

Conversion Type	Equation	Reference
<i>For COPEPOD database biomass</i>		
Displacement Volume (DV) to Carbon Mass (CM)	$\log \text{CM} = (\log \text{DV} + 1.434)/0.820$	Wiebe (1988)
Wet Mass (WM) to CM	$\log \text{CM} = (\log \text{WM} + 1.537)/0.852$	Wiebe (1988)
Dry Mass (DM) to CM	$\log \text{CM} = (\log \text{DM} - 0.499)/0.991$	Wiebe (1988)
Settled Volume (SV) to DM	$\log \text{DM} = 0.843 \cdot \log \text{SV} + 1.417$	Balvay (1987)
Ash-free Dry Mass (AFDM) to CM	$\log \text{CM} = (\log \text{AFDM} - 0.410)/0.963$	Bode et al. (1998)
<i>For COPEPOD database mesh size – Converted to 333um mesh equivalent</i>		
CM sampled with 200um net	$\text{Log CMM333} = 0.6195 \cdot \log \text{CMM200}$	O'Brien (2005)
CM sampled with 505um net	$\text{Log CMM333} = 1.2107 \cdot \log \text{CMM505}$	O'Brien (2005)
<i>For Continuous Plankton Recorder – Abundance x TL to DM/WM conversions</i>		
Total Length (TL) to WM Copepoda & Ostracoda	$\text{WM} = 0.08 \cdot \text{TL}^{2.1}$	Richardson et al., (2006)
TL to DM Amphipod	$\log \text{DM} = 2.71 \cdot \log \text{TL} + 0.76$	Yamada (2002)
TL to DM Chaetognatha	$\log \text{DM} = 2.80 \cdot \log \text{TL} - 0.6$	Imao (2005)
TL to DM Annelida	$\log \text{DM} = 1.53 \cdot \log \text{TL} + 1.49$	Imao (2005)
TL to DM Euphausiacea		
<i>Euphausia spp.</i>	$\log \text{DM} = 2.64 \cdot \log \text{TL} + 0.92$	Lindley et al., (1999)
<i>Nyctiphanes spp.</i>	$\log \text{DM} = 2.23 \cdot \log \text{TL} + 1.12$	Lindley et al., (1999)
<i>Thysanoessa spp.</i>	$\log \text{DM} = 2.73 \cdot \log \text{TL} + 0.881$	Lindley et al., (1999)
Calypotes (All)	$\log \text{DM} = 1.40 \cdot \log \text{TL} + 1.194$	Lindley et al., (1999)
TL to DM Appendicularia	$\text{DM} = 2.47 \cdot \log \text{TL} - 6.1$	Hopcroft et al., (1998)
TL to DM Pteropoda	$\text{DM} = 0.1365 \cdot \text{TL}^{1.501}$	Bednarsek et al., (2012)

Supplementary Table 1: The list of conversion equations for transforming the various methods for calculating zooplankton biomass into a standard unit of Carbon mass. Also shown are the mesh size conversions where the carbon mass measured from 200um and 505um mesh size nets were standardised to a 333um mesh size equivalent. The final lists of equations are the length-weight relationships for the dominant groups within the Continuous Plankton Recorder database. The total biomass within a sample is obtained by multiplying the total number of individuals by the mass of the individuals within a sample.

