

# Kinetic and Thermodynamic Investigation of Lipase-Catalyzed Hydrolysis of (*R,S*)-3-Phenylbutyl Azolides

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## Supplementary Information

### 1. EXPERIMENTAL SECTION

The synthesized substrates are confirmed from the retention time in HPLC analysis (Table S1) or <sup>1</sup>H NMR spectra recorded at 500 MHz on Bruker spectrometer (Avance DRX 500) in DMSO-*d*<sub>6</sub> solution with TMS as an internal standard as follows:

(*R,S*)-3-Phenylbutyl 4-methylpyrazolide (**1**). <sup>1</sup>HNMR (DMSO-*d*<sub>6</sub>/TMS) δ: 1.21-1.28 (3H, q), 2.04 (3H, s), 3.28-3.46 (1H and 2H, m), 7.25-7.29 (5H, m), 7.71 (1H, s), 8.09 (1H, s). The abbreviations d, q, m and s are the peak multiplicities of doublet, quartet, multiplet and single, respectively.

(*R,S*)-3-Phenylbutyl 4-bromopyrazolide (**2**). <sup>1</sup>HNMR (DMSO-*d*<sub>6</sub>/TMS) δ: 1.27-1.29 (3H, d), 3.32-3.47 (1H and 2H, m), 7.25-7.30 (5H, m), 8.00 (1H, s), 8.62 (1H, s).

(*R,S*)-3-Phenylbutyl 4-nitropyrazolide (**3**). <sup>1</sup>HNMR (DMSO-*d*<sub>6</sub>/TMS) δ: 1.30-1.31 (3H, d), 3.35-3.56 (1H and 2H, m), 7.25-7.31 (5H, m), 8.57 (1H, s), 9.36 (1H, s).

(*R,S*)-3-Phenylbutyl 1,2,4-triazolide (**8**). <sup>1</sup>HNMR (DMSO-*d*<sub>6</sub>/TMS) δ: 1.30-1.31 (3H, d), 3.36-3.57 (1H and 2H, m), 7.26-7.35 (5H, m), 8.28 (1H, s), 9.27 (1H, s).

(*R,S*)-Methyl 3-phenylbutyrate (**9**). <sup>1</sup>HNMR (DMSO-*d*<sub>6</sub>/TMS) δ: 1.22-1.23 (3H, m), 2.61 (2H, d), 3.15-3.21 (1H, q), 3.54 (3H, s), 7.17-7.36 (5H, m).

(*R,S*)-3-(Boc-amino)-3-phenylpropionyl 4-methylpyrazolide (**10**). <sup>1</sup>HNMR (DMSO-*d*<sub>6</sub>/TMS) δ: 1.32 (9H, s), 2.03 (3H, s), 3.44-3.45 (2H, d), 5.01 (1H, s), 7.28-7.35 (5H, m), 7.51 (1H, s), 7.70 (1H, s), 8.10 (NH, s).

## 2. KINETIC ANALYSIS

At the initial stage, eq 1 for the fast-reacting substrate without and with adding the acid product is, respectively, reduced to

$$V_R = \frac{k_{2R}(S_R)_0(E_t)K_{mR}^{-1}}{1 + (S_R)_0[K_{mR}^{-1} + K_{mS}^{-1}]} \quad (S1)$$

$$V_R = \frac{k_{2R}(S_R)_0(E_t)K_{mR}^{-1}}{1 + (S_R)_0[K_{mR}^{-1} + K_{mS}^{-1}] + (I)K_I^{-1}} \quad (S2)$$

Moreover if the lipase has high (*R*)-enantioselectivity, the (*R*)-enantiomer will completely convert to the acid product (i.e. (*I*) = (*S<sub>R</sub>*)<sub>0</sub>) when estimating the initial rate *V<sub>S</sub>*. Therefore, the initial rate equation for the slow-reacting substrate is expressed as

$$V_S = \frac{k_{2S}(S_S)_0(E_t)K_{mS}^{-1}}{1 + (S_S)_0[K_{mR}^{-1} + K_I^{-1}]} \quad (S3)$$

