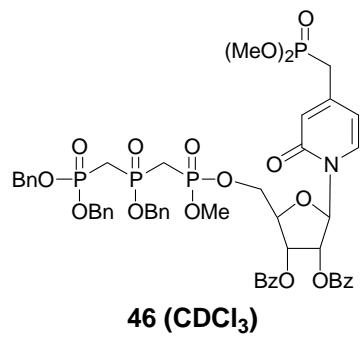


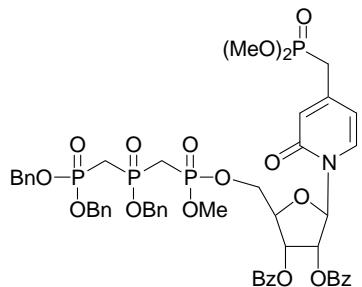


**46 (CDCl<sub>3</sub>)**

<sup>1</sup>H

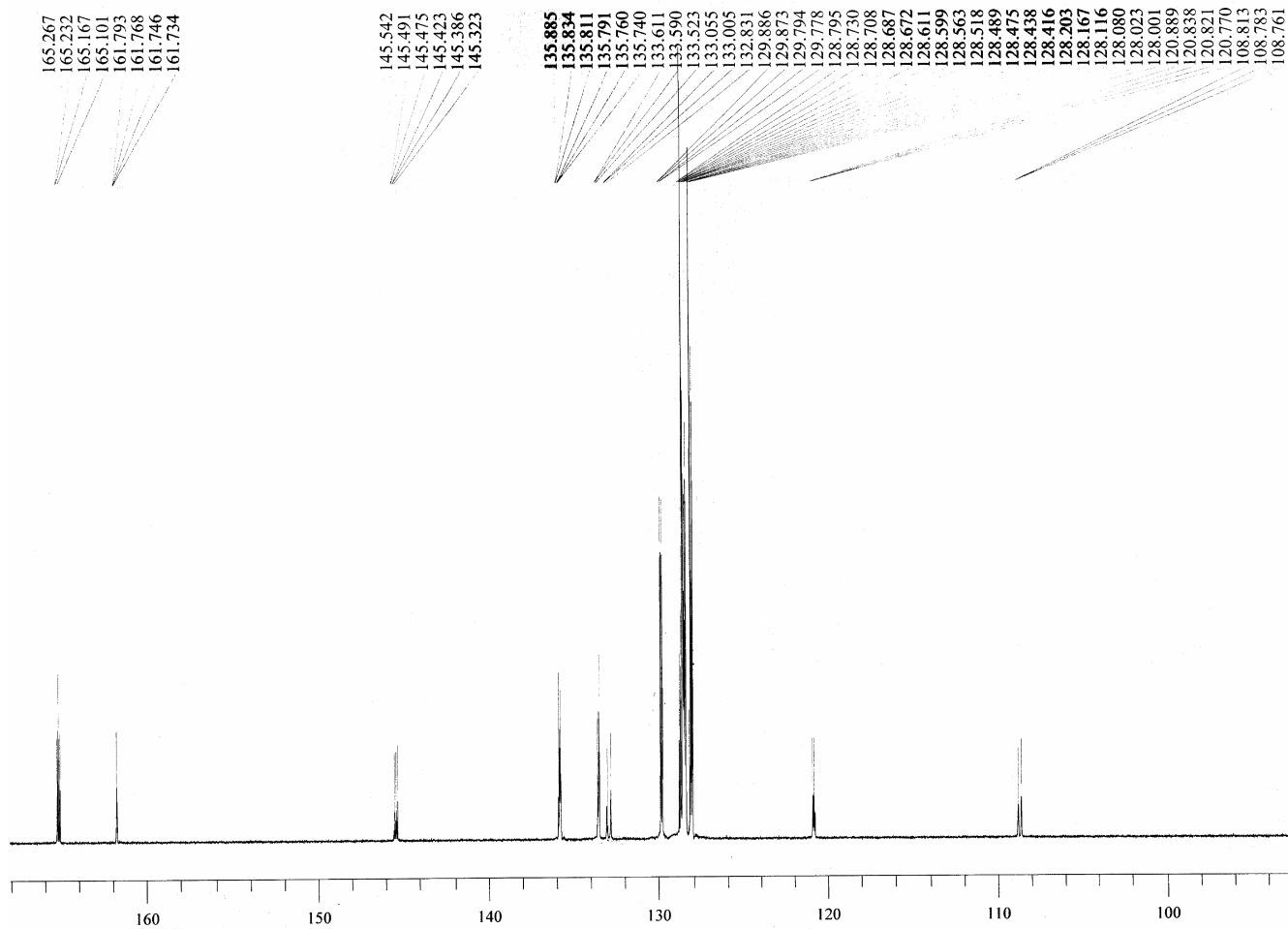


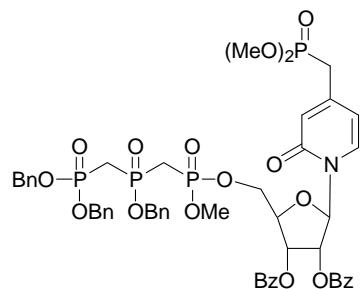




**46 (CDCl<sub>3</sub>)**

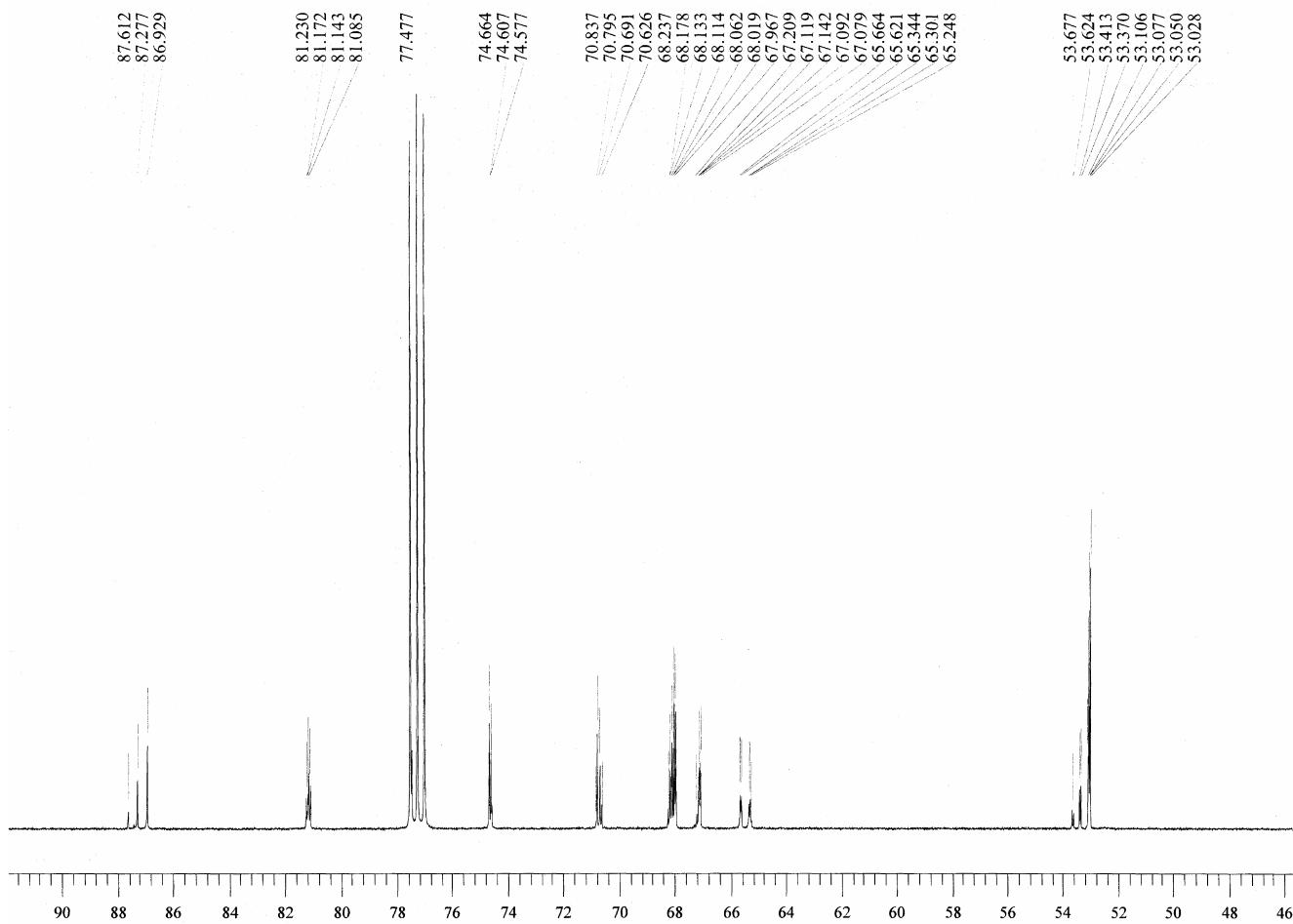
<sup>13</sup>C (expanded)

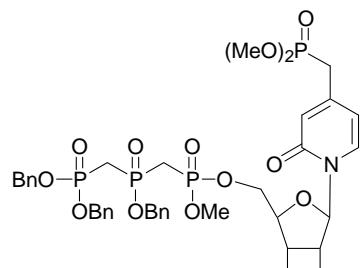




**46 (CDCl<sub>3</sub>)**

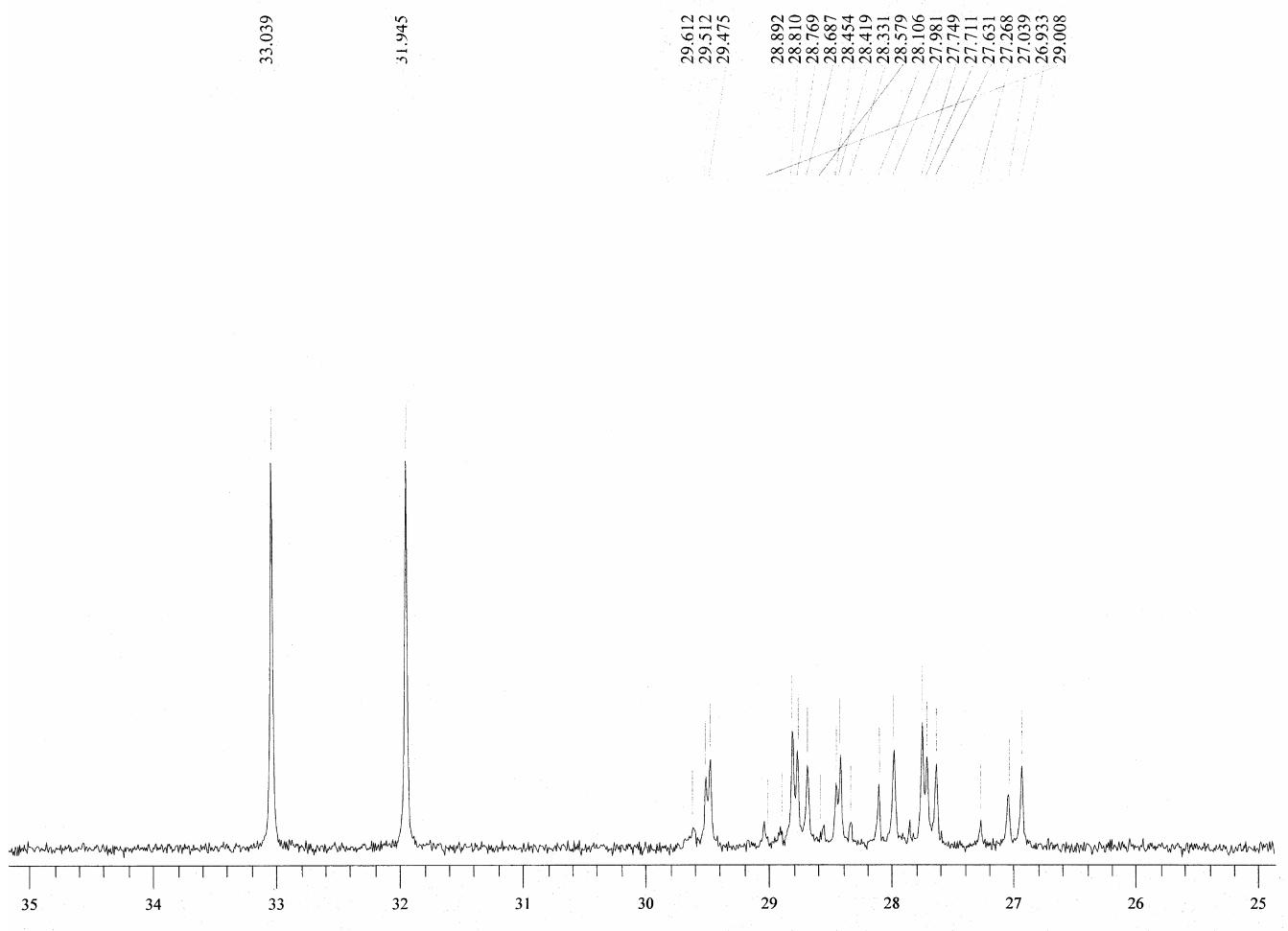
<sup>13</sup>C (expanded)

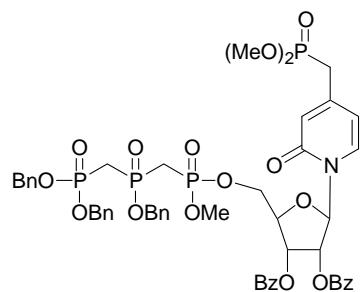




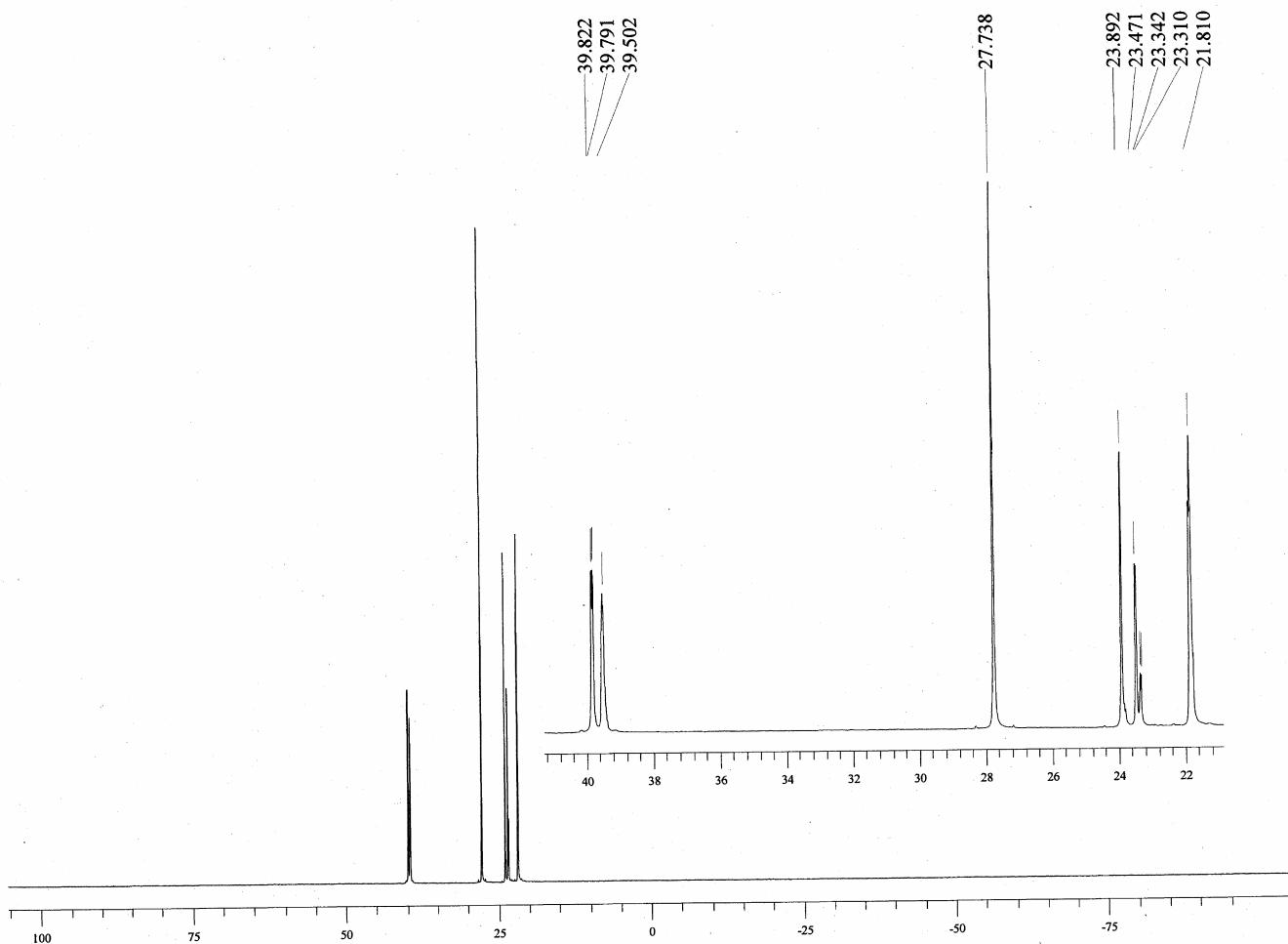
**46 ( $\text{CDCl}_3$ )**

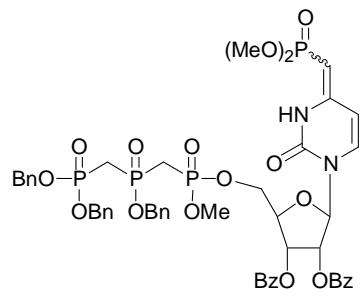
$^{13}\text{C}$  (expanded)





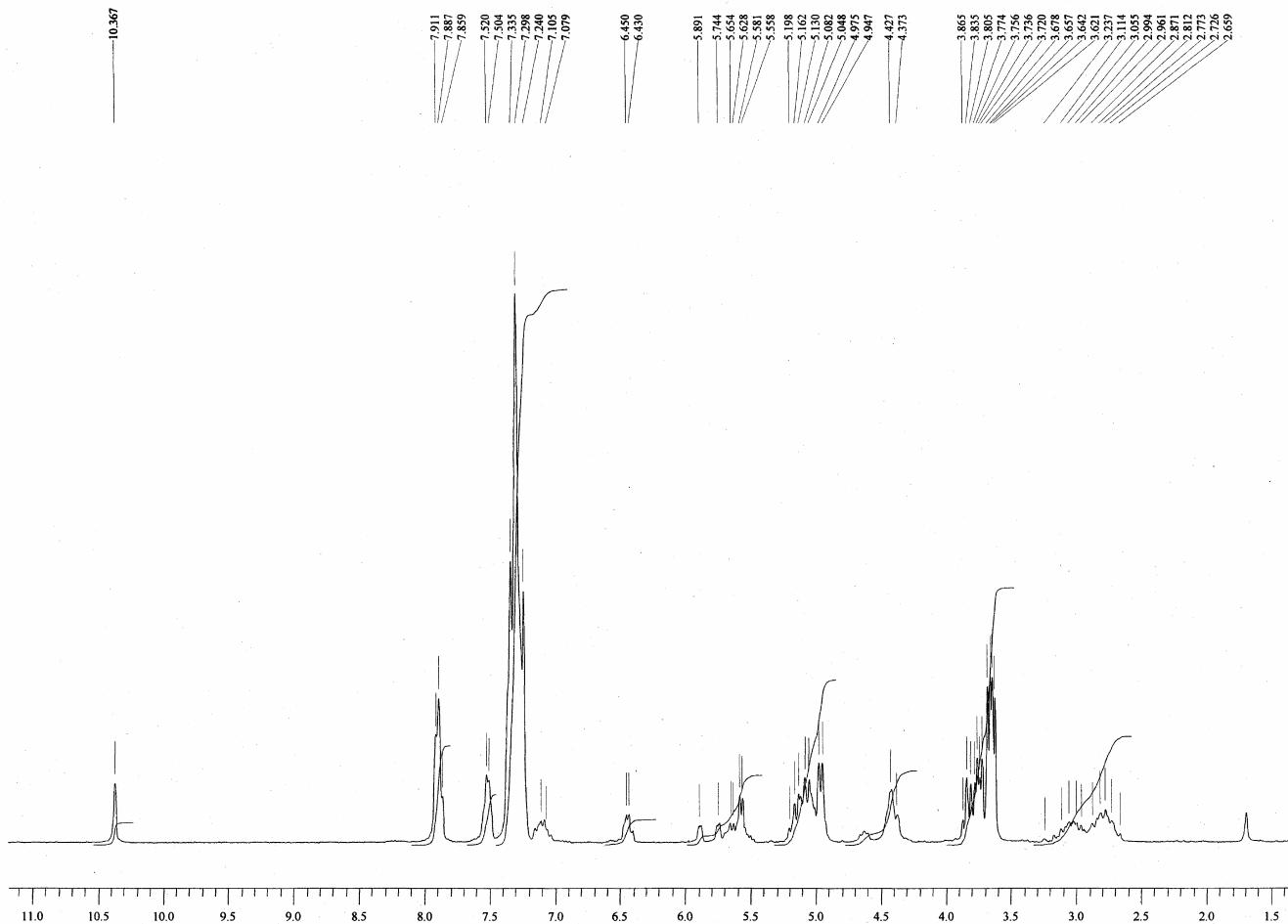
$^{31}\text{P}$

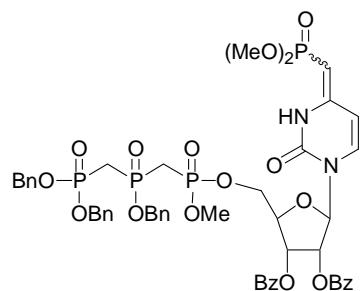




**47 ( $\text{CDCl}_3$ )**

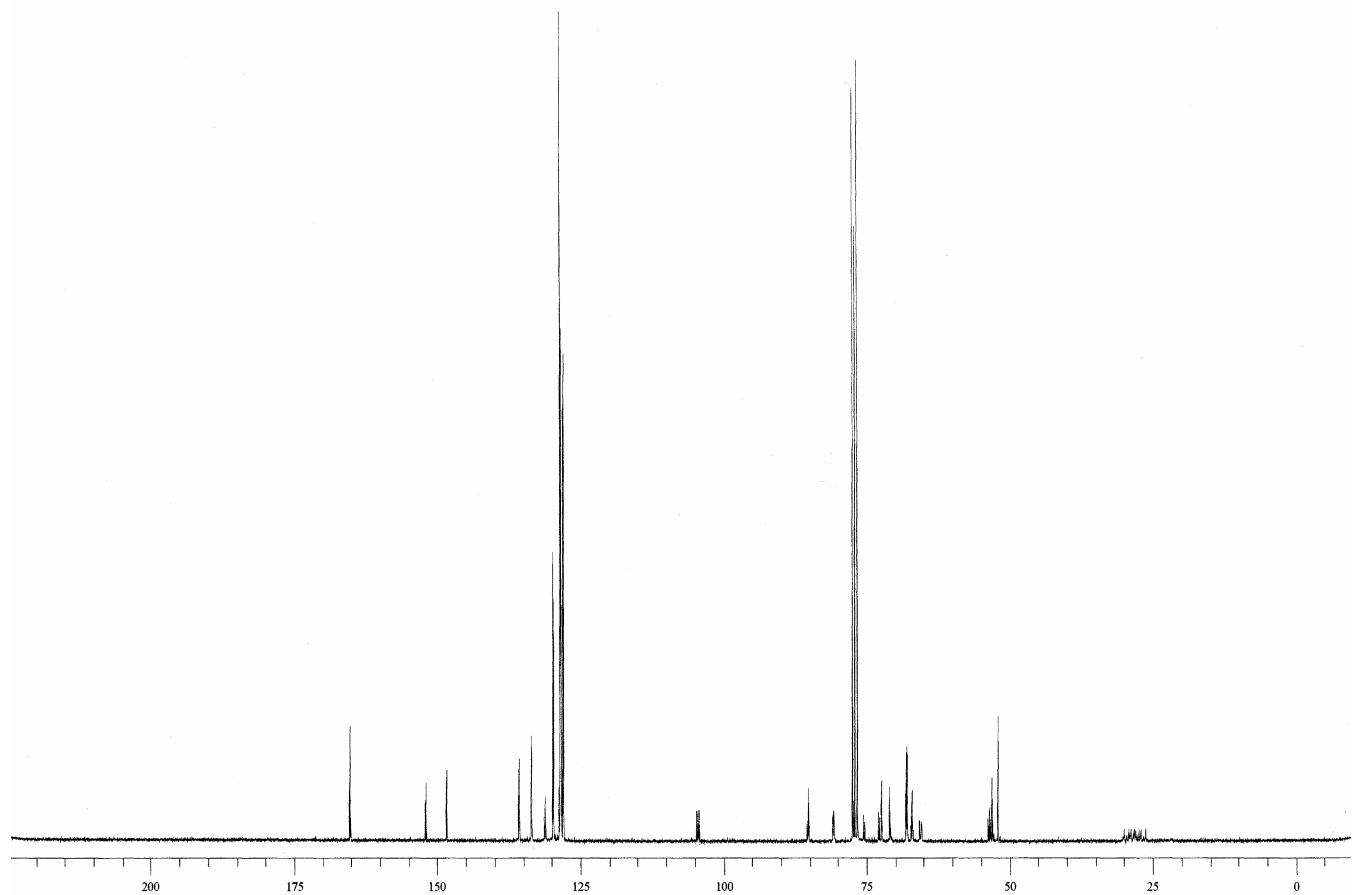
$^1\text{H}$

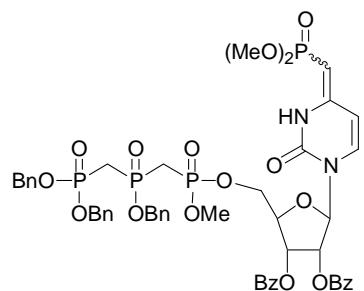




47 ( $\text{CDCl}_3$ )