Group	Species	Length (mm)	Reference
Amphipod	Hyperiella sp.	6	See notes ¹
Amphipod	Hyperiididae indet	12	McHardy 1970 ²
Amphipod	<i>Primno macropa</i>	12	Bowman 1978
Amphipod	<i>Primno spp.</i>	12	McHardy 1970 ²
Annelida	<i>Tomopteris sp.</i>	20	See notes ³
Appendicularia	Appendicularia	0.8	Uye & Ichino, 1995 ⁴
Appendicularia	<i>Oikopleura sp.</i>	0.8	Uye & Ichino, 1995 ⁴
Chaetognatha	Chaetognatha indet	3	Bainbridge 1963 ⁵
Chaetognatha	<i>Eukrohnia hamata</i>	10.2	Bielecka et al., 2016
Chaetognatha	<i>Pseudosagitta gazellae</i>	20.1	Bielecka et al., 2016
Chaetognatha	<i>Serratosagitta tasmanica</i>	13.3	Sotelo et al., 2018
Copepoda	<i>Acartia danae</i>	1.08	Brun et al., 2017
Copepoda	<i>Acartia negligens</i>	1.04	Brun et al., 2017
Copepoda	<i>Acartia spp.</i>	1.04	Brun et al., 2017
Copepoda	<i>Acartiura tranteri</i>	1.03	Brun et al., 2017
Copepoda	<i>Acrocalanus spp.</i>	1	Brun et al., 2017
Copepoda	<i>Aetideus armatus</i>	1.73	Brun et al., 2017
Copepoda	<i>Alteutha spp</i>	0.95	Brun et al., 2017
Copepoda	<i>Anomalocera patersoni</i>	3.3	Brun et al., 2017
Copepoda	Calanidae	2	Brun et al., 2017
Copepoda	<i>Calanoides acutus</i>	4.1	Brun et al., 2017
Copepoda	<i>Calanoides carinatus</i>	1.74	Brun et al., 2017
Copepoda	<i>Calanoides spp.</i>	3.5	Brun et al., 2017
Copepoda	<i>Calanus australis</i>	2.62	Brun et al., 2017
Copepoda	<i>Calanus finmarchicus</i>	2.7	Brun et al., 2017
Copepoda	<i>Calanus glacialis</i>	4.6	Brun et al., 2017
Copepoda	<i>Calanus helgolandicus</i>	2.68	Brun et al., 2017
Copepoda	<i>Calanus hyperboreus</i>	6.95	Brun et al., 2017
Copepoda	<i>Calanus IV</i>	1.65	Brun et al., 2017
Copepoda	<i>Calanus propinquus</i>	5.1	Brun et al., 2017
Copepoda	<i>Calanus simillimus</i>	3.1	Brun et al., 2017
Copepoda	<i>Calocalanus pavo</i>	0.74	Brun et al., 2017
Copepoda	<i>Calocalanus sp</i>	0.84	Brun et al., 2017
Copepoda	<i>Calocalanus spp</i>	0.75	Brun et al., 2017
Copepoda	<i>Calocalanus spp.</i>	0.75	Brun et al., 2017
Copepoda	<i>Candacia armata</i>	2.18	Brun et al., 2017
Copepoda	<i>Candacia IV</i>	1.4	Brun et al., 2017
Copepoda	<i>Candacia maxima</i>	2.15	Brun et al., 2017
Copepoda	<i>Candacia spp.</i>	1.95	Brun et al., 2017
Copepoda	<i>Canthocalanus pauper</i>	1.32	Brun et al., 2017
Copepoda	<i>Centropages australiensis</i>	1.12	Brun et al., 2017
Copepoda	<i>Centropages bradyi</i>	1.85	Brun et al., 2017
Copepoda	<i>Centropages chierchiae</i>	3.25	Brun et al., 2017
Copepoda	<i>Centropages furcatus</i>	1.5	Brun et al., 2017

