SUPPORTING INFORMATION

Carboxylation and Decarboxylation of Active Site Lys 84 Controls the Activity of OXA-24 β-Lactamase of *Acinetobacter baumannii*: Raman Crystallographic and Solution Evidence

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Supplemental Figure Captions

Figure S1. Overall difference Raman spectra of OXA-24 β -lactamase complexed with SA-1-204 for selected time points.

Figure S2. Overall kinetic depiction of the main peaks intensities over time. A. Kinetic depiction of the substrate peaks intensities at 1695 cm⁻¹, 1588 cm⁻¹, 1570 cm⁻¹, 1216 cm⁻¹ over time. B. Kinetic depiction of the intermediate peaks intensities at 1659 cm⁻¹, 1550 cm⁻¹, 1447 cm⁻¹, 1322 cm⁻¹ over time.

Figure S3. Difference absorbance spectrum of SA-1-204 in OXA-24 solution with 10 mM HEPES (pH 7.5)s. The spectrum of OXA-24 and SA-1-204 were first acquired separately. Once they were mixed, spectrum was acquired at the intervals of 30 seconds in a 30 minutes time range. A. The reaction undergoes one cycle in the absence of NaHCO₃. 20 μ M OXA-24 and 60 μ M SA-1-204 were mixed. Without NaHCO₃, the spectrum from 1 minutes to 15 minutes are from the mixture of product and unreacted substrates. B. The reaction undergoes more than one cycle in the presence of 100 mM NaHCO₃. With NaHCO3 in solution, the reaction happens quickly and all the substrates (black line) are transformed into product (from 1 minute to 15 minutes).

Figure S4. Two views of the energy-minimized structure of intermediate X in Scheme 3. Color representations: white: hydrogen, gray: carbon, blue: nitrogen, red: oxygen, yellow: sulfur. A, from the front view; B, from the bottom view. The structure is very similar to that of the products shown in the active site of OXA-24 in Figure 2 of Bou *et al.*¹

S2





Wavenumber (cm⁻¹)

Figure S2











Supplemental Text

1). Acquisition of Raman difference spectrum

For the spectrum of unreacted inhibitor SA-1-204, a buffer spectrum (W/O 28% PEG 2000) was obtained in a 5 µL hanging drop with a power of 80 mW (Spectrum 1). Next, spectrum of 5 mM inhibitor SA-1-204 was obtained in the same buffer as the above control, respectively (W/O 28% PEG 2000) (Spectrum 2). The Raman difference spectrum of inhibitor SA-1-204 was obtained by subtracting Spectrum 1 from Spectrum 2. Spectrum subtraction was performed using GRAMS/32 software (Galactic Industries, Inc.).

For the spectrum of bound inhibitor SA-1-204 in the active site of OXA-24, a spectrum of OXA-24 crystal in holding solutions containing 0.1 M HEPES (pH 7.5), 0.1M sodium acetate and 28% PEG 2000 was firstly recorded (Spectrum 3). Then, a spectrum of its mother liquor, obtained by shifting the hanging drop ~50 µm so that the laser focal point is away from the crystal (Spectrum 4). After Spectrum 4 was subtracted from Spectrum 3, the Raman difference spectrum of OXA-24 crystal was obtained (Spectrum 5). After inhibitor SA-1-204 was soaked in (final concentration is 5 mM), the time-dependent spectrum was recorded and the laser focal point on the crystal was at the same point as that before soak in. The spectrum of crystal after soak in (Spectrum 6) and new buffer (Spectrum 7) were obtained, respectively. A similar spectral subtraction was performed to give Raman difference spectrum of crystal after soak in (Spectrum 8). Finally, a Raman spectrum of bound SA-1-204 in the OXA-24 crystal plus any conformational changes of the enzyme was obtained by subtracting the following two spectra:

Raman difference spectrum of bound ligand = Spectrum 8 - Spectrum 5 = [(Spectrum 6 + Spectrum 7) - (Spectrum 7)] - [(Spectrum 3 + Spectrum 4) - Spectrum 4] In this final trace, the intensities of the Raman peaks were ratiod against 1000 cm⁻¹ phenyl ring mode to enable us to compare intensities among spectra. The intensities of the modes in Fig. 2 are about 3% compared to the intense peaks in the acyl-enzyme mother spectrum.

2). Concentration of active sites in a single crystal is calculated as follows:

number of asymmetric subunits per unit cell /(
$$6.02 \times 10^{23} \text{ Mole}^{-1}$$
) volume per unit cell

Bou, G.; Santillana, E.; Sheri, A.; Beceiro, A.; Sampson, J. M.; Kalp, M.; Bethel, C. R.;
Distler, A. M.; Drawz, S. M.; Pagadala, S. R.; van den Akker, F.; Bonomo, R. A.; Romero, A.; Buynak, J.
D. J Am Chem Soc 2010, 132, 13320.