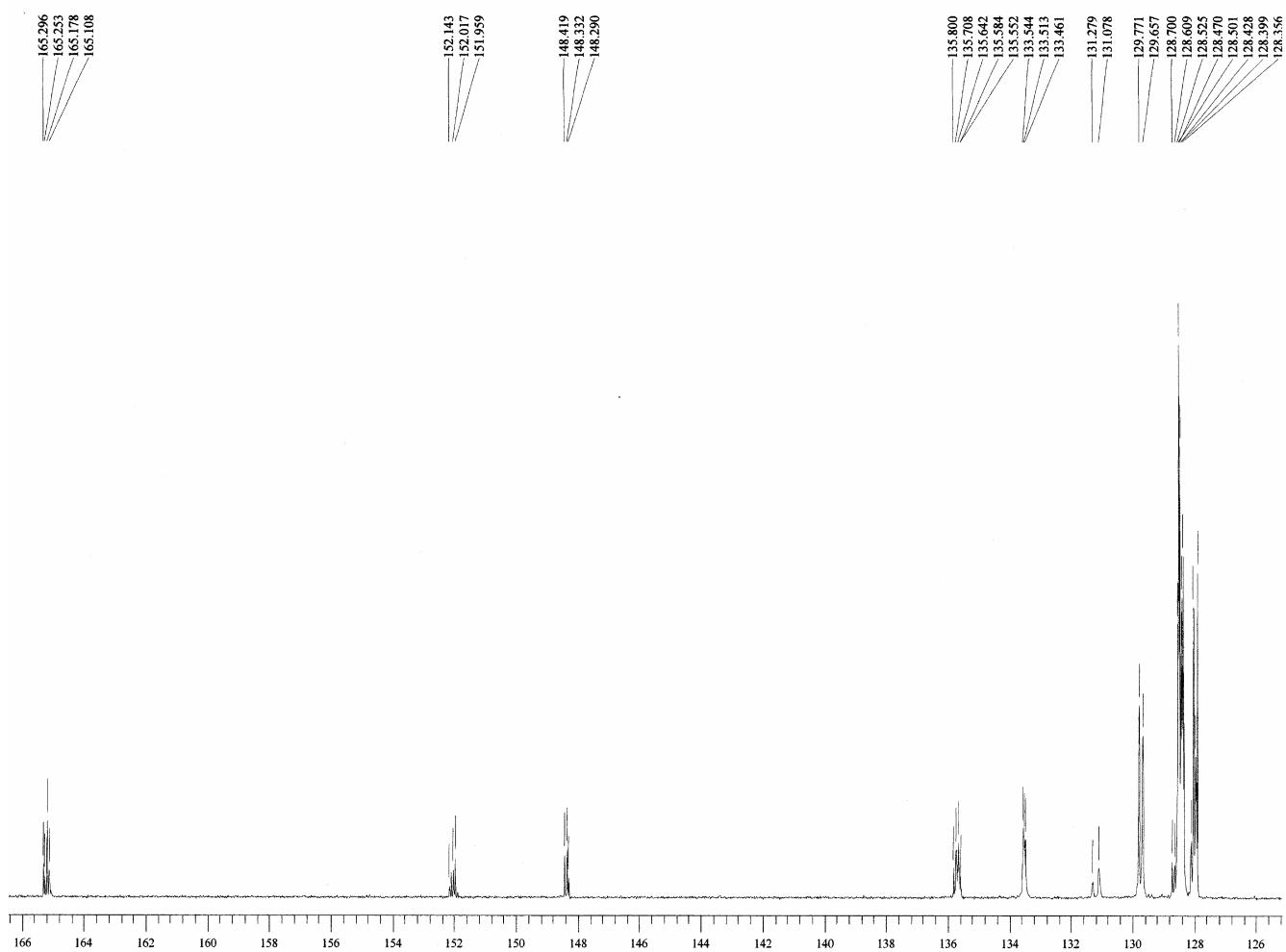
$^{13}\text{C}$

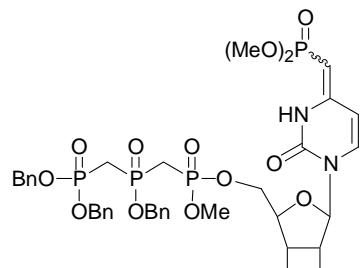




**47 (CDCl<sub>3</sub>)**

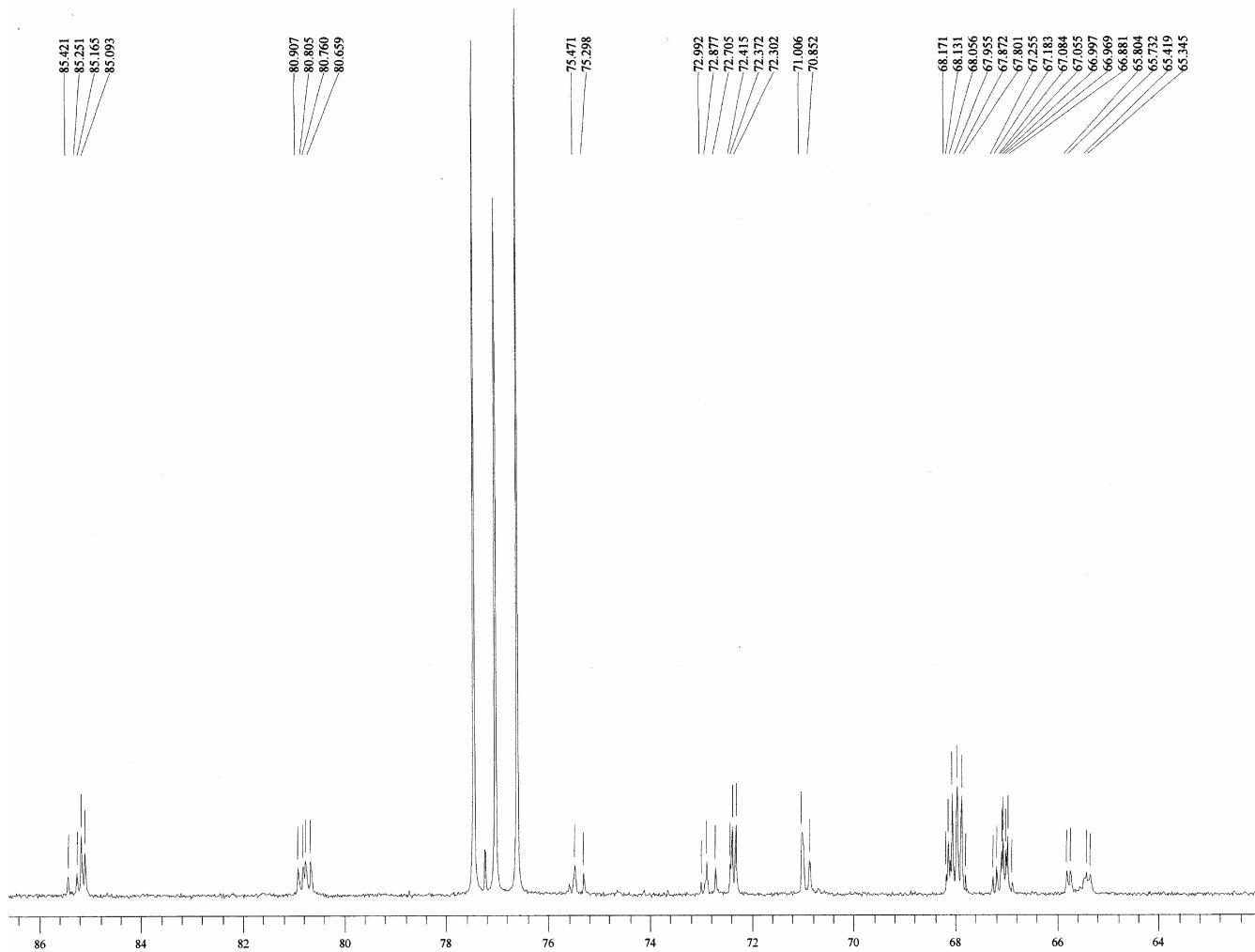
<sup>13</sup>C (expanded)

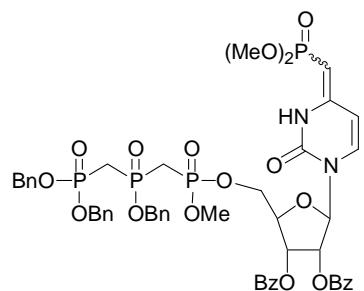




**47 ( $\text{CDCl}_3$ )**

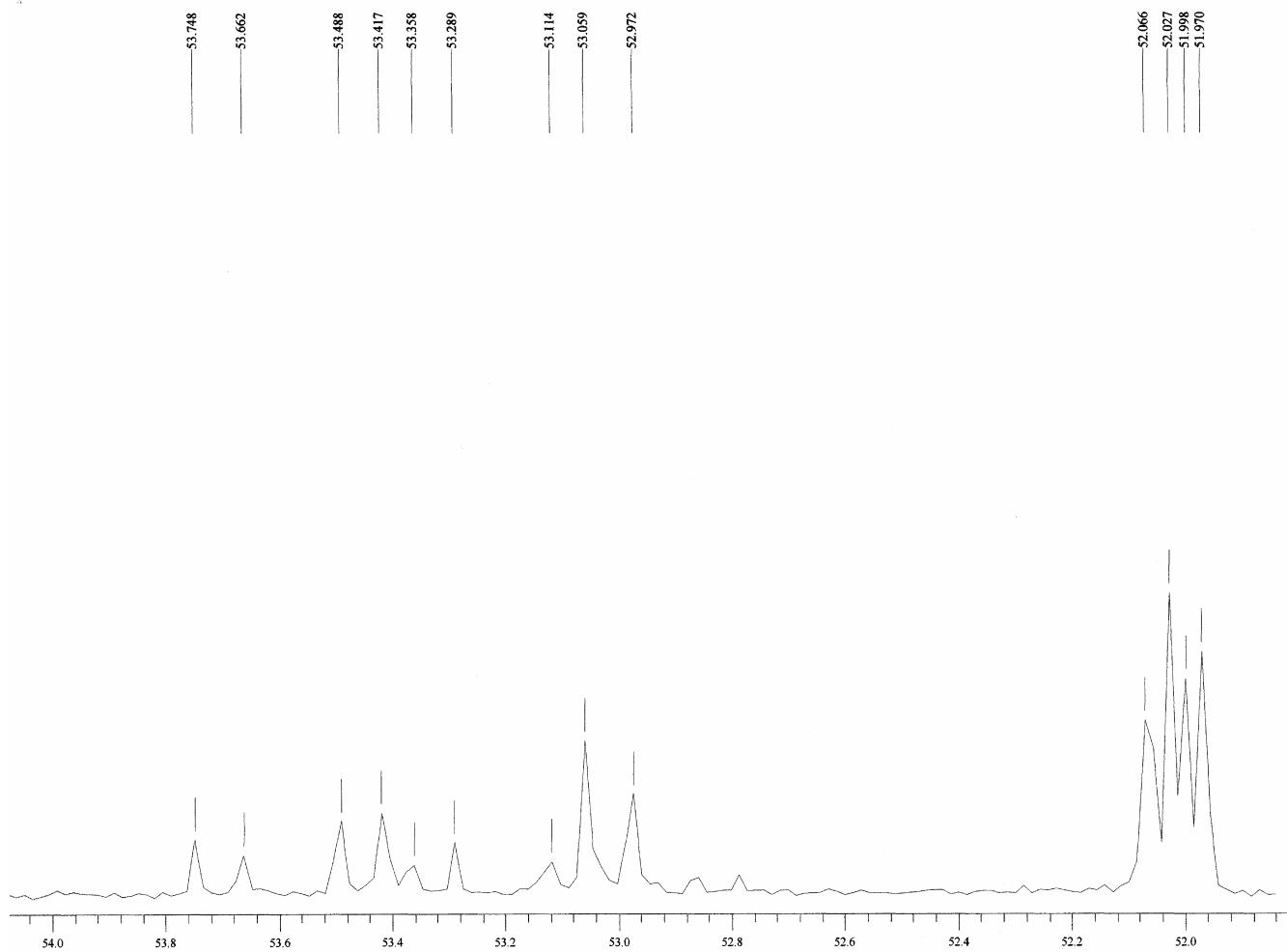
$^{13}\text{C}$  (expanded)

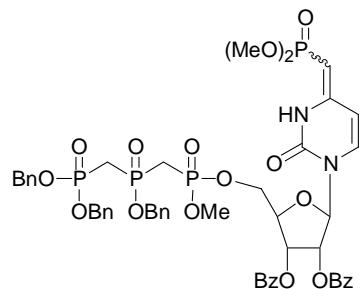




**47 (CDCl<sub>3</sub>)**

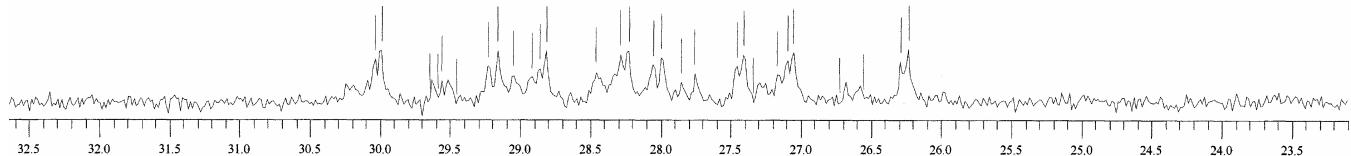
<sup>13</sup>C (expanded)

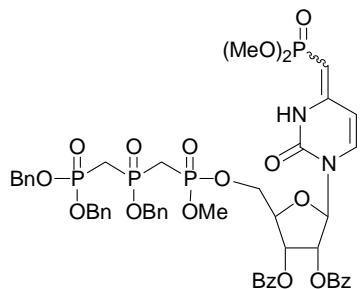




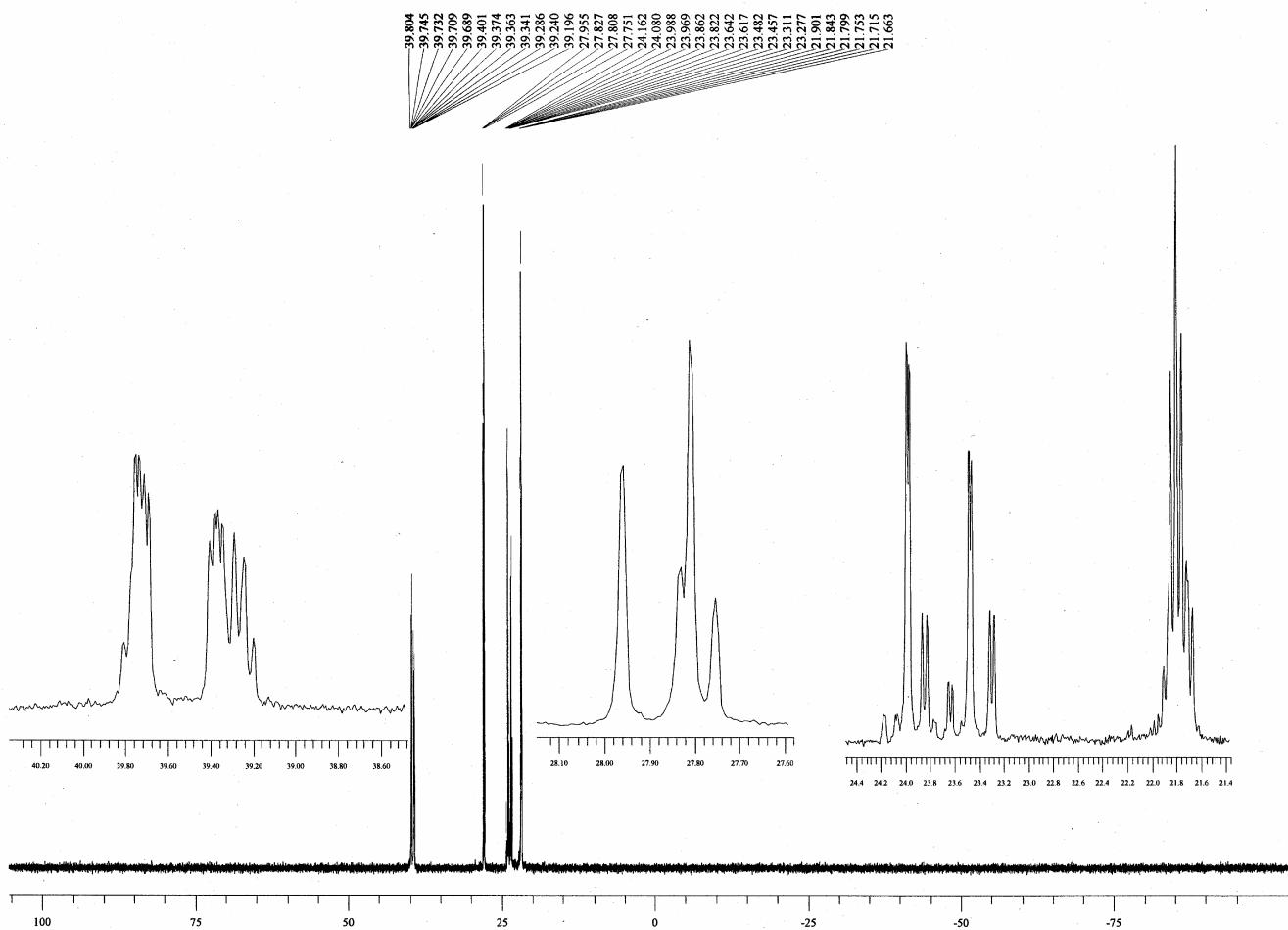
**47 (CDCl<sub>3</sub>)**

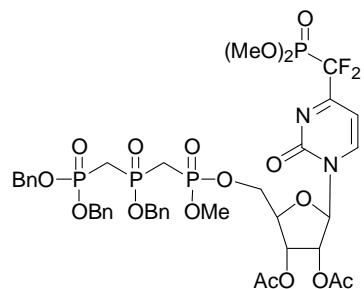
<sup>13</sup>C (expanded)