Copepoda	<i>Centropages gracilis</i>	1.74	Brun et al., 2017
Copepoda	<i>Centropages hamatus</i>	1.3	Brun et al., 2017
Copepoda	<i>Centropages spp.</i>	1.63	Brun et al., 2017
Copepoda	<i>Centropages typicus</i>	1.55	Brun et al., 2017
Copepoda	<i>Centropages violaceus</i>	1.8	Brun et al., 2017
Copepoda	<i>Clausocalanus brevipes</i>	1.15	Brun et al., 2017
Copepoda	<i>Clausocalanus laticeps</i>	1.15	Brun et al., 2017
Copepoda	<i>Clausocalanus spp.</i>	1.15	Brun et al., 2017
Copepoda	<i>Clausocalanus spp.</i>	0.68	Brun et al., 2017
Copepoda	Copepod nauplius	0.04	Brun et al., 2017
Copepoda	<i>Corycaeus spp.</i>	0.65	Brun et al., 2017
Copepoda	<i>Cosmocalanus darwinii</i>	2.12	Brun et al., 2017
Copepoda	<i>Ctenocalanus citer</i>	0.8	Brun et al., 2017
Copepoda	<i>Ctenocalanus vanus</i>	0.78	Brun et al., 2017
Copepoda	<i>Eucalanus elongatus</i>	5	Brun et al., 2017
Copepoda	<i>Eucalanus hyalinus</i>	3.4	Brun et al., 2017
Copepoda	<i>Euchaeta acuta</i>	3.84	Brun et al., 2017
Copepoda	<i>Euchaeta marina</i>	2.7	Brun et al., 2017
Copepoda	<i>Euchaetidae</i>	4	Brun et al., 2017
Copepoda	<i>Euchirella spp.</i>	2	Brun et al., 2017
Copepoda	<i>Haloptilus oxycephalus</i>	8	Brun et al., 2017
Copepoda	<i>Heterorhabdus norvegicus</i>	2.49	Brun et al., 2017
Copepoda	<i>Heterorhabdus papilliger</i>	2.5	Brun et al., 2017
Copepoda	<i>Isias clavipes</i>	1.25	Brun et al., 2017
Copepoda	<i>Labidocera spp.</i>	1.95	Brun et al., 2017
Copepoda	<i>Labidocera wollastoni</i>	3	Brun et al., 2017
Copepoda	<i>Lubbockia spp.</i>	1.2	Brun et al., 2017
Copepoda	<i>Lucicutia spp.</i>	5.2	Brun et al., 2017
Copepoda	<i>Lucifer spp.</i>	1.3	Brun et al., 2017
Copepoda	<i>Macrosetella spp.</i>	0.98	Brun et al., 2017
Copepoda	<i>Mecynocera clausi</i>	1	Brun et al., 2017
Copepoda	<i>Mecynocera spp.</i>	0.84	Brun et al., 2017
Copepoda	<i>Mesocalanus spp.</i>	1.75	Brun et al., 2017
Copepoda	<i>Mesocalanus tenuicornis</i>	1.95	Brun et al., 2017
Copepoda	<i>Metridia gerlachei</i>	0.9	Brun et al., 2017
Copepoda	<i>Metridia IV</i>	0.93	Brun et al., 2017
Copepoda	<i>Metridia longa</i>	4.1	Brun et al., 2017
Copepoda	<i>Metridia lucens</i>	3.5	Brun et al., 2017
Copepoda	<i>Metridia sp</i>	2.27	Brun et al., 2017
Copepoda	<i>Microsetella sp</i>	2	Brun et al., 2017
Copepoda	<i>Microsetella spp</i>	0.55	Brun et al., 2017
Copepoda	<i>Nannocalanus minor</i>	1.71	Brun et al., 2017
Copepoda	<i>Nauplii Rhincalanus</i>	3	Brun et al., 2017
Copepoda	<i>Neocalanus spp.</i>	7	Brun et al., 2017
Copepoda	<i>Neocalanus tonsus</i>	0.5	Brun et al., 2017
Copepoda	<i>Odontacartia pacifica</i>	1.15	Brun et al., 2017
Copepoda	<i>Oithona nana</i>	0.71	Brun et al., 2017

