



Supporting Information

Cu-Catalyzed Asymmetric Borylative Cyclization of Cyclohexadienone-Containing 1,6-Enynes**

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¹H NMR, ¹³C NMR, HSQC, HMBC, DEPT, H-H COSY & NOSEY COPIES

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TRANSFORMATIONS

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1. GENERAL INFORMATION

All solvents were dried before use following the standard procedures. Unless otherwise indicated, all starting materials purchased from commercial suppliers were used without further purification. The ^1H and ^{13}C NMR spectra were recorded on Bruker AV-400 MHz in the indicated solvents. Chemical shifts are reported in δ (ppm) referenced to an internal TMS standard for ^1H NMR and CDCl_3 ($\delta = 77.10$ ppm) for ^{13}C NMR. Coupling constants (J) are quoted in Hz. Optical rotations were measured on a JASCO P-1030 polarimeter. IR spectra were recorded on Nicolet iN 10 MX. ESI mass spectra were recorded on Agilent1200/G6100A. HRMS of boron-containing compounds is based on ^{10}B .

2. PRELIMINARY REACTION INVESTIGATION

Initially, various ligands were investigated in Cu-catalyzed tandem borylative cyclization of the model substrate **1a**.

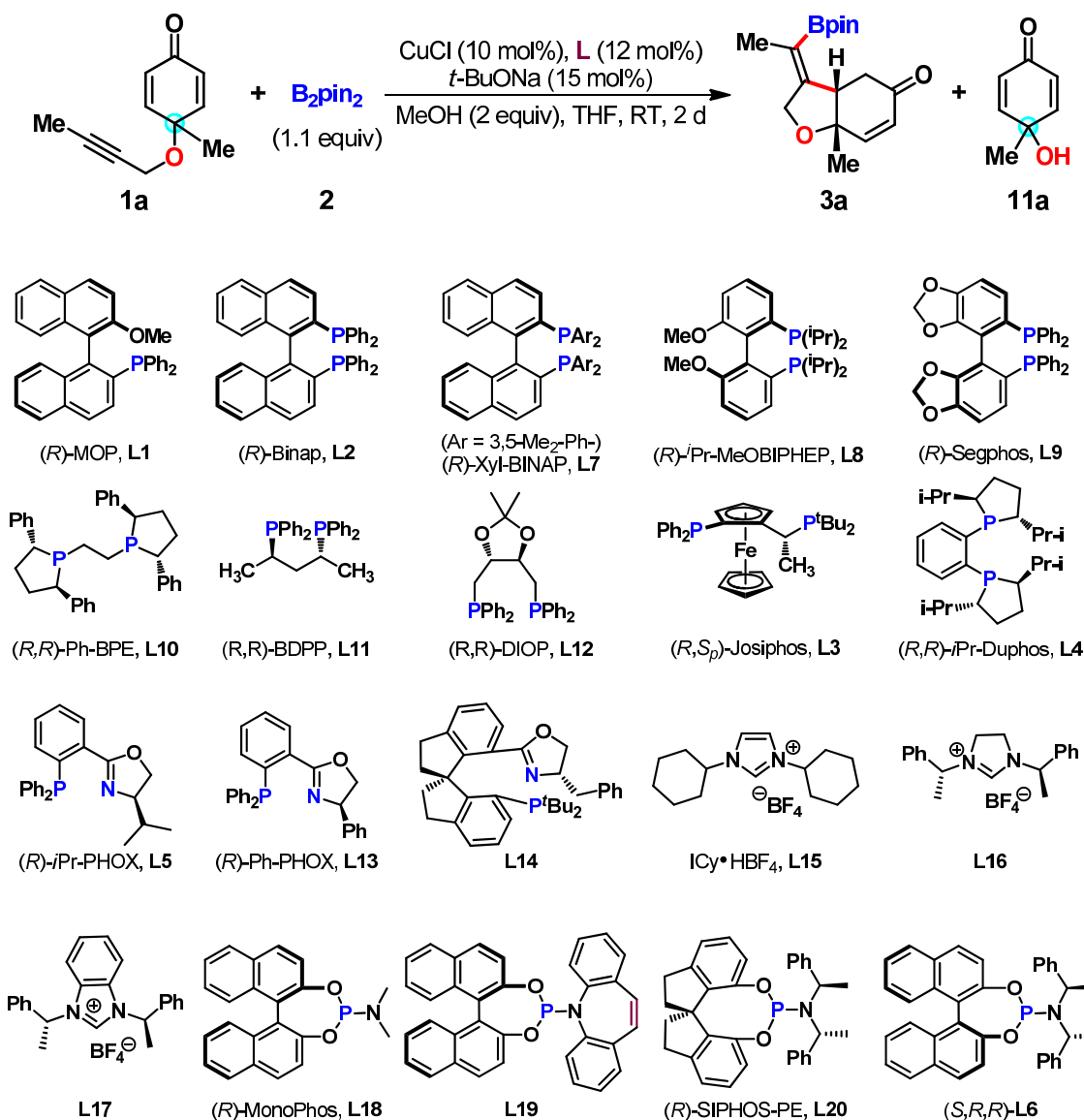


Table 3. Evaluation of Various Ligands for Cu-Catalyzed Tandem Borylative Cyclization of **1a**.^a

Entry	Ligand	Yield of Recovered 1a (%) ^b	Conversion (%) ^c	Yield of 11a (%) ^b	Yield of 3a (%) ^b	ee of 3a (%) ^d
1	Ph ₃ P	25	75	0	49	--
2 ^{e, g}	L15 , ICy·HBF ₄	14	86	33	27	--
3	L1 , (<i>R</i>)-MOP	16	84	0	47	-2
4	L2 , (<i>R</i>)-Binap	45	55	0	15	56
5 ^f	L2 , (<i>R</i>)-Binap	43	57	0	10	47
6	L7 , (<i>R</i>)-Xyl-Binap	57	43	0	12	49
7	L8 , (<i>R</i>)- <i>i</i> Pr-MeO-Biphep	57	43	0	18	52
8	L9 , (<i>R</i>)-Segphos	61	39	0	20	48
9	L10 , (<i>R,R</i>)-Ph-BPE	62	38	0	10	-1
10	L11 , (<i>R,R</i>)-BDPP	49	51	0	3	39
11	L12 , (<i>R,R</i>)-DIOP	47	53	0	10	40
12	L3 , (<i>R,Sp</i>)-Josiphos	60	40	0	12	-21
13	L4 , (<i>R,R</i>)- <i>i</i> Pr-Duphos	57	43	0	7	-65
14	L5 , (<i>R</i>)- <i>i</i> Pr-PHOX	35	65	0	33	-10
15	L13 , (<i>R</i>)-Ph-PHOX	52	48	0	45	1
16	L14	61	39	0	37	-4
17 ^e	L16	24	76	0	47	-35
18 ^e	L17	66	34	0	7	-2
19	L18 , (<i>R</i>)-MonoPhos	24	76	0	64	-45
20	L19	59	41	0	26	-78
21	L20 , (<i>R</i>)-SIPHOS-PE	42	58	0	54	88
22	L6	24	76	0	70	94
23 ^{h,j}	L6	27	73	0	58	90
24 ^{i,j}	L6	33	67	0	51	87

[a] The reaction was carried out with **1a** (0.1 mmol), **2** (0.11 mmol), CuCl (10 mol%), **L** (12 mol%), *t*-BuONa (15 mol%), and MeOH (2 equiv) in THF (2 mL) at room temperature under N₂ atmosphere.

[b] Yield of isolated yield.

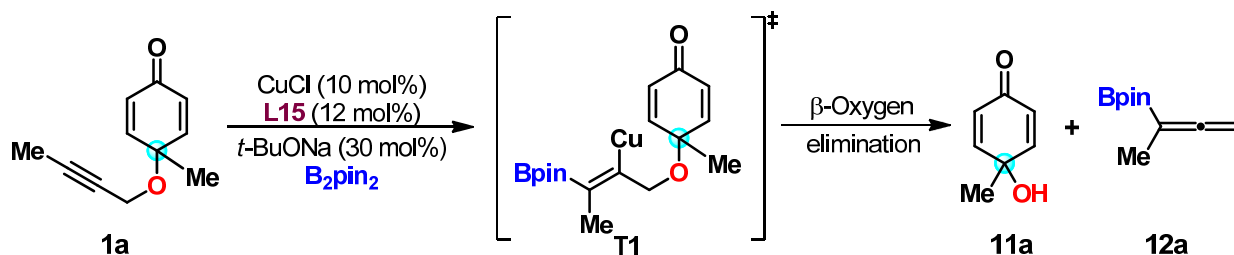
[c] Conversion was determined by the recovered **1a**.

[d] Determined by HPLC analysis using a chiral stationary phase.

[e] *t*-BuONa (30 mol%) was used.

[f] At 60°C.

[g] Using ICyHBF₄ (**L15**) as ligand, a side product **11a** was observed, which might be caused by the β-oxygen elimination of intermediate **T1**. (Scheme 4)

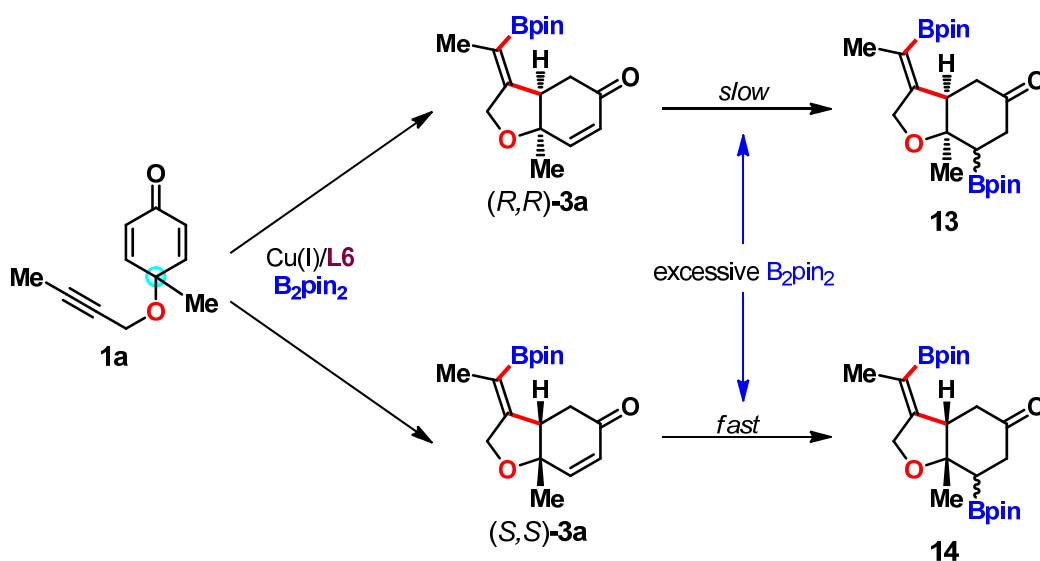


Scheme 4

[h] **B₂pin₂** (**2**, 1.2 equiv) was used.

[i] **B₂pin₂** (**2**, 1.3 equiv) was used.

[j] Increasing **B₂pin₂** (**2**) loading led to different levels of erosion in both yields and enantioselectivities (Table 3, entries 23-24), which might be caused by the further conjugate borylation of the excessive **B₂pin₂** to **3a**. (Scheme 5)

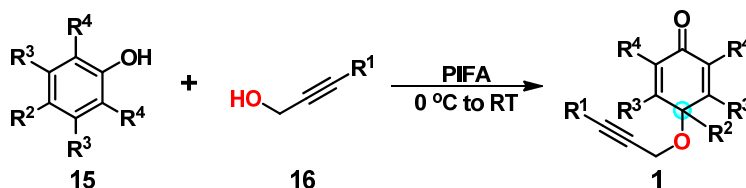


Scheme 5

Optimal Reaction Conditions: The reaction was carried out with **1a** (0.1 mmol), **2** (0.11 mmol), CuCl (10 mol%), **L6** (12 mol%), *t*-BuONa (15 mol%), and MeOH (2 equiv) in THF (2 mL) at room temperature under N₂ atmosphere.

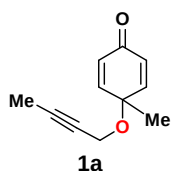
3. SUBSTRATE PREPARATION

3.1 General Procedure for Preparation of Substrates 1a-1c and 1g-1r.^[1]



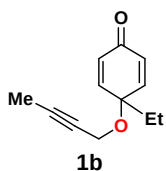
A well-stirred solution of substituted phenol **15** (1.0 mmol) in 1 mL of 3-substituted propargyl alcohol **16** was cooled to 0 °C and treated with phenyliodine(III)bis(trifluoroacetate) (PIFA, 516 mg, 1.2 mmol, 1.2 equiv, for substrates **1a**, **1b**, **1c**, **1g**, **1i**, **1j**, **1m**, **1n**, **1o**) or phenyliodine (III) diacetate (PIDA, 387 mg, 1.2 mmol, 1.2 equiv, for substrates **1h**, **1k**, **1l**, **1p**, **1q**, **1r**) in several portions. The resulting mixture was warmed to room temperature and stirred overnight. Then it was diluted with water (30 mL) and extracted with ethyl acetate (30 mL × 3). The combined organic phases were washed with brine (30 mL), dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography using hexane/ethyl acetate eluent to afford the desired product **1**.

4-(But-2-yn-1-yloxy)-4-methylcyclohexa-2,5-dienone (**1a**)^[1]



White solid. 76 mg, 43% yield. mp 81 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.83 (d, *J* = 10.4 Hz, 2H), 6.30 (d, *J* = 10.4 Hz, 2H), 3.96 (q, *J* = 2.4 Hz, 2H), 1.84 (t, *J* = 2.0 Hz, 3H), 1.48 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 185.05, 151.09, 130.38, 83.33, 75.58, 72.97, 54.37, 26.43, 3.73; ESI-MS: [M+Na]⁺ 199.1; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₁H₁₂O₂Na⁺ 199.0730, found 199.0732; IR (KBr) ν (cm⁻¹) 2975, 2928, 1665, 1630, 1380, 1309, 1081, 1033, 900, 865.

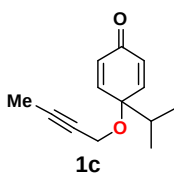
4-(But-2-yn-1-yloxy)-4-ethylcyclohexa-2,5-dienone (**1b**)^[1c]



Yellow solid. 57 mg, 30% yield. mp 61-62 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.77 (d, *J* = 10.4 Hz, 2H), 6.36 (d, *J* = 10.4 Hz, 2H), 3.98 (q, *J* = 2.4 Hz, 2H), 1.85-1.80 (m, 5H), 0.83 (t, *J* = 7.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 185.41, 150.29, 131.60, 83.17, 75.74, 54.22, 32.24, 7.83, 3.71; ESI-MS: [M+Na]⁺ 213.1; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₂H₁₄O₂Na⁺ 213.0886, found 213.0891; IR (KBr) ν (cm⁻¹) 3035, 2974, 2928, 1670, 1604, 1390, 1051, 924, 876,

809.

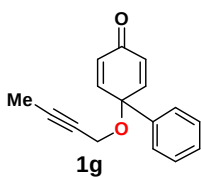
4-(But-2-yn-1-yloxy)-4-isopropylcyclohexa-2,5-dienone (**1c**)^[1c]



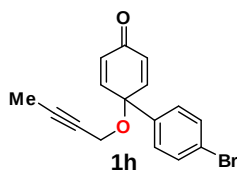
Yellow solid. 37 mg, 18% yield. mp 56 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.78 (d, *J* = 10.4 Hz, 2H), 6.39 (d, *J* = 10.4 Hz, 2H), 3.96 (q, *J* = 2.4 Hz, 2H), 2.06-2.03 (m, 1H), 1.84 (t, *J* = 2.4 Hz, 3H), 0.94 (d, *J* = 6.8 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 185.59, 149.51, 132.19, 82.97, 78.75, 76.00, 54.18, 36.59, 17.13, 3.76; ESI-MS: [M+Na]⁺ 227.1; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₃H₁₆O₂Na⁺ 227.1043, found 227.1049; IR (KBr) ν (cm⁻¹) 3045, 2997, 2968,

1661, 1622, 1388, 1276, 1066, 924, 862, 802.

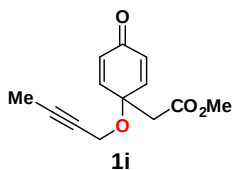
[1] (a) Tello-Aburto, R.; Harned, A. M. *Org. Lett.* **2009**, *11*, 3998. (b) Keilitz, J.; Newman, S. G.; Lautens, M. *Org. Lett.* **2013**, *15*, 1148. (c) He, Z.-T.; Tian, B.; Fukui, Y.; Tong, X.; Tian, P.; Lin, G.-Q. *Angew. Chem. Int. Ed.* **2013**, *52*, 5314.

1-(But-2-yn-1-yloxy)-[1,1'-biphenyl]-4(1H)-one (1g)^[1c,2]

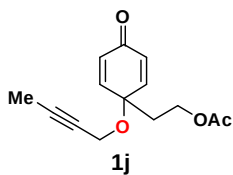
Red solid. 52 mg, 22% yield. mp 97 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.48 (d, *J* = 7.2 Hz, 2H), 7.38-7.30 (m, 3H), 6.88 (d, *J* = 10.4 Hz, 2H), 6.38 (d, *J* = 10.4 Hz, 2H), 4.20 (q, *J* = 2.4 Hz, 2H), 1.86 (t, *J* = 2.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 185.58, 149.96, 137.86, 129.88, 128.90, 128.53, 125.88, 83.47, 76.79, 75.86, 54.13, 3.78; ESI-MS: [M+Na]⁺ 261.1; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₆H₁₄O₂Na⁺ 261.0886, found 261.0890; IR (KBr) ν (cm⁻¹) 3065, 3039, 2925, 2863, 1668, 1489, 1448, 1279, 1051, 927, 753, 697.

4'-Bromo-1-(but-2-yn-1-yloxy)-[1,1'-biphenyl]-4(1H)-one (1h)

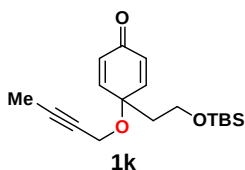
Light yellow solid. 30 mg, 10% yield. mp 126-127 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.48 (d, *J* = 8.8 Hz, 2H), 7.35 (d, *J* = 8.8 Hz, 2H), 6.82 (d, *J* = 10.0 Hz, 2H), 6.39 (d, *J* = 10.0 Hz, 2H), 4.19 (q, *J* = 2.4 Hz, 2H), 1.86 (t, *J* = 2.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 185.29, 149.38, 137.02, 132.01, 130.15, 127.67, 122.68, 83.73, 76.48, 75.67, 54.24, 3.79; ESI-MS: [M(C₁₆H₁₃⁷⁹BrO₂)+Na]⁺ 339.1; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₆H₁₃⁷⁹BrO₂Na⁺ 338.9997, found 338.9991; IR (KBr) ν (cm⁻¹) 3037, 2977, 2932, 2869, 2240, 1667, 1630, 1455, 1384, 1118, 1095, 1082, 867.

Methyl 2-(1-(but-2-yn-1-yloxy)-4-oxocyclohexa-2,5-dien-1-yl)acetate (1i)

White solid. 9 mg, 4% yield. mp 59-60 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.97 (d, *J* = 10.4 Hz, 2H), 6.37 (d, *J* = 10.4 Hz, 2H), 3.98 (q, *J* = 2.4 Hz, 2H), 3.66 (s, 3H), 2.78 (s, 2H), 1.83 (t, *J* = 2.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 184.88, 168.59, 148.37, 131.47, 83.70, 75.48, 73.12, 54.20, 52.07, 44.55, 3.73; ESI-MS: [M+Na]⁺ 257.0; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₃H₁₄O₄Na⁺ 257.0790, found 257.0784; IR (KBr) ν (cm⁻¹) 3084, 2988, 2919, 2954, 2868, 2320, 2239, 1732, 1704, 1666, 1625, 1460, 1440, 1390, 1371, 1357, 1290, 1244, 1185, 1166, 1095, 1062, 997, 873.

2-(1-(But-2-yn-1-yloxy)-4-oxocyclohexa-2,5-dien-1-yl)ethyl acetate (1j)^[1c]

Yellow oil. 62 mg, 25% yield. ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.83 (d, *J* = 10.0 Hz, 2H), 6.36 (d, *J* = 10.0 Hz, 2H), 4.15 (t, *J* = 6.4 Hz, 2H), 3.97 (q, *J* = 2.4 Hz, 2H), 2.12 (t, *J* = 6.8 Hz, 2H), 2.01 (s, 3H), 1.84 (t, *J* = 2.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 184.94, 170.69, 149.46, 131.33, 83.52, 75.55, 74.31, 59.40, 54.10, 38.62, 20.88, 3.70; ESI-MS: [M+Na]⁺ 271.2; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₄H₁₆O₄Na⁺ 271.0941, found 271.0946; IR (KBr) ν (cm⁻¹) 2983, 2930, 1690, 1671, 1630, 1525, 1368, 1254, 1166, 1053, 863, 779.

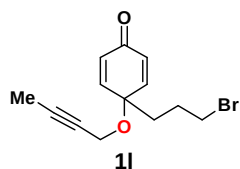
4-(But-2-yn-1-yloxy)-4-(2-((tert-butyldimethylsilyl)oxy)ethyl)cyclohexa-2,5-dienone (1k)

Light yellow solid. 44 mg, 14% yield. mp 70-72 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.86 (d, *J* = 10.0 Hz, 2H), 6.31 (d, *J* = 10.0 Hz, 2H), 3.95 (q, *J* = 2.0 Hz, 2H), 3.68 (t, *J* = 6.0 Hz, 2H), 1.99 (t, *J* = 6.0 Hz, 2H), 1.82 (t, *J* = 2.0 Hz, 3H), 0.85 (s, 9H), 0.00 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 185.40, 150.45, 130.60,

[2] J. K. Hexum, R. Tello-Aburto, N. B. Struntz, A. M. Harned, D. A. Harki, *ACS Med. Chem. Lett.* **2012**, 3, 459.

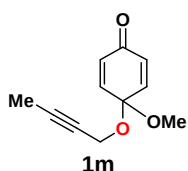
83.15, 75.70, 74.76, 57.84, 53.85, 42.86, 25.77, 18.07, 3.70, -5.50; ESI-MS: $[M+Na]^+$ 343.1; HRMS (FTMS-ESI): $[M+Na]^+$ calcd for $C_{18}H_{28}O_3SiNa^+$ 343.1705, found 343.1700; IR (KBr) ν (cm^{-1}) 3080, 3035, 2954, 2929, 2896, 2859, 2323, 2242, 1740, 1664, 1625, 1604, 1472, 1461, 1437, 1401, 1390, 1277, 1255, 1185, 1155, 1084, 1084, 1038, 1005, 924, 880.

4-(3-Bromopropyl)-4-(but-2-yn-1-yloxy)cyclohexa-2,5-dienone (1l)



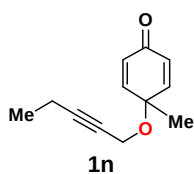
Light yellow solid. 42 mg, 15% yield. mp 71-73 °C; 1H NMR (400 MHz, $CDCl_3$) δ (ppm) 6.80 (d, J = 10.4 Hz, 2H), 6.37 (d, J = 10.4 Hz, 2H), 3.98 (q, J = 2.4 Hz, 2H), 3.37 (t, J = 12.8 Hz, 2H), 1.91-1.96 (m, 2H), 1.84-1.89 (m, 5H). ^{13}C NMR (100 MHz, $CDCl_3$) δ (ppm) 185.09, 149.83, 131.69, 83.49, 75.65, 75.47, 54.29, 38.21, 33.06, 26.94, 3.78; ESI-MS: $[M+Na]^+$ 305.1/307.1 (1:1); HRMS (FTMS-ESI): $[M+Na]^+$ calcd for $C_{13}H_{15}^{79}BrO_2Na^+$ 305.0153, found 305.0148; IR (KBr) ν (cm^{-1}) 3038, 2968, 2943, 2919, 2890, 2857, 2241, 1694, 1672, 1629, 1604, 1450, 1385, 1285, 1176, 1062, 865.

4-(But-2-yn-1-yloxy)-4-methoxycyclohexa-2,5-dienone (1m)



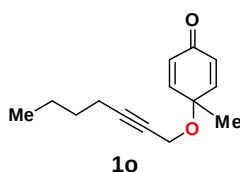
Light yellow Oil. 8 mg, 4% yield. 1H NMR (400 MHz, $CDCl_3$) δ (ppm) 6.87 (d, J = 10.0 Hz, 2H), 6.26 (d, J = 10.0 Hz, 2H), 4.27 (q, J = 2.0 Hz, 2H), 3.40 (s, 3H), 1.85 (t, 3H, J = 2.4 Hz). ^{13}C NMR (100 MHz, $CDCl_3$) δ (ppm) 185.15, 142.83, 129.76, 92.66, 83.22, 75.11, 51.49, 50.70, 3.72. ESI-MS: $[M+Na]^+$ 215.1; HRMS (FTMS-ESI): $[M+Na]^+$ calcd for $C_{11}H_{12}O_3Na^+$ 215.0684, found 215.0679; IR (KBr) ν (cm^{-1}) 3055, 2942, 2922, 2834, 2305, 2239, 1670, 1640, 1617, 1459, 1386, 1317, 1247, 1181, 1155, 1104, 1063, 1027, 967, 952, 915, 857.

4-Methyl-4-(pent-2-yn-1-yloxy)cyclohexa-2,5-dienone (1n)



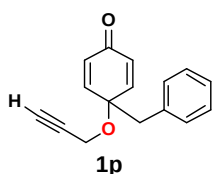
White Solid. 80 mg, 42% yield. mp 53 °C; 1H NMR (400 MHz, $CDCl_3$) δ (ppm) 6.84 (d, J = 10.0 Hz, 2H), 6.30 (d, J = 10.0 Hz, 2H), 3.98 (t, J = 2.0 Hz, 2H), 2.19-2.25 (m, 2H), 1.13 (t, J = 7.6 Hz, 3H). ^{13}C NMR (100 MHz, $CDCl_3$) δ (ppm) 185.14, 151.21, 130.38, 89.13, 75.84, 73.03, 54.44, 26.48, 13.61, 12.58; ESI-MS: $[M+Na]^+$ 213.0; HRMS (FTMS-ESI): $[M+Na]^+$ calcd for $C_{12}H_{14}O_2Na^+$ 213.0891, found 213.0886; IR (KBr) ν (cm^{-1}) 3068, 3041, 2926, 2866, 2241, 1670, 1627, 1487, 1389, 1052, 1023, 1005, 869.

4-(Hept-2-yn-1-yloxy)-4-methylcyclohexa-2,5-dienone (1o)



Yellow solid. 46 mg, 21% yield. mp 43-44 °C; 1H NMR (400 MHz, $CDCl_3$) δ (ppm) 6.84 (d, J = 10.0 Hz, 2H), 6.29 (d, J = 10.0 Hz, 2H), 3.99 (t, J = 2.0 Hz, 2H), 2.22-2.18 (m, 2H), 1.50-1.36 (m, 7H), 0.90 (t, J = 7.2 Hz, 3H). ^{13}C NMR (100 MHz, $CDCl_3$) δ (ppm) 185.16, 151.27, 130.35, 87.91, 76.55, 73.07, 54.48, 30.57, 26.50, 22.01, 18.59, 13.61; ESI-MS: $[M+Na]^+$ 241.1; HRMS (FTMS-ESI): $[M+Na]^+$ calcd for $C_{14}H_{18}O_2Na^+$ 241.1199, found 241.1203; IR (KBr) ν (cm^{-1}) 3036, 2960, 2930, 1709, 1664, 1626, 1456, 1398, 1195, 1083, 878.

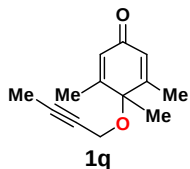
4-Benzyl-4-(prop-2-yn-1-yloxy)cyclohexa-2,5-dienone (1p)



Light yellow solid. 76 mg, 32% yield. mp 94-95 °C; 1H NMR (400 MHz, $CDCl_3$) δ (ppm) 7.23-7.28 (m, 2H), 7.14-7.17 (m, 2H), 6.79 (d, J = 10.0 Hz, 2H), 6.28 (d, J = 10.0 Hz, 2H), 4.00 (d, J = 2.8 Hz, 2H), 3.06 (s, 2H), 2.45 (t, J = 2.4 Hz, 1H). ^{13}C

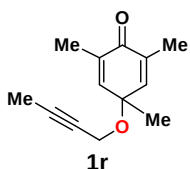
NMR (100 MHz, CDCl₃) δ (ppm) 185.01, 149.54, 134.44, 131.55, 130.78, 128.12, 127.26, 80.45, 76.37, 75.01, 53.80, 46.31; ESI-MS: [M+Na]⁺ 261.1; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₆H₁₄O₂Na⁺ 261.0886, found 261.0876; IR (KBr) ν (cm⁻¹) 3259, 3026, 2946, 2922, 2903, 2860, 2126, 1699, 1668, 1629, 1609, 1494, 1453, 1398, 1386, 1374, 1256, 1085, 1050, 959, 871.

4-(But-2-yn-1-yloxy)-3,4,5-trimethylcyclohexa-2,5-dienone (**1q**)



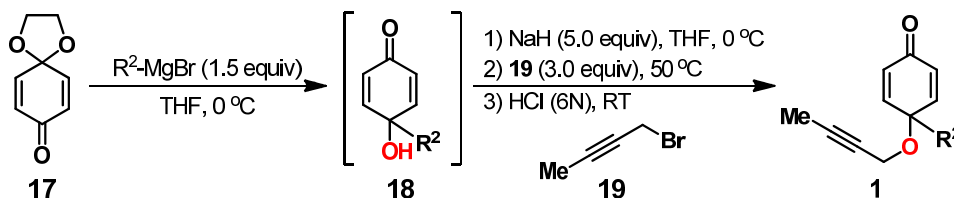
White solid. 64 mg, 31% yield. mp 60-61 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.17 (s, 2H), 3.65 (q, *J* = 2.0 Hz, 2H), 2.05 (s, 6H), 1.85 (t, *J* = 2.0 Hz, 3H), 1.45 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 185.12, 160.23, 129.35, 82.99, 77.04, 74.62, 53.58, 25.11, 18.01, 3.76; ESI-MS: [M+Na]⁺ 227.2; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₃H₁₆O₂Na⁺ 227.1048, found 227.1043; IR (KBr) ν (cm⁻¹) 3252, 3049, 2998, 2980, 2955, 2918, 2902, 2863, 2233, 1675, 1638, 1623, 1444, 1386, 1307, 1173, 1083, 1036, 902, 890.

4-(But-2-yn-1-yloxy)-2,4,6-trimethylcyclohexa-2,5-dienone (**1r**)



Colorless Oil. 87 mg, 43% yield. ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.55 (s, 2H), 3.91 (q, *J* = 2.4 Hz, 2H), 1.91 (s, 6H), 1.84 (t, *J* = 2.4 Hz, 3H), 1.42 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 186.57, 146.19, 136.77, 82.89, 76.02, 72.89, 53.79, 26.70, 16.02, 3.81; ESI-MS: [M+Na]⁺ 227.1; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₃H₁₆O₂Na⁺ 227.1048, found 227.1043; IR (KBr) ν (cm⁻¹) 3272, 2978, 2923, 2857, 2242, 1675, 1644, 1447, 1372, 1074, 1037, 905.

3.2 General Procedure for Preparation of Substrates **1d-1f**.

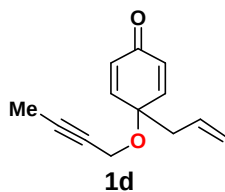


To a well-stirred solution of 1,4-dioxaspiro[4.5]deca-6,9-dien-8-one **17**^[3] (1.0 mmol) in THF (1 mL) was added Grignard reagent (1.5 mmol, 1.5 equiv) dropwise at 0 °C under argon atmosphere. The resulting mixture was stirred for 15 min, then quenched by water (50 mL) and extracted with DCM (50 mL $\times$ 3). The combined organic phases were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure to give a crude product **18**, which was used in next step without further purification.

To a solution of the above crude product **18** in THF (10 mL) was added NaH (60% in Mineral oil, 5.0 mmol, 5.0 equiv) in several portions at 0 °C under argon atmosphere, followed by the addition of 1-bromo-2-butyne (**19**, 3.0 mmol, 3.0 equiv). The resulting mixture was heated at 50 °C overnight. The mixture was quenched by water (10 mL) at 0 °C and subsequently acidified with 6N HCl (0.9 mL). The resulting mixture was stirred at room temperature for 2 h to hydrolyze the ketal. Then it was extracted with DCM (50 mL $\times$ 3). The combined organic phases were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography using hexane/ethyl acetate eluent to afford the pure substrates **1**.

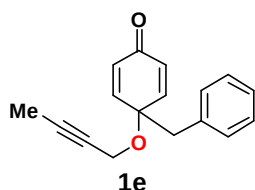
[3] J. Wegner, S. V. Ley, A. Kirschning, A.-L. Hansen, J. M. Garcia, I. R. Baxendale, *Org. Lett.* **2012**, *14*, 696.

4-Allyl-4-(but-2-yn-1-yloxy)cyclohexa-2,5-dienone (1d)



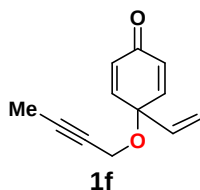
Colorless oil, 54.7 mg, 27% yield (3 steps). ^1H NMR (400 MHz, CDCl_3) δ (ppm) 6.79 (d, $J = 10.4$ Hz, 2H), (6.34 d, $J = 10.4$ Hz, 2H), 5.62-5.73 (m, 1H), 5.04-5.12 (m, 2H), 3.99 (q, $J = 2.0$ Hz, 2H), 2.53 (d, $J = 7.2$ Hz, 2H), 1.84 (t, $J = 2.0$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ (ppm) 185.25, 149.96, 131.39, 130.84, 119.80, 83.39, 75.68, 75.60, 54.31, 43.92, 3.72; ESI-MS: $[\text{M}+\text{Na}]^+$ 225.0; HRMS (FTMS-ESI): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{13}\text{H}_{14}\text{O}_2\text{Na}^+$ 225.0891, found 225.0886; IR (KBr) ν (cm^{-1}) 3078, 2979, 2920, 2859, 2230, 1670, 1630, 1507, 1440, 1381, 1254, 1198, 1173, 1157, 1142, 1051, 997, 942, 861.

4-Benzyl-4-(but-2-yn-1-yloxy)cyclohexa-2, 5-dienone (1e)



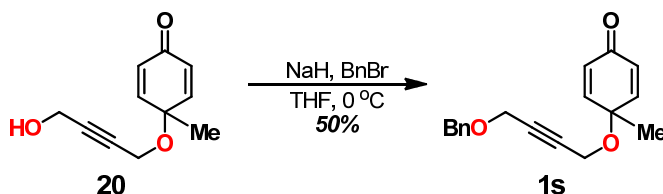
White solid, 47.9 mg, 19% yield (3 steps). mp 59-60 $^\circ\text{C}$; ^1H NMR (400 MHz, CDCl_3) δ (ppm) 7.27-7.12 (m, 5H), 6.79 (d, $J = 10.0$ Hz, 2H), 6.25 (d, $J = 10.0$ Hz, 2H), 3.97 (q, $J = 2.0$ Hz, 2H), 3.06 (s, 2H), 1.84 (t, $J = 2.0$ Hz, 3H), ^{13}C NMR (100 MHz, CDCl_3) δ (ppm) 185.09, 149.92, 134.52, 131.28, 130.73, 128.04, 127.17, 83.32, 76.15, 75.81, 54.41, 46.34, 3.75; ESI-MS: $[\text{M}+\text{Na}]^+$ 275.0; HRMS (FTMS-ESI): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{17}\text{H}_{17}\text{O}_2^+$ 253.1223, found 253.1228; IR (KBr) ν (cm^{-1}) 3062, 3030, 2920, 2858, 1670, 1629, 1495, 1455, 1382, 1049, 860, 770, 704.

4-(But-2-yn-1-yloxy)-4-vinylcyclohexa-2, 5-dienone (1f)



Pale yellow solid, 65.9 mg, 35% yield (3 steps). mp 52-54 $^\circ\text{C}$; ^1H NMR (400 MHz, CDCl_3) δ (ppm) 6.81 (d, $J = 10.0$ Hz, 2H), 6.34 (d, $J = 10.0$ Hz, 2H), 5.76 (dd, $J=10.4, 17.6$ Hz, 1H), 5.44 (d, $J = 17.6$ Hz, 1H), 5.27 (d, $J = 10.4$ Hz, 1H), 4.07 (q, $J = 2.4$ Hz, 2H), 1.84 (t, $J = 2.4$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ (ppm) 185.20, 148.71, 135.92, 130.43, 117.44, 83.53, 75.77, 75.65, 54.03, 3.74; ESI-MS: $[\text{M}+\text{Na}]^+$ 211.1; HRMS (FTMS-ESI): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{12}\text{H}_{12}\text{O}_2\text{Na}^+$ 211.0730, found 211.0733; IR (KBr) ν (cm^{-1}) 3042, 2975, 2923, 2865, 1670, 1627, 1507, 1387, 1283, 1050, 926, 878, 691.

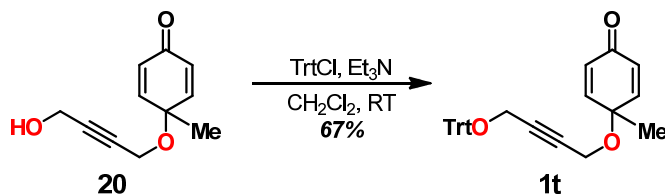
3.3 Preparation of Substrates 1s and 1t.



4-((4-(Benzyloxy)but-2-yn-1-yl)oxy)-4-methylcyclohexa-2,5-dienone (1s) To a well-stirred solution of alcohol **20**^[4] (1.5 g, 7.8 mmol) in dry THF (30 mL) was added NaH (60% in mineral oil, 1.6 g, 39.0 mmol, 5.0 equiv) in several portions at 0 $^\circ\text{C}$ under argon atmosphere. Then benzyl bromide (2.7 g, 15.6 mmol, 2 equiv) was added. The resulting mixture was warmed to room temperature and stirred for 2 h. Then it was diluted with water (30 mL) and extracted with ether (30 mL $\times$ 3). The combined organic phases were dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The residue was purified by flash column chromatography using hexane/ethyl acetate eluent to afford the desired product **1s** (1.1 g, 50% yield) as colorless oil. ^1H NMR (400 MHz, CDCl_3) δ (ppm) 7.36-7.25 (m, 5H), 6.82 (d, $J = 10.0$ Hz, 2H),

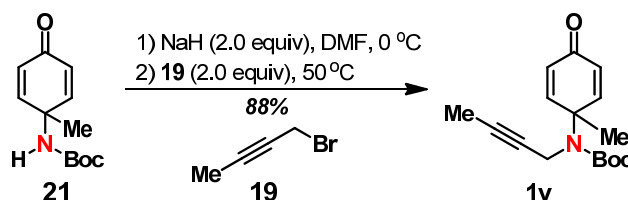
[4] S.-Y. Cai, Z. Liu, W.-B. Zhang, X.-Y. Zhao, D. Z. Wang, *Angew. Chem. Int. Ed.* **2011**, *50*, 11133.

6.30 (d, $J = 10.0$ Hz, 2H), 4.56 (s, 2H), 4.19 (s, 2H), 4.05 (s, 2H), 1.47 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ (ppm) 184.84, 150.70, 137.28, 130.43, 128.39, 127.93, 127.84, 83.01, 82.89, 73.12, 71.63, 57.38, 53.94, 26.29; ESI-MS: $[\text{M}+\text{Na}]^{\oplus}$ 305.1; HRMS (FTMS-ESI): $[\text{M}+\text{Na}]^{\oplus}$ calcd for $\text{C}_{18}\text{H}_{18}\text{O}_3\text{Na}^{\oplus}$ 305.1148, found 305.1136; IR (KBr) ν (cm^{-1}) 3032, 2981, 2930, 2856, 1671, 1630, 1497, 1454, 1383, 1352, 1300, 1242, 1180, 1125, 1076, 1032, 861, 738, 698.



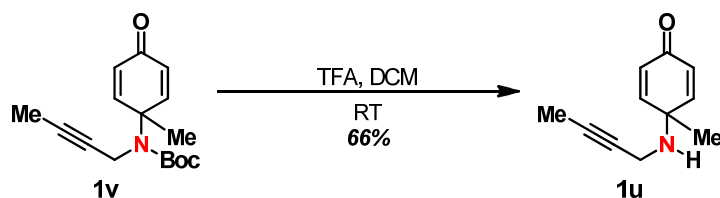
4-Methyl-4-((4-(trityloxy)but-2-yn-1-yl)oxy)cyclohexa-2,5-dienone (1t) A solution of alcohol **20**^[4] (100 mg, 0.52 mmol), trityl chloride (189 mg, 0.68 mmol, 1.3 equiv.) and Et_3N (79 mg, 0.78 mmol, 1.5 equiv.) in DCM (30 mL) was stirred for 2 d at room temperature. Then it was diluted with water (30 mL) and extracted with DCM (30 mL $\times$ 3). The combined organic phases were dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The residue was purified by flash column chromatography using hexane/ethyl acetate eluent to afford the desired product **1t** (151 mg, 67% yield) as colorless oil. ^1H NMR (400 MHz, CDCl_3) δ (ppm) 6.47-7.42 (m, 6H), 7.33-7.21 (m, 9H), 6.83 (d, $J = 10.0$ Hz, 2H), 6.30 (d, $J = 10.0$ Hz, 2H), 4.02 (s, 2H), 3.78 (s, 2H), 1.47 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ (ppm) 185.09, 150.97, 143.45, 130.12, 128.67, 128.02, 127.28, 87.51, 83.73, 81.99, 73.24, 54.23, 53.15, 26.45; ESI-MS: $[\text{M}+\text{Na}]^{\oplus}$ 457.1; HRMS (FTMS-ESI): $[\text{M}+\text{Na}]^{\oplus}$ calcd for $\text{C}_{30}\text{H}_{26}\text{O}_3\text{Na}^{\oplus}$ 457.1774, found 457.1762; IR (KBr) ν (cm^{-1}) 3084, 3063, 2983, 2912, 2856, 1673, 1630, 1489, 1448, 1383, 1368, 1184, 1164, 1058, 1029, 900, 864, 746, 704, 632.

3.4 Preparation of Substrates 1u, 1v and 1w.

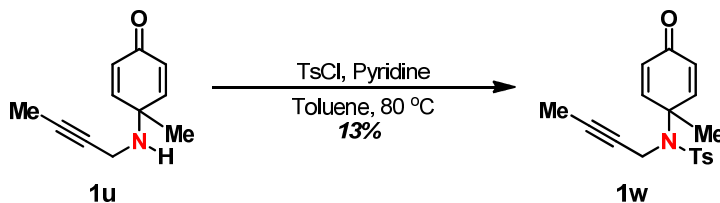


tert-Butyl but-2-yn-1-yl(1-methyl-4-oxocyclohexa-2,5-dien-1-yl)carbamate (1v) To a well-stirred solution of substrate **21**^[5] (3.3 g, 14.8 mmol) in DMF (30 mL) was added NaH (60% in mineral oil, 1182 mg, 29.6 mmol, 2 equiv.) in several portions at 0 °C under argon atmosphere. Then 1-bromo-2-butyne (**19**, 3931 mg, 29.6 mmol, 2 equiv.) was added and the resulting mixture was stirred at 0 °C for another 1 min. The reaction was quenched by sat. NH_4Cl aq. (50 mL) and extracted with AcOEt (100 mL $\times$ 3). The combined organic phases were dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The residue was purified by flash column chromatography using hexane/ethyl acetate eluent to afford the pure **1v** (3.575 g, 88% yield) as pale yellow oil. ^1H NMR (400 MHz, CDCl_3) δ (ppm) 7.07 (d, $J = 10.0$ Hz, 2H), 6.16 (d, $J = 10.4$ Hz, 2H), 4.16 (d, $J = 2.4$ Hz, 2H), 1.83 (t, $J = 2.4$ Hz, 3H), 1.69 (s, 3H), 1.38 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3) δ (ppm) 185.30, 154.96, 153.92, 126.63, 81.71, 79.58, 76.07, 57.48, 34.39, 28.13, 26.19, 3.56; ESI-MS: $[\text{M}+\text{Na}]^{\oplus}$ 298.1; HRMS (FTMS-ESI): $[\text{M}+\text{Na}]^{\oplus}$ calcd for $\text{C}_{16}\text{H}_{21}\text{NO}_3\text{Na}^{\oplus}$ 298.1414, found 298.1425; IR (KBr) ν (cm^{-1}) 3042, 2979, 2923, 1698, 1668, 1629, 1605, 1456, 1432, 1390, 1367, 1281, 1246, 1162, 1127, 1088, 1031, 874, 854.

[5] Carreño, M. C.; Luzón, C. G.; Ribagorda, M. *Chem. Eur. J.* **2002**, *8*, 208.

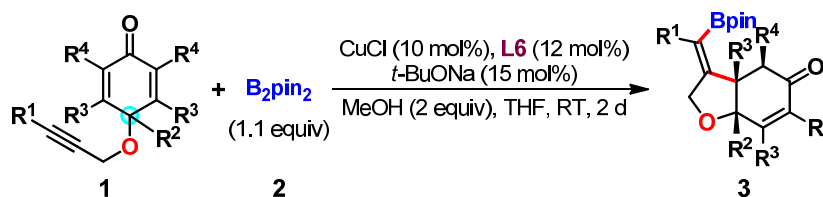


4-(But-2-ynylamino)-4-methylcyclohexa-2,5-dienone (1u). To a well-stirred solution of **1v** (2.5 g, 9.1 mmol) in DCM (45 mL) was added TFA (4.5 mL). The resulting mixture was stirred for 5 h at room temperature, and then concentrated under reduced pressure. The residue was diluted with sat. Na₂CO₃ aq. (30 mL) and extracted with AcOEt (30 mL × 4). The combined organic phases were dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography using hexane/ethyl acetate eluent to afford the pure **1u** (1.045 g, 66% yield) as pale yellow solid. mp 58-59 °C; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.78 (d, *J* = 10.0 Hz, 2H), 6.27 (d, *J* = 10.0 Hz, 2H), 3.20-3.15 (m, 2H), 1.81-1.78 (m, 3H), 1.67 (br, 1H), 1.37 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 185.49, 153.99, 129.48, 79.91, 77.19, 55.27, 34.21, 26.67, 3.47; ESI-MS: [M+Na]⁺ 198.2; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₁H₁₃NONa⁺ 198.0889, found 198.0880; IR (KBr) ν (cm⁻¹) 3279, 2976, 2804, 1663, 1621, 1497, 1397, 1383, 1293, 1182, 1110, 1092, 861, 827, 789, 704, 469.



N-(But-2-ynyl)-4-methyl-N-(1-methyl-4-oxocyclohexa-2,5-dienyl)benzenesulfonamide (1w) To a well-stirred solution of pyridine (911 mg, 11.41 mmol, 4 equiv.) and **1u** (500 mg, 2.85 mmol) in toluene (30 mL) was added TsCl (1632 mg, 8.56 mmol, 3 equiv) at room temperature. The resulting mixture was heated to 80 °C overnight. Then the mixture was quenched by HCl (2N, 20 mL) at room temperature and subsequently extracted with AcOEt (30 mL × 2). The combined organic phases were washed with sat. NaHCO₃, dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography using hexane/ethyl acetate eluent to afford the pure **1w** (121.3 mg, 13% yield) as pale yellow oil. ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.79 (d, *J* = 8.4 Hz, 2H), 7.29 (d, *J* = 8.4 Hz, 2H), 7.03 (d, *J* = 10.0 Hz, 2H), 6.13 (d, *J* = 10.0 Hz, 2H), 4.25-4.24 (m, 2H), 2.43 (s, 3H), 1.78-1.76 (m, 3H), 1.61 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 184.54, 151.72, 143.78, 138.38, 129.36, 127.82, 127.48, 81.70, 75.43, 36.77, 26.08, 21.48, 3.43; ESI-MS: [M+Na]⁺ 352.2; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₈H₁₉NO₃SN⁺ 352.0978, found 352.09736; IR (KBr) ν (cm⁻¹) 2921, 1670, 1629, 1598, 1448, 1393, 1334, 1247, 1189, 1160, 1091, 891, 862, 812, 812, 706, 688, 591, 574, 549.

4. SCOPE OF THE SUBSTRATES



For **1a** to **1m**: R¹ = Me-, R³ = R⁴ = H

1a: R² = Me-

1f: R² = CH₂=CH-

1k: R² = TBSO-(CH₂)₂-

For **1n** to **1p**: R³ = R⁴ = H

1n: R¹ = Et-, R² = Me-

1b: R² = Et-

1g: R² = Ph-

1l: R² = Br-(CH₂)₃-

1o: R¹ = *n*-Bu-, R² = Me-

1c: R² = *i*Pr-

1h: R² = 4-Br-Ph-

1m: R² = MeO-

1p: R¹ = H-, R² = PhCH₂-

1d: R² = CH₂=CH-CH₂-

1i: R² = MeOC(O)-CH₂-

For **1q** to **1r**: R¹ = R² = Me-

1q: R³ = Me-, R⁴ = H-

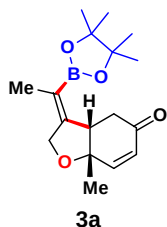
1e: R² = PhCH₂-

1j: R² = AcO-(CH₂)₂-

1r: R³ = H-, R⁴ = Me-

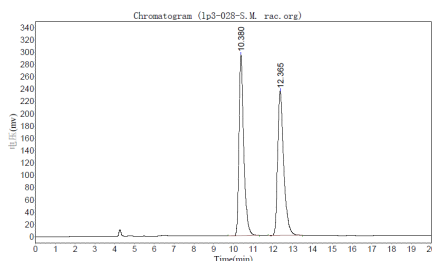
GENERAL PROCEDURE: A dried Schlenk flask was charged with CuCl (1.5 mg, 0.015 mmol, 10 mol%), ligand **L6** (9.7 mg, 0.018 mmol, 12 mol%), B₂pin₂ (**2**, 41.9 mg, 0.165 mmol, 1.1 equiv; except for substrate **1j**, 1.2 equiv of **2**; for substrates **1l**, **1m** and **1o**, 1.3 equiv of **2**), *t*-BuONa (2.2 mg, 0.0225 mmol, 15 mol%) and anhydrous THF (2 mL) under nitrogen atmosphere. After the mixture was stirred at room temperature for 30 min, a solution of cyclohexadienone-containing 1,6-enyne substrate **1** (0.15 mmol) in anhydrous THF (1.5 mL) was added, followed by anhydrous MeOH (12.2 μL, 0.30 mmol, 2.0 equiv). The resulting mixture was stirred at room temperature for 2 days. The reaction mixture was quenched with 1 N NaHCO₃ (5 mL), and extracted with EtOAc (15 mL × 3). The combined organic phases were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by silica gel (300-400 mesh) column chromatography to afford the desired product **3**.

(3a*S*,7a*S*,*Z*)-7a-Methyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3a)

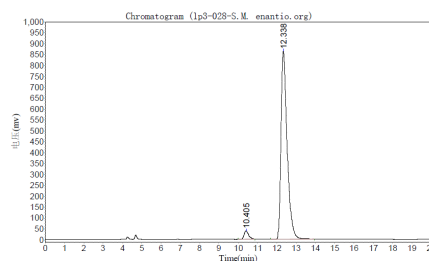


White solid. 32.0 mg, 70% yield; mp 73-75 °C; [α]_D²⁵ -91.2 (c 0.50, CHCl₃) for 94% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.63 (d, *J* = 10.4 Hz, 1H), 6.00 (d, *J* = 10.4 Hz, 1H), 4.49 (s, 2H), 3.36 (t, *J* = 7.6 Hz, 1H), 2.55-2.67 (m, 2H), 1.63 (s, 3H), 1.38 (s, 3H), 1.28 (s, 6H), 1.27 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 199.04, 157.23, 149.01, 129.72, 83.47, 79.04, 69.83, 47.57, 39.75, 25.03, 24.88, 24.87, 16.95; ESI-MS: [M+Na]⁺ 327.1; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₇H₂₅¹⁰BO₄Na⁺ 326.1780, found 326.1774; IR (KBr) ν (cm⁻¹) 2975, 2929, 2862,

1682, 1656, 1367, 1306, 1218, 1148, 1018, 890; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 10.4 min (*R*, *R*-isomer), 12.3 min (*S*,*S*-isomer).

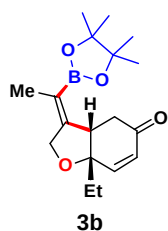


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		10.380	294853.906	4947239.000
2		12.365	234990.703	4980714.500
Total			529844.609	9927953.500



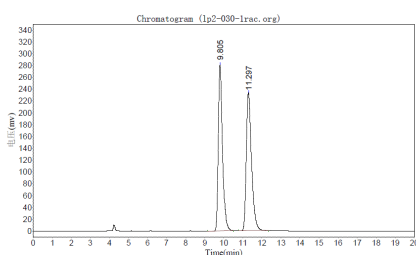
Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		10.405	37951.035	614142.250
2		12.338	863158.875	19085218.000
Total			901109.910	19699360.250

(3a*S*,7a*S*,*Z*)-7a-Ethyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3b)

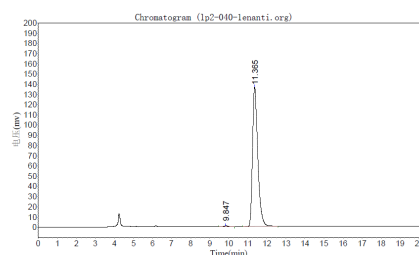


White solid. 33.6 mg, 70% yield; mp 74-75 °C; $[\alpha]_D^{25}$ -94.8 (c 0.29, CHCl₃) for 99% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.66 (d, *J* = 10.4 Hz, 1H), 6.07 (d, *J* = 10.4 Hz, 1H), 4.47 (AB, *J*_{AB} = 14.8 Hz, 2H), 3.46 (t, *J* = 7.6 Hz, 1H), 2.60 (t, *J* = 7.6 Hz, 2H), 1.65-1.71 (m, 2H), 1.62 (s, 3H), 1.28 (s, 6H), 1.26 (s, 6H), 0.93 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 199.21, 157.66, 147.80, 130.72, 83.45, 81.64, 69.69, 45.13, 40.04, 30.29, 24.98, 24.94, 16.93, 8.35; ESI-MS: [M+Na]⁺ 341.0; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₈H₂₇¹⁰BO₄Na⁺

340.1936, found 340.1931; IR (KBr) ν (cm⁻¹) 2970, 2927, 2857, 1690, 1653, 1460, 1364, 1313, 1208, 1144, 1023, 848; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 9.8 min (*R*, *R*-isomer), 11.4 min (*S*,*S*-isomer).

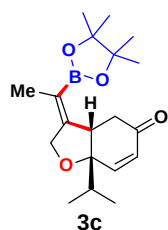


Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		9.805	281324.813	4308706.500	49.2042
2		11.297	233794.516	4448080.000	50.7958
Total			515119.328	8756786.500	100.0000



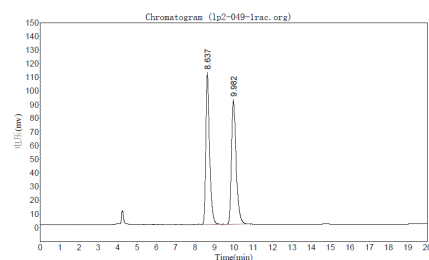
Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		9.847	909.089	14255.102	0.5423
2		11.365	136688.750	2614560.750	99.4577
Total			137597.839	2628815.852	100.0000

(3a*S*,7a*S*,*Z*)-7a-Isopropyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3c)

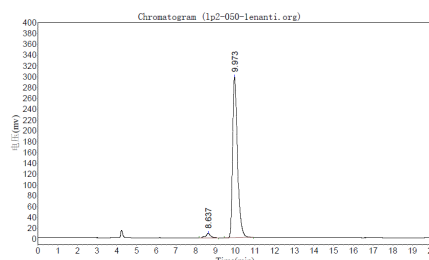


White solid. 32.6 mg, 65% yield; mp 104-106 °C; $[\alpha]_D^{25}$ -111.0 (c 0.50, CHCl₃) for 95% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.67 (d, *J* = 10.4 Hz, 1H), 6.12 (d, *J* = 10.4 Hz, 1H), 4.40-4.48 (m, 2H), 3.60 (t, *J* = 7.6 Hz, 1H), 2.54-2.66 (m, 2H), 1.89-2.00 (m, 1H), 1.62 (s, 3H), 1.28 (s, 6H), 1.26 (s, 6H), 1.04 (d, *J* = 6.8 Hz, 3H), 0.88 (d, *J* = 6.8 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 199.34, 158.38, 145.93, 131.48, 83.82, 83.42, 69.35, 43.91, 40.89, 33.72, 24.97, 24.94, 17.81, 16.92, 16.55; ESI-MS: [M+Na]⁺ 355.3; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₉H₂₉¹⁰BO₄Na⁺

354.2093, found 354.2087; IR (KBr) ν (cm⁻¹) 2961, 2930, 2874, 2856, 1730, 1690, 1652, 1468, 1363, 1305, 1210, 1144, 1027, 965, 849; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 8.6 min (*R*, *R*-isomer), 10.0 min (*S*,*S*-isomer).

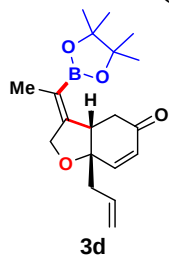


Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		8.637	109802.844	1481303.875	49.4667
2		9.982	89955.102	1513241.250	50.5333
Total			199757.945	2994545.125	100.0000



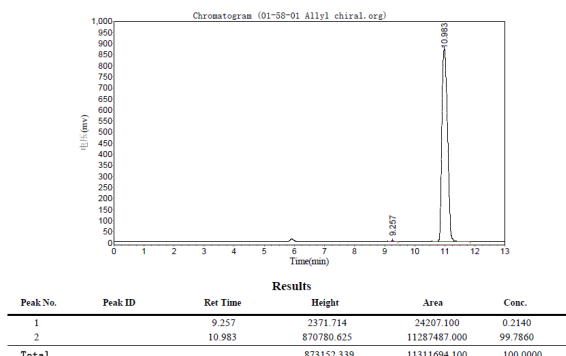
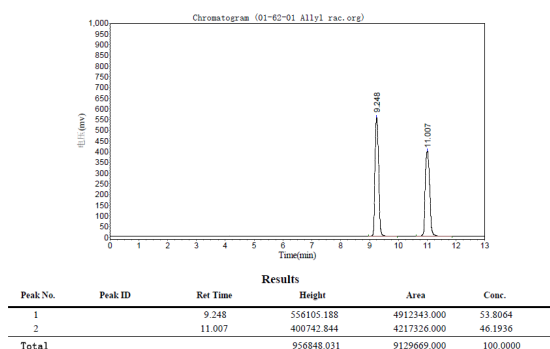
Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		8.637	8451.410	143471.906	2.7453
2		9.973	296869.750	5082527.000	97.2546
Total			305321.160	5225998.906	100.0000

(3a*S*,7a*S*,*Z*)-7a-allyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3d)

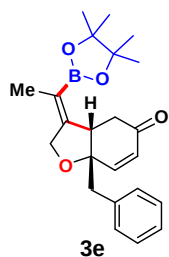


3d

Colorless oil. 29.7 mg, 60% yield; $[\alpha]_D^{25}$ -85.1 (*c* 0.52, CHCl₃) for >99% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.68 (d, *J* = 10.0 Hz, 1H), 6.05 (d, *J* = 10.0 Hz, 1H), 5.84-5.70 (m, 1H), 5.17-5.08 (m, 2H), 4.51 (AB, *J*_{AB} = 14.8 Hz, 2H), 3.51 (t, *J* = 8.0 Hz, 1H), 2.65-2.52 (m, 2H), 2.50-2.43 (m, 1H), 2.38-2.30 (m, 1H), 1.63 (s, 3H), 1.28 (s, 6H), 1.26 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 198.97, 157.12, 147.35, 132.50, 130.62, 119.19, 83.44, 80.71, 69.83, 45.54, 42.25, 39.80, 24.92, 24.90, 16.88; ESI-MS: $[M+H]^+$ 331.1; HRMS (FTMS-ESI): $[M+Na]^+$ calcd for C₁₉H₂₇BO₄Na⁺ 352.1931, found 352.1934; IR (KBr) ν (cm⁻¹) 2978, 2967, 2853, 1691, 165.4, 1364, 1307, 1274, 1212, 1145, 1094, 1022, 968, 928, 849, 786, 685; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 9.3 min (*R*, *R*-isomer), 11.0min (*S*,*S*-isomer).

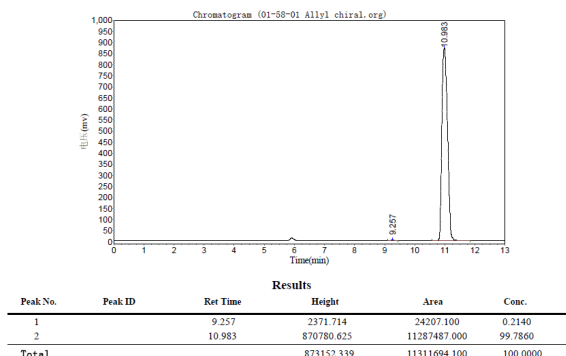
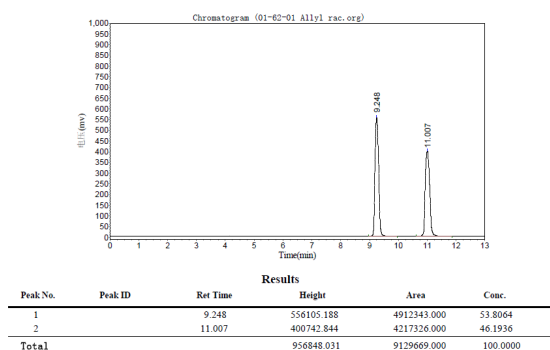


(3a*S*,7a*S*,*Z*)-7a-Benzyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3e)

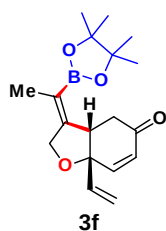


3e

White solid, 32.2 mg, 58% yield; mp 92-94 °C; $[\alpha]_D^{25}$ -126.1 (*c* 0.64, CHCl₃) for >99% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.30-7.16 (m, 5H), 6.65 (d, *J* = 10.0 Hz, 1H), 5.99 (d, *J* = 10.0 Hz, 1H), 4.57 (AB, *J*_{AB} = 14.8 Hz, 2H), 3.61 (t, *J* = 7.6 Hz, 1H), 3.01 (d, *J* = 13.6 Hz, 1H), 2.87 (d, *J* = 13.6 Hz, 1H), 2.59-2.42 (m, 2H), 1.63 (s, 3H), 1.28 (s, 6H), 1.27 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 198.91, 157.08, 147.36, 135.91, 130.56, 130.50, 128.29, 126.90, 83.44, 81.58, 69.93, 45.47, 44.08, 39.73, 24.04, 16.91; ESI-MS: $[M+H]^+$ 381.2; HRMS (FTMS-ESI): $[M+H]^+$ calcd for C₂₃H₃₀BO₄⁺ 380.2268, found 380.2278; IR (KBr) ν (cm⁻¹) 3028, 2976, 2922, 2857, 1683, 1496, 1455, 1366, 1224, 1146, 1112, 1089, 1013, 938, 849, 700, 687; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 11.2 min (*R*, *R*-isomer), 13.3min (*S*,*S*-isomer).

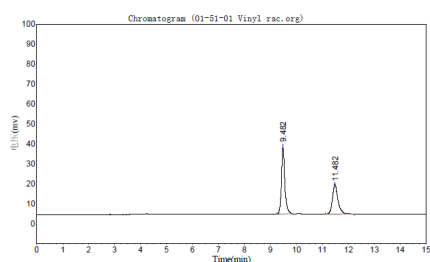


(3a*S*,7a*S*,*Z*)-3-(1-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-7a-vinyl-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3f)

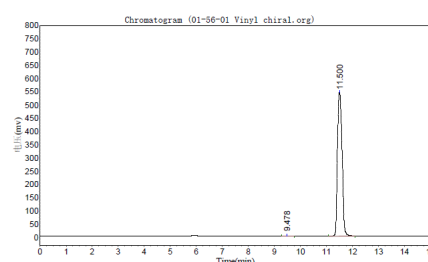


White solid. 29.9 mg, 63% yield; mp 75-77 °C; $[\alpha]_D^{25}$ -142.4 (*c* 0.78, CHCl₃) for >99% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.56 (d, *J* = 10.0 Hz, 1H), 6.07 (d, *J* = 10.0 Hz, 1H), 5.85 (dd, *J* = 10.8, 17.4 Hz, 1H), 5.36 (d, *J* = 17.6 Hz, 1H), 5.27 (d, *J* = 10.8 Hz, 1H), 4.50 (AB, *J*_{AB} = 14.8 Hz, 2H), 3.55 (t, *J* = 7.6 Hz, 1H), 2.70-2.57 (m, 2H), 1.61 (s, 3H), 1.28 (s, 6H), 1.26 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 198.88, 156.54, 146.59, 139.07, 130.49, 116.38, 83.46, 81.49, 70.32, 46.39, 39.32, 24.98, 24.87, 16.90; ESI-MS: [M+H]⁺ 317.3; HRMS (FTMS-ESI): [M+H]⁺

calcd for C₁₈H₂₆BO₄⁺ 316.1955, found 316.1966; IR (KBr) ν (cm⁻¹) 2980, 2945, 2846, 1678, 1657, 1401, 1370, 1326, 1308, 1268, 1215, 1168, 1149, 1114, 1021, 932, 848, 782, 684; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 9.5 min (*R,R*-isomer), 11.5min (*S,S*-isomer).

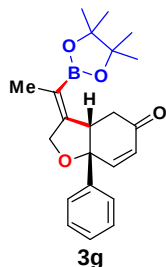


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		9.482	33770.172	296600.500
2		11.482	15125.902	217305.109
Total			48896.074	513905.609



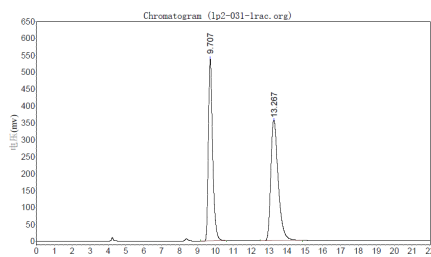
Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		9.478	922.498	11507.348
2		11.500	542222.500	6678938.000
Total			543144.998	6690445.348

(3a*S*,7a*S*,*Z*)-7a-Phenyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3g)

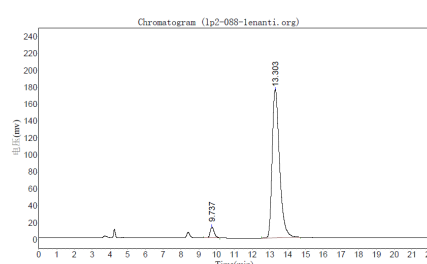


Colorless oil. 35.7 mg, 65% yield; $[\alpha]_D^{25}$ -143.6 (*c* 1.00, CHCl₃) for 92% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.43 (d, *J* = 7.2 Hz, 2H), 7.38 (t, *J* = 7.2 Hz, 2H), 7.31 (t, *J* = 7.2 Hz, 1H), 6.59 (d, *J* = 10.4 Hz, 1H), 6.10 (d, *J* = 10.4 Hz, 1H), 4.59 (AB, *J*_{AB} = 14.8 Hz, 2H), 3.88 (t, *J* = 7.2 Hz, 1H), 2.68-2.81 (m, 2H), 1.57 (s, 3H), 1.26 (s, 6H), 1.24 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 199.04, 156.28, 147.92, 142.48, 129.39, 128.79, 127.90, 125.59, 83.45, 83.30, 70.47, 48.61, 39.79, 24.94, 24.87, 16.96; ESI-MS: [M+Na]⁺ 389.1; HRMS (FTMS-ESI): [M+Na]⁺

calcd for C₂₂H₂₇¹⁰BO₄Na⁺ 388.1936, found 388.1931; IR (KBr) ν (cm⁻¹) 3029, 2979, 2923, 2858, 1684, 1654, 1449, 1402, 1365, 1314, 1266, 1214, 1146, 1106, 1017, 971, 851; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 9.7 min (*R,R*-isomer), 13.3 min (*S,S*-isomer).

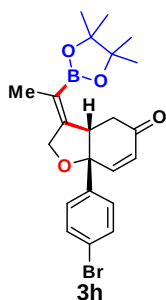


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		9.707	538782.375	8701880.000
2		13.267	356791.719	10156837.000
Total			895574.094	18858717.000

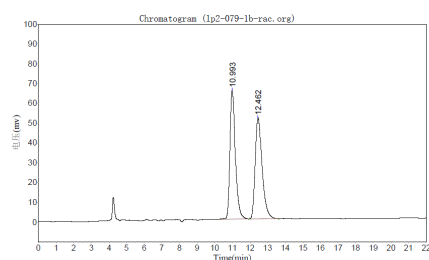


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		9.737	13185.913	205913.594
2		13.303	175222.031	4972531.000
Total			188407.944	5178444.594

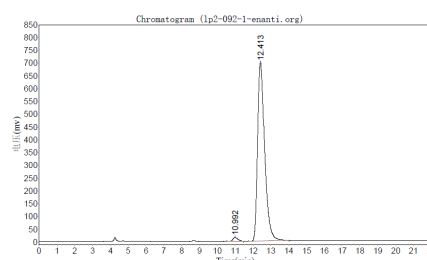
(3a*S*,7a*S*,*Z*)-7a-(4-Bromophenyl)-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3h)



Colorless oil. 42.8 mg, 64% yield; $[\alpha]_D^{25}$ -137.4 (*c* 1.04, CHCl₃) for 97% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.50 (d, *J* = 8.0 Hz, 2H), 7.30 (d, *J* = 8.0 Hz, 2H), 6.54 (d, *J* = 10.4 Hz, 1H), 6.10 (d, *J* = 10.4 Hz, 1H), 4.57 (AB, *J*_{AB} = 14.8 Hz, 2H), 3.85 (t, *J* = 7.2 Hz, 1H), 2.66-2.77 (m, 2H), 1.57 (s, 3H), 1.26 (s, 6H), 1.24 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 198.71, 155.72, 147.14, 141.75, 131.93, 129.61, 127.42, 122.02, 83.49, 82.92, 70.44, 48.56, 39.74, 24.92, 24.88, 16.93; ESI-MS: [M+Na]⁺ 467.1/469.1 (1:1); HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₂₂H₂₆¹⁰B⁷⁹BrO₄Na⁺ 466.1036, found 466.1053; IR (KBr) ν (cm⁻¹) 2978, 2929, 2852, 1690, 1655, 1589, 1485, 1451, 1363, 1310, 1265, 1213, 1144, 1095, 1073, 1009, 968, 849; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 11.0 min (*R*, *R*-isomer), 12.4 min (*S*, *S*-isomer).

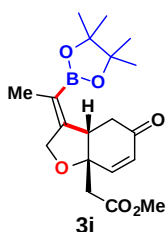


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		10.993	65333.145	1387081.250
2		12.462	51156.801	1396542.375
Total			116489.945	2783623.625

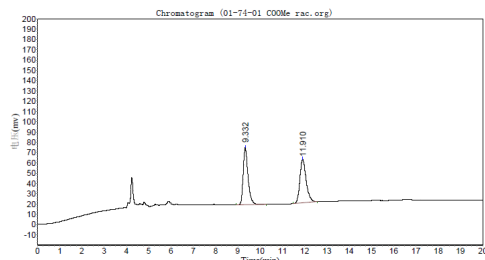


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		10.992	14229.114	279739.781
2		12.413	697066.313	19277728.000
Total			711295.427	19557467.781

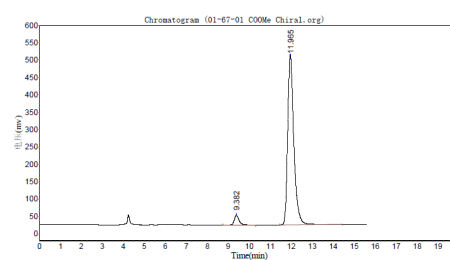
Methyl 2-((3a*S*,7a*S*,*Z*)-5-oxo-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4,5,7a-hexahydrobenzofuran-7a-yl)acetate (3i)



White solid. 27.5 mg, 51% yield; mp 86-87 °C; $[\alpha]_D^{25}$ -72.2 (*c* 0.62, CHCl₃) for 91% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.98 (d, *J* = 10.0 Hz, 1H), 6.08 (d, *J* = 10.0 Hz, 1H), 4.55 (AB, *J*_{AB} = 14.8 Hz, 2H), 3.69 (s, 3H), 3.67-3.60 (m, 1H), 2.79-2.48 (m, 4H), 1.64 (s, 3H), 1.28 (s, 6H), 1.26 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 198.45, 169.97, 156.00, 145.80, 130.51, 83.56, 79.23, 70.12, 51.98, 46.57, 42.78, 39.56, 24.96, 24.91, 16.93; ESI-MS: [M+Na]⁺ 385.2; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₉H₂₇BO₆Na⁺ 384.1829, found 384.1832; IR (KBr) ν (cm⁻¹) 2978, 2851, 1739, 1691, 1655, 1438, 1364, 1307, 1273, 1214, 1145, 1019, 968, 849, 685; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 80/20; flow rate = 0.7 mL/min; Retention time: 9.3 min (*R*, *R*-isomer), 11.9min (*S*, *S*-isomer).

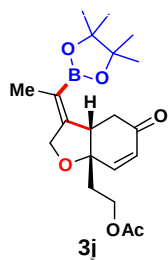


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		9.332	55686.215	838556.938
2		11.910	42394.301	828163.250
Total			98080.516	1666720.188

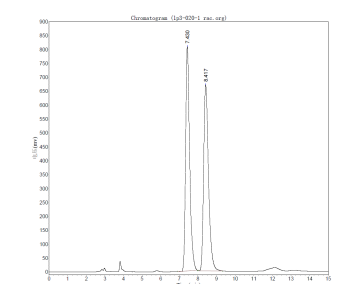


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		9.382	28314.559	435198.594
2		11.965	488818.438	9786368.000
Total			517132.996	10221566.594

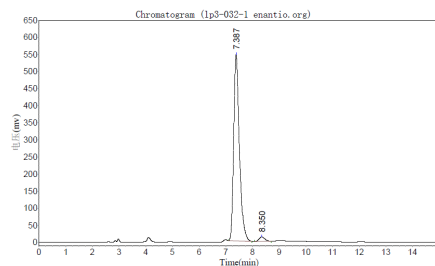
2-((3a*S*,7a*S*,*Z*)-5-oxo-3-(1-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4,5,7a-hexahydrobenzofuran-7a-yl)ethyl acetate (3j)



Colorless oil. 33.6 mg, 60% yield; $[\alpha]_D^{25}$ -100.6 (*c* 0.94, CHCl₃) for 95% *ee* (1.2 equiv. of B₂pin₂ was used); ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.72 (d, *J* = 10.4 Hz, 1H), 6.06 (d, *J* = 10.4 Hz, 1H), 4.47 (AB, *J*_{AB} = 14.8 Hz, 2H), 4.15-4.29 (m, 2H), 3.50 (t, *J* = 7.2 Hz, 1H), 2.57-2.68 (m, 2H), 1.95-2.11 (m, 5H), 1.63 (s, 3H), 1.28 (s, 6H), 1.27 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 198.54, 170.86, 156.63, 146.80, 130.53, 83.52, 80.03, 69.77, 60.19, 46.22, 39.79, 36.12, 25.02, 24.84, 21.02, 17.00; ESI-MS: [M+Na]⁺ 399.1; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₂₀H₂₉¹⁰BO₆Na⁺ 398.1991, found 398.1986; IR (KBr) ν (cm⁻¹) 2978, 2933, 1740, 1687, 1608, 1474, 1455, 1368, 1238, 1145, 1095, 1040, 983, 966, 871, 850; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 70/30; flow rate = 1.0 mL/min; Retention time: 7.4 min (*R,R*-isomer), 8.4 min (*S,S*-isomer).

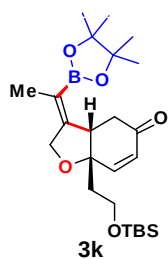


Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1	7.408	7.408	86825.513	1323237.000	50.4157
2	8.417	8.417	68889.438	1261486.000	49.5843
Total			147694.951	2424723.000	100.0000

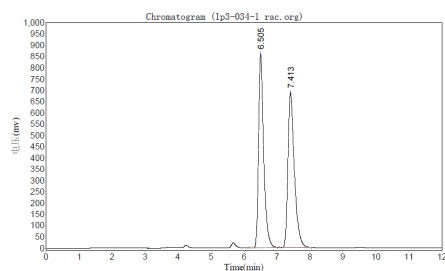


Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1	7.387	7.387	545290.500	8120843.500	97.5096
2	8.350	8.350	13361.660	207406.281	2.4904
Total			558652.160	8328249.781	100.0000

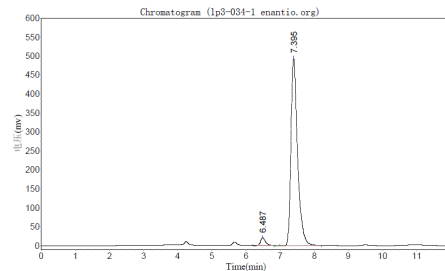
(3a*S*,7a*S*,*Z*)-7a-(2-((*tert*-Butyldimethylsilyl)oxy)ethyl)-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3k)



Colorless oil. 37.3 mg, 55% yield; $[\alpha]_D^{25}$ -80.6 (*c* 0.98, CHCl₃) for 94% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.75 (d, *J* = 10.4 Hz, 1H), 6.02 (d, *J* = 10.4 Hz, 1H), 4.44 (s, 2H), 3.71-3.81 (m, 2H), 3.52 (t, *J* = 6.8 Hz, 1H), 2.61-2.72 (m, 2H), 1.92 (t, *J* = 6.4 Hz, 2H), 1.62 (s, 3H), 1.28 (s, 6H), 1.27 (s, 6H), 0.89 (s, 9H), 0.05 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 199.05, 157.30, 148.40, 129.99, 83.44, 80.71, 69.63, 58.86, 45.98, 40.26, 39.79, 26.00, 25.01, 24.85, 18.30, 17.03, -5.34, -5.37; ESI-MS: [M+Na]⁺ 471.3; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₂₄H₄₁¹⁰BO₅SiNa⁺ 470.2750, found 470.2745; IR (KBr) ν (cm⁻¹) 2955, 2930, 2857, 1721, 1687, 1607, 1472, 1450, 1363, 1312, 1255, 1214, 1145, 1097, 1008, 984, 966, 938, 837; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 6.5 min (*R,R*-isomer), 7.4 min (*S,S*-isomer).