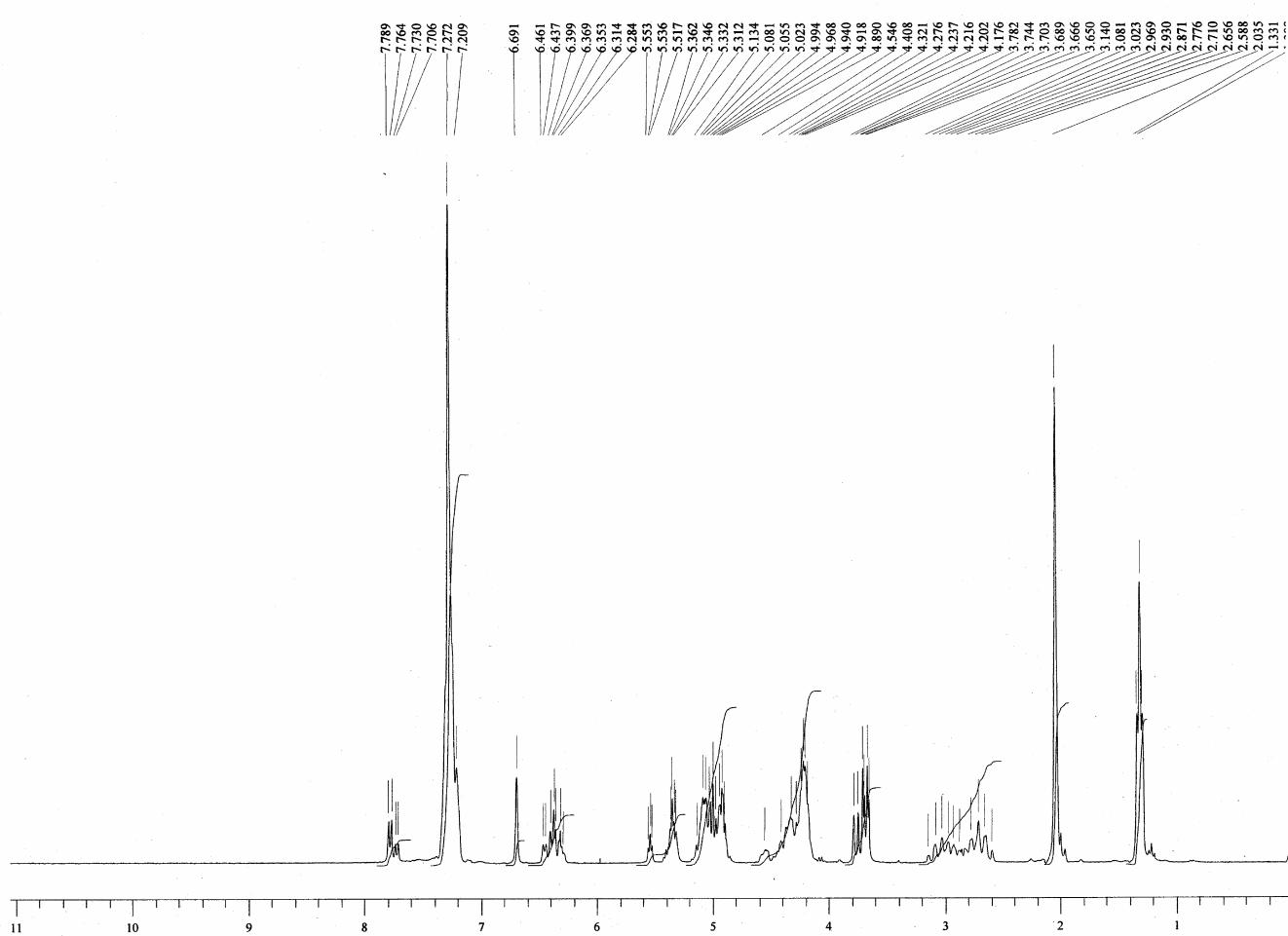


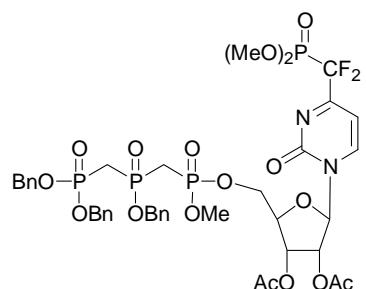
$^{31}\text{P}$





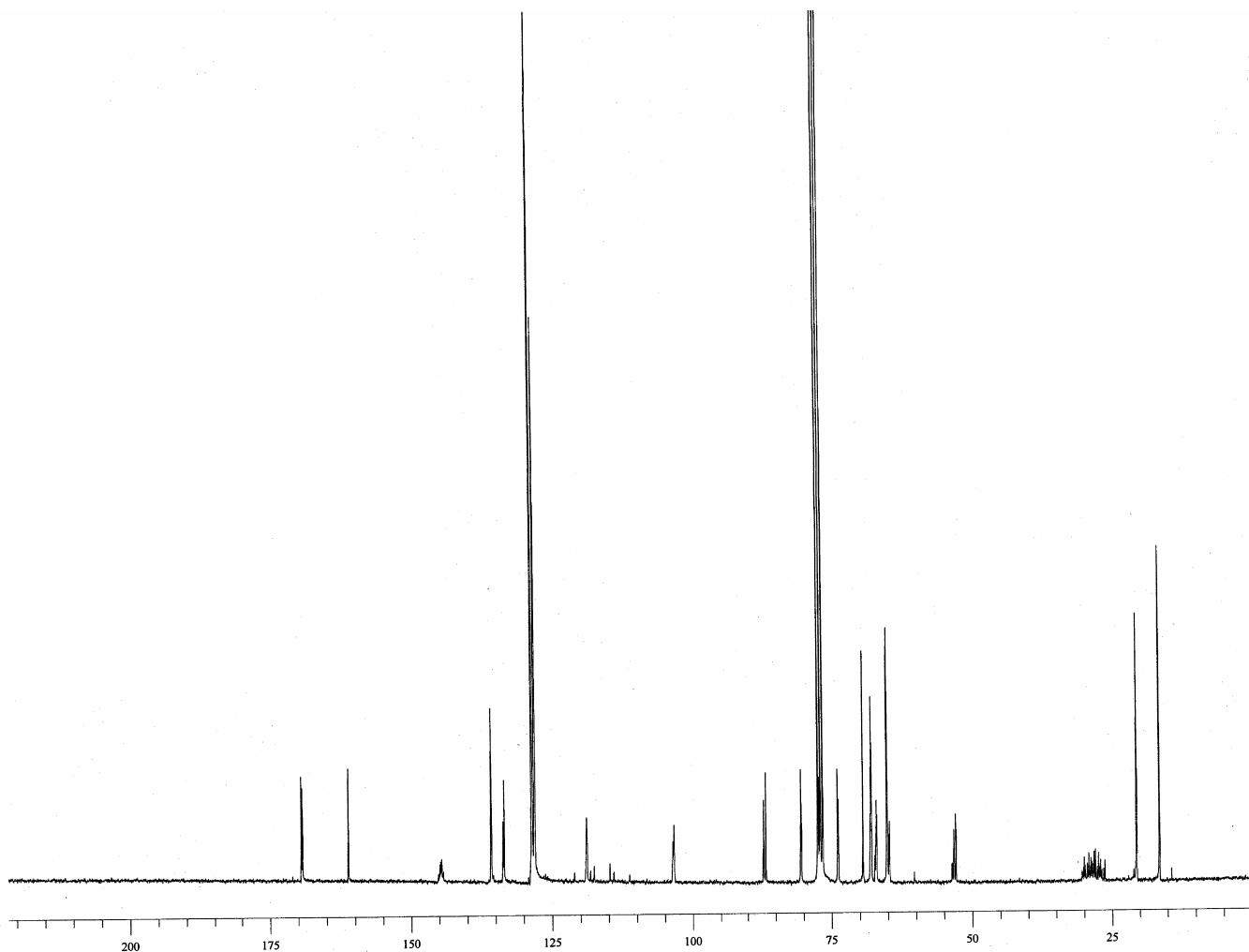
<sup>1</sup>H

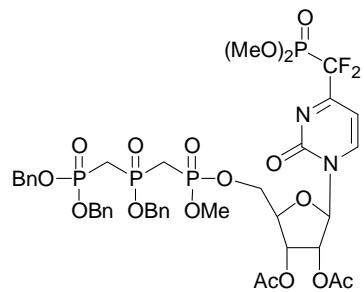




48 (CDCl<sub>3</sub>)

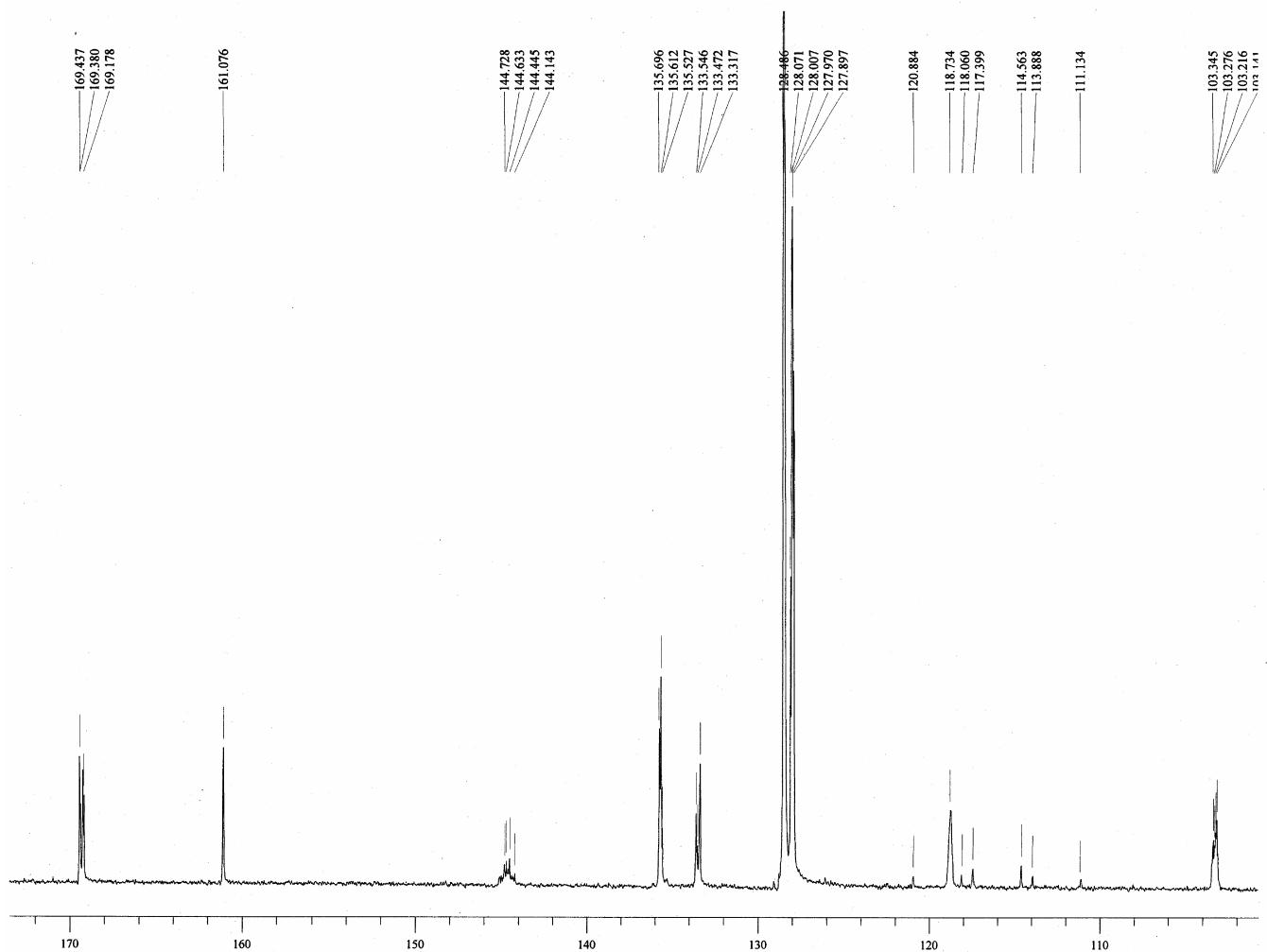
<sup>13</sup>C

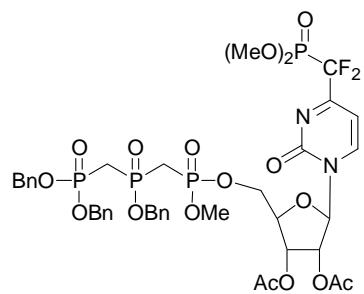




**48 (CDCl<sub>3</sub>)**

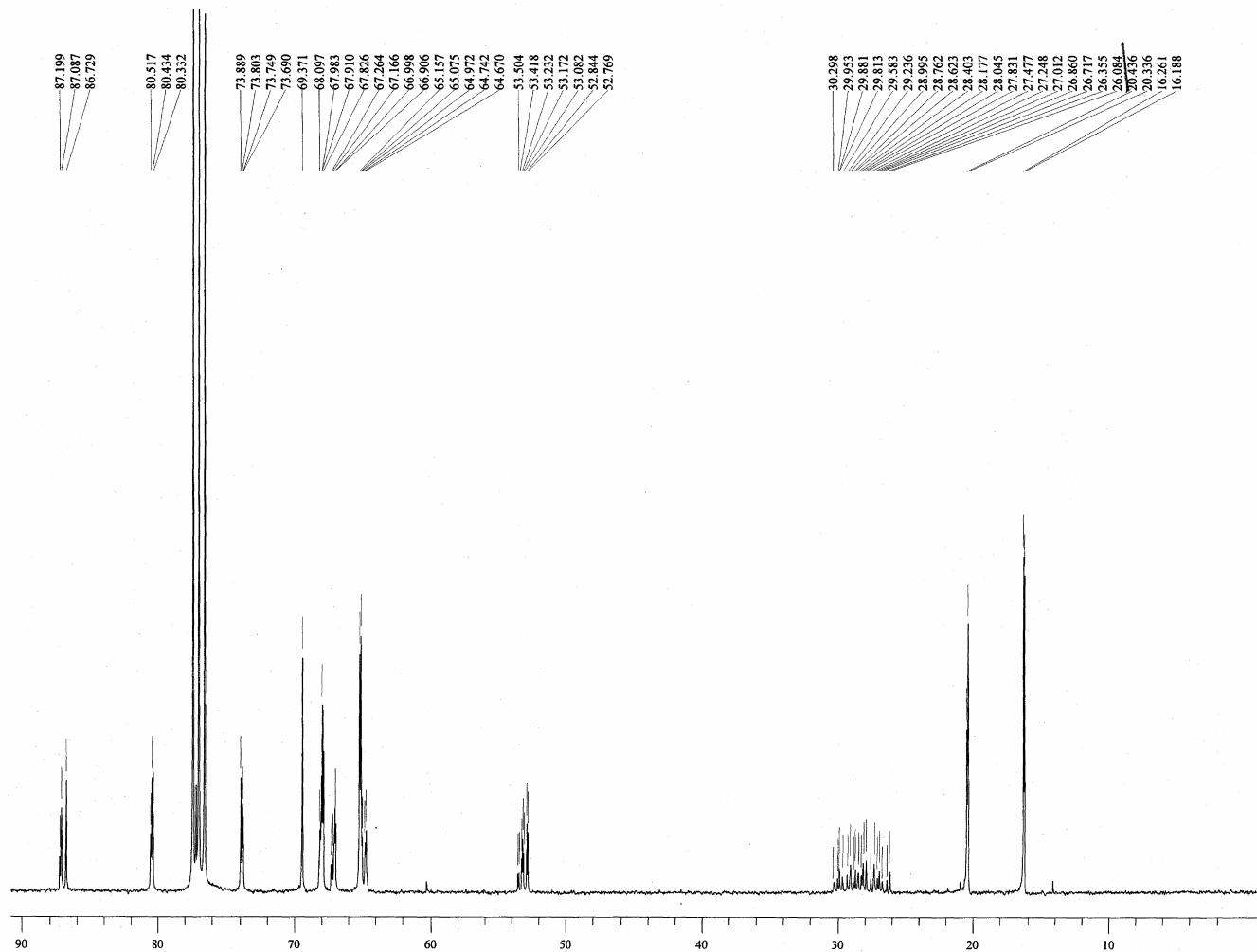
<sup>13</sup>C (expanded)

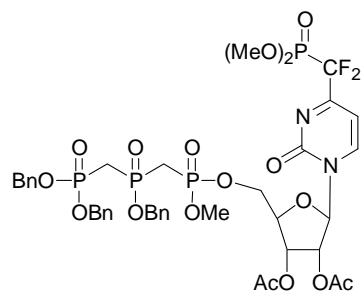




**48 (CDCl<sub>3</sub>)**

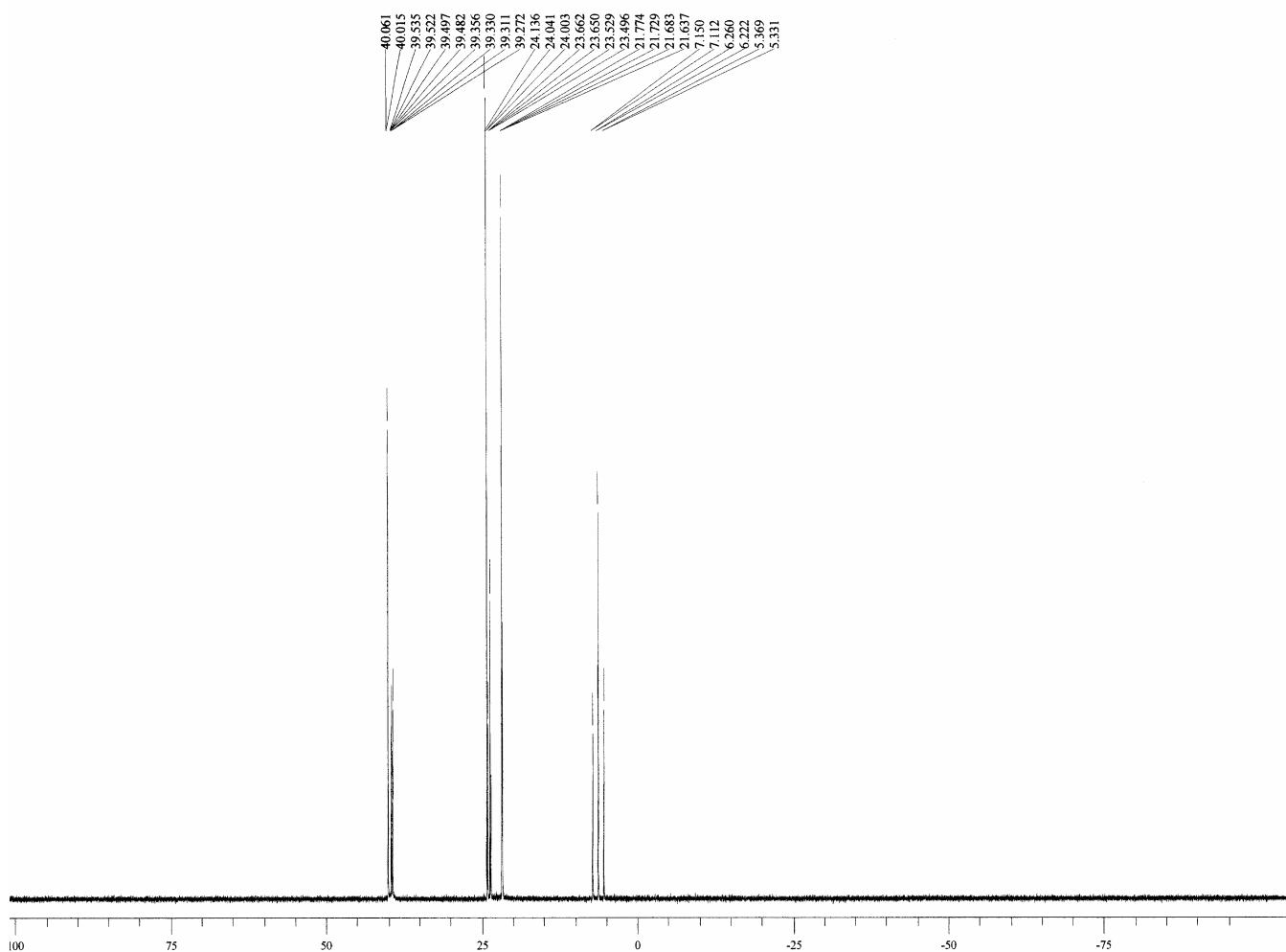
<sup>13</sup>C (expanded)

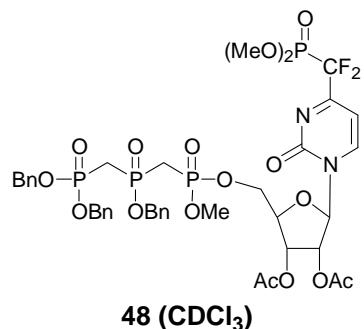




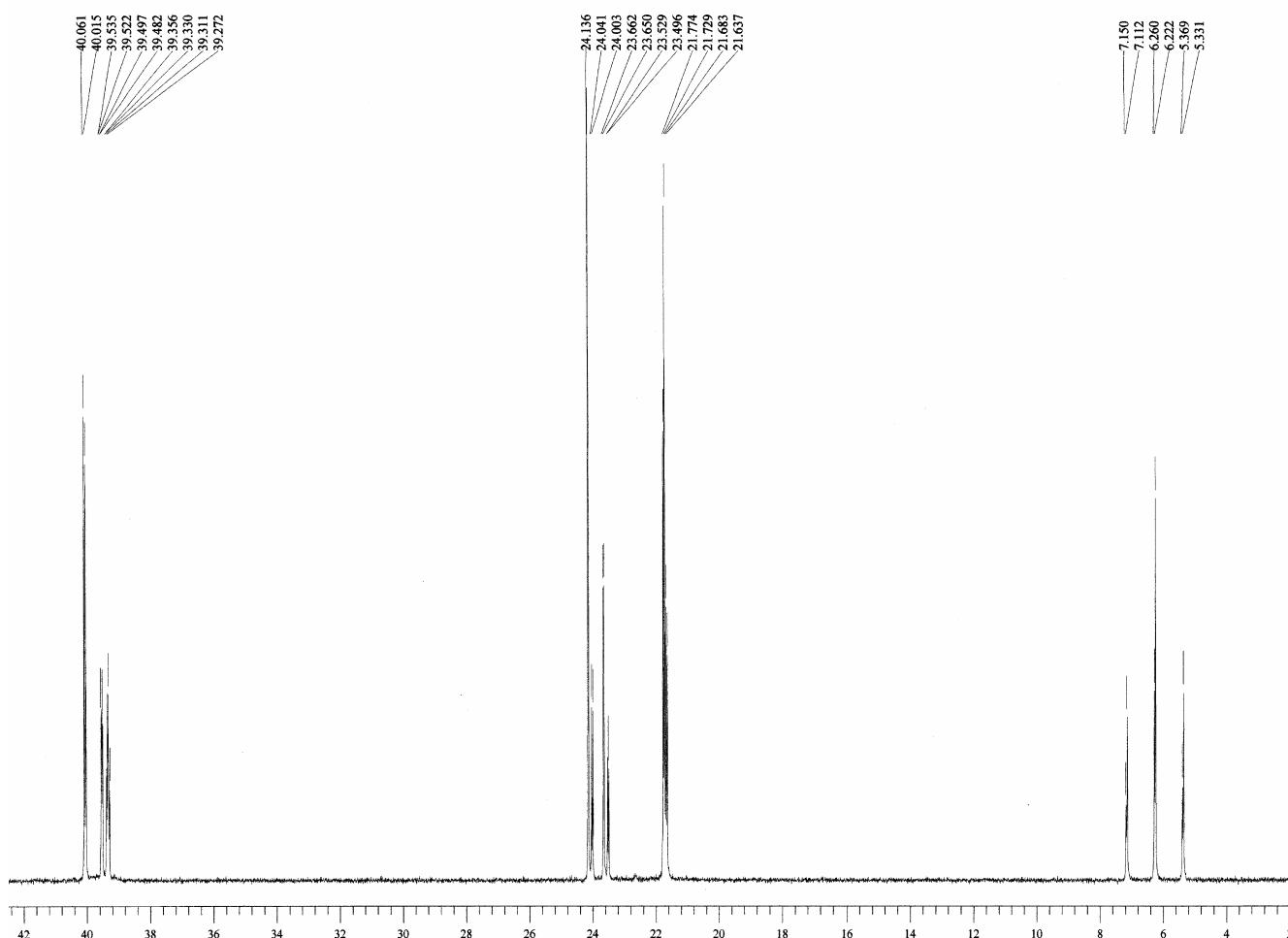
**48 (CDCl<sub>3</sub>)**

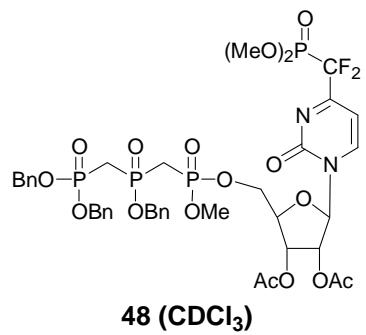
<sup>31</sup>P



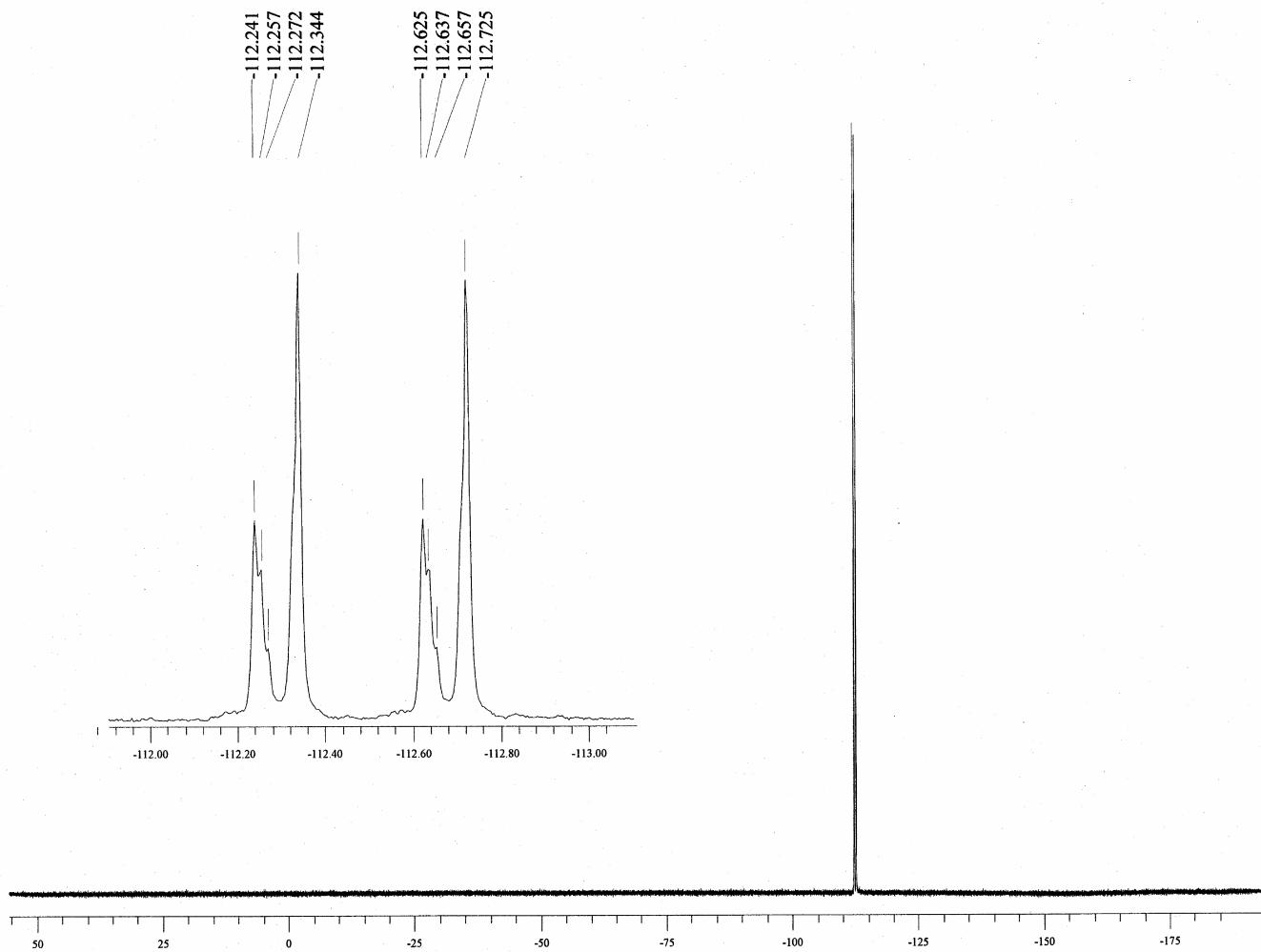


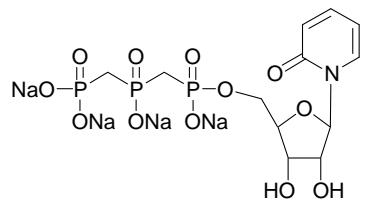
<sup>31</sup>P (expanded)