Copepoda	<i>Oithona similis</i>	0.5	Brun et al., 2017
Copepoda	<i>Oithona</i> spp.	0.68	Brun et al., 2017
Copepoda	<i>Oncaea clevei</i>	0.86	Brun et al., 2017
Copepoda	<i>Oncaea media</i>	0.86	Brun et al., 2017
Copepoda	<i>Oncaea scottodicarloi</i>	0.86	Brun et al., 2017
Copepoda	<i>Oncaea</i> spp.	0.86	Brun et al., 2017
Copepoda	<i>Oncaea venusta</i>	0.86	Brun et al., 2017
Copepoda	<i>Para Pseudocalanus</i>	0.75	Brun et al., 2017
Copepoda	<i>Paracalanus indicus</i>	0.75	Brun et al., 2017
Copepoda	<i>Paracalanus</i> spp.	0.75	Brun et al., 2017
Copepoda	<i>Paraeuchaeta exigua</i>	0.7	Brun et al., 2017
Copepoda	<i>Paraeuchaeta hebes</i>	2.4	Brun et al., 2017
Copepoda	<i>Paraeuchaeta norvegica</i>	7	Brun et al., 2017
Copepoda	<i>Paraeuchaeta</i> spp.	7	Brun et al., 2017
Copepoda	<i>Pareucalanus</i> spp.	5.89	Brun et al., 2017
Copepoda	<i>Pleuromamma abdominalis</i>	2.67	Brun et al., 2017
Copepoda	<i>Pleuromamma borealis</i>	1.97	Brun et al., 2017
Copepoda	<i>Pleuromamma gracilis</i>	1.76	Brun et al., 2017
Copepoda	<i>Pleuromamma piseki</i>	1.73	Brun et al., 2017
Copepoda	<i>Pleuromamma robusta</i>	1.73	Brun et al., 2017
Copepoda	<i>Pleuromamma</i> spp.	2.56	Brun et al., 2017
Copepoda	<i>Pleuromamma xiphias</i>	4.13	Brun et al., 2017
Copepoda	<i>Pseudocalanus</i> spp	1.2	Brun et al., 2017
Copepoda	<i>Rhincalanus gigas</i>	15	Brun et al., 2017
Copepoda	<i>Rhincalanus nasutus</i>	3.99	Brun et al., 2017
Copepoda	<i>Rhincalanus</i> spp.	4.5	Brun et al., 2017
Copepoda	<i>Scolecithricella minor</i>	3	Brun et al., 2017
Copepoda	<i>Scolecithricella</i> spp	1.4	Brun et al., 2017
Copepoda	<i>Scolecithrix danae</i>	1.16	Brun et al., 2017
Copepoda	Small Calanoid	1.34	Brun et al., 2017
Copepoda	<i>Subeucalanus crassus</i>	1.1	Brun et al., 2017
Copepoda	<i>Subeucalanus longiceps</i>	1	Brun et al., 2017
Copepoda	<i>Subeucalanus</i> sp	4.1	Brun et al., 2017
Copepoda	<i>Subeucalanus subcrassus</i>	2.2	Brun et al., 2017
Copepoda	<i>Temora discaudata</i>	2	Brun et al., 2017
Copepoda	<i>Temora longicornis</i>	0.75	Brun et al., 2017
Copepoda	<i>Temora</i> spp.	1.5	Brun et al., 2017
Copepoda	<i>Temora stylifera</i>	1.45	Brun et al., 2017
Copepoda	<i>Temora turbinata</i>	1.3	Brun et al., 2017
Copepoda	<i>Themisto gaudichaudii</i>	14	Brun et al., 2017
Copepoda	<i>Tortanus</i> spp.	0.72	Brun et al., 2017
Copepoda	<i>Triconia conifera</i>	0.65	Brun et al., 2017
Copepoda	<i>Undeuchaeta plumosa</i>	3.18	Brun et al., 2017
Copepoda	<i>Undinula vulgaris</i>	1.76	Brun et al., 2017
Euphausiid	<i>Euphausia</i>	9	Lindley 1977 ⁶
Euphausiid	<i>Euphausia frigida</i>	15	Siegel 1987
Euphausiid	<i>Euphausia longirostris</i>	20	Gurney et al., 2002