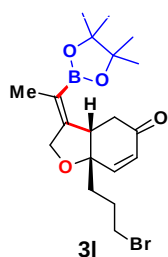


Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1	6.505	6.505	857385.125	9316013.000	50.0154
2	7.413	7.413	684924.125	9310281.000	49.9846
Total			1542309.250	18626294.000	100.0000

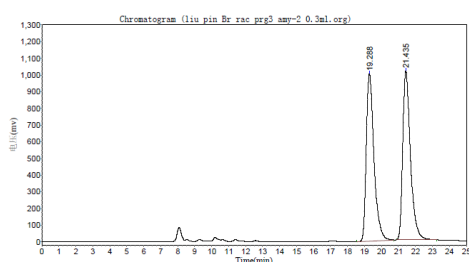


Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1	6.487	6.487	20077.881	209476.594	3.0504
2	7.395	7.395	493566.313	6657657.000	96.9496
Total			513644.193	6867133.594	100.0000

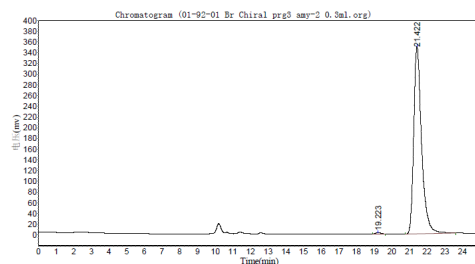
(3a*S*,7a*S*,*Z*)-7a-(3-bromopropyl)-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3l)



White solid. 178 mg, 72% yield (0.6 mmol of **1l** and 1.3 equiv. of B₂pin₂ were used); mp 96-98 °C; [α]_D²⁵ -77.4 (c 0.89, CHCl₃) for 99% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.65 (d, *J* = 10.0 Hz, 1H), 6.06 (d, *J* = 10.0 Hz, 1H), 4.47 (s, 2H), 3.47-3.40 (m, 3H), 2.69-2.55 (m, 2H), 2.05-1.72 (m, 4H), 1.63 (s, 3H), 1.29 (s, 6H), 1.27 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 198.72, 156.86, 147.05, 130.75, 83.51, 80.79, 69.78, 45.88, 39.82, 35.98, 33.69, 27.36, 24.98, 24.90, 16.99; ESI-MS: [M+Na]⁺ 433.2/434.9 (1:1); HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₉H₂₈¹⁰B⁷⁹BrO₄Na⁺ 432.1193, found 432.1193; IR (KBr) ν (cm⁻¹) 3390, 2977, 2931, 1755, 1688, 1473, 1453, 1361, 1214, 1143, 1010, 983, 966, 8551, 674 ; HPLC: Phenomenex Lux 5u Amylose-2 (PA-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 90/10; flow rate = 0.3 mL/min; Retention time: 19.3 min (*R,R*-isomer), 21.4 min (*S,S*-isomer)

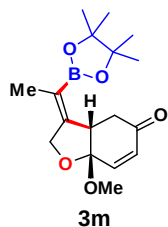


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		19.288	1005998.250	31453262.000
2		21.435	1014638.025	31924626.000
Total			2020656.875	63377888.000

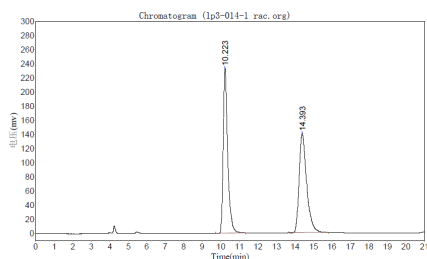


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		19.223	1820.068	39867.750
2		21.422	349268.906	10778526.000
Total			351088.974	10818393.750

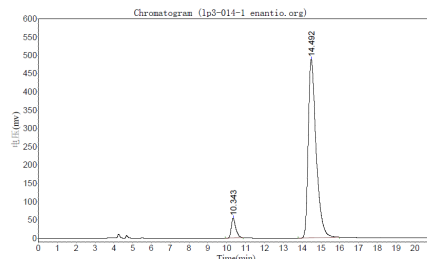
(3a*S*,7a*S*,*Z*)-7a-Methoxy-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3m)



White solid. 23.4 mg, 49% yield; mp 74-76 °C; [α]_D²⁵ -128.3 (c 0.51, CHCl₃) for 88% *ee* (1.3 equiv. of B₂pin₂ was used); ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.94 (d, *J* = 10.4 Hz, 1H), 6.08 (d, *J* = 10.4 Hz, 1H), 4.63 (AB, *J*_{AB} = 14.8 Hz, 2H), 3.68 (dd, *J* = 12.4 Hz, *J* = 6.0 Hz, 1H), 3.38 (s, 3H), 2.77 (dd, *J* = 16.0 Hz, *J* = 6.0 Hz, 1H), 2.29 (dd, *J* = 16.0 Hz, *J* = 12.4 Hz, 1H), 1.65 (s, 3H), 1.26 (s, 6H), 1.25 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 198.40, 155.59, 139.70, 130.62, 103.63, 83.46, 70.94, 48.91, 41.49, 25.10, 24.80, 16.61; ESI-MS: [M+Na]⁺ 343.2; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₇H₂₅¹⁰BO₅Na⁺ 342.1729, found 342.1724; IR (KBr) ν (cm⁻¹) 2978, 2929, 2854, 1693, 1659, 1454, 1366, 1308, 1273, 1216, 1146, 1130, 1102, 1071, 1011, 927, 849; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 10.3 min (*R,R*-isomer), 14.5 min (*S,S*-isomer).

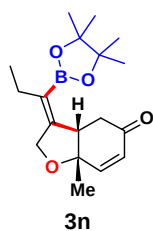


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		10.223	233194.094	3974547.500
2		14.393	140039.672	3953609.000
Total			373233.766	7928156.500

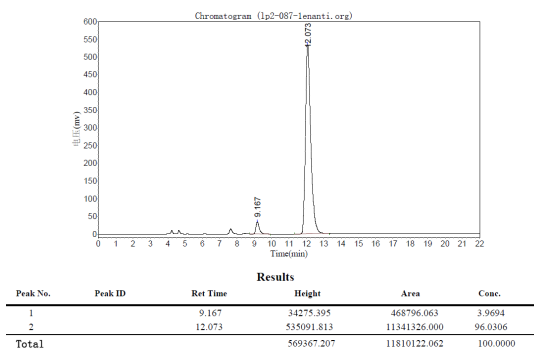
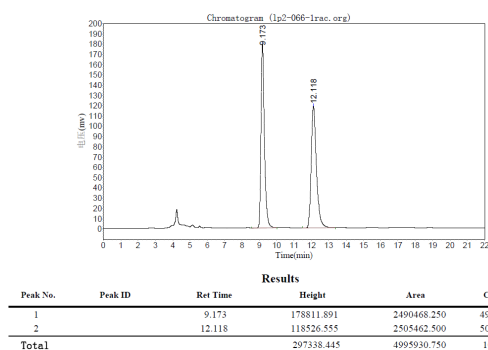


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		10.343	54305.719	907745.063
2		14.492	488614.188	14236909.000
Total			542919.906	15144654.063

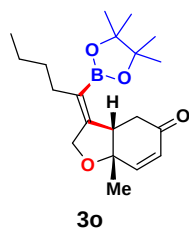
(3a*S*,7a*S*,*Z*)-7a-Methyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)propylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3n)



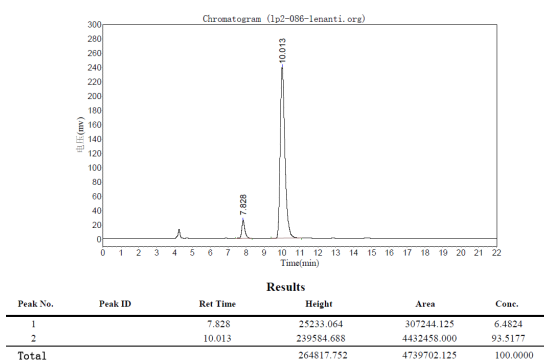
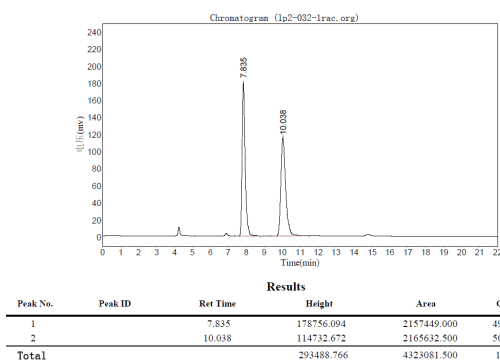
White solid. 26.4 mg, 55% yield; mp 108-110 °C; $[\alpha]_D^{25}$ -109.4 (c 0.98, CHCl₃) for 92% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.62 (d, *J* = 10.0 Hz, 1H), 6.00 (d, *J* = 10.0 Hz, 1H), 4.53 (s, 2H), 3.35 (t, *J* = 7.2 Hz, 1H), 2.55-2.66 (m, 2H), 1.93-2.07 (m, 2H), 1.39 (s, 3H), 1.29 (s, 6H), 1.27 (s, 6H), 0.94 (t, *J* = 7.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 198.95, 155.86, 149.04, 129.71, 83.33, 78.77, 69.02, 47.45, 39.87, 25.37, 25.02, 24.77, 24.67, 14.00; ESI-MS: [M+Na]⁺ 341.2; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₈H₂₇¹⁰BO₄Na⁺ 340.1936, found 340.1931; IR (KBr) ν (cm⁻¹) 2977, 2931, 1720, 1686, 1604, 1474, 1456, 1373, 1273, 1215, 1143, 1067, 1010, 983, 852; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 9.2 min (*R,R*-isomer), 12.1 min (*S,S*-isomer).



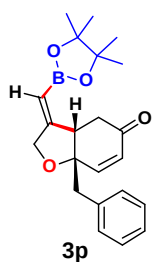
(3a*S*,7a*S*,*Z*)-7a-Methyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)pentylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3o)



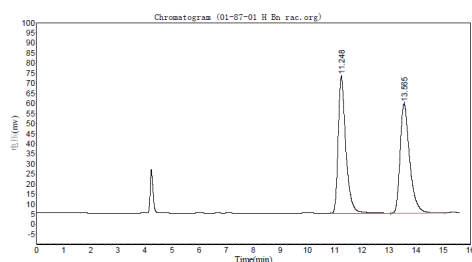
Colorless oil. 24.8 mg, 47% yield; $[\alpha]_D^{25}$ -100.7 (c 0.49, CHCl₃) for 87% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.62 (d, *J* = 10.0 Hz, 1H), 6.00 (d, *J* = 10.0 Hz, 1H), 4.53 (s, 2H), 3.36 (t, *J* = 7.6 Hz, 1H), 2.59 (d, *J* = 7.6 Hz, 2H), 1.92-2.05 (m, 2H), 1.38 (s, 3H), 1.26-1.32 (m, 16H), 0.89 (t, *J* = 6.8 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 199.02, 156.10, 149.02, 129.72, 83.33, 78.71, 69.30, 47.55, 39.95, 31.91, 31.83, 25.01, 24.79, 24.74, 22.70, 14.19; ESI-MS: [M+Na]⁺ 369.3; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₂₀H₃₁¹⁰BO₄Na⁺ 368.2249, found 368.2244; IR (KBr) ν (cm⁻¹) 2959, 2930, 2871, 1690, 1516, 1456, 1371, 1307, 1276, 1218, 1143, 1116, 1056, 1009, 982, 852; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 7.8 min (*R,R*-isomer), 10.0 min (*S,S*-isomer).



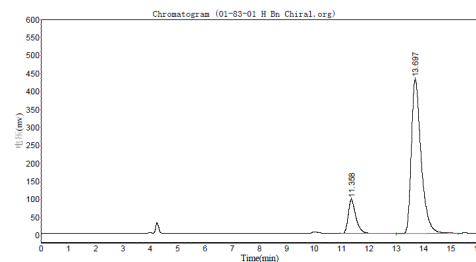
(3a*S*,7a*S*,*E*)-7a-benzyl-3-((4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)methylene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3p)



White solid. 34.4 mg, 63% yield; mp 80-82 °C; $[\alpha]_D^{25}$ -51.9 (c 0.85, CHCl₃) for 70% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.31-7.19 (m, 5H), 6.57 (d, *J* = 10.0 Hz, 1H), 5.99 Hz (d, *J* = 10.0 Hz, 1H), 5.31-5.28 (m, 1H), 4.45 (s, 2H), 3.43 (t, *J* = 6.0 Hz, 1H), 3.09-2.87 (m, 3H), 2.42 (dd, *J* = 6.0 and 16.8 Hz, 1H), 1.29 (s, 6H), 1.27 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 198.23, 165.45, 148.22, 135.67, 130.69, 130.47, 128.36, 127.02, 83.51, 82.29, 72.14, 45.83, 44.05, 38.45, 24.91, 24.89; ESI-MS: [M+Na]⁺ 389.2; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₂₂H₂₇¹⁰BO₄Na⁺ 388.1931, found 388.1943; IR (KBr) ν (cm⁻¹) 2977, 2926, 2852, 1690, 1660, 1496, 1455, 1373, 1332, 1262, 1143, 1043, 969, 850, 763, 701; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 90/10; flow rate = 0.5 mL/min; Retention time: 11.3 min (*R,R*-isomer), 13.6 min (*S,S*-isomer).

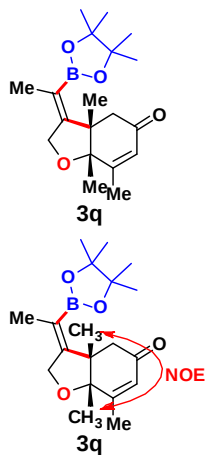


Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		11.248	67731.797	1331678.125	50.1518
2		13.565	54315.250	1323617.750	49.8482
Total			122047.047	2655295.875	100.0000

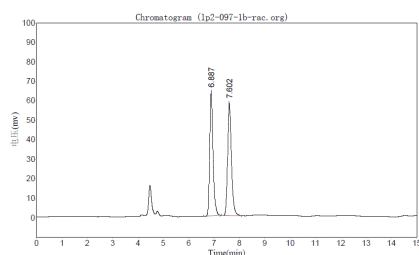


Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		11.358	94183.813	1852301.750	14.8834
2		13.697	428040.531	10593118.000	85.1166
Total			522224.344	12445419.750	100.0000

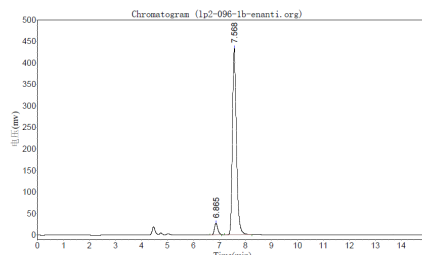
(3a*S*,7a*S*,*Z*)-3a,7,7a-trimethyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3q)



White solid. 32.2 mg, 65% yield; mp 92-94 °C; $[\alpha]_D^{25}$ +20.1 (c 0.96, CHCl₃) for 90% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 5.80 (s, 1H), 4.18 (AB, *J*_{AB} = 14.8 Hz, 2H), 3.58 (d, *J* = 17.2 Hz, 1H), 2.21 (d, *J* = 17.2 Hz, 1H), 1.94 (s, 3H), 1.55 (s, 3H), 1.36 (s, 3H), 1.34 (s, 6H), 1.30 (s, 6H), 1.23 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 197.34, 162.76, 157.68, 129.05, 85.69, 83.61, 69.33, 49.41, 43.31, 25.04, 24.97, 23.06, 17.93, 17.84, 17.31; ESI-MS: [M+Na]⁺ 355.3; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₉H₂₉¹⁰BO₄Na⁺ 354.2093, found 354.2087; IR (KBr) ν (cm⁻¹) 2978, 2930, 2829, 1674, 1450, 1373, 1347, 1299, 1214, 1172, 1140, 1119, 1042, 967, 882, 845; HPLC: Chiracel AD-H Column (250 mm); detected at 214 nm; *n*-hexane / *i*-propanol = 97/03; flow = 0.7 mL/min; Retention time: 6.9 min (*R,R*-isomer), 7.6 min (*S,S*-isomer). The relative configuration was determined by the NOSEY (See Page S114).

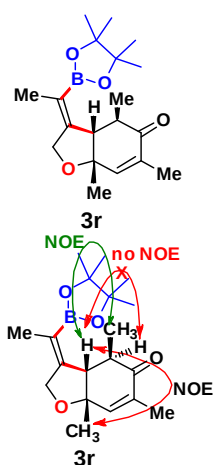


Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		6.887	63436.336	591200.563	49.6524
2		7.602	57728.906	599477.688	50.3476
Total			121165.242	1190678.250	100.0000



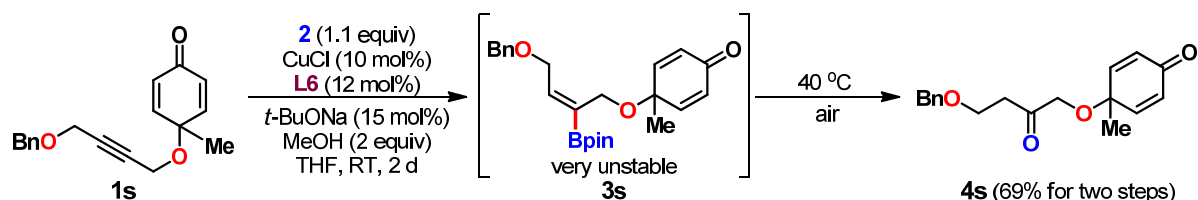
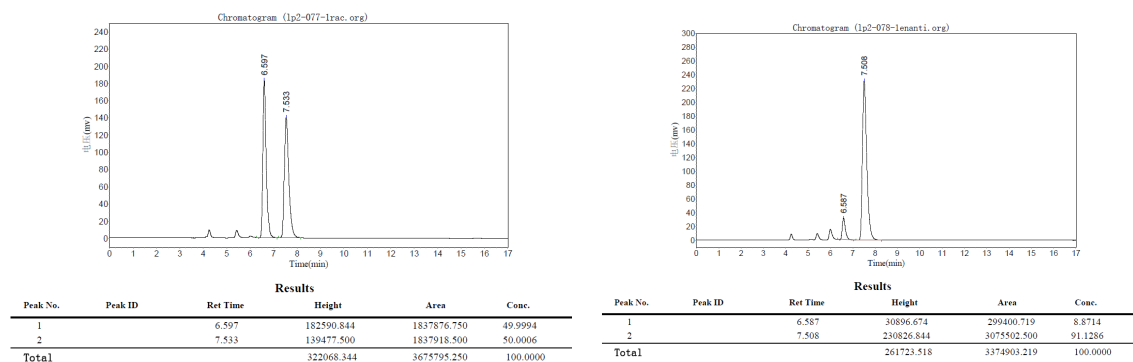
Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		6.865	27131.273	237277.813	4.9851
2		7.568	434355.438	4522500.500	95.0149
Total			461486.711	4759778.313	100.0000

(3a*S*,4*R*,7a*S*,*Z*)-4,6,7a-Trimethyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3r)

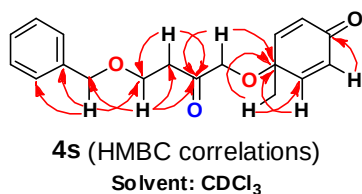


White solid. 30.6 mg, 61% yield; mp 127-129 °C; $[\alpha]_D^{25}$ -79.3 (c 0.51, CHCl₃) for 82% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.37 (d, *J* = 1.6 Hz, 1H), 4.46 (AB, *J*_{AB} = 14.8 Hz, 2H), 3.26 (d, *J* = 9.2 Hz, 1H), 2.55-2.63 (m, 1H), 1.81 (d, *J* = 1.2 Hz, 3H), 1.66 (s, 3H), 1.33 (s, 3H), 1.26 (s, 6H), 1.24 (s, 6H), 1.13 (d, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 202.10, 156.44, 142.79, 135.57, 83.35, 78.88, 69.17, 54.01, 43.35, 26.24, 25.09, 24.85, 17.02, 16.08, 13.64; ESI-MS: [M+Na]⁺ 355.2; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₉H₂₉¹⁰BO₄Na⁺ 354.2093, found 354.2087; IR (KBr) ν (cm⁻¹) 2985, 2942, 2924, 2900, 2851, 1679, 1652, 1449, 1398, 1373, 1360, 1303, 1290, 1220, 1151, 1114, 1088, 1013, 968, 861; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7 mL/min; Retention time: 6.6 min (minor), 7.5 min (major). The relative configurations were determined by the NOSEY (See Page

S117).

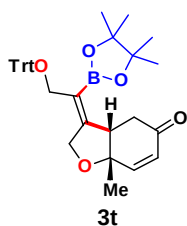


4-(4-(Benzyloxy)-2-oxobutoxy)-4-methylcyclohexa-2,5-dienone (4s) After the general procedure for Cu-catalyzed asymmetric borylative cyclization, the reaction mixture was passed through silica gel (300-400 mesh) and washed with EtOAc. Then silica gel (500 mg) was added to the solution. The mixture was concentrated in vacuo. The resulting residue was stirred at 40 °C for 7 d under air atmosphere. The residue was purified by silica gel (300-400 mesh) column chromatography to afford **4s** (31.0 mg, 69% yield) as colorless oil. ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.37-7.25 (m, 5H), 6.73 (d, *J* = 10.0 Hz, 2H), 6.26 (d, *J* = 10.0 Hz, 2H), 4.78 (s, 2H), 3.97 (s, 2H), 3.73 (t, *J* = 6.0 Hz, 2H), 2.67 (t, *J* = 6.0 Hz, 2H), 1.52 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 205.93, 184.86, 150.40, 137.96, 130.69, 128.52, 127.87, 127.82, 73.41, 73.14, 71.38, 65.15, 39.58, 26.15; ESI-MS: [M+Na]⁺ 323.3; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₈H₂₀O₄Na⁺ 323.1254, found 323.1252; IR (KBr) ν (cm⁻¹) 3032, 2962, 2919, 2850, 1724, 1674, 1632, 1454, 1384, 1365, 1261, 1091, 1027, 863, 801, 740, 700. The position of carbonyl group (blue) in **4s** was determined by HMBC (See Page

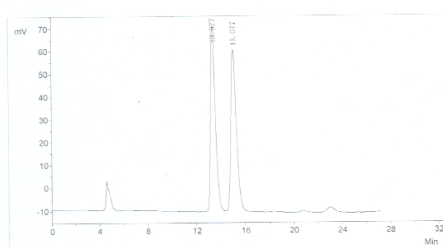


S120-122).

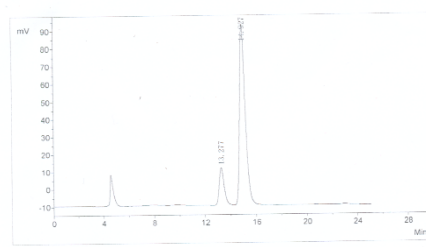
(3a*S*,7a*S*,*Z*)-7a-methyl-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-2-(trityloxy)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (3t)



Colorless oil. 44.5 mg, 53%; $[\alpha]_D^{25}$ -80.3 (*c* 1.02, CHCl₃) for 70% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) . 7.50-7.45 (m, 6H), 7.31-7.18 (m, 9H), 6.56 (d, *J*= 10.0 Hz, 1H), 5.97 (m, *J* = 10.0 Hz, 1H), 4.21 (s, 2H), 3.59 (d, *J* = 9.6 Hz, 1H), 3.48 (d, *J* = 9.6 Hz, 1H), 3.37 (t, *J* = 7.2 Hz, 1H), 2.71-2.55 (m, 2H), 1.37 (s, 3H), 1.33 (s, 6H), 1.32 (s, 6H). ¹³C NMR (100 MHz, acetone-*d*₆) δ (ppm) 197.74, 162.41, 150.27, 145.35, 130.26, 129.69, 128.68, 127.93, 87.51, 84.50, 79.85, 69.51, 63.56, 48.92, 40.14, 25.52, 25.41, 24.51; ESI-MS: $[M+Na]^+$ 585.3; HRMS (FTMS-ESI): $[M+Na]^+$ calcd for C₃₆H₃₉¹⁰BO₅Na⁺ 584.2819, found 584.2792; IR (KBr) ν (cm⁻¹) 2968, 2924, 2852, 1689, 1597, 1490, 1449, 1372, 1308, 1277, 1218, 1142, 1113, 1048, 953, 898, 850, 799, 746, 705, 632; HPLC: DAICEL CHIRALCEL IC-H Column; detected at 214 nm; *n*-hexane / *i*-propanol = 95/05; flow rate = 0.7mL/min; Retention time: 13.4min (*R, R*-isomer), 15.1 min (*S, S*-isomer).



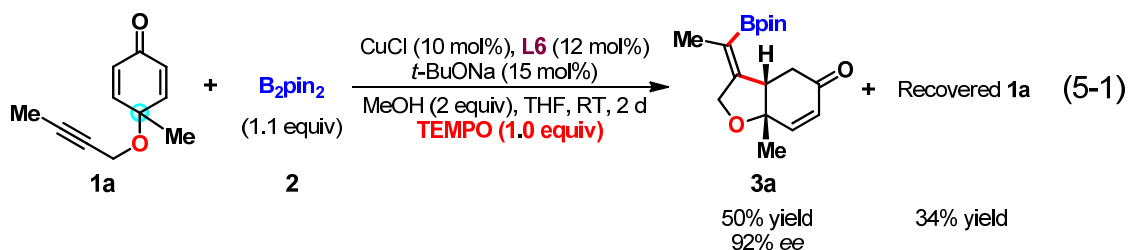
No.	PeakNo	ID_Name	R_Time	PeakHeight	PeakArea	PerCent
1	1		13.377	81647.9	2259405.4	49.5620
2	2		15.077	71165.9	2299340.6	50.4380
Total				152813.7	4558746.1	100.0000



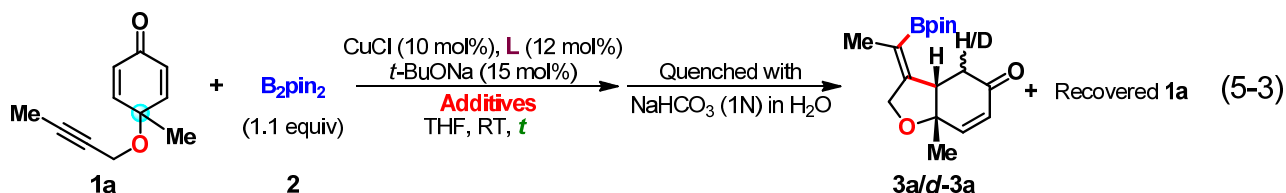
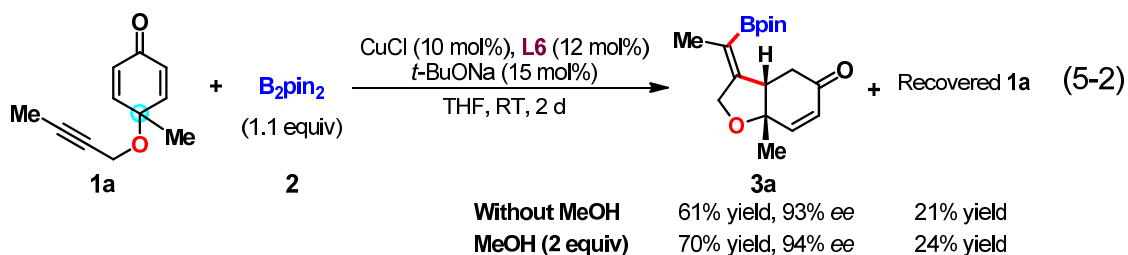
No.	PeakNo	ID_Name	R_Time	PeakHeight	PeakArea	PerCent
1	1		13.277	21214.0	582744.5	15.2389
2	2		14.927	101829.3	3241302.1	84.7611
Total				123043.3	3824046.6	100.0000

5. MECHANISM INVESTIGATION

To probe the reaction mechanism, several controlled experiments were designed and investigated.



PROCEDURE: A dried Schlenk flask was charged with CuCl (1.0 mg, 0.01 mmol, 10 mol%), ligand **L6** (6.5 mg, 0.012 mmol, 12 mol%), B₂pin₂ (27.9 mg, 0.11 mmol, 1.1 equiv), *t*-BuONa (1.4 mg, 0.015 mmol, 15 mol%) and anhydrous THF (1.0 mL) under nitrogen atmosphere. After the mixture was stirred at room temperature for 30 min, a solution of substrate **1a** (17.6 mg, 0.1 mmol) and TEMPO (15.6 mg, 0.1 mmol) in anhydrous THF (1.0 mL) was added, followed by anhydrous MeOH (8.1 μ L, 0.20 mmol, 2.0 equiv) was added. The resulting mixture was stirred at room temperature for 2 days. The reaction mixture was quenched with NaHCO₃ (1 N, 5 mL), and extracted with EtOAc (15 mL $\times$ 3). The combined organic phases were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by silica gel (300-400 mesh) column chromatography to afford the desired product **3a**.

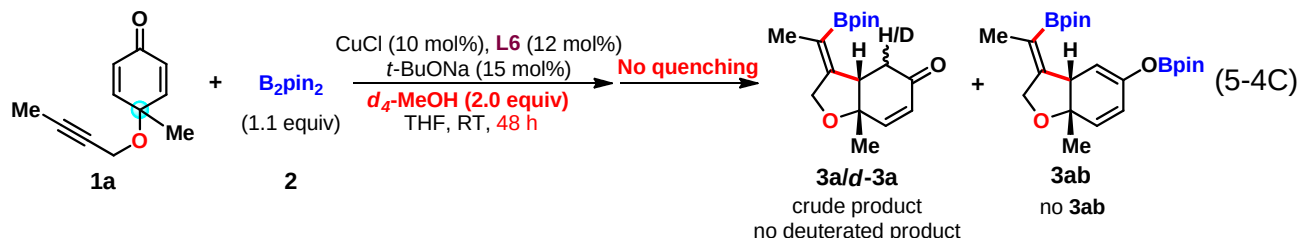
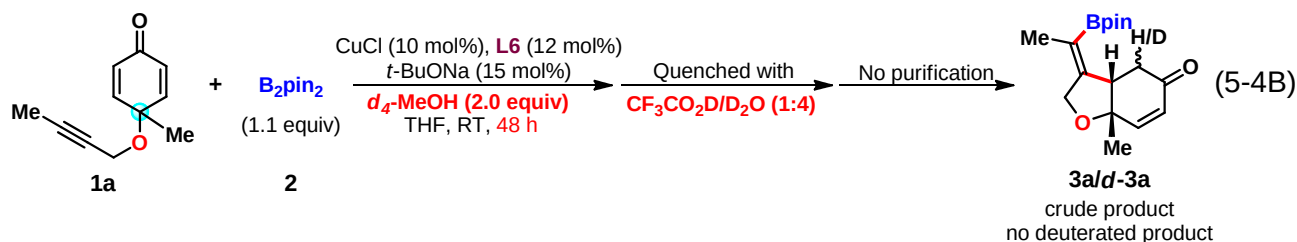
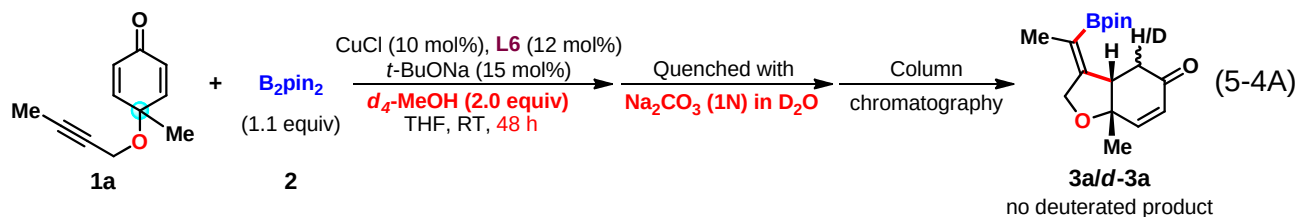


1a (0.1 mmol), THF (2.0 mL), **L** = **L6**, ***d*₄-MeOH (2.0 equiv)**, **48 h**
 64% yield (no deuterated product) 22% yield (5-3A)

1a (0.1 mmol), THF (2.0 mL), **L** = **L6**, ***D*₂O (2.0 equiv)**, **48 h**
 13% yield (no deuterated product) 68% yield (5-3B)

1a (0.3 mmol), THF (2.0 mL), **L** = **L6**, ***d*₄-MeOH (2.0 equiv)**, **48 h**
Operated in glove box. 65% yield (17% deuterated product) 25% yield (5-3C)

1a (0.3 mmol), THF (2.0 mL), **L** = **Ph₃P**, ***d*₄-MeOH (2.0 equiv)**, **18 h**
Operated in glove box. 43% yield (60% deuterated product) 34% yield (5-3D)

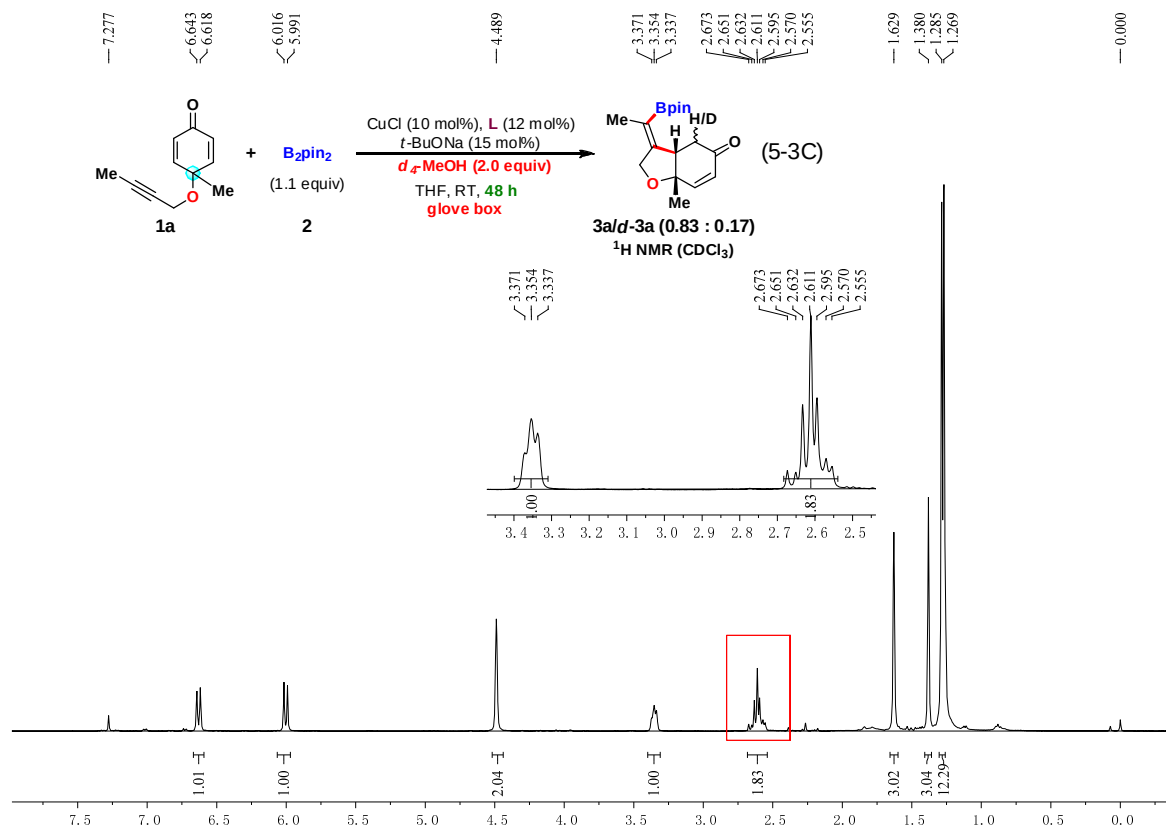


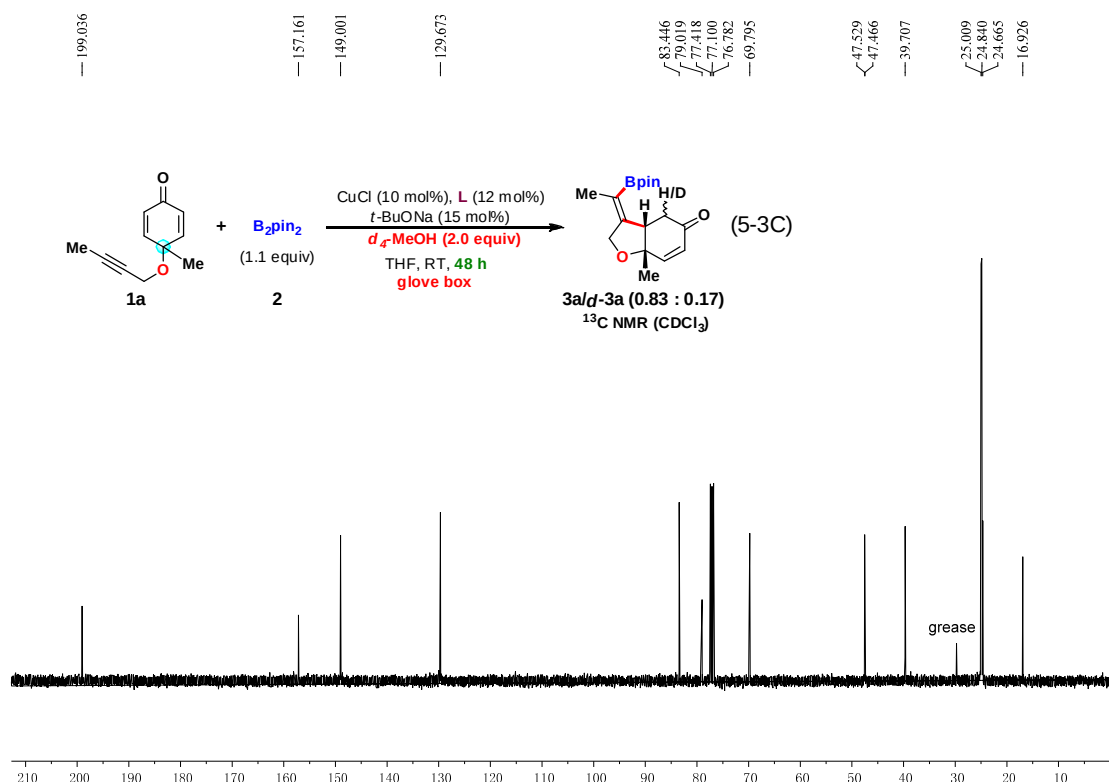
PROCEDURE for 5-3A and 5-3B: A dried Schlenk flask was charged with CuCl (1.0 mg, 0.01 mmol, 10 mol%), ligand **L6** (6.5 mg, 0.012 mmol, 12 mol%), B₂pin₂ (27.9 mg, 0.11 mmol, 1.1 equiv), *t*-BuONa (1.4 mg, 0.015 mmol, 15 mol%) and anhydrous THF (1.0 mL) under nitrogen atmosphere. After the mixture was stirred at room temperature for 30 min, a solution of substrate **1a** (17.6 mg, 0.1 mmol) in anhydrous THF (1.0 mL) was added, followed by anhydrous *d*₄-MeOH (8.1 μL, 0.20 mmol, 2.0 equiv, for

eq 5-3A) or D₂O (3.6 μ L, 0.20 mmol, 2.0 equiv, for eq 5-3B) was added. The resulting mixture was stirred at room temperature for 2 days. The reaction mixture was quenched with NaHCO₃ (1 N, 5 mL), and extracted with EtOAc (15 mL $\times$ 3). The combined organic phases were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by silica gel (300-400 mesh) column chromatography to afford the desired product **3a**.

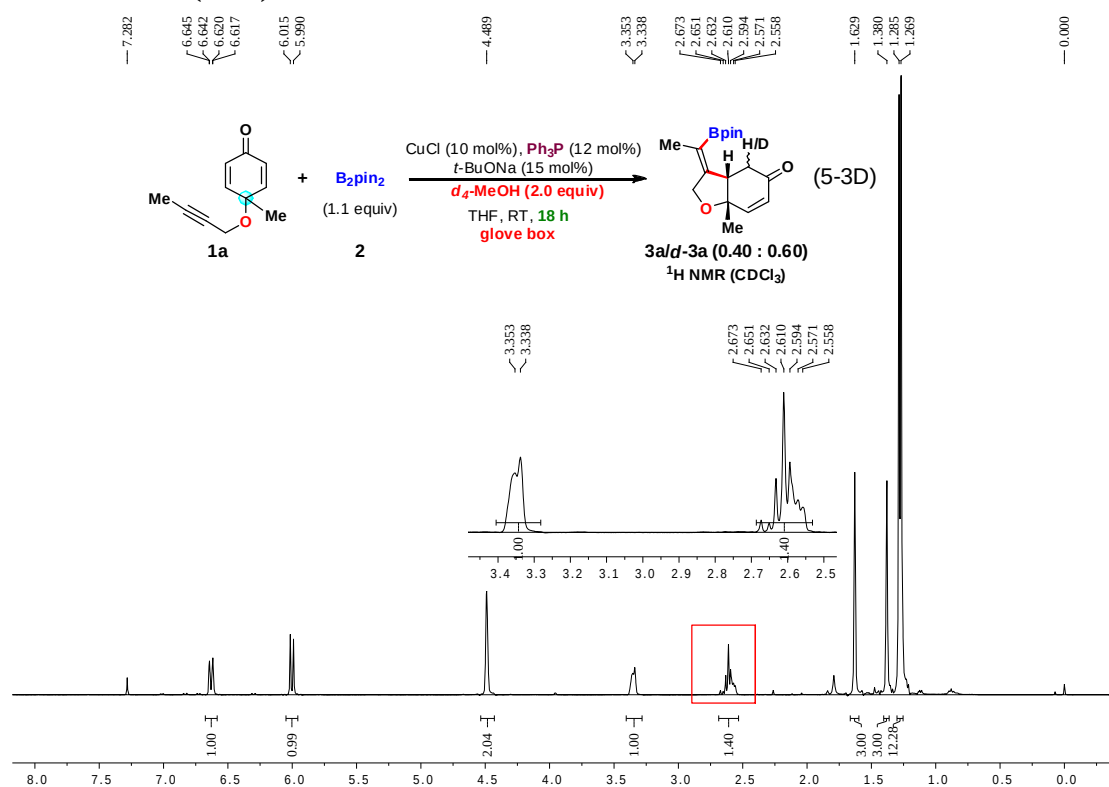
PROCEDURE for 5-3C and 5-3D: In a glove box, a dried Schlenk flask was charged with CuCl (3.0 mg, 0.03 mmol, 10 mol%), ligand **L6** (19.5 mg, 0.036 mmol, 12 mol%, for eq 5-3C) or Ph₃P (9.4 mg, 0.036 mmol, 12 mol%, for eq 5-3D), B₂pin₂ (83.7 mg, 0.33 mmol, 1.1 equiv), *t*-BuONa (4.2 mg, 0.045 mmol, 15 mol%) and anhydrous THF (1.0 mL). After the mixture was stirred at room temperature for 30 min, a solution of substrate **1a** (52.8 mg, 0.3 mmol) in anhydrous THF (1.0 mL) was added, followed by anhydrous *d*₄-MeOH (24.3 μ L, 0.60 mmol, 2.0 equiv) was added. The resulting mixture was stirred at room temperature for 2 days (for eq 5-3C) or 18 h (for eq 5-3D). The reaction mixture was quenched with NaHCO₃ (1 N, 15 mL), and extracted with EtOAc (45 mL $\times$ 3). The combined organic phases were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by silica gel (300-400 mesh) column chromatography to afford a mixture of the desired products **3a** and **d-3a**.

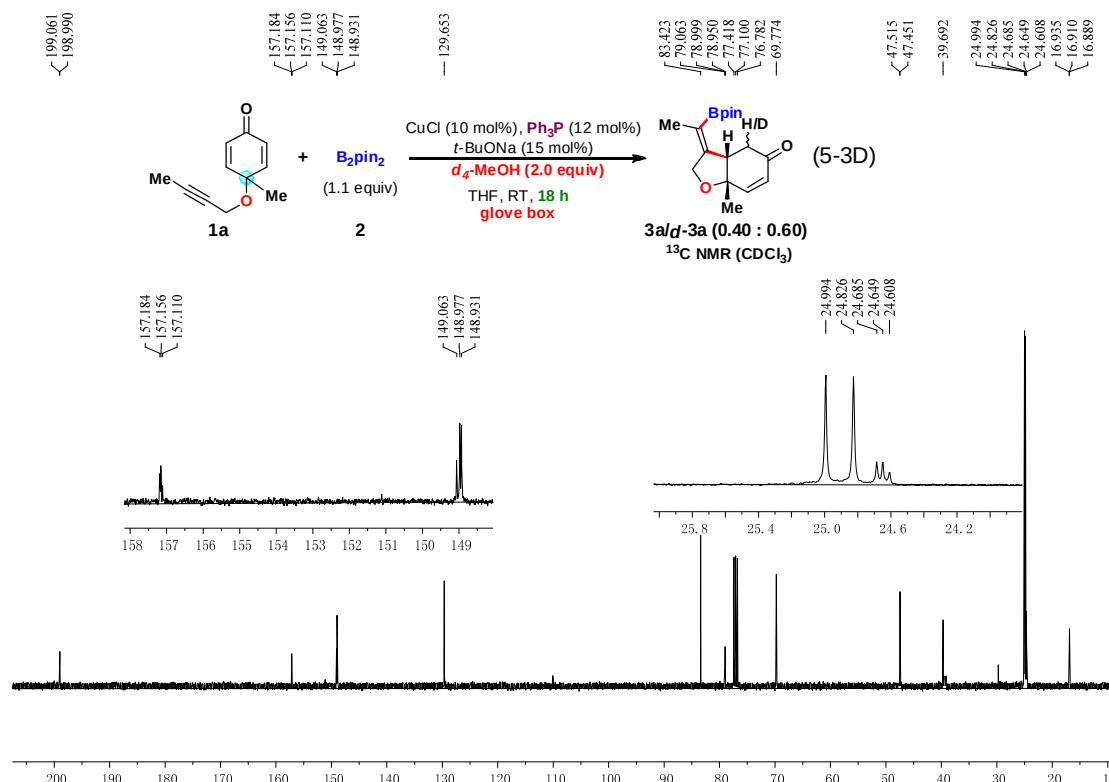
Spectral data for eq 5-3C: Colorless Oil. **3a** and **d-3a** (0.83 : 0.17, determined by ¹H NMR; 59.7 mg, 65% yield); ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.63 (d, *J* = 10.0 Hz, 1H), 6.00 (d, *J* = 10.0 Hz, 1H), 4.49 (s, 2H), 3.37-3.34 (m, 1H), 2.67-2.56 (m, 1.83 H), 1.63 (s, 3H), 1.38 (s, 3H), 1.29 (s, 6H), 1.27 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 199.04, 157.16, 149.00, 129.67, 83.45, 79.02, 69.79, 47.53, 47.47, 39.71, 25.01, 24.84, 24.67, 16.93; ESI-MS: [M+Na]⁺ 327.2 (100%, **3a**), 328.1 (39%, **d-3a**); HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₇H₂₅¹⁰BO₄Na⁺ 326.1774 (**3a**), found 326.1777 (**3a**); [M+Na]⁺ calcd for C₁₇H₂₄D¹⁰BO₄Na⁺ 327.1837 (**d-3a**), found 327.1842 (**d-3a**).



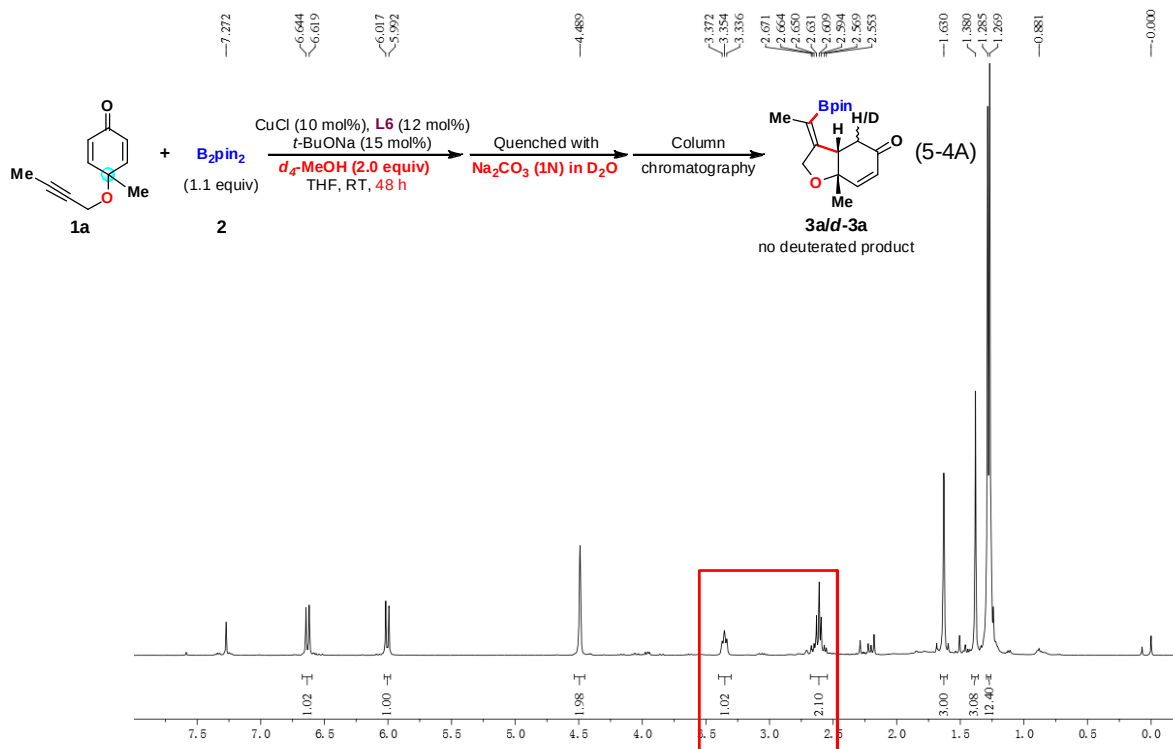


Spectral data for eq 5-3D: Colorless Oil. **3a** and **d-3a** (0.40 : 0.60, determined by ¹H-NMR; 43.2 mg, 43% yield); ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.65-6.62 (m, 1H), 6.00 (d, *J* = 10.0 Hz, 1H), 4.49 (s, 2H), 3.35-3.34 (m, 1H), 2.67-2.56 (m, 1.40 H), 1.63 (s, 3H), 1.38 (s, 3H), 1.29 (s, 6H), 1.27 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 199.06, 198.99, 157.18, 157.16, 157.11, 149.06, 148.98, 148.93, 83.42, 79.06, 79.00, 78.95, 69.77, 47.51, 47.45, 39.69, 24.99, 24.83, 24.69, 24.65, 24.61, 16.93, 16.91, 16.89; ESI-MS: [M+Na]⁺ 327.2 (57%, **3a**); 328.1 (100%, **d-3a**); HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₇H₂₅¹⁰BO₄Na⁺ 326.1774 (**3a**), found 326.1775 (**3a**); [M+Na]⁺ calcd for C₁₇H₂₄D¹⁰BO₄Na⁺ 327.1837 (**d-3a**), found 327.1839 (**d-3a**).

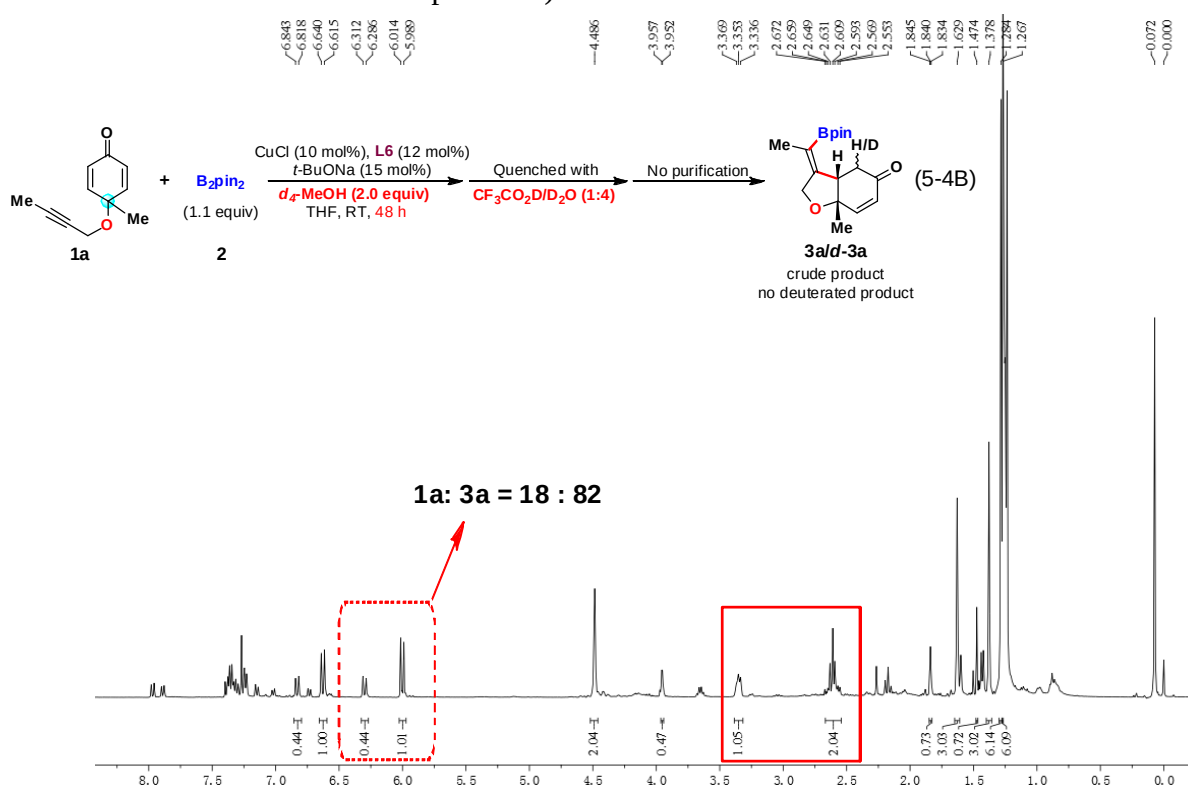




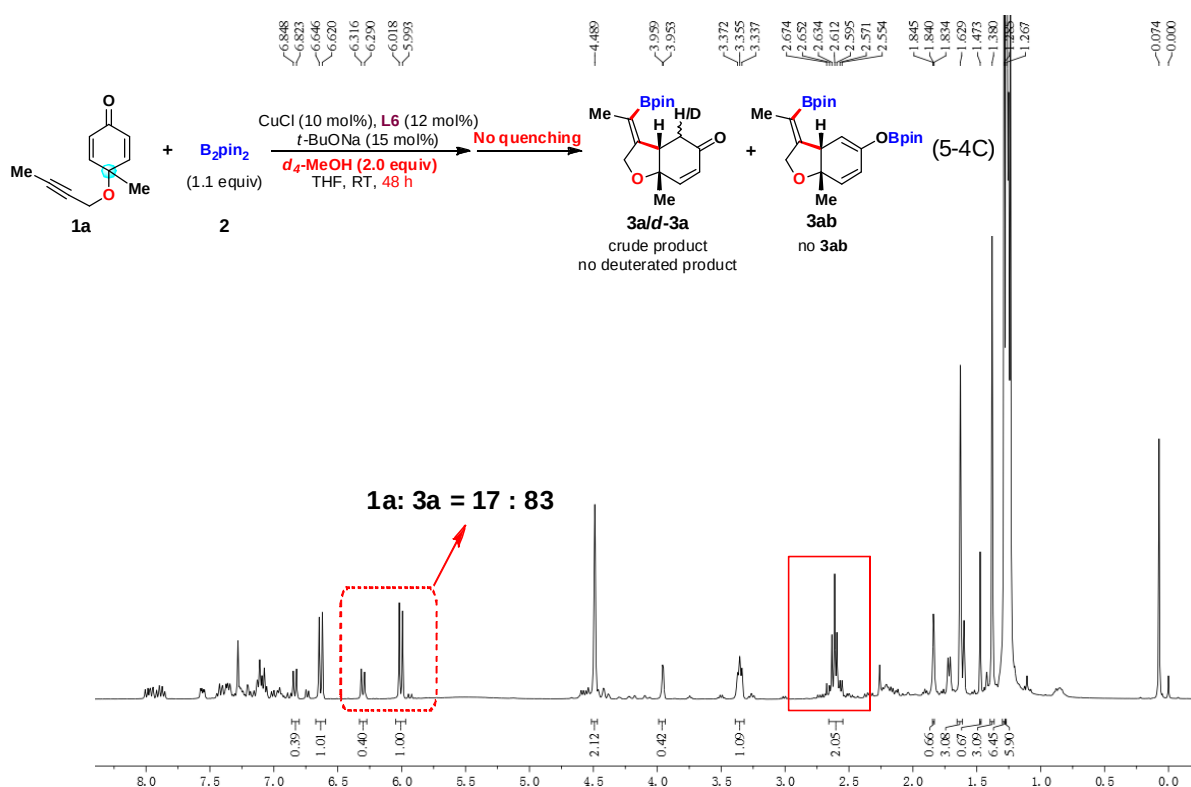
PROCEDURE for 5-4A: A dried Schlenk flask was charged with CuCl (1.0 mg, 0.01 mmol, 10 mol%), ligand **L6** (6.5 mg, 0.012 mmol, 12 mol%), B₂pin₂ (27.9 mg, 0.11 mmol, 1.1 equiv), *t*-BuONa (1.4 mg, 0.015 mmol, 15 mol%) and anhydrous THF (1.0 mL) under nitrogen atmosphere. After the mixture was stirred at room temperature for 30 min, a solution of substrate **1a** (17.6 mg, 0.1 mmol) in anhydrous THF (1.0 mL) was added, followed by anhydrous *d*₄-MeOH (8.1 μL, 0.20 mmol, 2.0 equiv) was added. The resulting mixture was stirred at room temperature for 2 days. The reaction mixture was quenched with a solution of Na₂CO₃ in D₂O (1 N, 2 mL), and extracted with EtOAc (15 mL × 3). The combined organic phases were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by silica gel column chromatography to afford the product **3a** (no *d*-3a).



PROCEDURE for 5-4B: A dried Schlenk flask was charged with CuCl (1.0 mg, 0.01 mmol, 10 mol%), ligand **L6** (6.5 mg, 0.012 mmol, 12 mol%), B₂pin₂ (27.9 mg, 0.11 mmol, 1.1 equiv), *t*-BuONa (1.4 mg, 0.015 mmol, 15 mol%) and anhydrous THF (1.0 mL) under nitrogen atmosphere. After the mixture was stirred at room temperature for 30 min, a solution of substrate **1a** (17.6 mg, 0.1 mmol) in anhydrous THF (1.0 mL) was added, followed by anhydrous *d*₄-MeOH (8.1 μL, 0.20 mmol, 2.0 equiv) was added. The resulting mixture was stirred at room temperature for 2 days. The reaction mixture was quenched with a solution of CF₃CO₂D (0.5 mL) in D₂O (2 mL) and concentrated in vacuo to give the crude product **3a** (No **d**-**3a** was observed in the ¹H NMR spectrum.)



PROCEDURE for 5-4C: A dried Schlenk flask was charged with CuCl (1.0 mg, 0.01 mmol, 10 mol%), ligand **L6** (6.5 mg, 0.012 mmol, 12 mol%), B₂pin₂ (27.9 mg, 0.11 mmol, 1.1 equiv), *t*-BuONa (1.4 mg, 0.015 mmol, 15 mol%) and anhydrous THF (1.0 mL) under nitrogen atmosphere. After the mixture was stirred at room temperature for 30 min, a solution of substrate **1a** (17.6 mg, 0.1 mmol) in anhydrous THF (1.0 mL) was added, followed by anhydrous *d*₄-MeOH (8.1 μL, 0.20 mmol, 2.0 equiv) was added. After the resulting mixture was stirred at room temperature for 2 days, it was concentrated in vacuo to give the crude product **3a** (No **3ab** was observed in the ¹H NMR and ESI-MS spectrums.)



Conclusions:

[1] The reaction performed well in the presence of TEMPO (1 equiv) affording the desired product **3a** in 50% yield and 92% *ee* (eq 5-1), which may exclude a radical process in this tandem reaction.

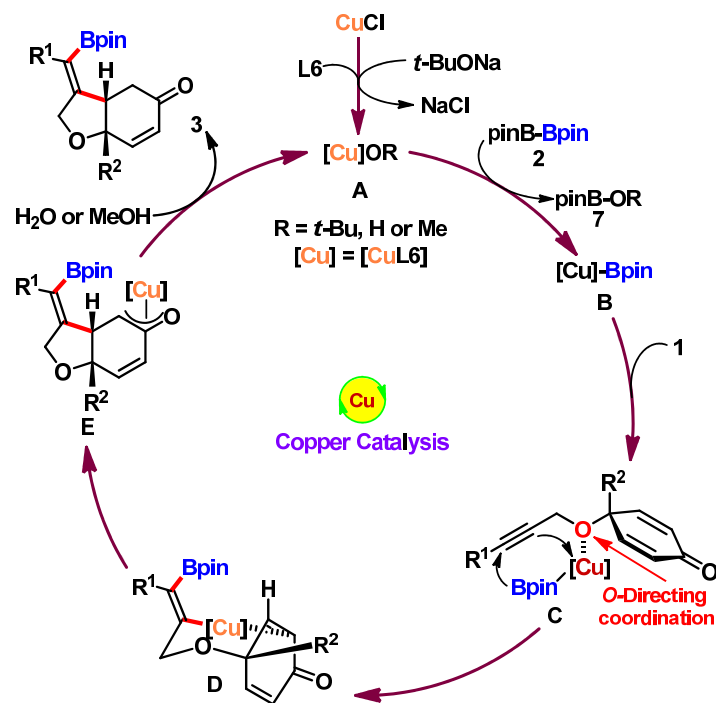
[2] Without methanol under standard condition, the reaction still worked well, albeit in a little lower yield (eq 5-2). The reaction system including dry THF solvent (distilled under Na, 0.8‰ water content, determined by Karl Fischer titration), still contained trace water, which could be acted as the proton source to promote the dissociation of copper. However, the reaction was remarkably inhibited in the presence of the excessive water (eq 5-3B).

[3] In a 0.1 mmol reaction scale, no deuterated product was observed with *d*₄-MeOH (2 equiv) or D₂O (2 equiv) as additive (eq 5-3A, 3B). The trace water in reaction system should provide the protons. In a 0.3 mmol reaction scale under glove box, 17% deuterated product was observed in the presence of *d*₄-MeOH (2 equiv, eq 5-3C). Using PPh₃ as ligand, the deuterated product was increased to 60% (eq 5-3D). The protons should come from the trace water and *d*₄-MeOH.

[4] Owing to the low deuteration rates in the product **3a**, we suspected that the enol boronate **3ab** was a stable intermediate, resulting from the direct transmetallation between enolate copper **E** (Scheme 5-1) and bis(pinacolato)diboron (B₂pin₂, **2**). However, no deuterated product (**d-3a**) or **3ab** was observed in the ¹H NMR spectrums, after the reaction mixture was quenched with a solution of Na₂CO₃ (1 N) in D₂O (eq 5-4A) or CF₃CO₂D in D₂O (eq 5-4B). Without quenching procedure (eq 5-4C), **3ab** was still not observed in the ¹H NMR and ESI-MS spectrums. Thus we supposed that **3ab** was not involved in the transmetallation step.

On the basis of the above results, a plausible mechanism was proposed in scheme 5-1. Initiation of the reaction through the transmetallation of a (pinacolato)boron group (Bpin) from boron to copper species **A** generated the borylated copper **B**, which was coordinated with substrate **1** and subsequently un-

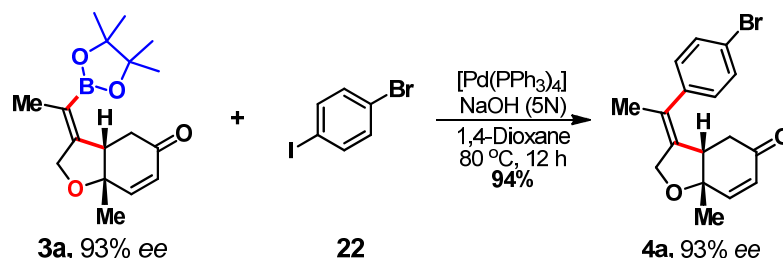
derwent *syn*-addition to carbon-carbon triple bond in the substrate **1** to afford borylated alkenyl-copper intermediate **D**. The favorable six-membered chair form with axial H atom and equatorial R² group in **D** enabled *syn*-migratory insertion of alkenyl-copper across the carbon-carbon double bond in cyclohexadienone, forming the oxa- π -allylcopper intermediate **E**, which was readily protonated by trace water or methanol to regenerate **A** and liberate the product **3**.



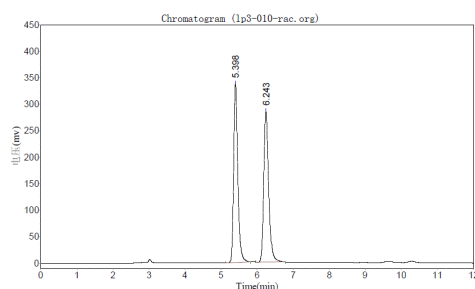
Scheme 5-1 Proposed mechanism

6. TRANSFORMATION OF THE CYCLIZATION PRODUCTS

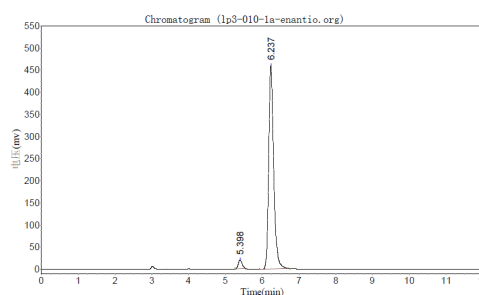
4.1 The Transformation of Cyclization Product 3a.



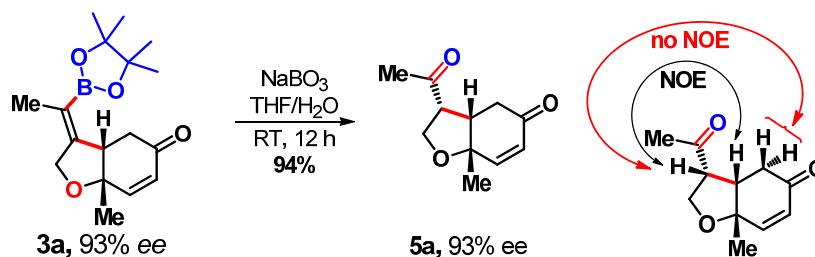
(3a*S*,7a*S*,*E*)-3-(1-(4-bromophenyl)ethylidene)-7a-methyl-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (4a)^{1c} A mixture of **3a** (30.4 mg, 0.1 mmol), 1-bromo-4-iodobenzene (**22**, 42.4 mg, 0.15 mmol), Pd(PPh₃)₄ (11.6 mg, 0.01 mmol), aq. NaOH (5N, 30 μL, 0.15 mmol) in dioxane (2 mL) was stirred at 80 °C under N₂ atmosphere overnight. After cooled to room temperature, the reaction mixture was filtered and washed with CH₂Cl₂. The filtrate was concentrated under reduced pressure and purified by flash column chromatography using hexane/ethyl acetate as eluent (20:1) to afford the desired product **4a**. White solid. 31.2 mg, 94% yield; mp 162-164 °C; [α]_D²⁵ +38.4 (*c* 0.50, CHCl₃) for 93% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.48 (d, *J* = 8.4 Hz, 2H), 7.02 (d, *J* = 8.4 Hz, 2H), 6.49 (d, *J* = 10.4 Hz, 1H), 5.92 (d, *J* = 10.4 Hz, 1H), 4.39 (AB, *J*_{AB} = 13.6 Hz, 2H), 3.09 (s, 1H), 2.16 (dd, *J* = 16.4 Hz, *J* = 5.6 Hz, 1H), 2.01 (dd, *J* = 16.8 Hz, *J* = 4.0 Hz, 1H), 1.84 (s, 3H), 1.48 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 197.41, 150.28, 141.17, 136.68, 131.86, 129.93, 129.20, 129.01, 121.22, 80.70, 69.73, 46.70, 36.91, 23.33, 21.92; ESI-MS: [M+Na]⁺ 355.0/357.0 (1:1); HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₇H₁₇⁷⁹BrO₂Na⁺ 355.0304, found 355.0300; IR (KBr) ν (cm⁻¹) 3029, 2977, 2968, 1676, 1482, 1450, 1395, 1229, 1040, 873, 822; HPLC: Chiracel AD-H Column (250 mm); detected at 214 nm; n-hexane / i-propanol = 90/10; flow = 1.0 mL/min; Retention time: 5.4 min (*R*, *R*-isomer), 6.2 min (*S*, *S*-isomer).



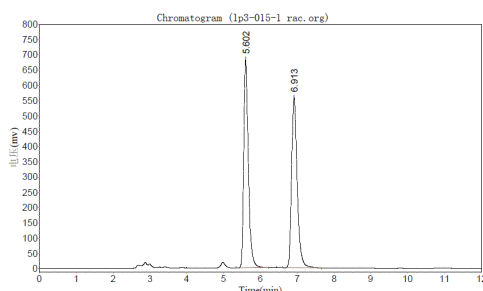
Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		5.398	338402.656	2726331.500
2		6.243	285770.219	2708528.500
Total			624172.875	5434860.000



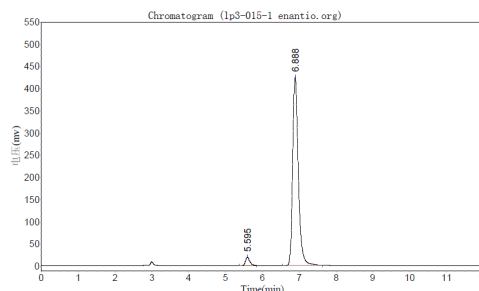
Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		5.398	20148.514	153971.500
2		6.237	459436.844	4344393.000
Total			479585.357	4498364.500



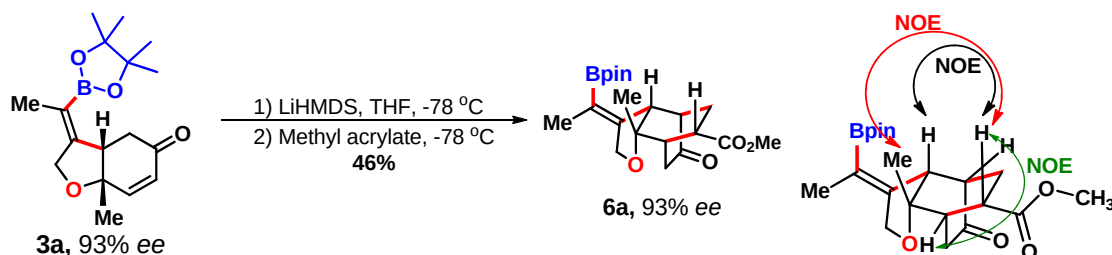
(3*R*,3*aS*,7*aS*)-3-acetyl-7*a*-methyl-2,3,3*a*,4-tetrahydrobenzofuran-5(7*aH*)-one (5a) To a solution of **3a** (25 mg, 0.08 mmol) in THF/H₂O (1:1, 4 mL) was added NaBO₃•4H₂O (124 mg, 0.8 mmol, 10.0 equiv.) and the reaction mixture was stirred at room temperature overnight. Then it was diluted with H₂O (10 mL) and extracted with ethyl acetate (15 mL × 3). The combined organic phases were washed with brine (10 mL), dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography using hexane/ethyl acetate eluent (6:1) to afford the desired product **5a**. Colorless oil. 14.7 mg, 94% yield; [α]_D²⁵ +28.5 (c 0.51, CHCl₃) for 93% ee; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.60 (d, *J* = 10.4 Hz, 1H), 5.99 (d, *J* = 10.4 Hz, 1H), 4.00-3.92 (m, 2H), 3.02-3.08 (m, 1H), 2.74-2.79 (m, 1H), 2.60-2.71 (m, 2H), 2.18 (s, 3H), 1.51 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 205.91, 196.87, 152.62, 129.07, 80.60, 67.57, 56.97, 45.75, 37.76, 29.79, 23.38; ESI-MS: [M+Na]⁺ 217.1; HRMS (FTMS-ESI): [M+Na]⁺ calcd for C₁₁H₁₄O₃Na⁺ 217.0841, found 217.0835; IR (KBr) ν (cm⁻¹) 2973, 2930, 2884, 1712, 1681, 1420, 1374, 1358, 1286, 1234, 1174, 1155, 1119, 1053, 1024, 876; HPLC: Chiracel AD-H Column (250 mm); detected at 214 nm; n-hexane / *i*-propanol = 70/30; flow = 1.0 mL/min; Retention time: 5.6 min (minor), 6.9 min (major). The relative configuration of acetyl substitution was determined by the NOSEY (See Page S132).



Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		5.602	680854.750	5821362.500	49.8891
2		6.913	556886.563	5847248.000	50.1109
Total			1237741.313	11668610.500	100.0000

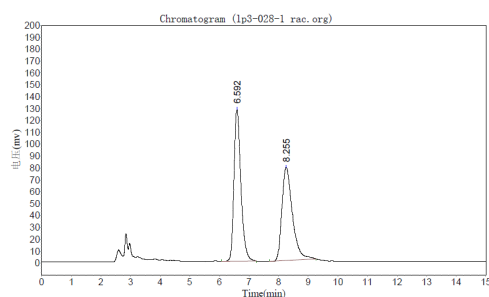


Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		5.595	18343.594	150452.203	3.2697
2		6.888	423764.688	4450891.000	96.7303
Total			442108.281	4601343.203	100.0000

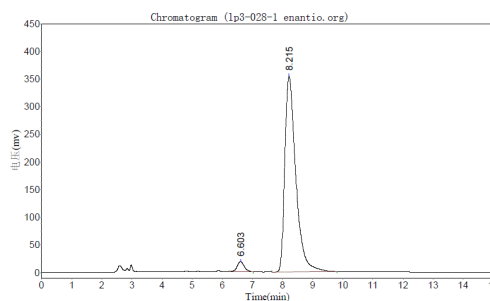


(3*aS*,4*R*,6*S*,7*S*,7*aS*,*Z*)-methyl-7*a*-methyl-9-oxo-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)eth ylidene)octahydro-4,7-ethanobenzofuran-6-carboxylate (6a) To a solution of LiHMDS (120 μ L, 1.0 N in THF, 0.12 mmol) in THF (1 mL) was added a solution of **3a** (30.4 mg, 0.1 mmol) in THF (1.5 mL) dropwise at -5 $^\circ$ C. After the reaction mixture was stirred for 30 min, methyl acrylate (12.9 mg, 0.15 mmol)

was added and the resulting reaction mixture was still stirred overnight at the same temperature. Then the reaction was quenched with aqueous saturated NH_4Cl (10 mL) and extracted with EtOAc (15 mL $\times$ 3). The combined organic phases were washed with brine (10 mL), dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The residue was purified by flash column chromatography using hexane/ethyl acetate eluent (10:1) to afford the recovered **3a** (5.2 mg, 17% yield) and the desired product **6a** (17.9 mg, 46% yield) as colorless oil. $[\alpha]_{\text{D}}^{25}$ -60.4 (c 0.99, CHCl_3) for 94% *ee*; ^1H NMR (400 MHz, CDCl_3) δ (ppm) 4.39 (s, 2H), 3.74 (s, 3H), 3.08 (s, 1H), 2.88(t, J = 9.2Hz, 1H), 2.62-2.67 (m, 2H), 2.51-2.52 (m, 1H), 2.08-2.21 (m, 3H), 1.61 (s, 3H), 1.36 (s, 3H), 1.26 (s, 6H), 1.24 (s, 6H). ^{13}C NMR (100 MHz, CDCl_3) δ 215.06, 174.92, 156.57, 84.25, 83.28, 70.49, 54.79, 52.28, 50.91, 41.93, 37.58, 36.32, 25.64, 25.15, 24.94, 22.63, 17.02; ESI-MS: $[\text{M}+\text{Na}]^+$ 413.3; HRMS (FTMS-ESI): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{21}\text{H}_{31}^{10}\text{BO}_6\text{Na}^+$ 412.2148, found 412.2142; IR (KBr) ν (cm^{-1}) 2976, 1731, 1649, 1438, 1402, 1364, 1310, 1282, 1213, 1177, 1145, 1099, 1058, 1029, 967, 856; HPLC: Phenomenex Lux 5u Cellulose-2 (PC-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 70/30; flow rate = 1.0 mL/min; Retention time: 6.6 min (minor), 8.2 min (major). The relative configurations were determined by the NOSEY (See Page S140-141).