By using the data of Figure 2,  $k_{2R}K_{mR}^{-1}$  and  $[K_{mR}^{-1} + K_{mS}^{-1}]$  are first estimated from eq S1, and then  $K_I$  from eq S2. Similarly, the kinetic constants  $k_{2S}$  and  $K_{mS}$  are estimated from eq S3, and then  $k_{2R}$ ,  $K_{mR}$ , and enantiomeric ratio *E* defined as  $k_{2R}k_{2S}^{-1}K_{mR}^{-1}K_{mS}$ . Therefore, the time-course variations of (*S<sub>R</sub>*)<sub>0</sub> and (*S<sub>S</sub>*)<sub>0</sub>, and hence *X<sub>R</sub>* and *X<sub>S</sub>* (Figures 1 and S1) are predicted from solving eqs 1-3 by using a fourth-order Runge-Kutta numerical method.

## 3. THERMODYNAMIC ANALYSIS

According to the transition theory with  $k_{-IR} \gg k_{2R}$  and  $k_{-IS} \gg k_{2S}$  for eqs 1 and 2 in the Michaelis-Menten kinetics, the logarithm of enantiomeric ratio can be expressed as  $R\ln(E) = -\Delta\Delta G/T = -\Delta\Delta H/T + \Delta\Delta S$ . Therefore from the variation of  $\ln(E)$  with  $T^{-1}$ ,

one may estimate  $-\Delta\Delta H$ ,  $-\Delta\Delta S$ , and hence  $-\Delta\Delta G$  at a specified temperature between the transition states of both enantiomers. The results of  $-\Delta\Delta H$  and  $-\Delta\Delta S$  along with others for Novozym 435-catalyzed resolution of (*R,S*)-azolides containing an  $\alpha$ -chiral center are represented in Table S2.

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- (5) Kao, M. F.; Lu, P. Y.; Kao, J. Y.; Wang, P. Y.; Wu, A. C.; Tsai, S. W. (*R,S*)-2-Chlorophenoxy pyrazolides as extreme substrates for improving lipase-catalyzed hydrolytic resolution. *Chirality*, **2011** (in press)