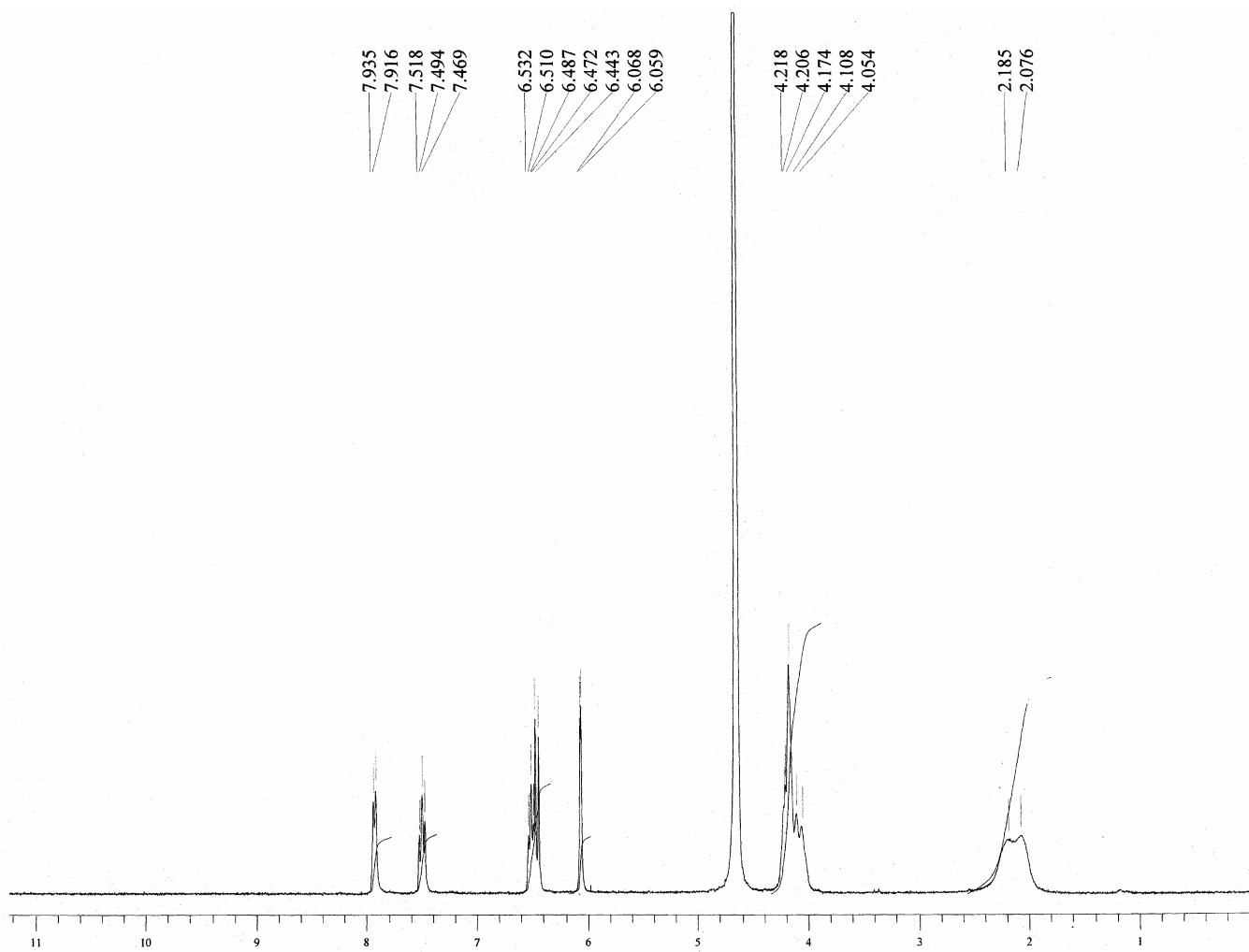
<sup>19</sup>F

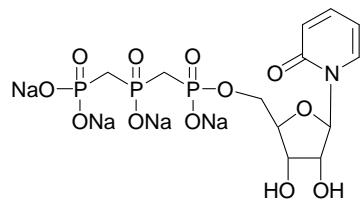




**49 ( $D_2O$ )**

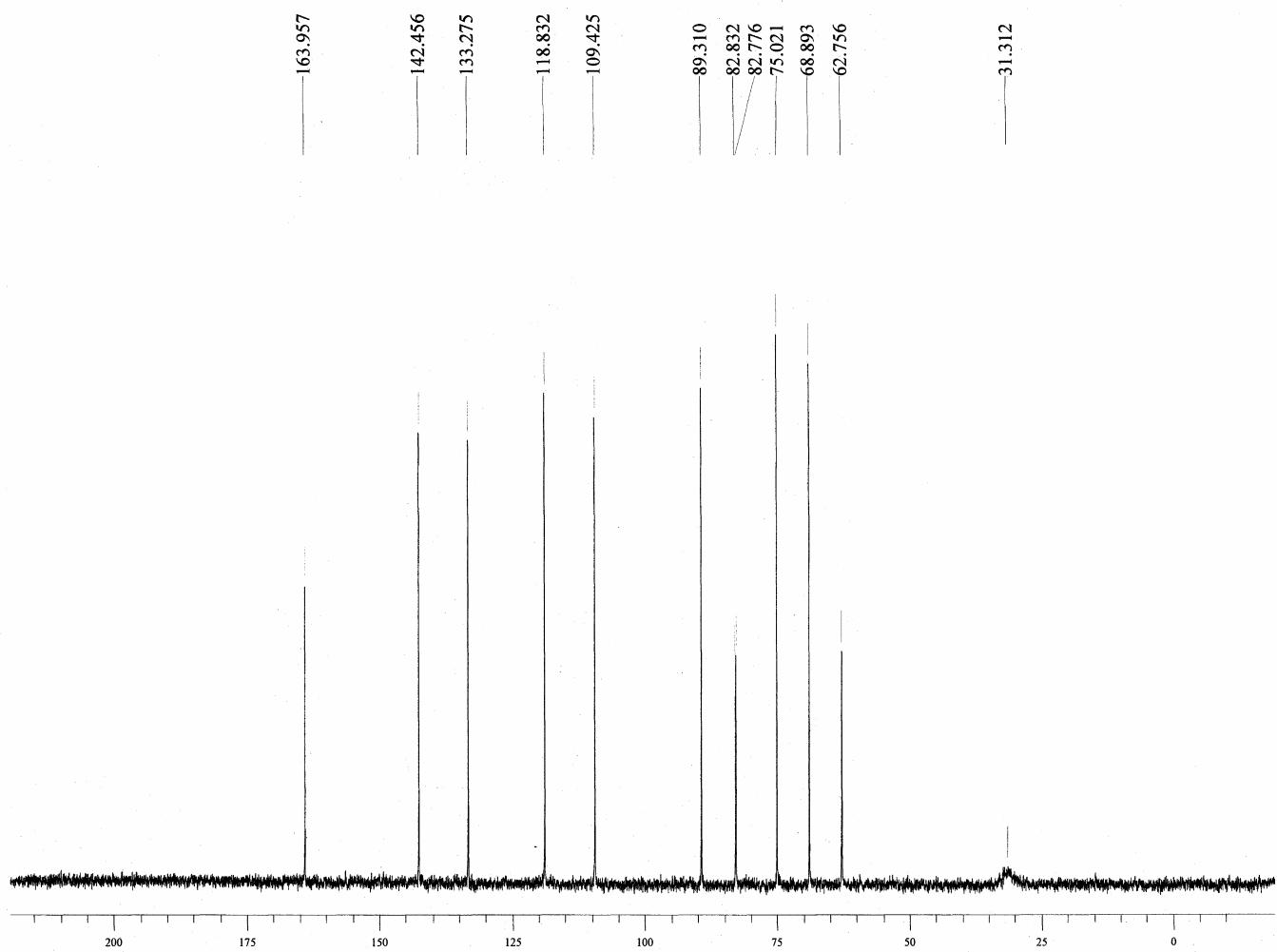
$^1H$

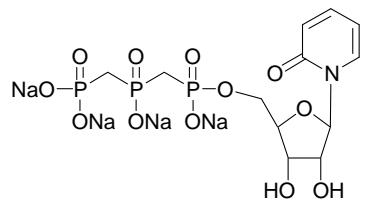




**49 ( $D_2O$ )**

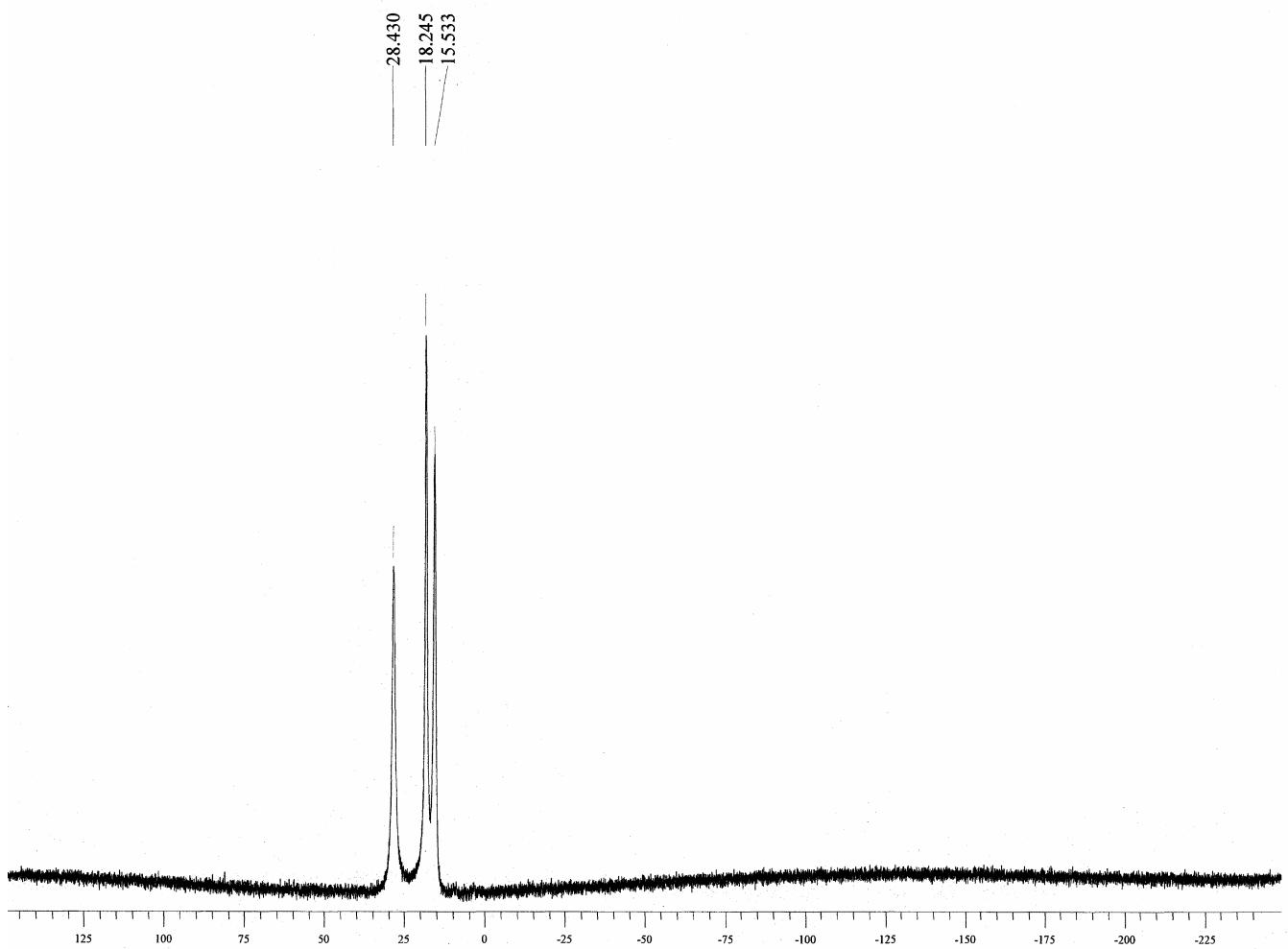
$^{13}C$

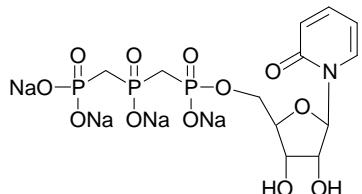




**49 ( $D_2O$ )**

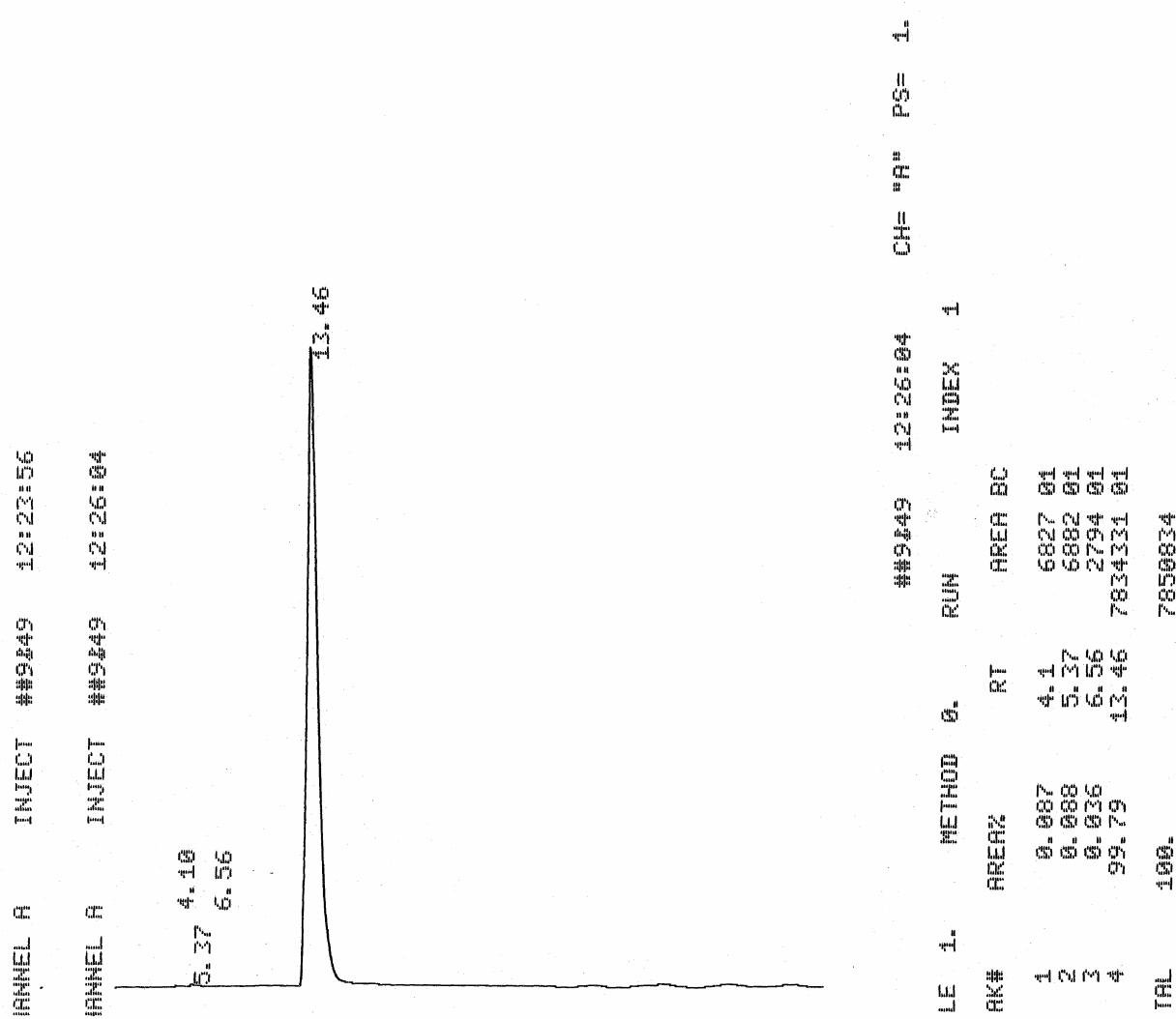
$^{31}P$



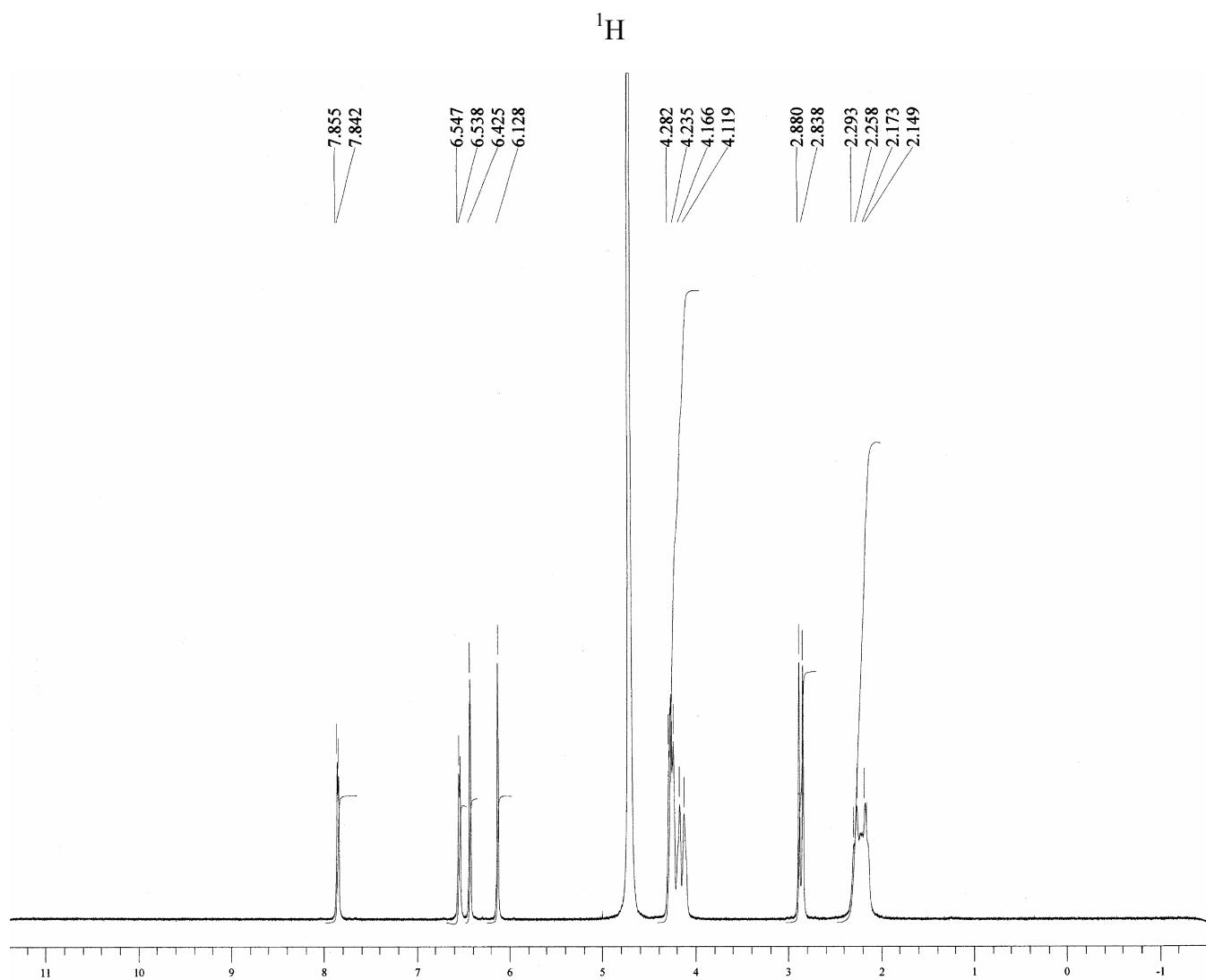
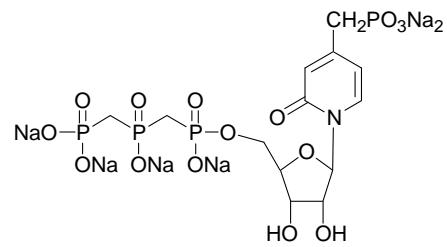


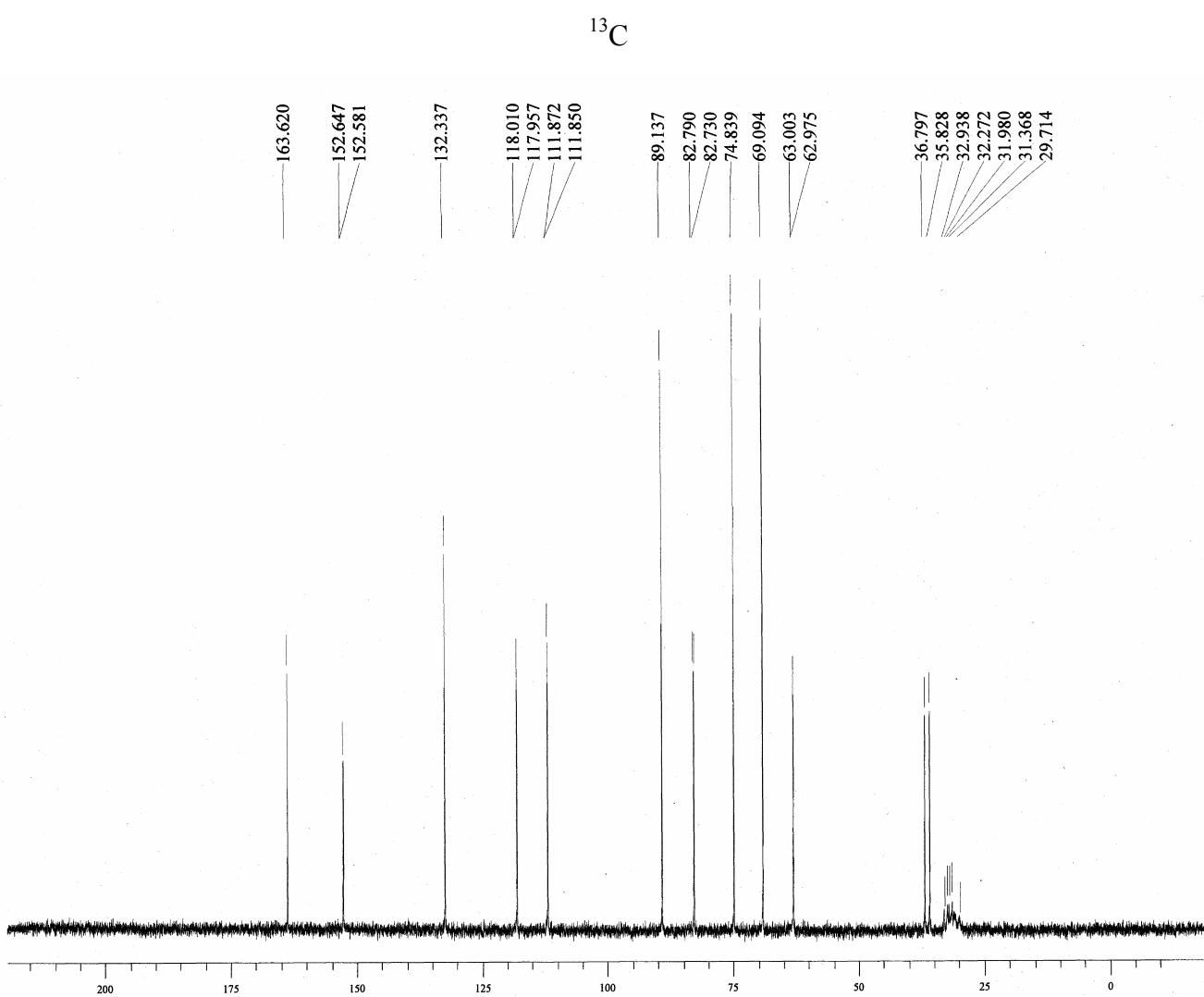
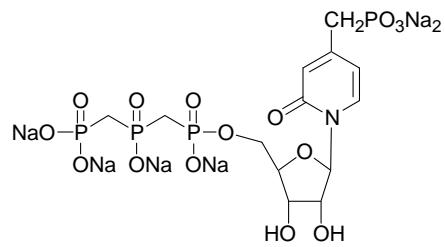
**49**

