Differential reactivity of purified bioactive coffee furans, cafestol and kahweol, with acidic nitrite: product characterization and factors controlling nitrosation versus ring opening pathways.

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### NMR spectra

Spectra were acquired using 256 equally spaced evolution-time period  $t_1$  values averaging 16 transients of 2048 points, with 6024 Hz of spectral width. Time-domain data matrices were all zero-filled to 4K in both dimensions, thus yielding a digital resolution of 2.94 Hz/pt. Prior to Fourier transformation, a Lorentz-to-Gauss window with different parameters was applied for both  $t_1$  and  $t_2$  dimensions for all the experiments. ROESY spectra were obtained with 200-ms mixing time, and the recycle time was set to 5 times the longest T1. TOCSY experiments were recorded with spinlock periods of 64 ms, achieved with the MLEV-17 pulse sequence. Linear prediction was applied to extend the data to twice their length in  $t_1$ . The <sup>1</sup>H and <sup>13</sup>C natural abundance HSQC and HMBC spectra were recorded at 300 K on the DRX-600 spectrometer, operating at 150.90 MHz for <sup>13</sup>C. One hundred twenty eight equally spaced evolution time period t<sub>1</sub> values were acquired, averaging 48 transients of 2048 points and using GARP4 for decoupling. The final data matrix was zero-filled to 4096 in both dimensions, and apodized before Fourier transformation by a shifted cosine window function in t<sub>2</sub> and in t<sub>1</sub>. Linear prediction was also applied to extend the data to twice their length in  $t_1$ .

<sup>1</sup>H NMR spectrum of kahweol in CD<sub>3</sub>OD





<sup>13</sup>C NMR spectrum of kahweol in CD<sub>3</sub>OD





COSY spectrum of kahweol in CD<sub>3</sub>OD





TOCSY spectrum of kahweol in CD<sub>3</sub>OD





ROESY spectrum of kahweol in CD<sub>3</sub>OD





<sup>1</sup>H, <sup>13</sup>C HSQC spectrum of kahweol in CD<sub>3</sub>OD





<sup>1</sup>H, <sup>13</sup>C HMBC spectrum of kahweol in CD<sub>3</sub>OD





<sup>1</sup>H NMR spectrum of compound **1** in acetone- $d_6$ 





<sup>13</sup>C NMR spectrum of compound **1** in acetone- $d_6$ 



# COSY spectrum of compound 1 in acetone- $d_6$





# TOCSY spectrum of compound 1 in acetone- $d_6$





# ROESY spectrum of compound 1 in acetone- $d_6$





<sup>1</sup>H, <sup>13</sup>C HSQC spectrum of compound **1** in acetone- $d_6$ 





<sup>1</sup>H, <sup>13</sup>C HMBC spectrum of compound **1** in acetone- $d_6$ 





<sup>1</sup>H NMR spectrum of compound **2** in CD<sub>3</sub>OD





<sup>13</sup>C NMR spectrum of compound **2** in CD<sub>3</sub>OD

















<sup>1</sup>H, <sup>13</sup>C HSQC spectrum of compound **2** in CD<sub>3</sub>OD





<sup>1</sup>H, <sup>13</sup>C HMBC spectrum of compound **2** in CD<sub>3</sub>OD





HPLC elution profile and ESI+/MS spectrum of kahweol isolated from green coffee beans. Elution conditions are described in the Experimental Procedures.



### **Kahweol**

HPLC elution profile and ESI+/MS spectrum of cafestol obtained from green coffee beans. Elution conditions are described in the Experimental Procedures.



### **Cafestol**

HPLC elution profiles of the ethyl acetate extract from the reaction mixture of coffee with nitrite ions (2 mM) at pH 3 (blue trace: 0 min; red trace: 2 h). Elution conditions are described in the Experimental Procedures.