**Table S1. HPLC analytical conditions for various substrates illustrated in Scheme 1.**

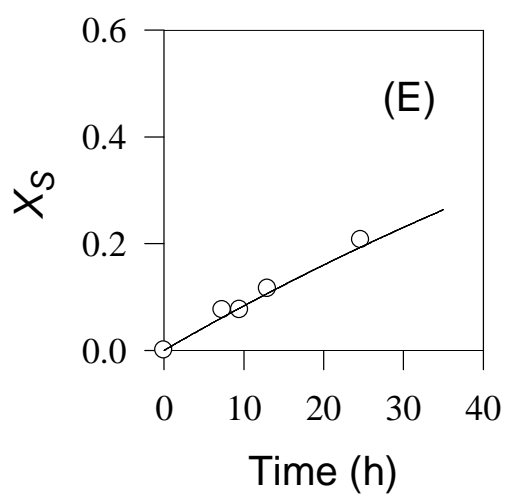
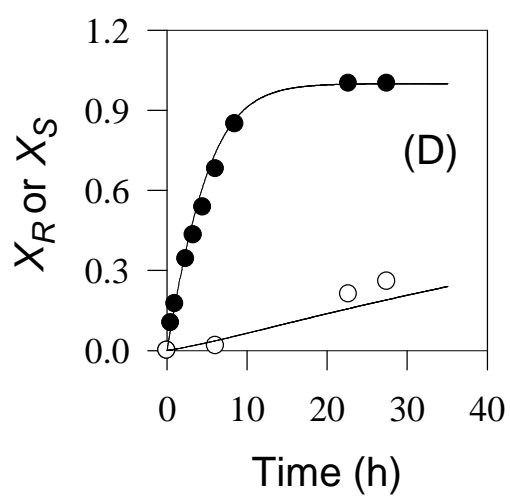
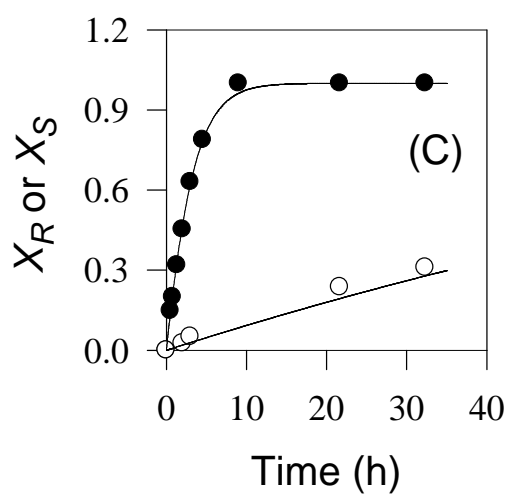
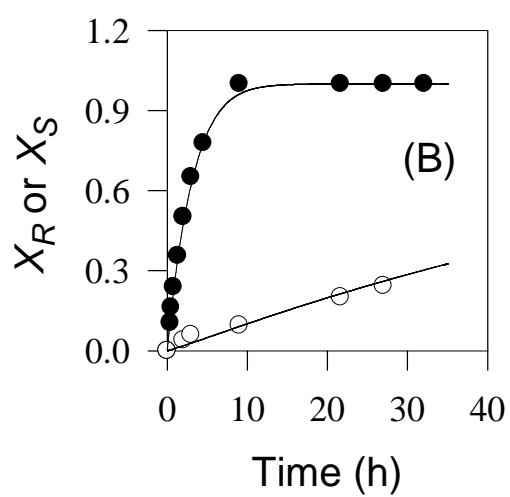
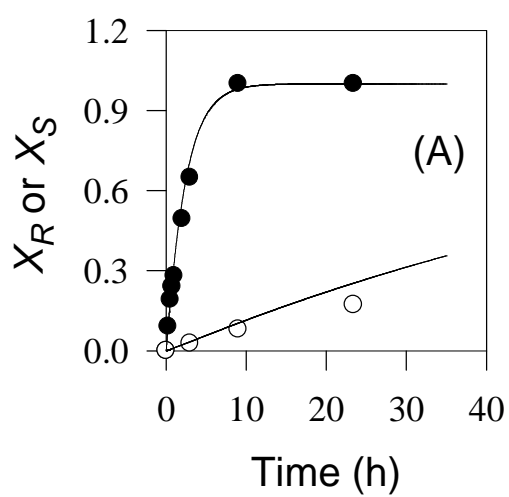
Substrate	Column	UV (nm)	Flow rate (mL min <sup>-1</sup> )	Internal standard <sup>[a]</sup>	Mobile phase <sup>[b]</sup> (HEX:IPA:AA, v/v)	Retention time (min)		
						Internal standard	( <i>R</i> )-substrate	( <i>S</i> )-substrate
<b>1</b>	OJ-H	270	2.0	A	89.5:10:0.5	13.5, 14.7	4.7	8.0
<b>2</b>	OJ	270	2.0	B	88:12:0:0	3.0	4.5	5.2
<b>3</b>	OJ-H	270	1.5	B	85:15:0	3.2	8.9	9.8
<b>4</b>	OJ-H	270	2.0	B	95:5:0	2.7	3.6	4.1
<b>5</b>	OJ-H	270	2.0	C	90:10:0	2.0	5.3	6.4
<b>6</b>	( <i>S,S</i> )-Whelk-O1	270	2.0	C	99:1:0	1.7	4.8	5.7
<b>7</b>	OJ-H	270	2.0	C	88:12:0	2.0	16.8	19.9
<b>8</b>	OD-H	220	1.0	C	50:50:0	3.6	11.1	6.4
<b>9</b>	OJ-H	220	2.0	B	95:5:0	3.4	4.4	3.2
<b>10</b>	OJ-H	270	2.0	C	90:10:0	2.0	5.1	7.9

<sup>[a]</sup> (A) (*R,S*)-Naproxen, (B) 2-nitrotoluene, (C) benzene. <sup>[b]</sup> IPA and AA as isopropyl alcohol and acetic acid, respectively.