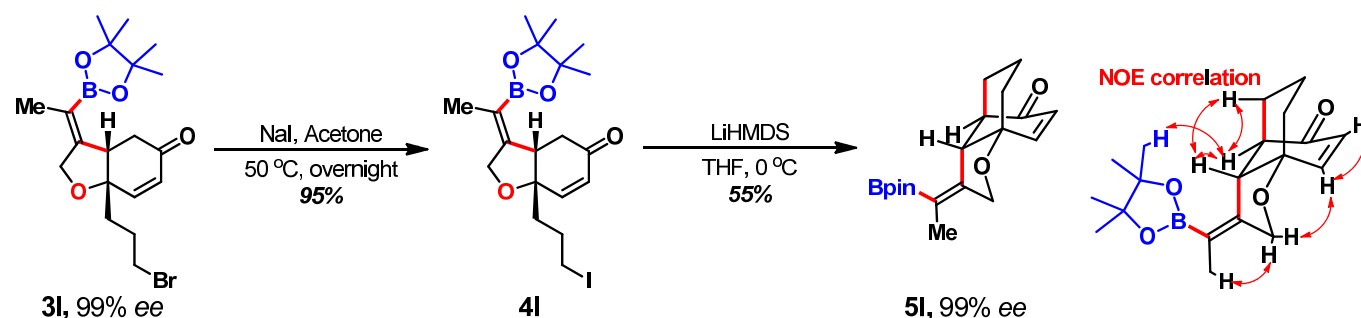


Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		6.592	127600.398	2035743.000	51.4219
2		8.255	78372.586	1923162.375	48.5781
Total			205972.984	3958905.375	100.0000



Results					
Peak No.	Peak ID	Ret Time	Height	Area	Conc.
1		6.603	18255.059	298599.906	3.2287
2		8.215	353955.719	8949618.000	96.7713
Total			372210.777	9248217.906	100.0000

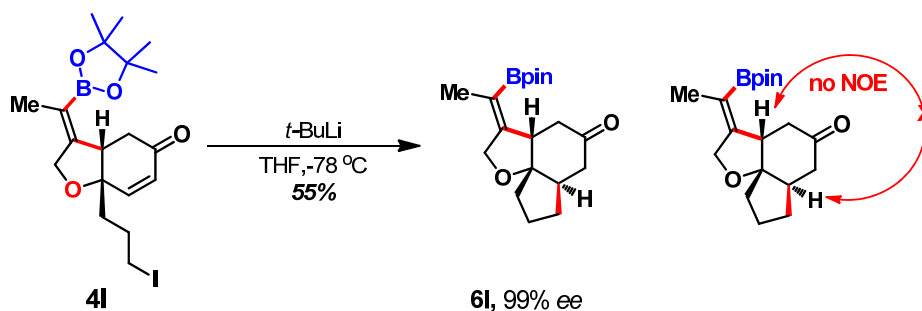
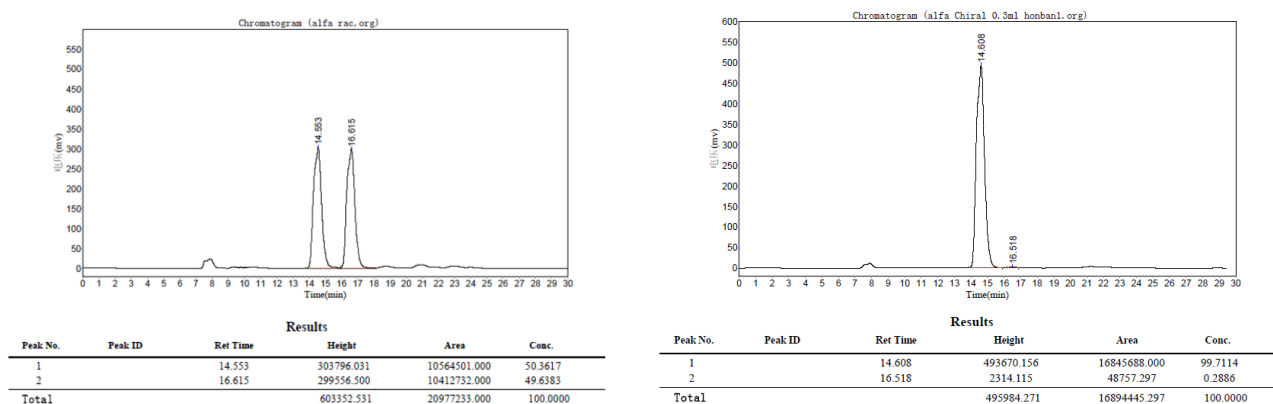
4.2 The Transformation of Cyclization Product 3l



(3a*S*,7a*S*,*Z*)-7a-(3-Iodopropyl)-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-2,3,3a,4-tetrahydrobenzofuran-5(7a*H*)-one (4l) A mixture of **3l** (50 mg, 0.122 mmol) and NaI (182 mg, 1.22 mmol, 10 equiv) in acetone (10 mL) was stirred for overnight at 50 °C. The resulting mixture was concentrated in vacuo and the residue was purified by silica gel (300-400 mesh) column chromatography to afford **4l** (53.2 mg, 95%) as white solid. mp 101-103 °C; ^1H NMR (400 MHz, CDCl_3) δ (ppm) 6.64 (d, J = 10.4 Hz, 1H), 6.06 (d, J = 10.4 Hz, 1H), 4.47 (s, 2H), 3.43 (t, J = 7.2 Hz, 1H), 3.21 (t, J = 6.4 Hz, 2H),

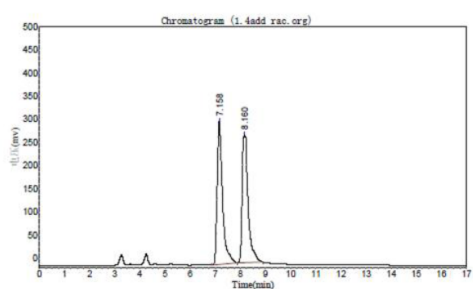
2.68-2.55 (m, 2H), 2.01-1.69 (m, 4H), 1.63 (s, 3H), 1.29 (s, 6H), 1.27 (s, 6H). ^{13}C NMR (100 MHz, CDCl_3) δ (ppm) 198.73, 156.86, 147.12, 130.78, 83.54, 80.74, 69.83, 45.91, 39.84, 38.30, 28.14, 25.01, 24.94, 17.01, 6.55; ESI-MS: $[\text{M}+\text{Na}]^+$ 481.1; HRMS (FTMS-ESI): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{19}\text{H}_{28}^{10}\text{BIO}_4\text{Na}^+$ 480.1054, found 480.1064; IR (KBr) ν (cm^{-1}) 2976, 2925, 2853, 1687, 1624, 1451, 1363, 1307, 1273, 1213, 1144, 1094, 1022, 966, 850, 685.

(3a*S*,4*R*,7a*S*,*Z*)-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)-3a,4-dihydro-2*H*-4,7a-propanobenzofuran-5(3*H*)-one (5*I*) To a solution of **4I** (30.6 mg, 0.067 mmol) in dry THF (1 mL) was added to a solution of LiHMDS (1 M THF solution, 0.344 mL, 5 equiv) in dry THF (1 mL) at 0 °C. The resulting mixture was warmed to room temperature and stirred for overnight. The reaction mixture was quenched by aqueous saturated NH_4Cl (10 mL) and extracted with AcOEt (20 mLx3). The combined organic phases were dried over anhydrous Na_2SO_4 , filtered and concentrated in vacuo. The residue was purified by silica gel (300-400 mesh) column chromatography to afford **5I** (12.1 mg, 55%) as white solid. mp 102-104 °C; $[\alpha]_D^{25}$ -69.8 (*c* 0.32, CHCl_3) for 99% *ee*; ^1H NMR (400 MHz, CDCl_3) δ (ppm) 6.53 (dd, *J* = 10.0, 2.0 Hz, 1H), 6.09 (d, *J* = 10.0 Hz, 1H), 4.35 (AB, *J*_{AB} = 14.4 Hz, 2H), 3.81-3.77 (m, 1H), 2.65 (m, 1H), 1.95-1.88 (m, 1H), 1.85-1.62 (m, 4H), 1.59 (s, 3H), 1.55-1.42 (m, 1H), 1.31 (s, 6H), 1.28 (s, 6H). ^{13}C NMR (100 MHz, CDCl_3) δ (ppm) 200.62, 152.15, 151.75, 133.15, 83.68, 82.23, 71.24, 54.07, 45.98, 31.18, 28.31, 25.06, 24.97, 19.78, 17.50; ESI-MS: $[\text{M}+\text{H}]^+$ 331.3; HRMS (FTMS-ESI): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{19}\text{H}_{27}^{10}\text{BO}_4\text{Na}^+$ 352.1931, found 352.1925; IR (KBr) ν (cm^{-1}) 2976, 2933, 2862, 1716, 1683, 1540, 1456, 1358, 1305, 1212, 1180, 1134, 1084, 1005, 965, 925, 849, 820, 690.; HPLC: Phenomenex Lux 5u Amylose-2 (PA-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 90/10; flow rate = 0.3 mL/min; Retention time: 14.6 min (major), 16.6 min (minor). The relative configurations were determined by the NOSEY (See Page S154-155).

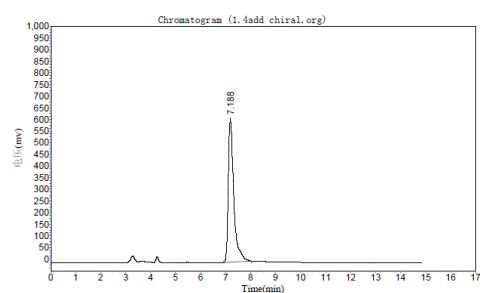


(3a*S*,6a*R*,91*S*,*Z*)-3-(1-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)octahydroindeno[4-*b*]furan-5(6*H*)-one (6*I*) To a solution of **4I** (34 mg, 0.074 mmol) in dry THF (2 mL) was added $t\text{-BuLi}$ (1.5M in THF, 0.148 mL, 0.223 mmol, 3 equiv.) dropwise at -78 °C. After the resulting mixture was

stirred for 2 min, it was quenched by MeOH (0.5 mL). The mixture was passed through silica gel (300-400 mesh), washed with EtOAc and concentrated in vacuo. The residue was purified by silica gel (300-400 mesh) column chromatography to afford **6I** (13.2 mg, 55%) as white solid. mp 111-112 °C; $[\alpha]_D^{25}$ -70.9 (*c* 0.34, CHCl₃) for 99% *ee*; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 4.42 (AB, J_{AB} = 15.2 Hz, 2H), 3.19 (dd, J = 12.8, 4.4 Hz, 1H), 2.48-2.32 (m, 3H), 2.23-2.09 (m, 2H), 2.09-1.99 (m, 1H), 1.92-1.84 (m, 1H), 1.83-1.66 (m, 2H), 1.63 (s, 3H), 1.48-1.39 (m, 1H), 1.32-1.23 (m, 12H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 213.10, 158.26, 93.11, 83.31, 68.57, 45.98, 44.04, 43.00, 42.00, 36.06, 31.86, 25.10, 24.76, 23.50, 16.59; ESI-MS: $[M+Na]^+$ 355.3; HRMS (FTMS-ESI): $[M+Na]^+$ calcd for C₁₉H₂₉¹⁰BO₄Na⁺ 354.2087, found 354.2079; IR (KBr) ν (cm⁻¹) 2954, 2868, 1716, 1652, 1456, 1403, 1364, 1306, 1212, 1145, 1112, 1029, 966, 850, 686.; HPLC: Phenomenex Lux 5u Amylose-2 (PA-2) Column; detected at 214 nm; *n*-hexane / *i*-propanol = 90/10; flow rate = 0.7 mL/min; Retention time: 7.2 min (major), 8.2 min (minor). The relative configuration was determined by the NOSEY (See Page S163).

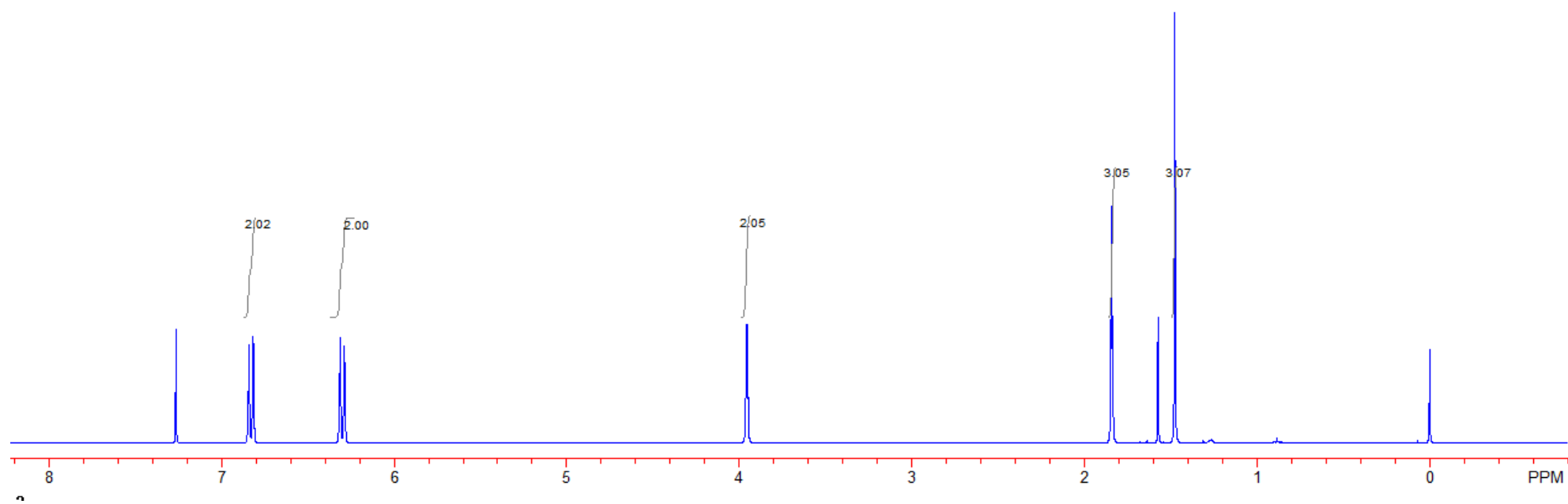
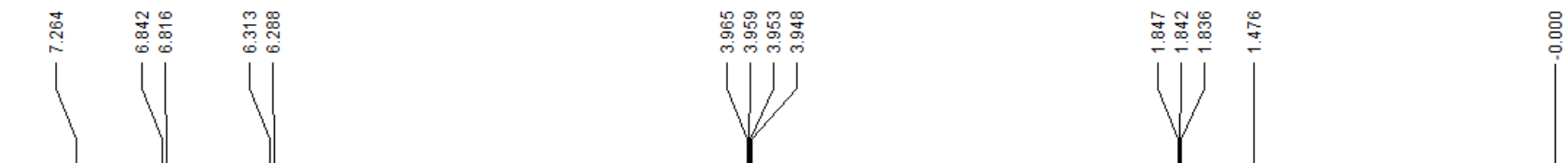


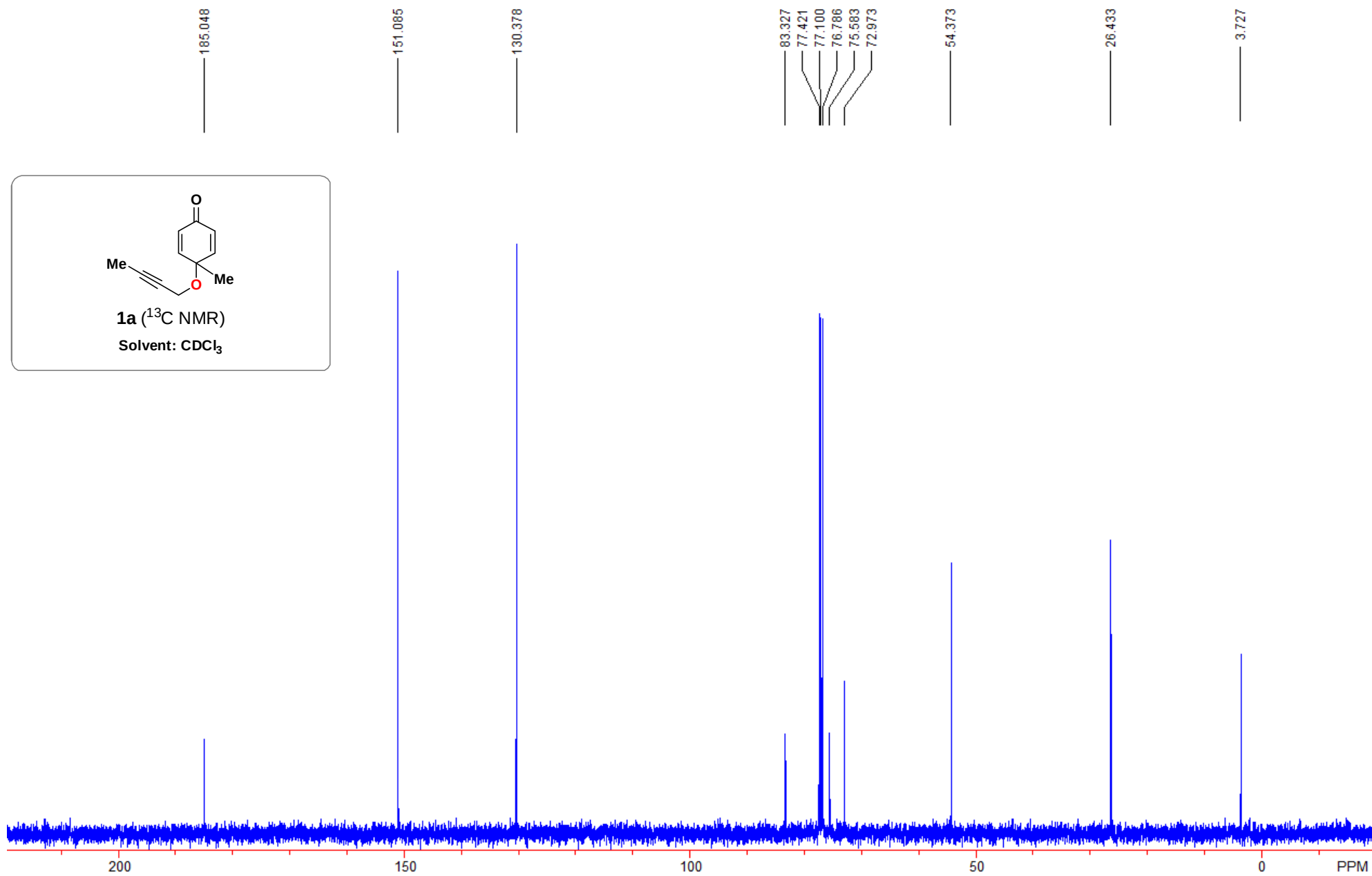
Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		7.158	309135.344	4564355.500
2		8.160	318772.125	4755568.000
Total			627907.469	9319923.500

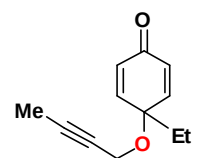
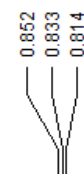
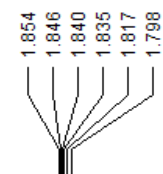
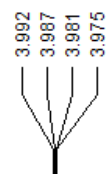
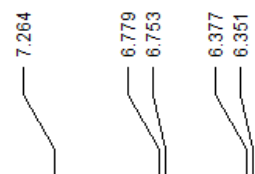


Results				
Peak No.	Peak ID	Ret Time	Height	Area
1		7.188	611045.313	9337060.000
Total			611045.313	9337060.000

7. ^1H NMR, ^{13}C NMR, HSQC, HMBC, DEPT, H-H COSY & NOSEY COPIES

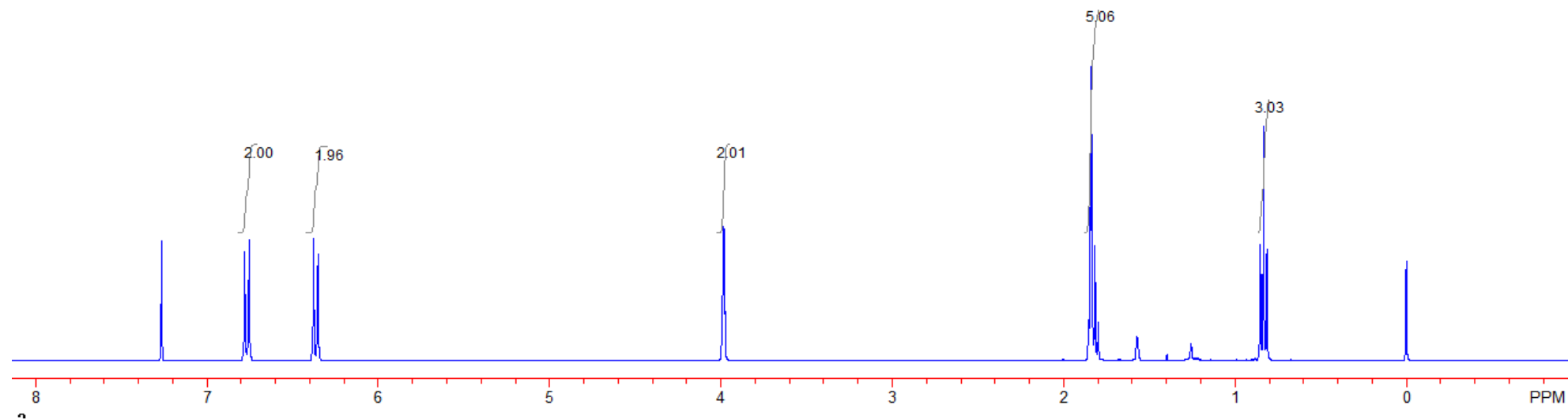


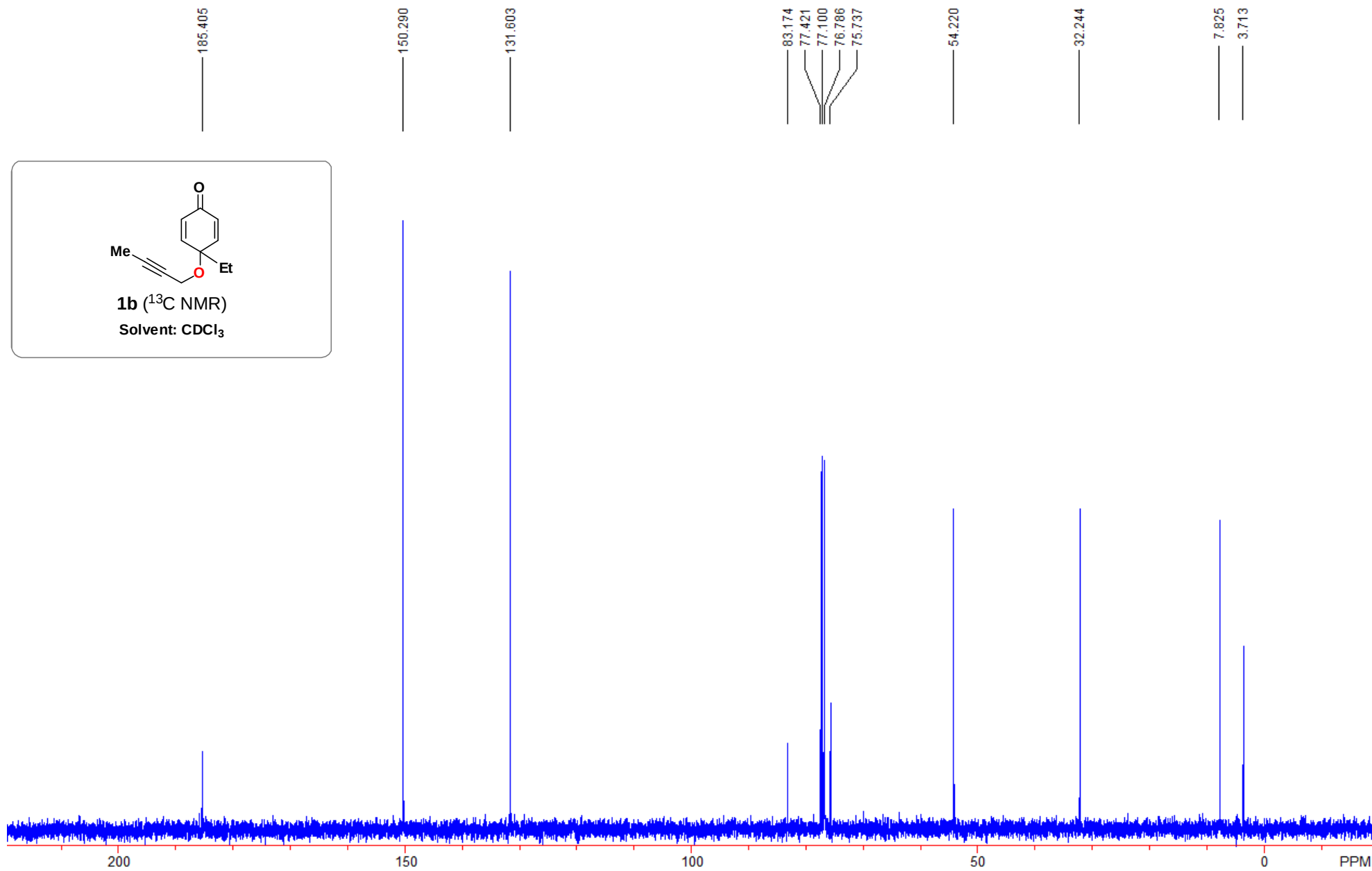


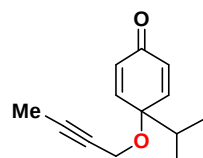


1b (¹H NMR)

Solvent: CDCl₃

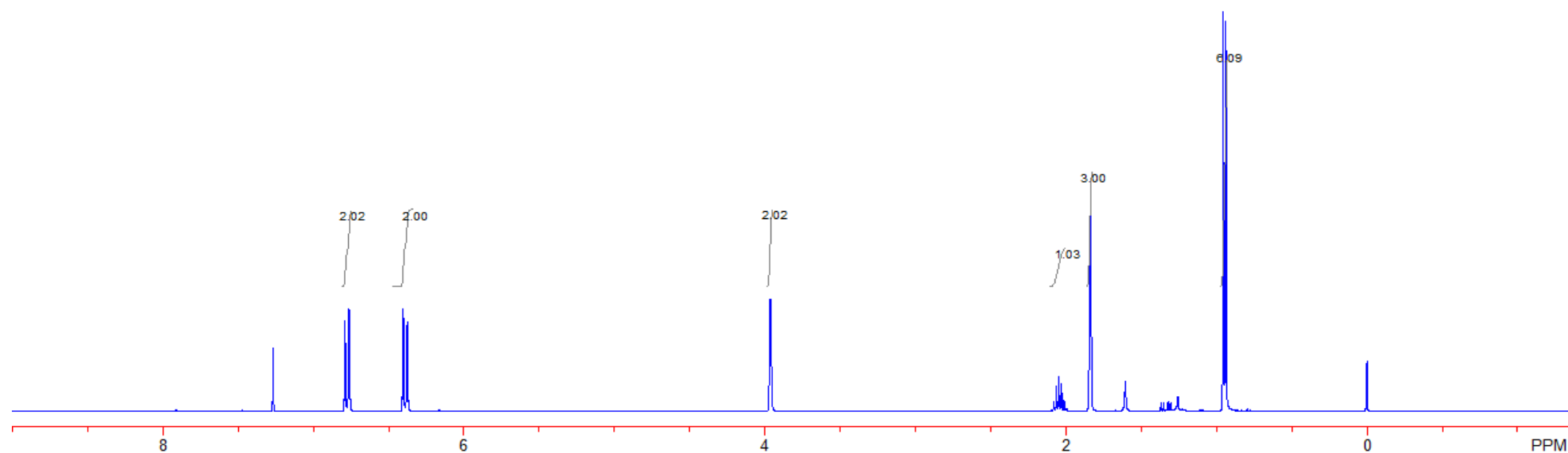
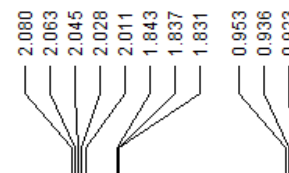
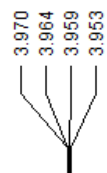
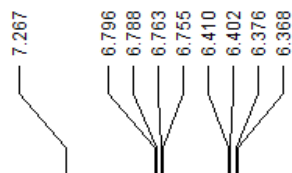


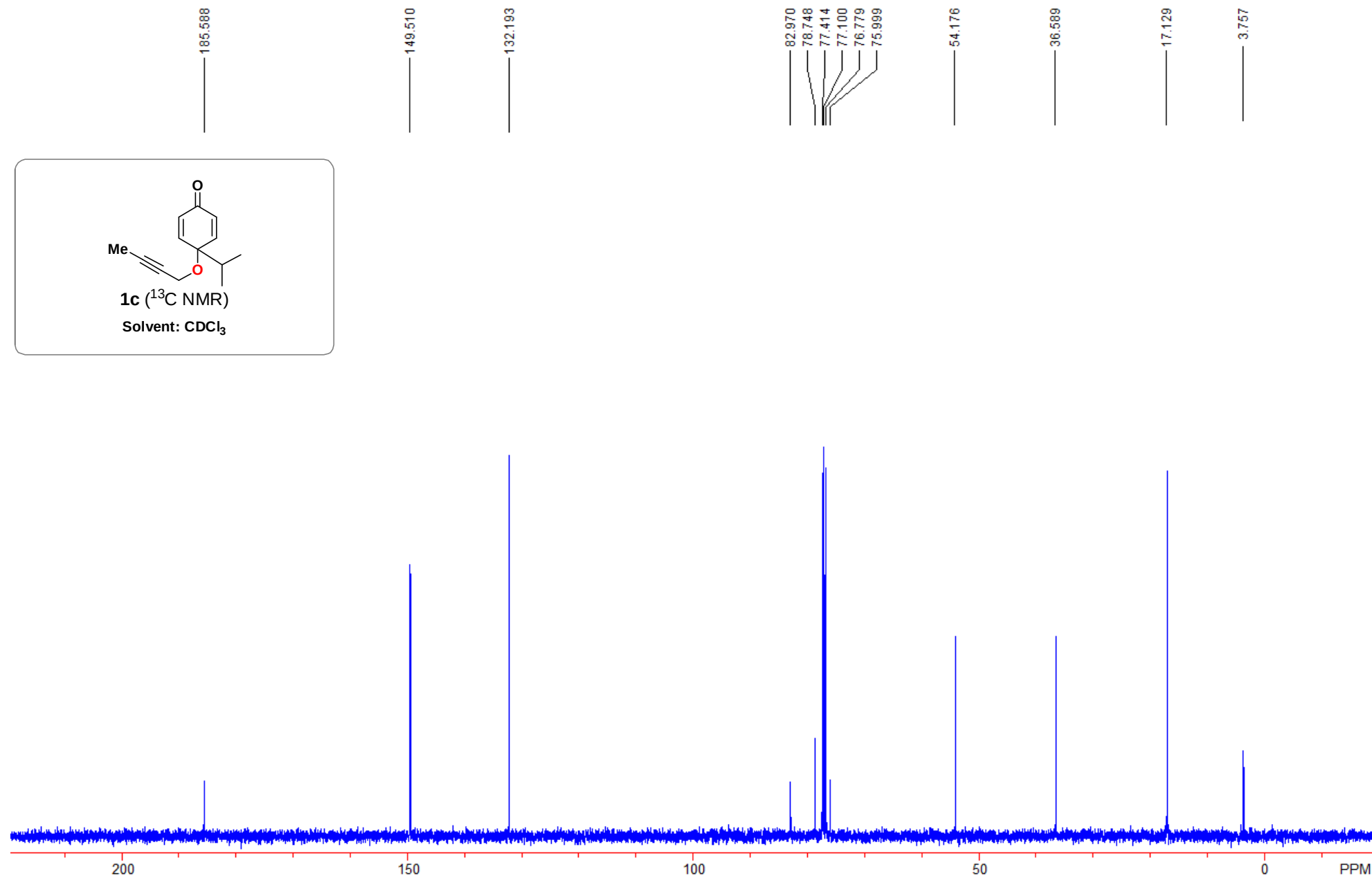
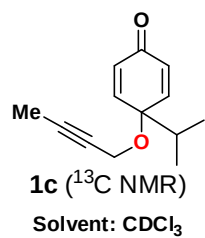


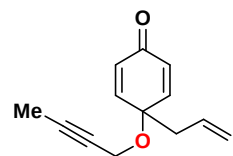
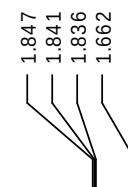
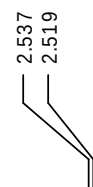
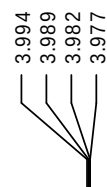
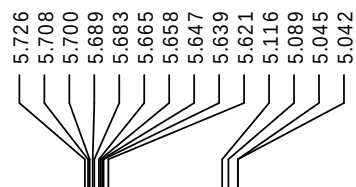
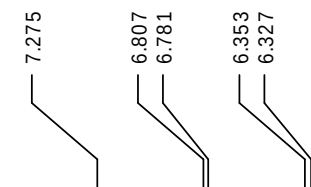


1c (^1H NMR)

Solvent: CDCl_3

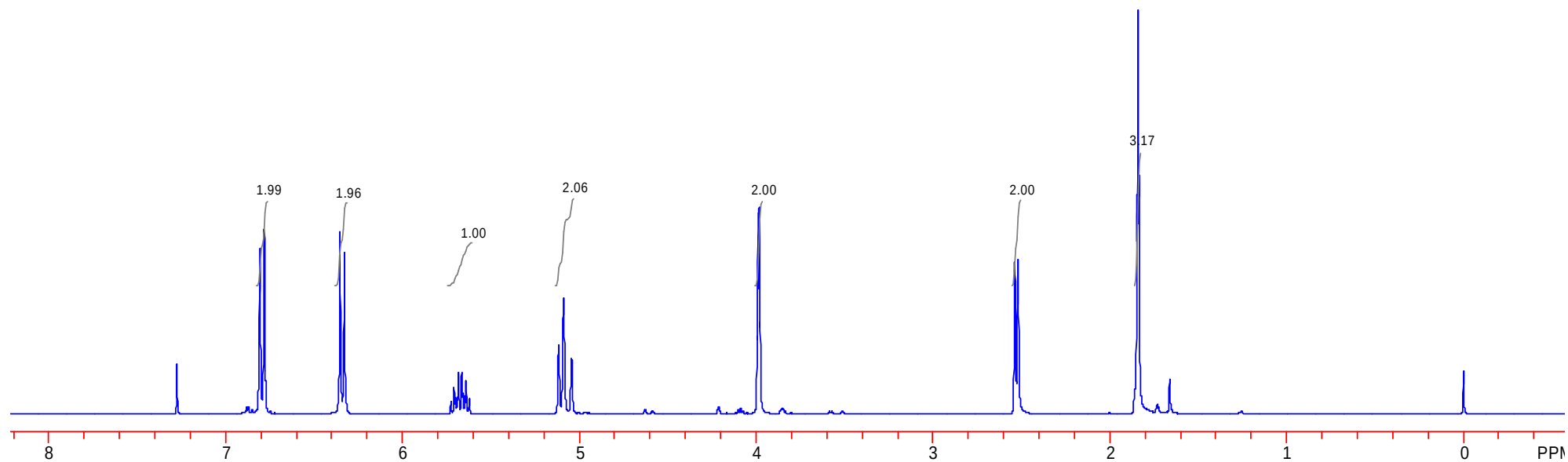




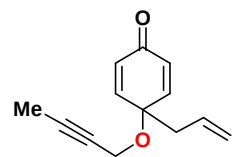


1d (^1H NMR)

Solvent: CDCl_3



S43



1d (^{13}C NMR)

Solvent: CDCl_3

185.249

149.961

131.391
130.844

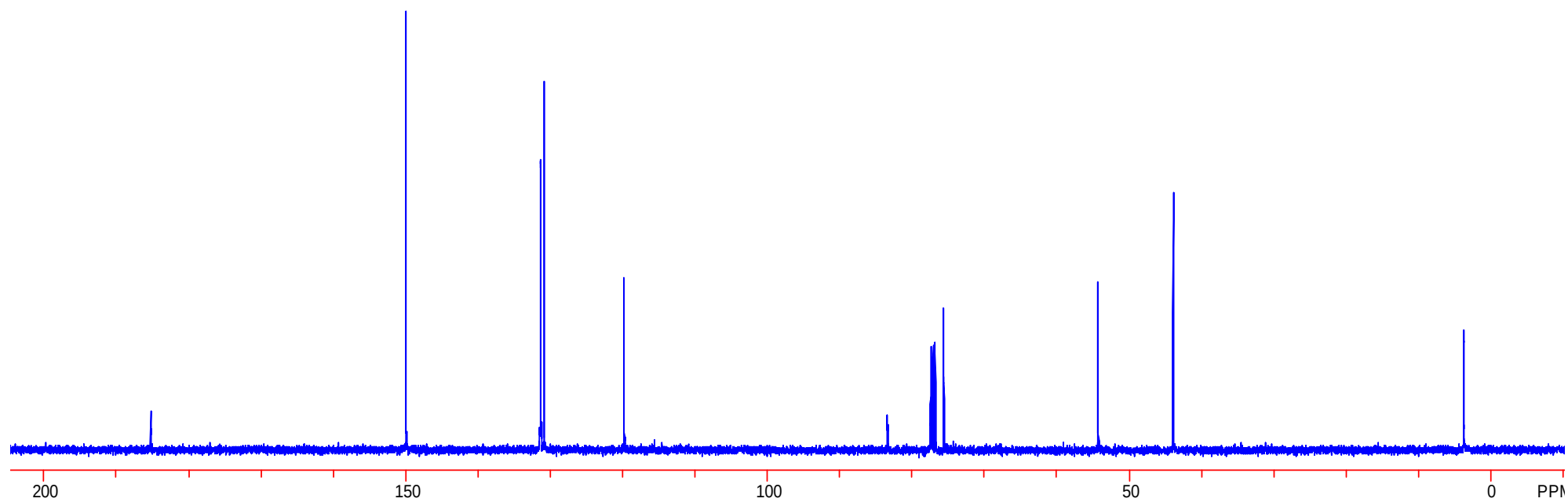
119.798

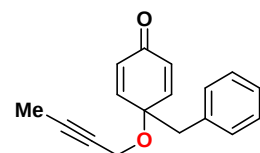
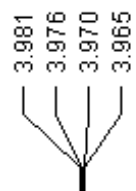
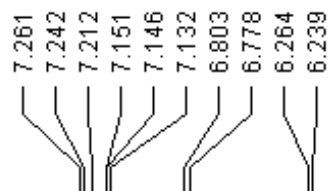
83.386
77.419
77.100
76.781
75.680
75.604

54.309

43.915

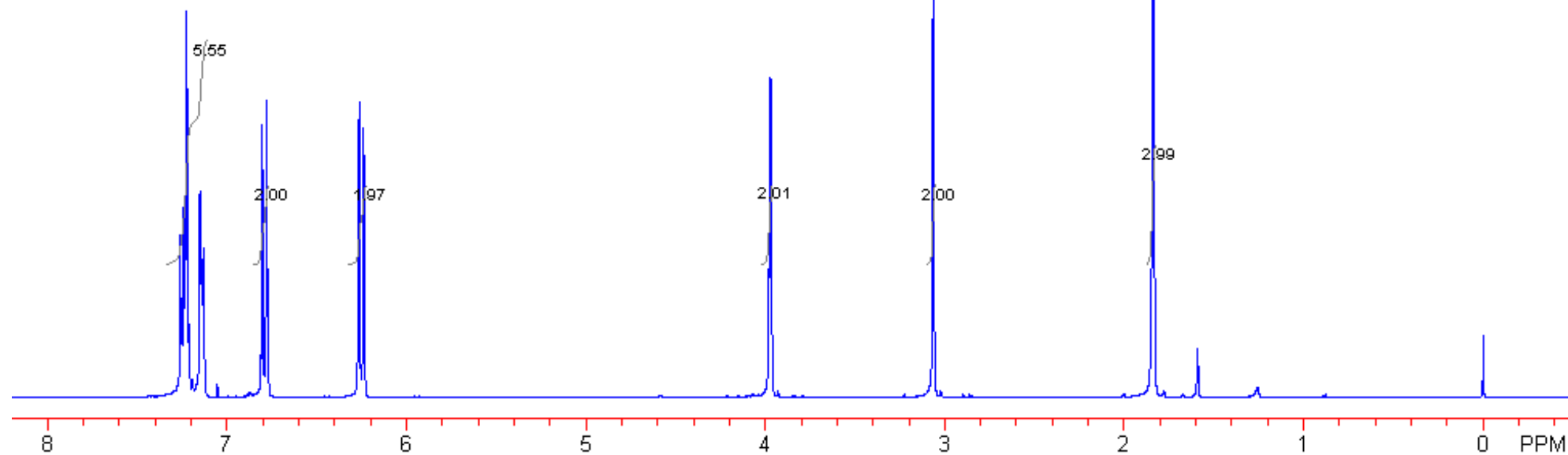
3.723

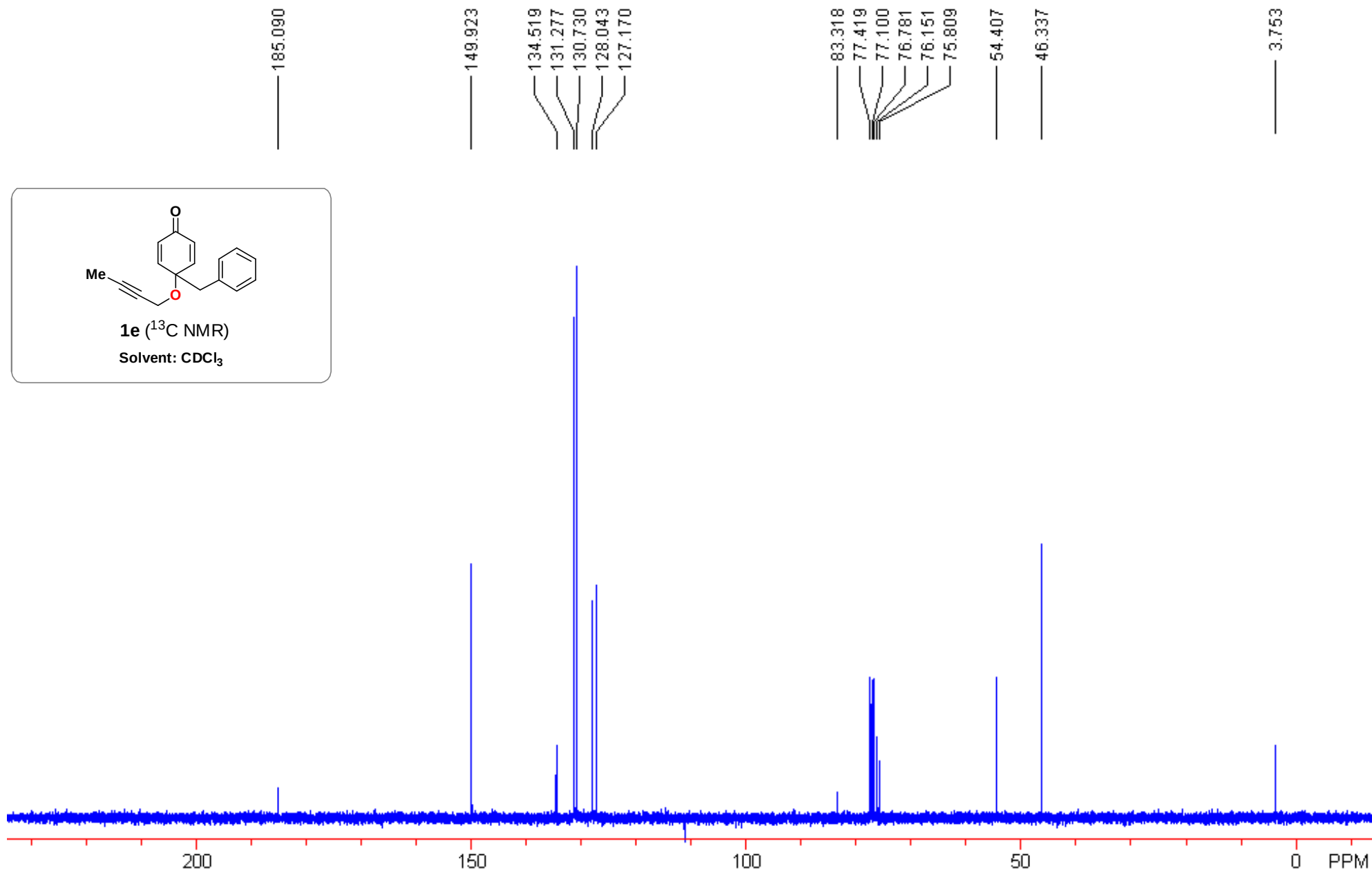


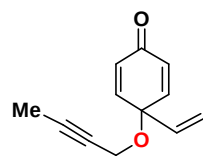


1e (^1H NMR)

Solvent: CDCl_3

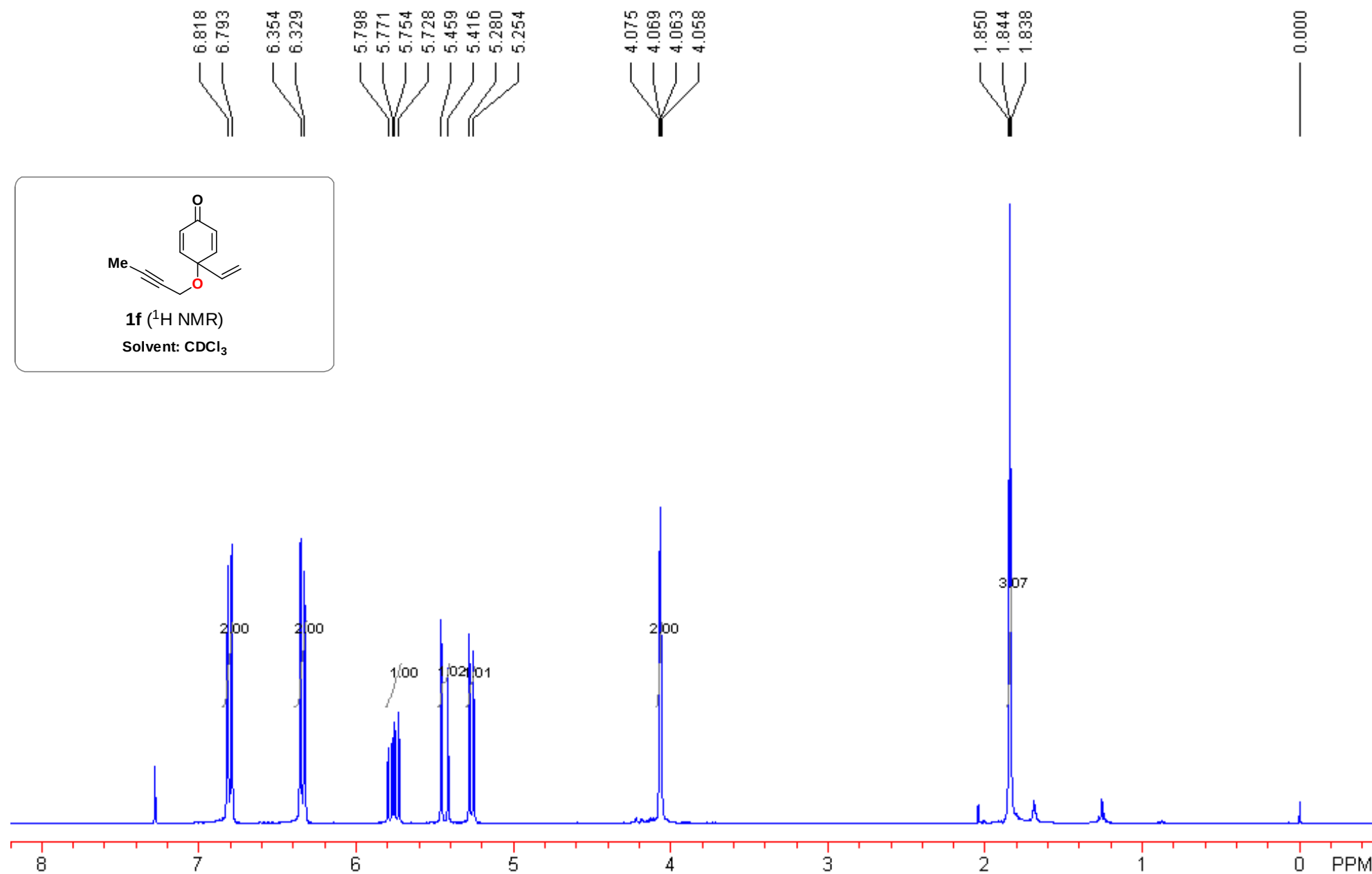


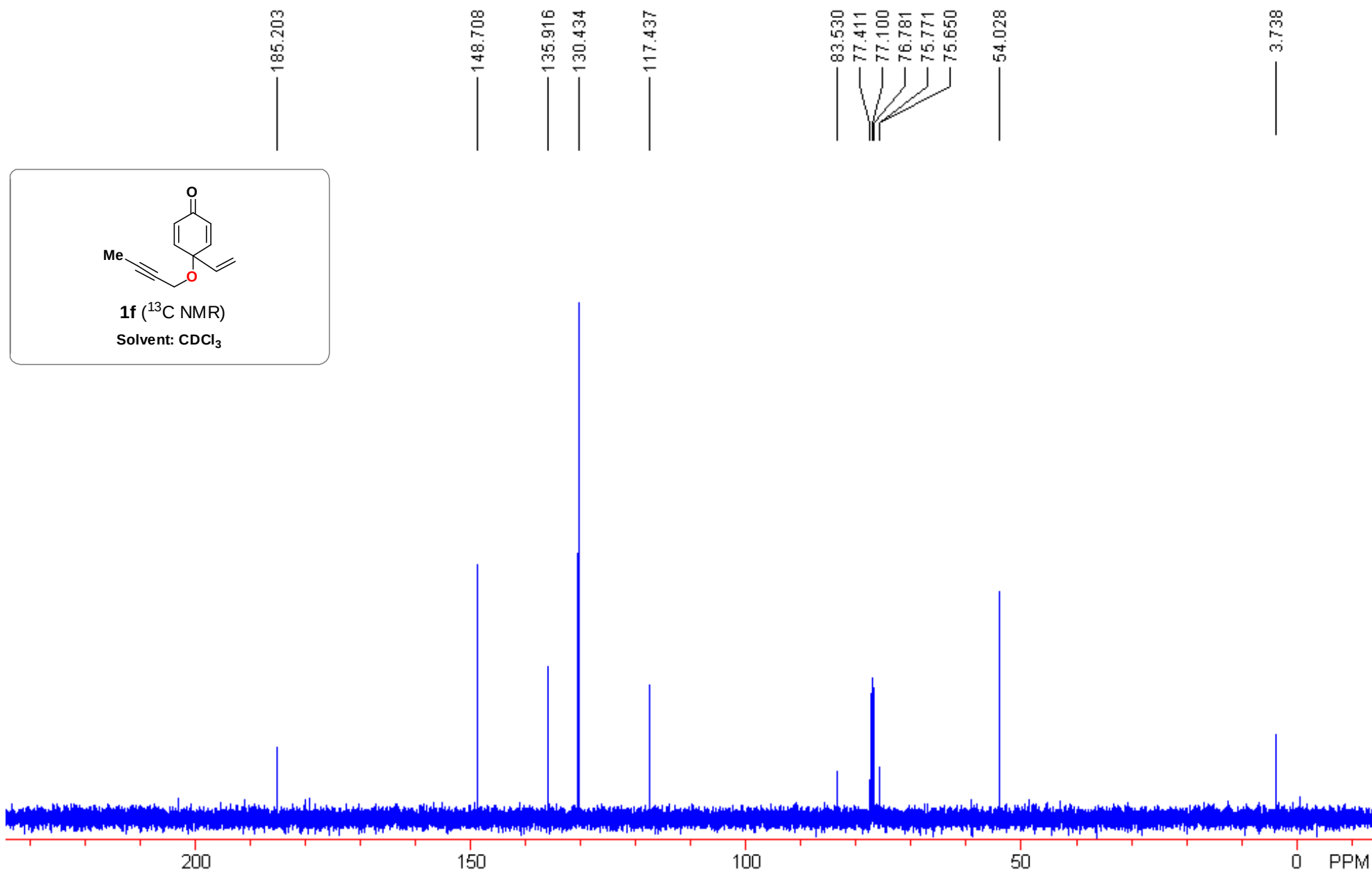


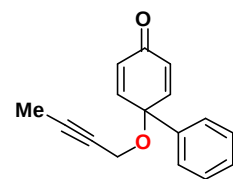
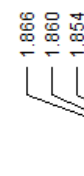
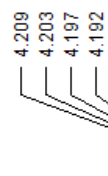
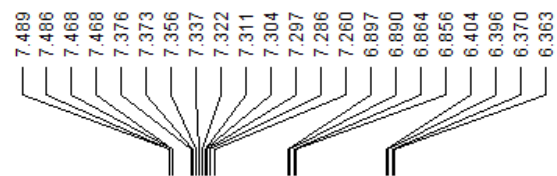


1f (^1H NMR)

Solvent: CDCl_3

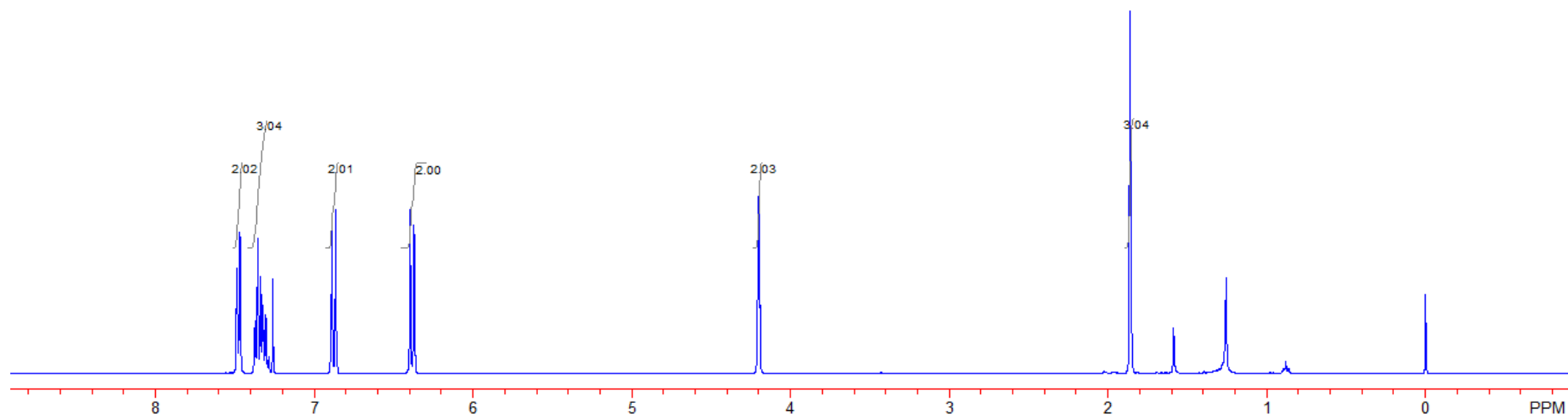


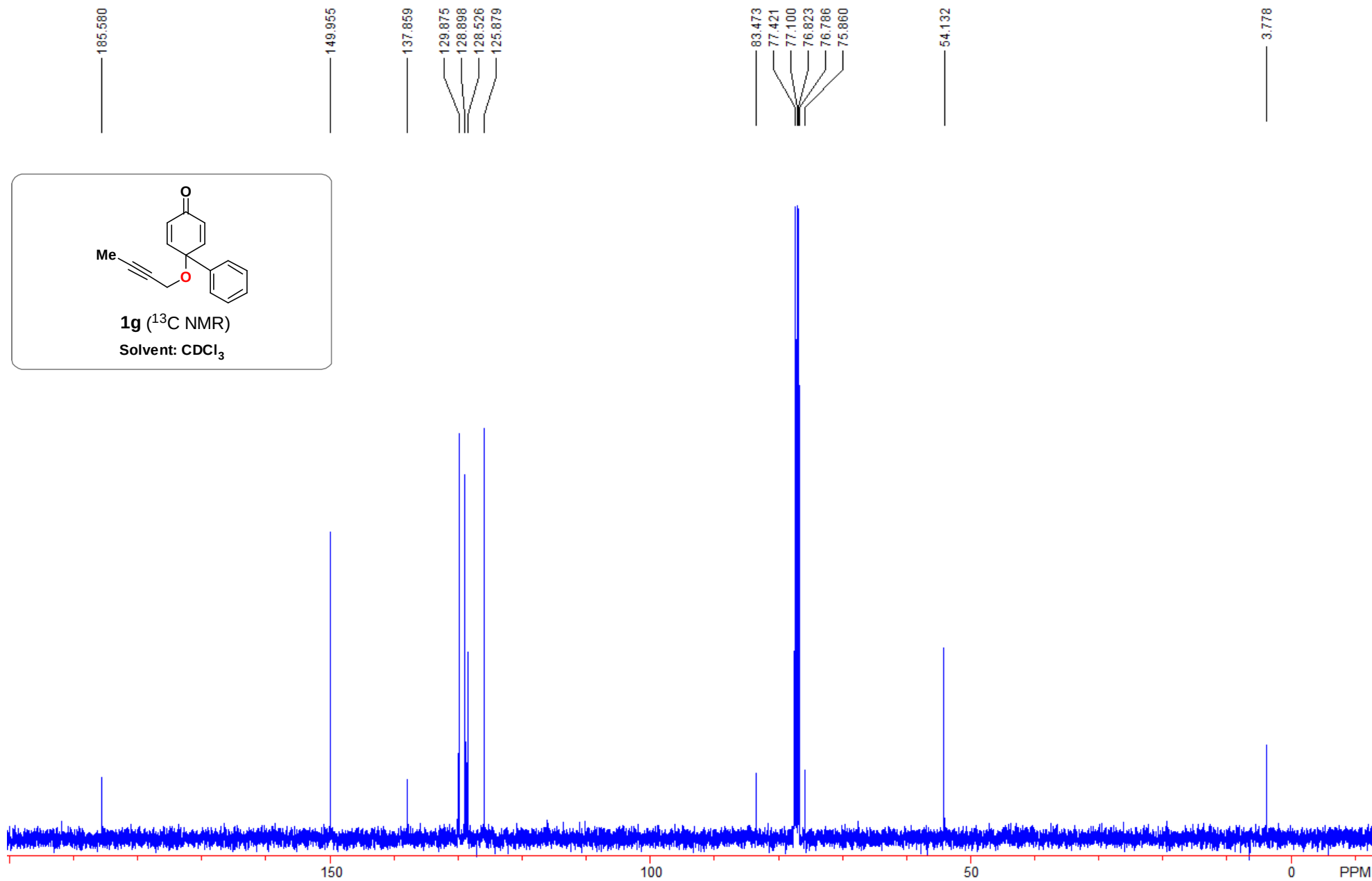


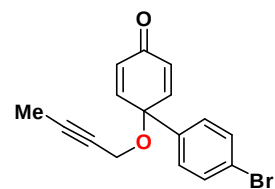
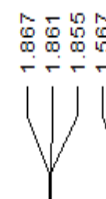
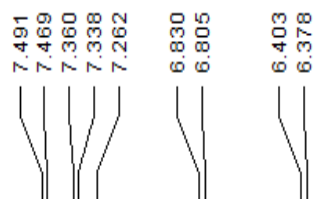


1g (^1H NMR)

Solvent: CDCl_3

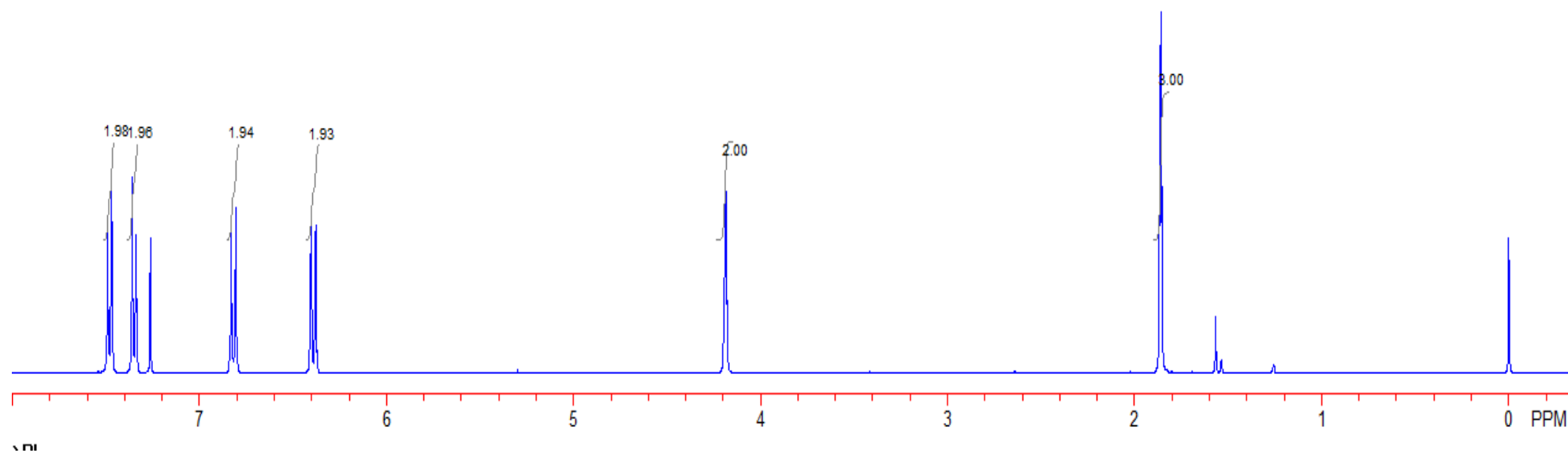


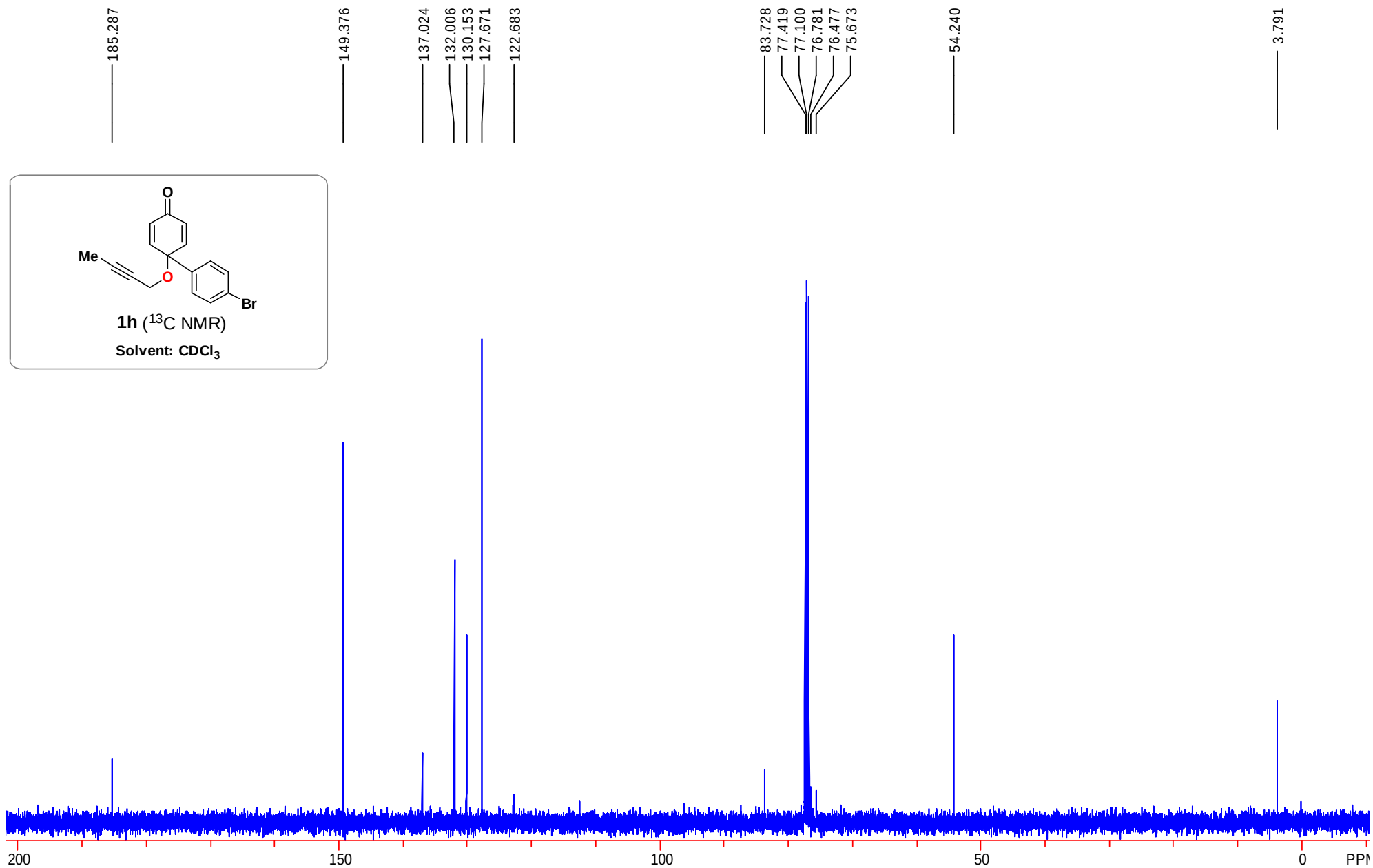


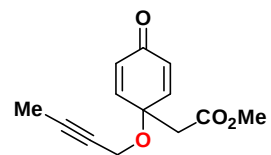


1h (^1H NMR)

Solvent: CDCl_3

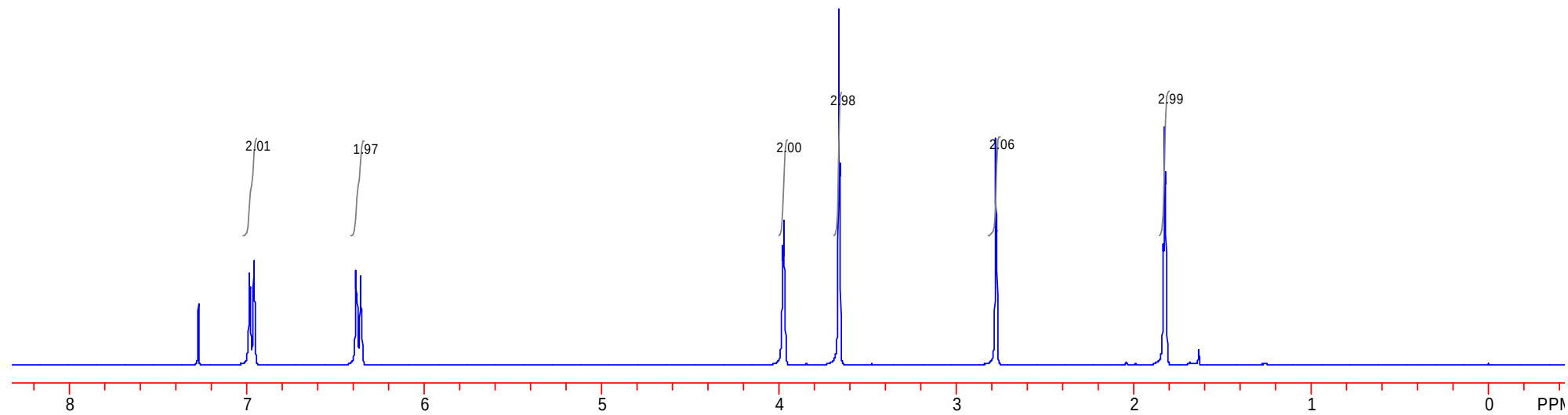
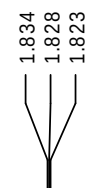
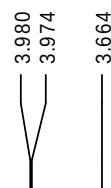
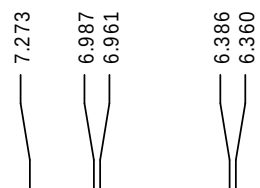




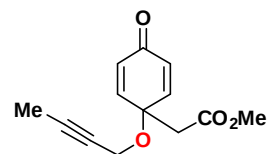


1i (^1H NMR)

Solvent: CDCl_3

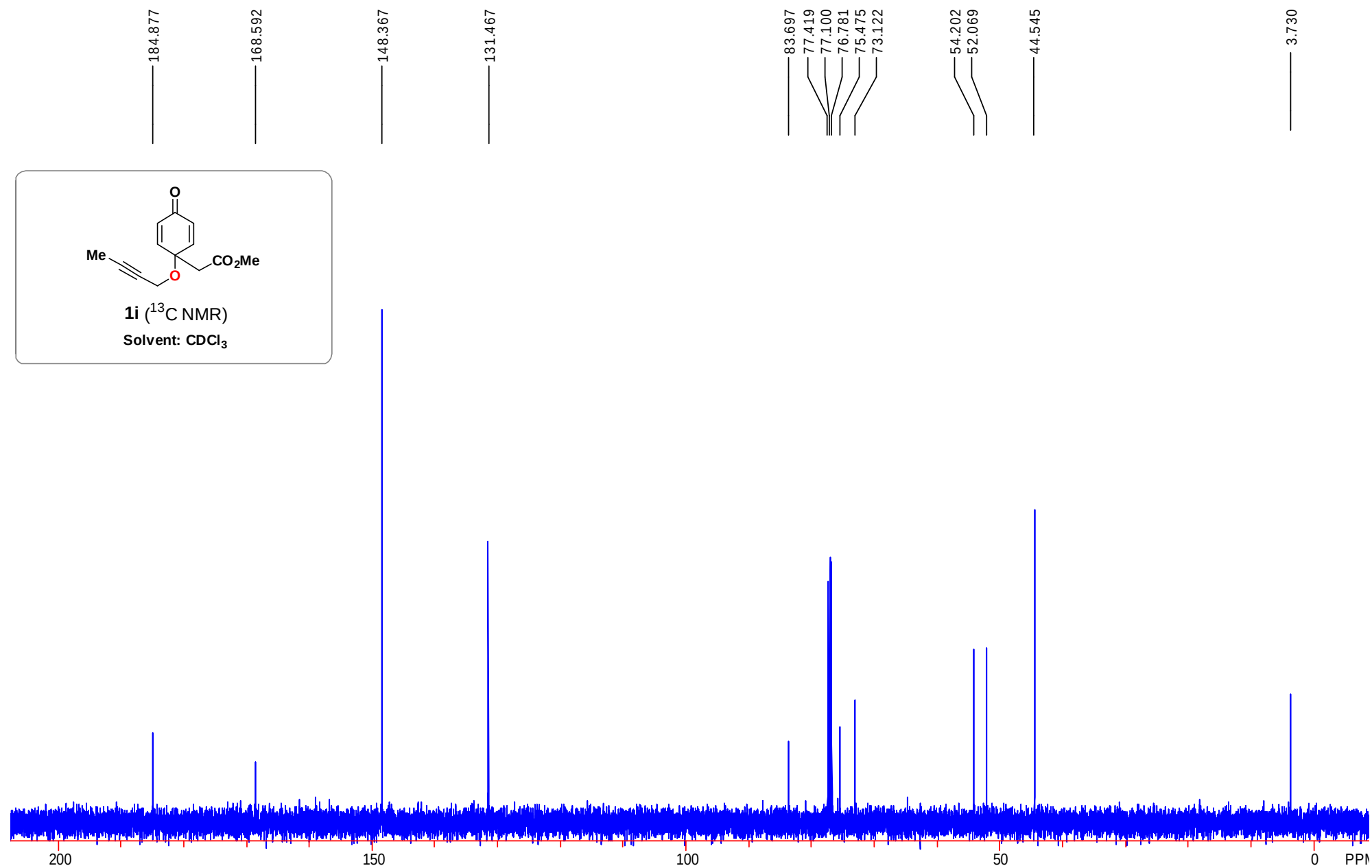


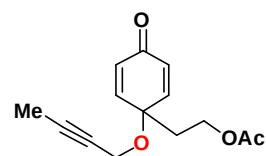
S53



1i (^{13}C NMR)

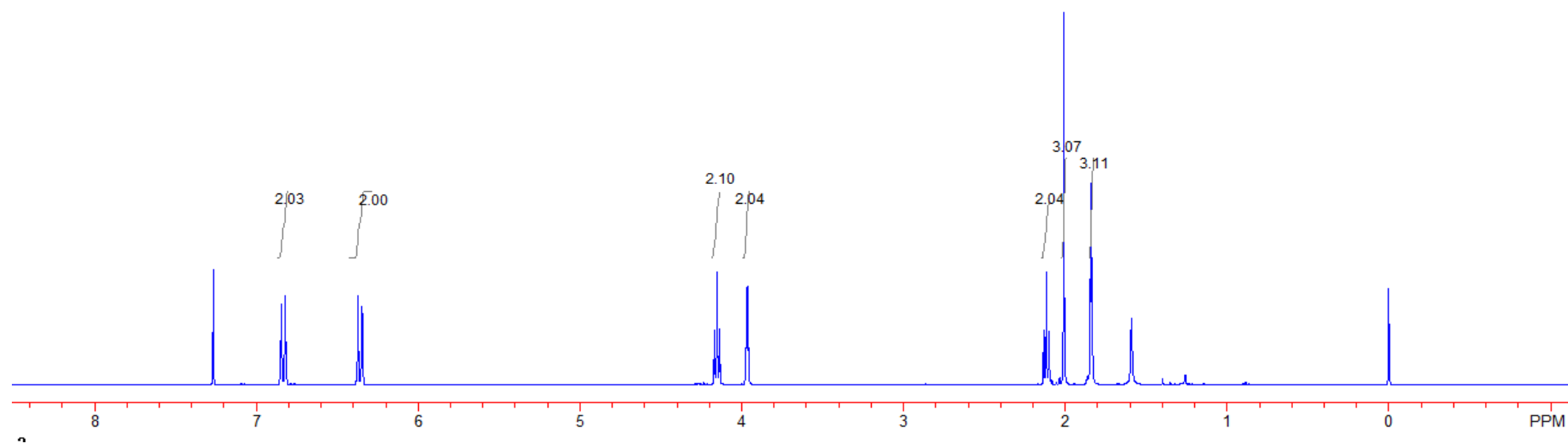
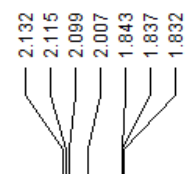
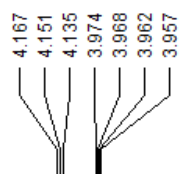
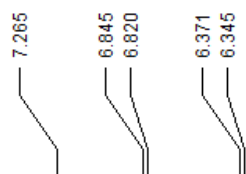
Solvent: CDCl_3

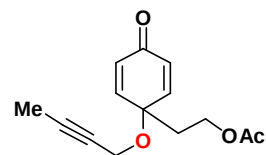




1j (^1H NMR)

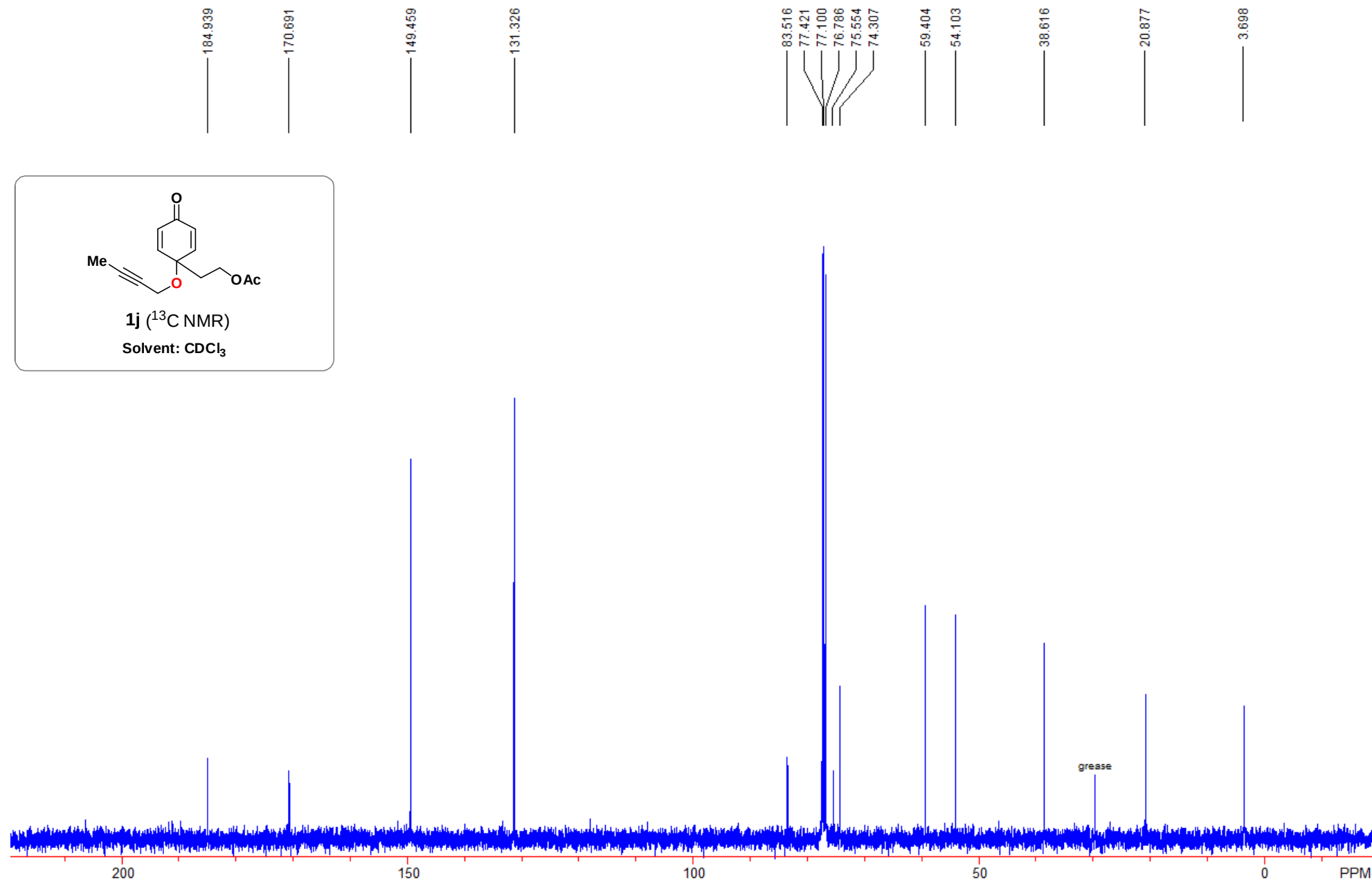
Solvent: CDCl_3





1j (^{13}C NMR)