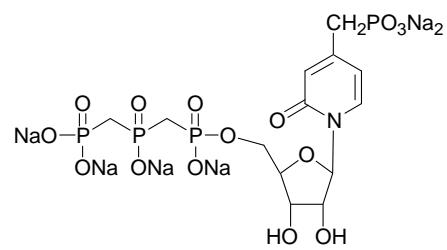
Analytical HPLC



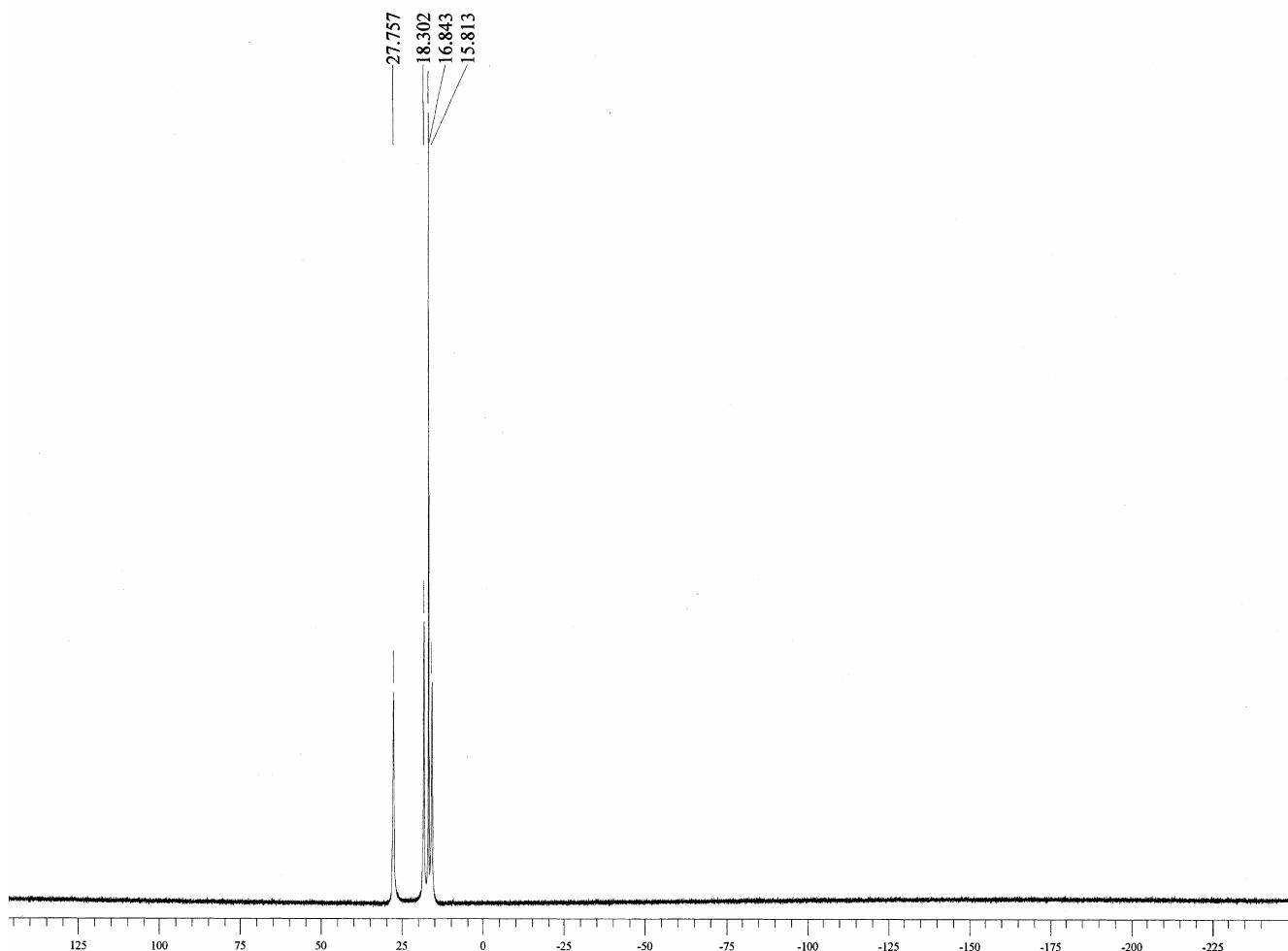
The analytical HPLC chromatogram of **49**. The following solvents were used: solvent A: 100 mM TEAA, pH 7.0, solvent B: CH<sub>3</sub>CN. The following elution profile was used: linear gradient of 99% A-1% B to 93% A-7% B over 50 minutes. Flow rate 1 mL/min,  $\lambda_{\text{detector}} = 260 \text{ nM}$ .

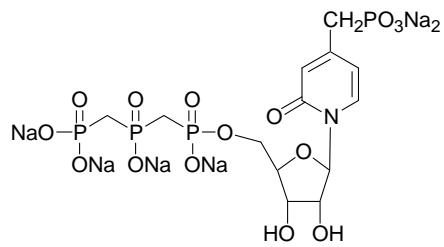






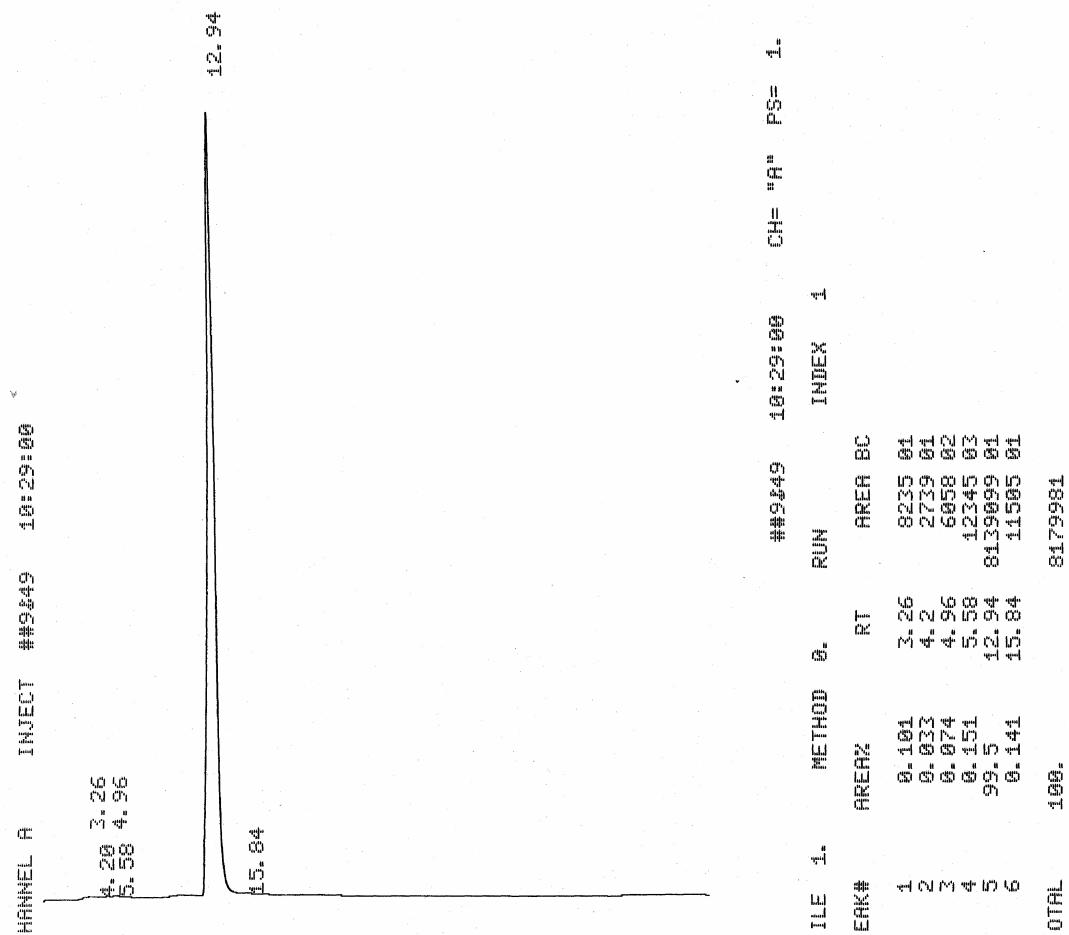
$^{31}P$



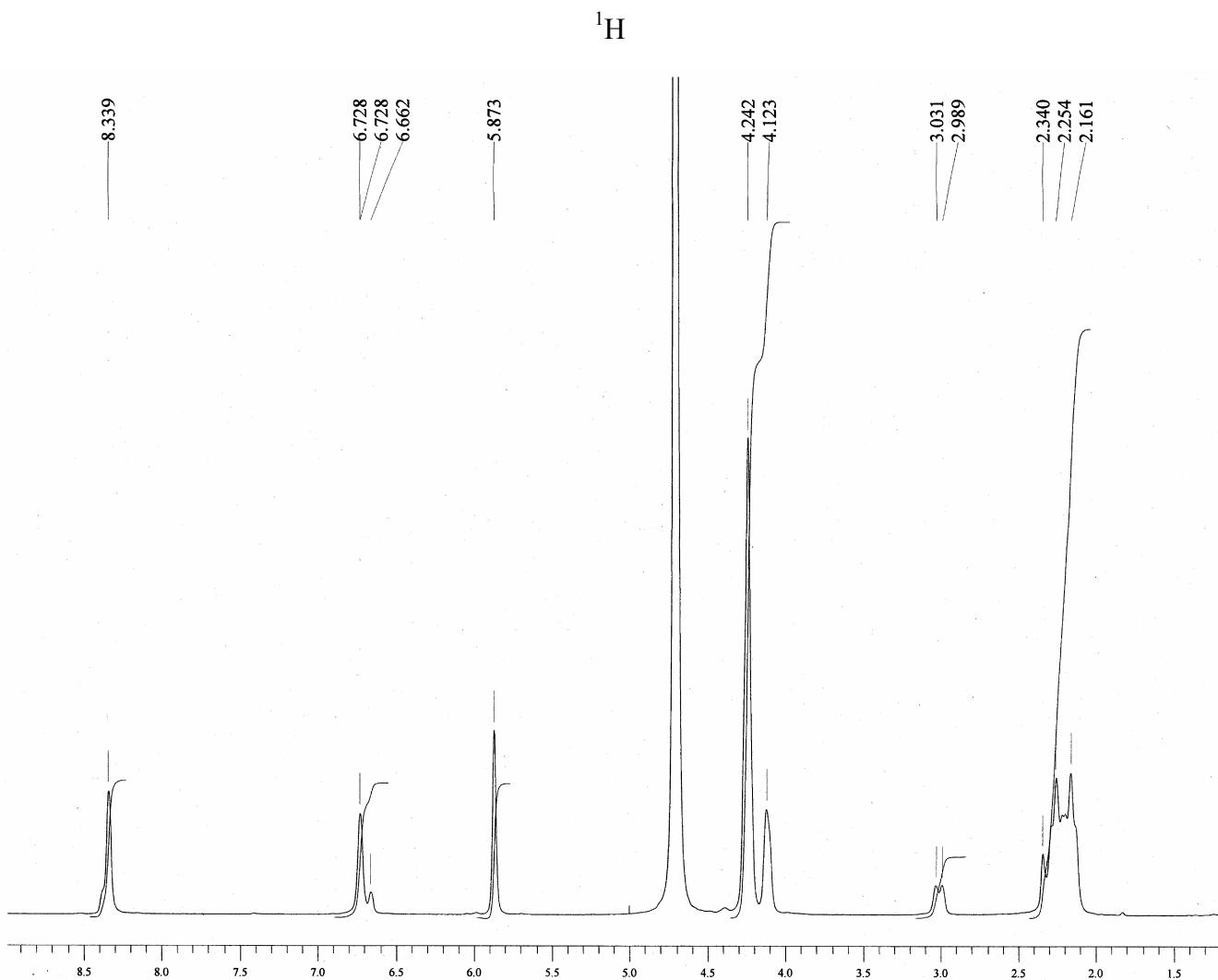
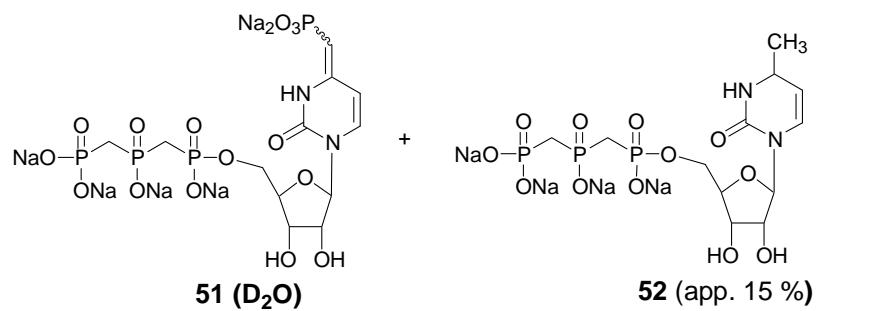


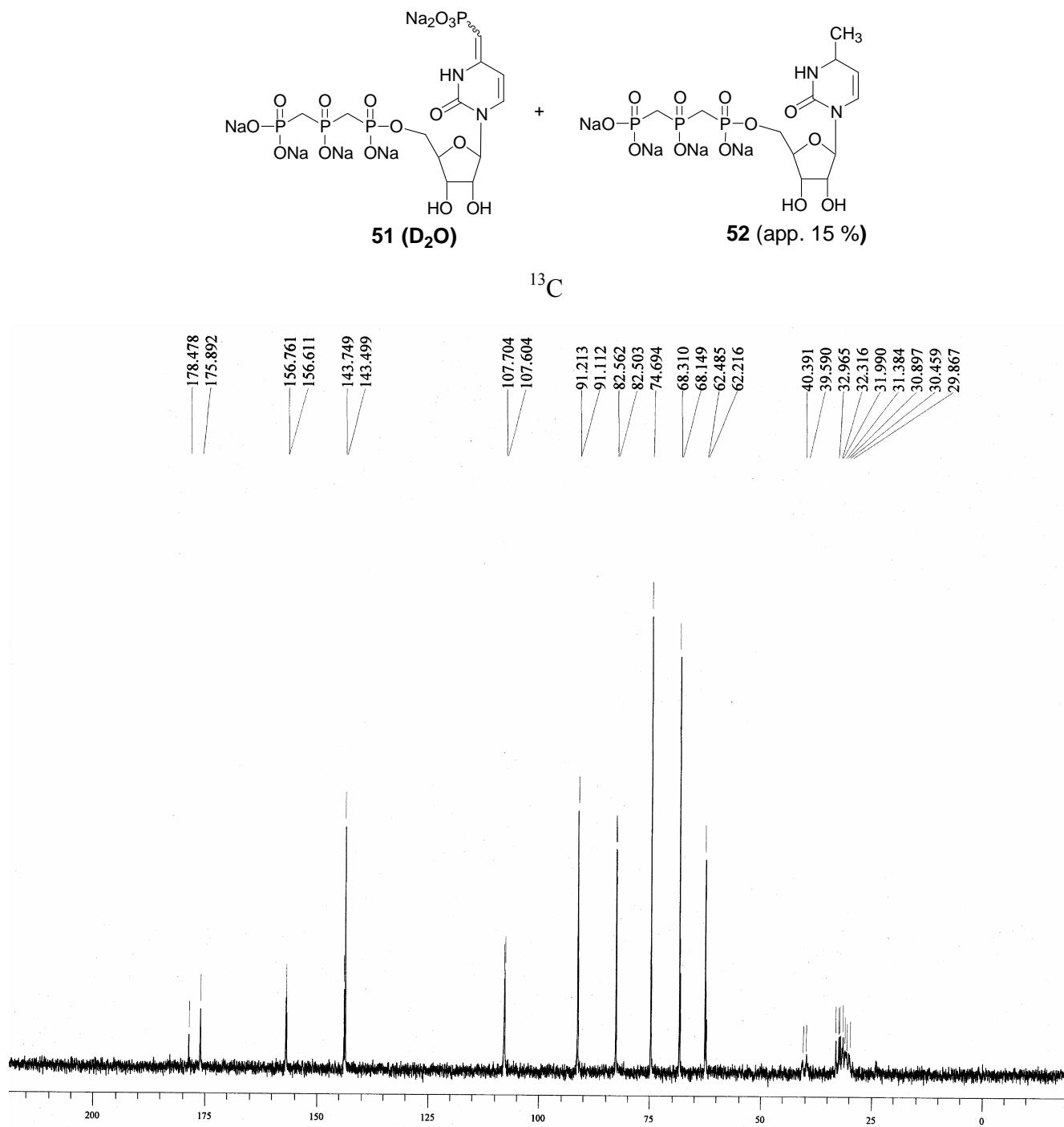
**50**

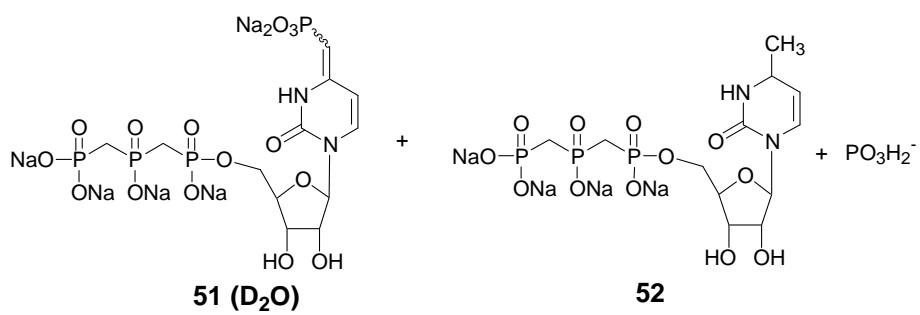
Analytical HPLC



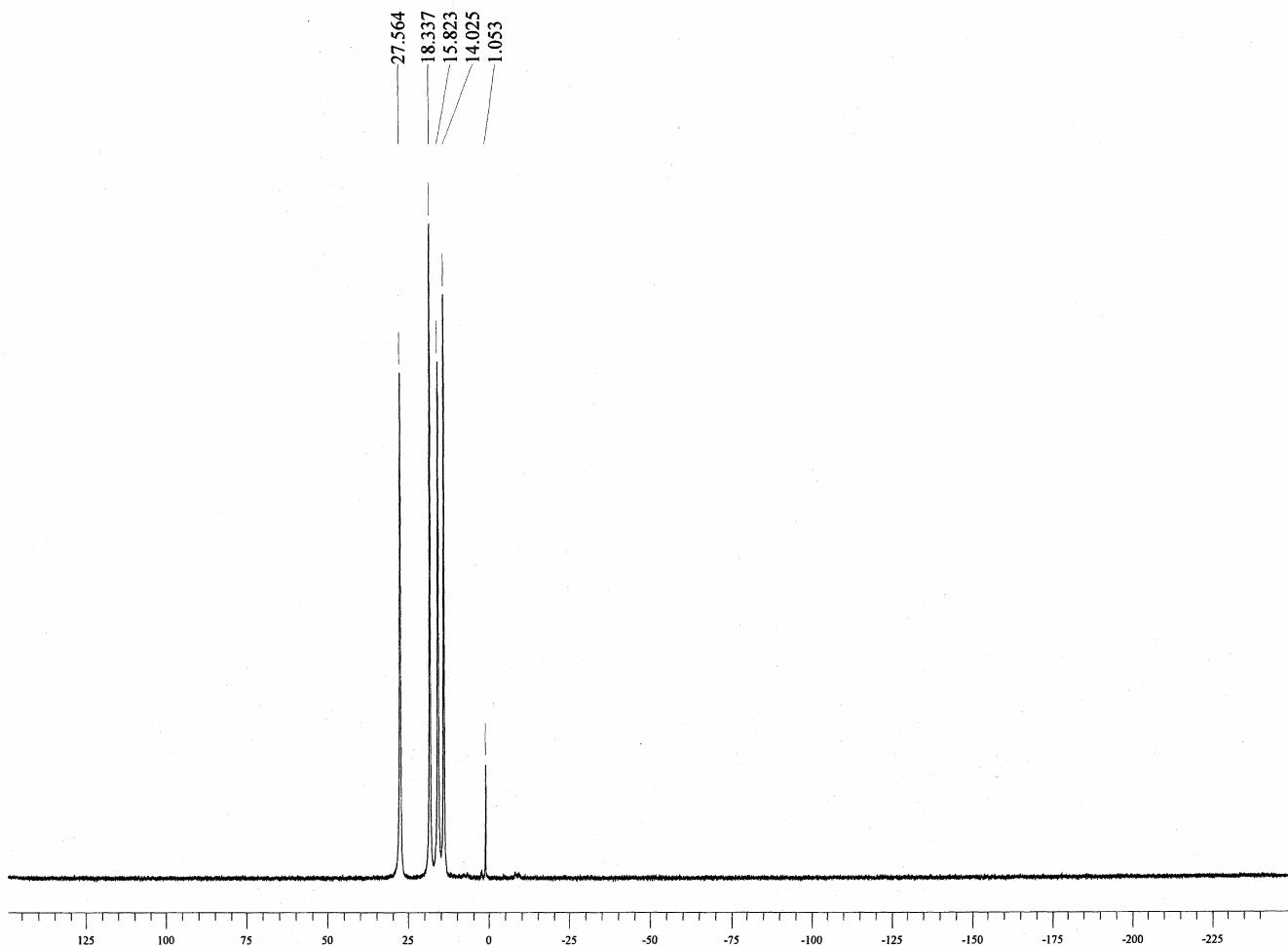
Analytical HPLC chromatogram of compound **50**. The following solvent system were used: solvent A: 100 mM TEAA, pH 7.0, solvent B: CH<sub>3</sub>CN. The following elution profile was used: 0-15 min: 99% A-1% B; 15-50 min: linear gradient of 99% A-1% B to 94% A-6% B. Flow rate = 1 mL/min.  $\lambda_{\text{detector}} = 260 \text{ nM}$ .