Euphausiid	<i>Euphausia superba</i>	33.9	Färber -Lora, 1994
Euphausiid	<i>Euphausia vallentini</i>	15	Siegel 1987
Euphausiid	<i>Euphausiacea calyptopis</i>	2	Lindley 1977 ⁷
Euphausiid	<i>Nyctiphanes australis</i>	12	Ritz and Hosie, 1982
Euphausiid	<i>Thysanoessa calyptopis</i>	3	Wallis 2018 ⁸
Euphausiid	<i>Thysanoessa gregaria</i>	15	Croxhall et al., 1985
Euphausiid	<i>Thysanoessa macrura</i>	17.2	Färber -Lora, 1994
Euphausiid	<i>Thysanoessa spp.</i>	15	T. gregaria
Ostracoda	<i>Evadne spp</i>	0.75	Richardson et al., 2006
Ostracoda	Ostracoda indet	0.7	Podon/Evadne sp.
Ostracoda	<i>Penilia avirostris</i>	1.1	Richardson et al., 2006
Ostracoda	<i>Podon spp.</i>	0.65	Richardson et al., 2006
Pteropoda	<i>Clio pyramidata</i>	7.43	Iersel et al., 1982
Pteropoda	<i>Limacina sp.</i>	1.5	Richardson et al., 2006 ⁹

Supplementary table 2: A list of the species and taxonomic groups used for the length-weight conversions within the Continuous Plankton Recorder dataset. All Copepoda lengths are taken from the trait database of marine copepods compiled by (Brun et al., 2017). The sizes are the mean lengths recorded for adult females. Notes: For broader taxonomic groups or life stages we assume that these belong to the more common species belonging to that group within the area. ¹ *Hypierella dilatata* ² *Parathemisto gaudichaudii* & *Primno macropa*. ³ *Tompteris helgolandica*. ⁴ Mean trunk length of the cosmopolitan species *Oikopleura dioica*. ⁵ *Sagitta setosa stage I* ⁶ *T. longicaudata* is most commonly found in samples of the North Atlantic CPR (Lindley, 1977) ⁷ *T. longicaudata calyptosis stage III* ⁸ *T. macrura calyptosis stage III*. ⁹ *Limacina retroversa*

References:

- Bainbridge, V. (1963). Continuous plankton records: contribution towards a plankton atlas of the north Atlantic and the North Sea. Part VIII: Chaetognatha. *Bulletins of Marine Ecology*, 6, 40–51.
- Balvay, P. G. 1987. Equivalence entre quelques parametres estimatifs del'abondance du zooplankton total, *Schweiz. Z. Hydrol.*, 49, 75–83.
- Bednarsek, N., Tarling, G., Fielding, S., and Bakker, D. 2002. Population dynamics and biogeochemical significance of *Limacina helicina antarctica* in the Scotia Sea (Southern Ocean), *Deep-Sea Research Pt. II*, 59–60, 105–116.
- Bielecka, L., Jerzak, B. and Złoch, I., 2016. Species composition, seasonal abundance and population structure of chaetognaths in Admiralty Bay (Antarctic). *Polish Polar Research*, 37(2), pp.303-324.
- Bode, A., Alvarez-Ossorio, M. T., and Gonzales, N. 1998. Estimation of mesozooplankton biomass in a coastal upwelling area off NW Spain, *Journal of Plankton Research*, 20, 1005–1014.
- Bowman, T.E., 1978. Revision of the pelagic amphipod genus *Primo* (Hyperidea: Phrosinidae).