Solvent: CDCl_3

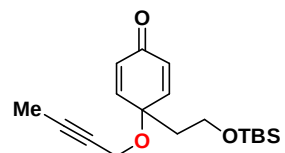
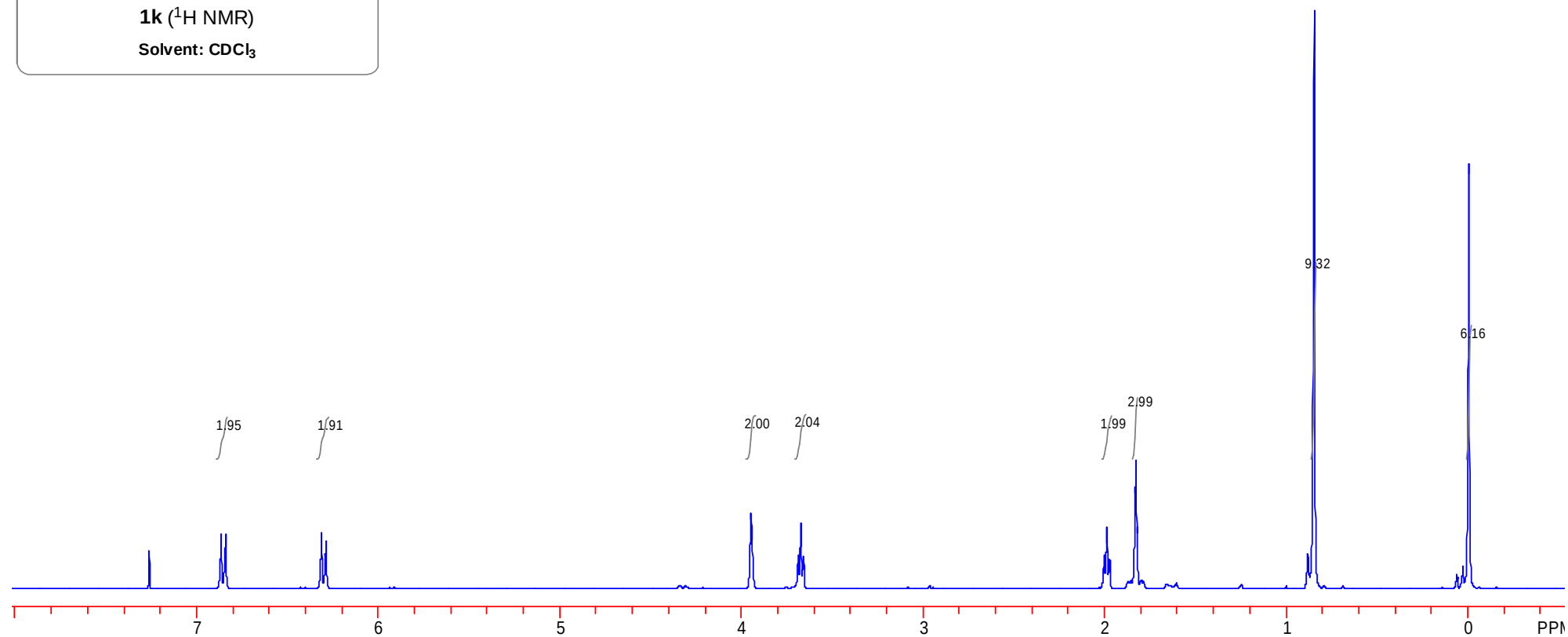


7.267

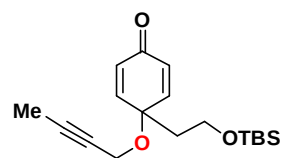
6.871
6.8456.318
6.2933.957
3.952
3.946
3.693
3.678
3.6632.007
1.992
1.977
1.837
1.831
1.826

0.850

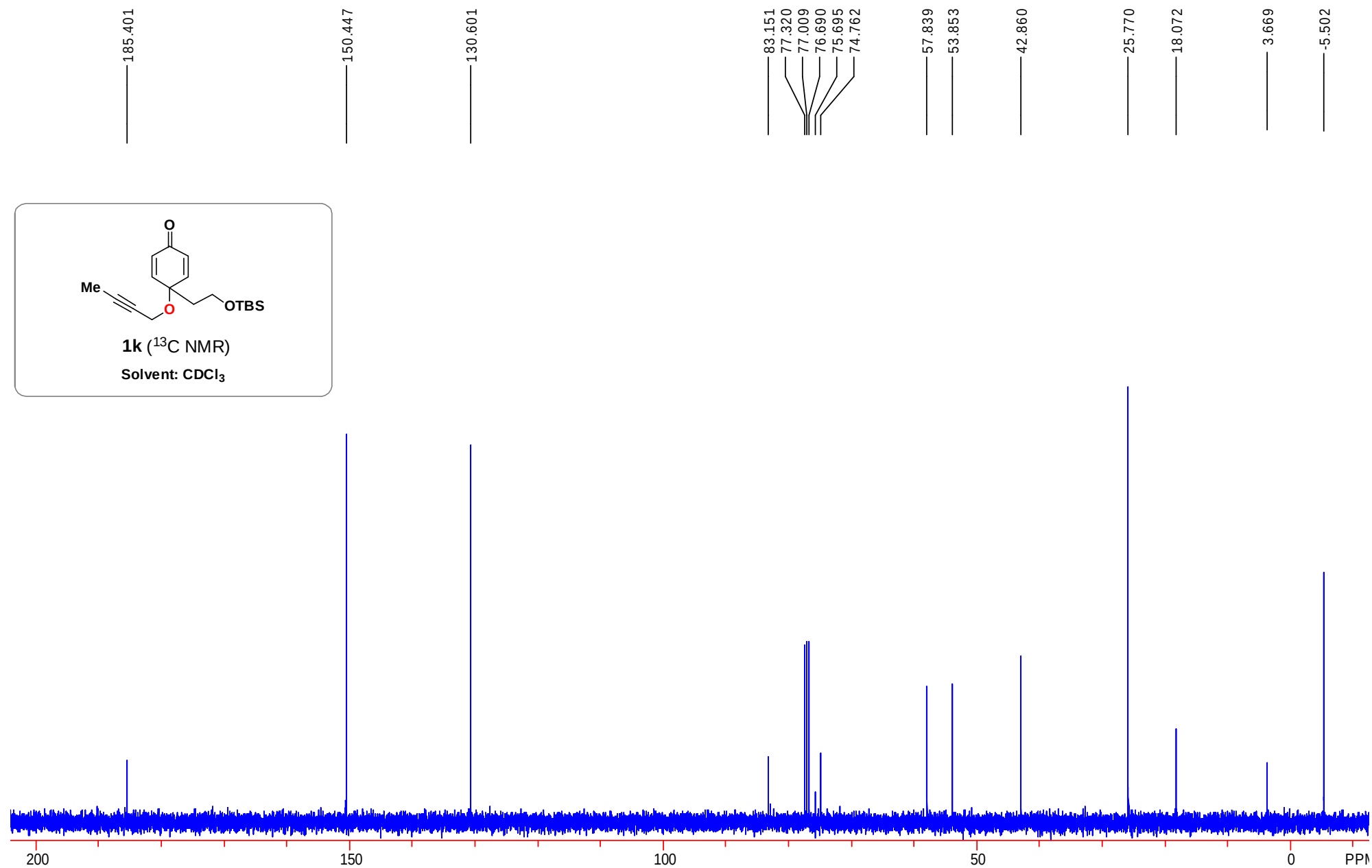
-0.001

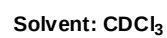
**1k** (^1H NMR)Solvent: CDCl_3 

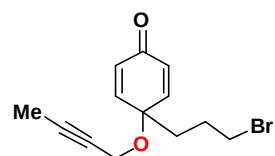
S57



1k (^{13}C NMR)
Solvent: CDCl_3

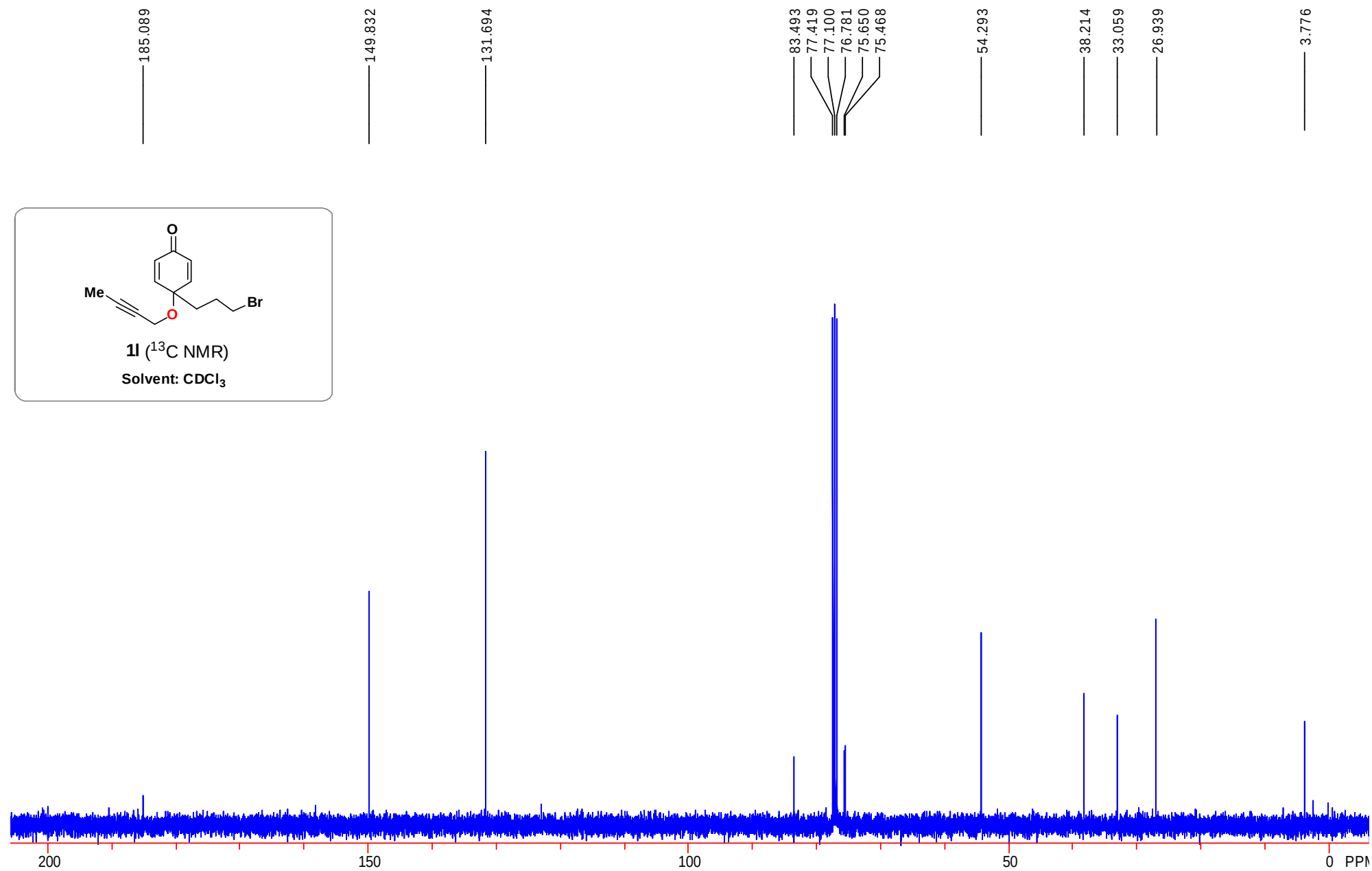






1l (^{13}C NMR)

Solvent: CDCl_3



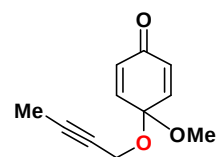
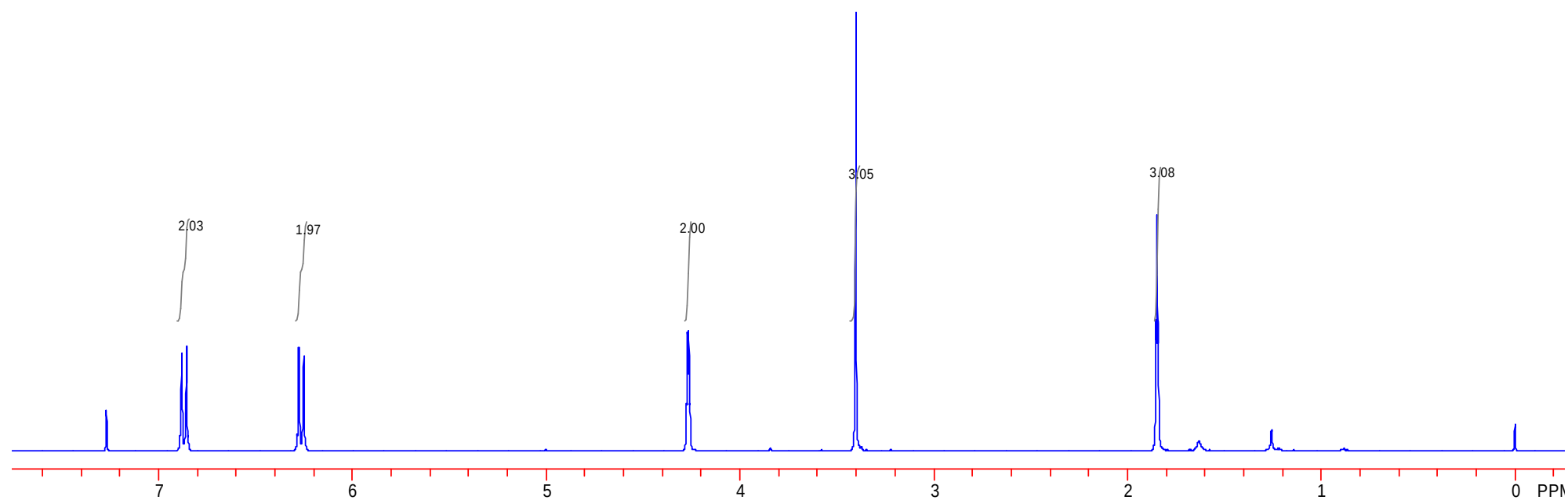
7.270

6.882
6.8576.277
6.2524.275
4.270
4.264
4.258

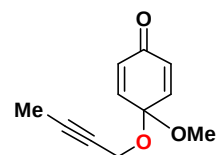
3.402

1.852
1.846
1.841
1.631

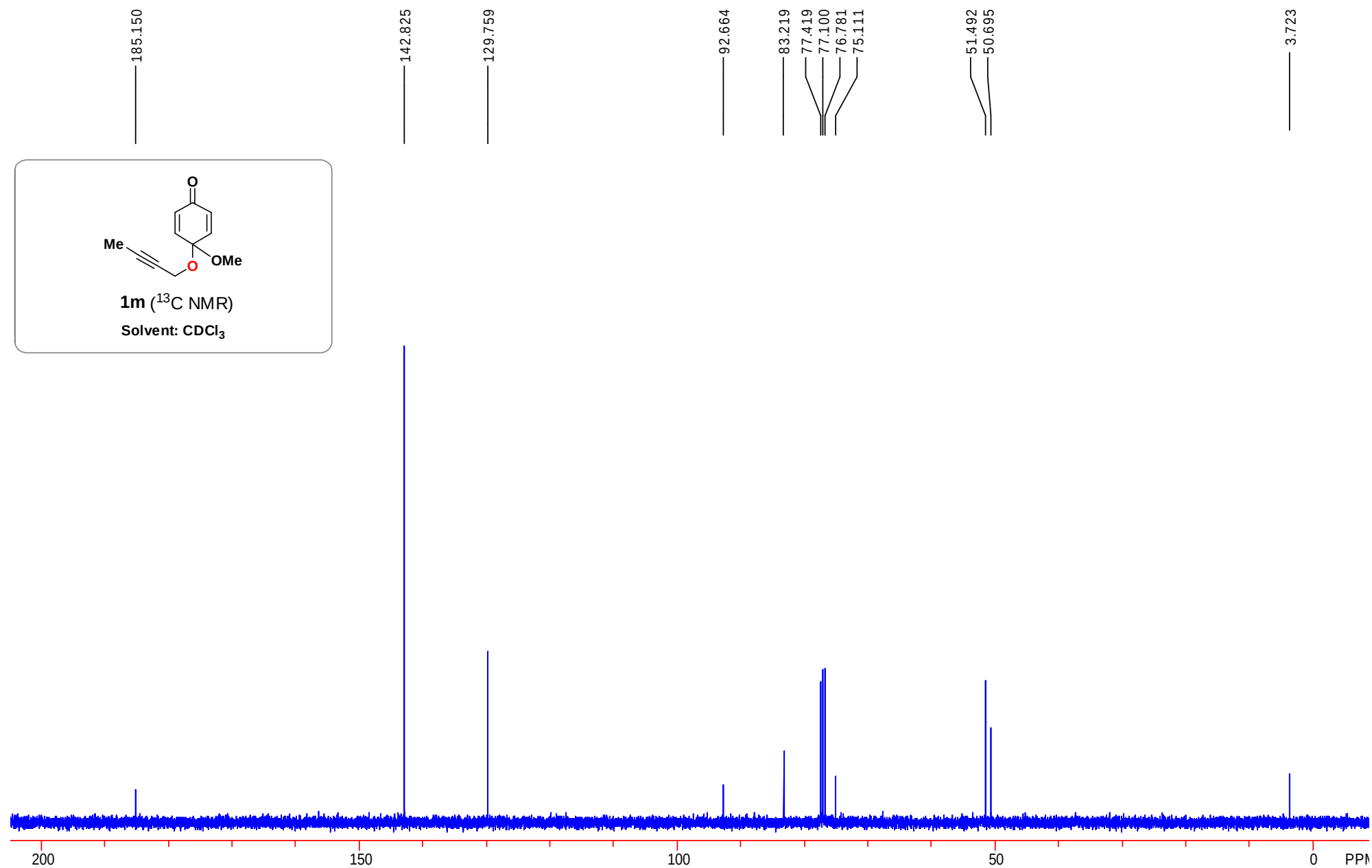
0.000

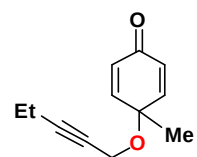
**1m** (^1H NMR)Solvent: CDCl_3 

S61



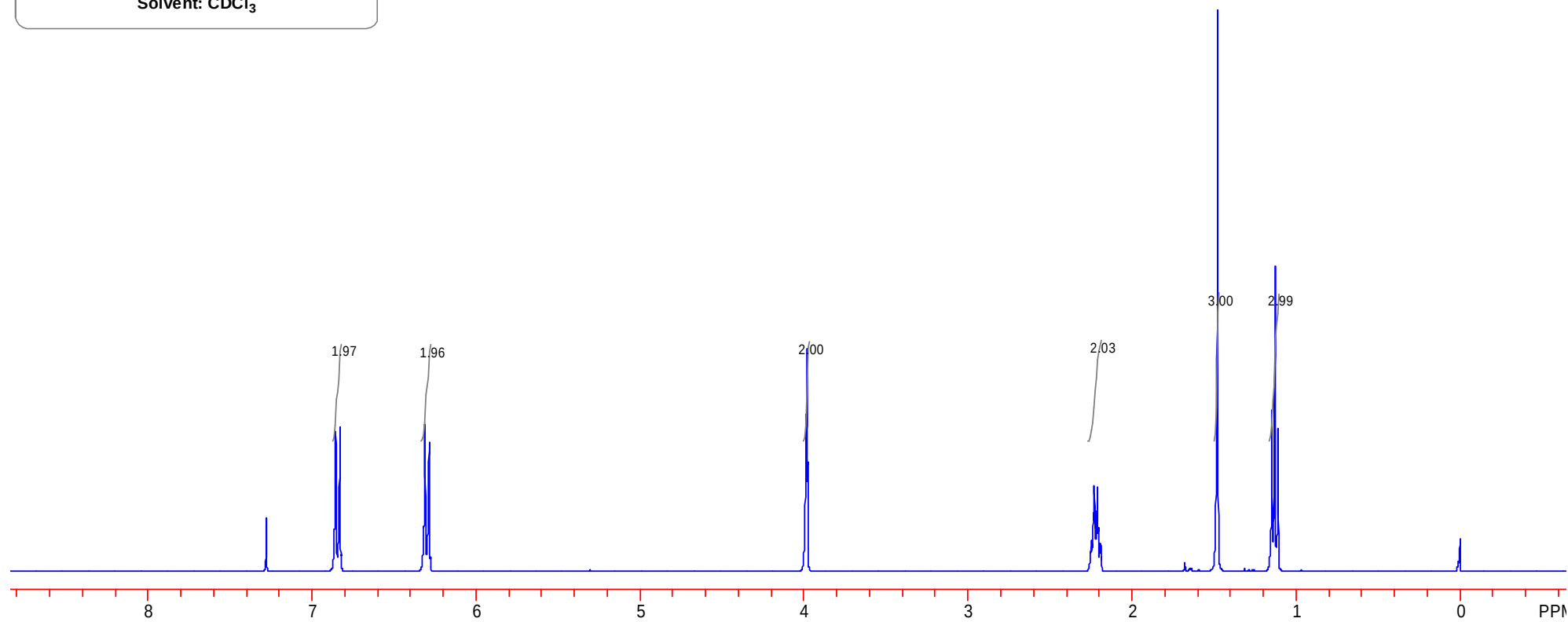
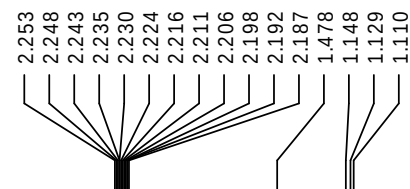
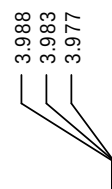
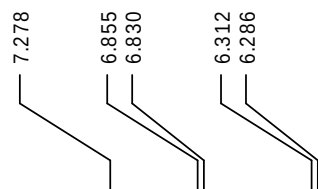
1m (^{13}C NMR)
Solvent: CDCl_3



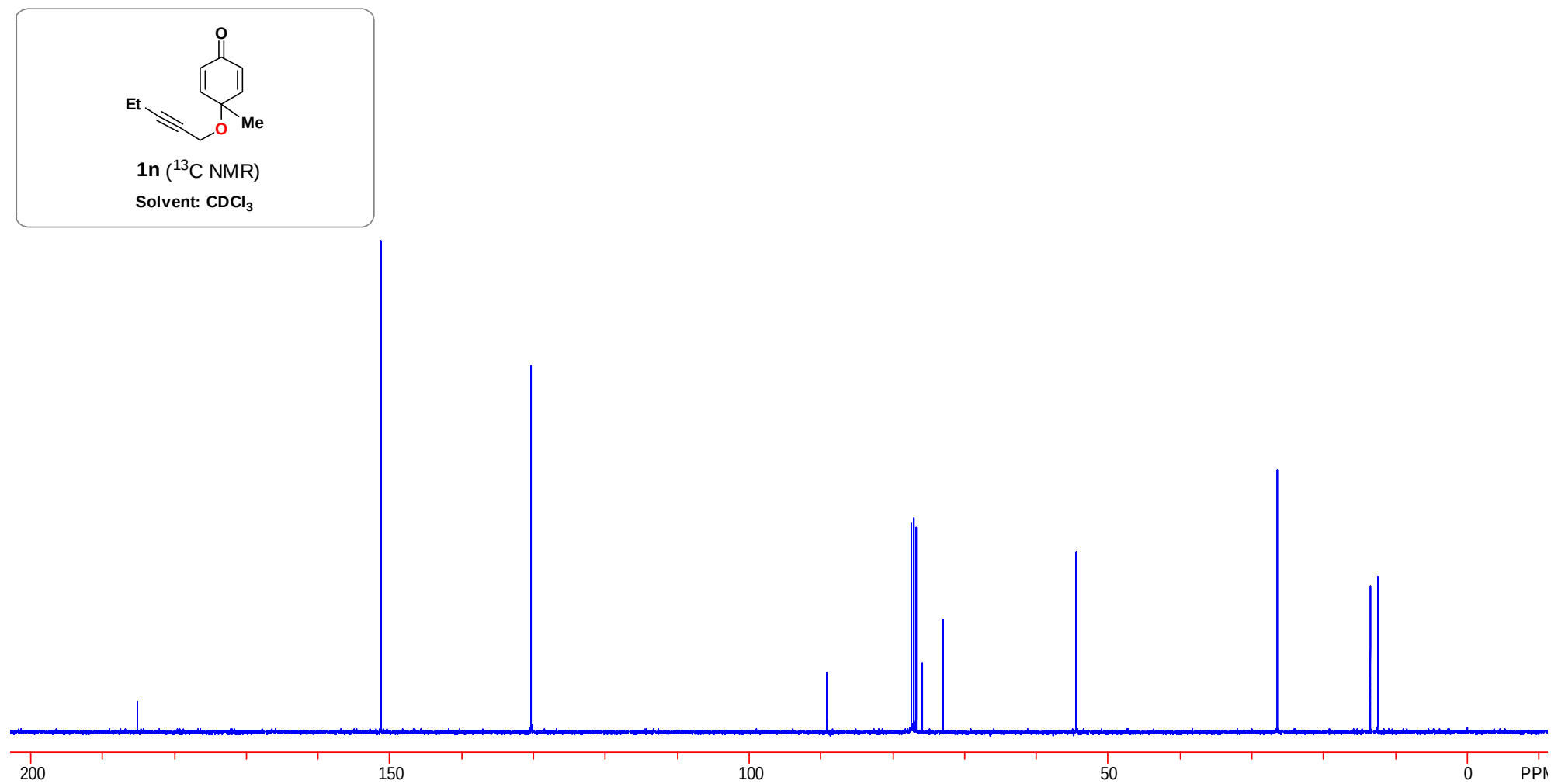


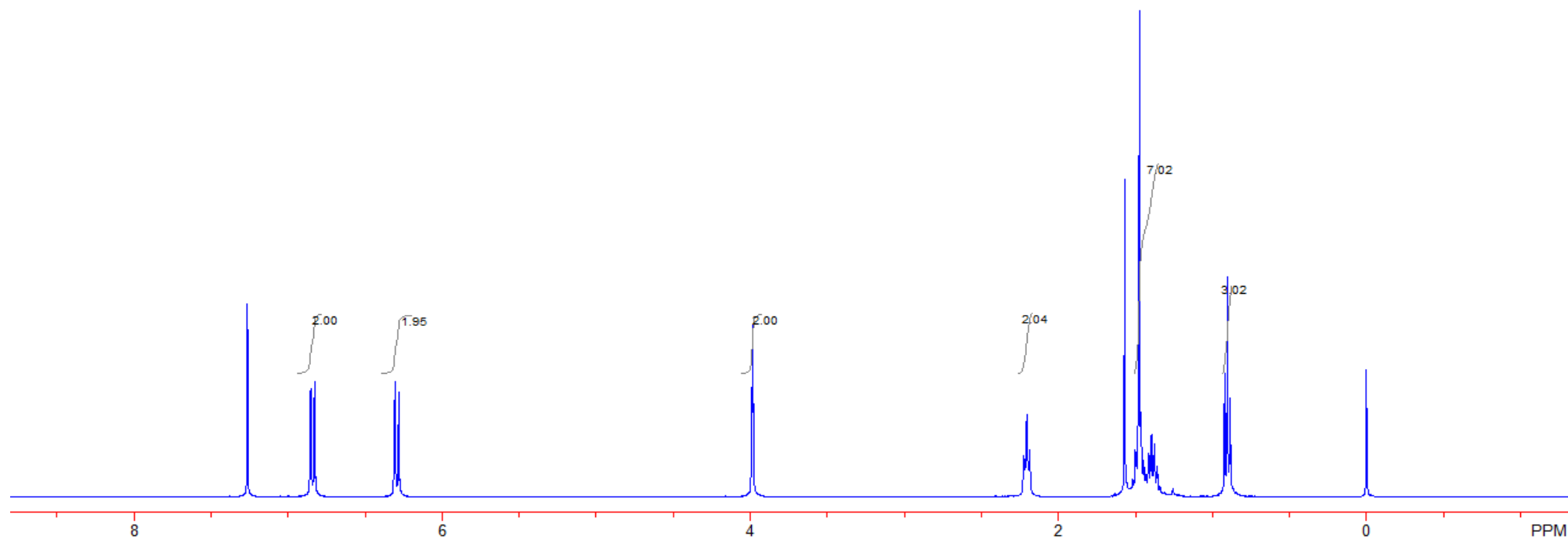
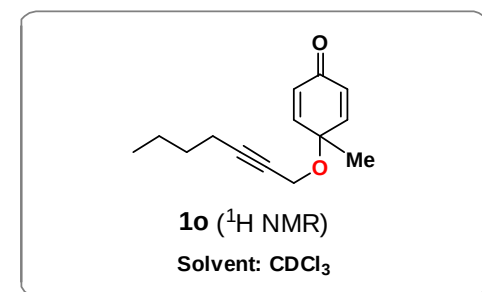
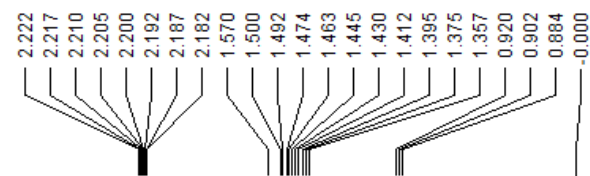
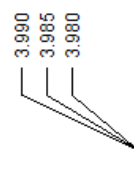
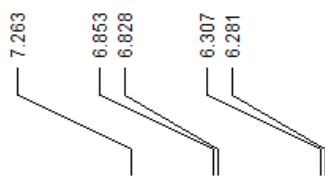
1n (^1H NMR)

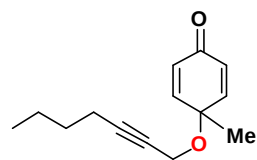
Solvent: CDCl_3



S63

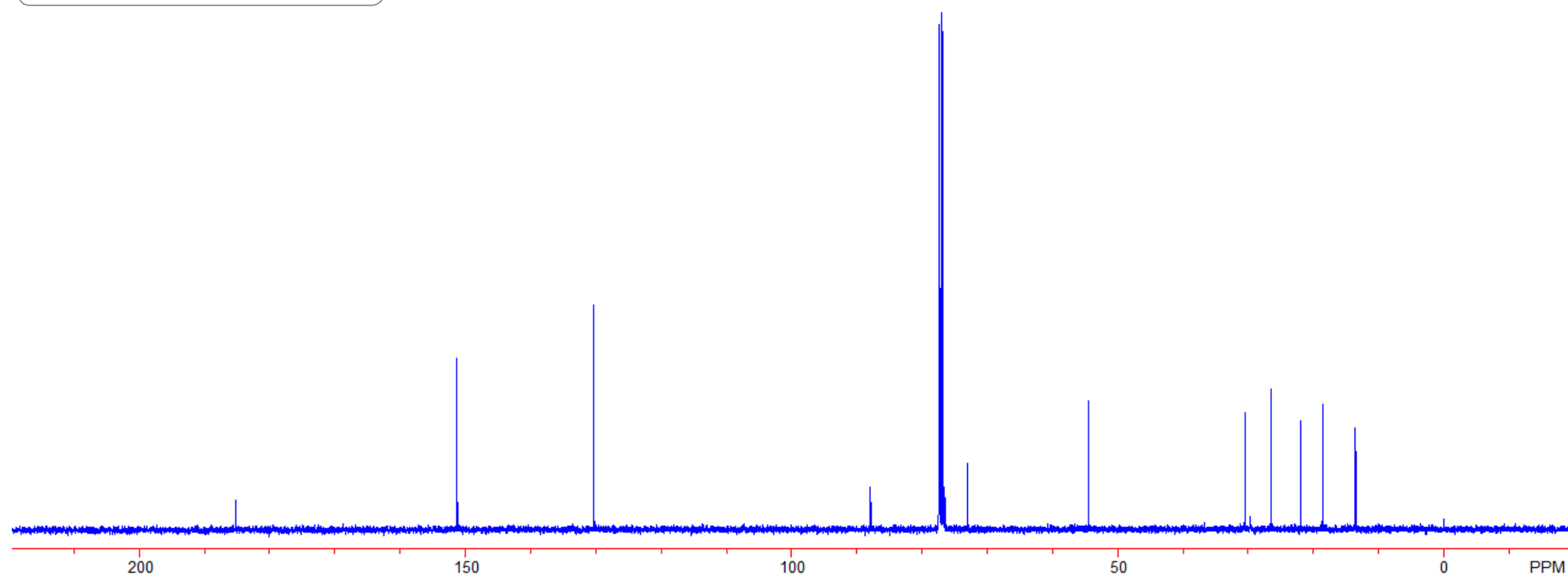


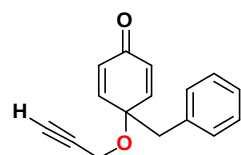
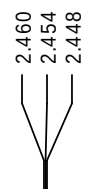
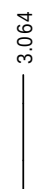
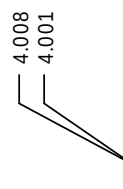
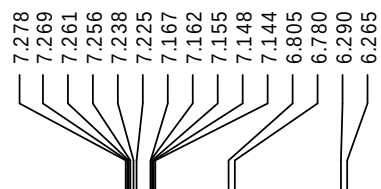




1o (^{13}C NMR)

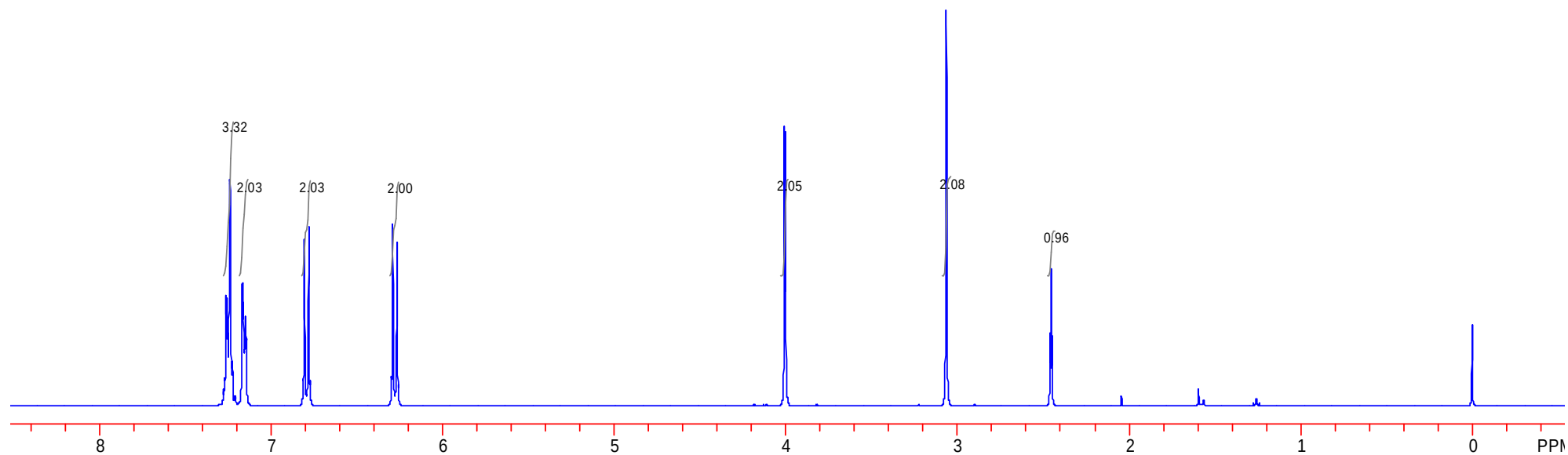
Solvent: CDCl_3



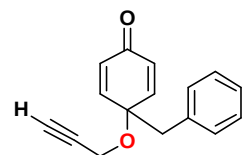


1p (^1H NMR)

Solvent: CDCl_3



S67



1p (^{13}C NMR)

Solvent: CDCl_3

185.013

149.536

134.443

131.550

130.783

128.119

127.261

80.448

77.419

77.100

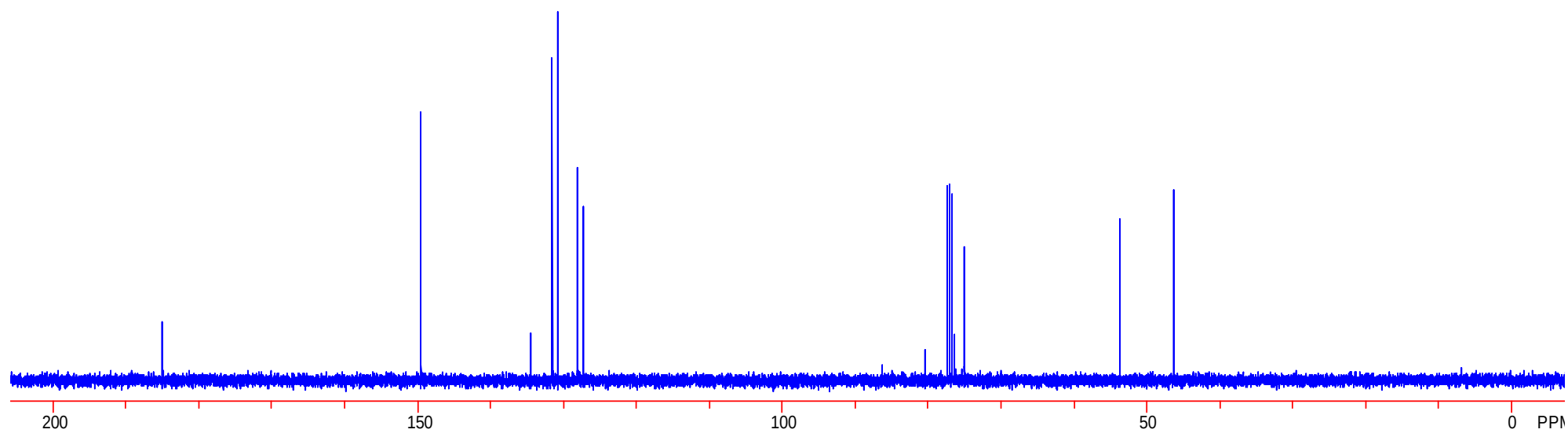
76.781

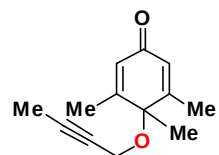
76.371

75.005

53.800

46.307





1q (^1H NMR)

Solvent: CDCl_3

7.272

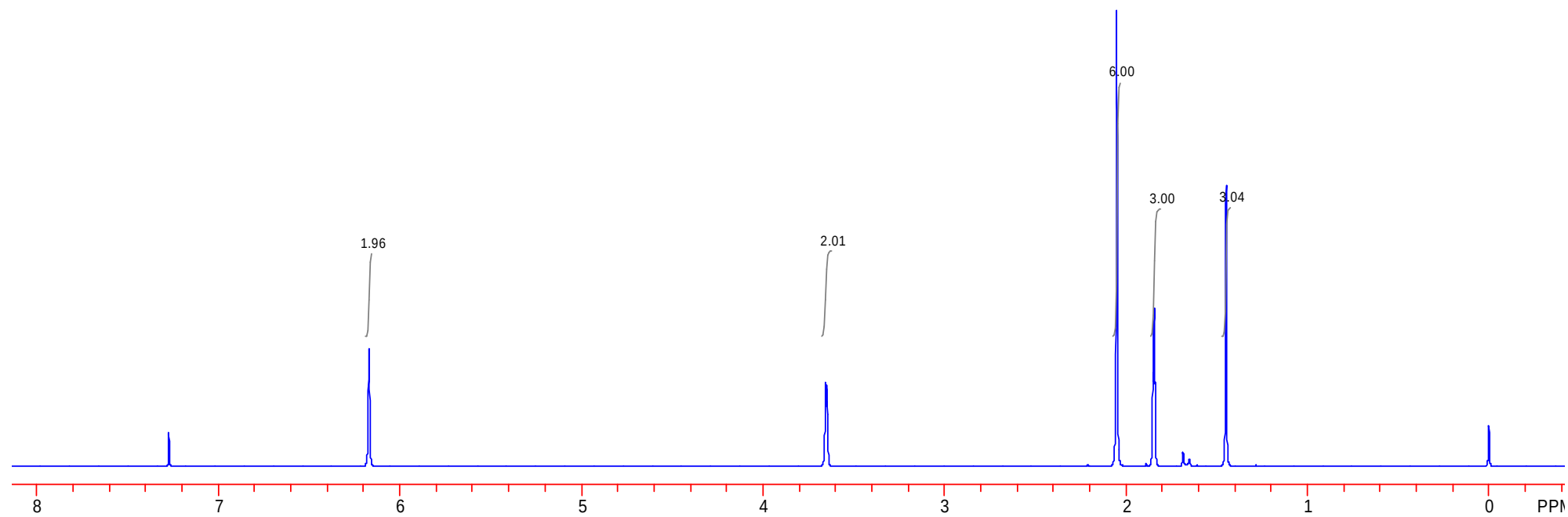
6.169

3.653
3.648

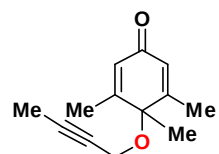
2.051
1.852
1.846
1.841

1.449

0.000

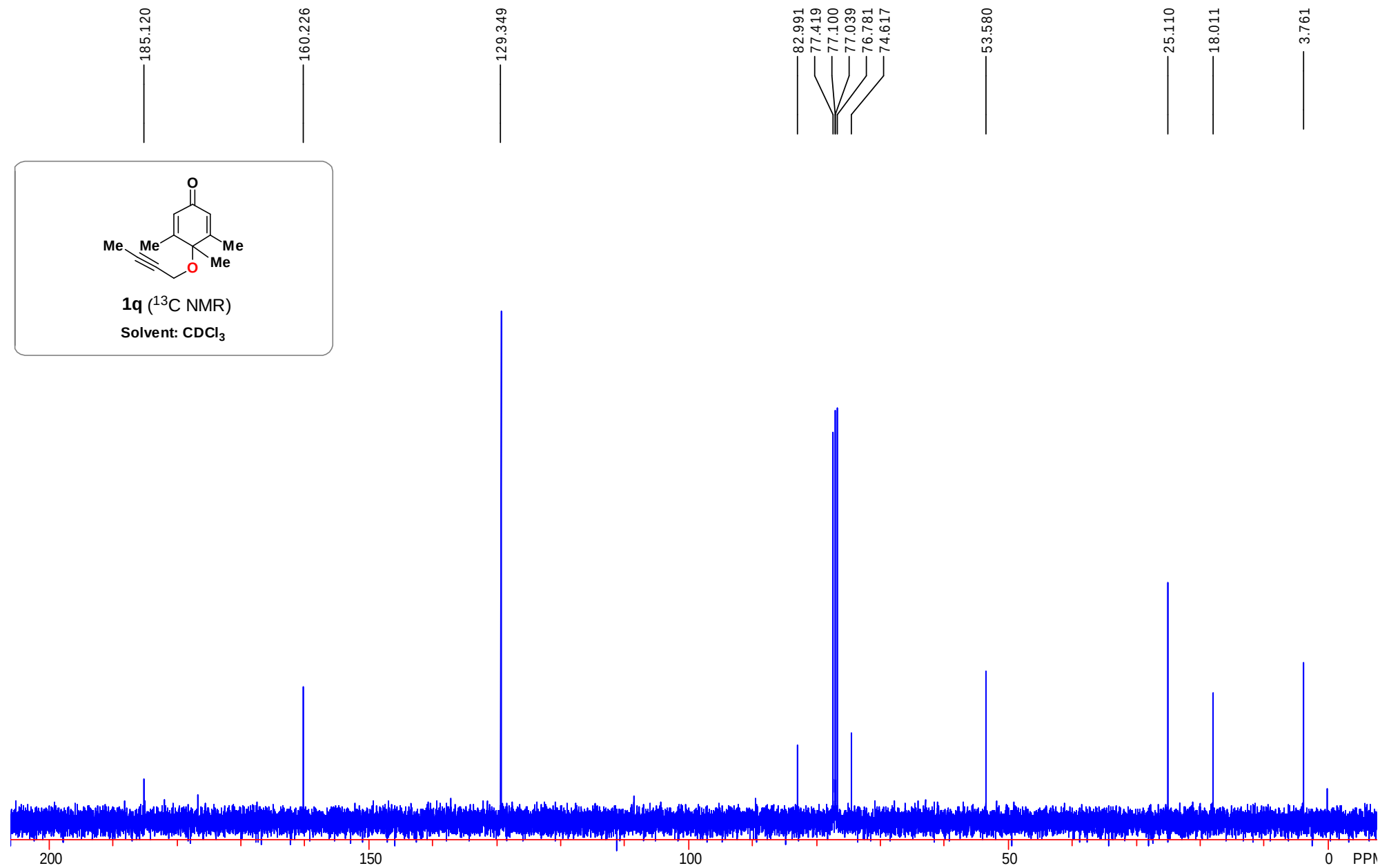


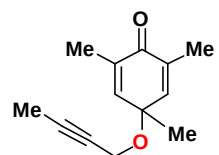
S69



1q (^{13}C NMR)

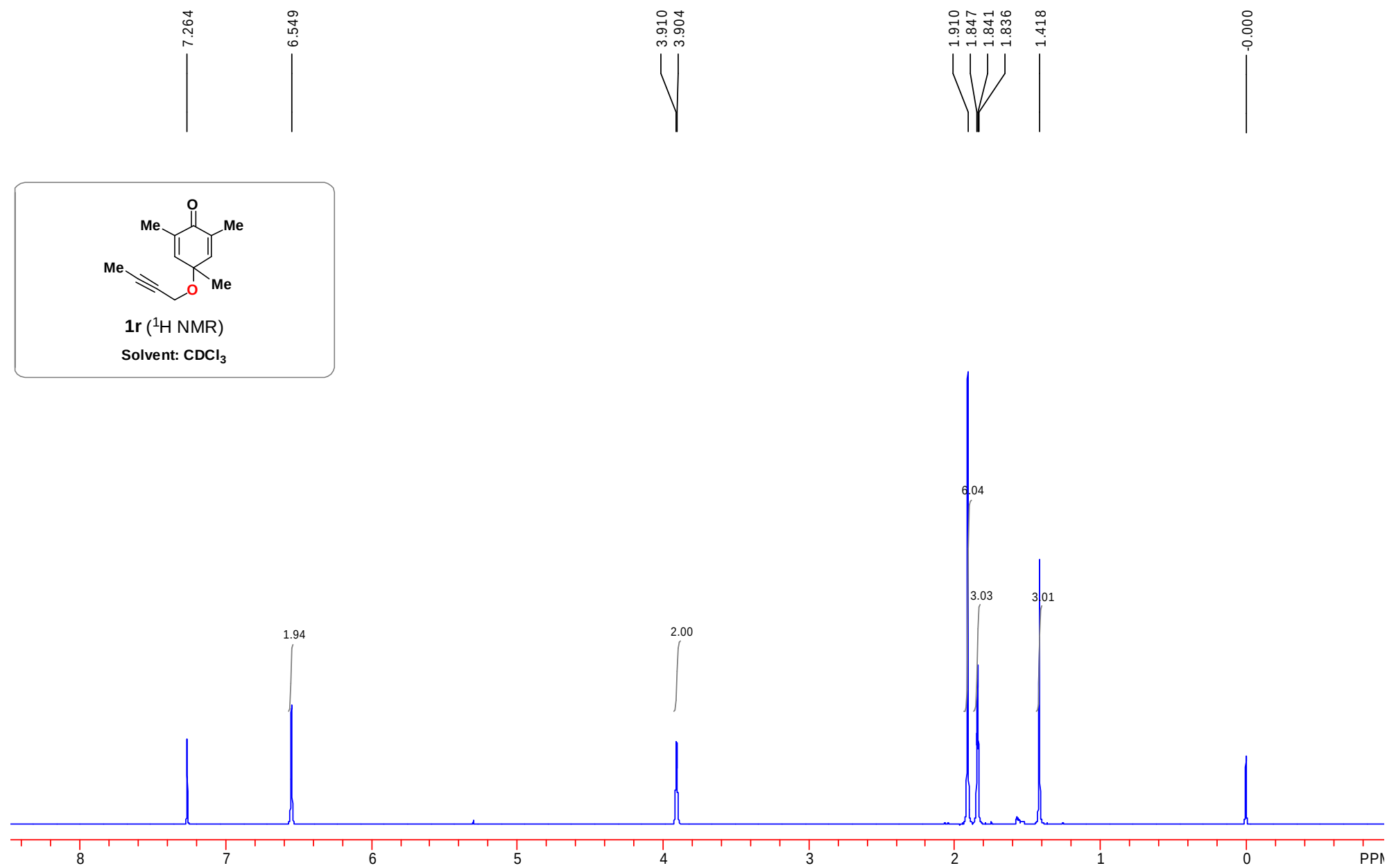
Solvent: CDCl_3

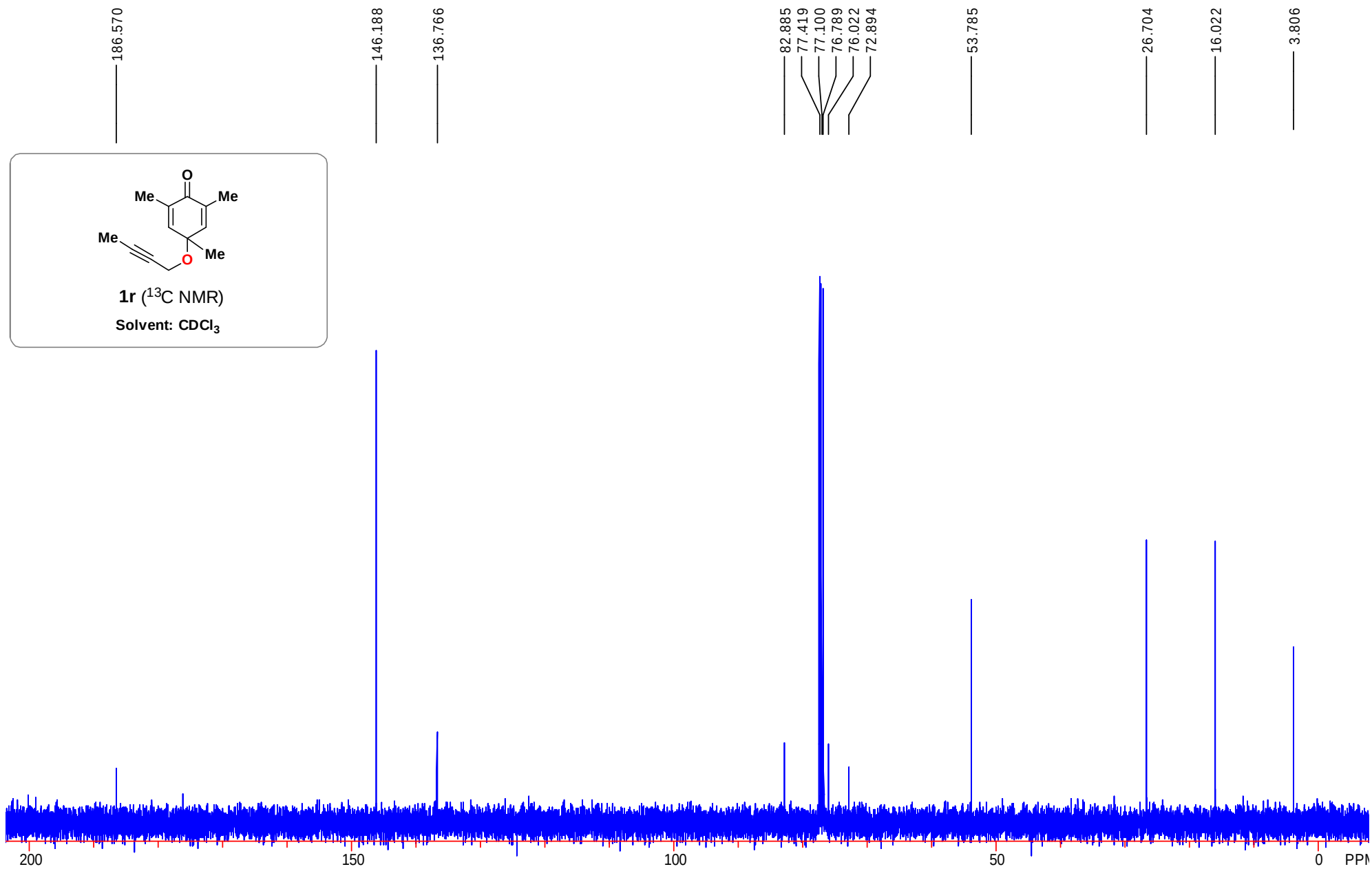


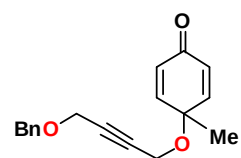


1r (^1H NMR)

Solvent: CDCl_3

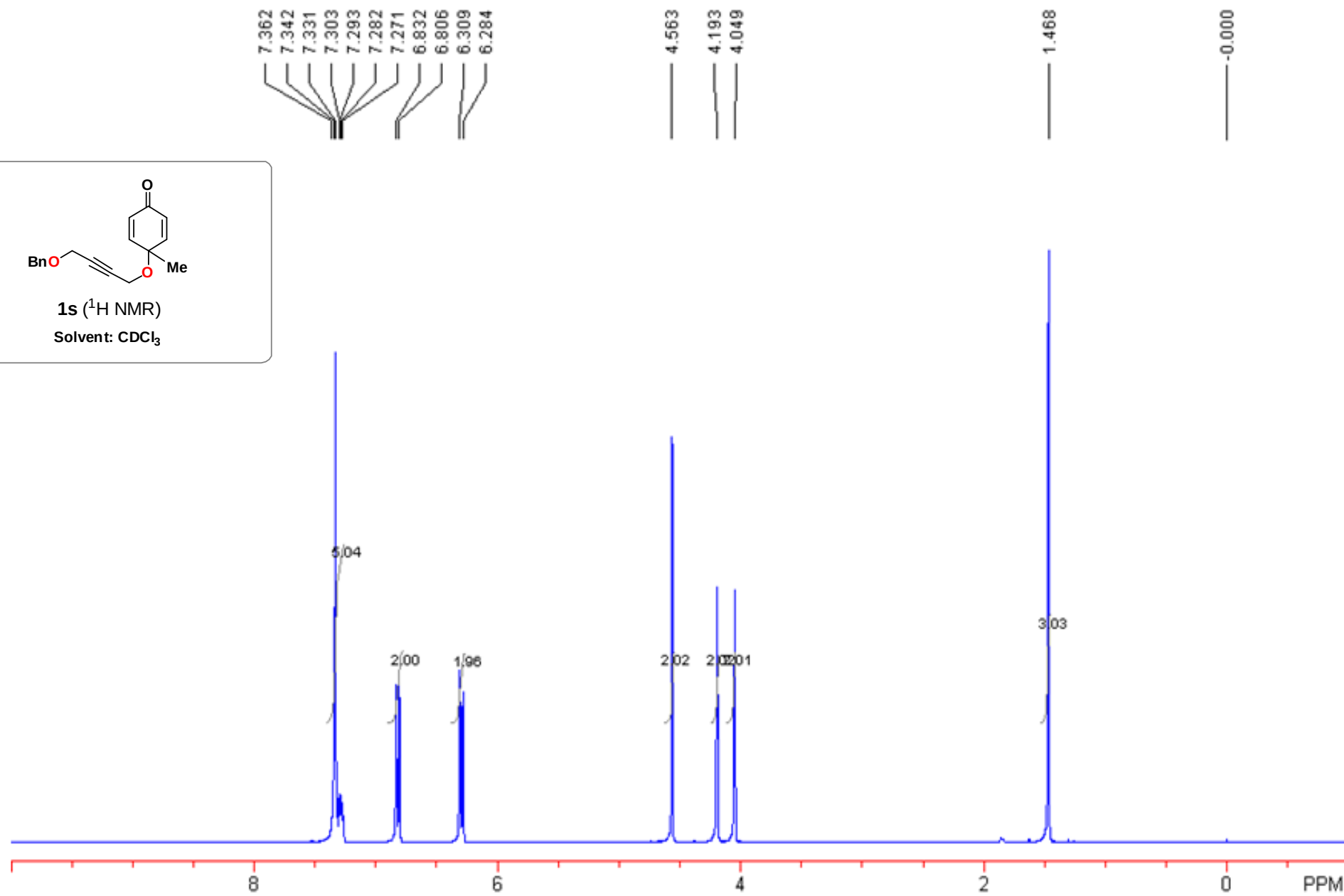


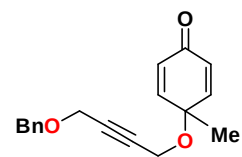




1s (^1H NMR)

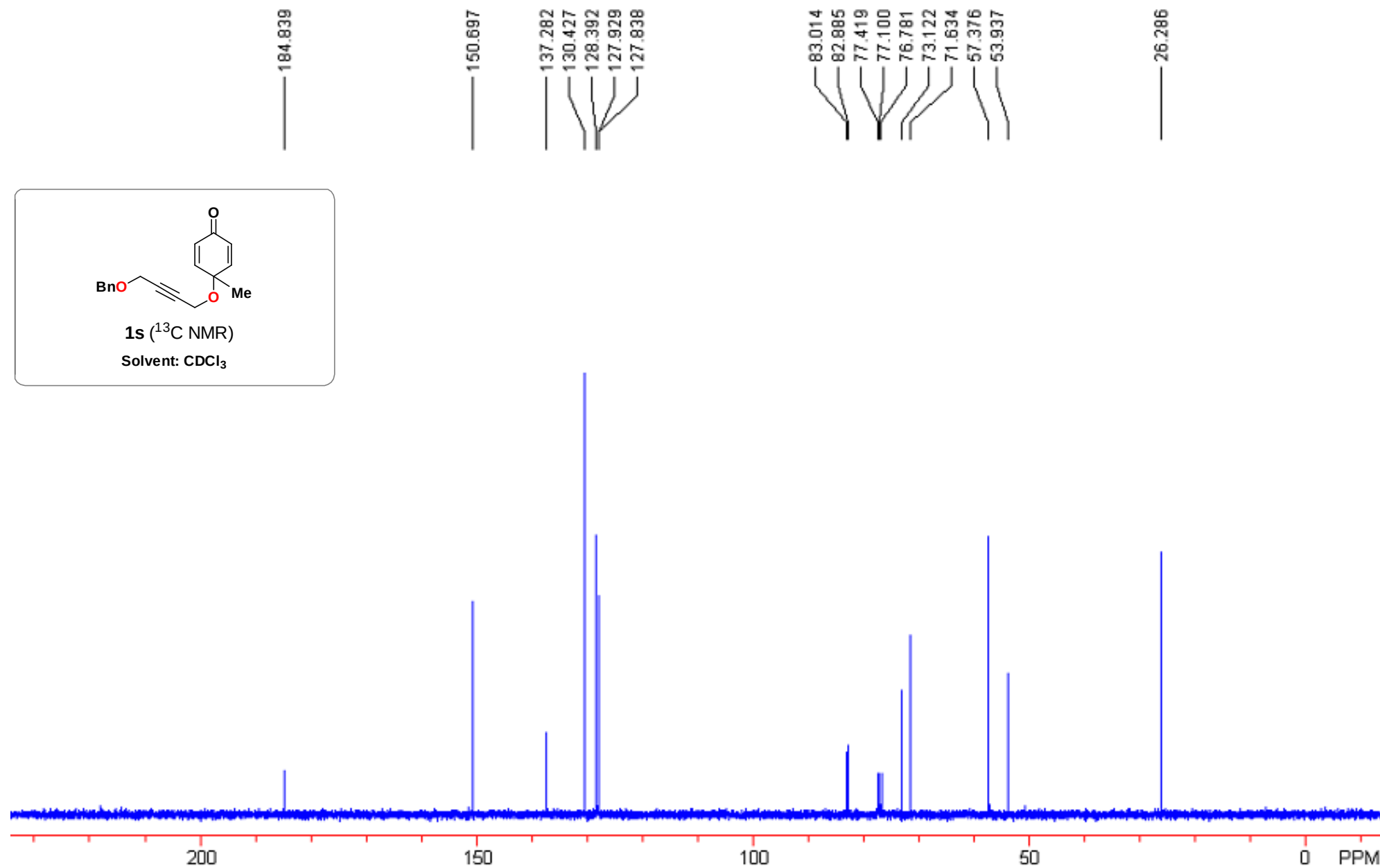
Solvent: CDCl_3

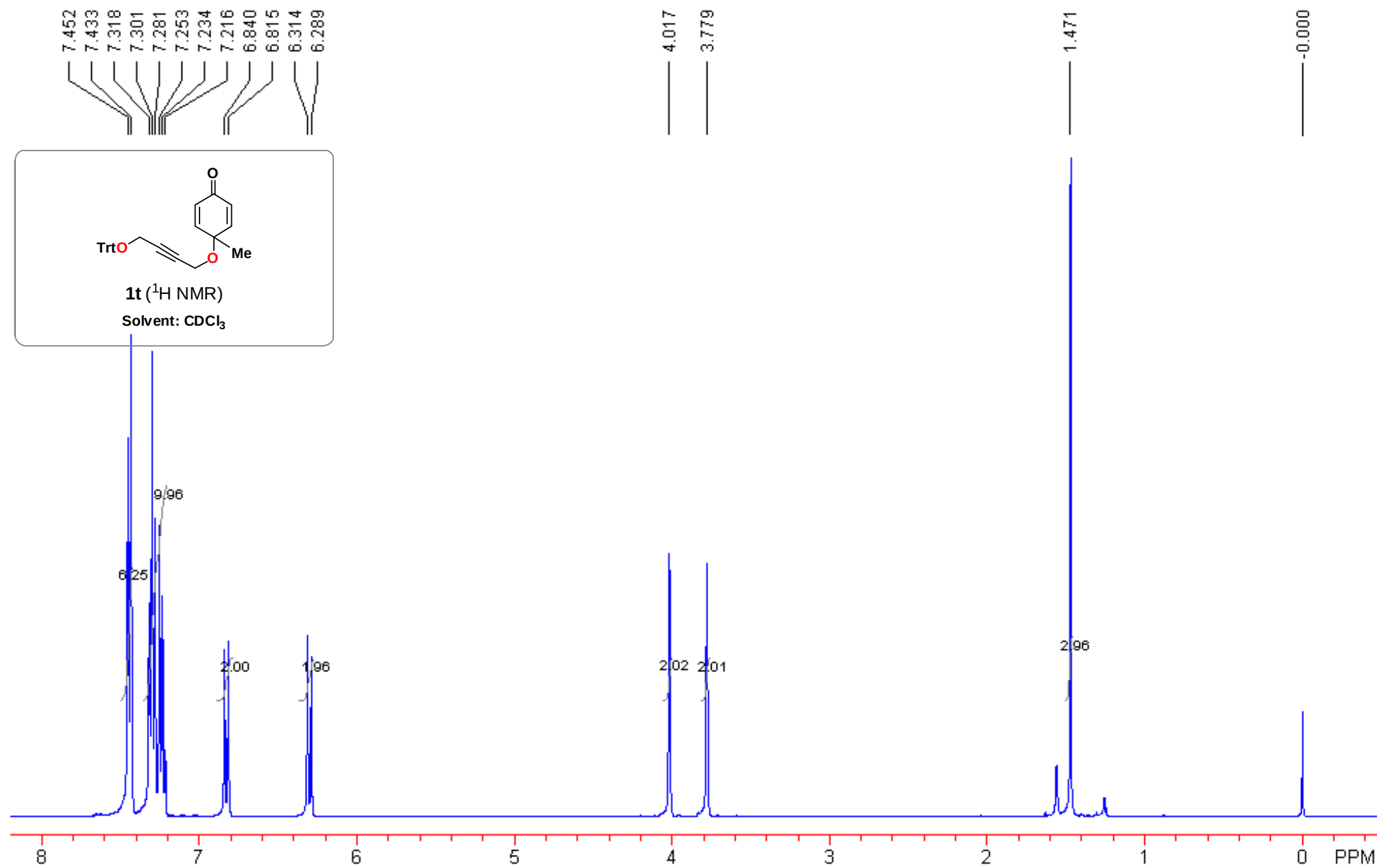


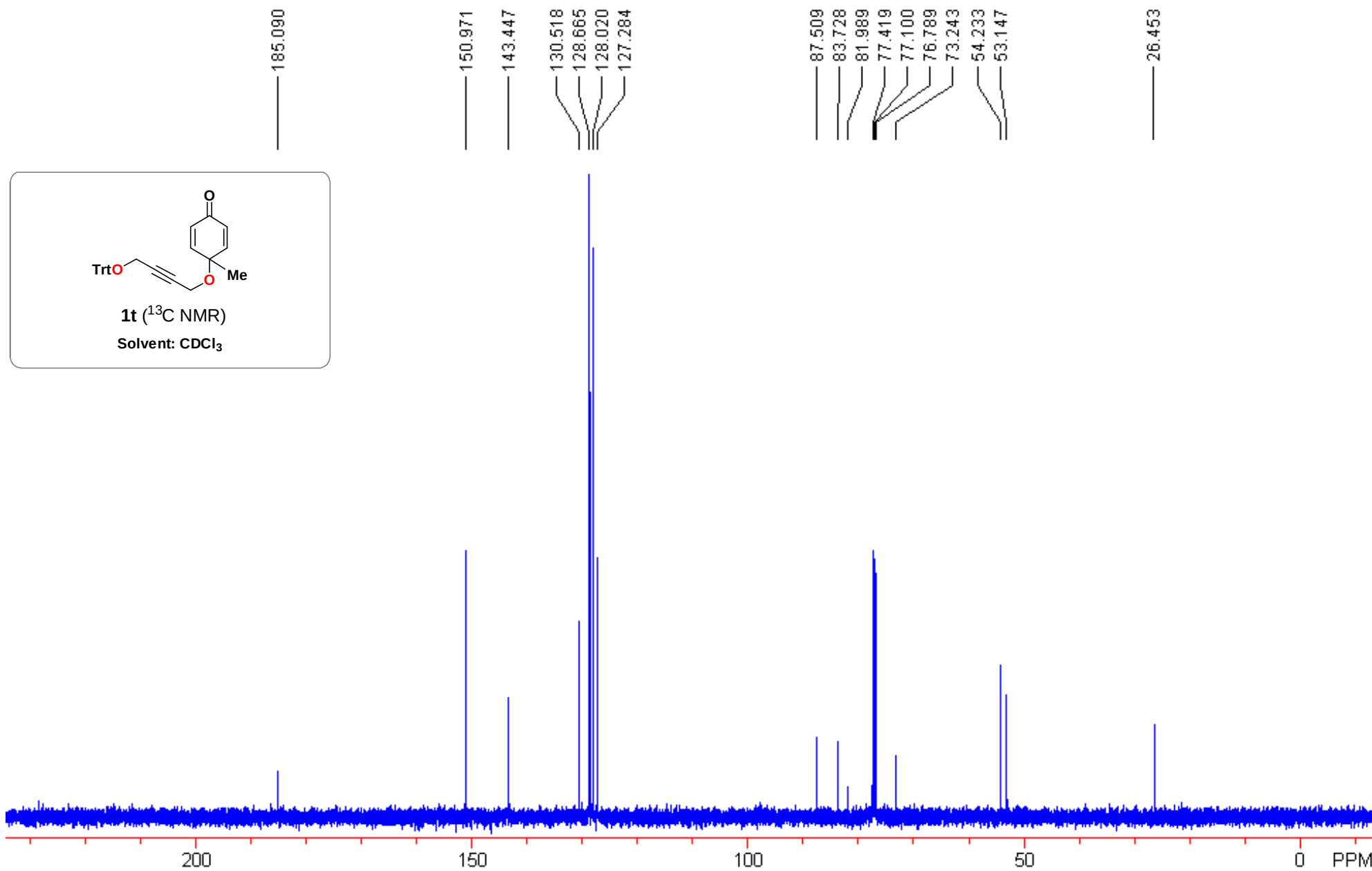


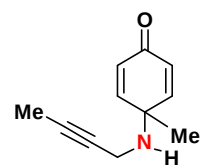
1s (^{13}C NMR)

Solvent: CDCl_3



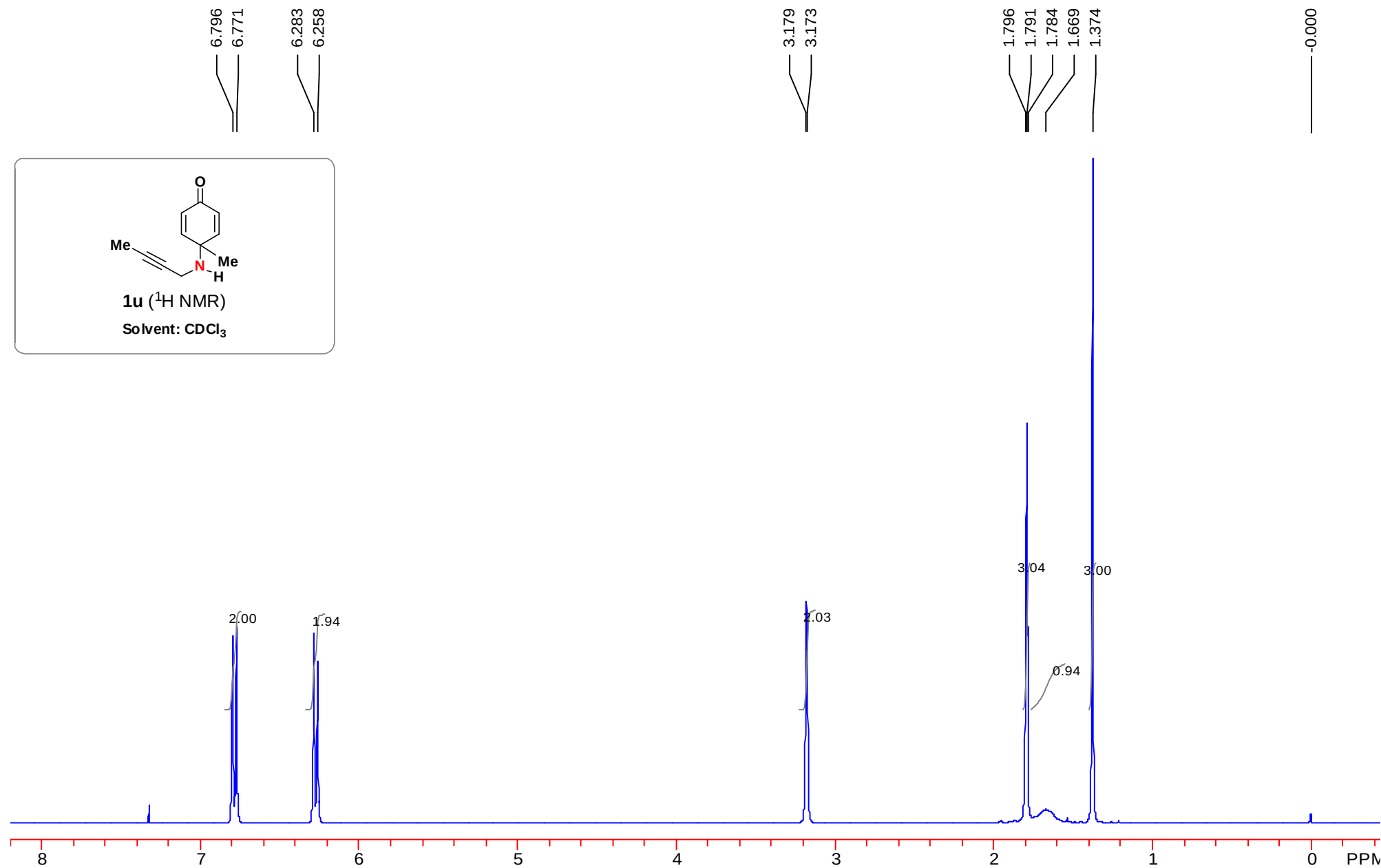


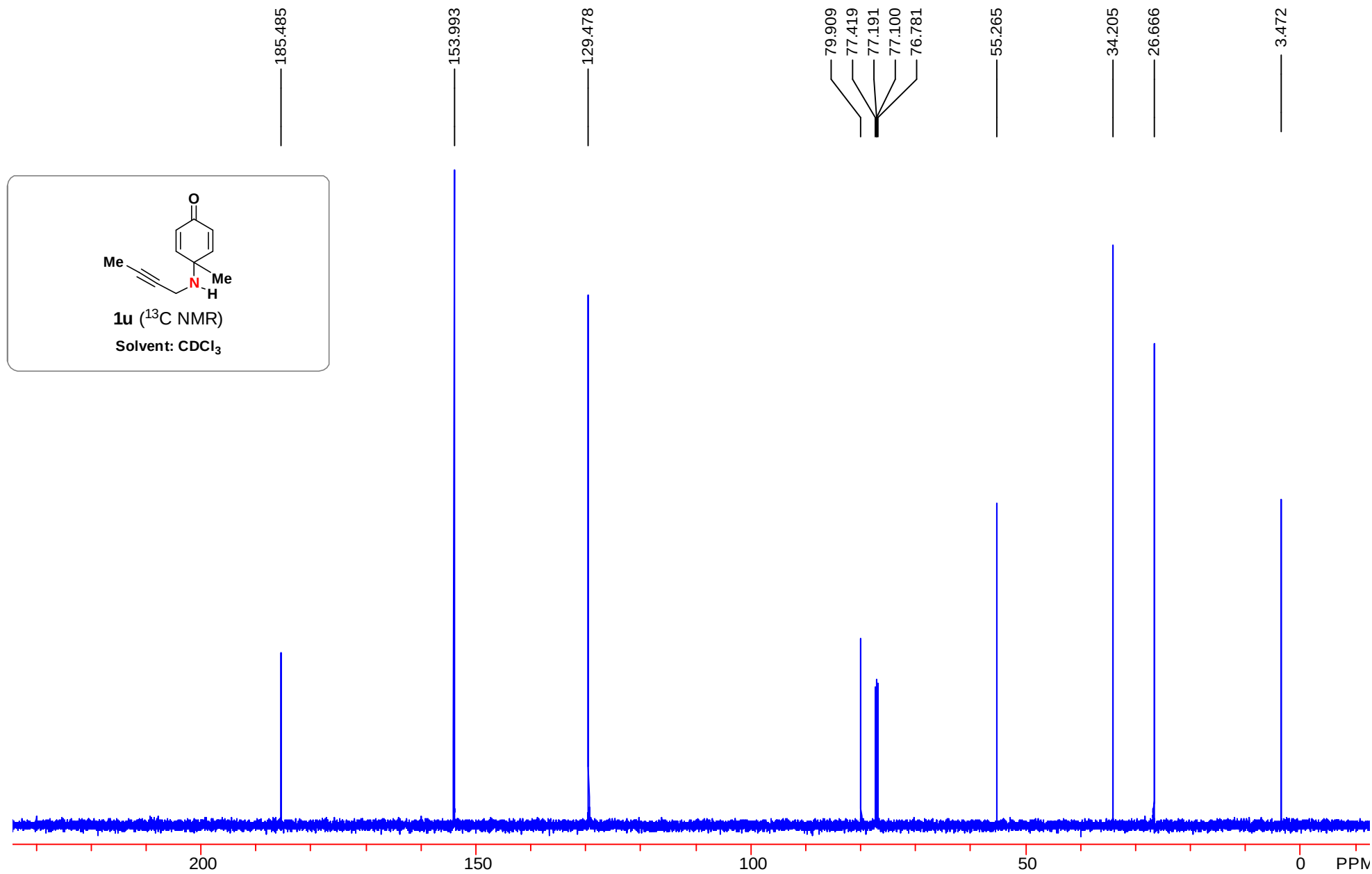


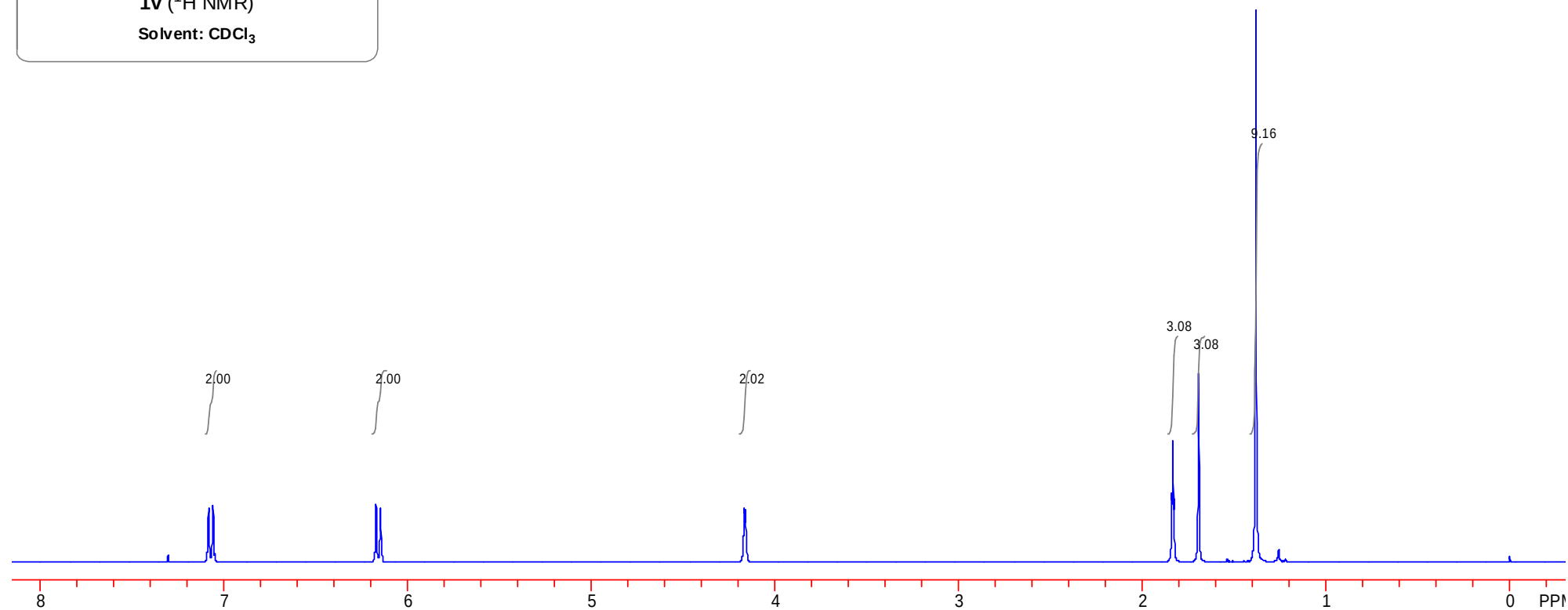
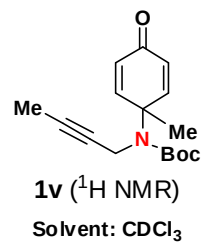
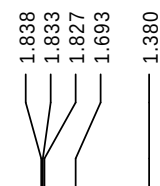
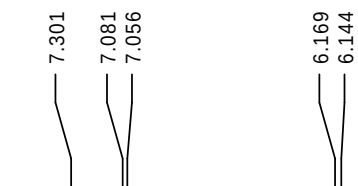