**Table S2. - $\Delta\Delta H$  and - $\Delta\Delta S$  for Novozym 435-catalyzed kinetic resolution of (*R,S*)-azolides.**

Substrates	$-\Delta\Delta H$ (kJ mol <sup>-1</sup> )	$-\Delta\Delta S$ (J mol <sup>-1</sup> K <sup>-1</sup> )	Reaction, solvent, stereo-preference
( <i>R,S</i> )-3-Phenylbutyl 4-methylpyrazolide	20.39	37.97	Hydrolysis, water-saturated CYC, R
( <i>R,S</i> )-3-Phenylbutyl 4-bromopyrazolide	17.61	26.74	Hydrolysis, water-saturated CYC, R
( <i>R,S</i> )-2-Phenylpropionyl 1,2,4-triazolide <sup>1</sup>	19.39	23.71	Hydrolysis, water-saturated MTBE, R
( <i>R,S</i> )-Naproxenyl 1,2,4-triazolide <sup>2</sup>	17.09	15.45	Alcoholysis by methanol, anhydrous MTBE, R
( <i>R,S</i> )-Flurbirpofenyl 1,2,4-triazolide <sup>3</sup>	8.43	-10.23	Alcoholysis by methanol, anhydrous MTBE, R
( <i>R,S</i> )-Flurbirpofenyl 4-bromopyrazolide <sup>3</sup>	7.54	-20.38	Alcoholysis by 2,3-dibromo-1-propanol, anhydrous MTBE, R
( <i>R,S</i> )-2-Methylheptyl 3-(2-pyridyl)pyrazolide <sup>4</sup>	27.17	51.12	Hydrolysis, water-saturated MTBE, R
( <i>R,S</i> )-2-Methylheptyl 3-(2-pyridyl)pyrazolide <sup>4</sup>	12.35	5.43	Hydrolysis, water-saturated IPE, R
( <i>R,S</i> )-2-Methylheptyl 3-(2-pyridyl)pyrazolide <sup>4</sup>	28.45	51.46	Alcoholysis by methanol by methanol, anhydrous MTBE, R
( <i>R,S</i> )-2-Methylheptyl 3-(2-pyridyl)pyrazolide <sup>4</sup>	59.10	151.0	Alcoholysis by methanol, anhydrous IPE, R
( <i>R,S</i> )-2-(2-Chlorophenoxy)propionyl 3-(2-pyridine)pyrazolide <sup>5</sup>	22.89	40.89	Hydrolysis, water-saturated MTBE, S
( <i>R,S</i> )-2-(3-Chlorophenoxy)propionyl 3-(2-pyridine)pyrazolide <sup>5</sup>	23.92	47.94	Hydrolysis, water-saturated MTBE, S
( <i>R,S</i> )-2-(4-Chlorophenoxy)propionyl 3-(2-pyridine)pyrazolide <sup>5</sup>	9.93	-13.70	Hydrolysis, water-saturated MTBE, S
( <i>R,S</i> )-2-(2,4-Dichlorophenoxy)propionyl	13.02	-5.51	Hydrolysis, water-saturated MTBE, S

3-(2-pyridine)pyrazolide<sup>5</sup>



**Figure S1.** Time-course conversions  $X_R$  (●) and  $X_S$  (○) at 25 °C in water-saturated CYC containing 40 mg mL<sup>-1</sup> of Novozym 435 and (A) 15 mM, (B) 30 mM, (C) 60 mM, and (D) 90 mM of **1**, or those of  $X_S$  (○) for (E) 75 mM of (*S*)-**1**; theoretical predictions via eqs 1-3 (—).