- Brun, P., Payne, M.R. and Kiørboe, T., 2017. A trait database for marine copepods. *Earth System Science Data*, 9(1), pp.99-113.
- Croxall, J.P., Prince, P.A., Baird, A. and Ward, P., 1985. The diet of the southern rockhopper penguin *Eudyptes chrysocome chrysocome* at Beauchene Island, Falkland Islands. *Journal of Zoology*, 206(4), pp.485-496.
- Färber-Lorda, J., 1994. Length-weight relationships and coefficient of condition of *Euphausia superba* and *Thysanoessa macrura* (Crustacea: Euphausiacea) in southwest Indian Ocean during summer. *Marine Biology*, 118, pp.645-650.
- Gurney, L.J., Froneman, P.W., Pakhomov, E.A. and McQuaid, C.D., 2002. Diel feeding patterns and daily ration estimates of three subantarctic euphausiids in the vicinity of the Prince Edward Islands (Southern Ocean). *Deep Sea Research Part II: Topical Studies in Oceanography*, 49(16), pp.3207-3227.
- Hopcroft, R.R., Roff, J.C., Bouman, H.A., 1998. Zooplankton growth rates: the larvaceans *Appendicularia*, *Fritillaria* and *Oikopleura* in tropical waters. *Journal of Plankton Research*, 20, 539-555.
- Lindley, J. A. (1977). Continuous plankton records: the distribution of the Euphausiacea (Crustacea: Malacostraca) in the North Atlantic and the North Sea, 1966–1967. *Journal of Biogeography*, 4, 121–133.
- Lindley, J.A., Robins, D.B., Williams, R. 1999. Dry weight carbon and nitrogen content of some euphausiids from the North Atlantic Ocean and the Celtic Sea. *Journal of Plankton Research* Vol.21 no.11 pp.2053–2066
- Iersel, P.V., 1982. Variation in *Diacria trispinosa* (De Blainville, 1821) and *Clio pyramidata* Linnaeus, 1767 (Pteropoda, Mollusca) of the Amsterdam Mid North Atlantic Plankton Expedition 1980. *Beaufortia*, 32(6), pp.97-115
- Imao F. 2005. Zooplankton community structure and functional role in carbon cycle in the Oyashio region, western North Pacific. Master's Thesis, Hokkaido University, pp. 42. (in Japanese)
- O'Brien, T. D. 2005. COPEPOD: A Global Plankton Database, US Dep. Commerce, NOAA Tech. Memo, 136 pp.
- Richardson, A. J., Walne, A. W., John, A. W. G., Jonas, T. D., Lindley, J. A., et al. 2006. Using continuous plankton recorder data. *Progress in Oceanography* 68, 27–74
- Ritz, D.A. and Hosie, G.W., 1982. Production of the euphausiid *Nyctiphanes australis* in Storm Bay, south-eastern Tasmania. *Marine biology*, 68, pp.103-108.
- Siegel, V., 1987. Age and growth of Antarctic Euphausiacea (Crustacea) under natural conditions. *Marine Biology*, 96, pp.483-495.
- Sotelo, I., Thompson, G., Sabatini, M., Reta, R. and Daponte, M.C., 2018. Mesoscale distribution and population structure of the chaetognath *Serratosagitta tasmanica* (Thomson, 1947) from Southwestern Atlantic Ocean. *Marine Biology Research*, 14(9-10), pp.945-960.

Uye, S.I. and Ichino, S., 1995. Seasonal variations in abundance, size composition, biomass and production rate of *Oikopleura dioica* (Fol)(Tunicata: Appendicularia) in a temperate eutrophic inlet. *Journal of Experimental Marine Biology and Ecology*, 189(1-2), pp.1-11.

Wallis, J.R., 2018. A description of the post-naupliar development of Southern Ocean krill (*Thysanoessa macrura*). *Polar Biology*, 41(12), pp.2399-2407.

Wiebe, P. H. 1988. Functional regression equations for zooplankton displacement volume, wet weight, dry weight, and carbon. A correction, *Fisheries Bulletin*, 86, 833–835.

Yamada Y, Ikeda T, Tsuda A. 2002. Abundance, growth and life cycle of the mesopelagic amphipod *Primno abyssalis* (Hyperidea: Phrosinidae) in the Oyashio region, western subarctic. *Pacific Journal of Marine Biology*, 141:333-341.