1u (^1H NMR)

Solvent: CDCl_3







185.302

154.956
153.924

126.631

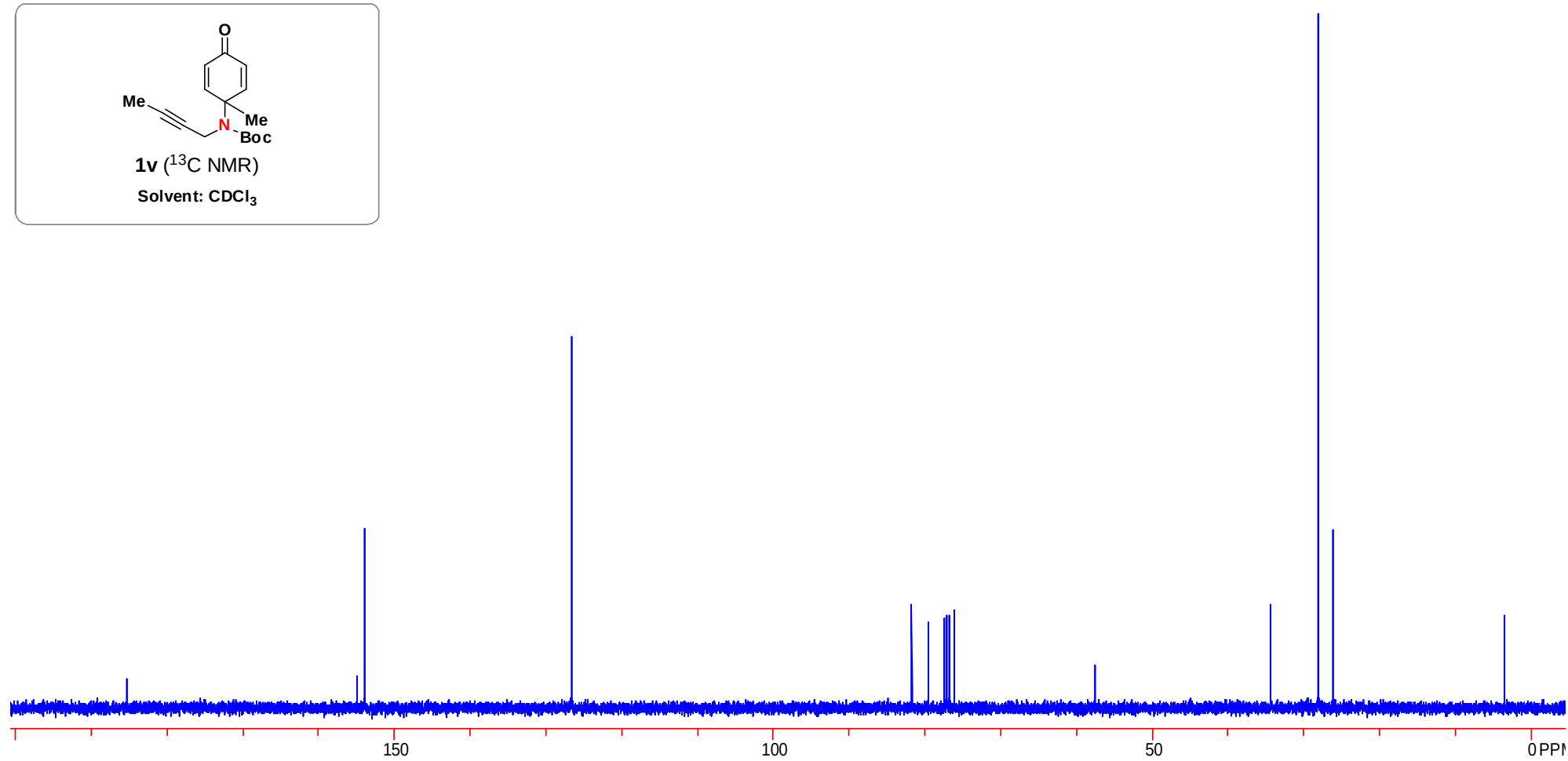
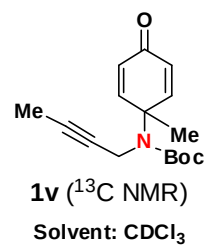
81.708
79.583
77.411
77.100
76.781
76.067

57.482

34.387

28.131
26.188

3.563



7.797
7.776

7.295
7.275

7.042
7.017

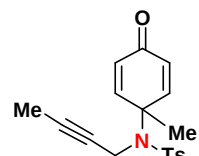
6.147
6.122

4.229
4.223

2.428

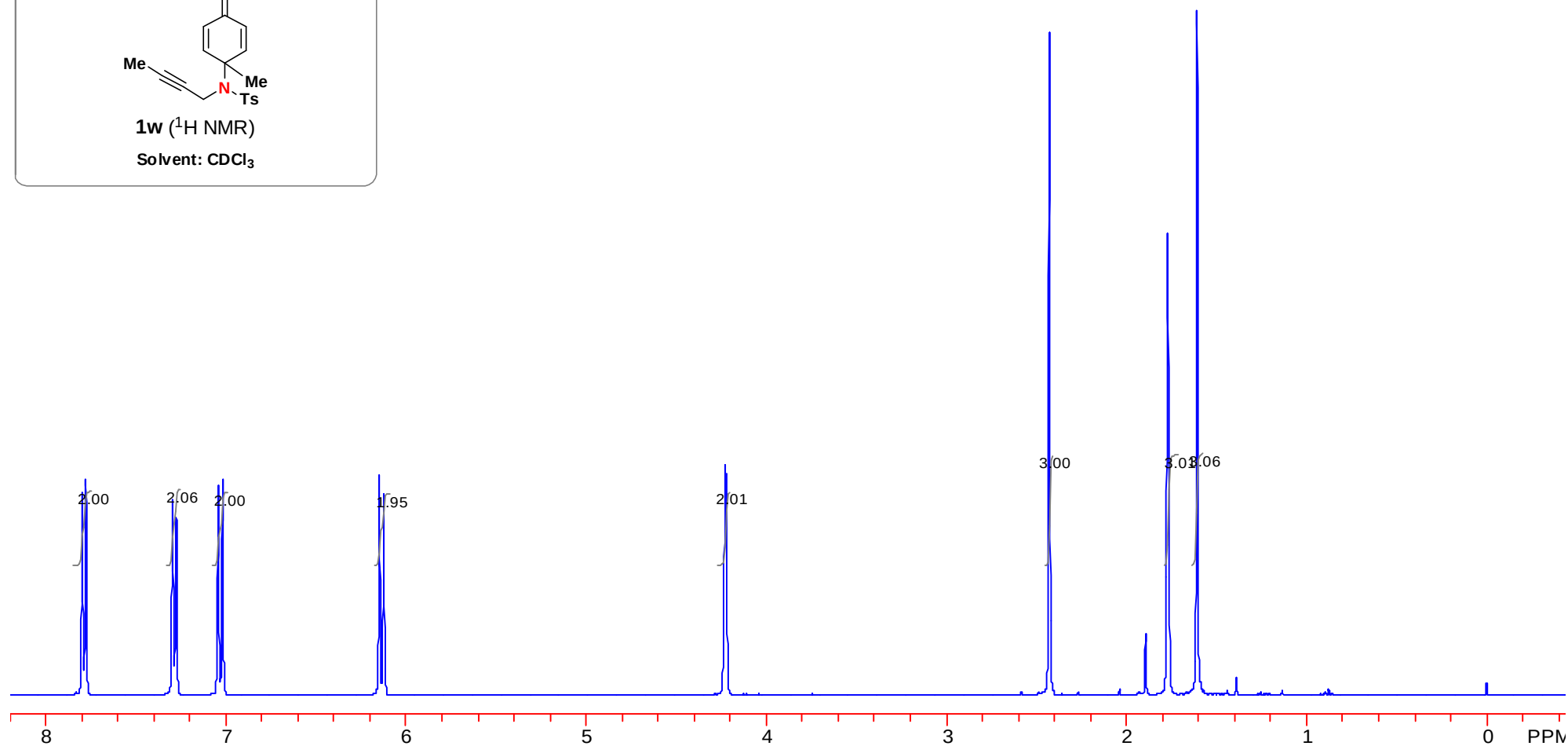
1.769
1.608

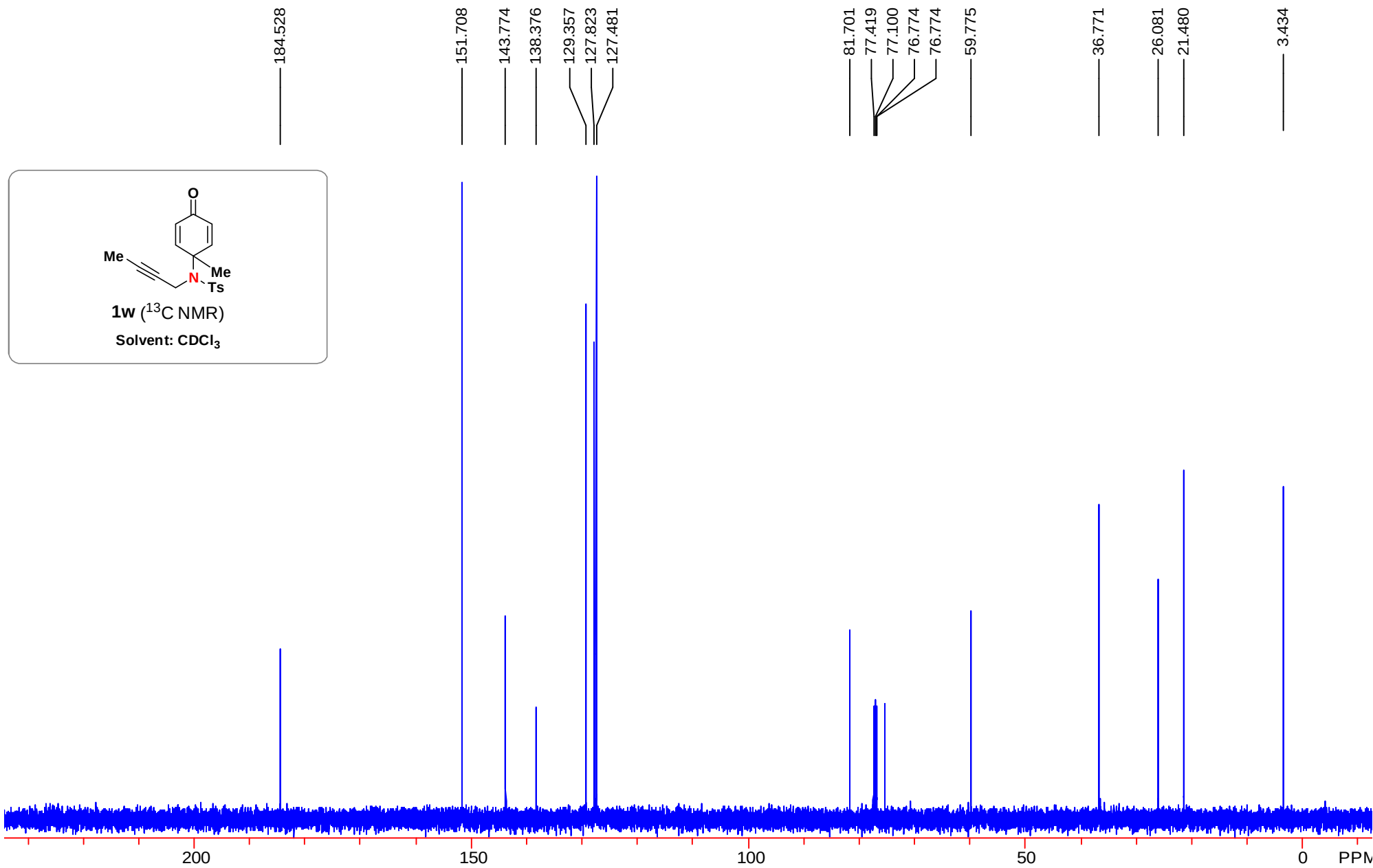
-0.000

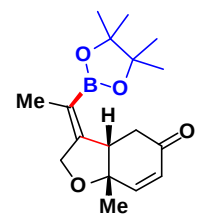
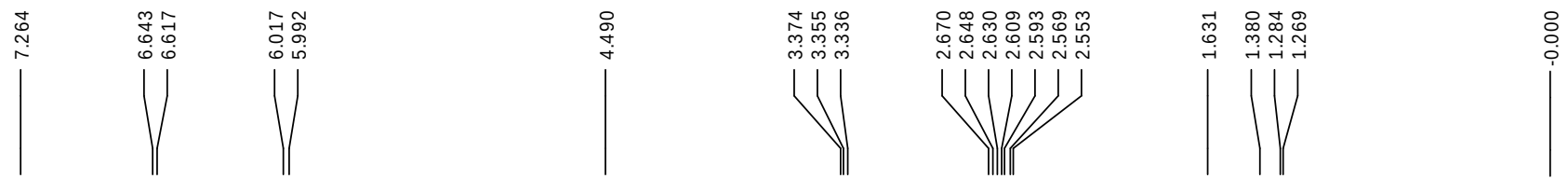


1w (^1H NMR)

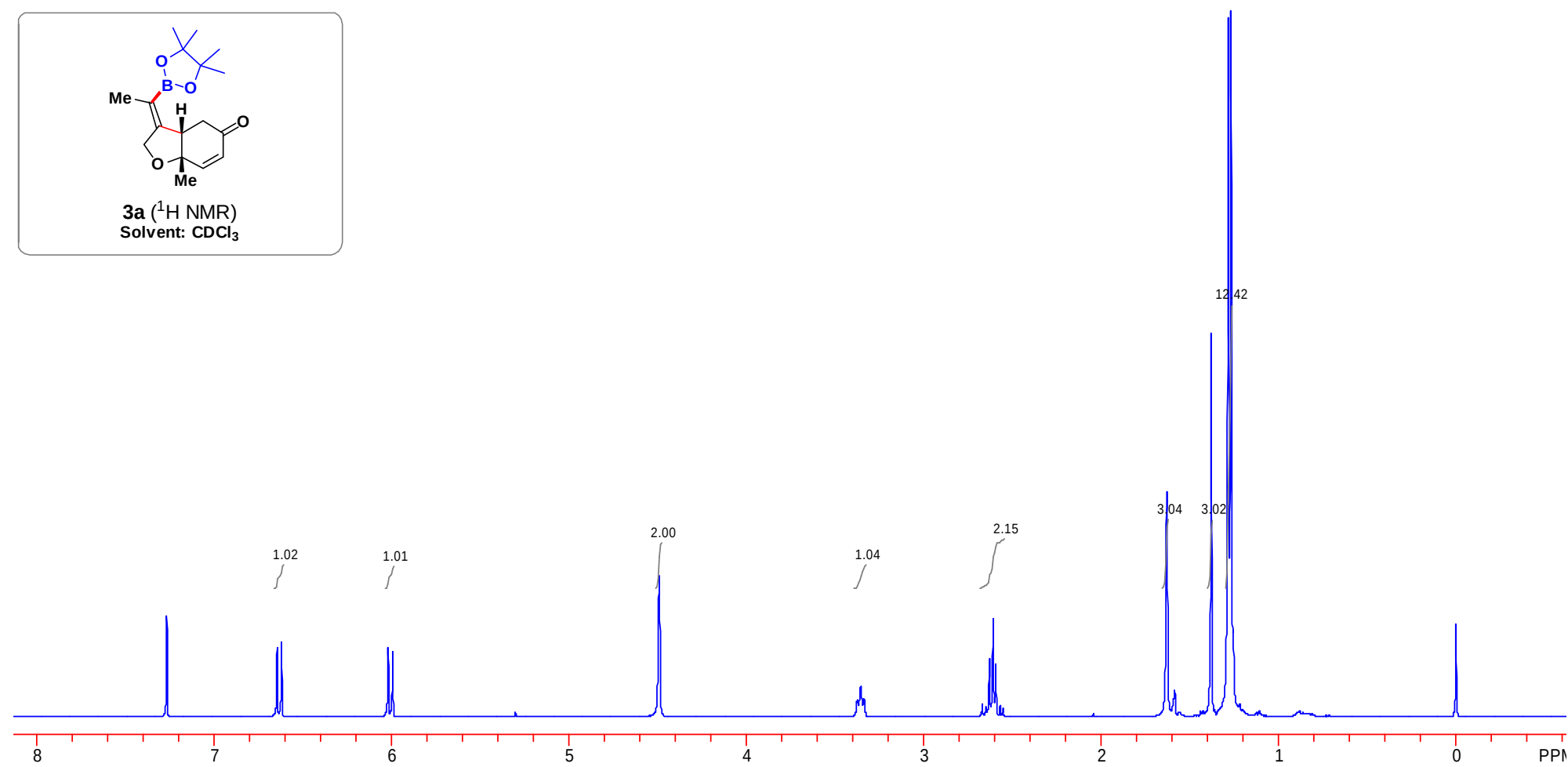
Solvent: CDCl_3

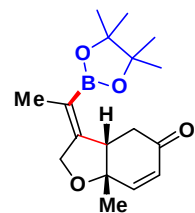






3a (^1H NMR)
Solvent: CDCl_3





3a (^{13}C NMR)
Solvent: CDCl_3

199.044

157.227

149.012

129.721

83.470

79.036

77.419

77.100

76.781

69.834

47.567

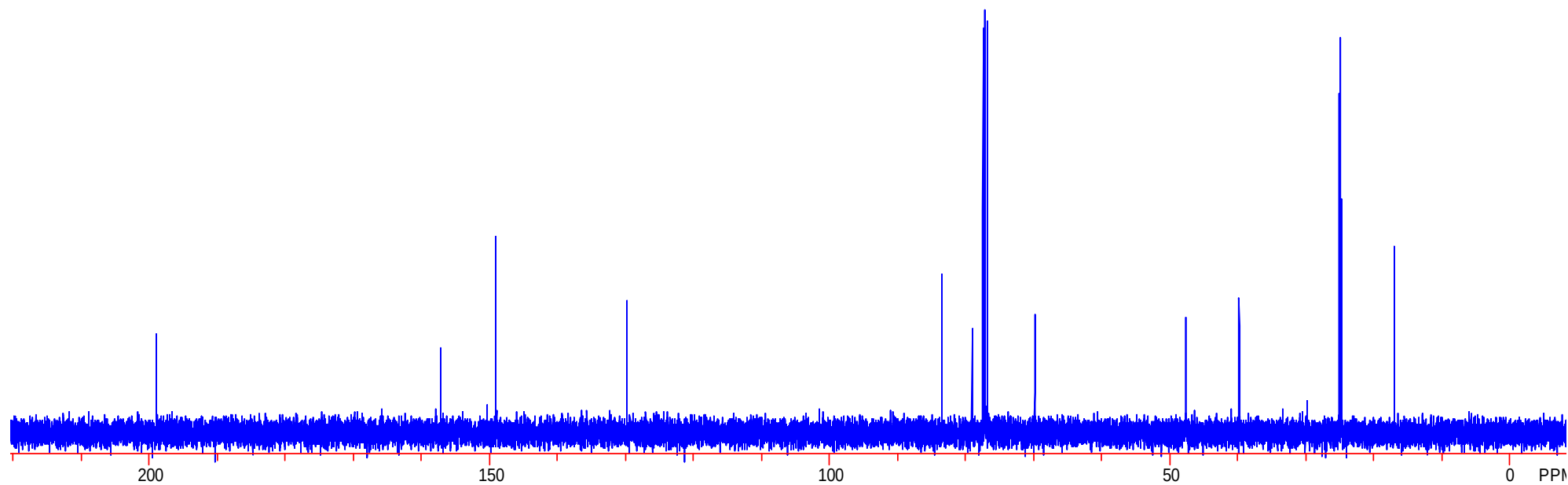
39.747

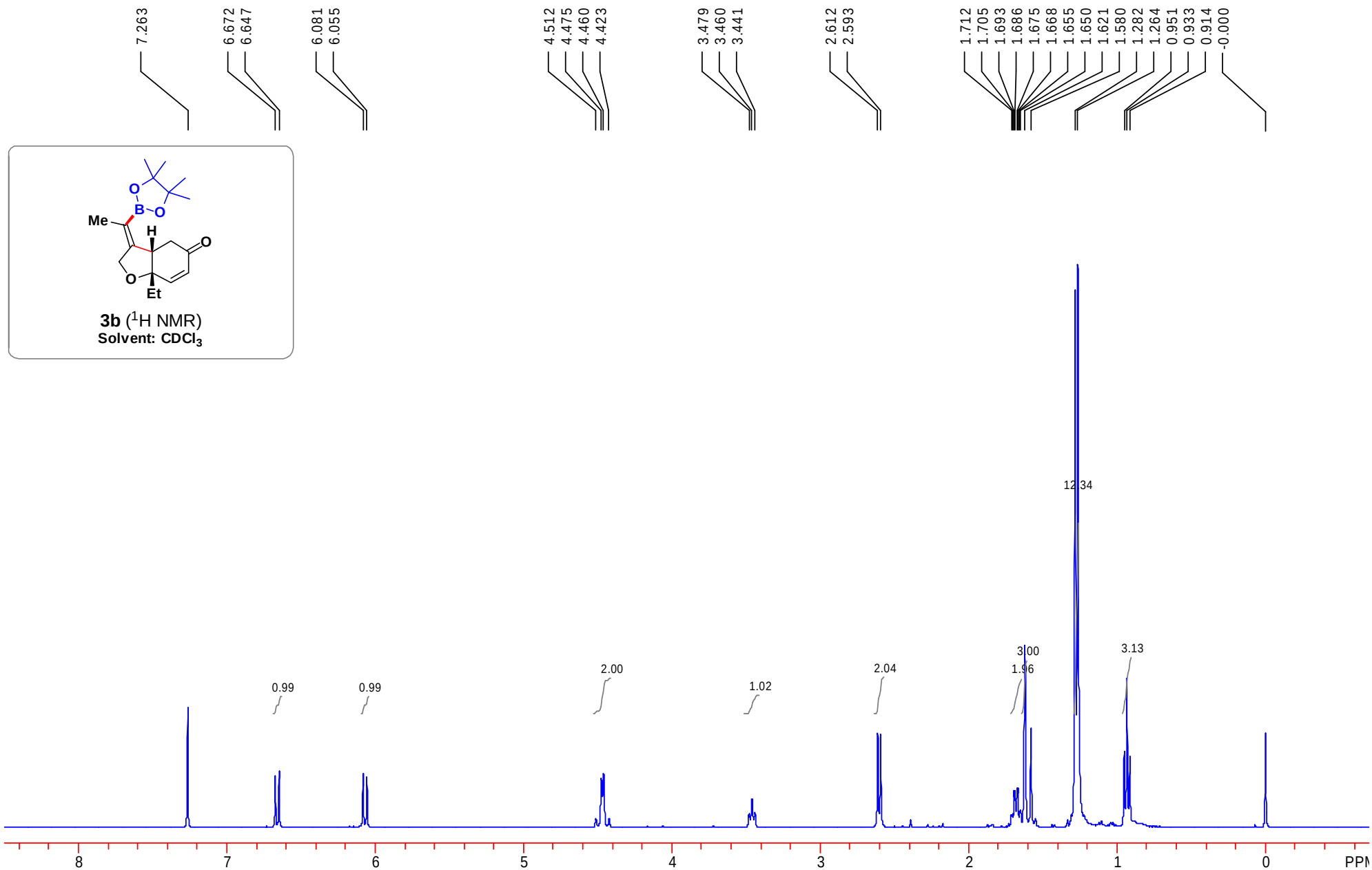
25.034

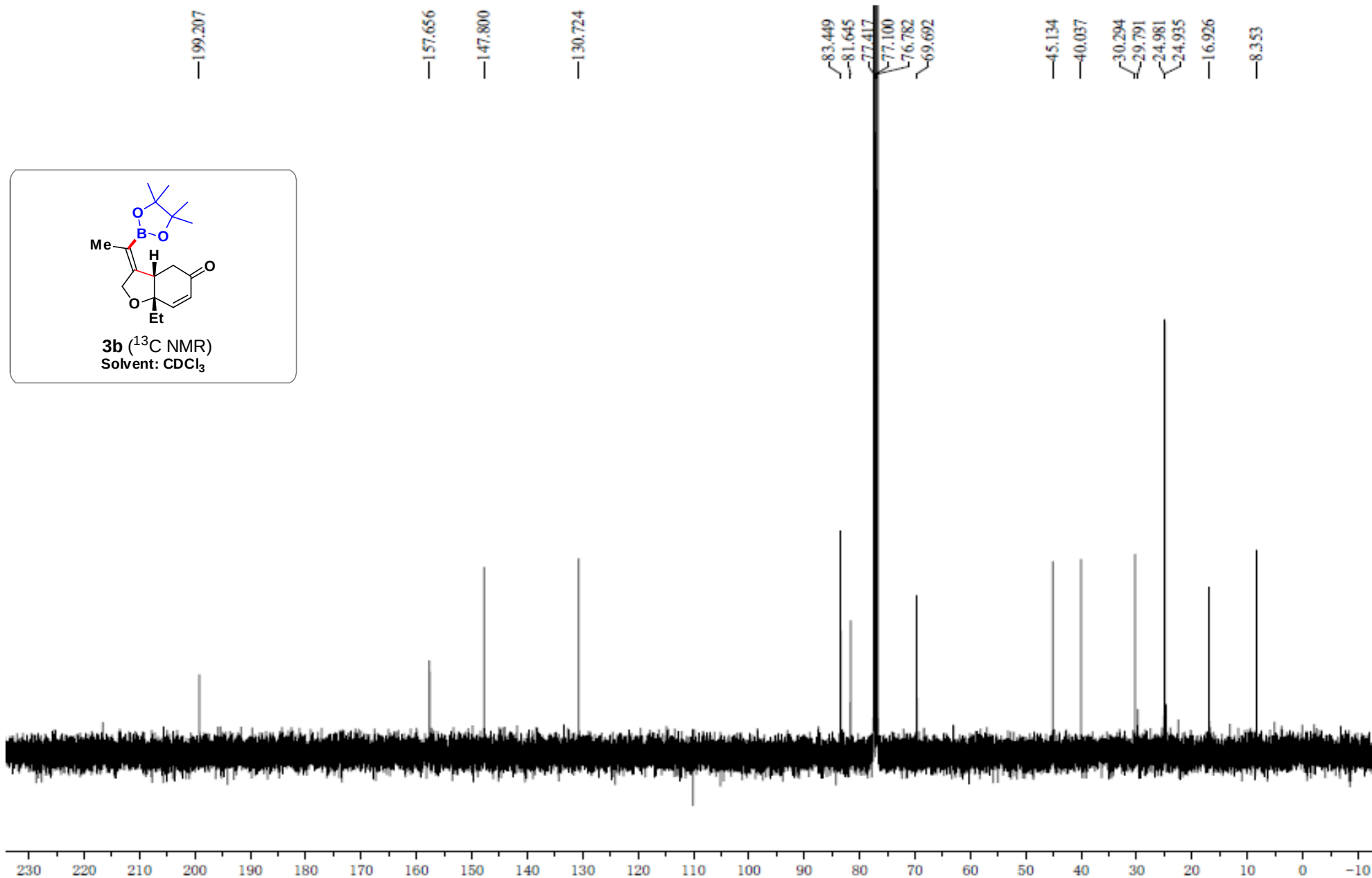
24.867

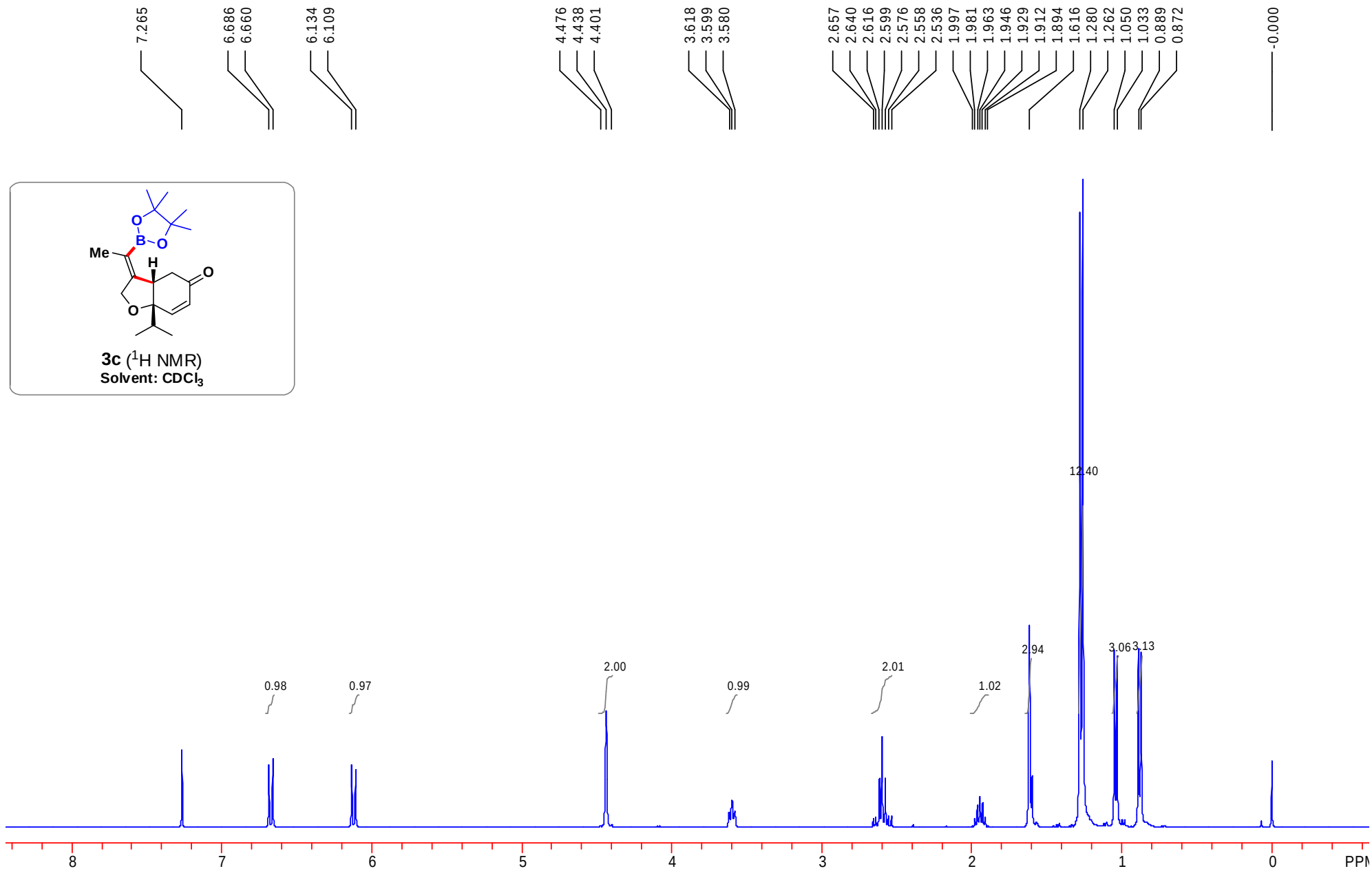
24.700

16.948









199.340

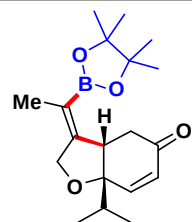
158.381

145.930

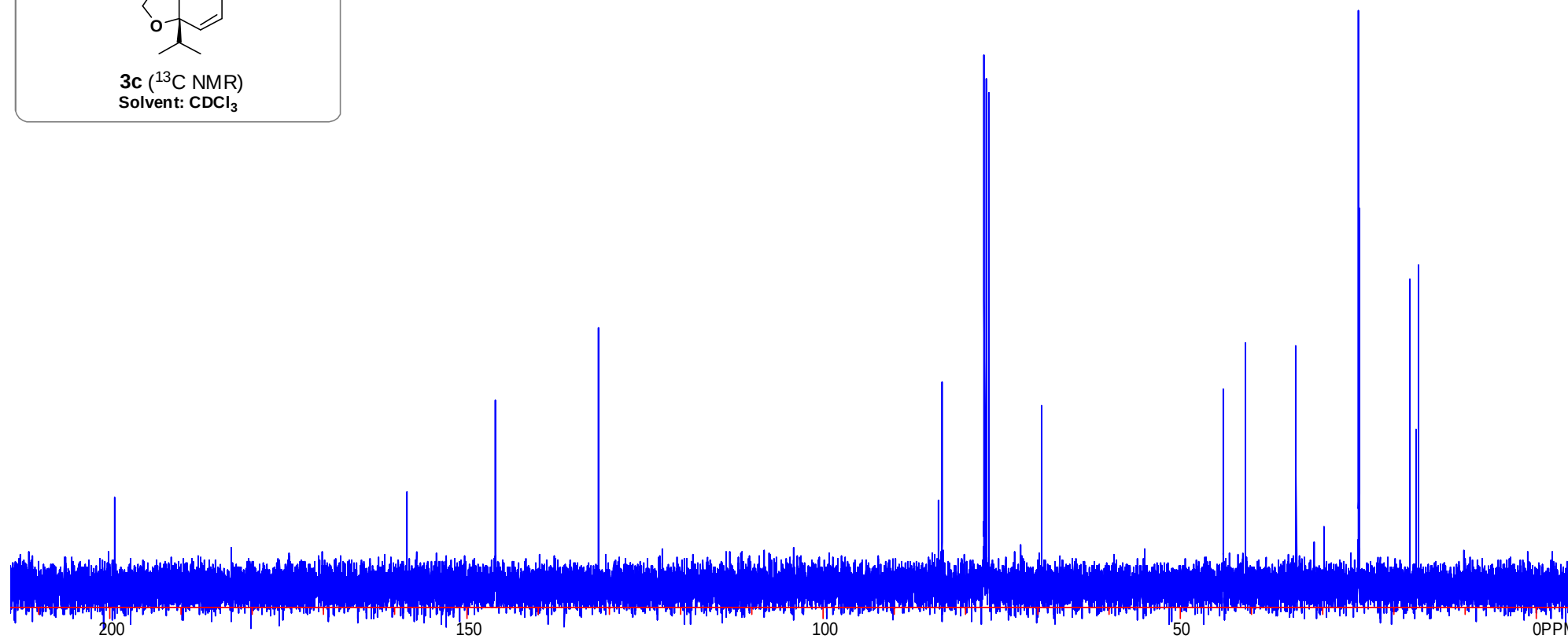
131.482

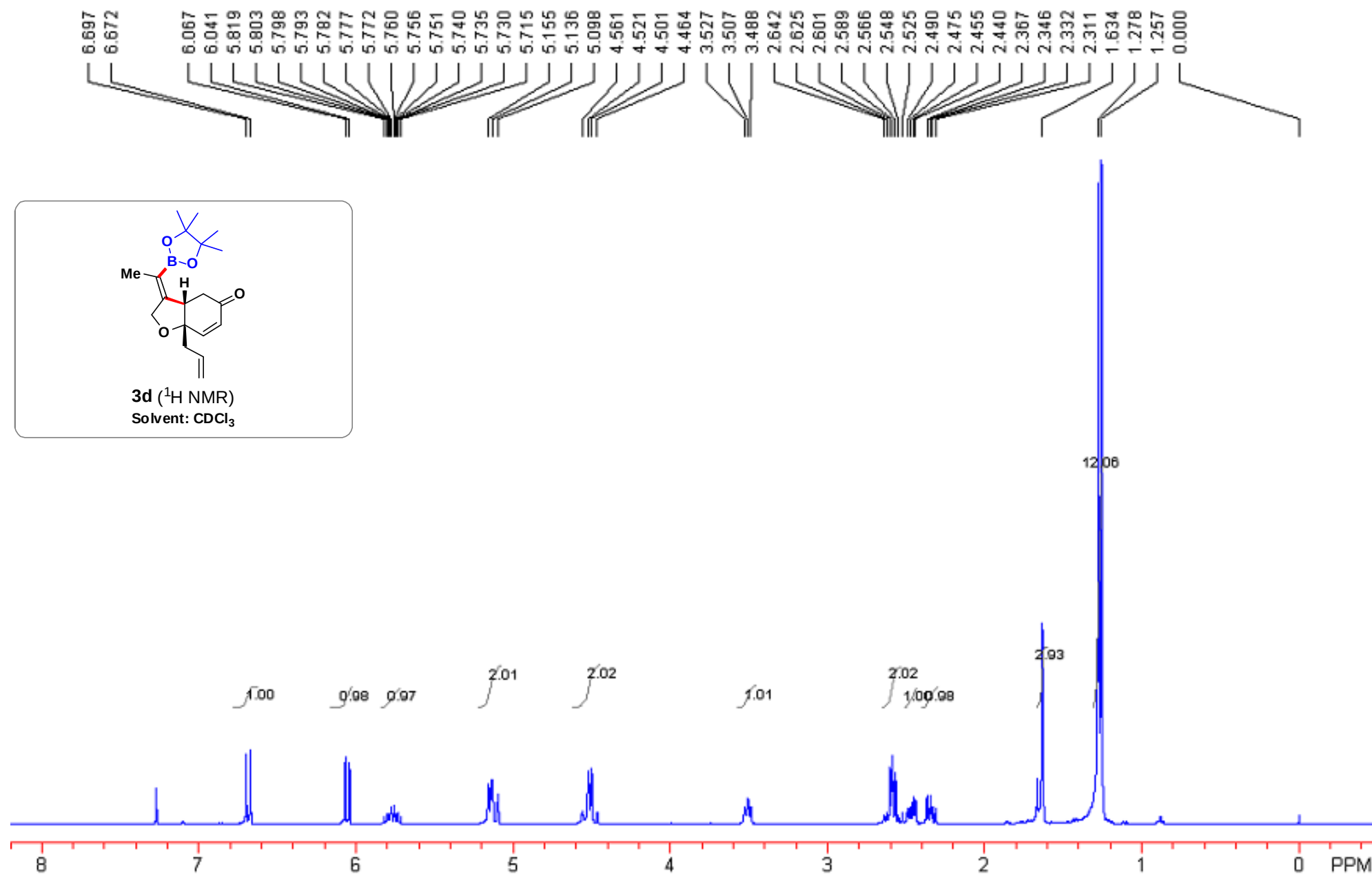
83.819
83.417
77.419
77.100
76.789
69.349

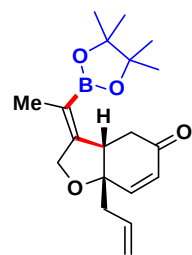
43.908
40.894
33.719
29.779
24.965
24.935
17.806
16.918
16.553



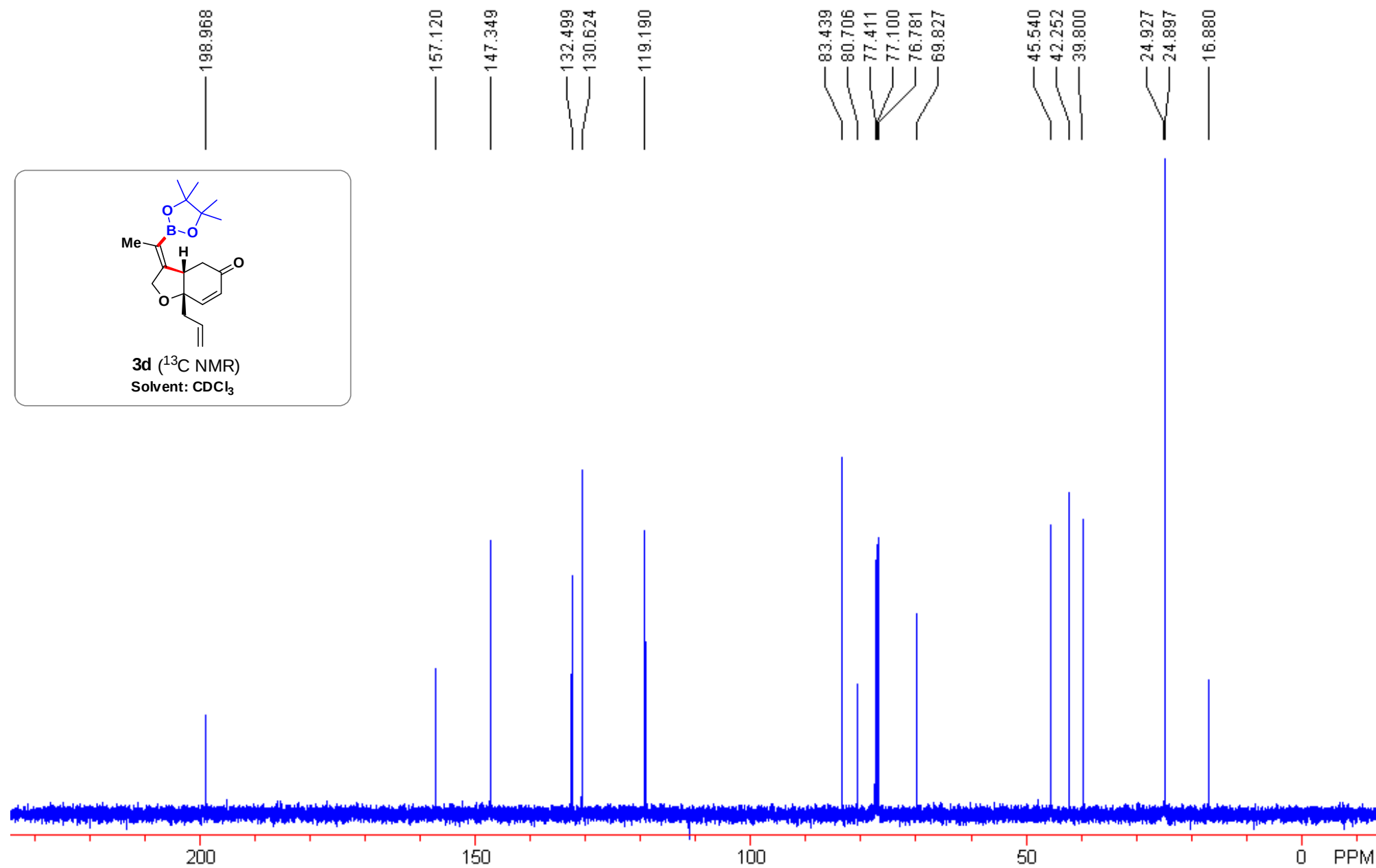
3c (^{13}C NMR)
Solvent: CDCl_3







3d (^{13}C NMR)
Solvent: CDCl_3



7.288
7.282
7.266
7.261
7.248
7.242
7.242
7.242
7.225
7.190
7.186
7.170
6.664
6.639
5.998
5.972

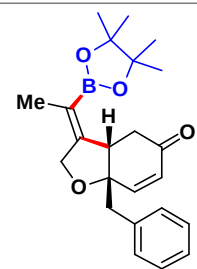
4.609
4.572
4.559
4.521

3.630
3.611
3.591

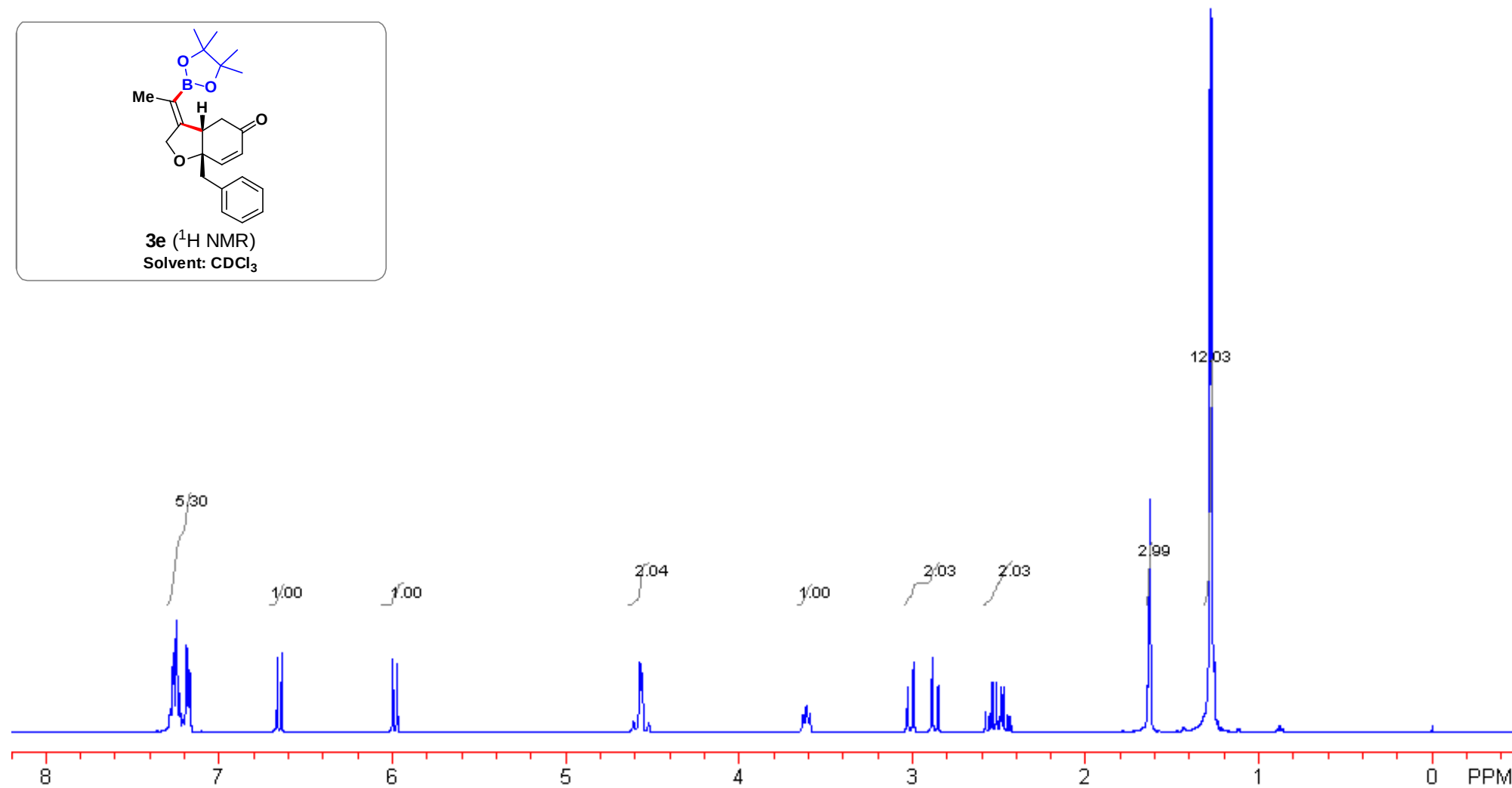
3.027
2.993
2.885
2.850
2.578
2.554
2.537
2.514

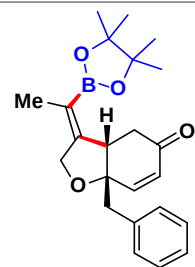
2.489
2.473
2.448
2.432
1.629
1.281
1.273

0.000

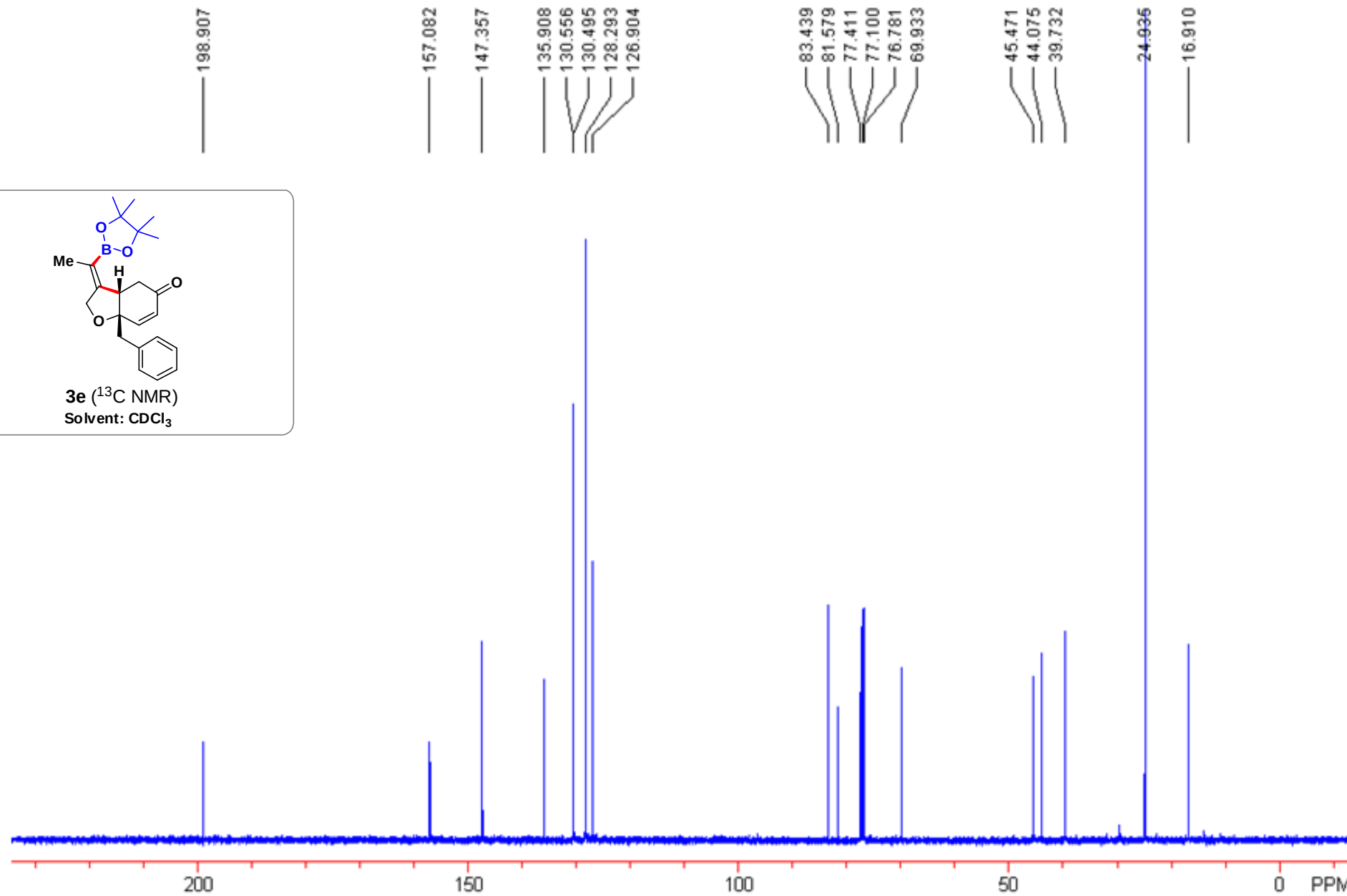


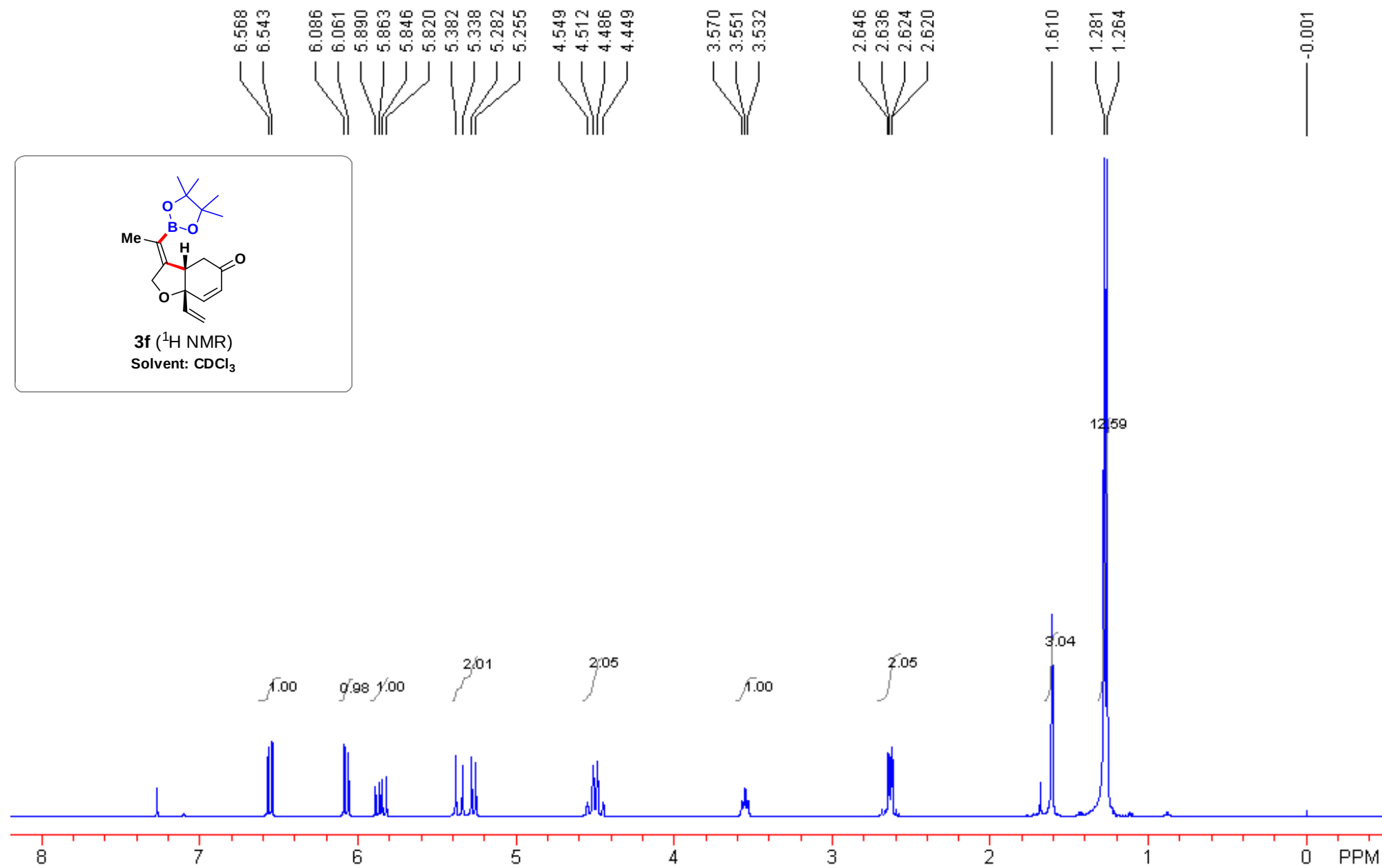
3e (^1H NMR)
Solvent: CDCl_3

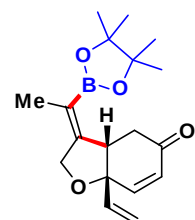




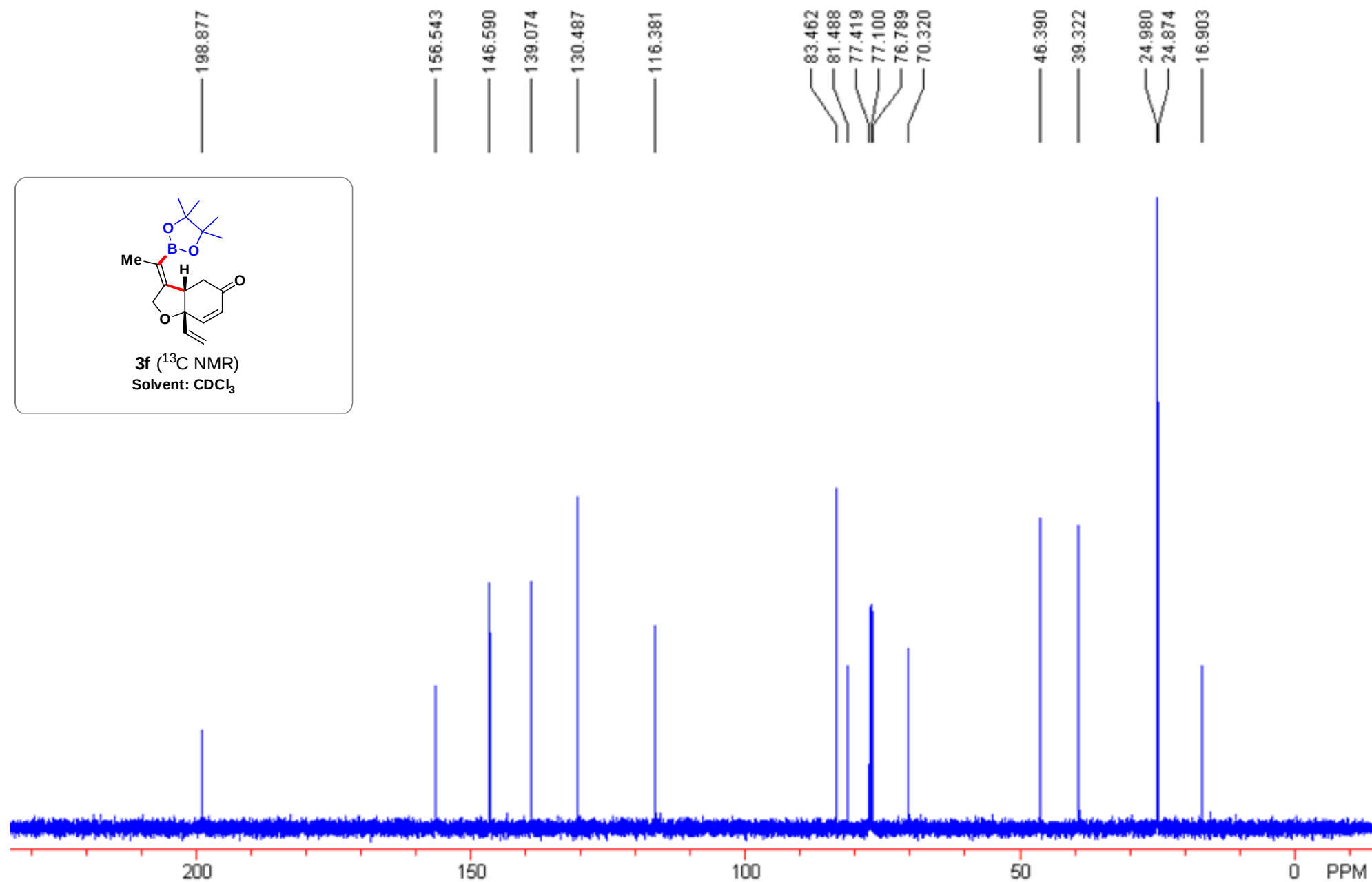
3e (^{13}C NMR)
Solvent: CDCl_3

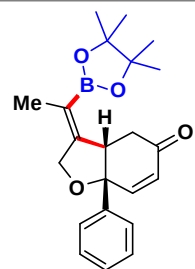
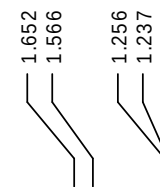
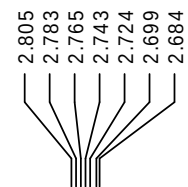
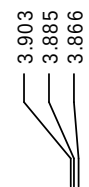
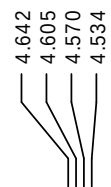
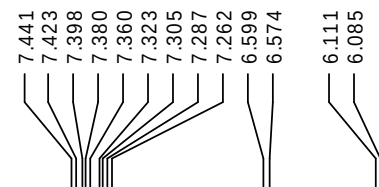






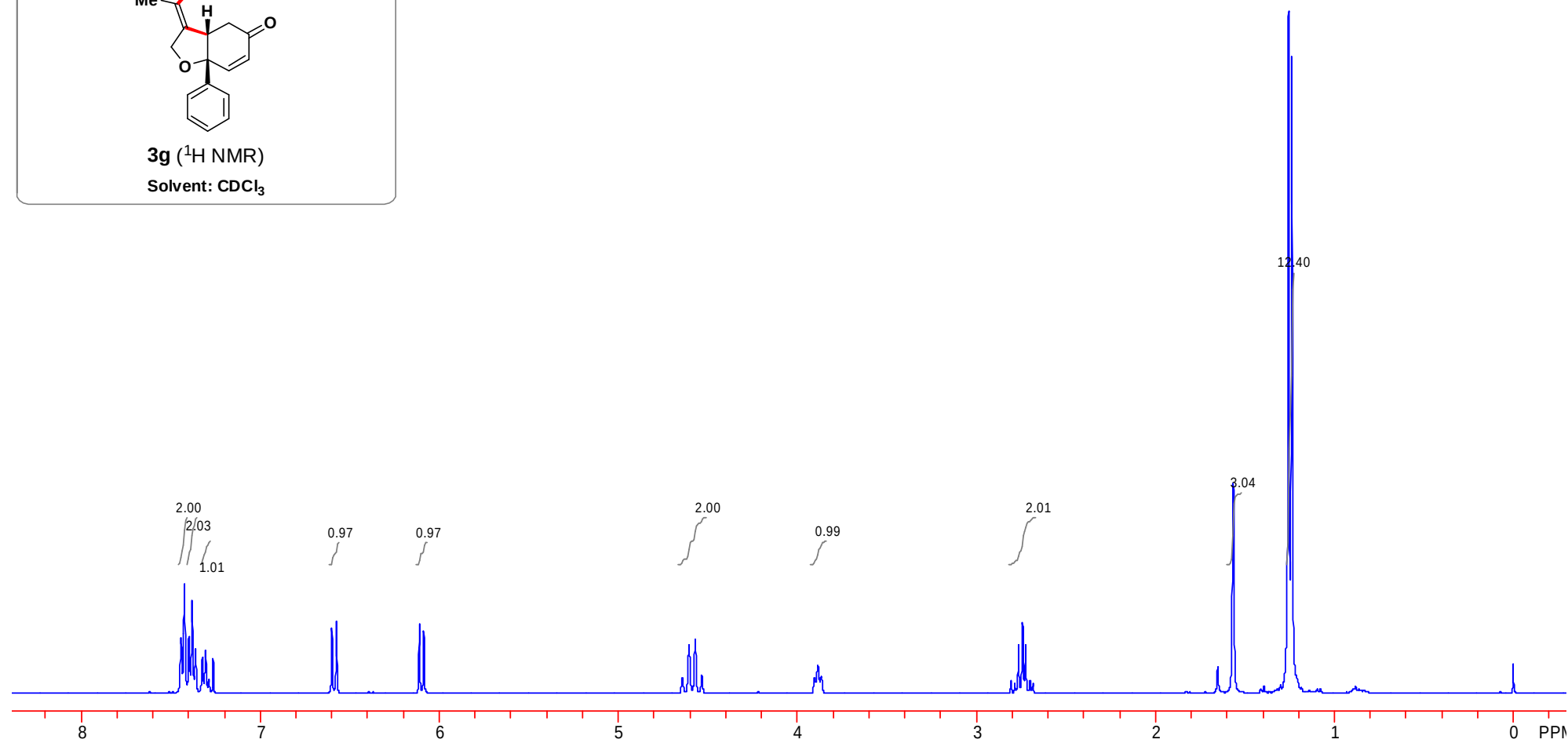
3f (^{13}C NMR)
Solvent: CDCl_3

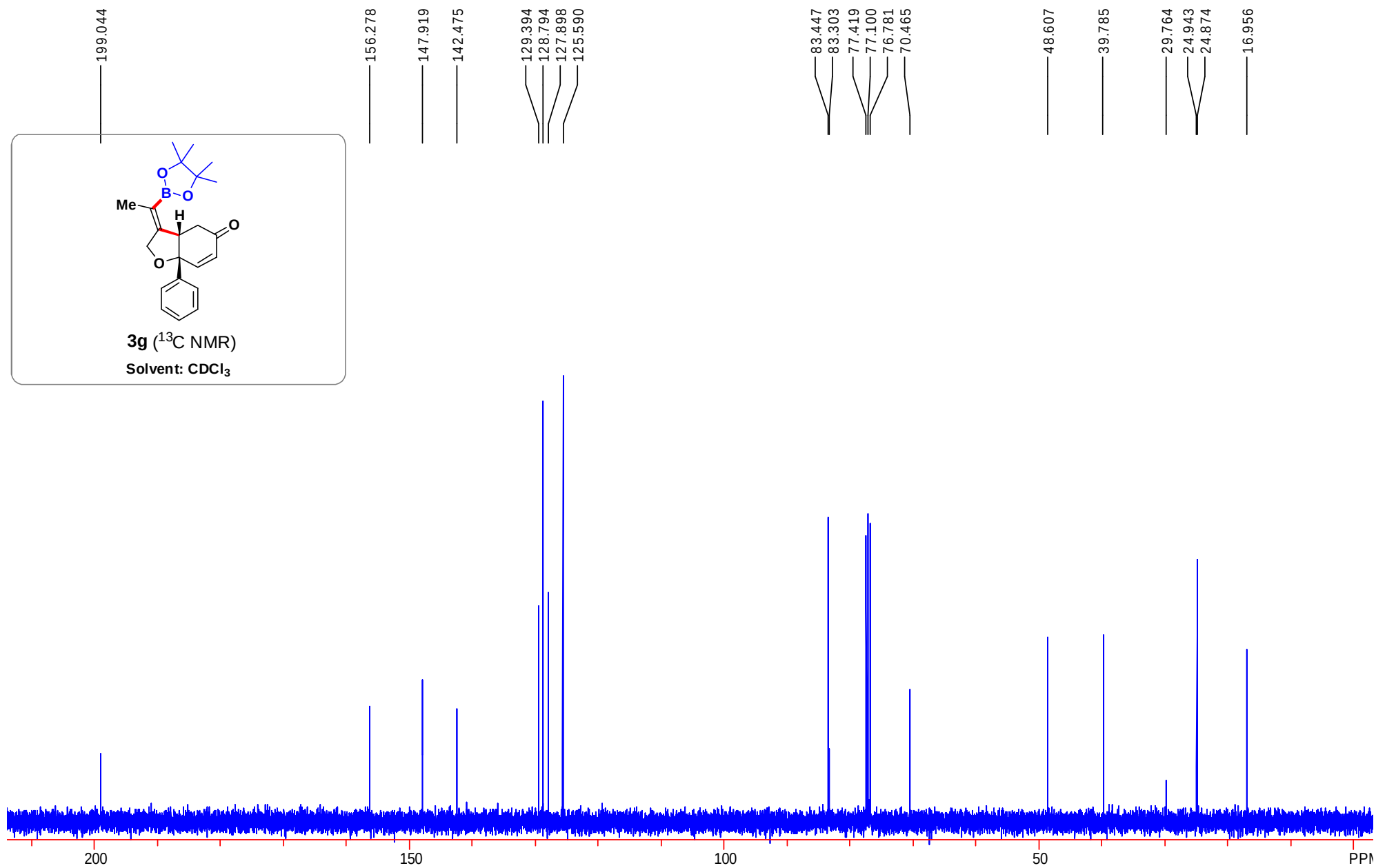


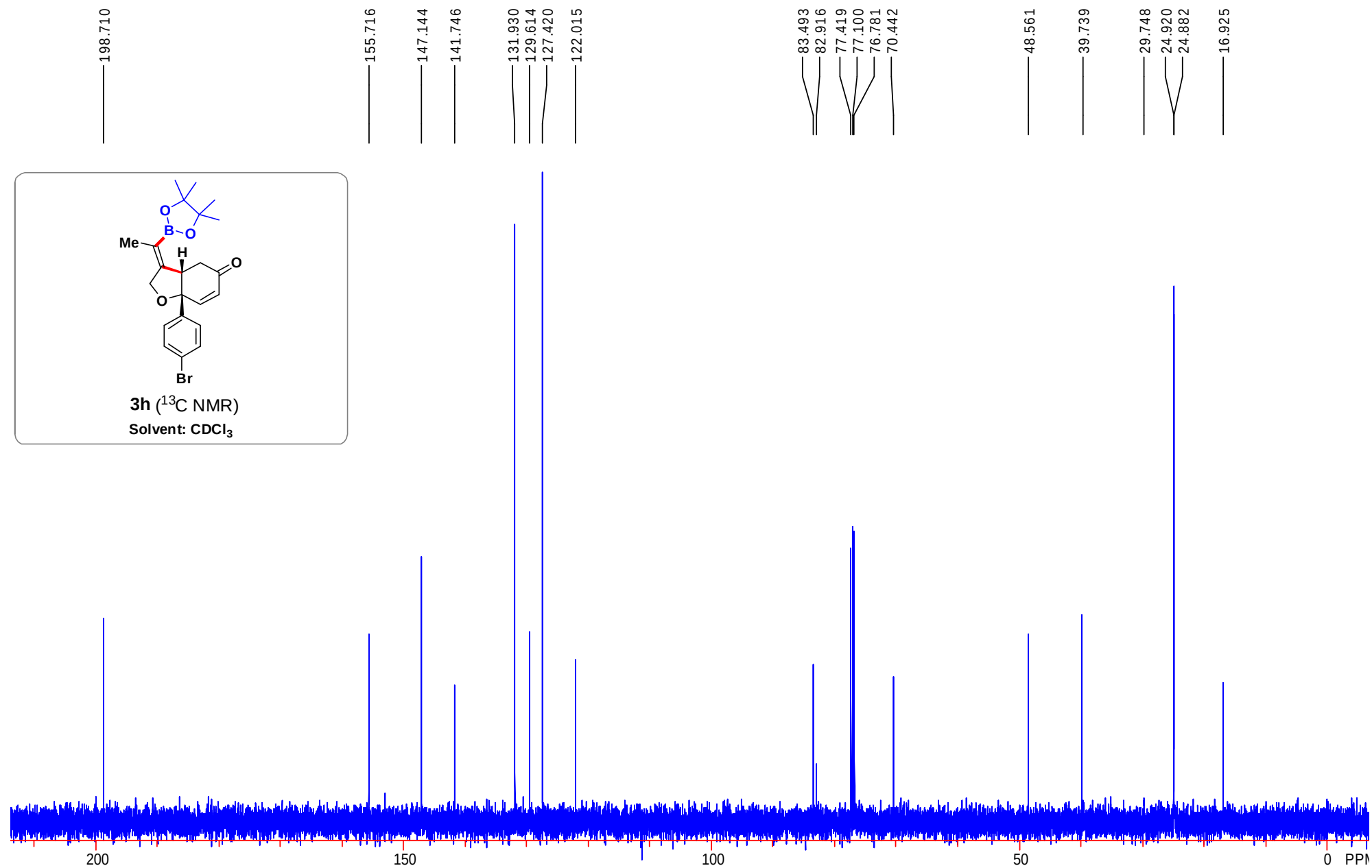
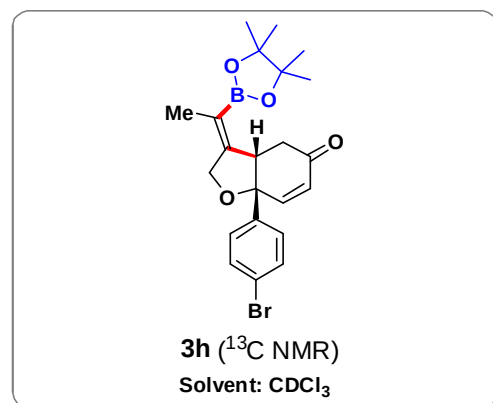


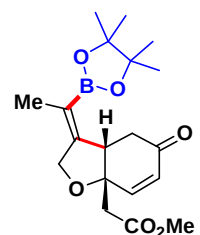
3g (¹H NMR)