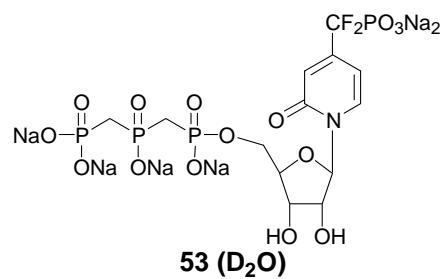




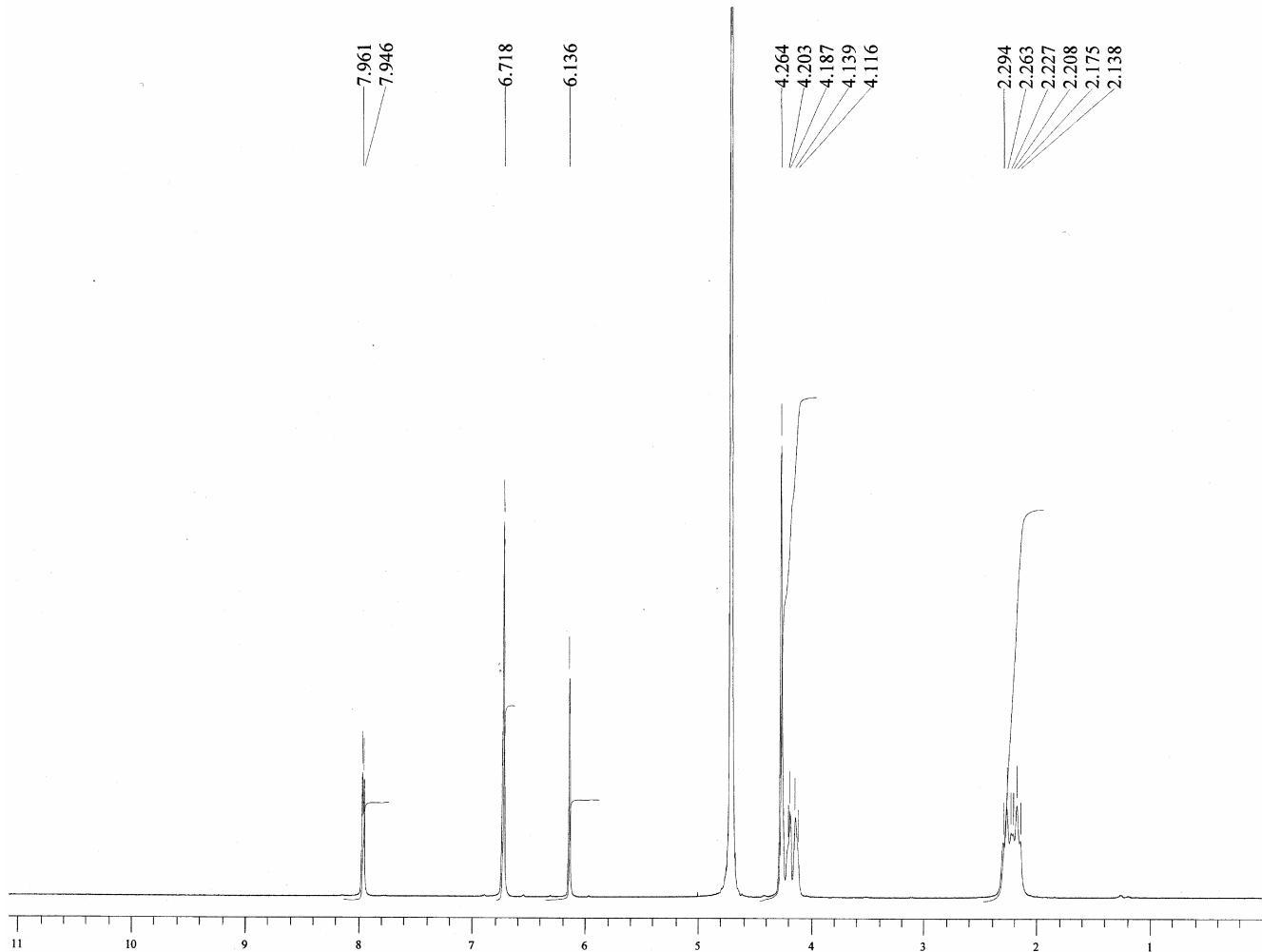


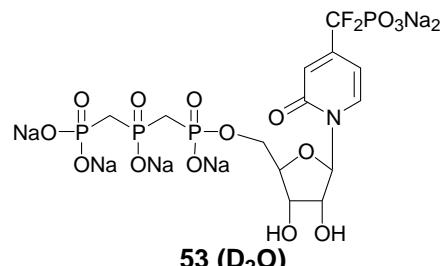
$^{31}P$



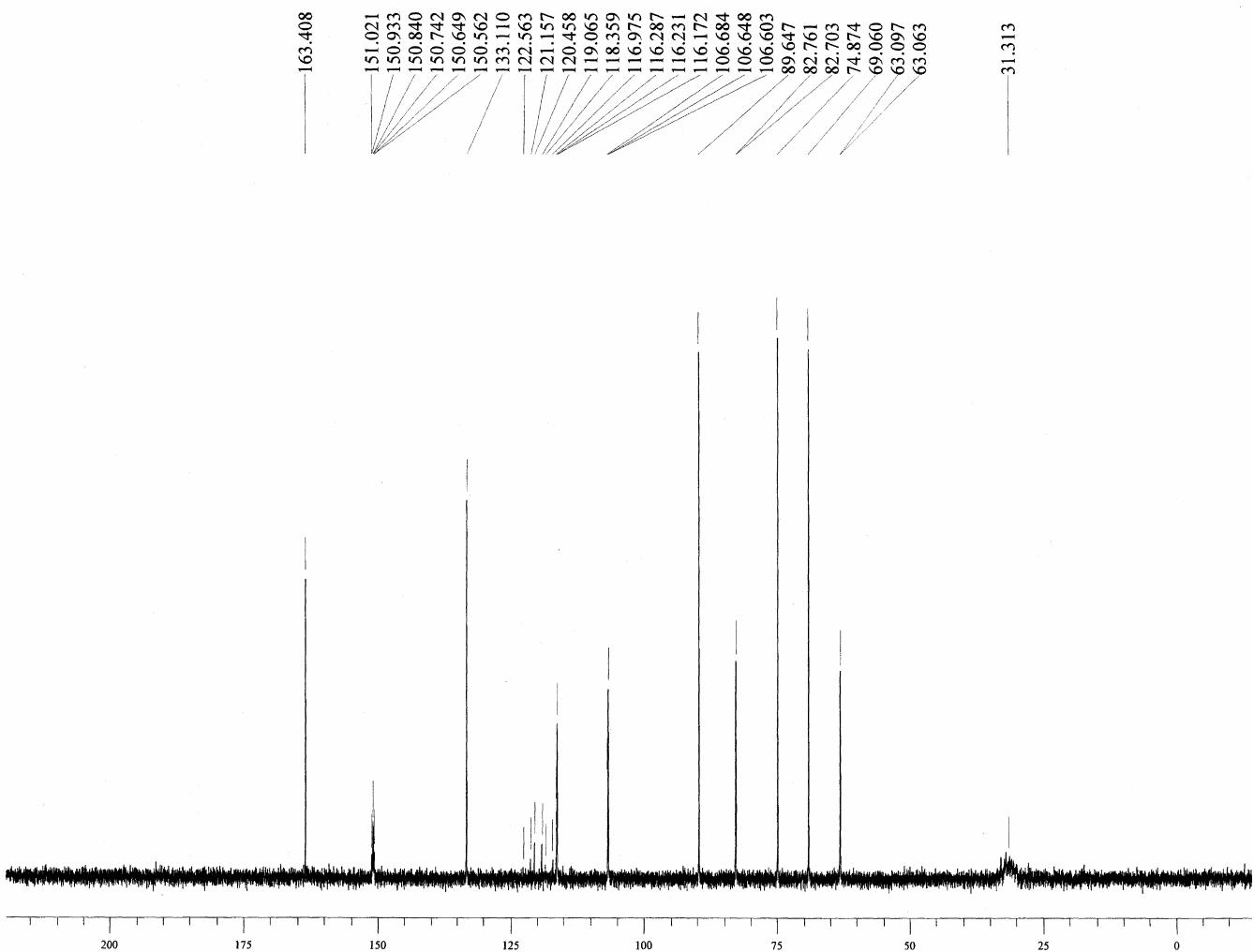


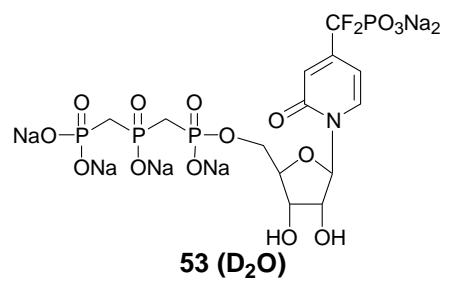
<sup>1</sup>H



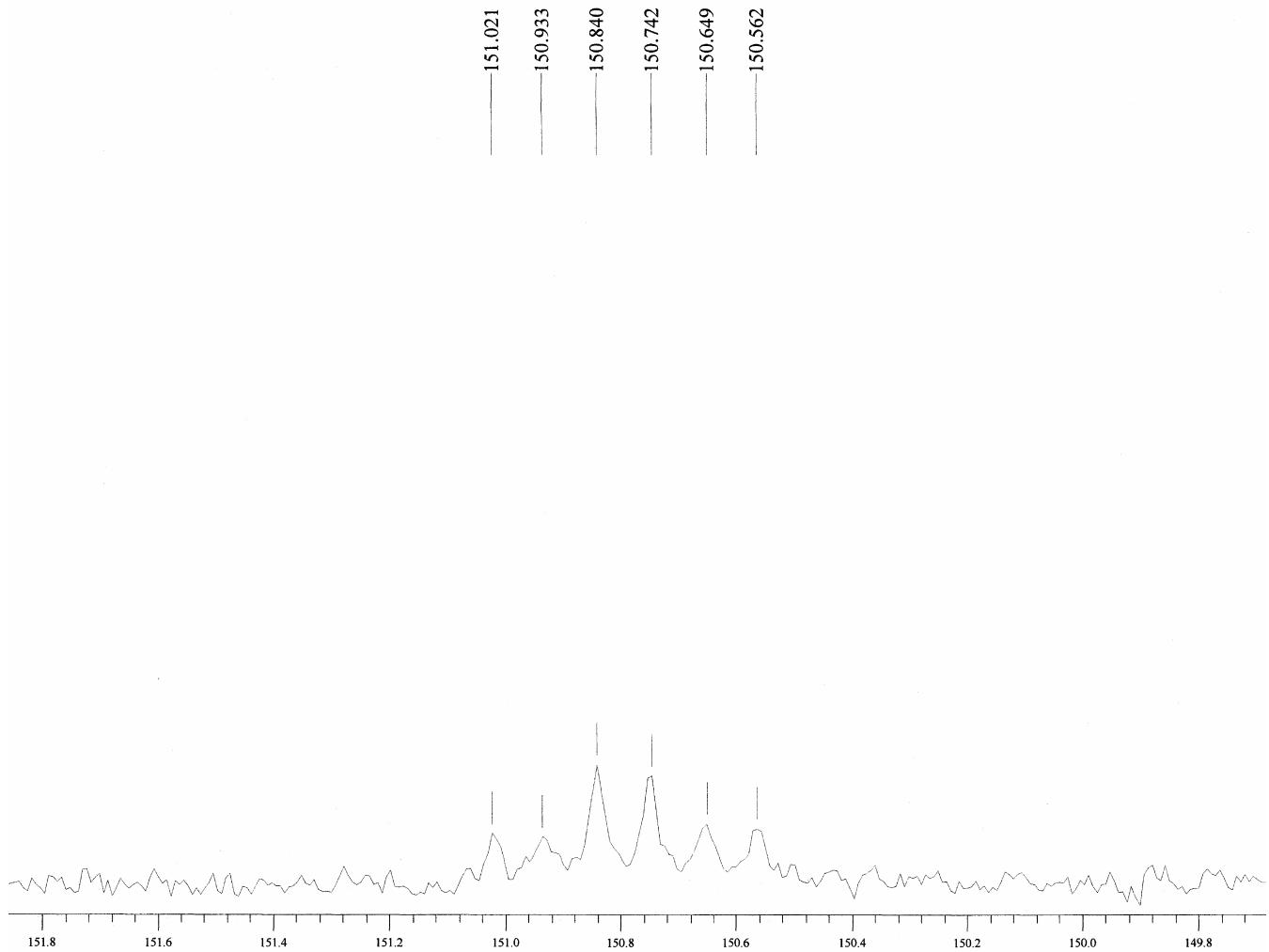


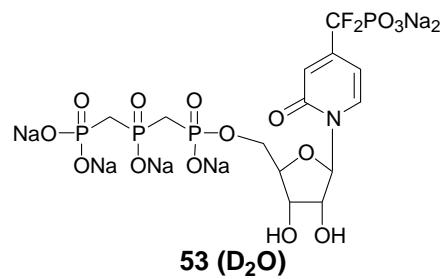
<sup>13</sup>C



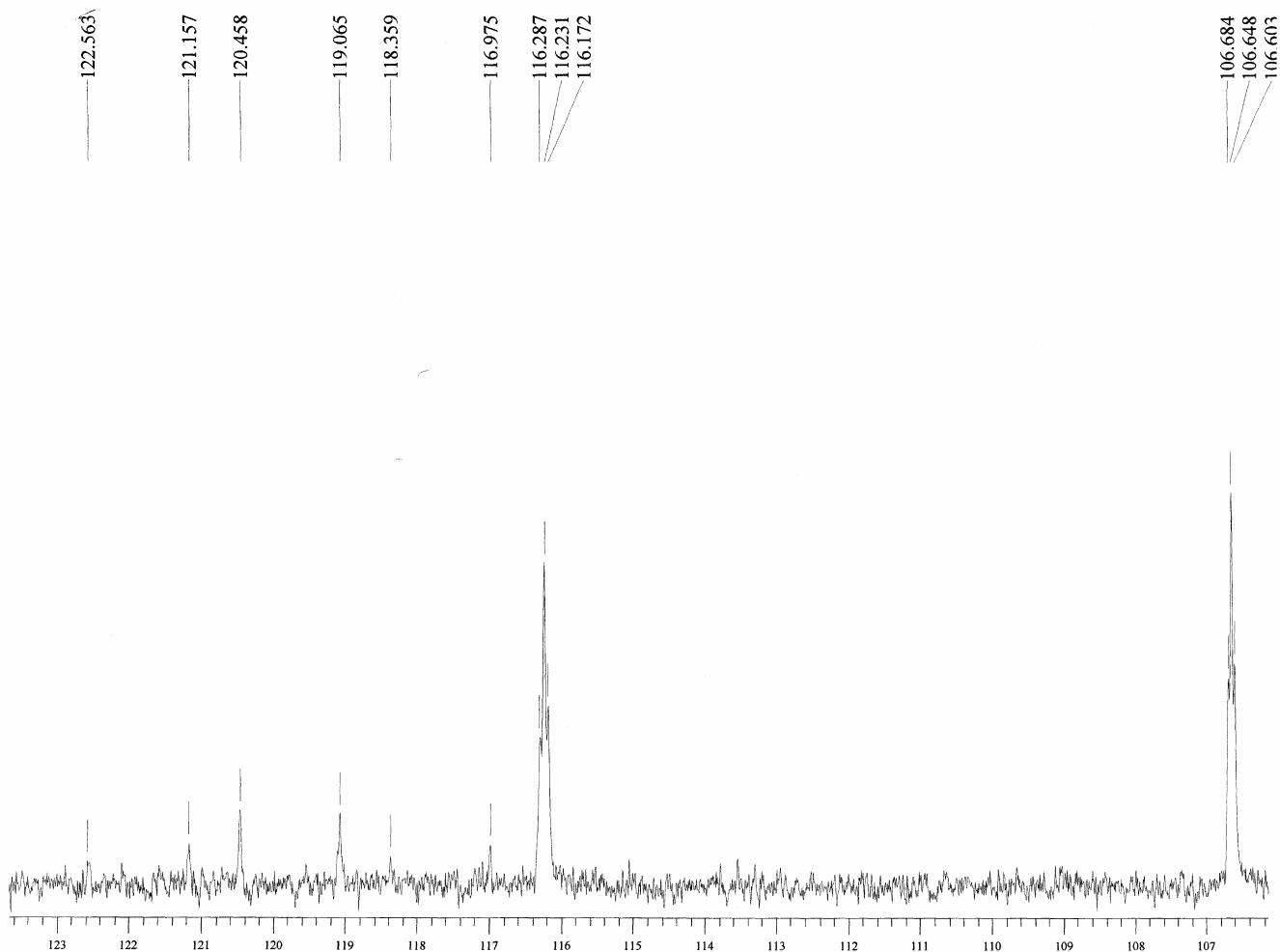


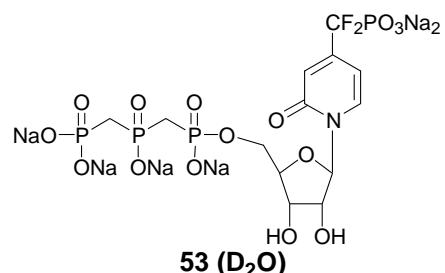
$^{13}C$  (expanded)



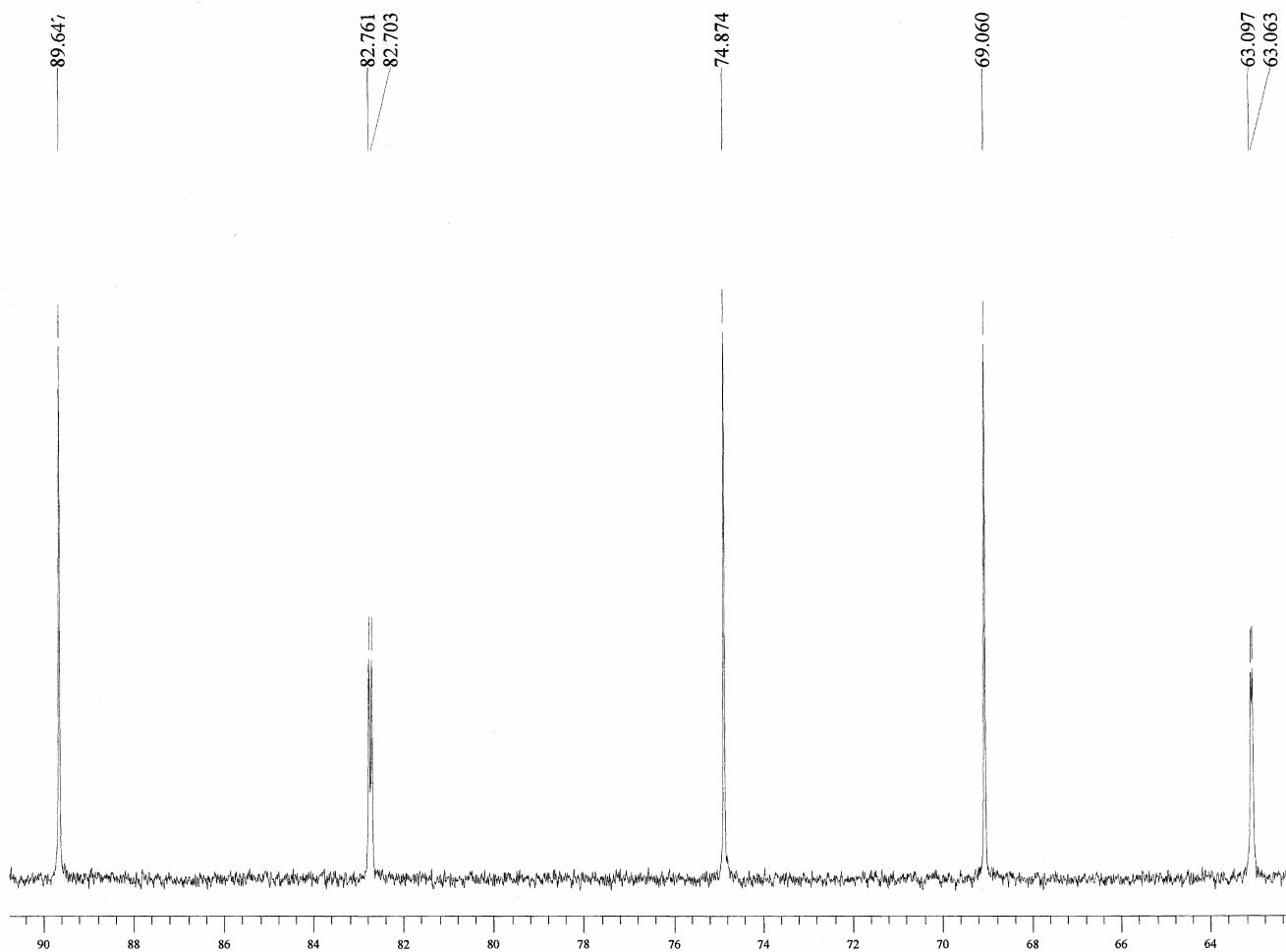


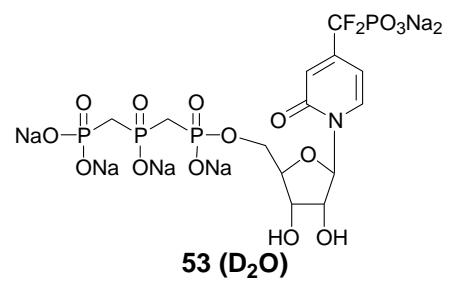
$^{13}C$  (expanded)





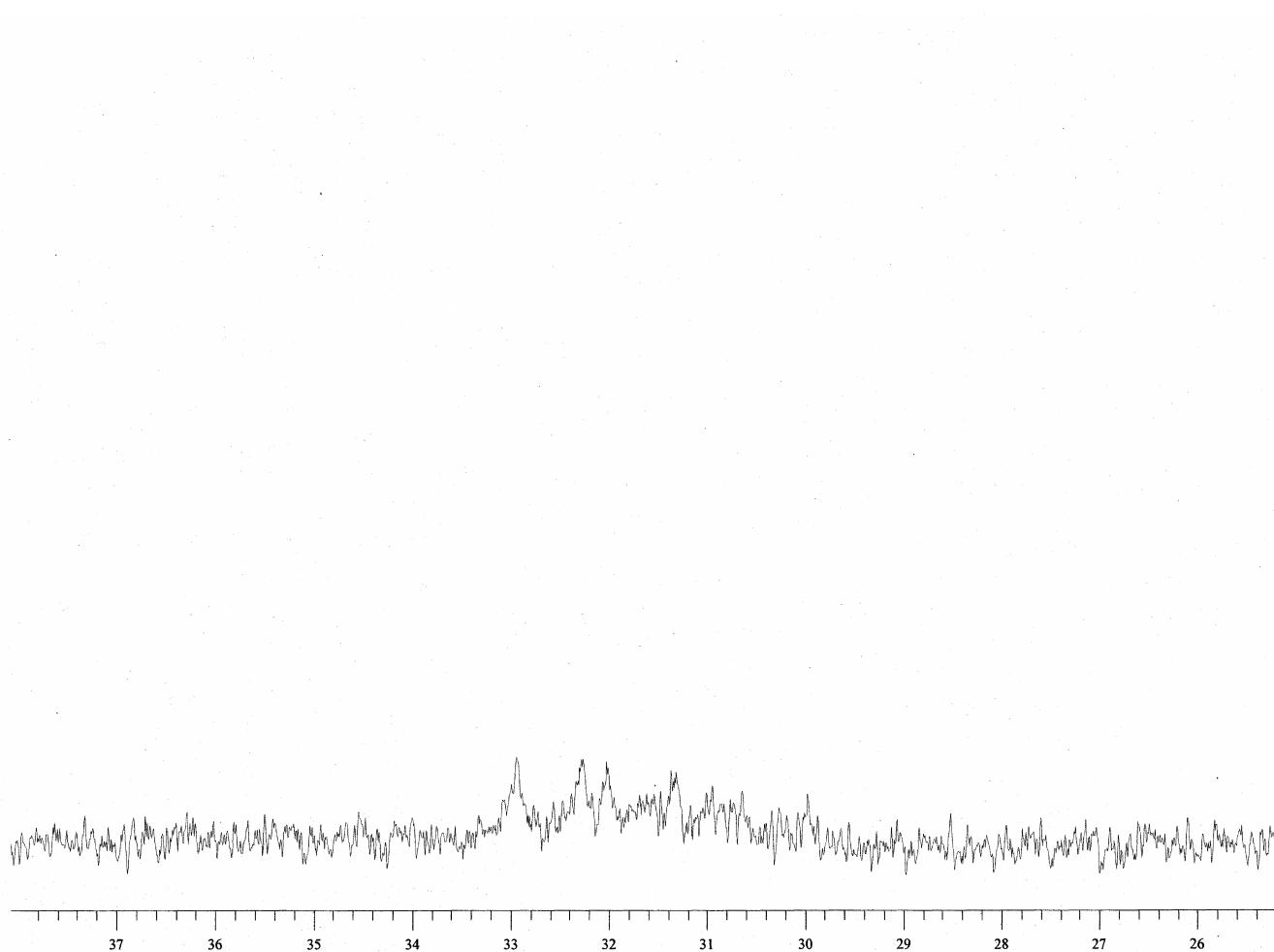
$^{13}C$  (expanded)