Solvent: CDCl₃

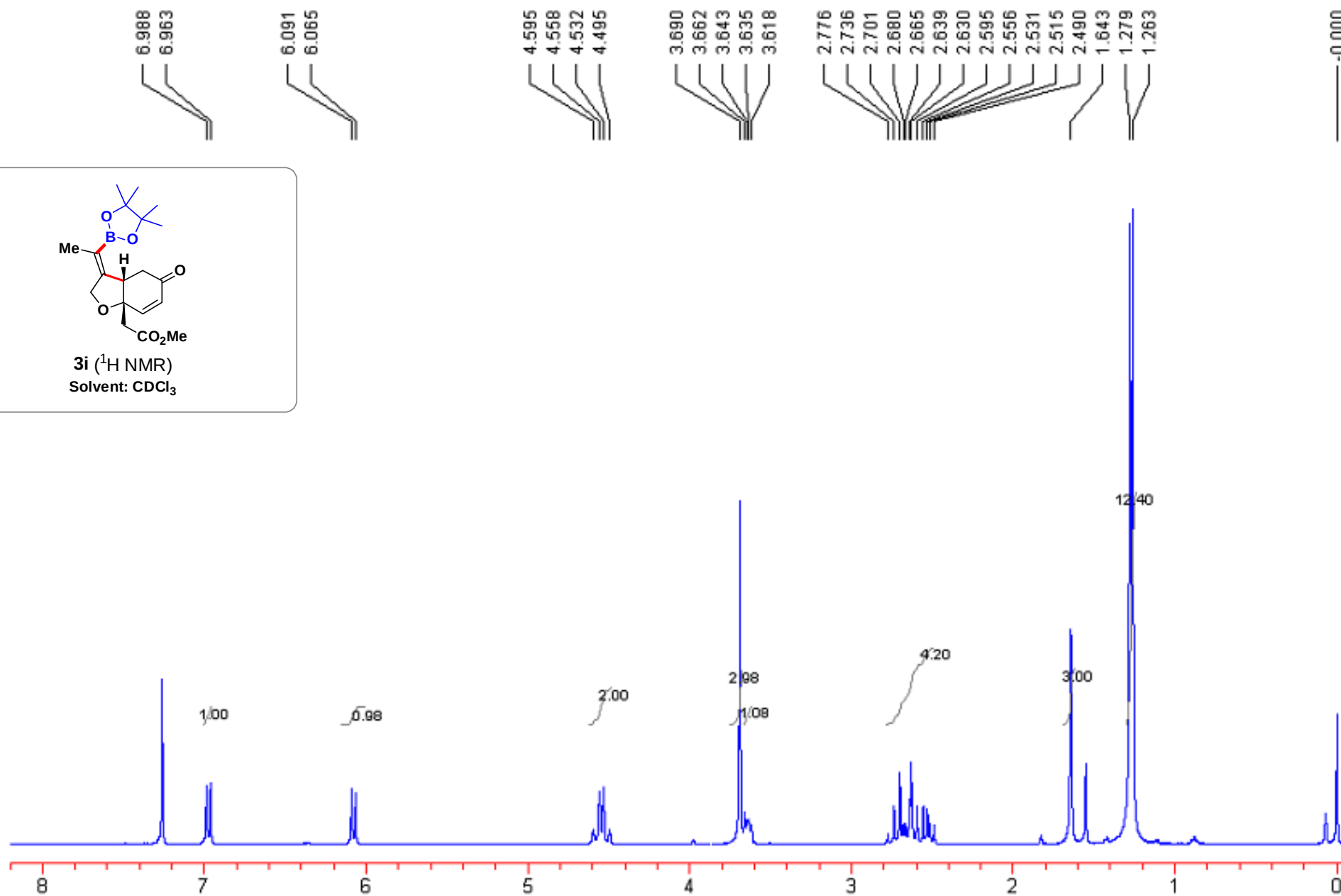


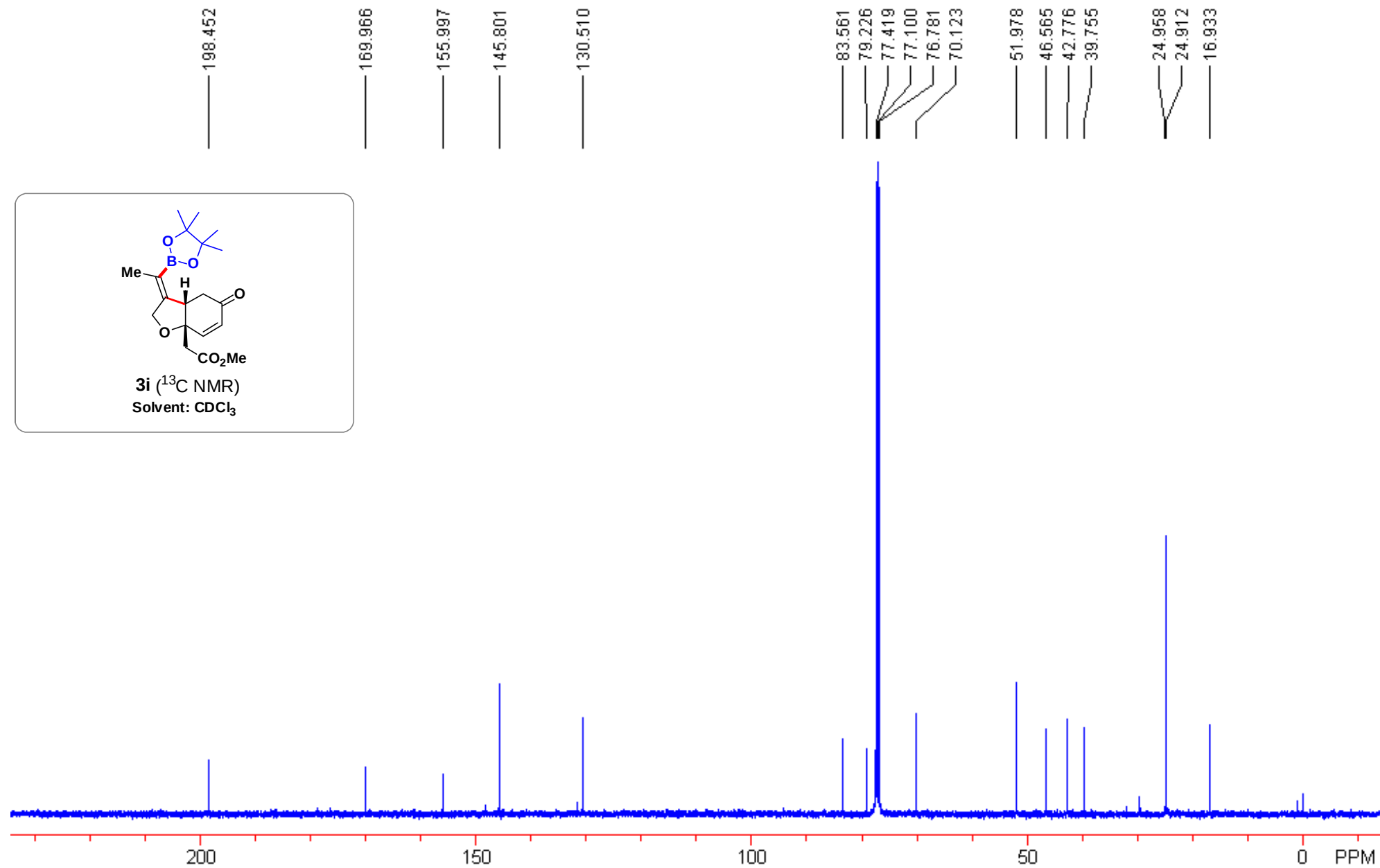


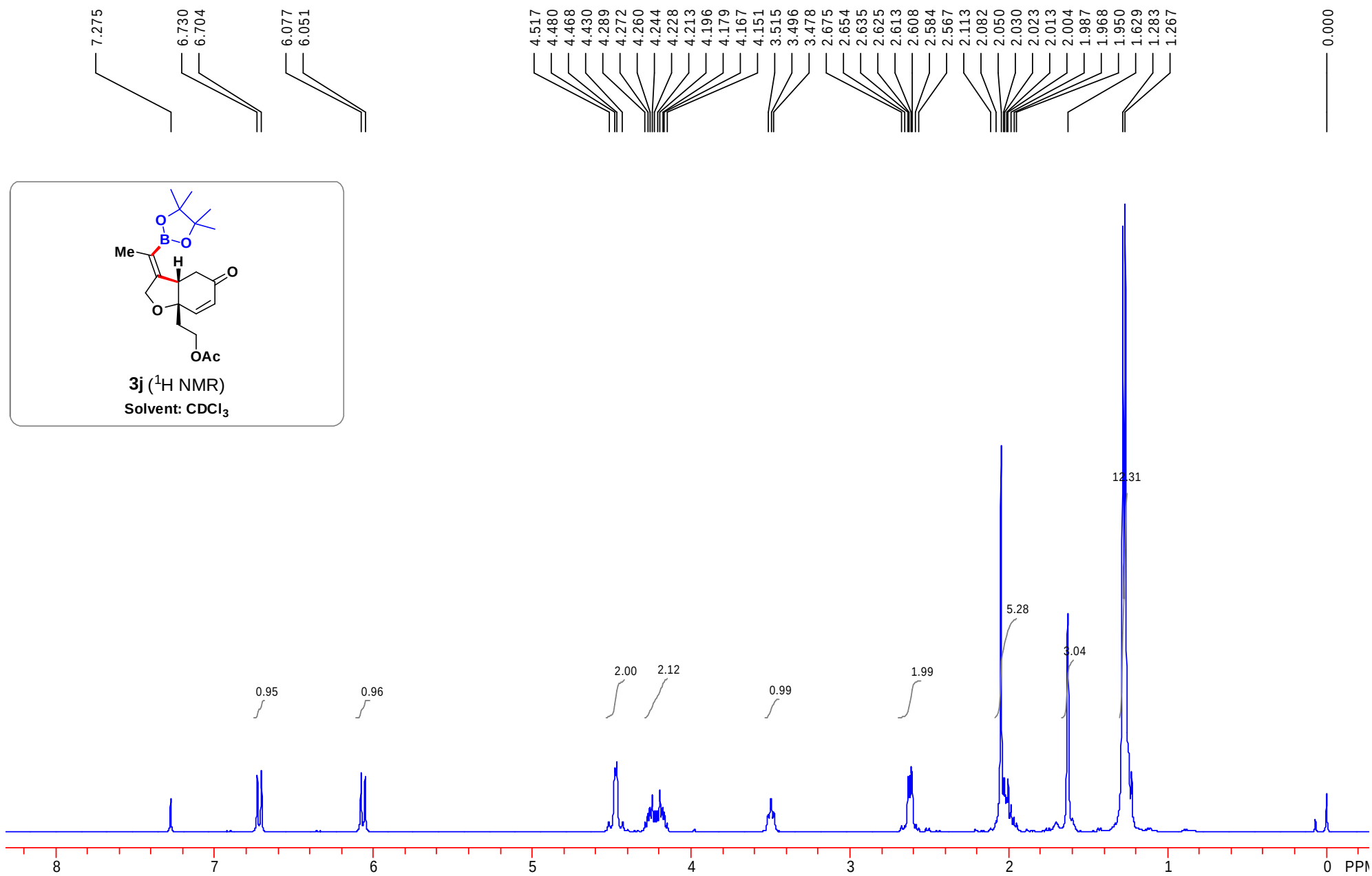




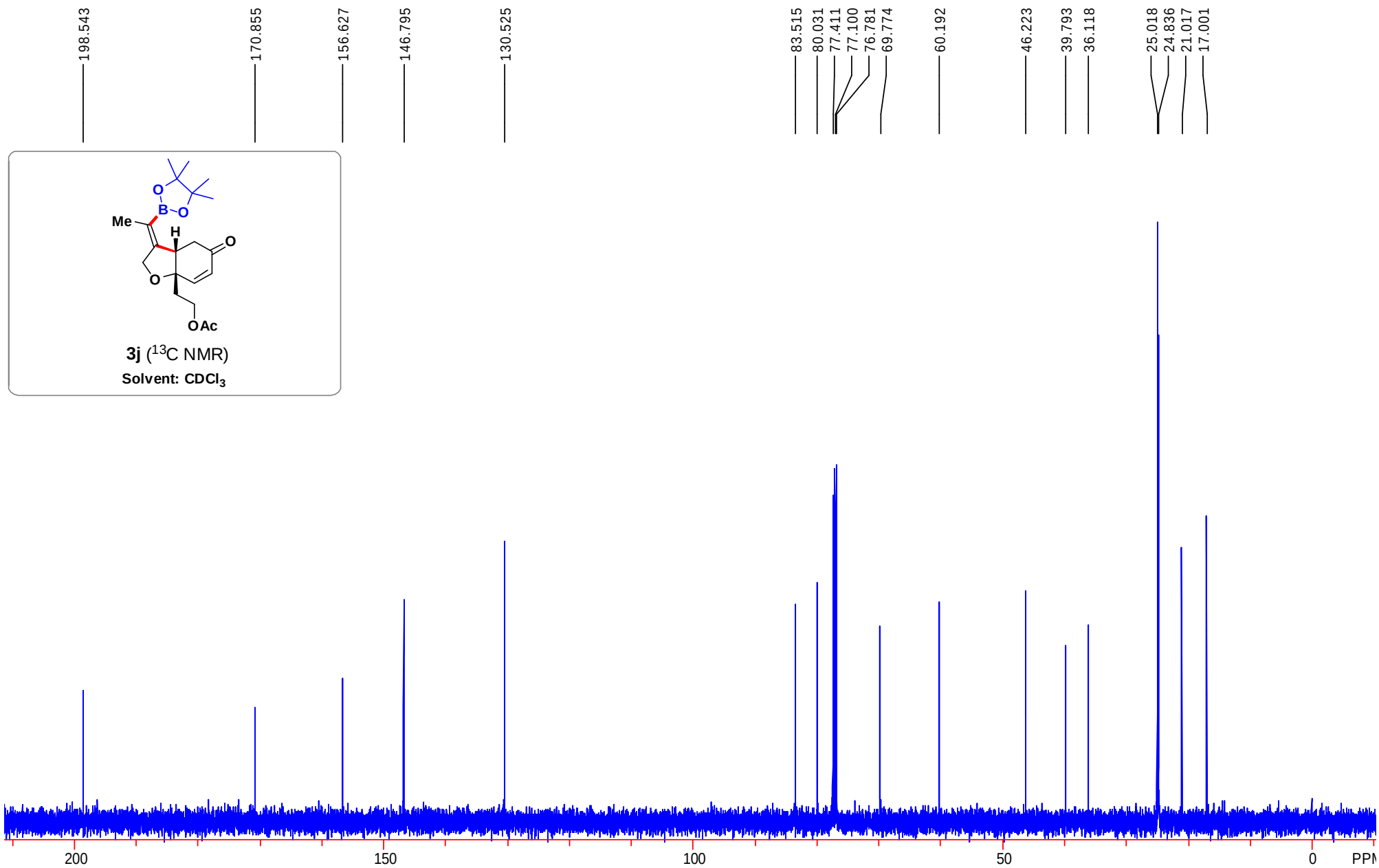
3i (^1H NMR)
Solvent: CDCl_3



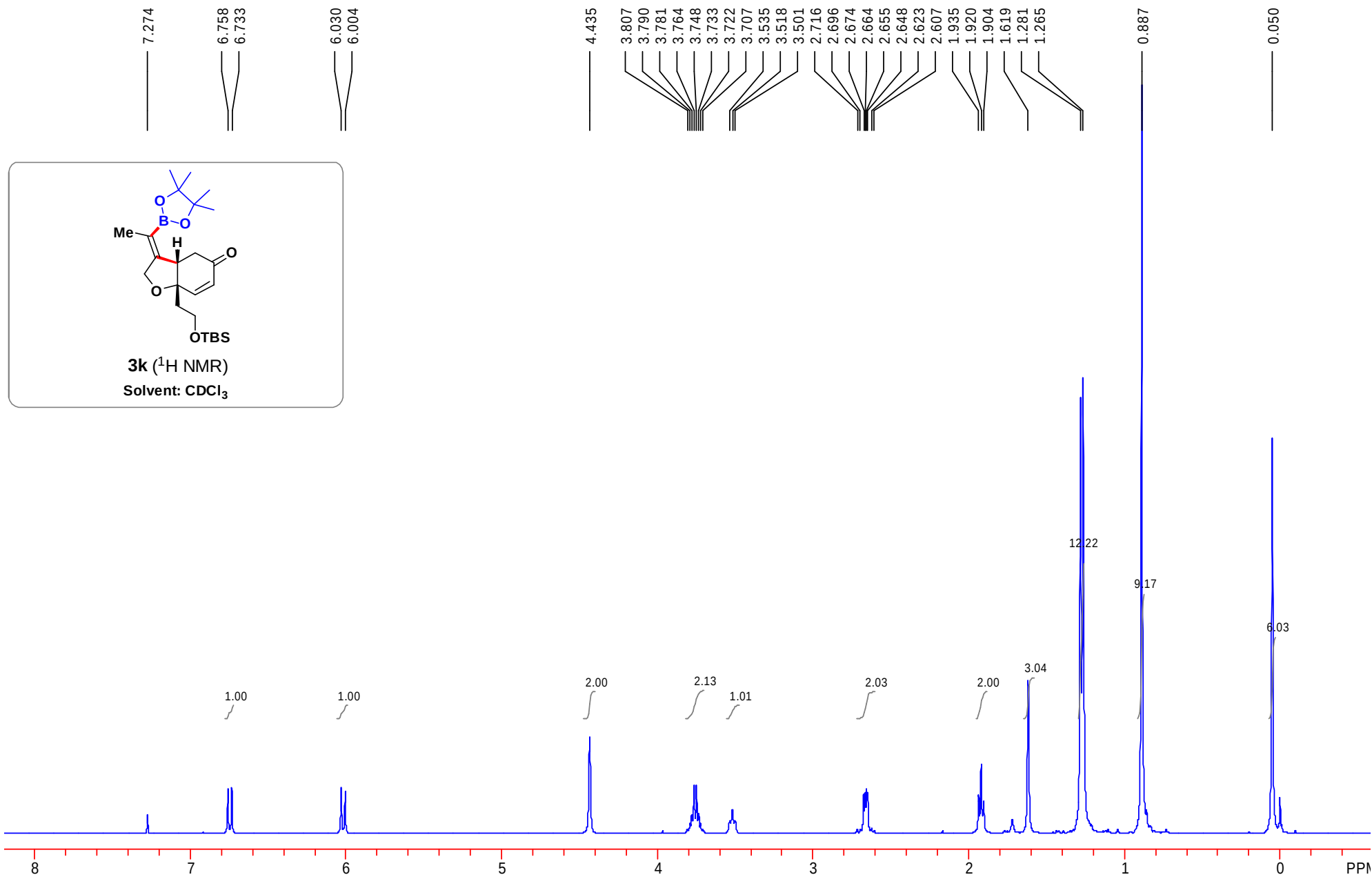




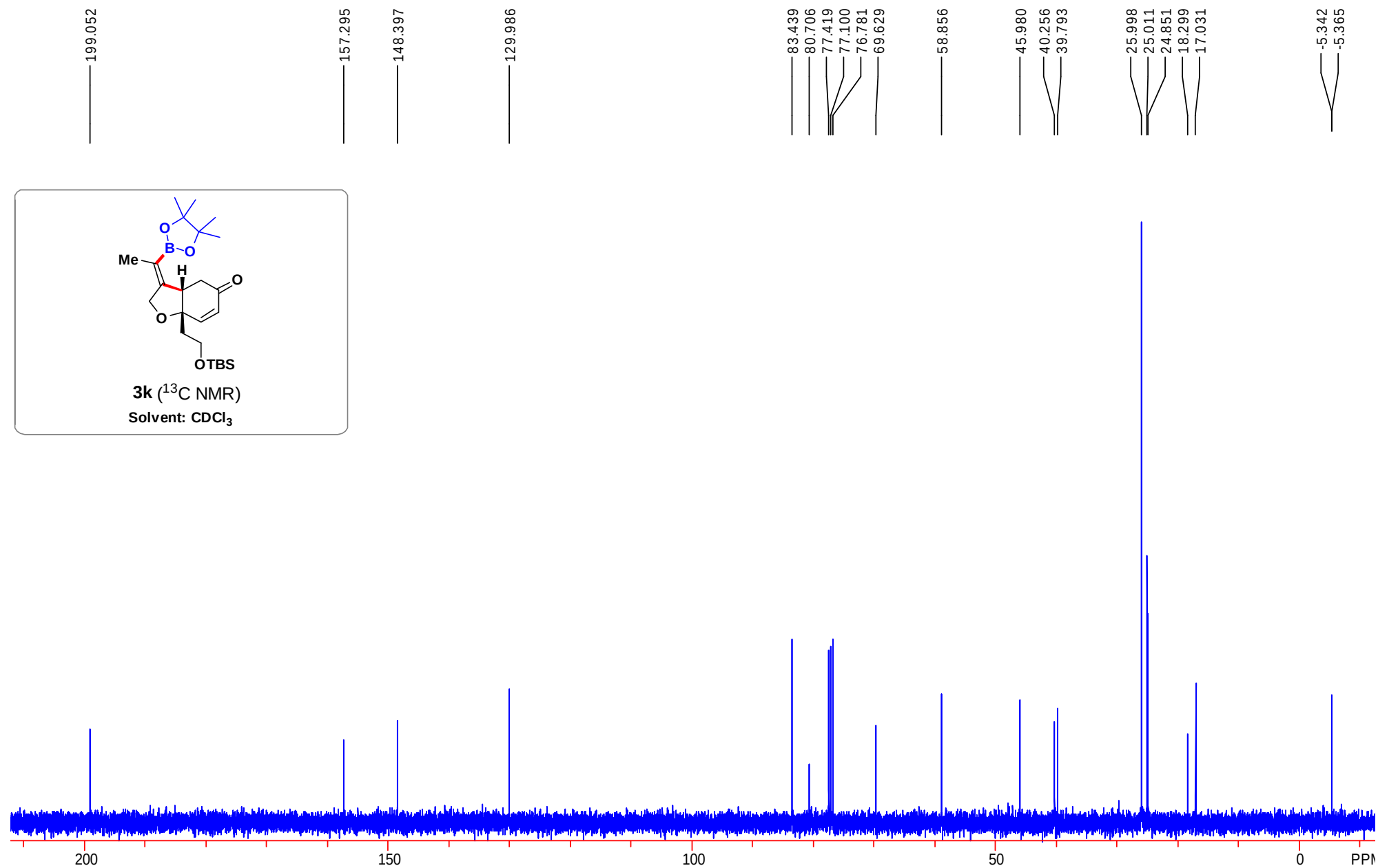
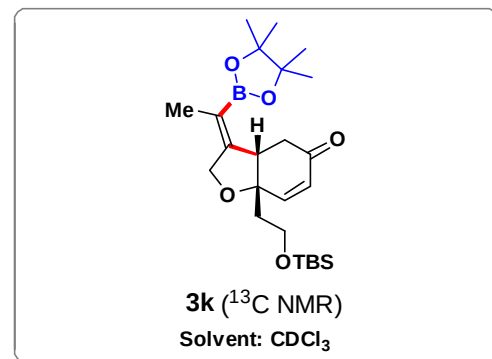
S101

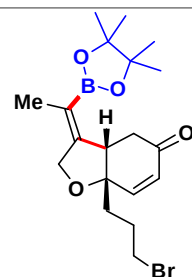


S102

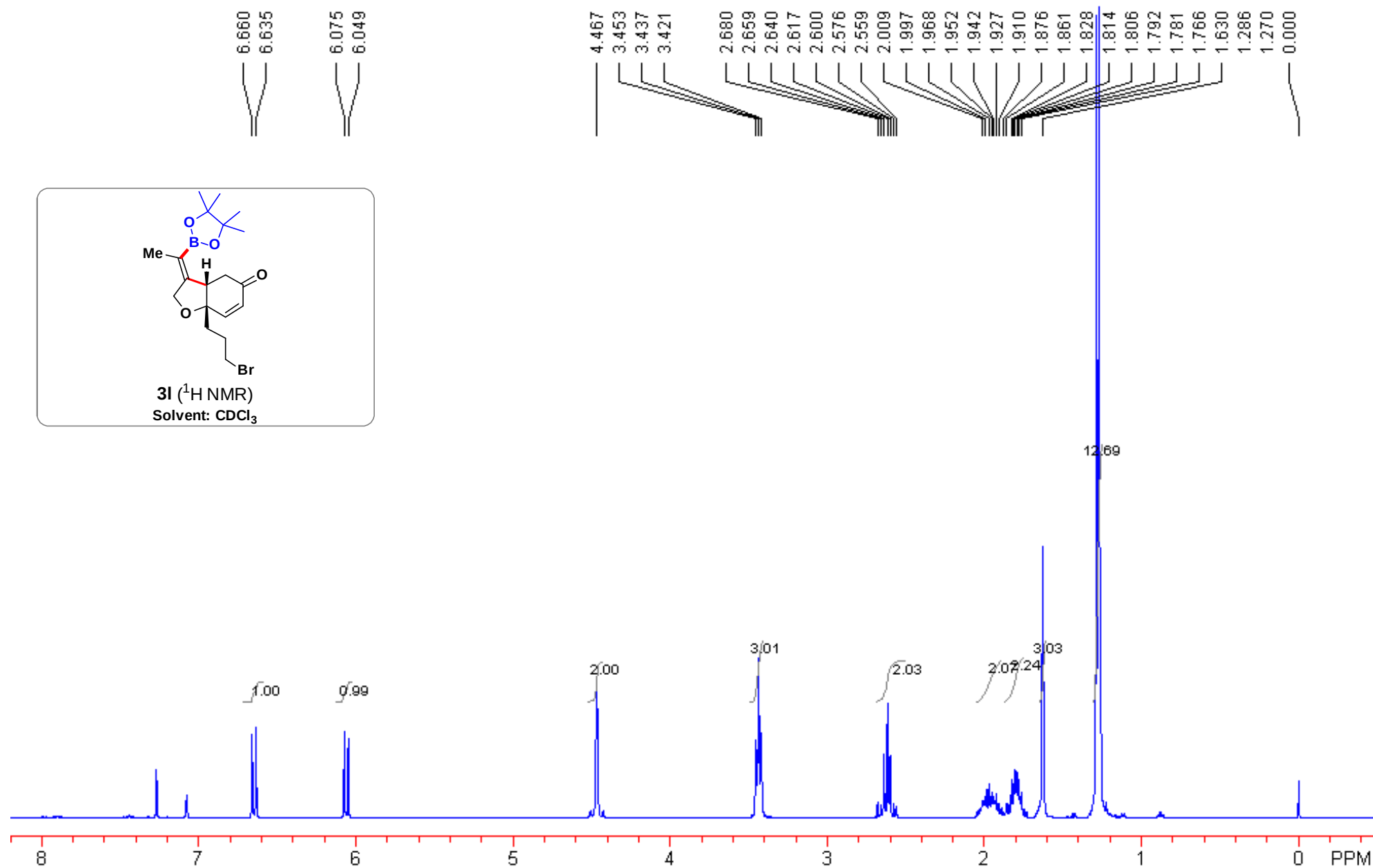


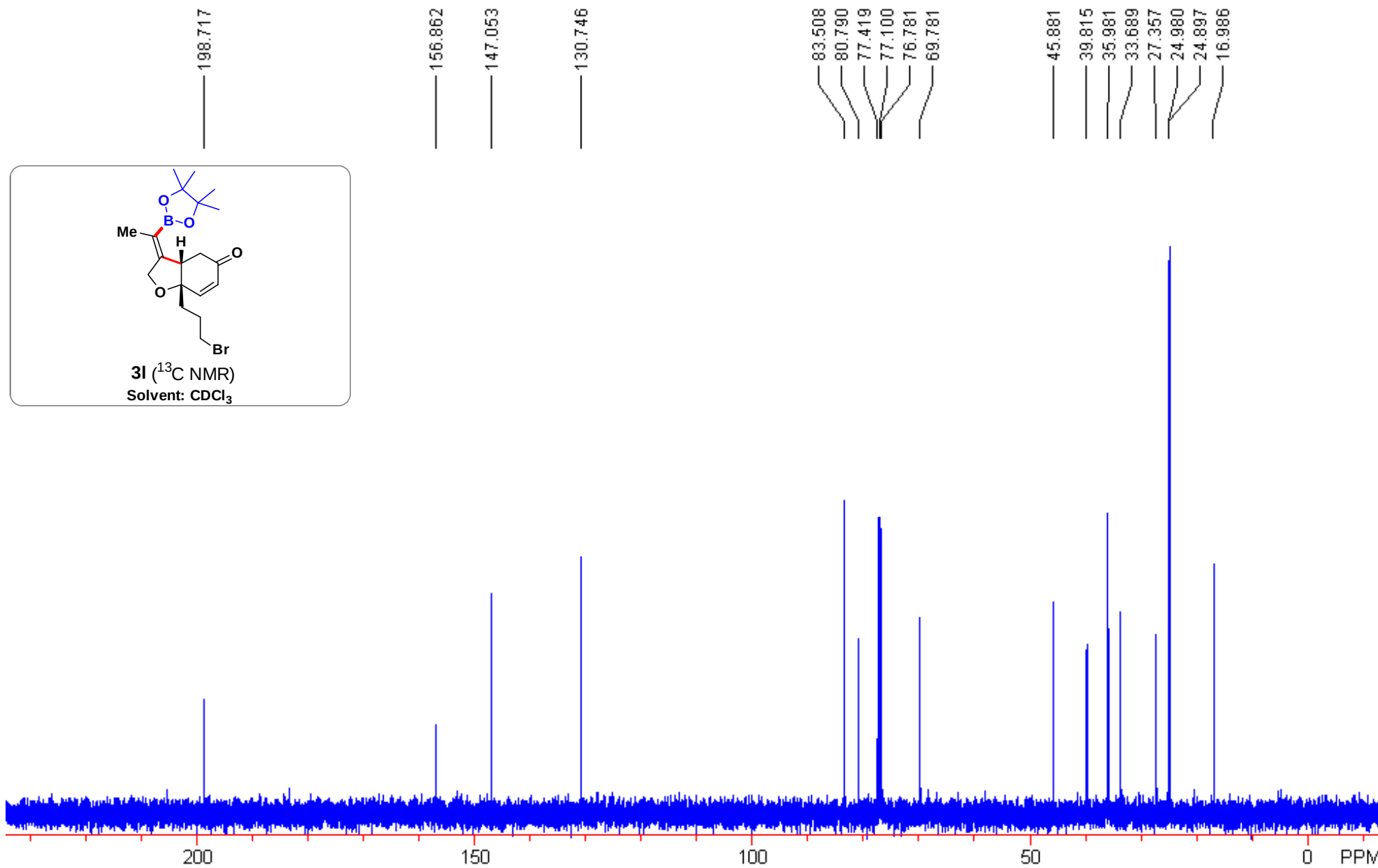
S103

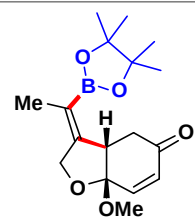
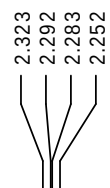
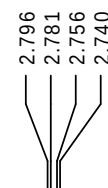
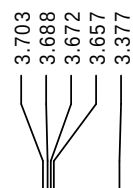
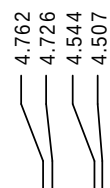
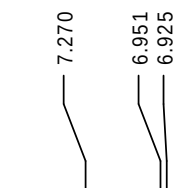




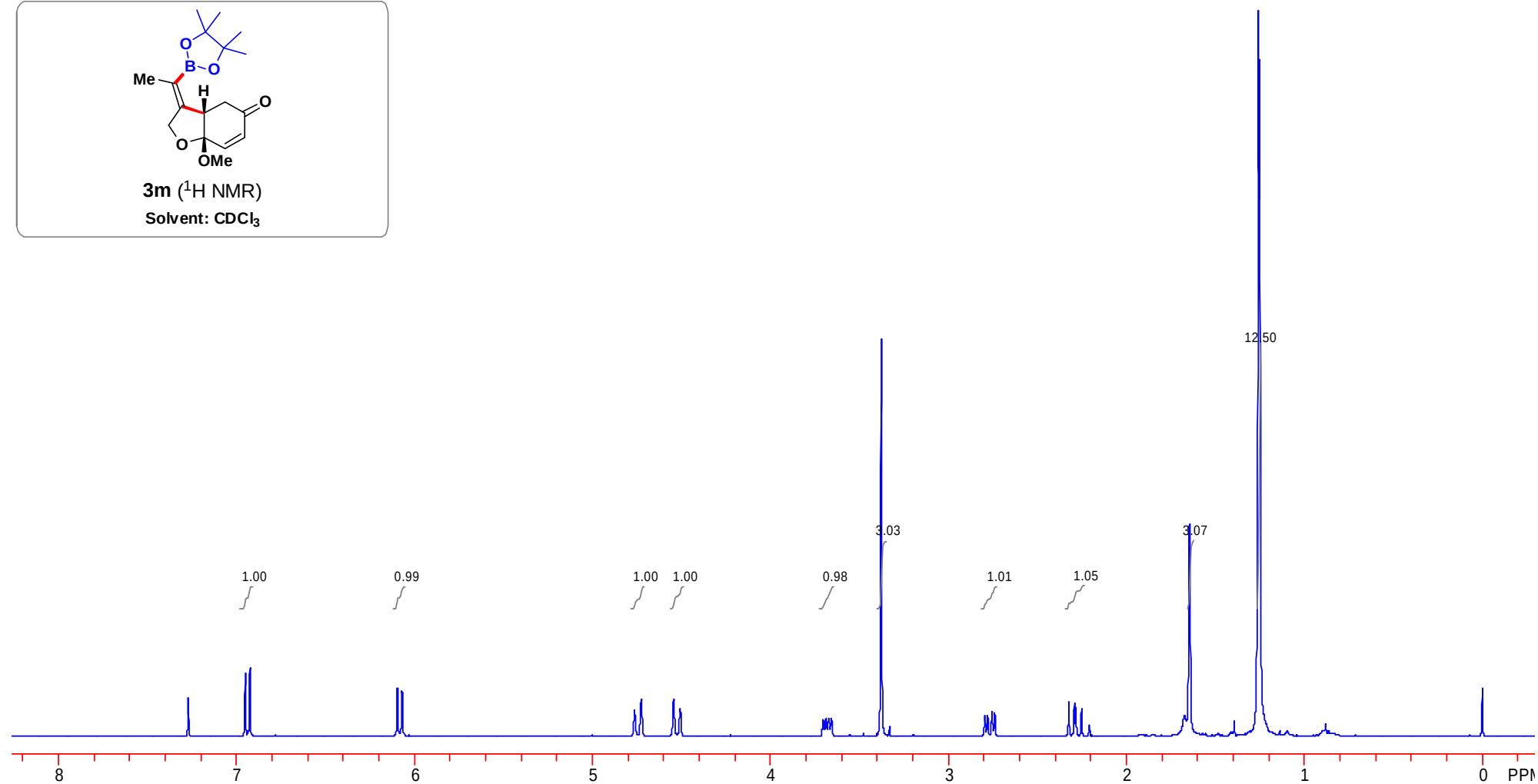
3I (^1H NMR)
Solvent: CDCl_3

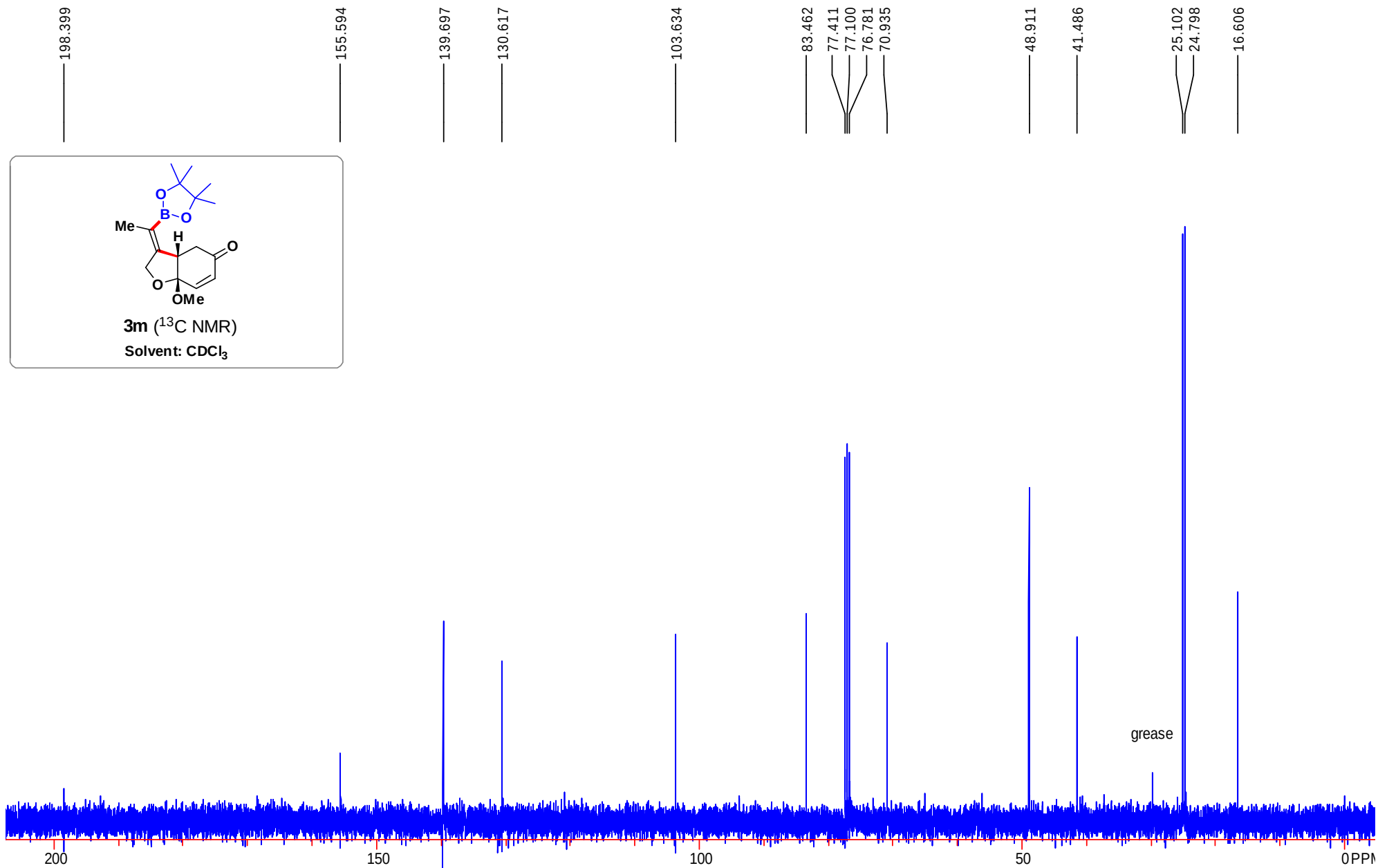


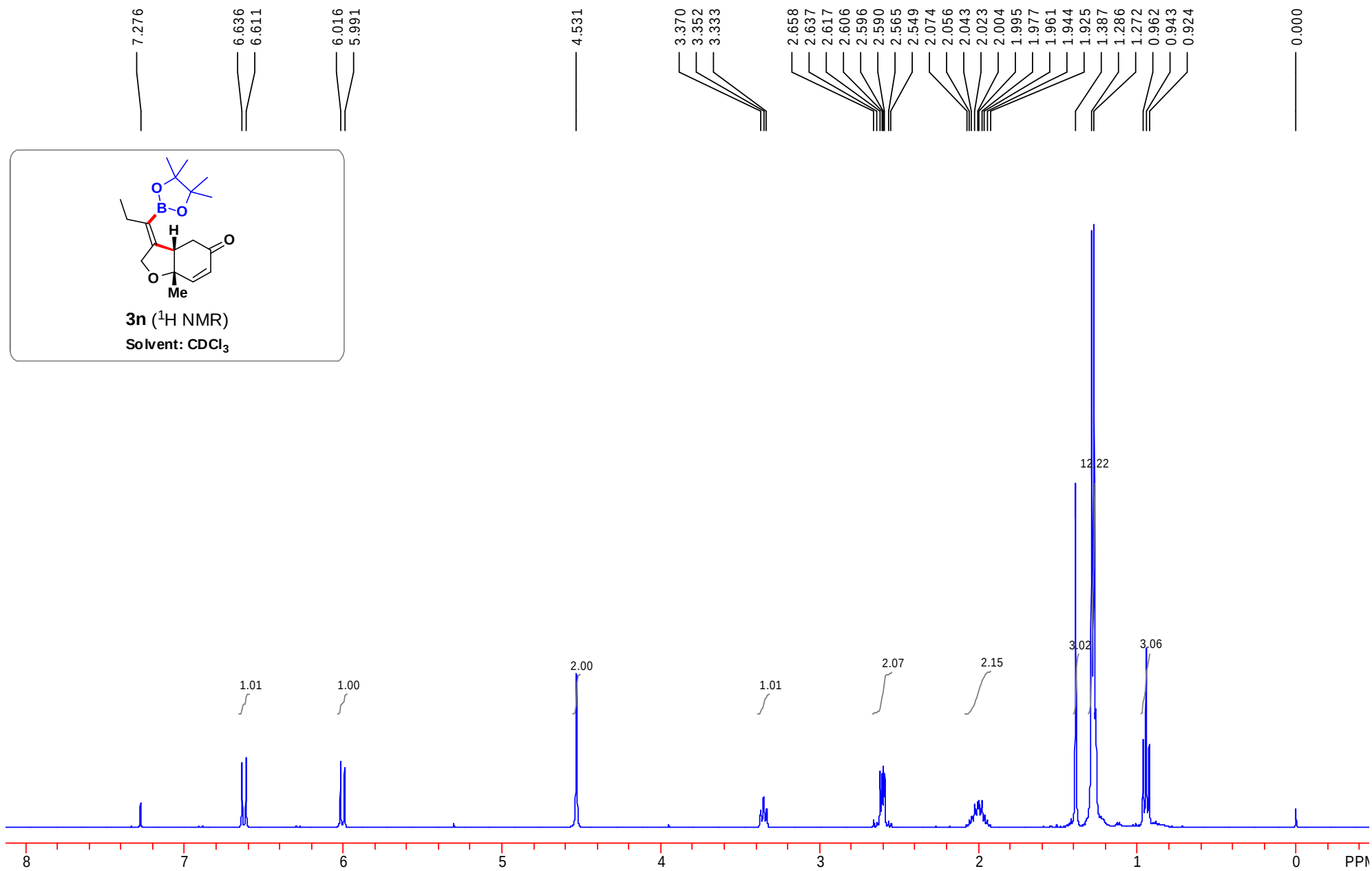


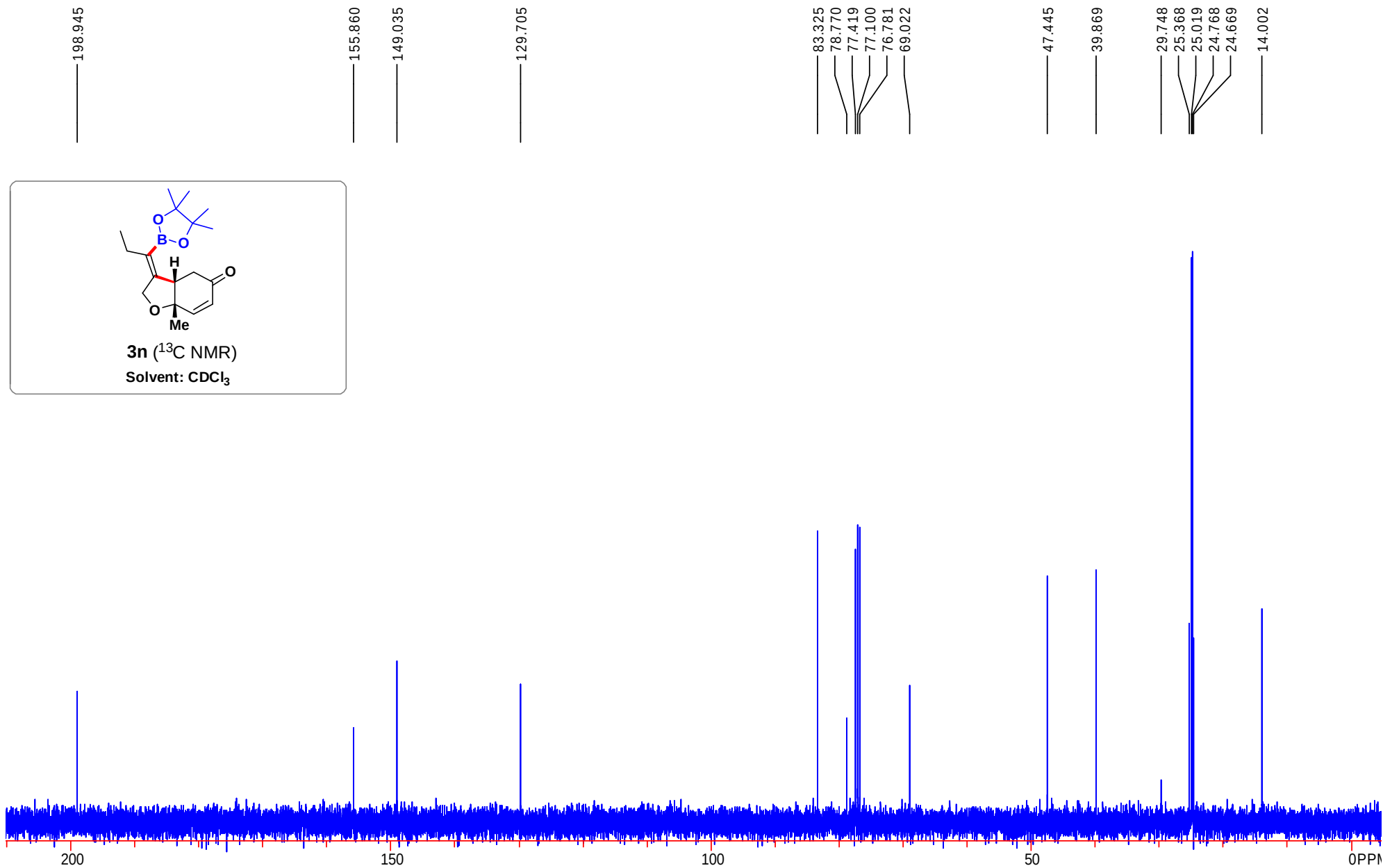


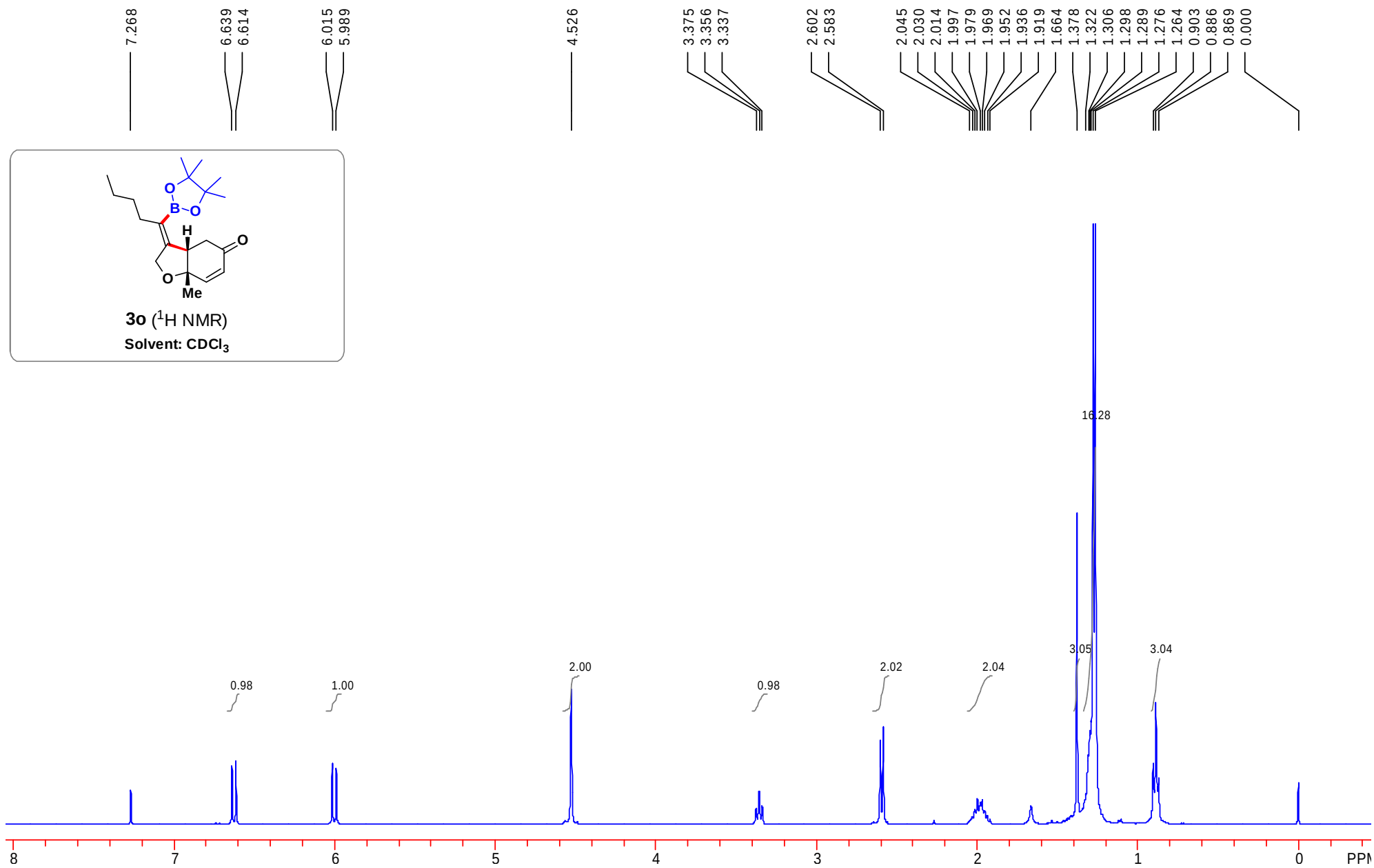
3m (¹H NMR)
Solvent: CDCl₃











199.021

156.095

149.020

129.721

83.326

78.710

77.419

77.100

76.781

69.303

47.552

39.952

31.912

31.829

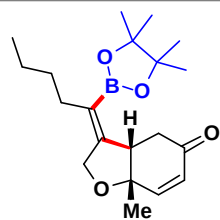
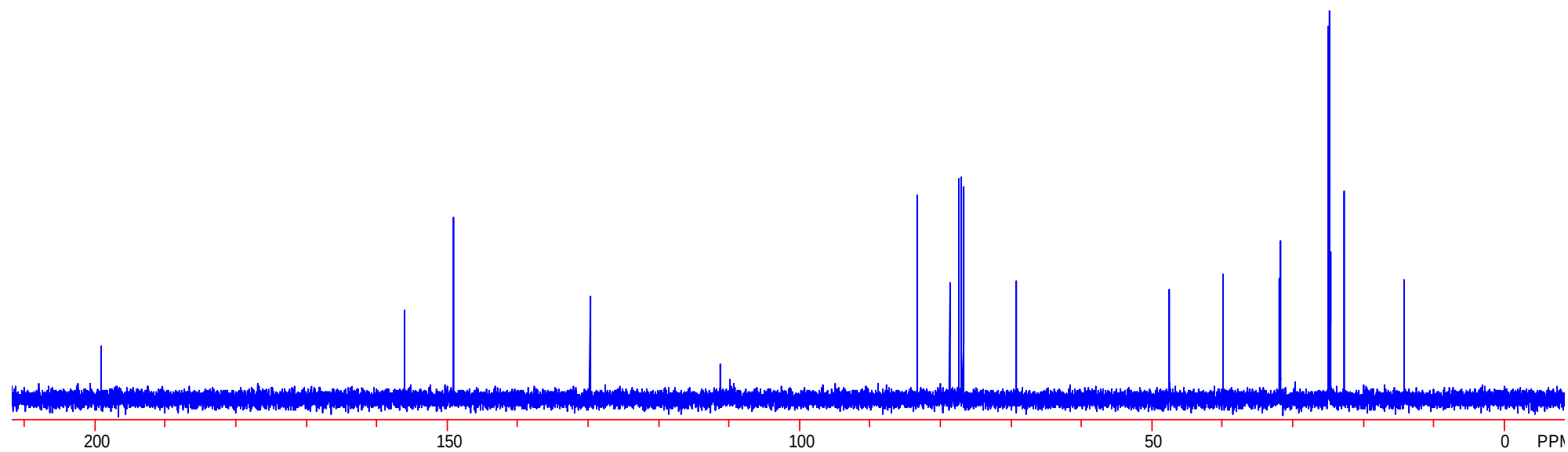
25.011

24.791

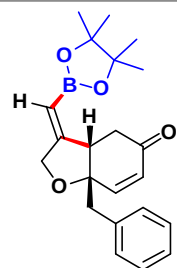
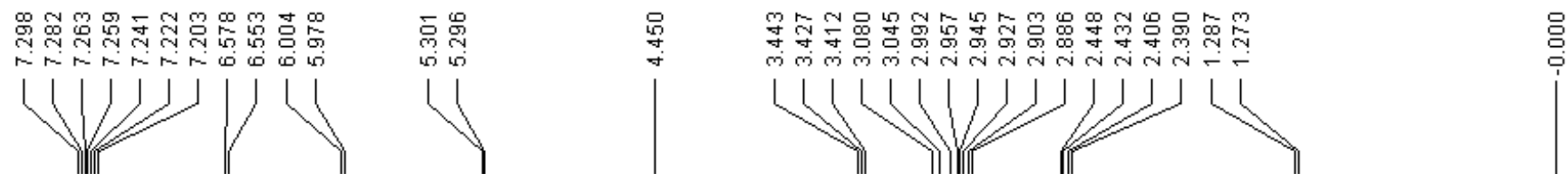
24.738

22.695

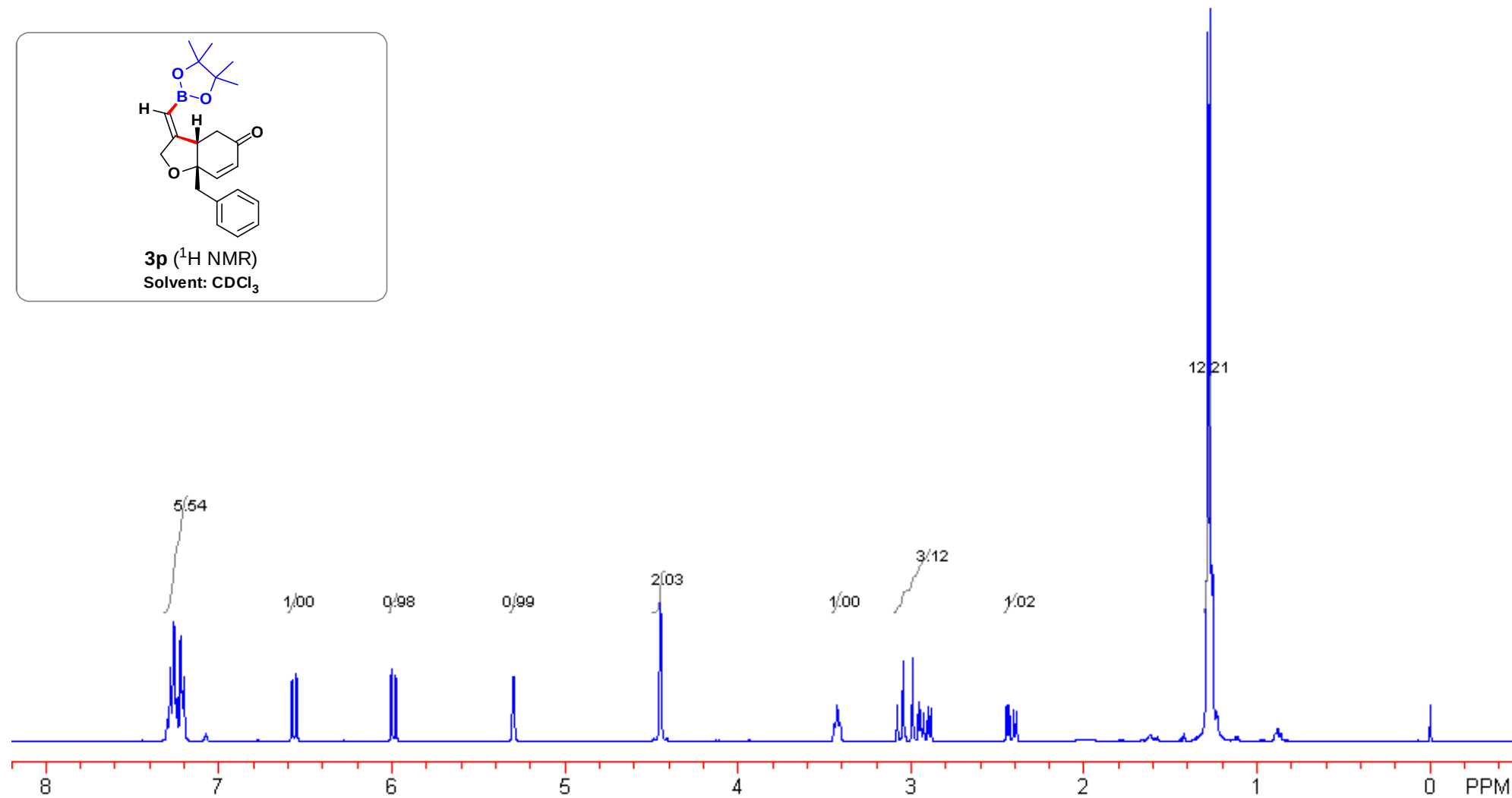
14.185

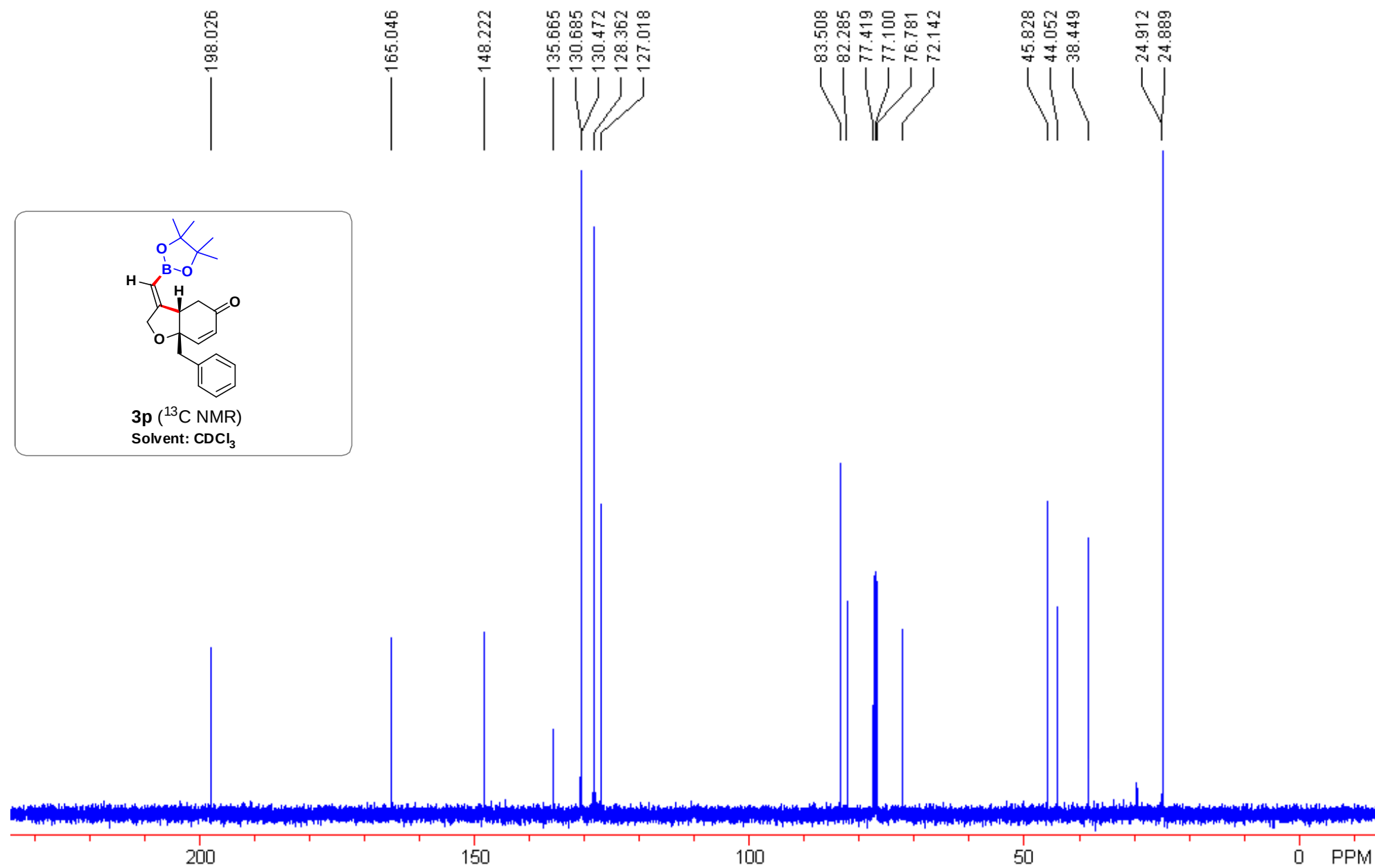
**30** (^1H NMR)Solvent: CDCl_3 

S112



3p (^1H NMR)
Solvent: CDCl_3



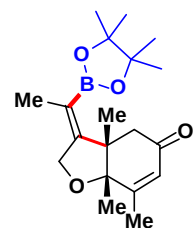


7.273

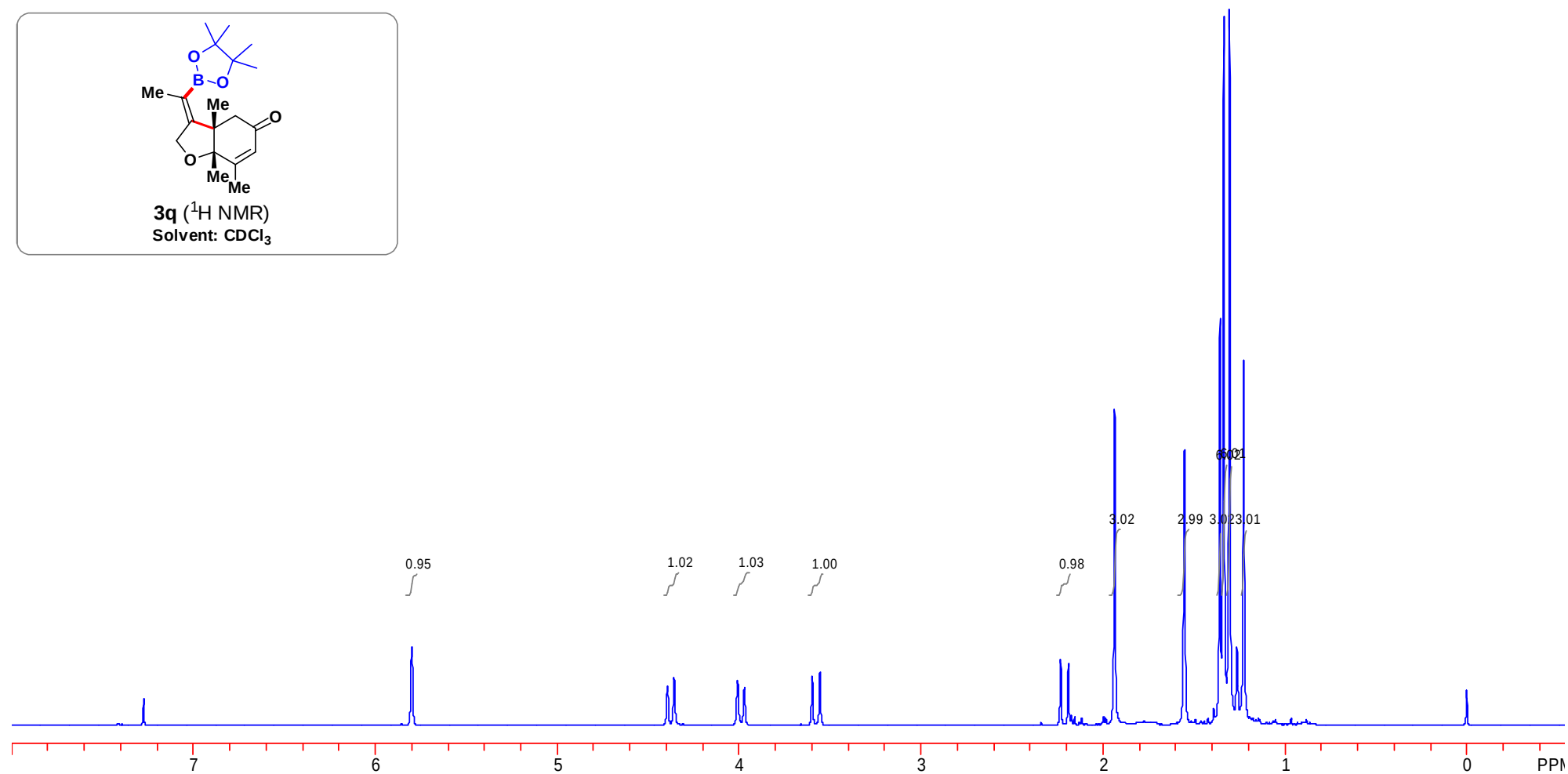
5.798

4.394
4.3574.009
3.9723.598
3.5552.232
2.189
1.9351.552
1.357
1.336
1.304
1.225

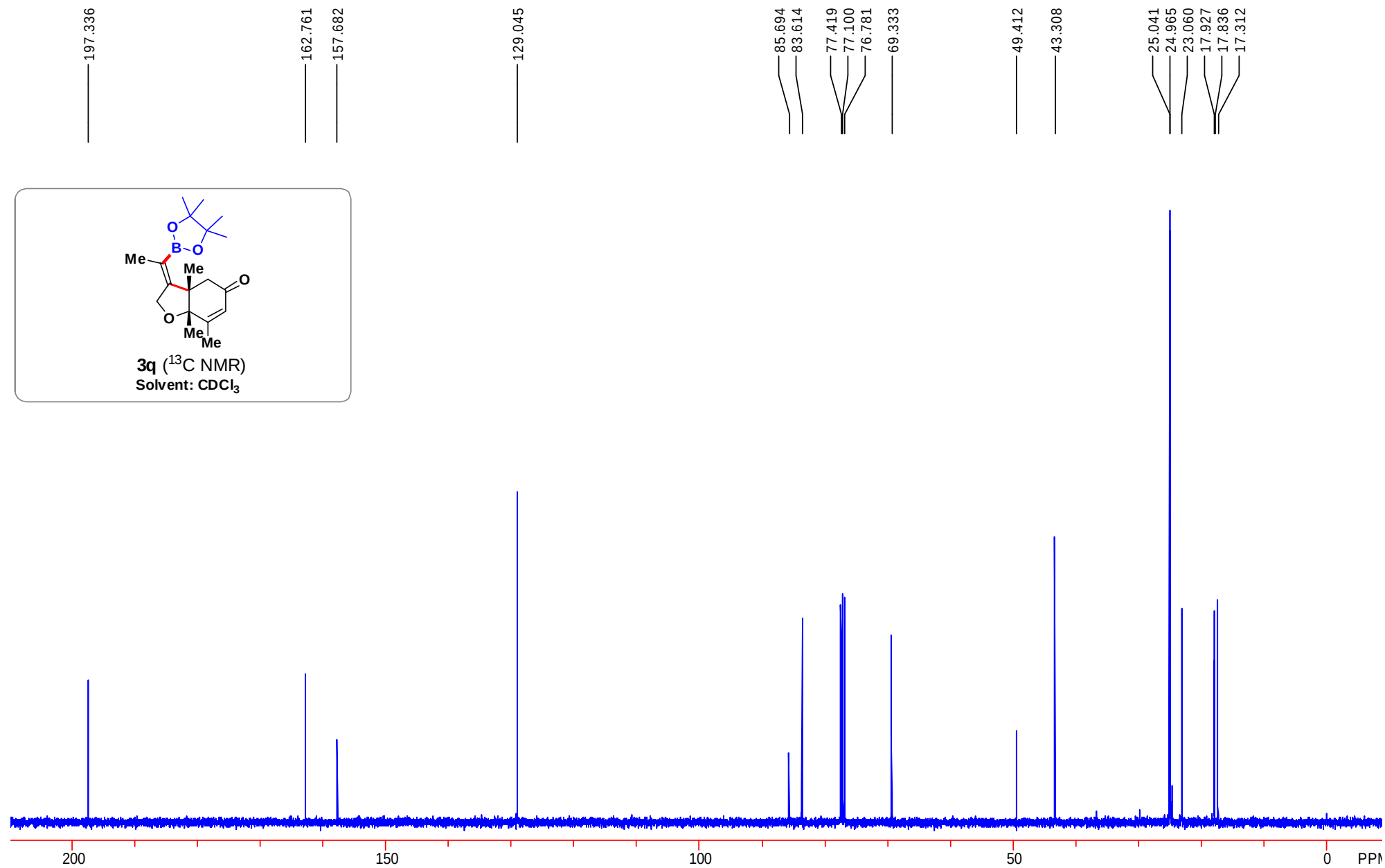
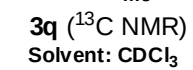
0.000



3q (^1H NMR)
Solvent: CDCl_3



S115



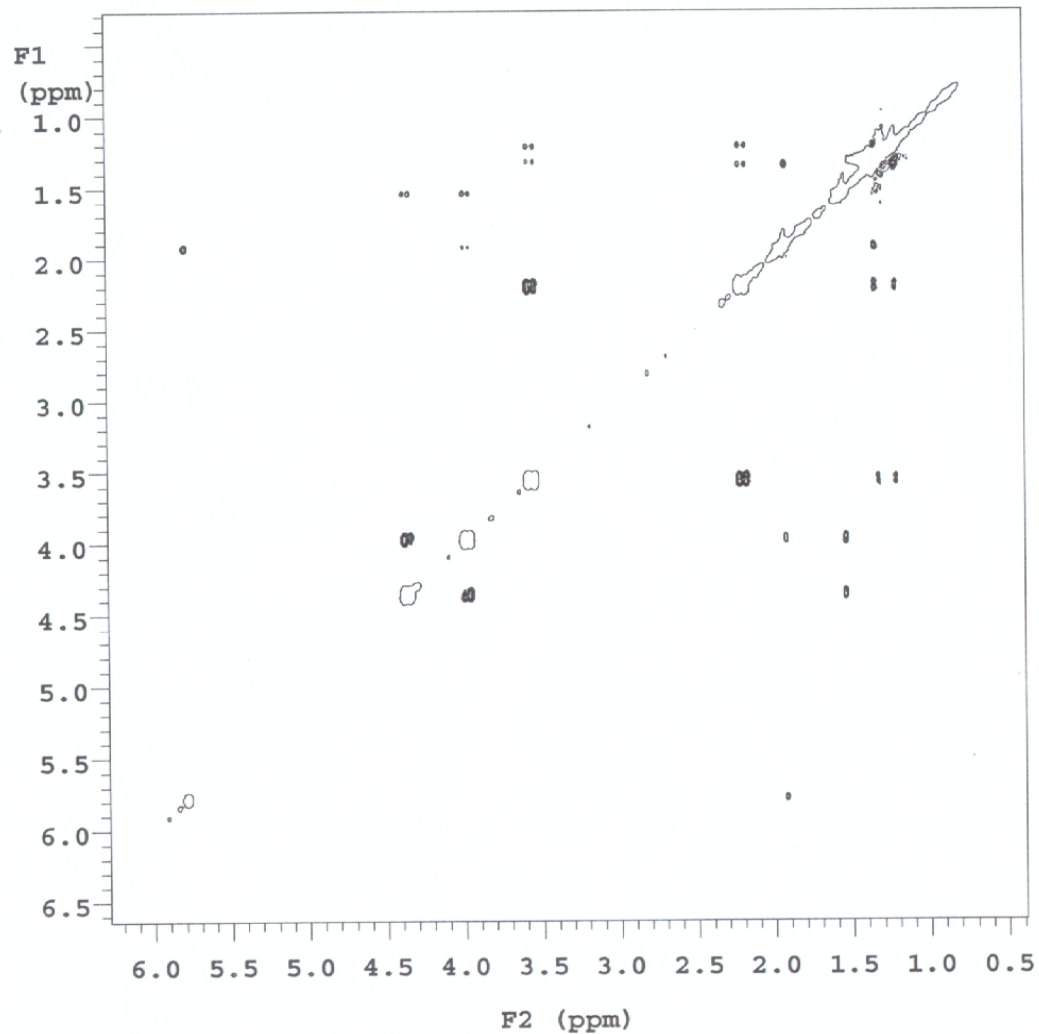
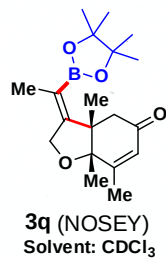
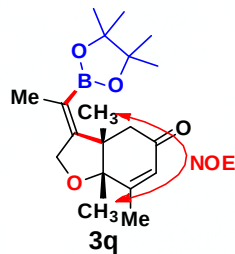
20120531p2-096-1b

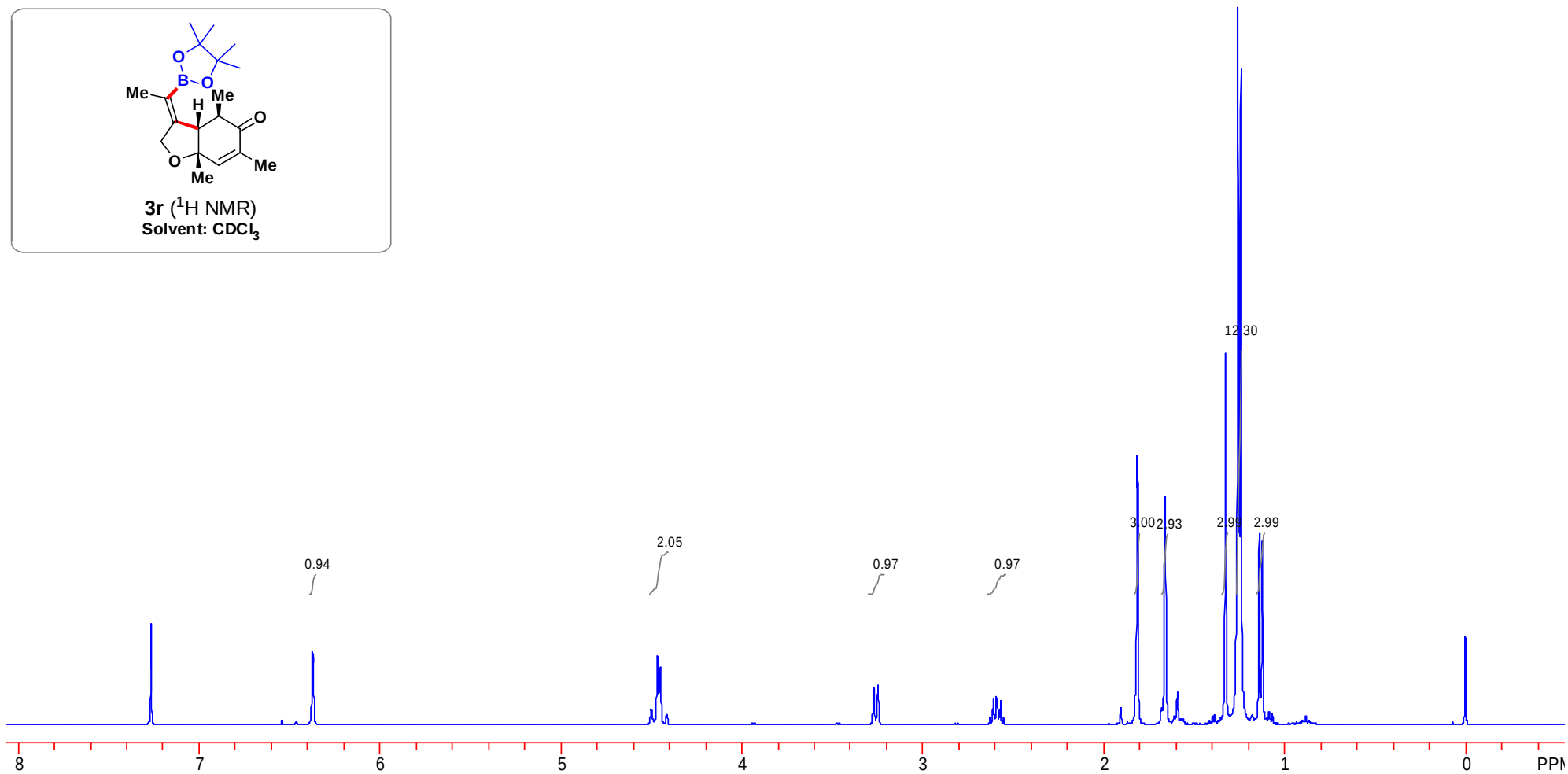
Sample Name:
20120531p2-096-1b
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p2-096-1b_20130228_01
FidFile: NOESY_01

Pulse Sequence: NOESY
Solvent: cdcl3
Data collected on: Feb 28 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3811.0 Hz
2D Width 3811.0 Hz
32 repetitions
2 x 256 increments
OBSERVE H1, 399.6607443 MHz
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.037 sec
FT size 2048 x 2048
Total time 8 hr, 36 min





202.103

156.437

142.794

135.574

83.348

78.884

77.419

77.100

76.781

69.174

54.005

43.353

26.241

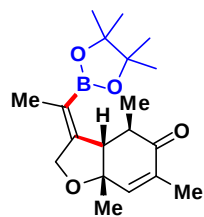
25.087

24.852

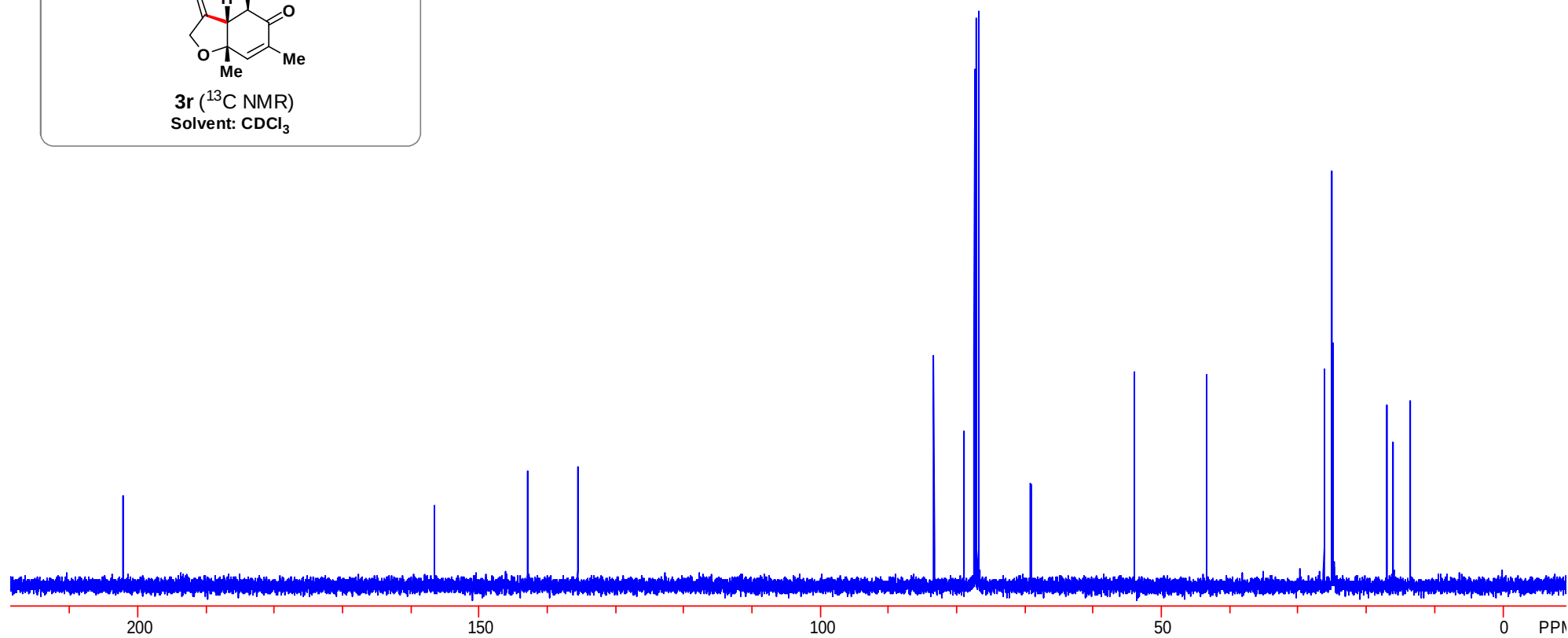
17.024

16.083

13.638



3r (^{13}C NMR)
Solvent: CDCl_3



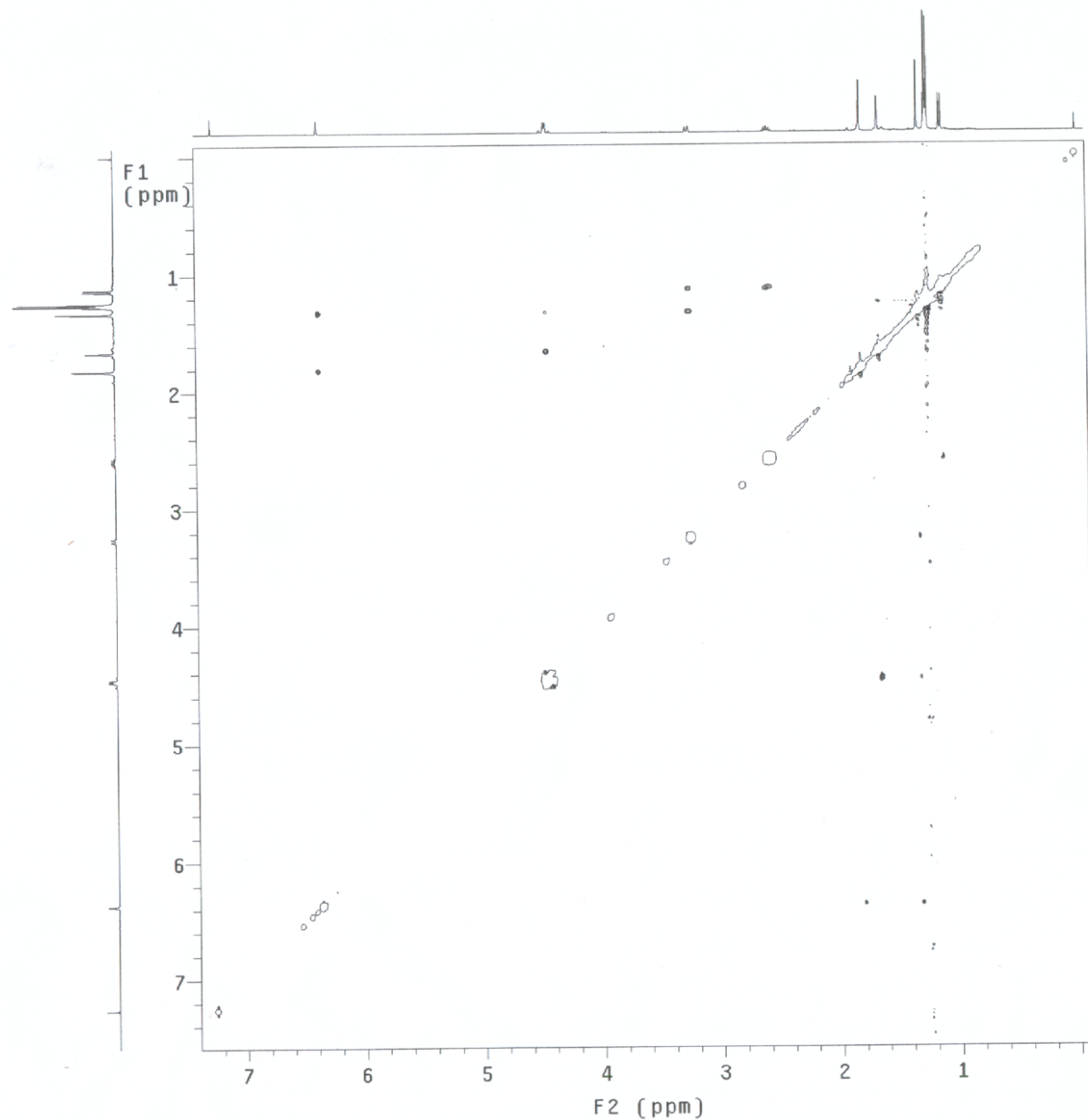
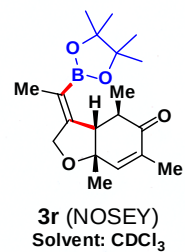
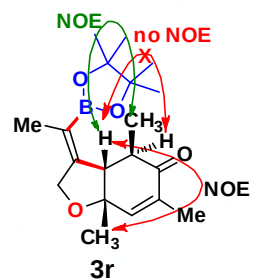
20120531p2-078-1a

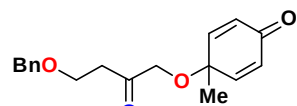
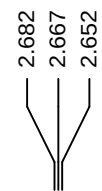
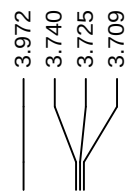
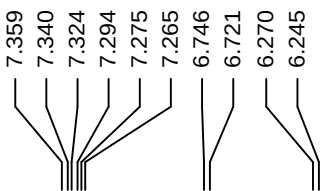
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20120531p2-078-1a
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p2-078-1a_20130128_01
FidFile: NOESY

Pulse Sequence: NOESY
Solvent: cdcl3
Data collected on: Jan 28 2013

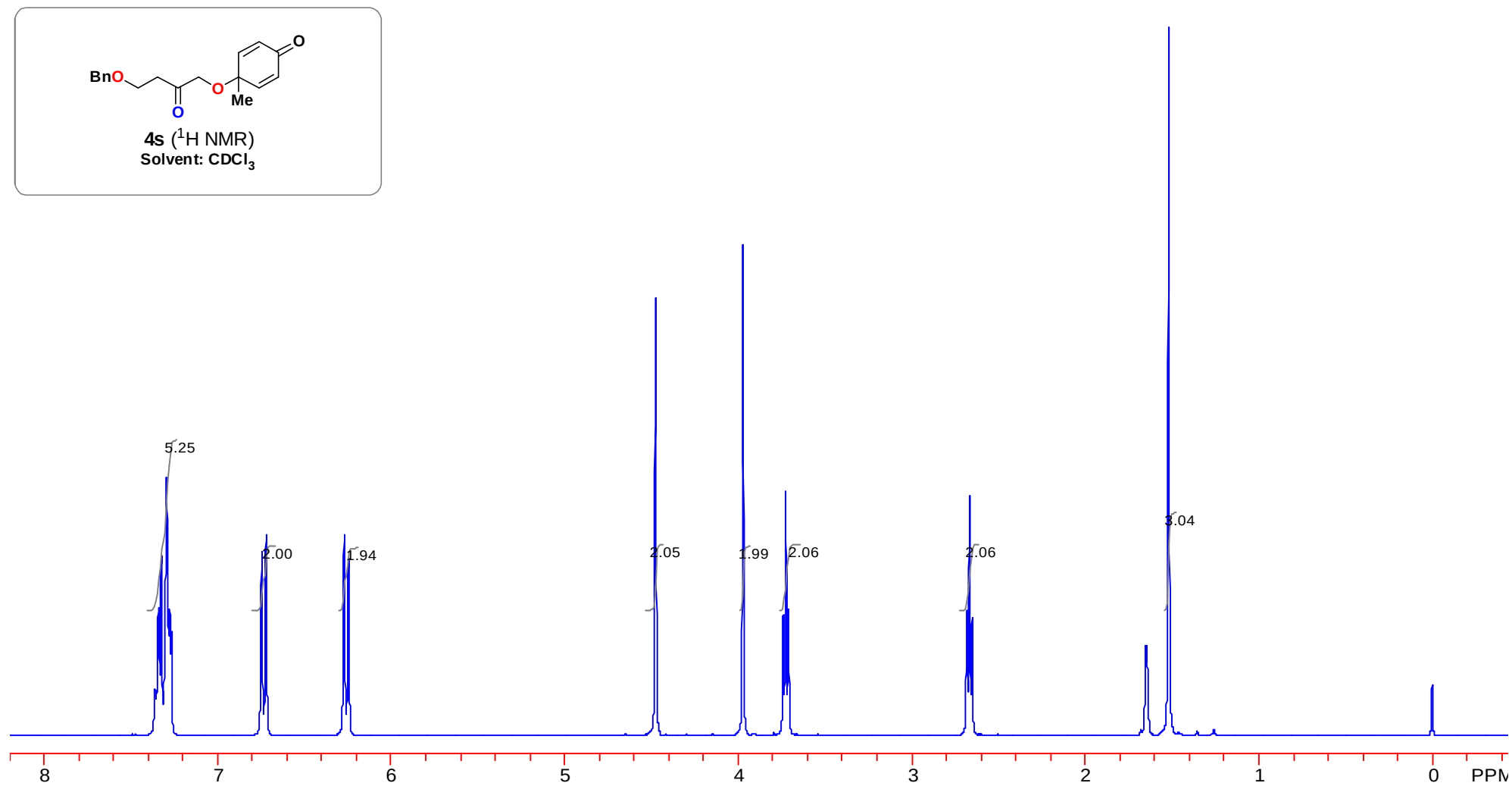
Temp. 25.0 C / 298.1 K
Operator: sioc

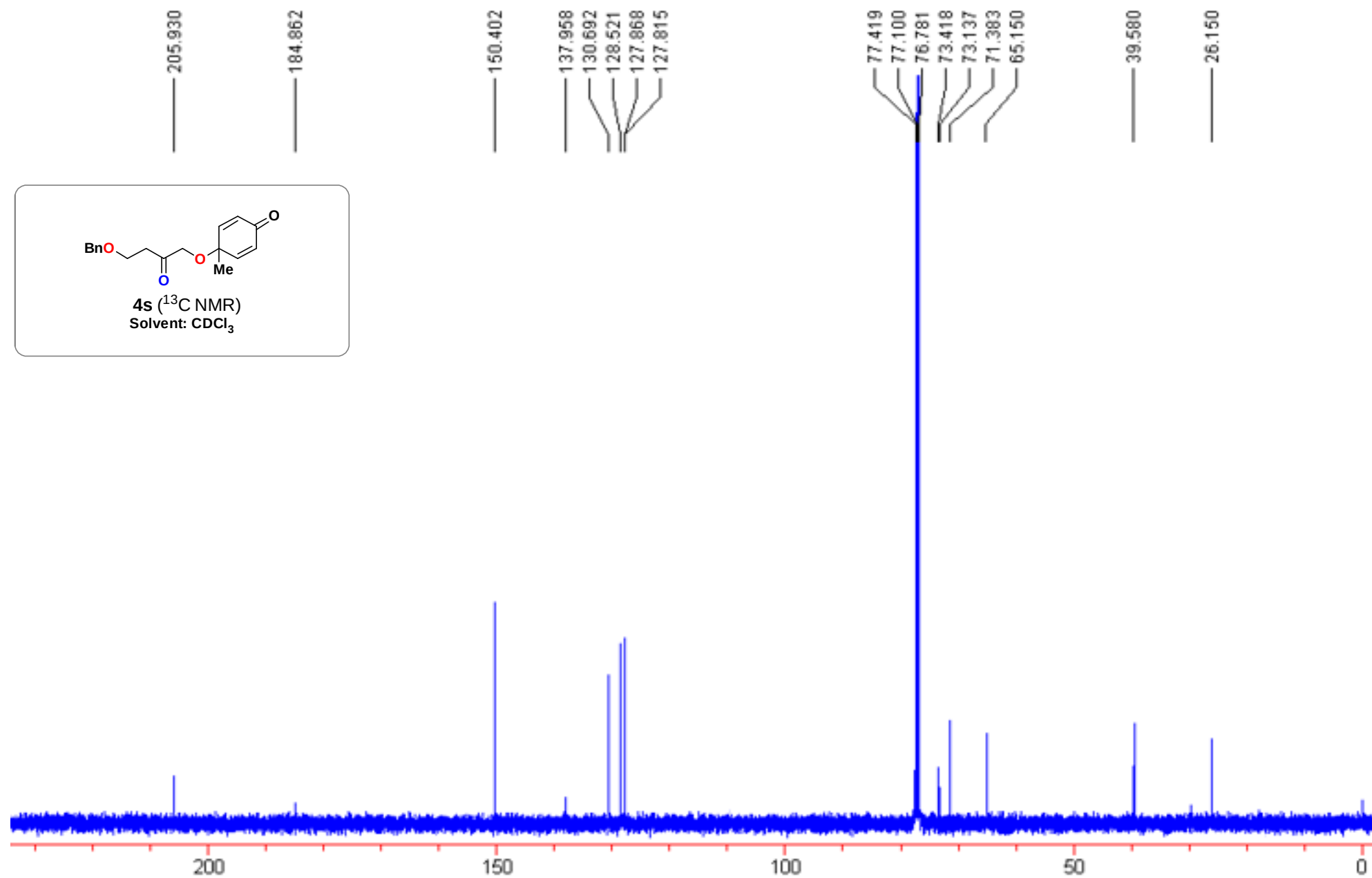
Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4401.4 Hz
2D Width 4401.4 Hz
8 repetitions
2 x 256 increments
OBSERVE H1, 399.6613957 MHz
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.048 sec
FT size 2048 x 2048
Total time 2 hr, 2 min





4s (¹H NMR)
Solvent: CDCl₃





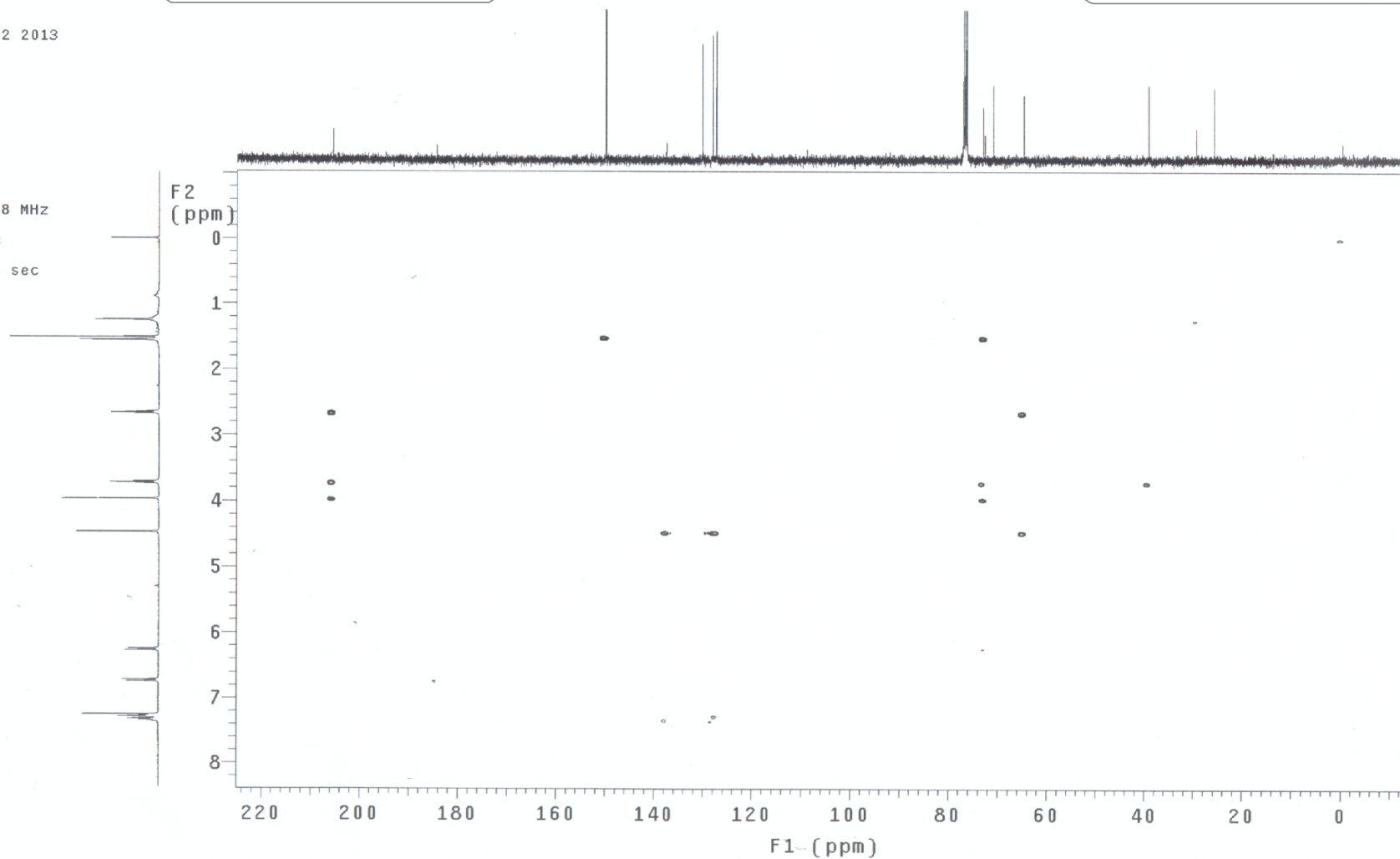
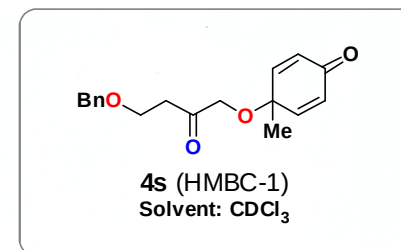
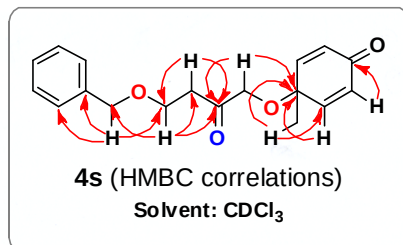
2012576yf02-10-01-01

Sample Name:
2012576yf02-10-01-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-10-01-01_20130402_01
FidFile: gHMBCAD_01

Pulse Sequence: gHMBCAD
Solvent: cdc13
Data collected on: Apr 2 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3765.1 Hz
2D Width 24118.2 Hz
8 repetitions
2 x 200 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.008 sec
FT size 2048 x 2048
Total time 1 hr, 6 min



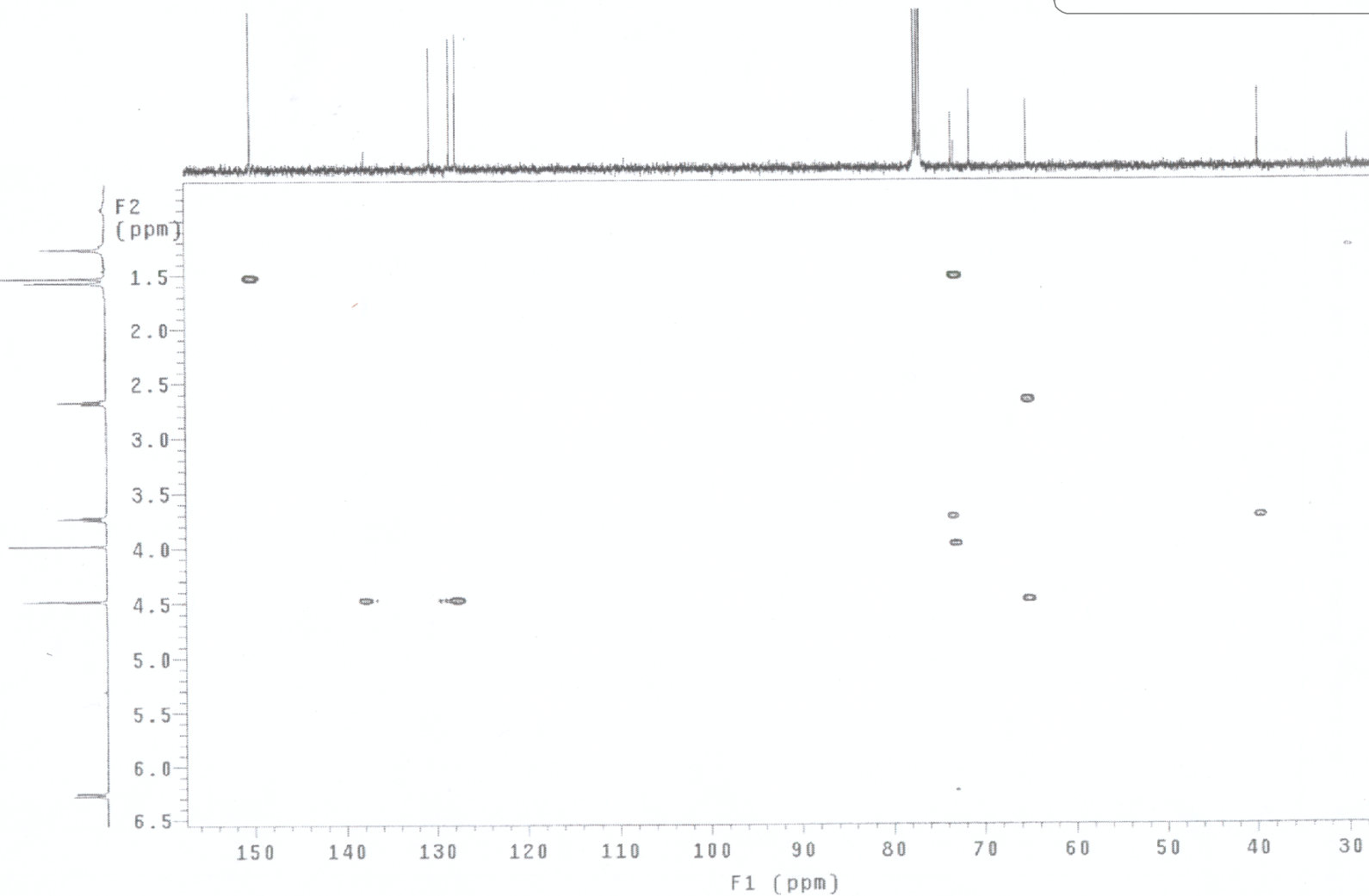
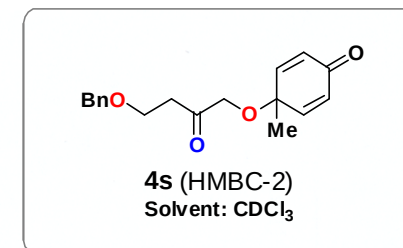
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Sample Name:
2012576yf02-10-01-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-10-01-01_20130402_01
FidFile: gHMBCAD_01

Pulse Sequence: gHMBCAD
Solvent: cdcl3
Data collected on: Apr 2 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3765.1 Hz
2D Width 24118.2 Hz
8 repetitions
2 x 200 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.008 sec
FT size 2048 x 2048
Total time 1 hr, 6 min



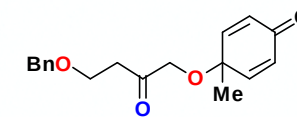
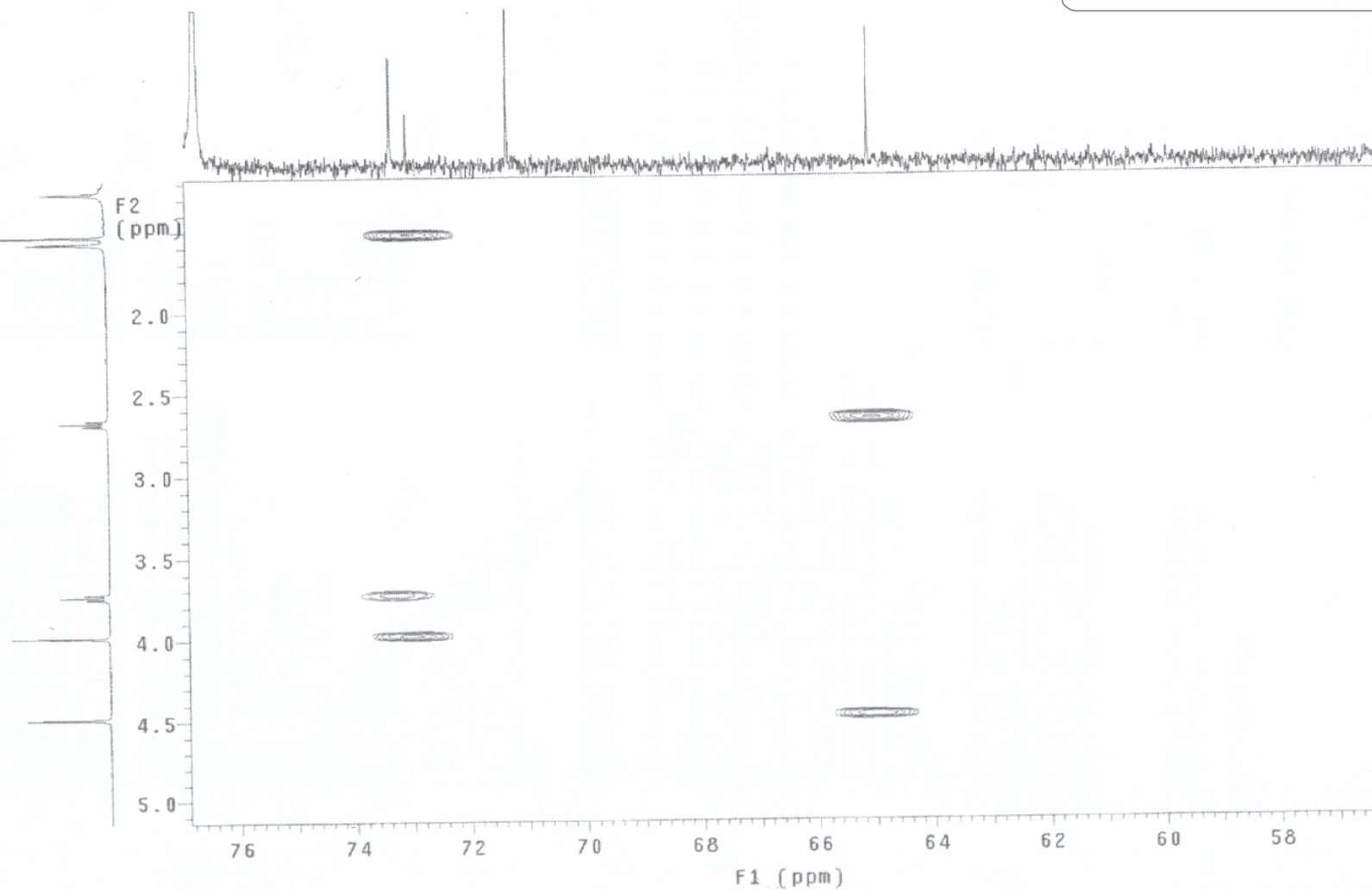
2012576yf02-10-01-01

Sample Name:
2012576yf02-10-01-01
Data Collected on:
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Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-10-01-01_20130402_01
FidFile: gHMBCAD_01

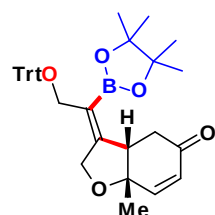
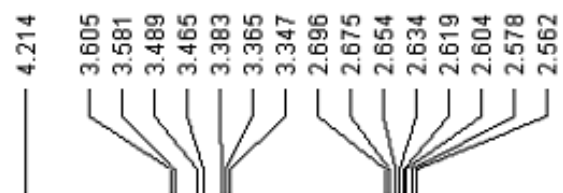
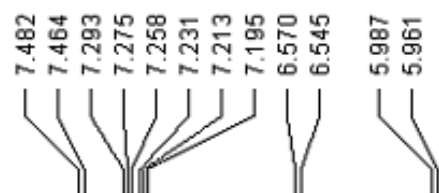
Pulse Sequence: gHMBCAD
Solvent: cdcl3
Data collected on: Apr 2 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

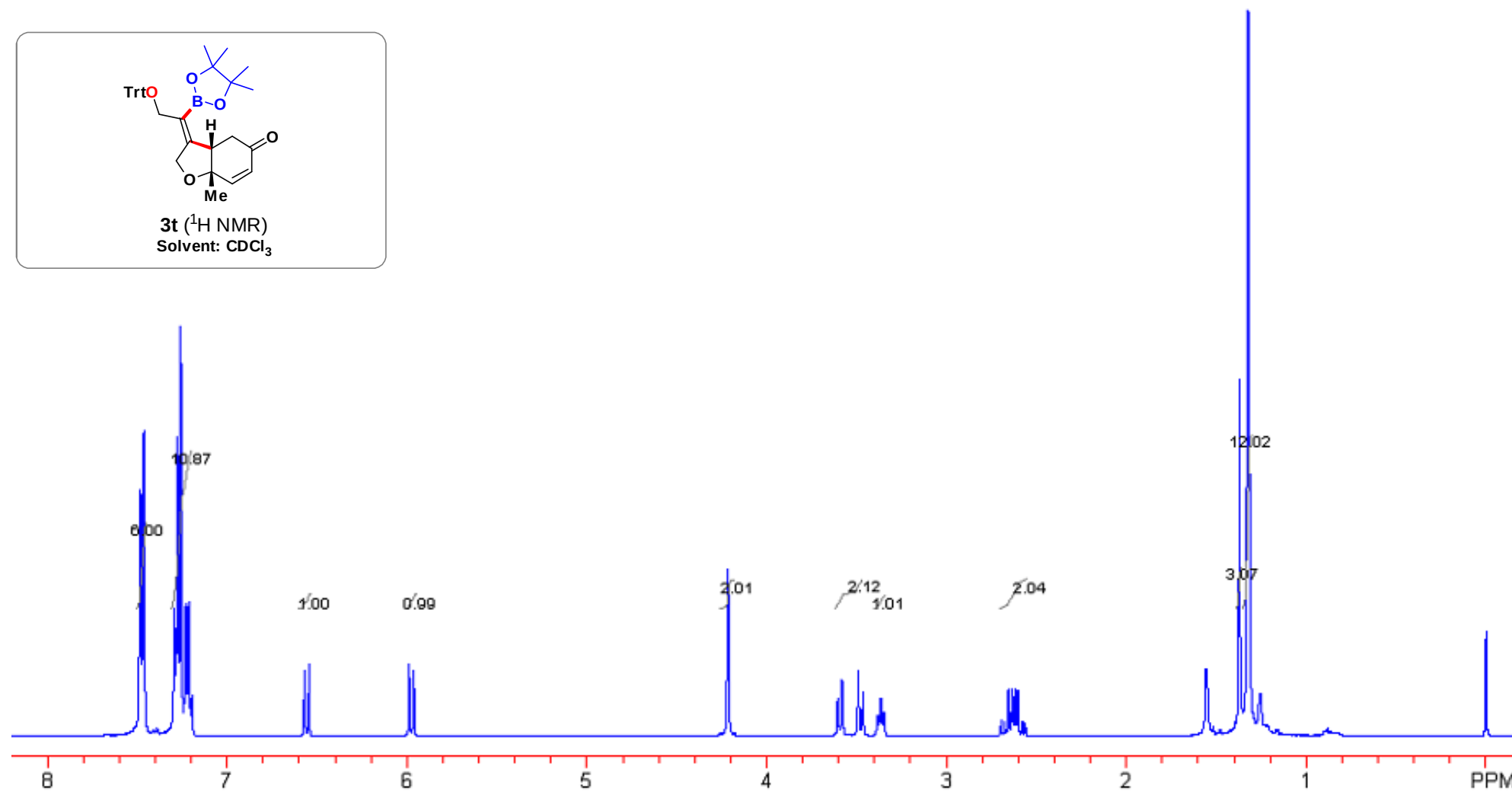
Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3765.1 Hz
2D Width 24118.2 Hz
8 repetitions
2 x 200 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.008 sec
FT size 2048 x 2048
Total time 1 hr, 6 min

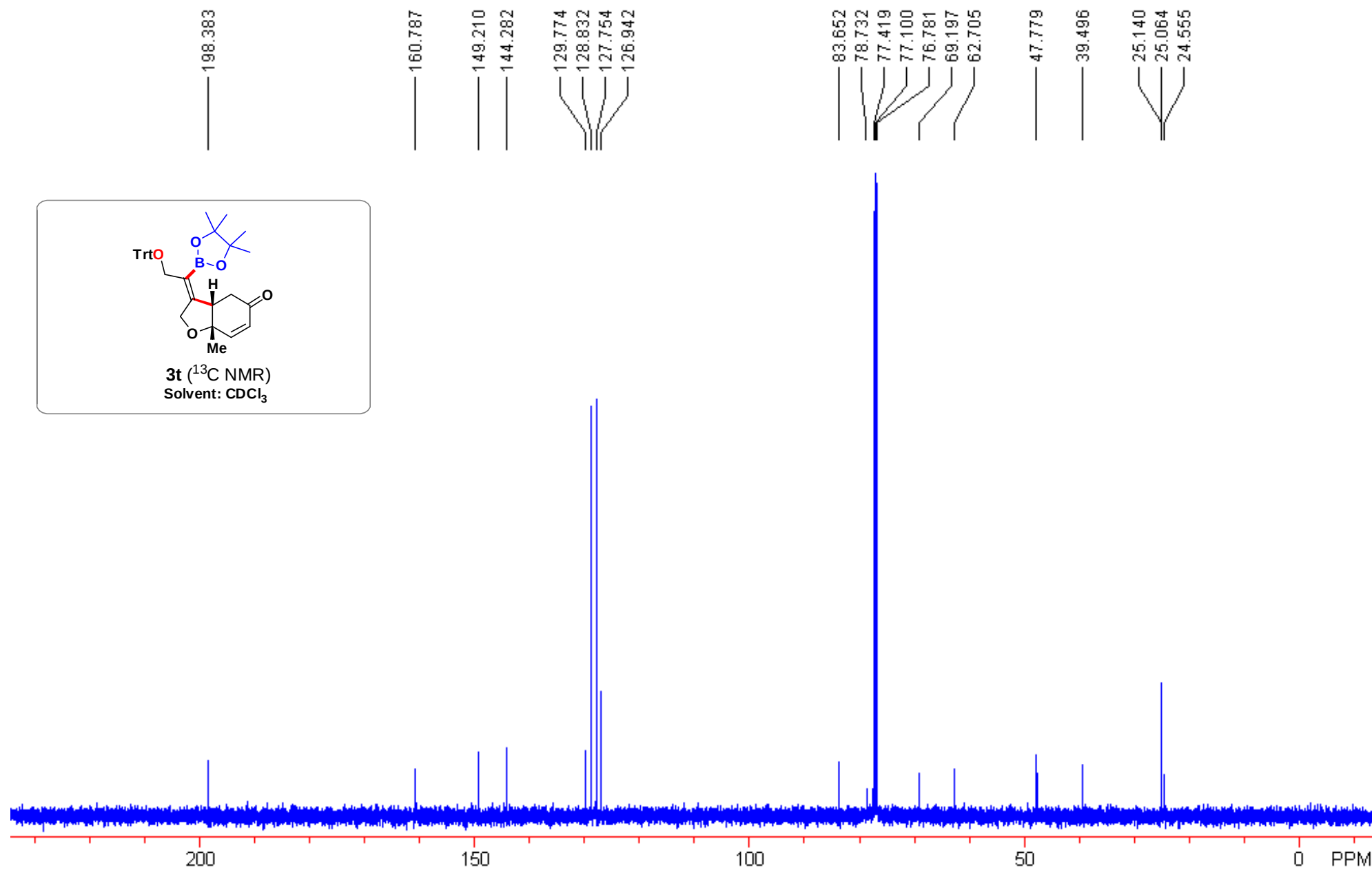


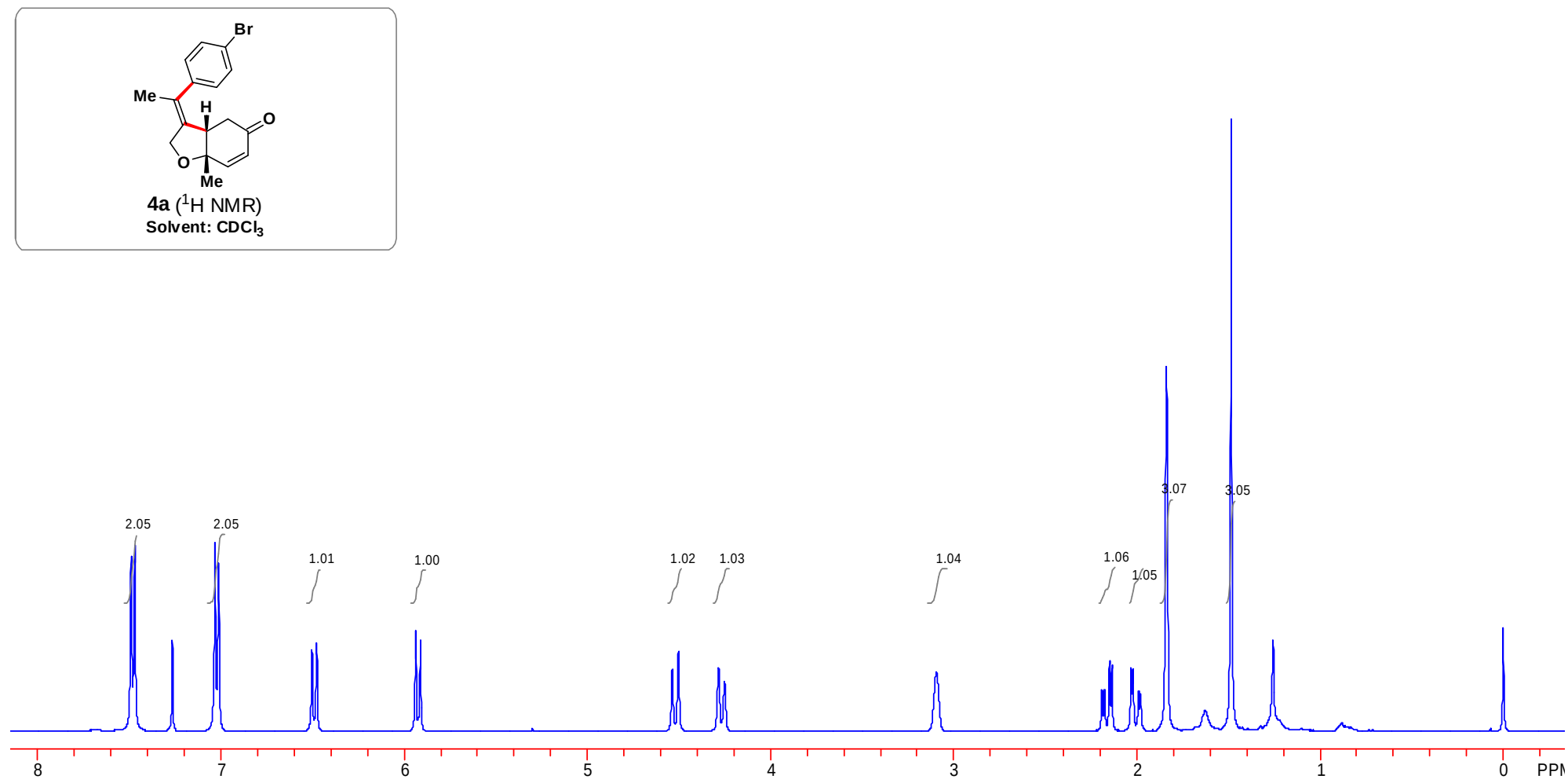
4s (HMBC-3)
Solvent: CDCl₃



3t (^1H NMR)
Solvent: CDCl_3







197.412

150.280

141.170

136.683

131.862

129.933

129.204

129.007

121.218

80.699

77.419

77.100

76.781

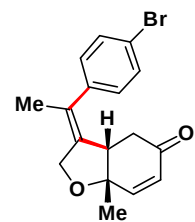
69.728

46.701

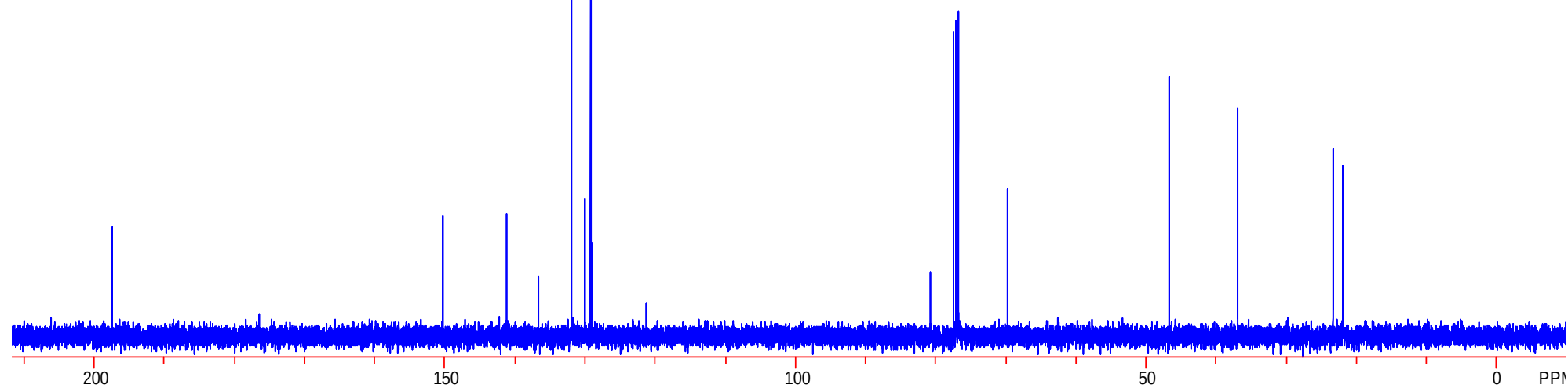
36.908

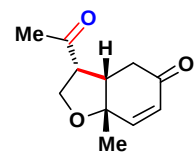
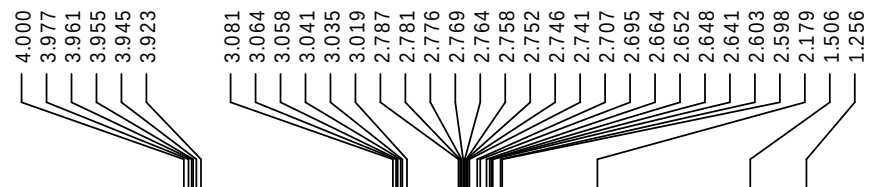
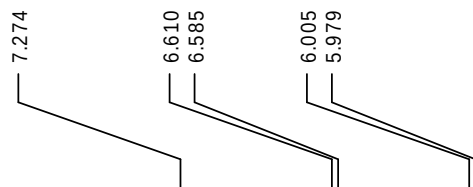
23.325

21.921

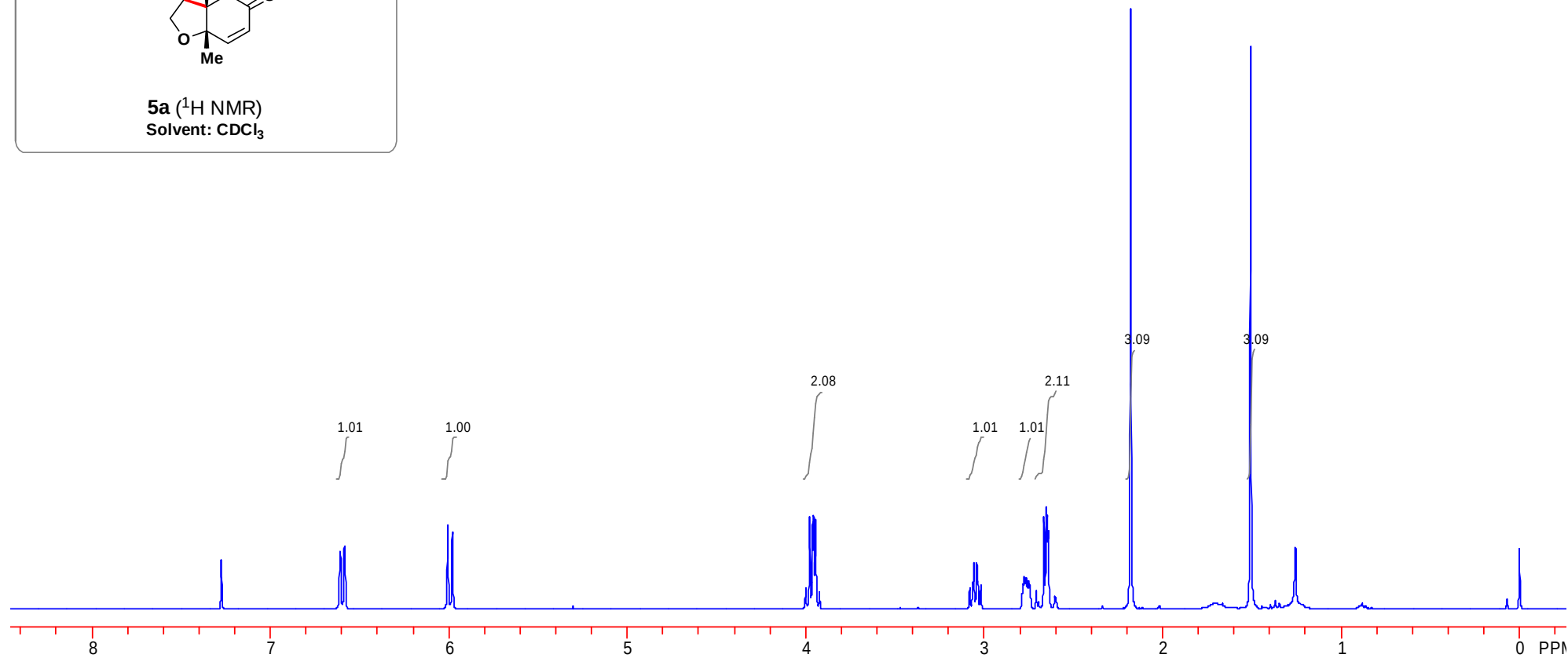


4a (^{13}C NMR)
Solvent: CDCl_3





5a (^1H NMR)
Solvent: CDCl_3



205.907

196.873

152.618

129.068

80.600

77.411

77.100

76.781

67.572

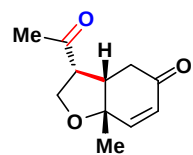
56.973

45.745

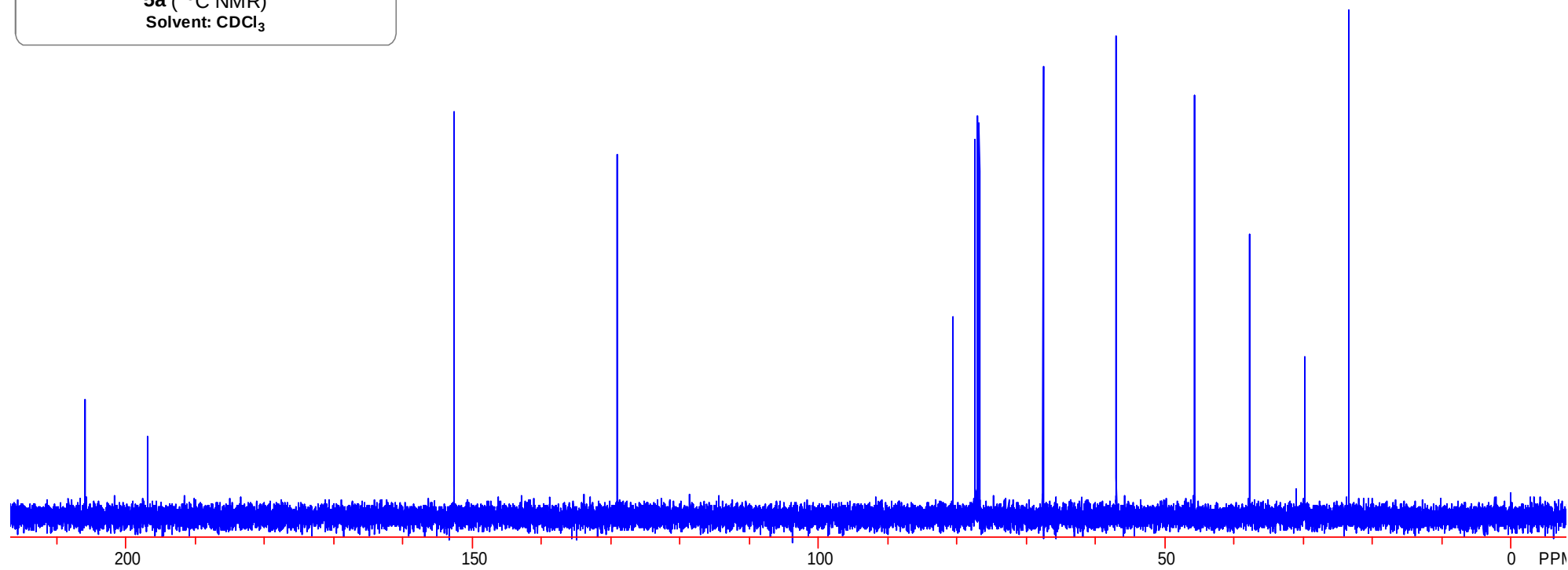
37.758

29.794

23.378



5a (^{13}C NMR)
Solvent: CDCl_3



```
Sample Name:
20120531p3-015-1a
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p3-015-1a_20130326_01
FidFile: gHMBCAD_01
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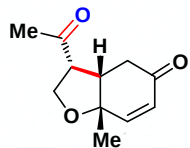
Pulse Sequence: gHMBCAD
Solvent: cdcl3
Data collected on: Mar 26 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

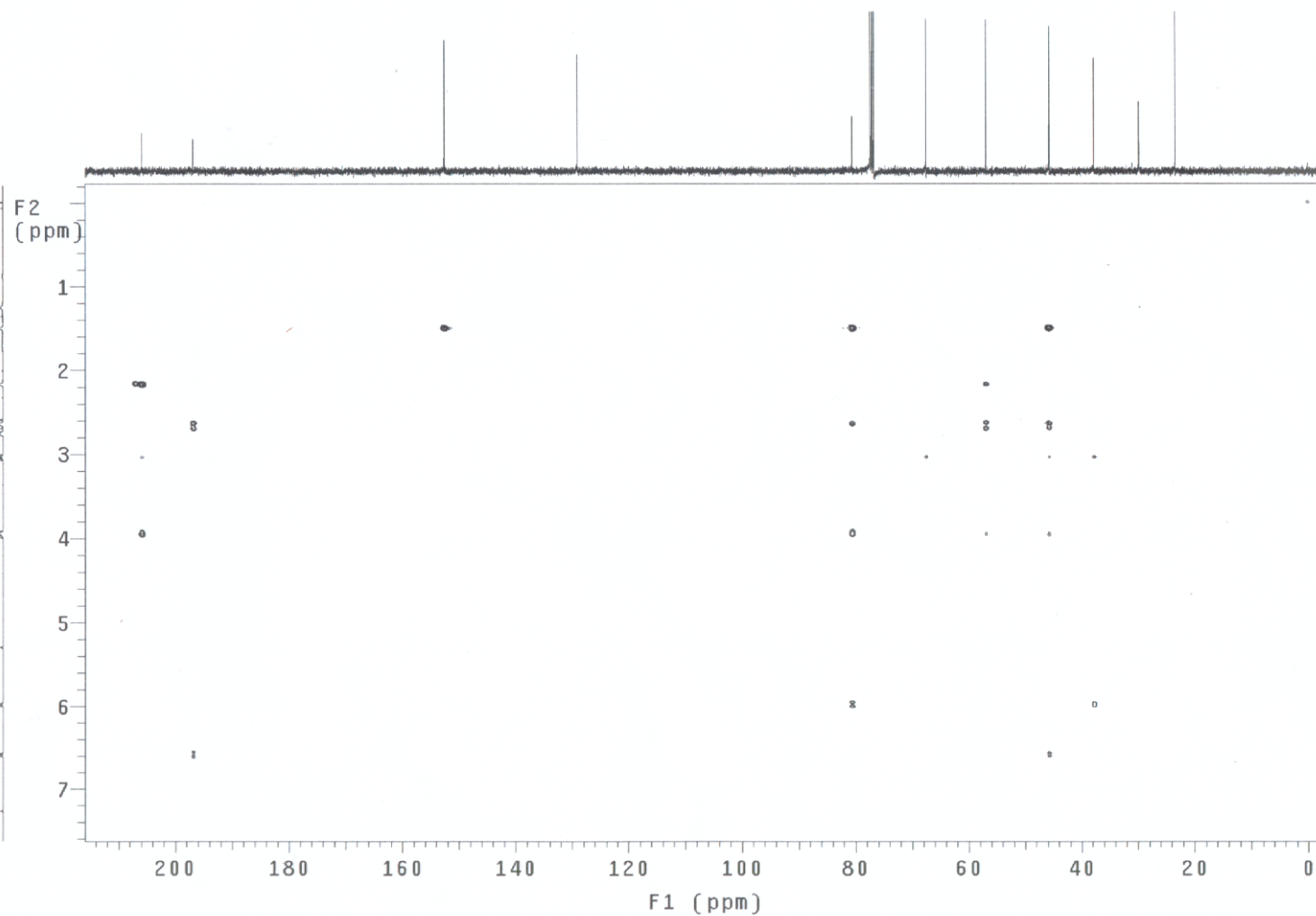
```