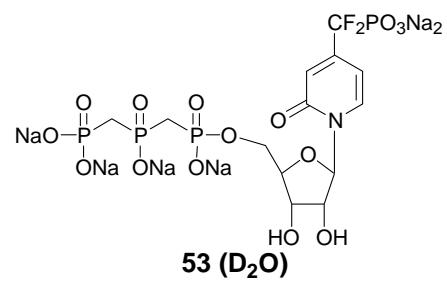




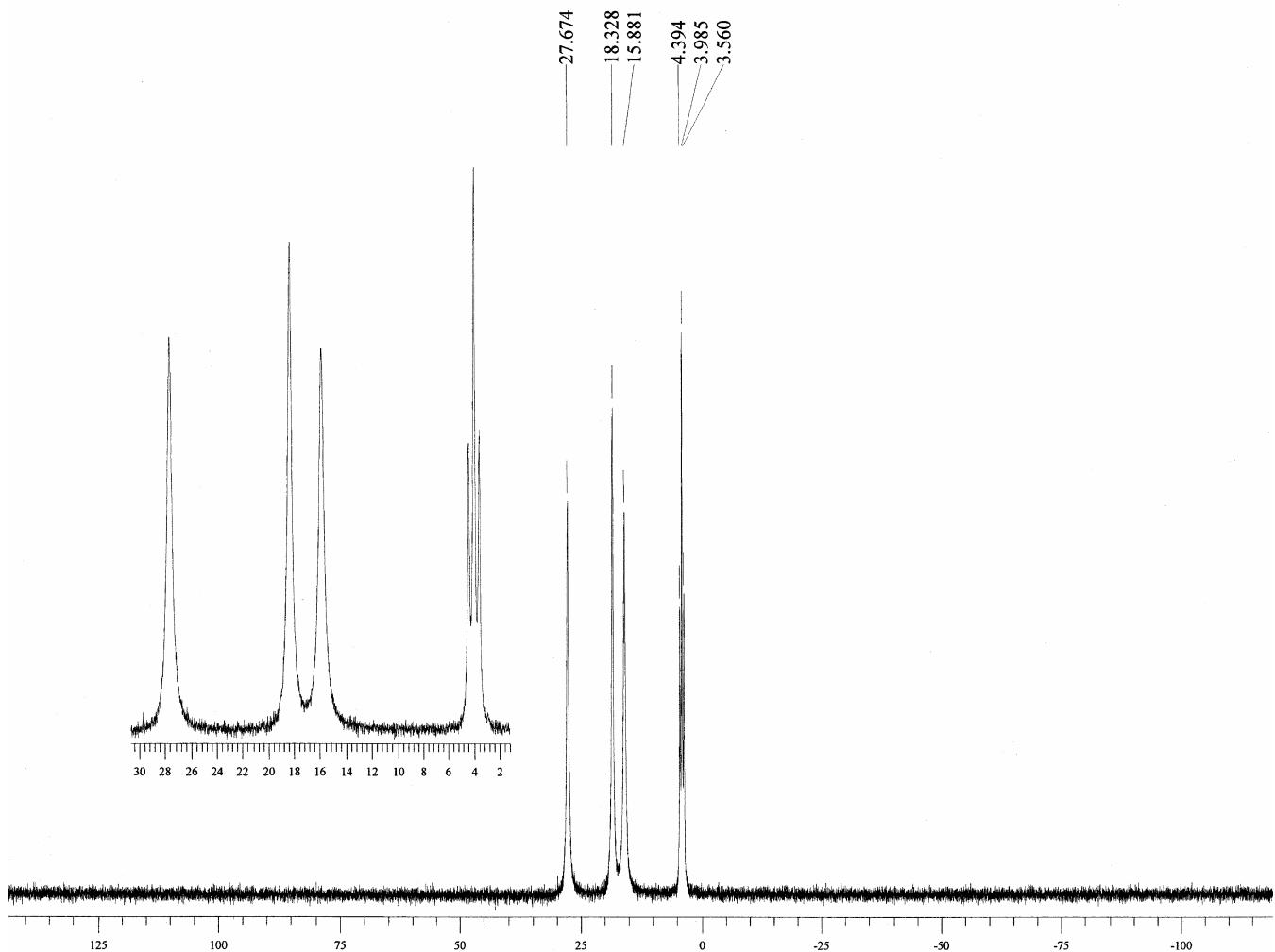
53 ( $\text{D}_2\text{O}$ )

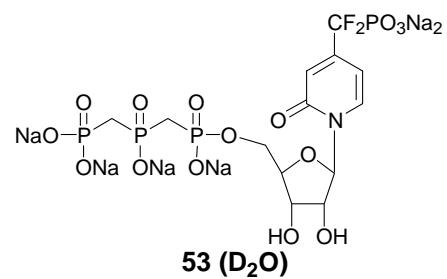
$^{13}\text{C}$  (expanded)



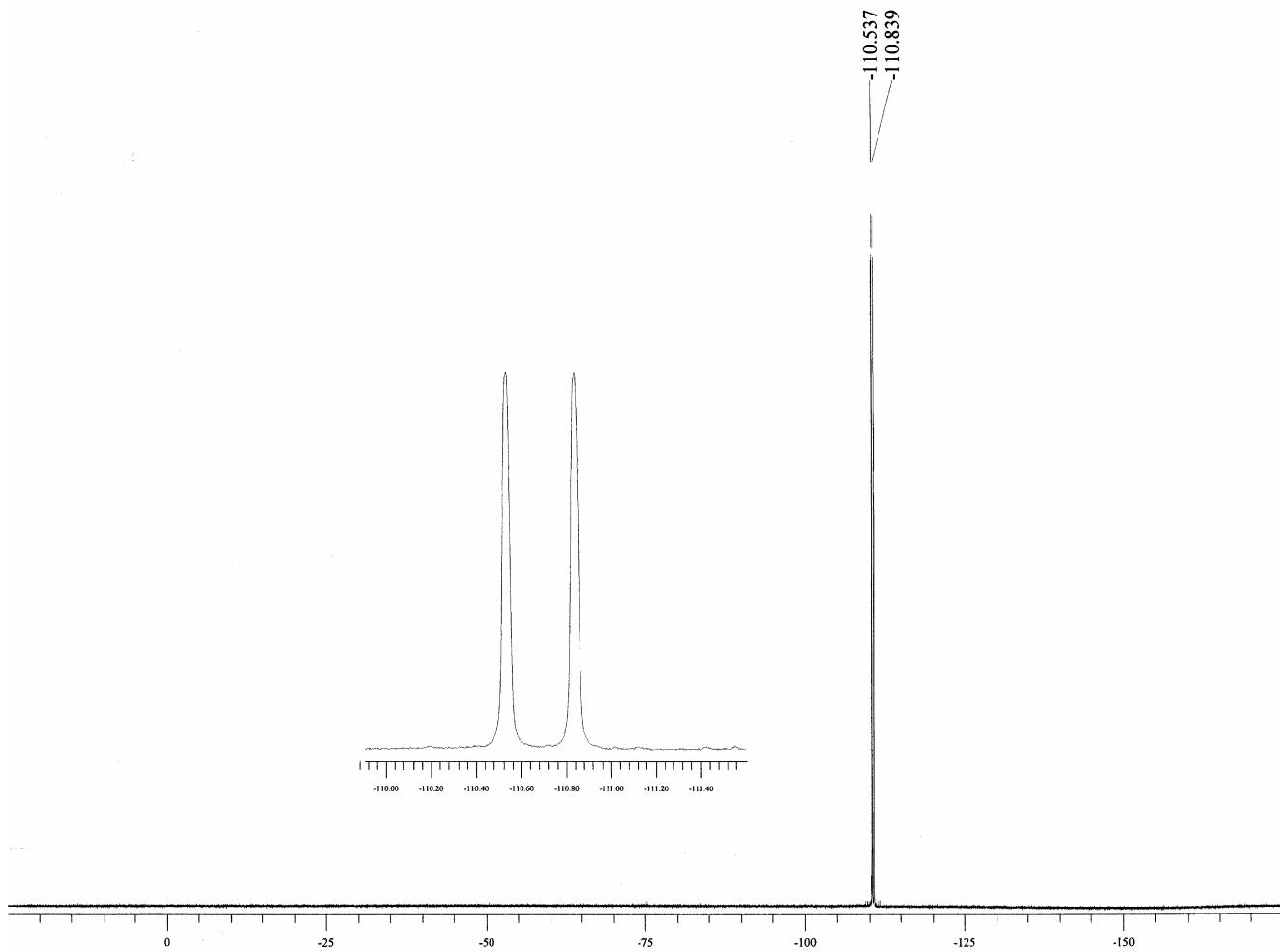


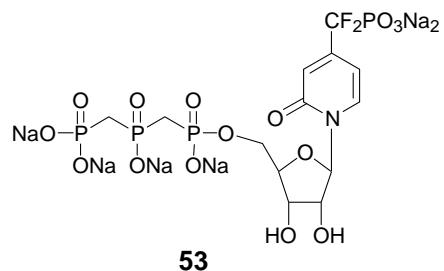
$^{31}P$



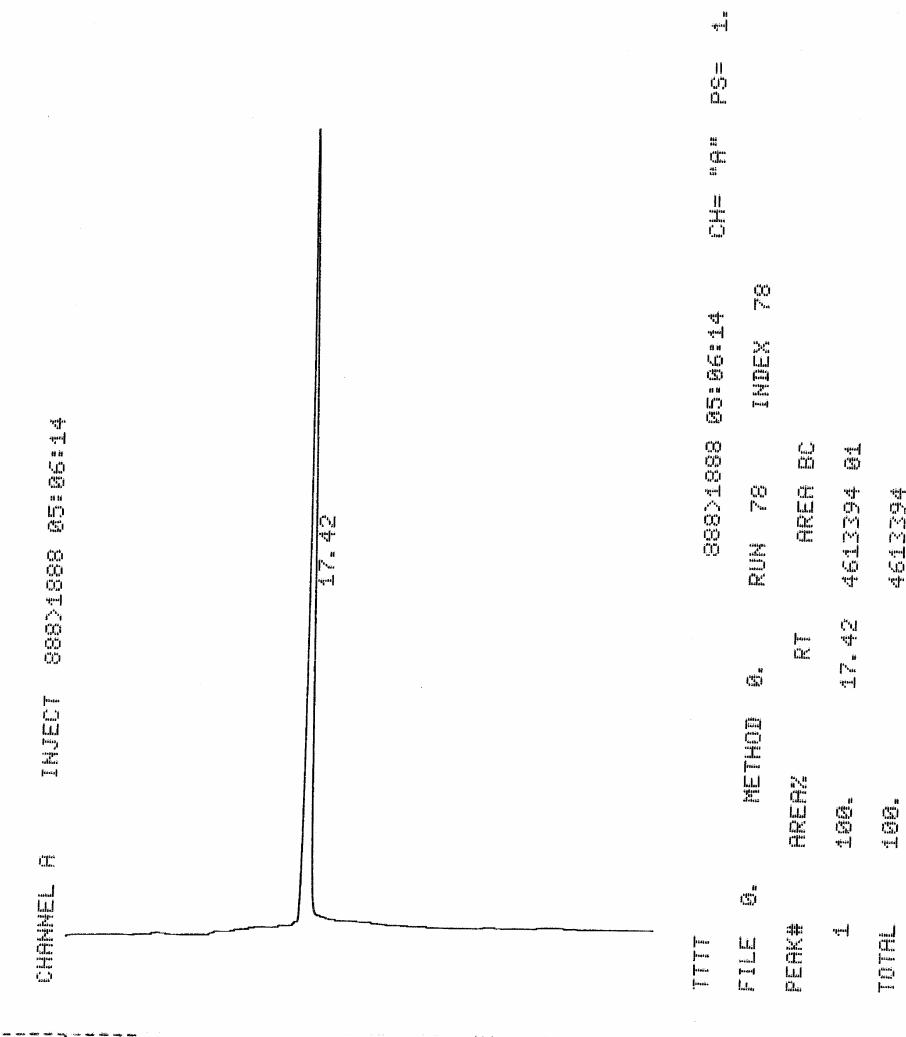


$^{19}F$

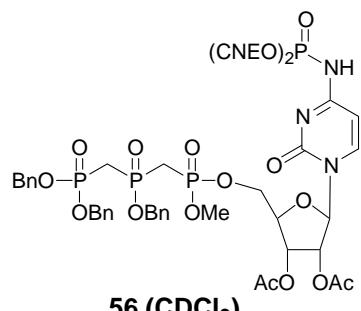




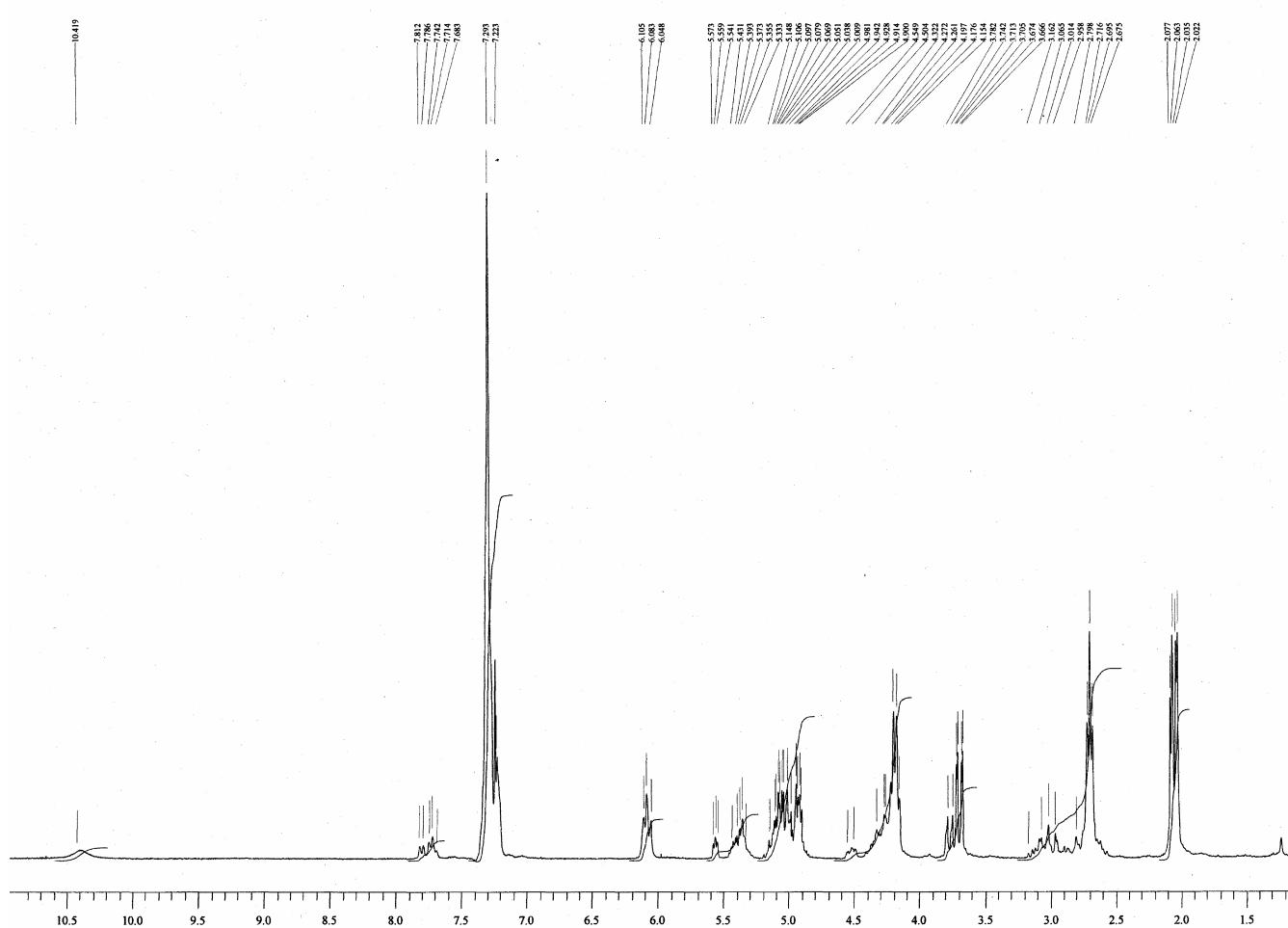
### Analytical HPLC

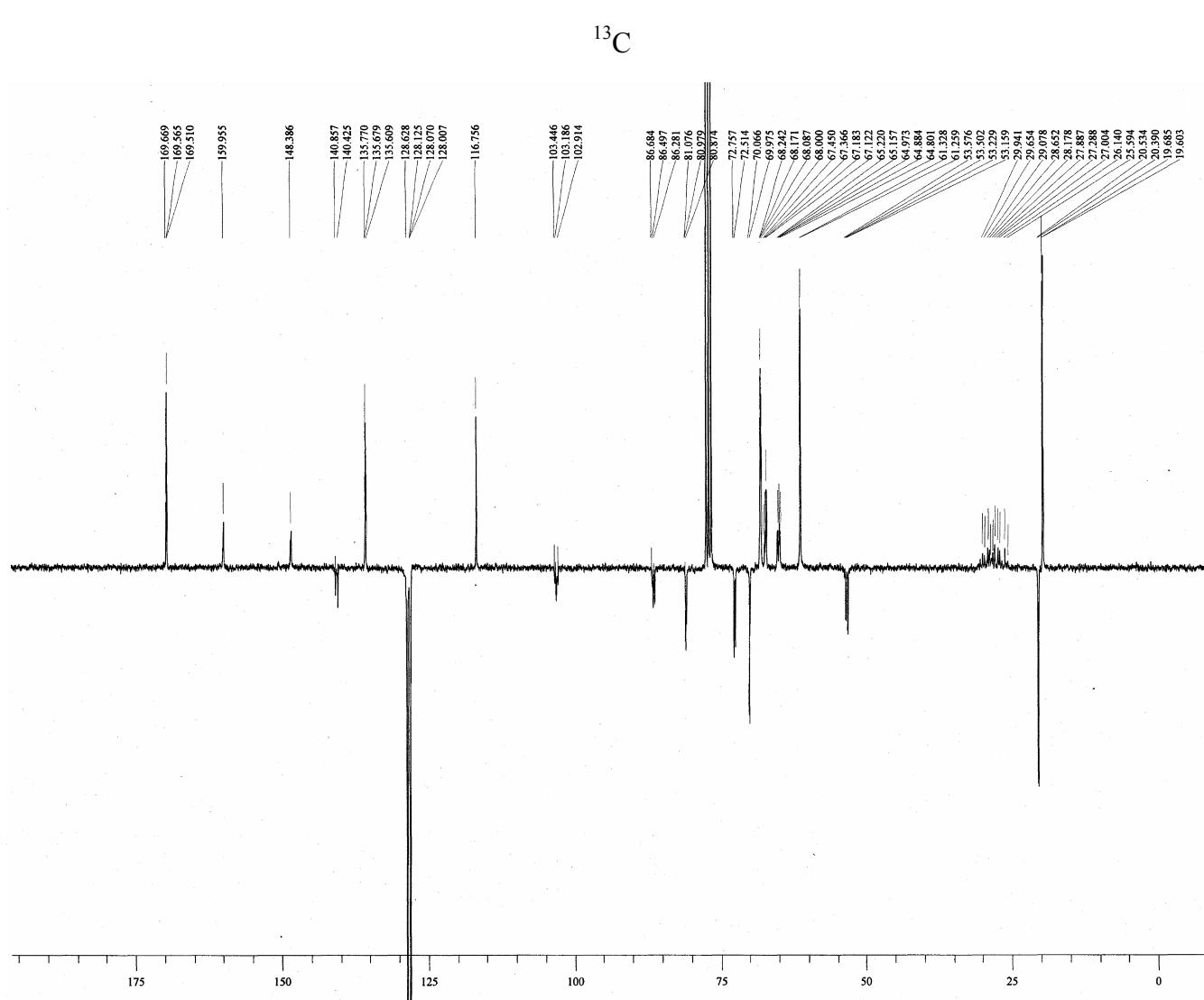
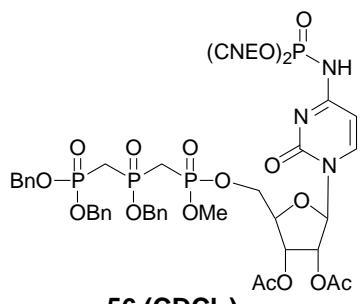


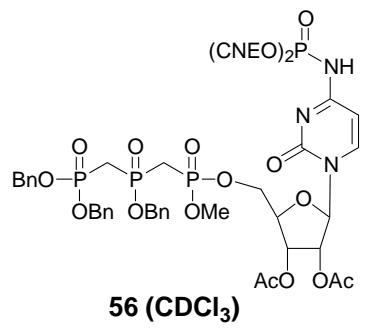
Analytical HPLC chromatogram of compound **53**. The following solvents were used: solvent A: 100 mM TEAA, pH 7.0, solvent B: CH<sub>3</sub>CN. The following elution profile was used: linear gradient of 99% A-1% B to 93% A-7% B over 40 minutes. Flow rate = 1 mL/min.  $\lambda_{\text{detector}} = 260 \text{ nM}$ .



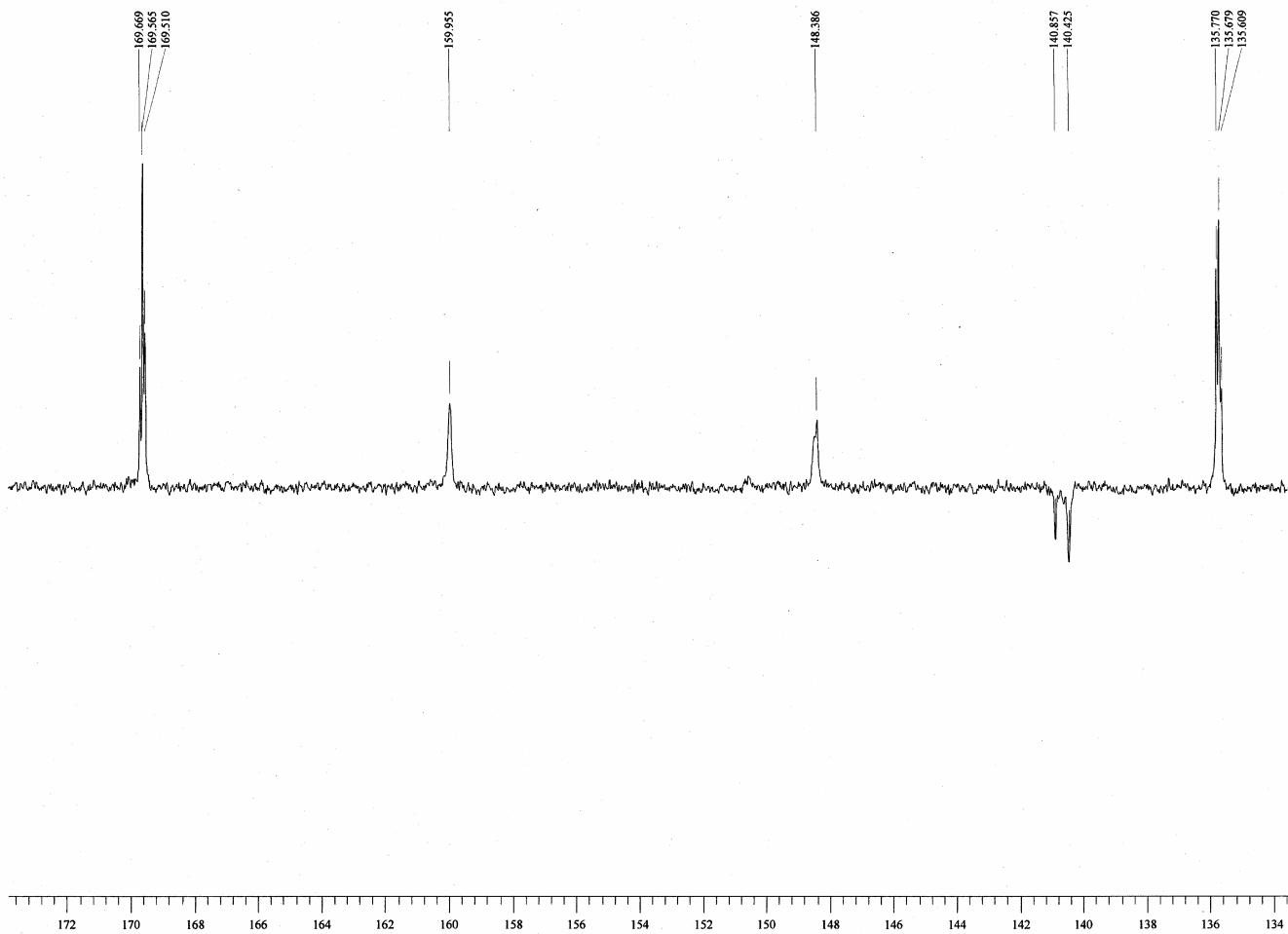
<sup>1</sup>H

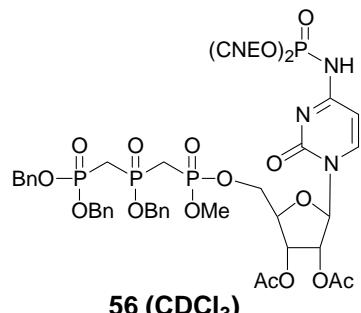




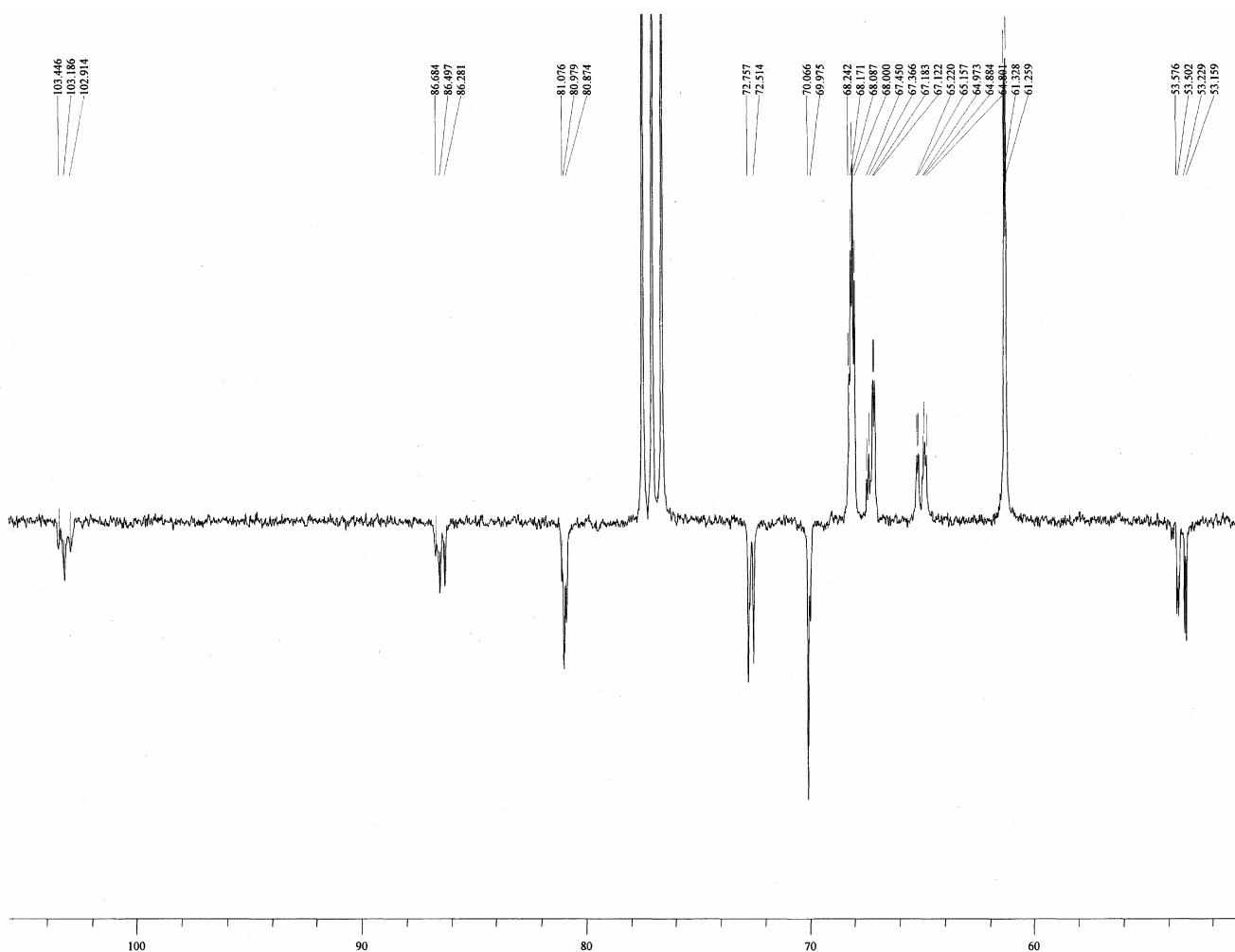


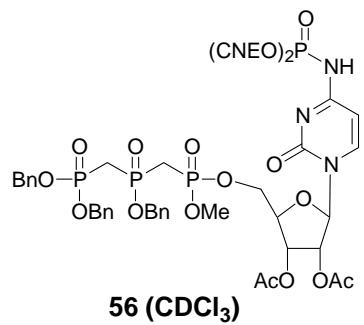
<sup>13</sup>C (expanded)



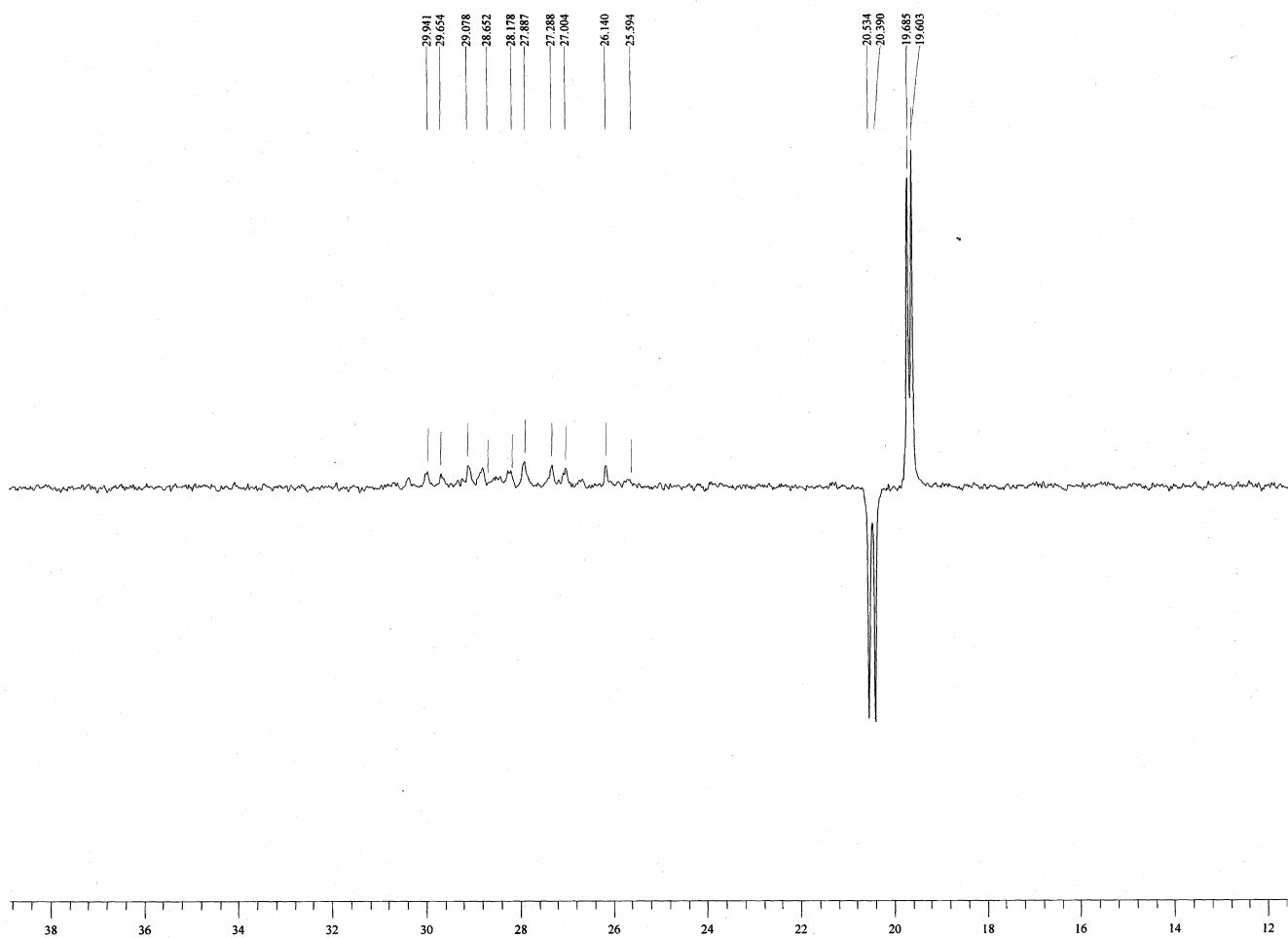


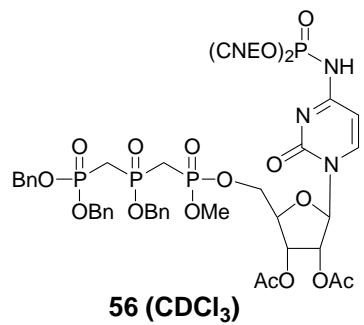
<sup>13</sup>C (expanded)



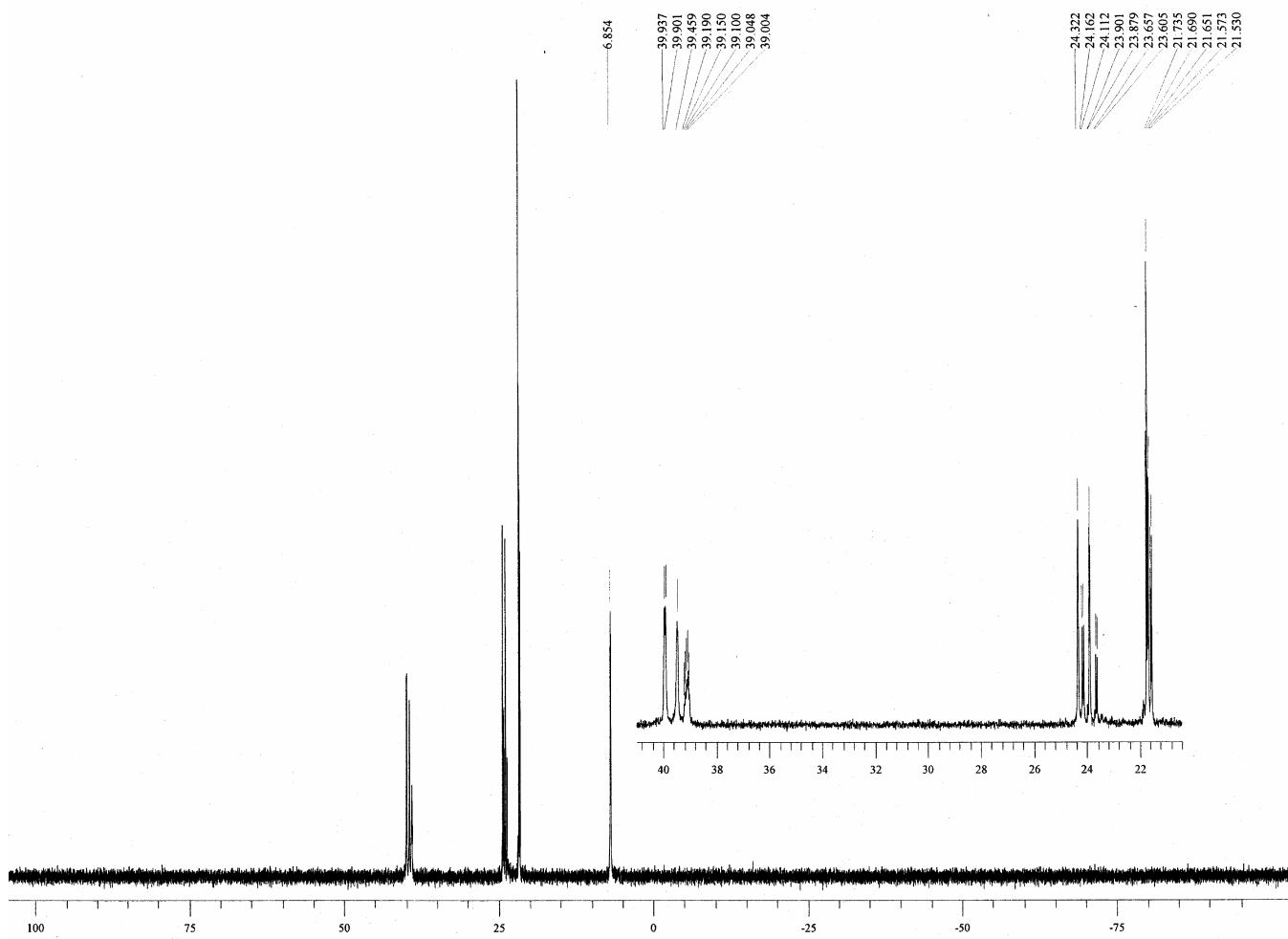


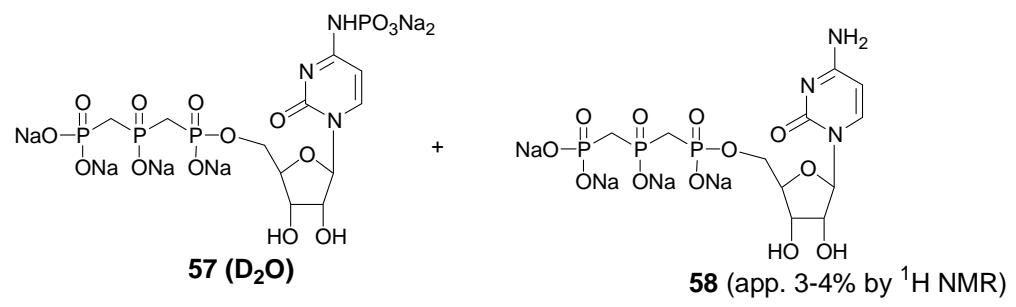
<sup>13</sup>C (expanded)



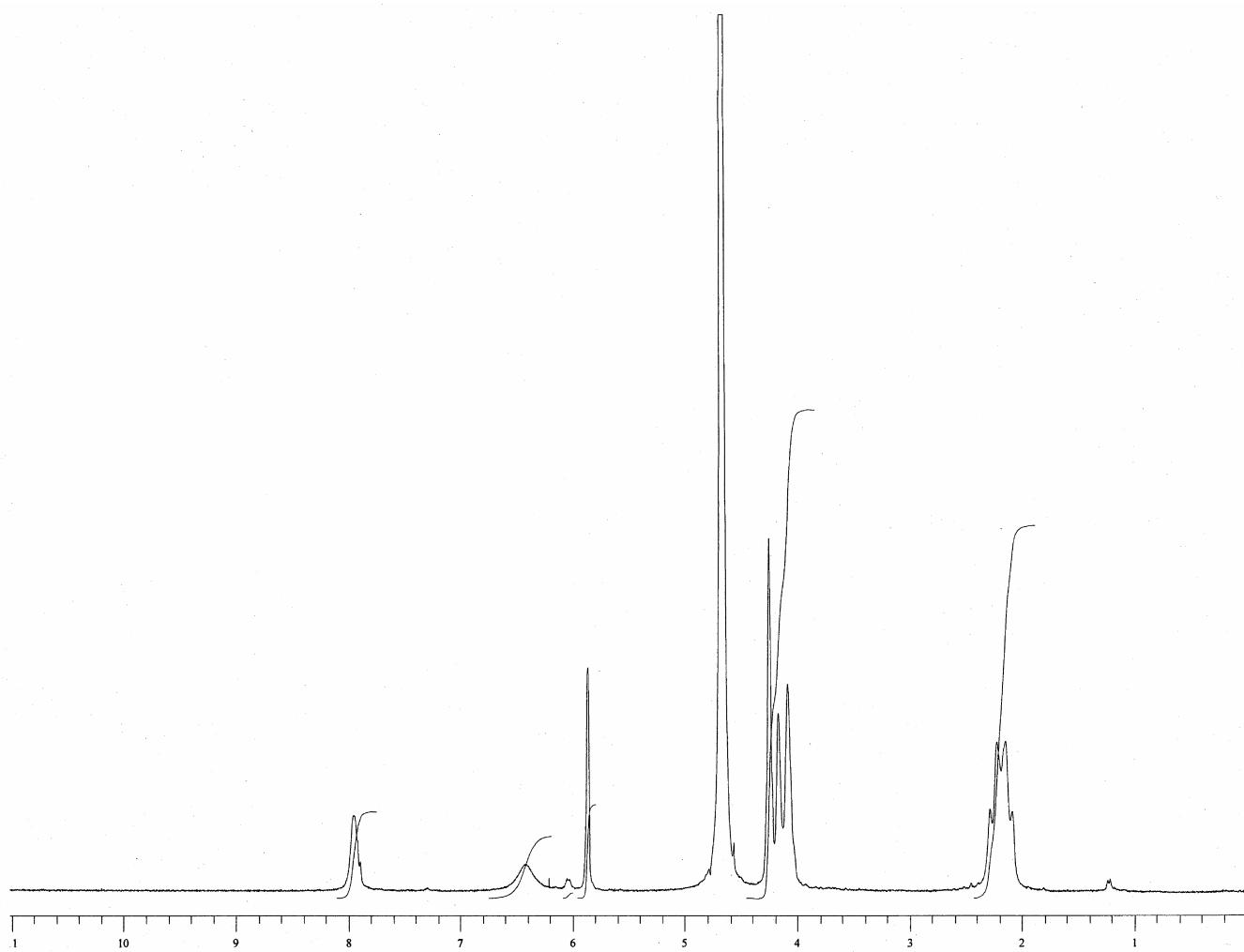


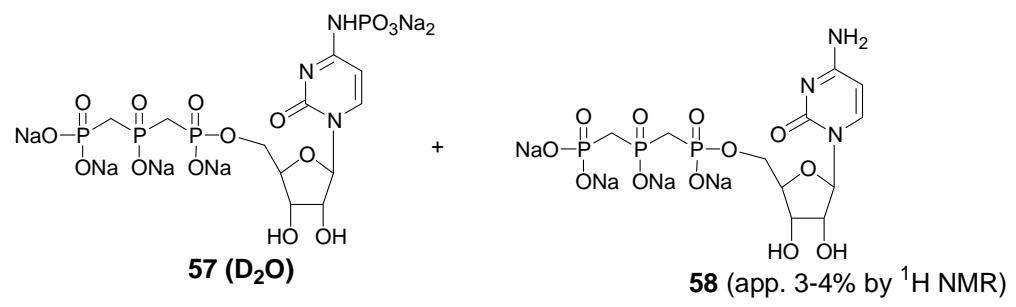
<sup>31</sup>P



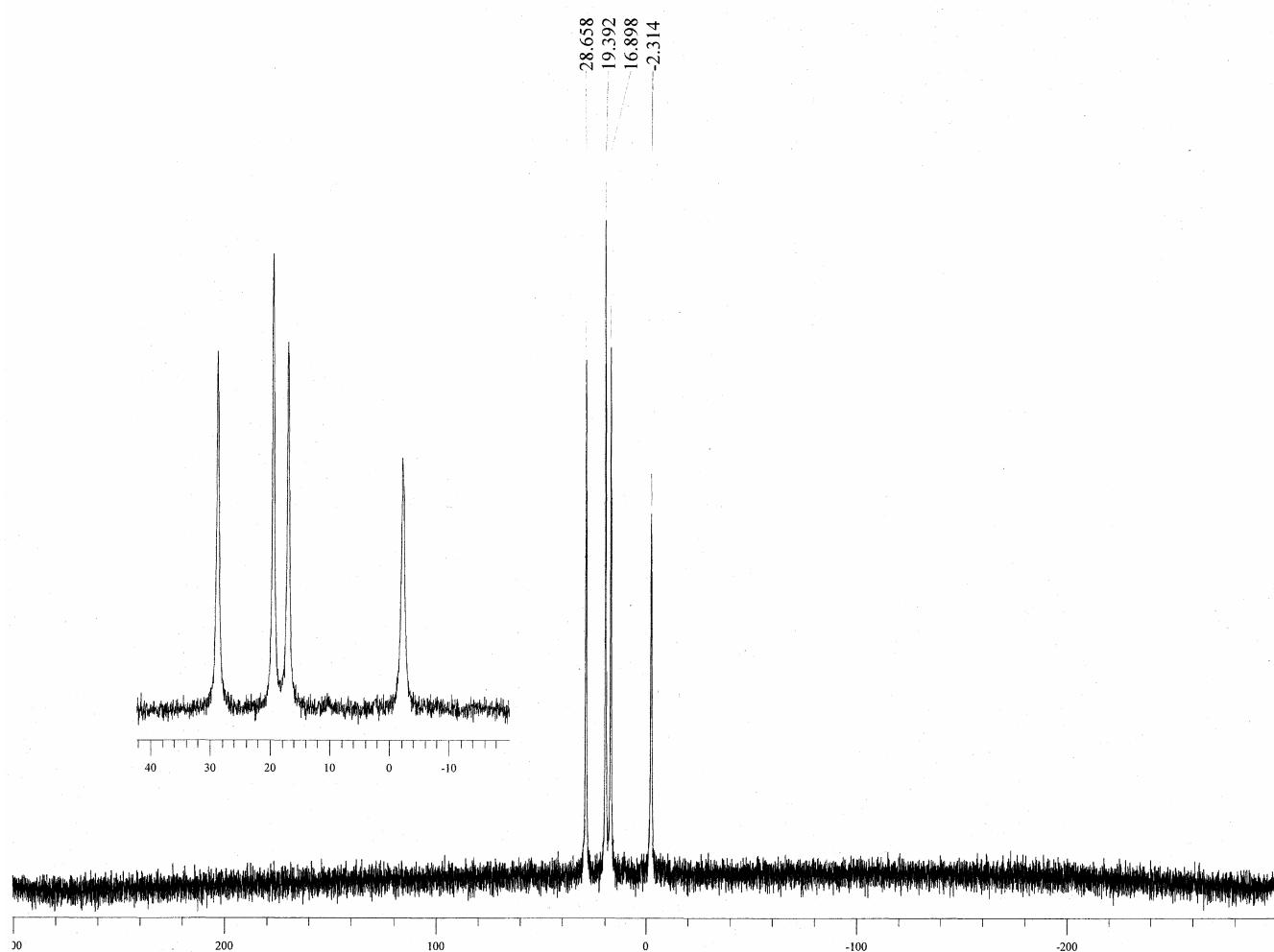


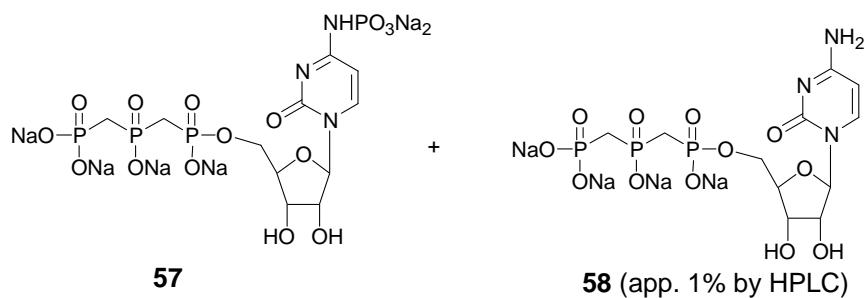
$^1H$



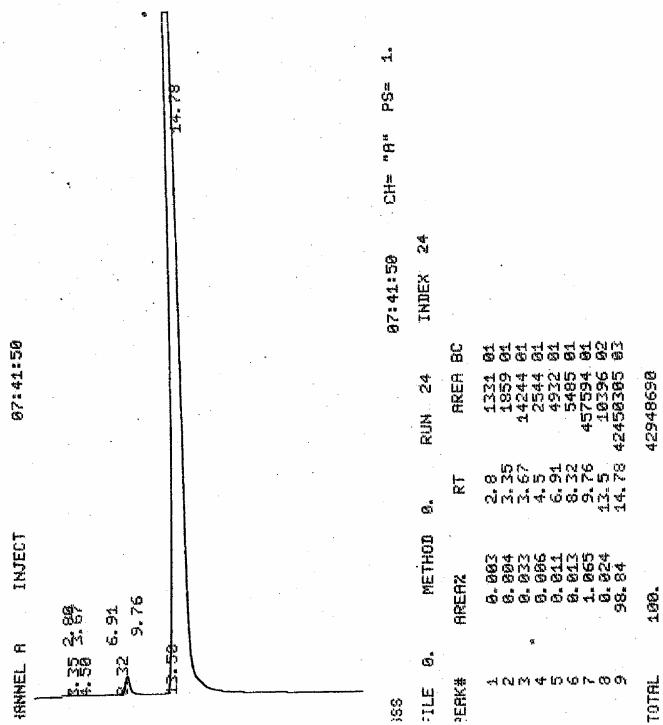


$^{31}P$





Analytical HPLC



Analytical HPLC chromatogram of compound **57**. The following elution profile was used: 100 mM TEAA, pH 9.0 for 30 min. Flow rate = 1 mL/min.  $\lambda_{\text{detector}} = 260 \text{ nM}$ .