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width      3765.1 Hz
2D Width 24118.2 Hz
4 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHZ
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.010 sec
FT size 2048 x 2048
Total time 42 min

```



5a (HMBC)
Solvent: CDCl₃



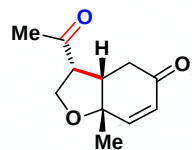
20120531p3-015-1a

Sample Name:
20120531p3-015-1a
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p3-015-1a_20130326_01
FidFile: gHMBCAD_01

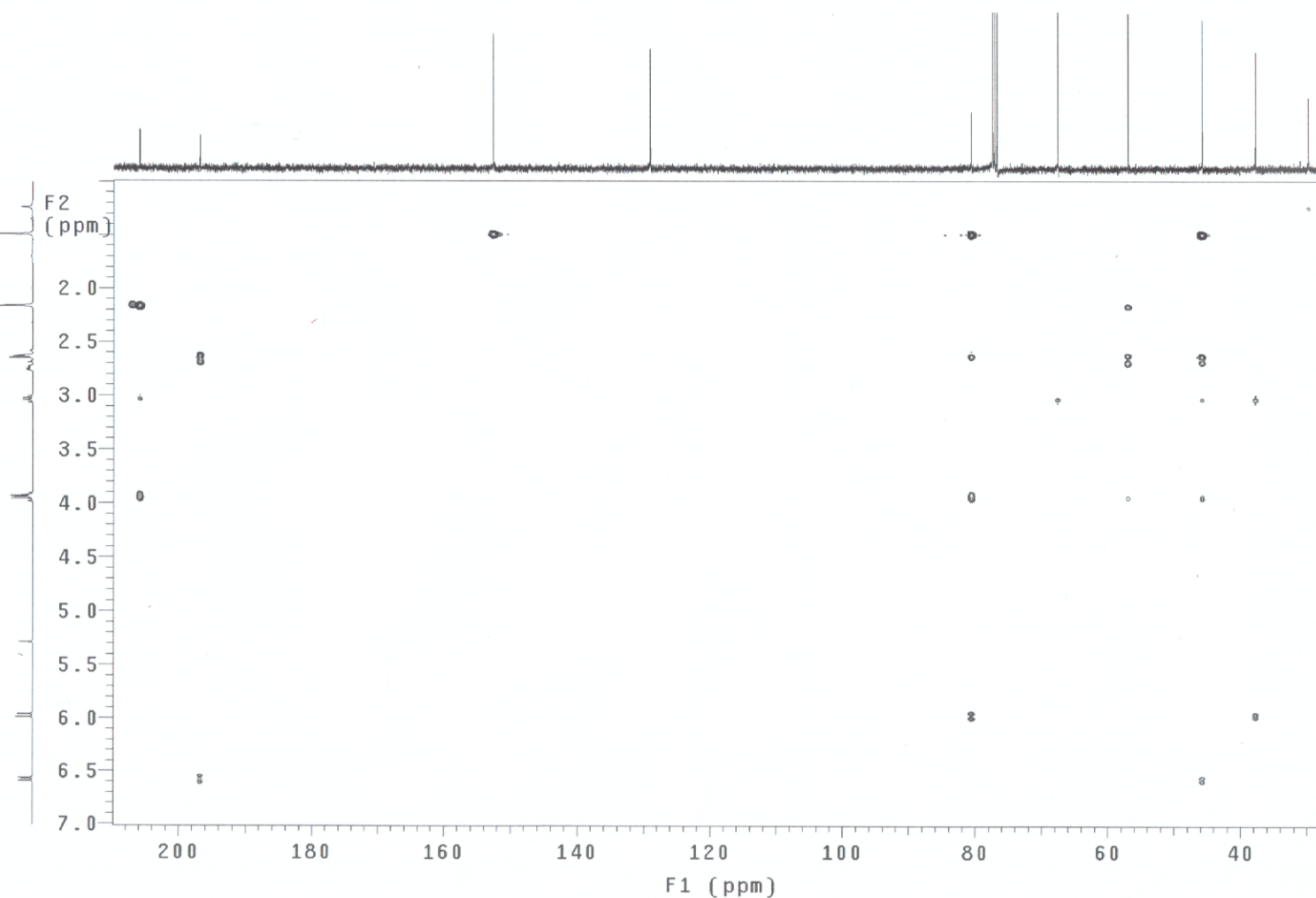
Pulse Sequence: gHMBCAD
Solvent: cdcl3
Data collected on: Mar 26 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3765.1 Hz
2D Width 24118.2 Hz
4 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.010 sec
FT size 2048 x 2048
Total time 42 min



5a (HMBC)
Solvent: CDCl₃



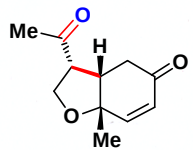
20120531p3-015-1a

Sample Name:
20120531p3-015-1a
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p3-015-1a_20130326_01
FidFile: gHSQCAD_01

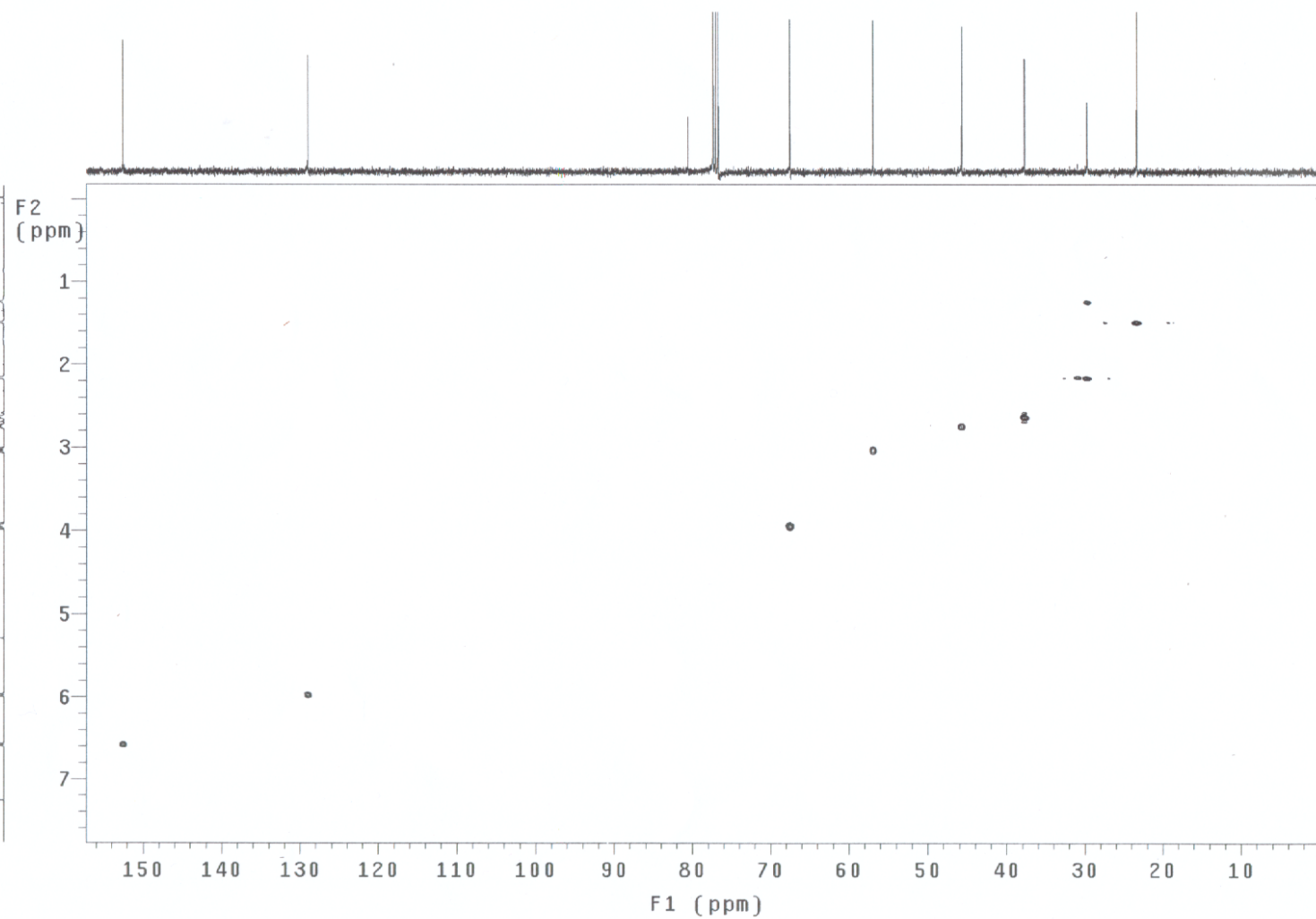
Pulse Sequence: gHSQCAD
Solvent: cdcl3
Data collected on: Mar 26 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3765.1 Hz
2D Width 20100.5 Hz
4 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DECOUPLE C13, 100.5036547 MHz
Power 37 dB
on during acquisition
off during delay
W40_ATB3 modulated
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.012 sec
FT size 2048 x 2048
Total time 41 min



5a (HSQC)
Solvent: CDCl₃



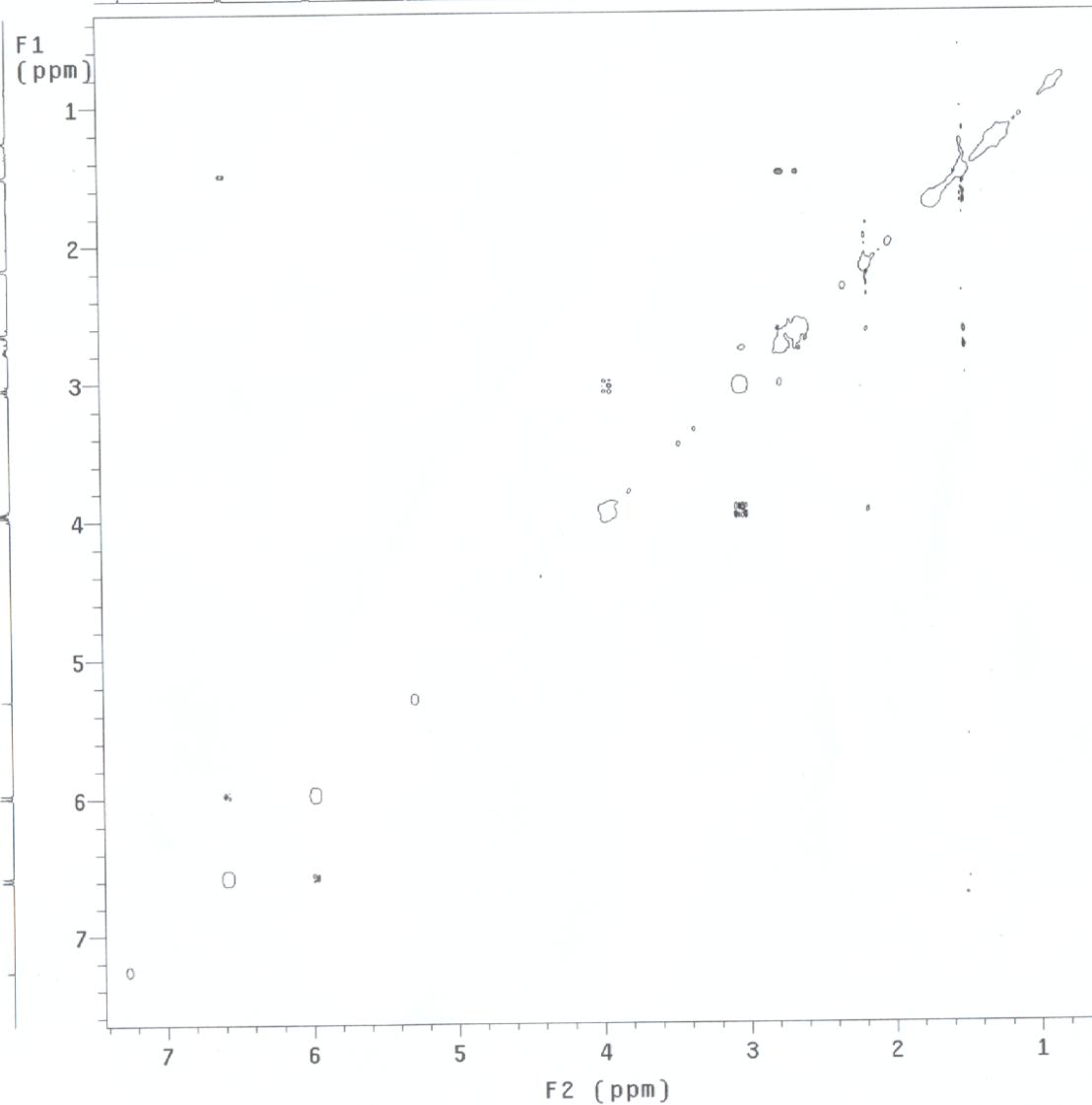
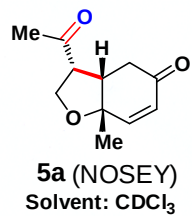
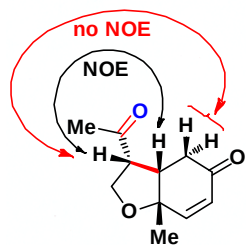
20120531p3-015-1a

Sample Name:
20120531p3-015-1a
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p3-015-1a_20130322_01
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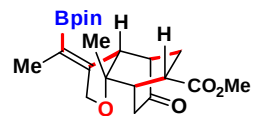
Pulse Sequence: NOESY
Solvent: cdc13
Data collected on: Mar 22 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

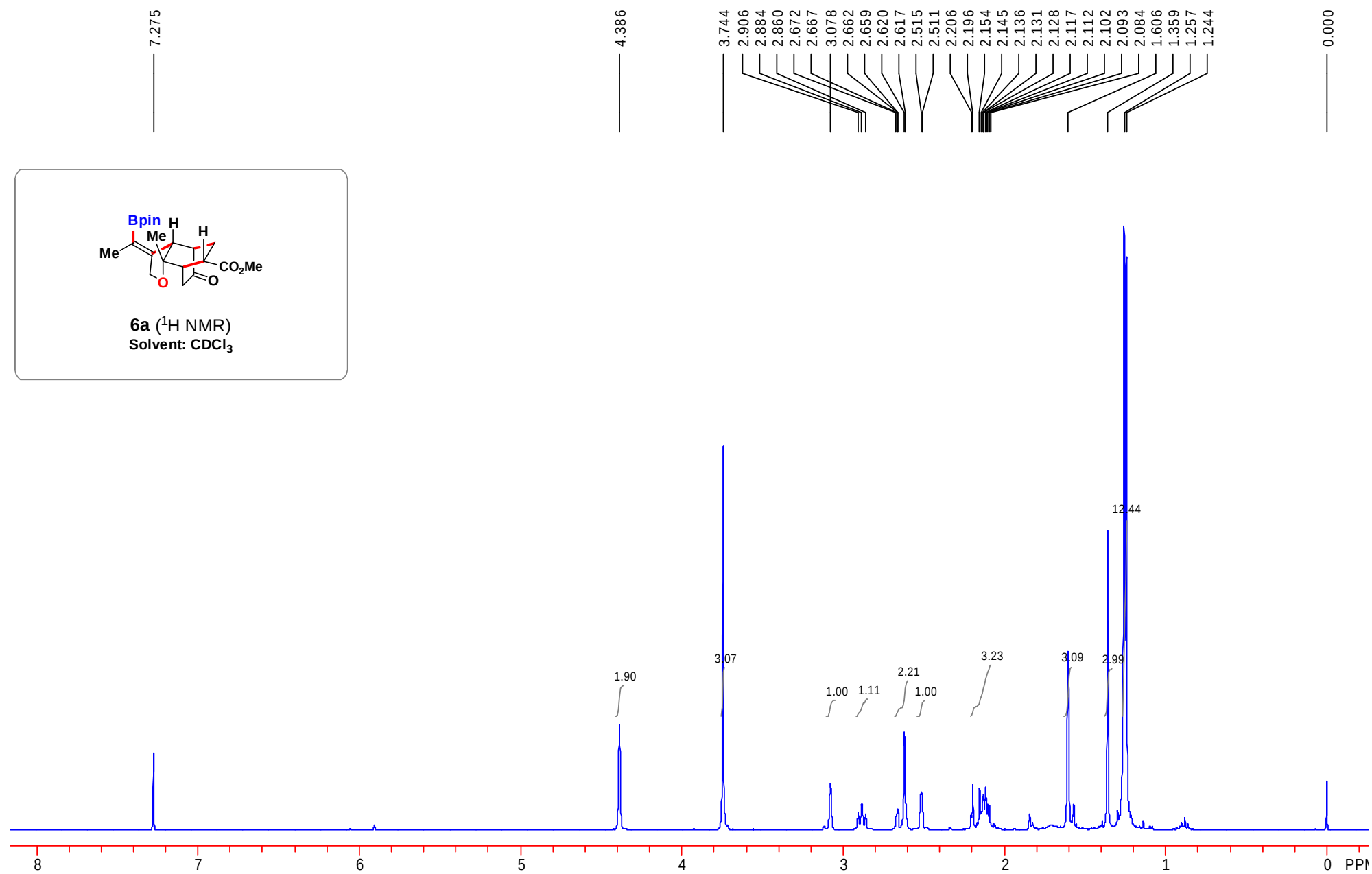
Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3720.2 Hz
2D Width 3720.2 Hz
8 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.037 sec
FT size 2048 x 2048
Total time 2 hr, 9 min

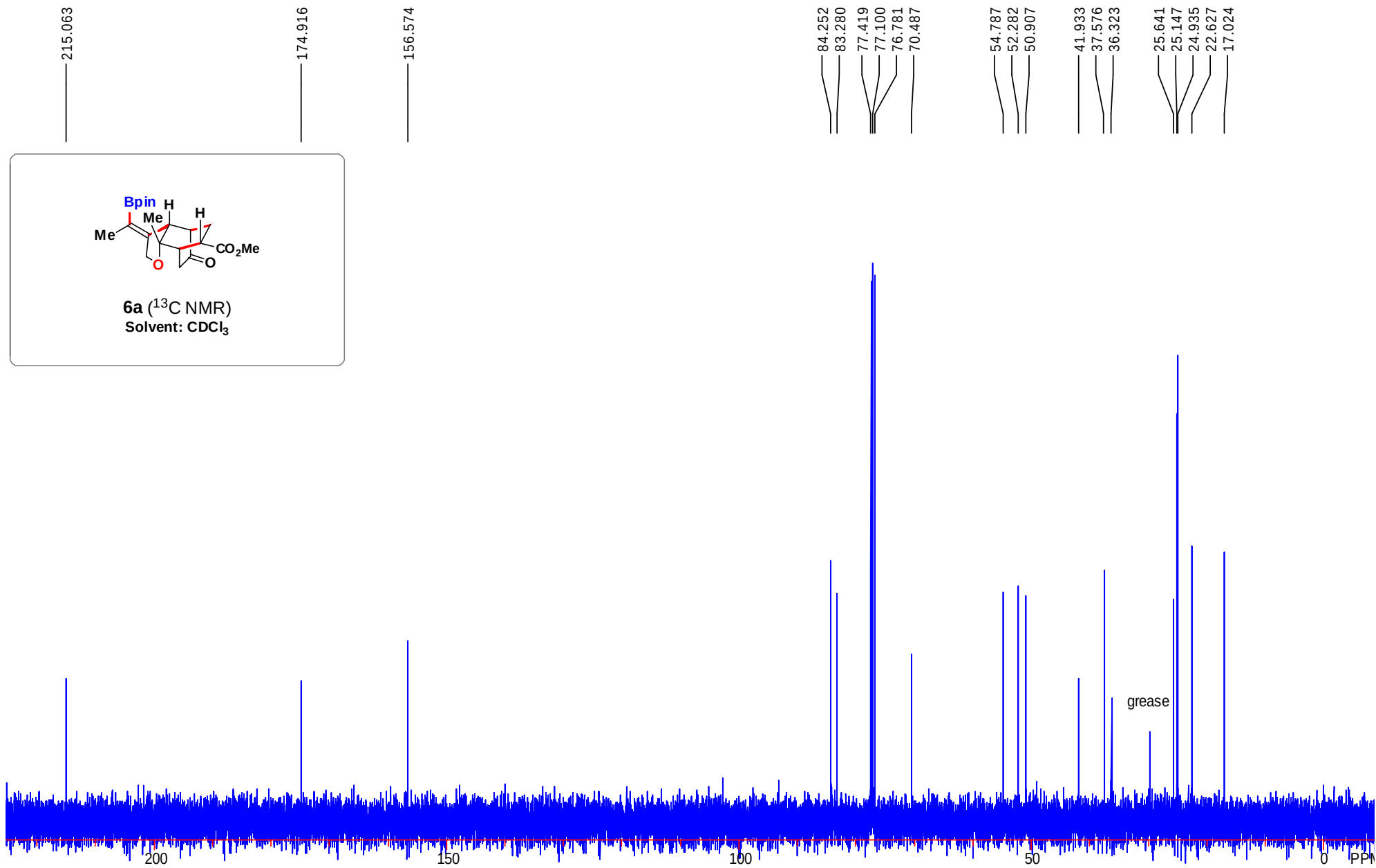


5153



6a (¹H NMR)
Solvent: CDCl₃





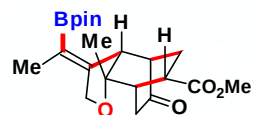
20120531p3-028-1

Sample Name:
20120531p3-028-1
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p3-028-1_20130401_01
FidFile: gHSQCAD_01

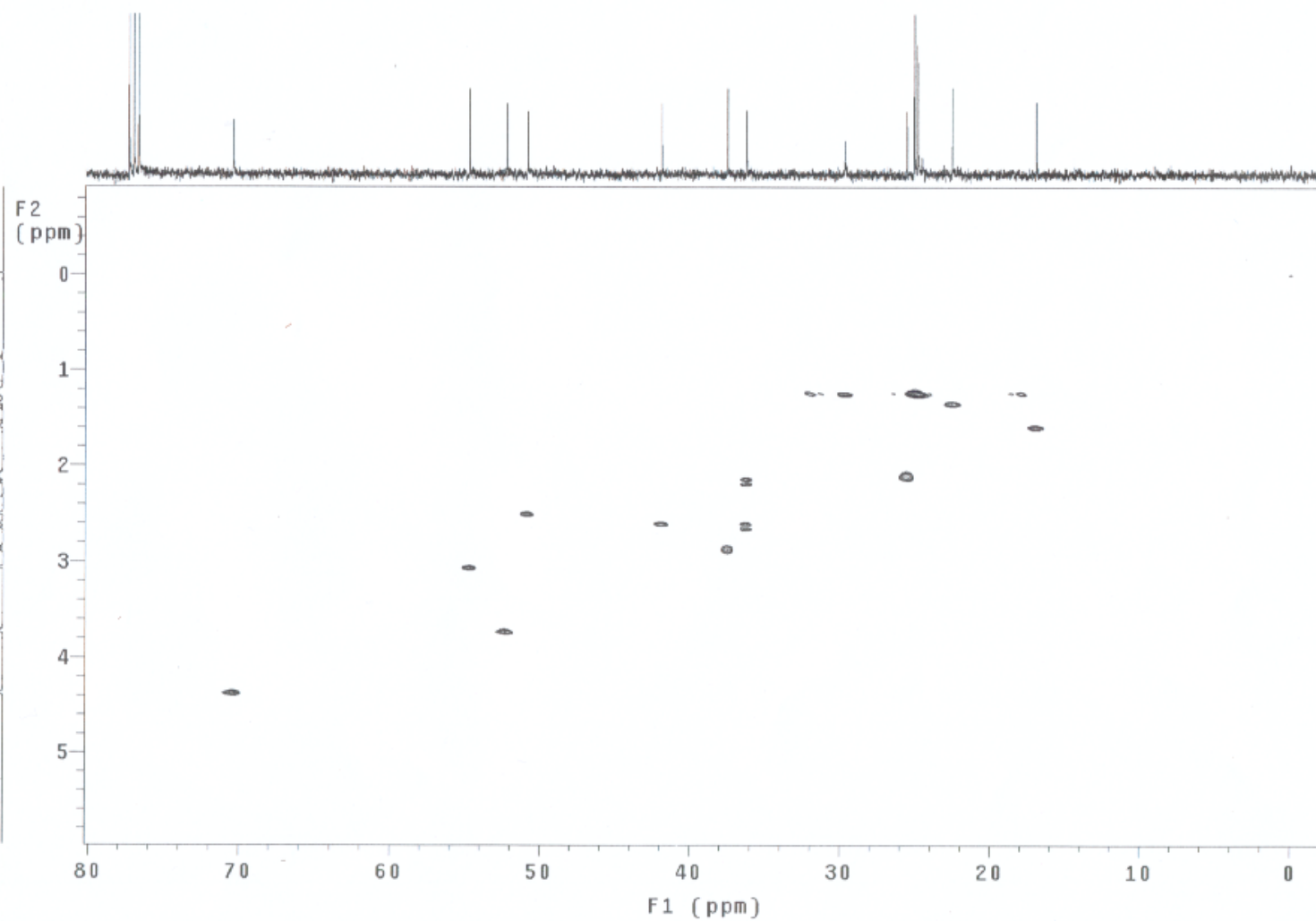
Pulse Sequence: gHSQCAD
Solvent: cdcl3
Data collected on: Apr 1 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4807.7 Hz
2D Width 20100.5 Hz
2 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DECOUPLE C13, 100.5036547 MHz
Power 37 dB
on during acquisition
off during delay
W40_ATB3 modulated
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.012 sec
FT size 2048 x 2048
Total time 21 min



6a (HSQC)
Solvent: CDCl₃



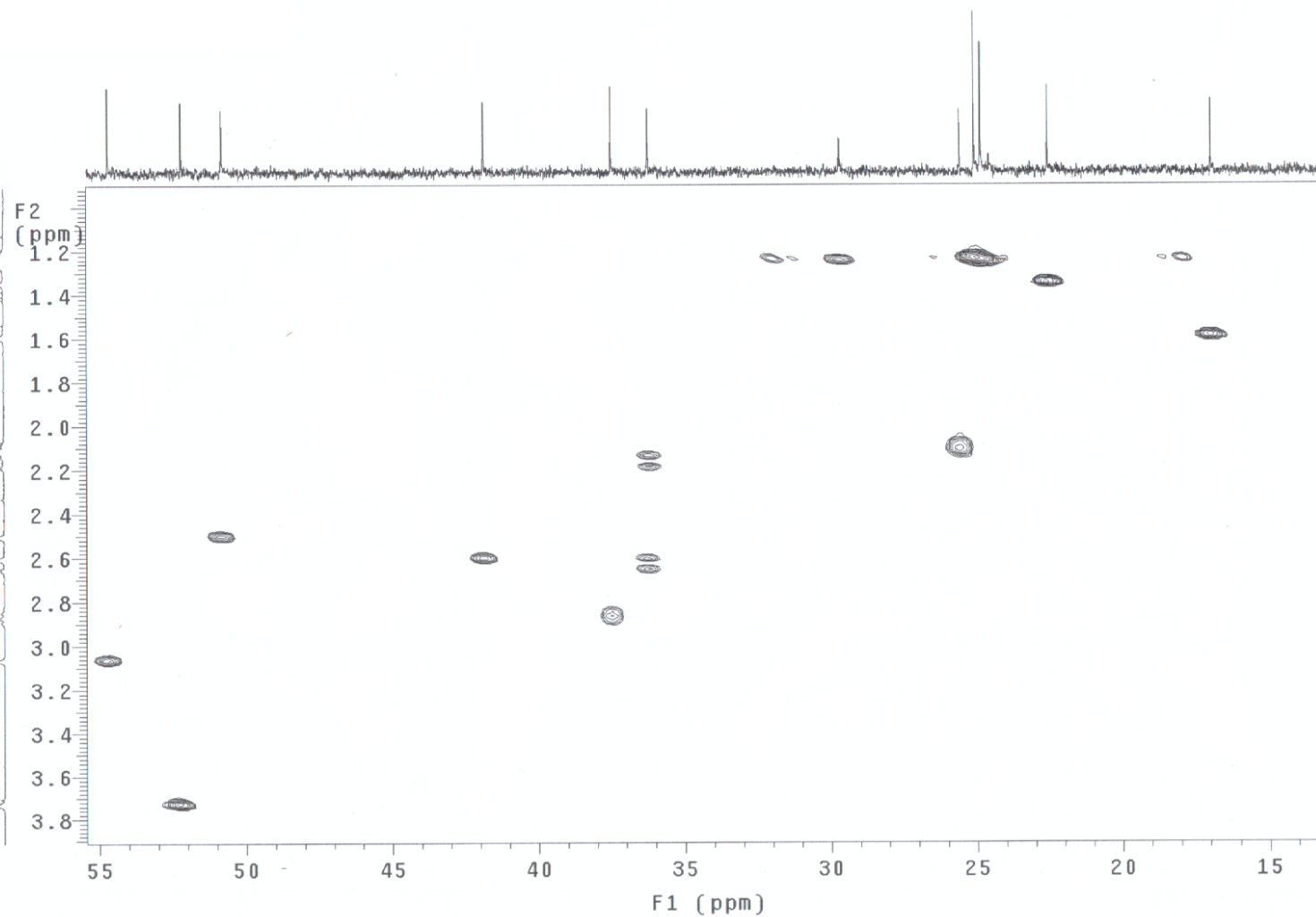
20120531p3-028-1

Sample Name:
20120531p3-028-1
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p3-028-1_20130401_01
FidFile: gHSQCAD_01

Pulse Sequence: gHSQCAD
Solvent: cdc13
Data collected on: Apr 1 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4807.7 Hz
2D Width 20100.5 Hz
2 repetitions
2 x 256 increments
OBSERVE H1, 399.660928 MHz
DECOUPLE C13, 100.5036547 MHz
Power 37 dB
on during acquisition
off during delay
W40_ATB3 modulated
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.012 sec
FT size 2048 x 2048
Total time 21 min



6a (HSQC)
Solvent: CDCl₃

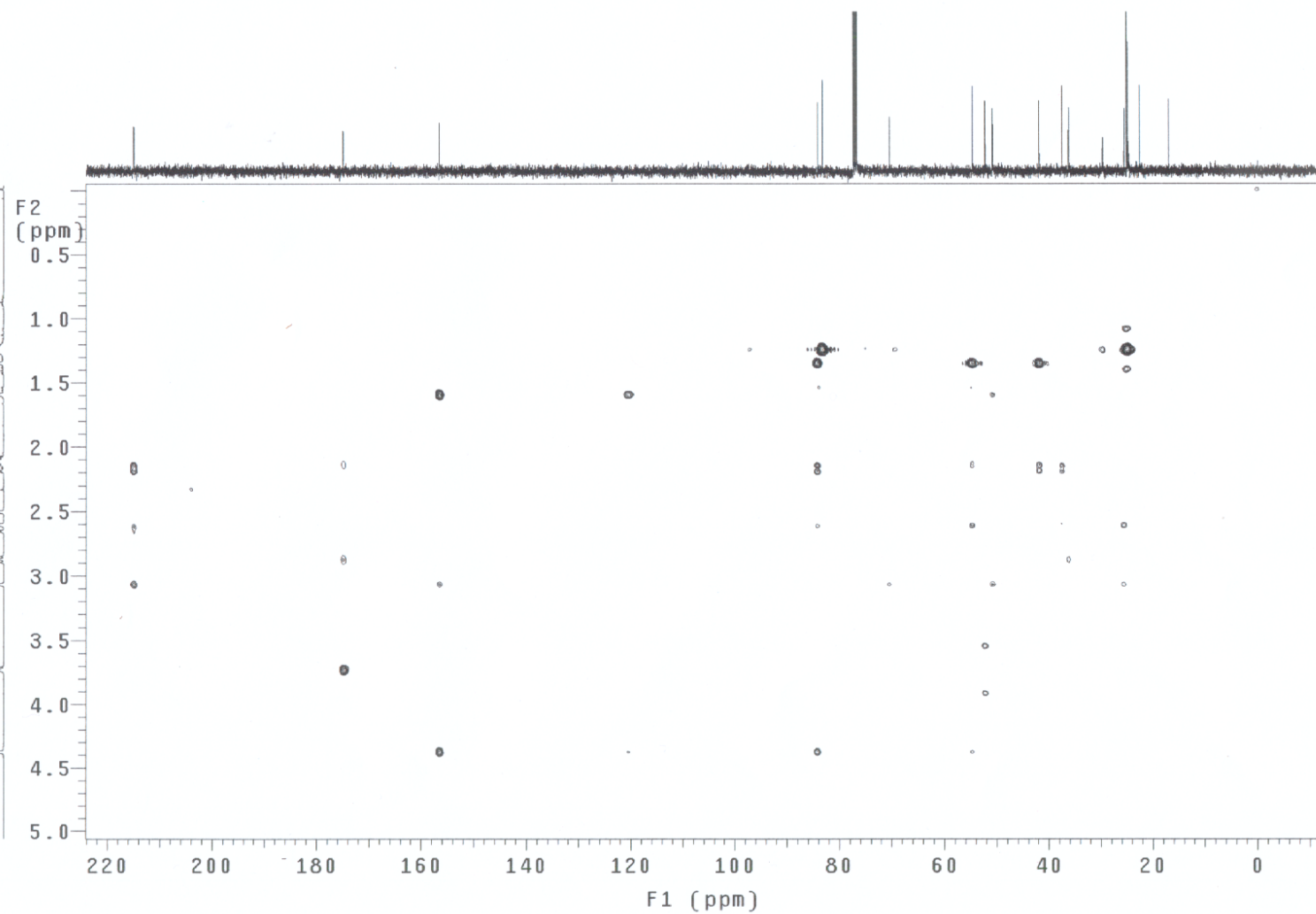
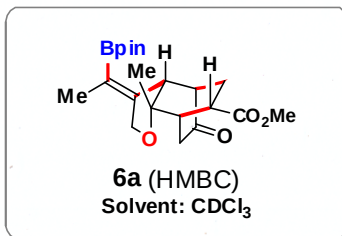
20120531p3-028-1

Sample Name:
20120531p3-028-1
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p3-028-1_20130401_01
FidFile: gHMBCAD_01

Pulse Sequence: gHMBCAD
Solvent: cdc13
Data collected on: Apr 1 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4807.7 Hz
2D Width 24118.2 Hz
4 repetitions
2 x 200 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.008 sec
FT size 2048 x 2048
Total time 33 min



20120531p3-028-1

Sample Name:
20120531p3-028-1
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p3-028-1_20130401_01
FidFile: gHMBCAD_01

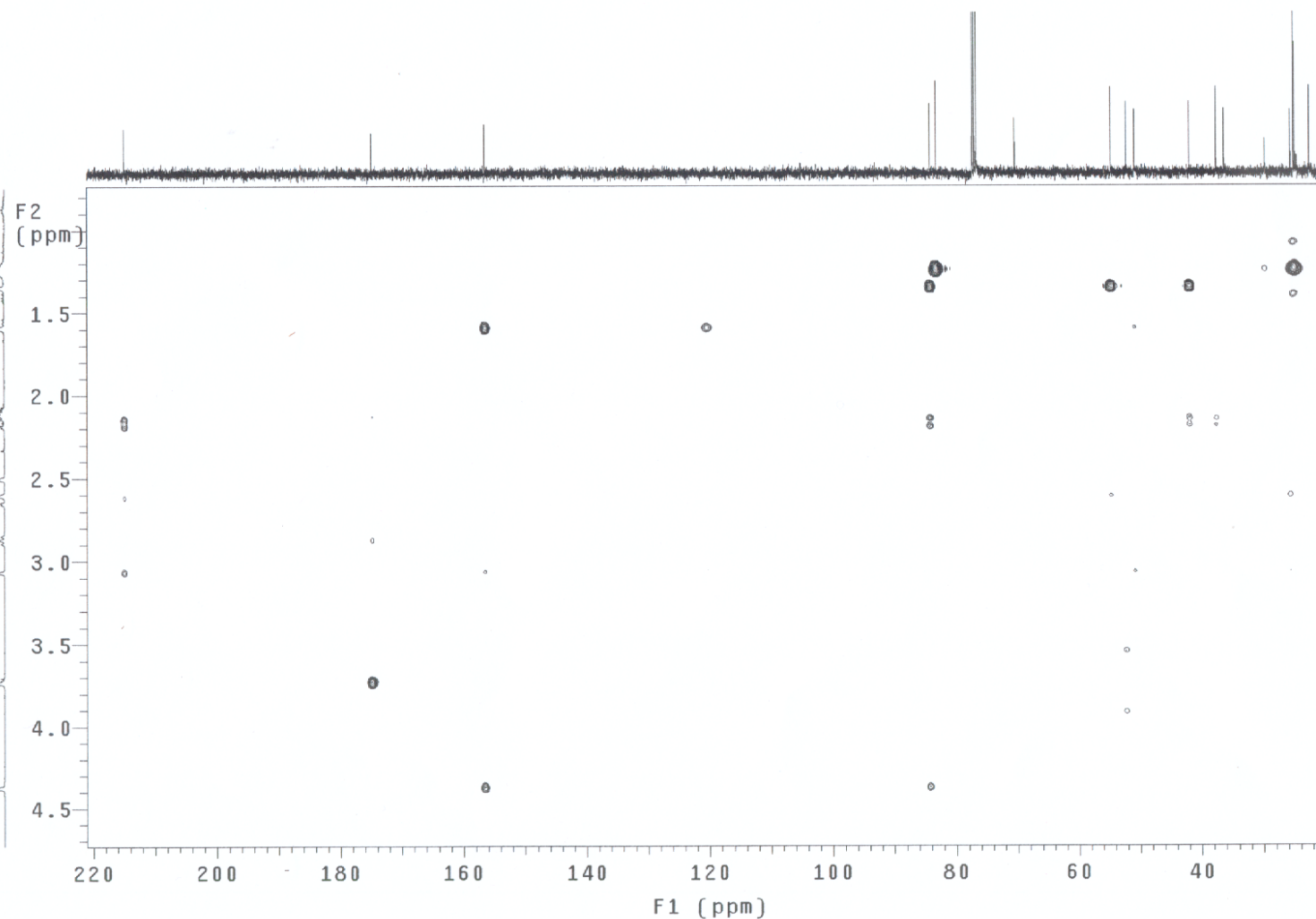
Pulse Sequence: gHMBCAD
Solvent: cdc13
Data collected on: Apr 1 2013

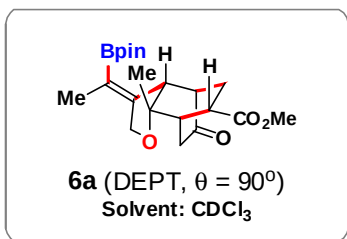
Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4807.7 Hz
2D Width 24118.2 Hz
4 repetitions
2 x 200 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.008 sec
FT size 2048 x 2048
Total time 33 min

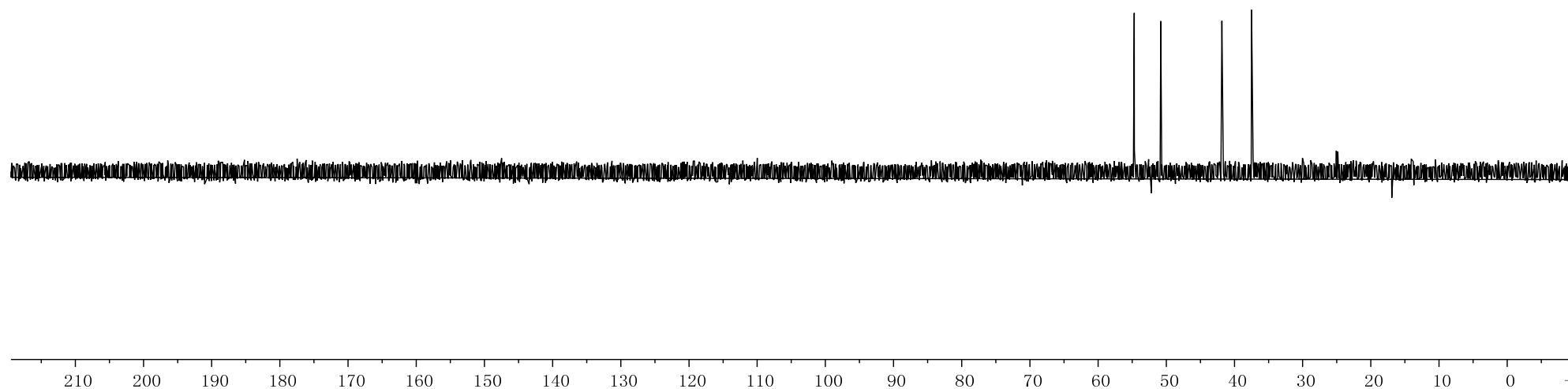


6a (HMBC)
Solvent: CDCl₃





— 54.695
— 50.813
— 41.846
— 37.485



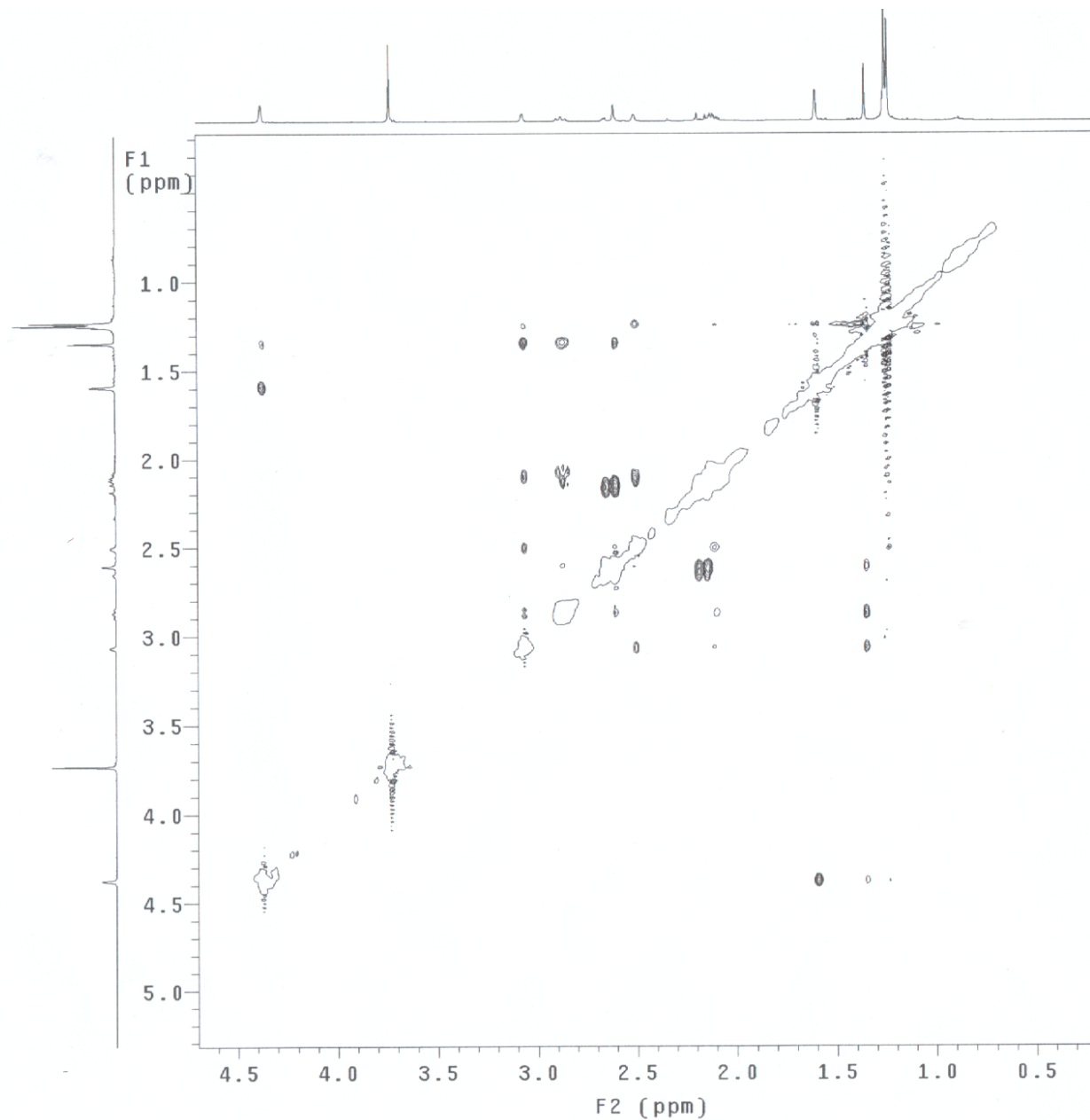
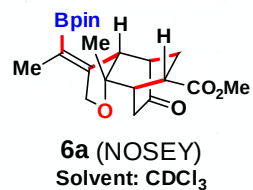
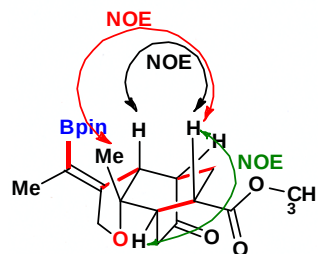
20120531p3-028-1

Sample Name:
20120531p3-028-1
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p3-028-1_20130401_0-
FidFile: NOESY_01

Pulse Sequence: NOESY
Solvent: cdcl3
Data collected on: Apr 1 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4807.7 Hz
2D Width 4807.7 Hz
8 repetitions
2 x 200 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.038 sec
FT size 2048 x 2048
Total time 1 hr, 41 min



20120531p3-028-1

Sample Name:
20120531p3-028-1
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
20120531p3-028-1_20130401_01
FidFile: NOESY_01

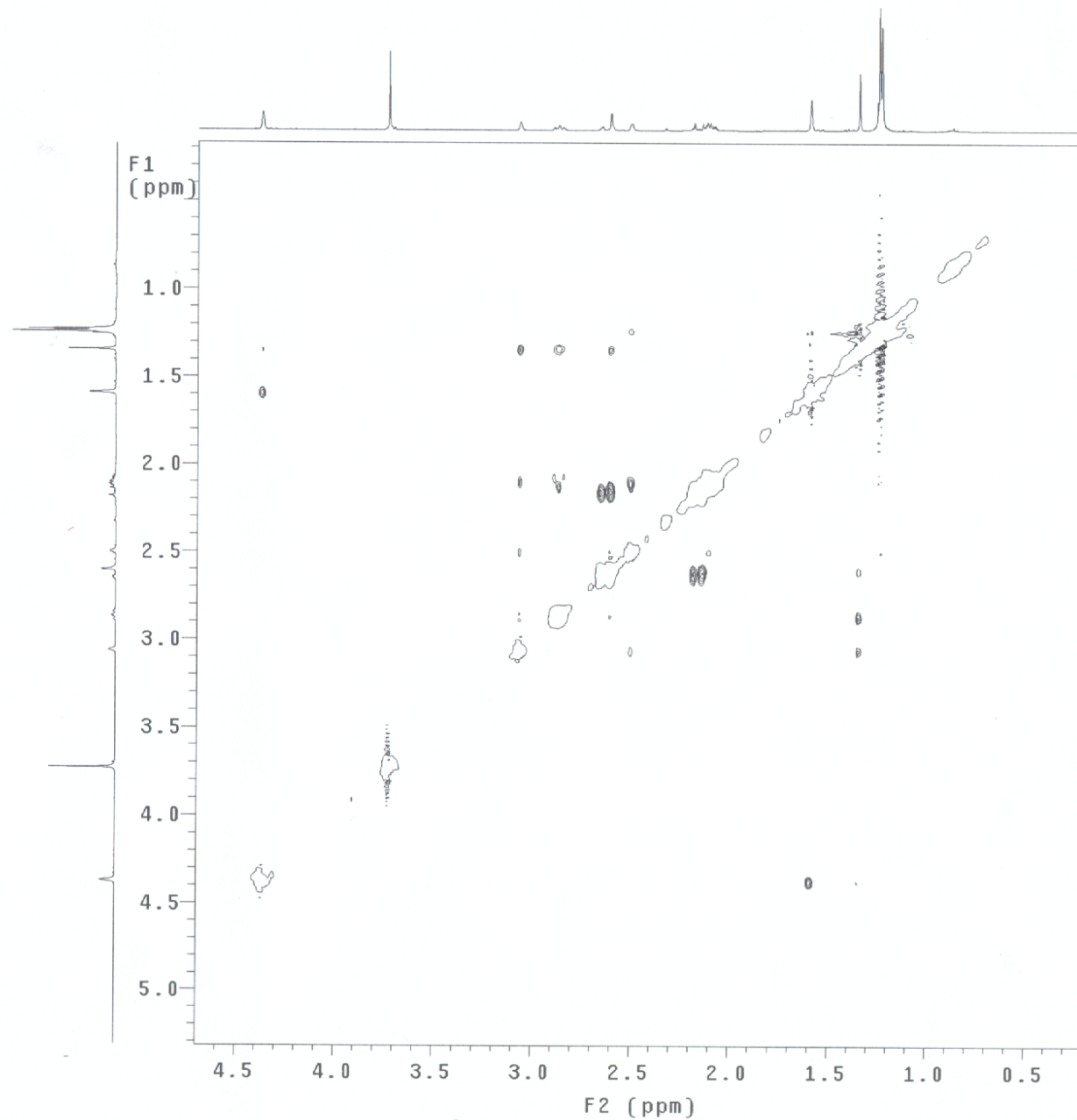
Pulse Sequence: NOESY
Solvent: cdc13
Data collected on: Apr 1 2013

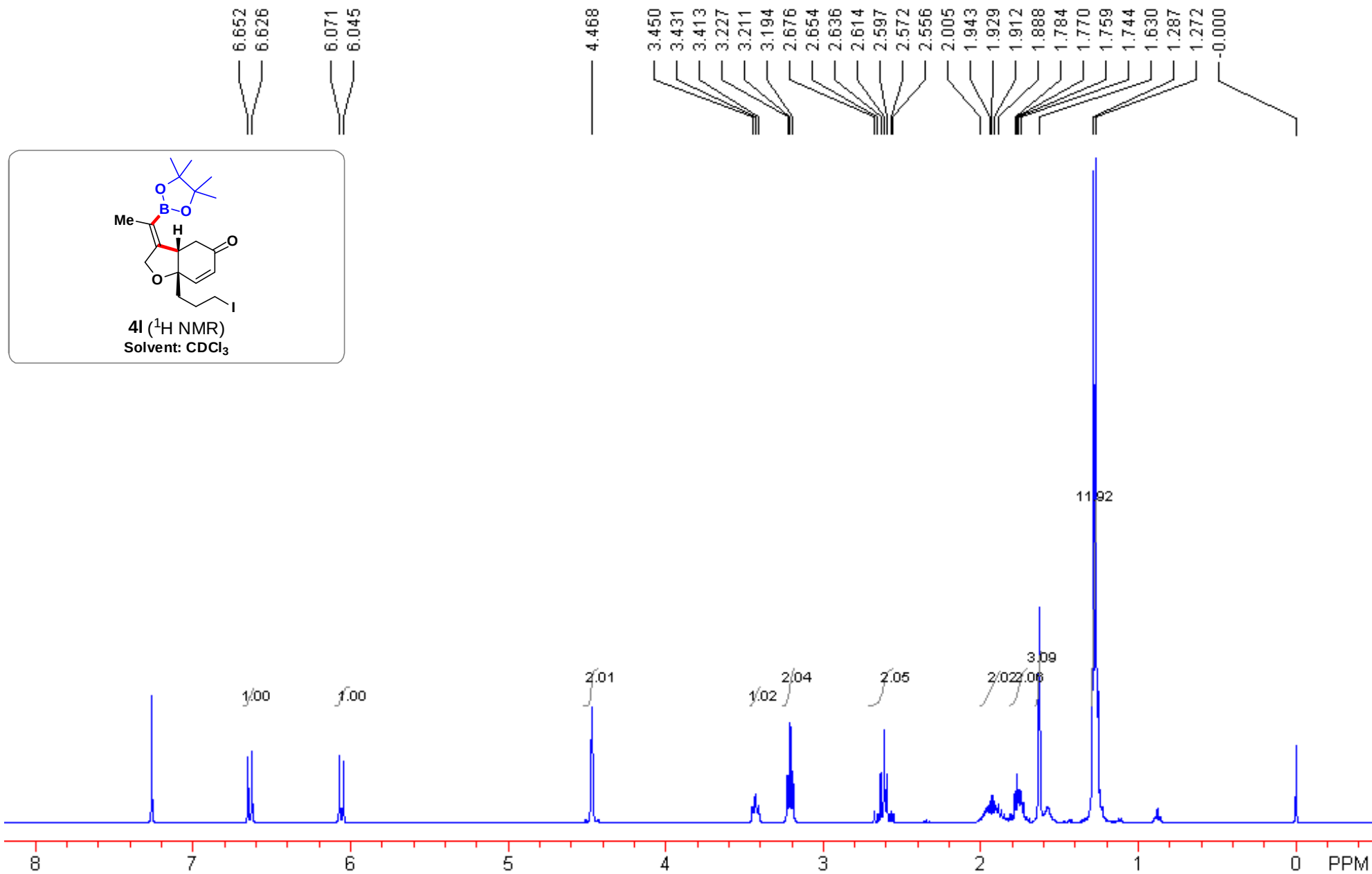
Temp. 25.0 C / 298.1 K
Operator: sioc

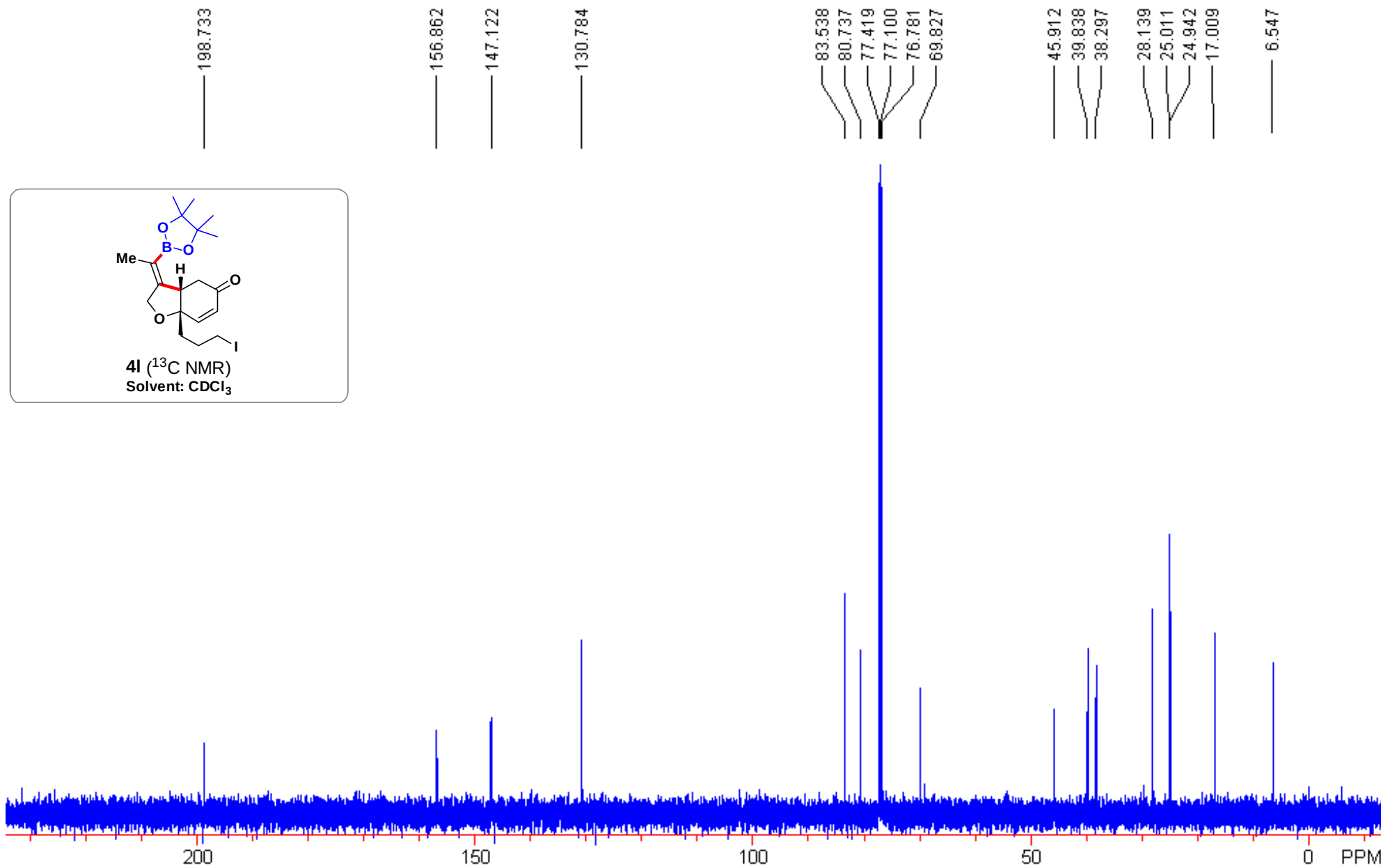
Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4807.7 Hz
2D Width 4807.7 Hz
8 repetitions
2 x 200 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.038 sec
FT size 2048 x 2048
Total time 1 hr, 41 min

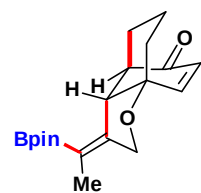


6a (NOSEY)
Solvent: CDCl₃

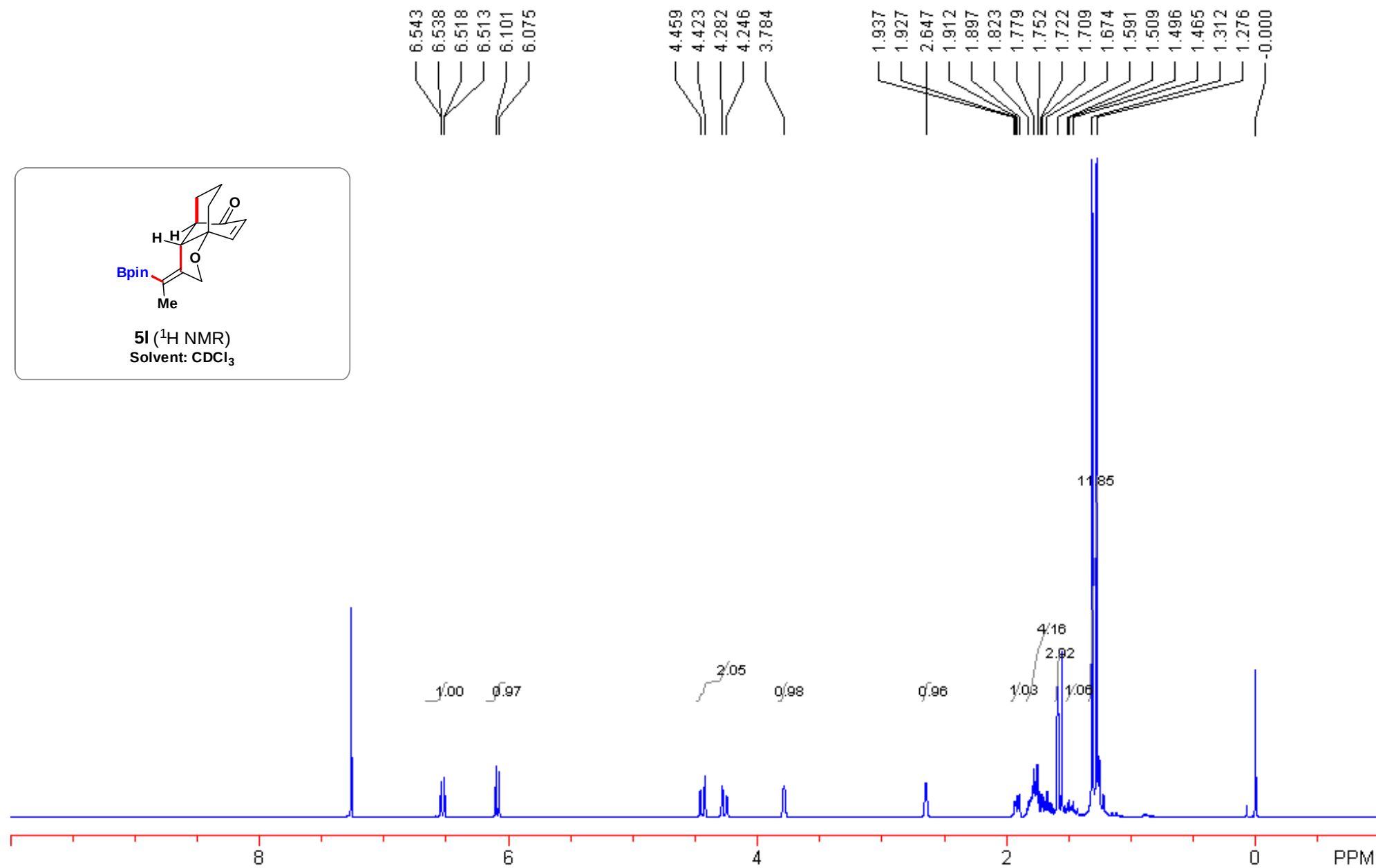


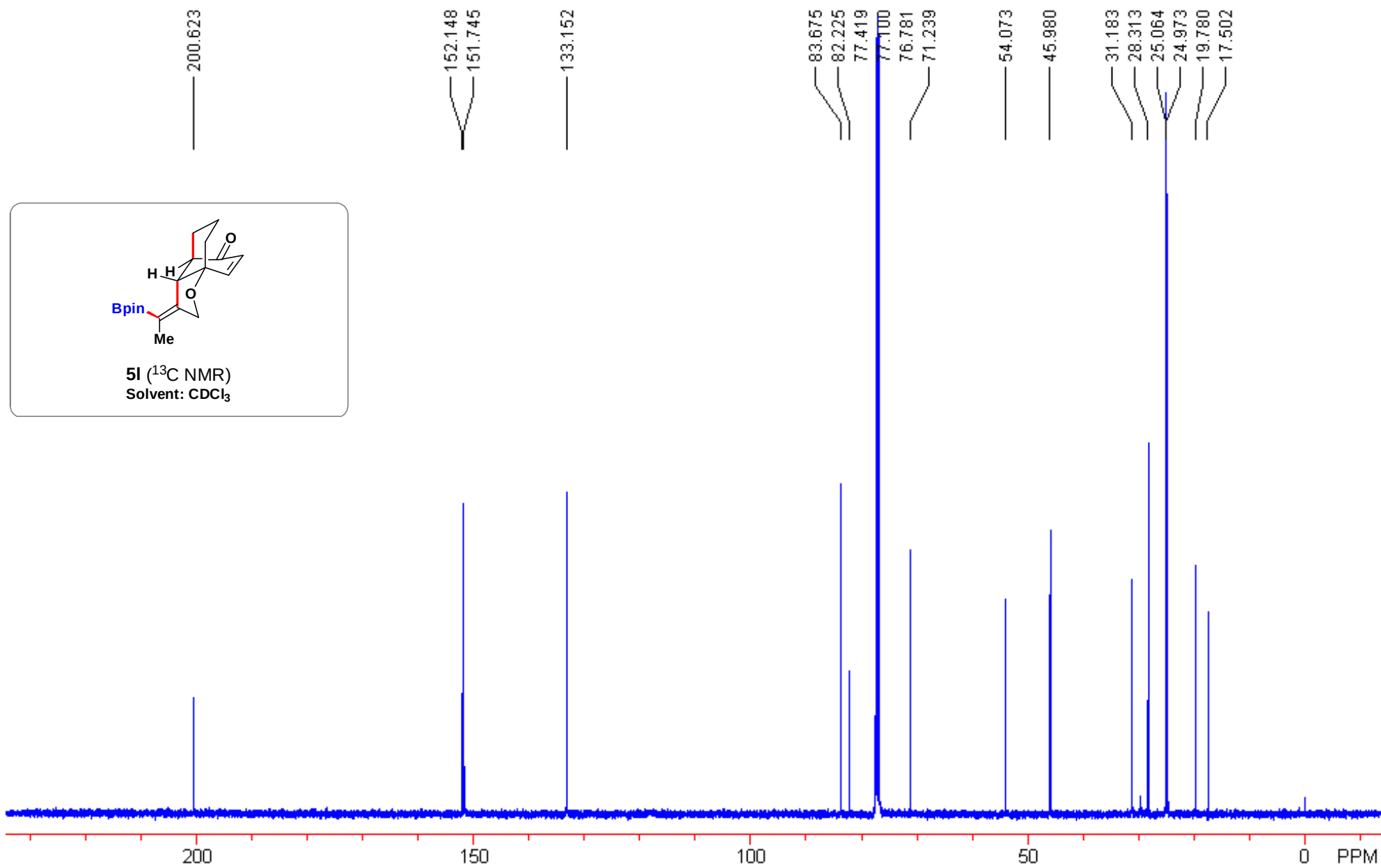


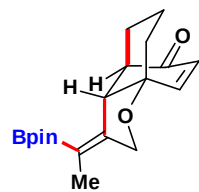




5I (^1H NMR)
Solvent: CDCl_3







5l (DEPT, $\theta = 135^\circ$)
Solvent: CDCl_3

151.760

133.152

71.246

54.073

45.987

31.183

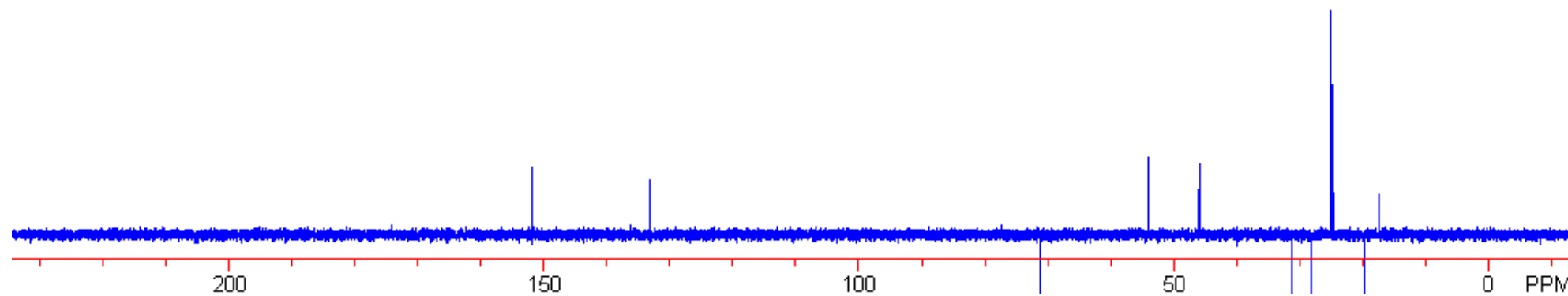
28.321

25.071

24.973

19.780

17.509



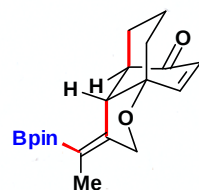
2012576yf02-16-01

Sample Name:
2012576yf02-16-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-16-01_20130408_01
FidFile: gCOSY_01

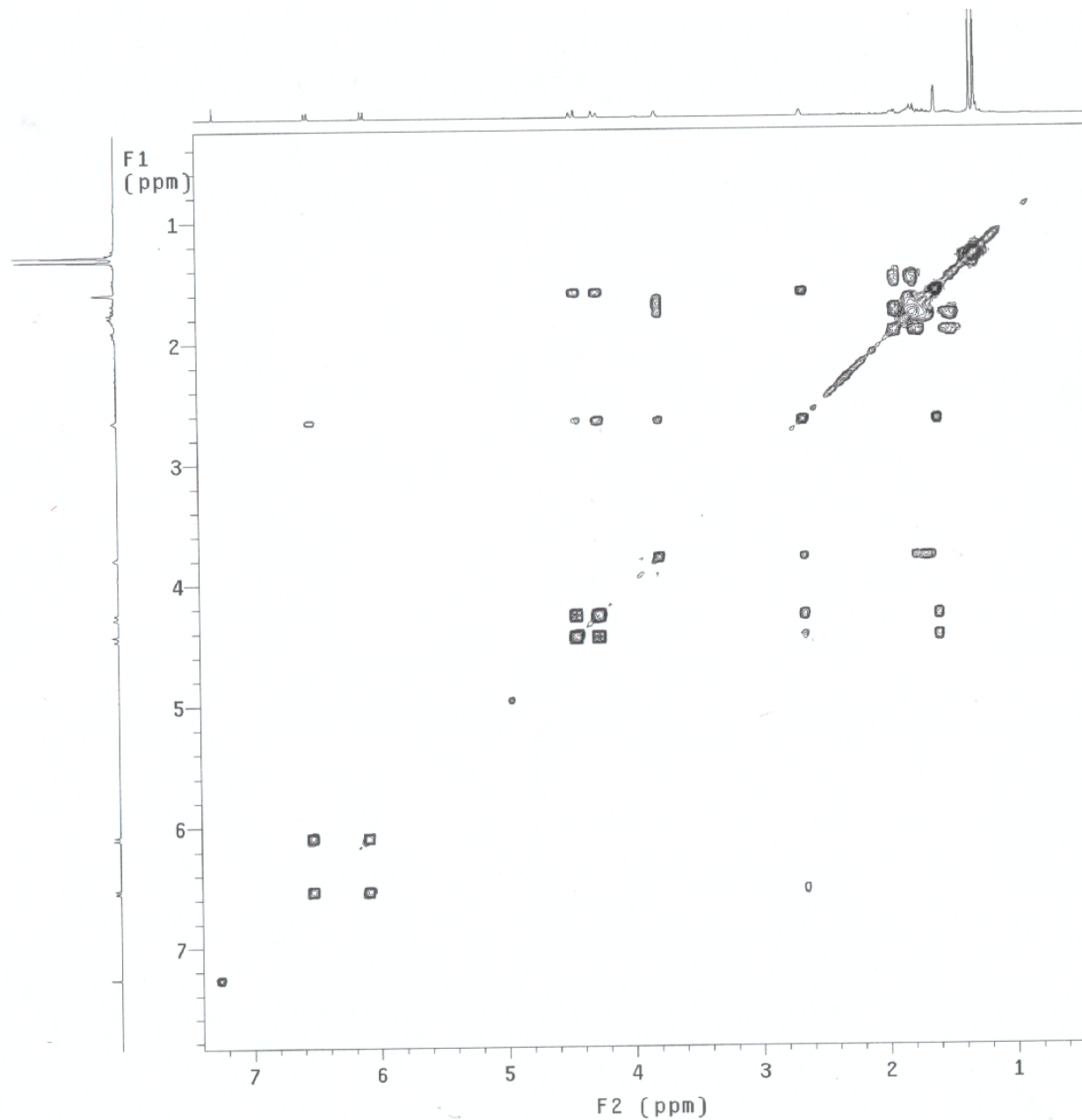
Pulse Sequence: gCOSY
Solvent: cdcl3
Data collected on: Apr 8 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4058.4 Hz
2D Width 4058.4 Hz
8 repetitions
256 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Sq. sine bell 0.040 sec
FT size 2048 x 2048
Total time 41 min



5I (H-H COSY)
Solvent: CDCl₃



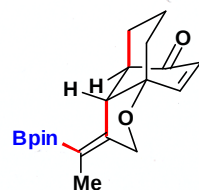
2012576yf02-16-01

Sample Name:
2012576yf02-16-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-16-01_20130408_01
FidFile: gCOSY_01

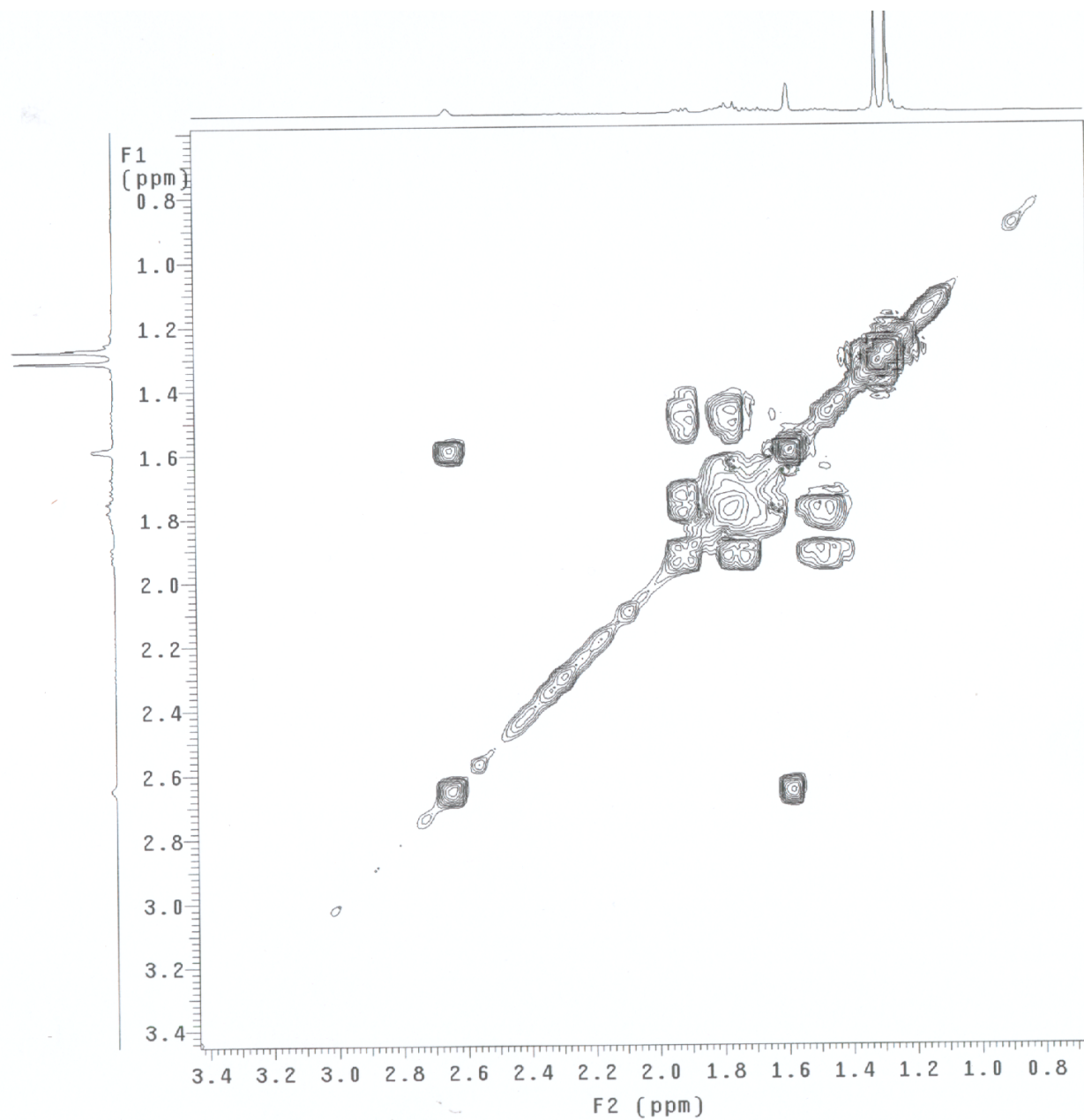
Pulse Sequence: gCOSY
Solvent: cdcl3
Data collected on: Apr 8 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4058.4 Hz
2D Width 4058.4 Hz
8 repetitions
256 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Sq. sine bell 0.040 sec
FT size 2048 x 2048
Total time 41 min



5I (H-H COSY)
Solvent: CDCl₃



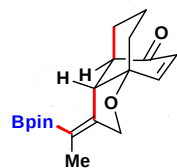
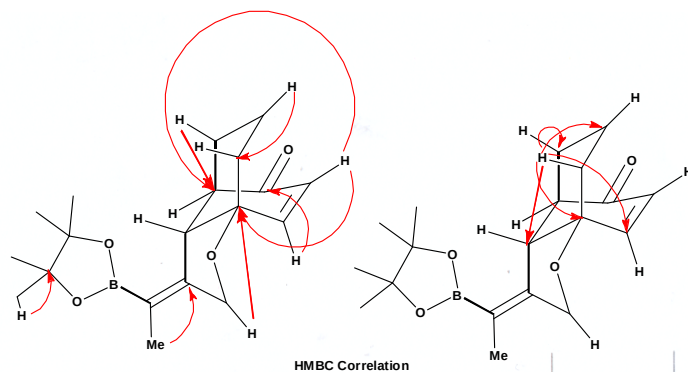
2012576yf02-16-01

Sample Name:
2012576yf02-16-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-16-01_20130403_01
FidFile: gHMBCAD_01

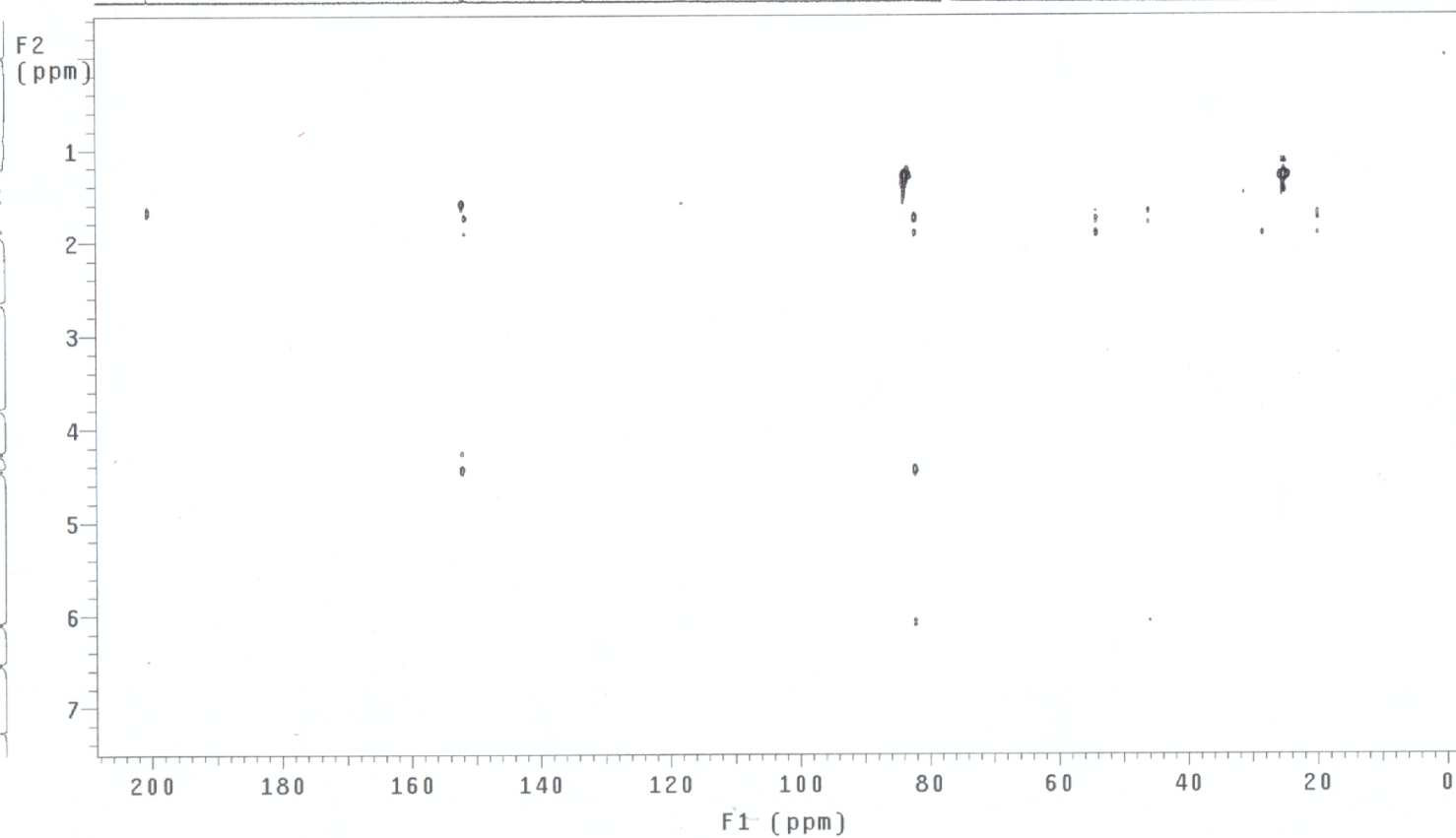
Pulse Sequence: gHMBCAD
Solvent: cdc13
Data collected on: Apr 4 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3742.5 Hz
2D Width 24118.2 Hz
64 repetitions
2 x 400 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.015 sec
FT size 2048 x 4096
Total time 17 hr, 39 min



5I (HMBC)
Solvent: CDCl₃



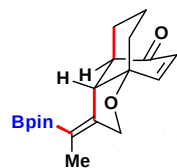
2012576yf02-16-01

Sample Name:
2012576yf02-16-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-16-01_20130403_01
FidFile: gHMBCAD_01

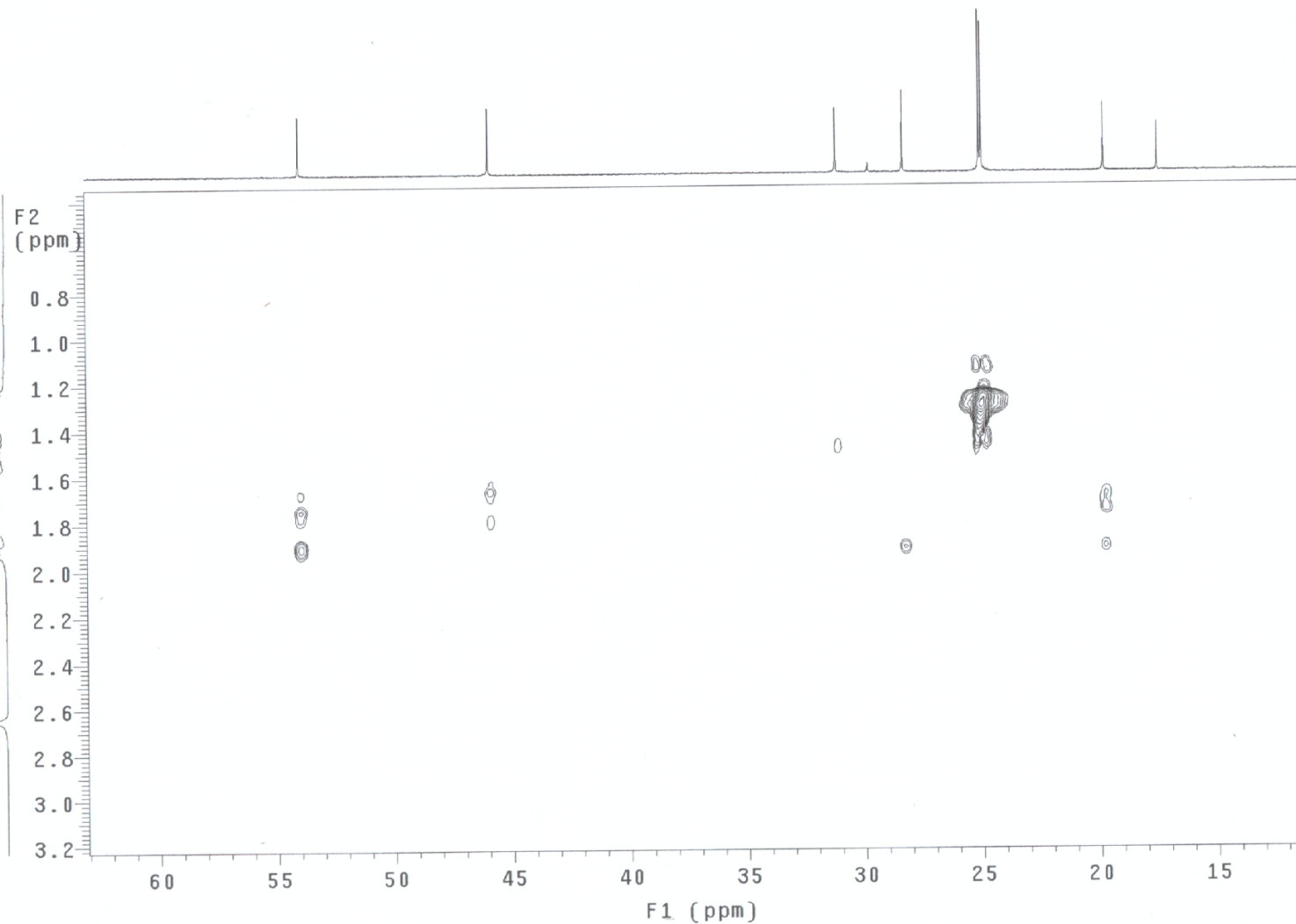
Pulse Sequence: gHMBCAD
Solvent: cdcl3
Data collected on: Apr 4 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3742.5 Hz
2D Width 24118.2 Hz
64 repetitions
2 x 400 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.015 sec
FT size 2048 x 4096
Total time 17 hr, 39 min



5I (HMBC)
Solvent: CDCl₃

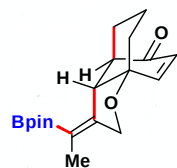


Sample Name:
2012576yf02-16-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-16-01_20130408_01
FidFile: gHSQCAD_01

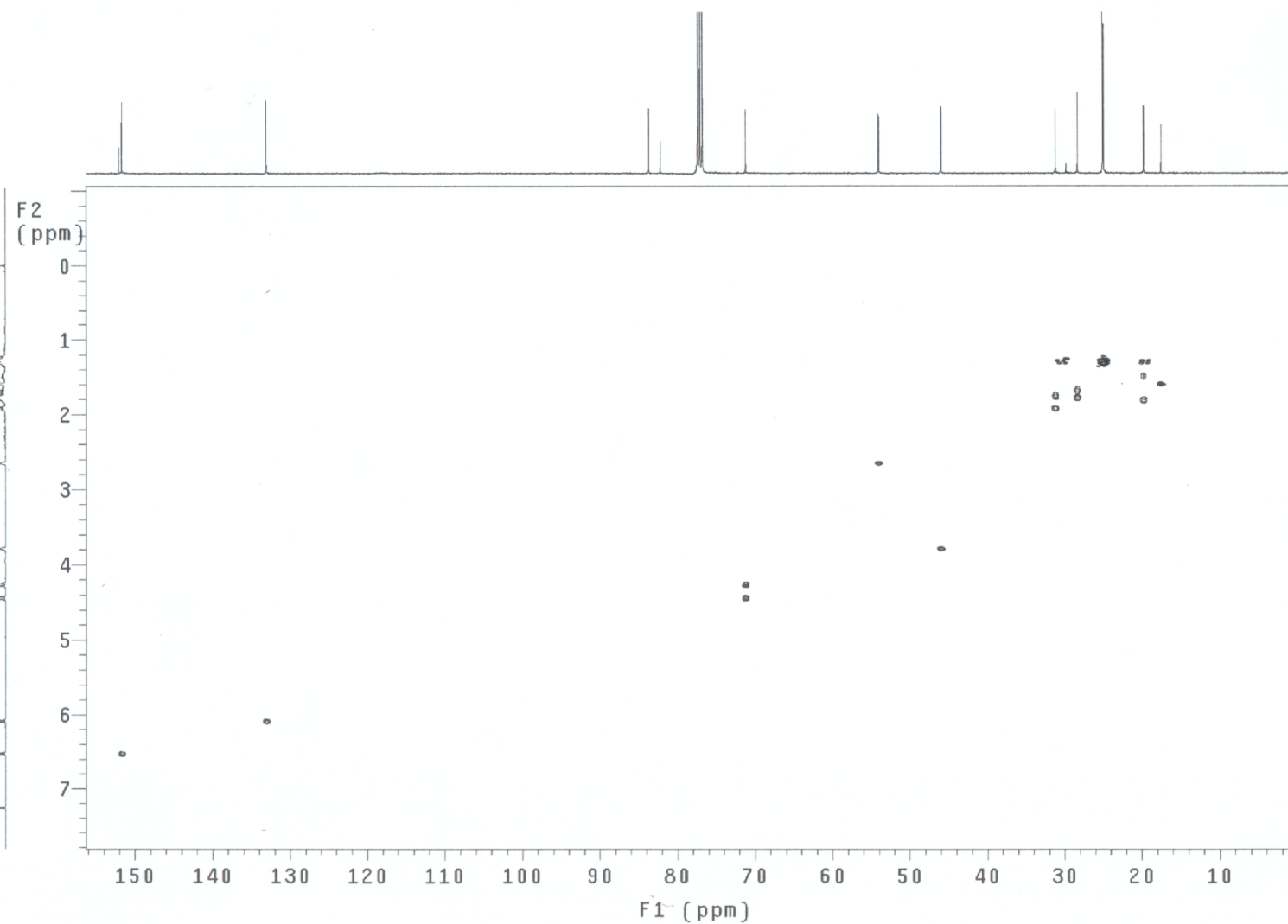
Pulse Sequence: gHSQCAD
Solvent: cdcl3
Data collected on: Apr 9 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4058.4 Hz
2D Width 20100.5 Hz
8 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DECOUPLE C13, 100.5036548 MHz
Power 37 dB
on during acquisition
off during delay
W40_ATB3 modulated
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.010 sec
FT size 2048 x 2048
Total time 1 hr, 22 min



5I (HMBC)
Solvent: CDCl₃



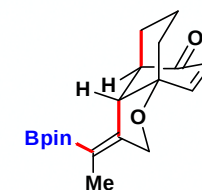
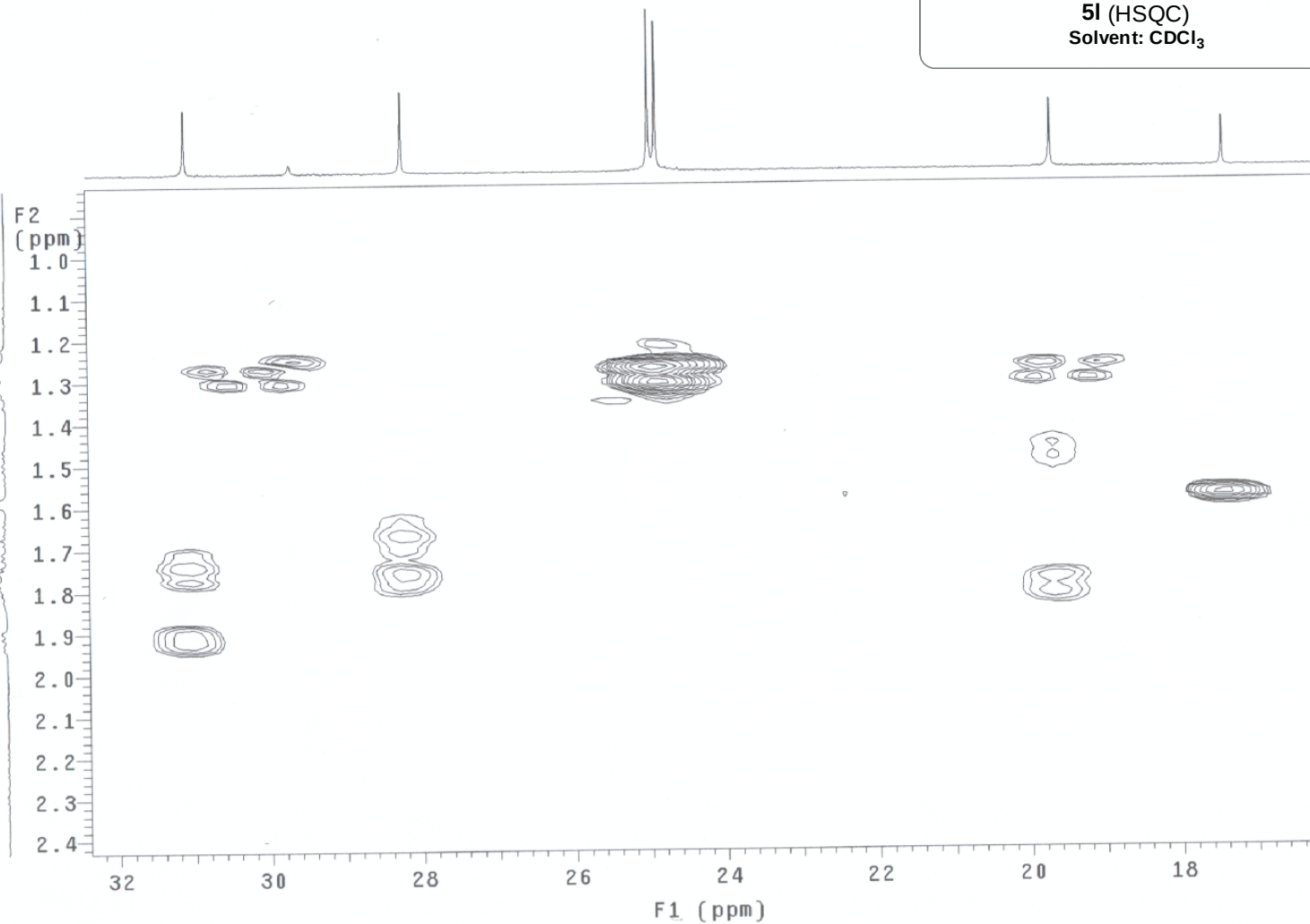
2012576yf02-16-01

Sample Name:
2012576yf02-16-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-16-01_20130408_01
FidFile: gHSQCAD_01

Pulse Sequence: gHSQCAD
Solvent: cdc13
Data collected on: Apr 9 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4058.4 Hz
2D Width 20100.5 Hz
8 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DECOUPLE C13, 100.5036548 MHz
Power 37 dB
on during acquisition
off during delay
W40_ATB3 modulated
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.010 sec
FT size 2048 x 2048
Total time 1 hr, 22 min



5I (HSQC)
Solvent: CDCl₃

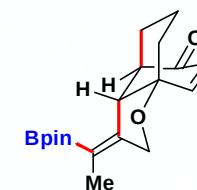
2012576yf02-16-01

Sample Name:
2012576yf02-16-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-16-01_20130408_01
FidFile: gHSQCAD_01

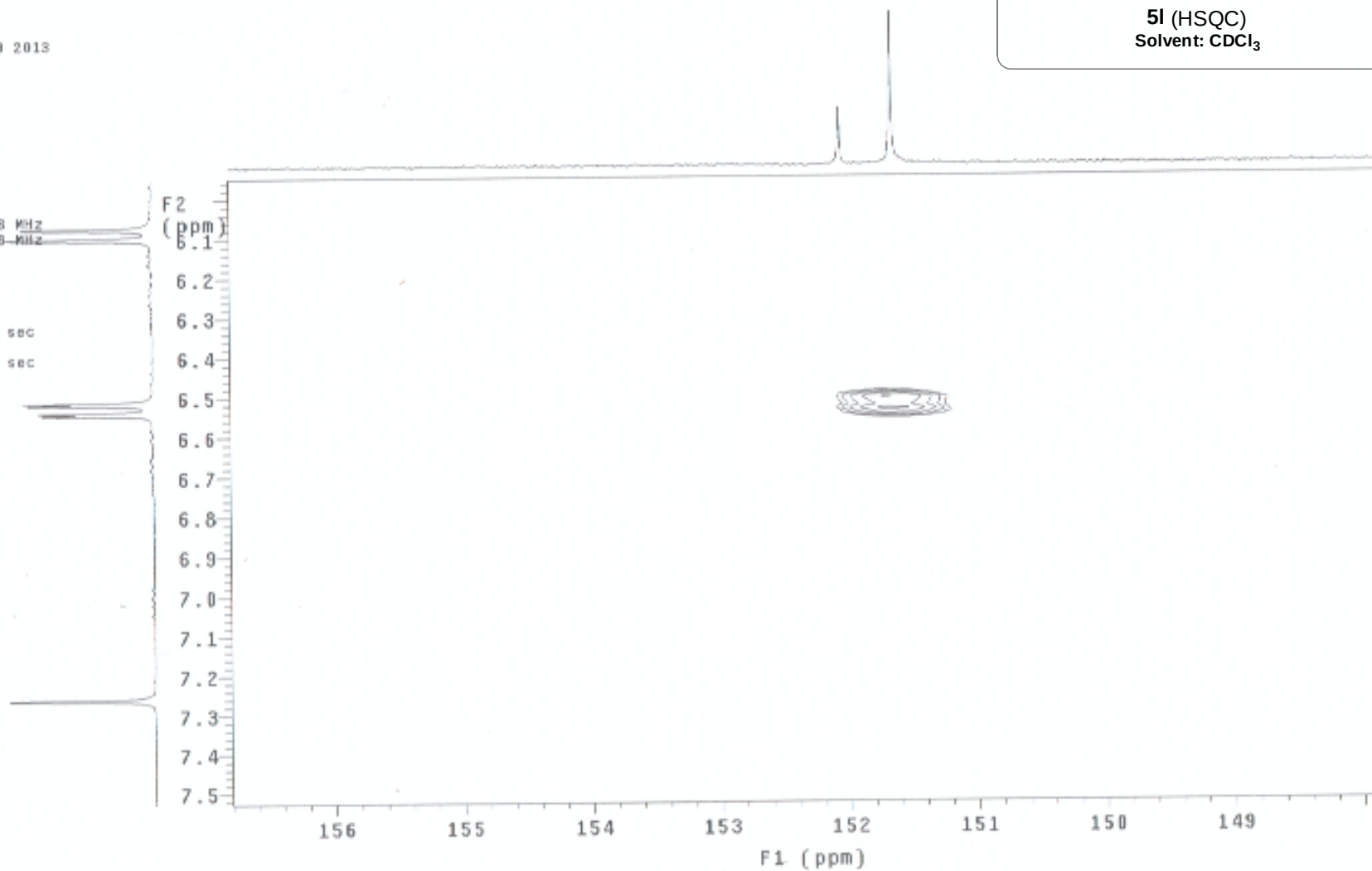
Pulse Sequence: gHSQCAD
Solvent: cdcl3
Data collected on: Apr 9 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4058.4 Hz
2D Width 20100.5 Hz
8 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DECOUPLE C13, 100.5036548 MHz
Power 37 dB
on during acquisition
off during delay
V40_AT83 modulated
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.010 sec
FT size 2048 x 2048
Total time 1 hr, 22 min



5I (HSQC)
Solvent: CDCl₃



2012576yf02-16-01

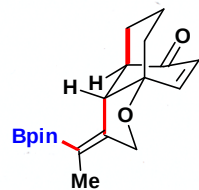
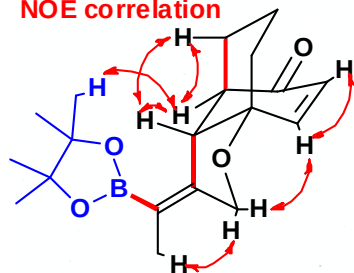
Sample Name:
2012576yf02-16-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-16-01_20130408_01
FidFile: NOESY_01

Pulse Sequence: NOESY
Solvent: cdc13
Data collected on: Apr 8 2013

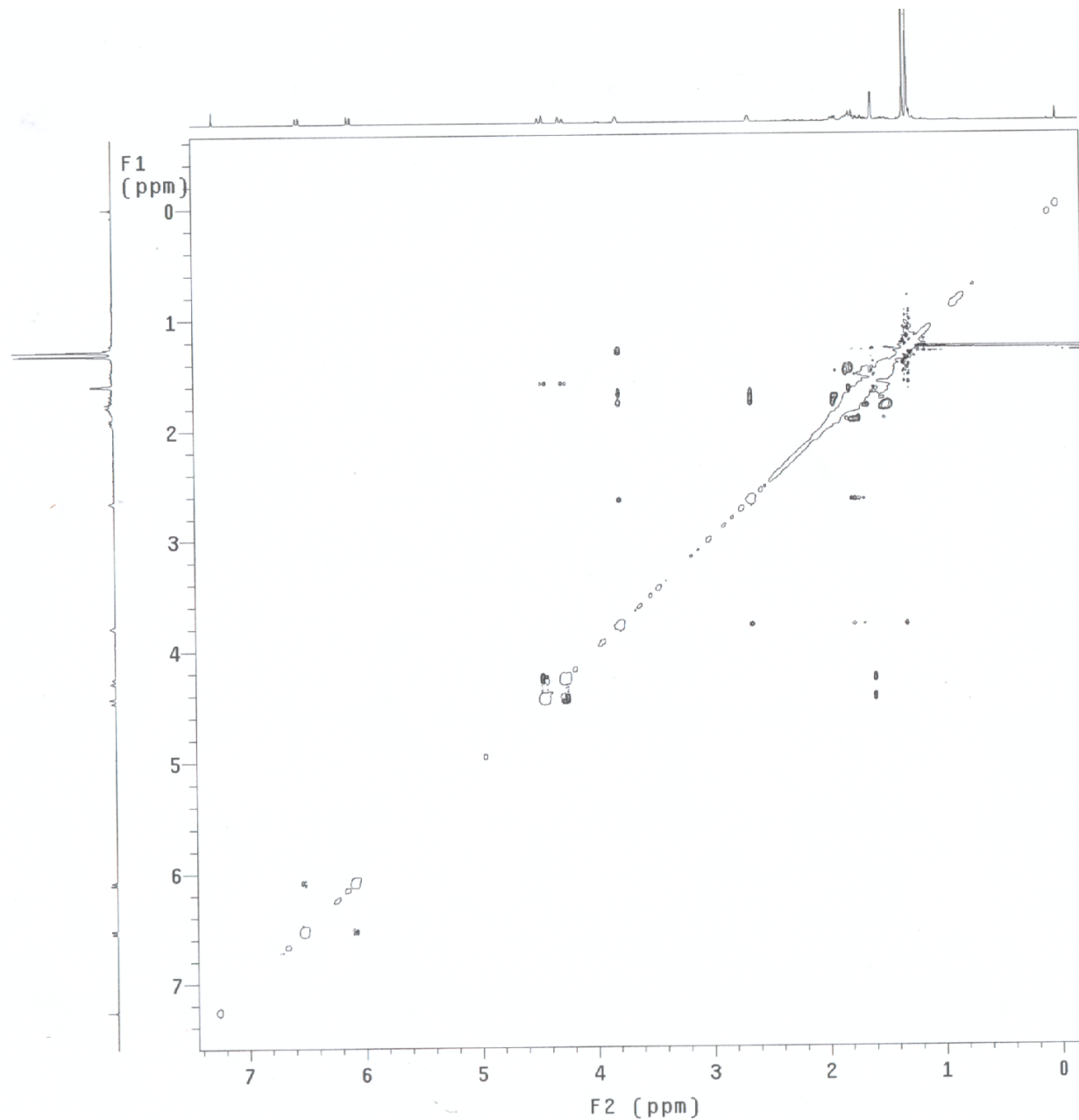
Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4058.4 Hz
2D Width 4058.4 Hz
64 repetitions
2 x 200 increments
OBSERVE H1, 399.6606928 MHz
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.045 sec
FT size 4096 x 4096
Total time 13 hr, 22 min

NOE correlation



5I (NOSEY)
Solvent: CDCl₃



2012576yf02-16-01

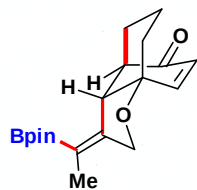
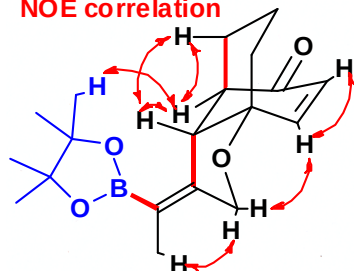
Sample Name:
2012576yf02-16-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-16-01_20130408_01
FidFile: NOESY_01

Pulse Sequence: NOESY
Solvent: cdc13
Data collected on: Apr 8 2013

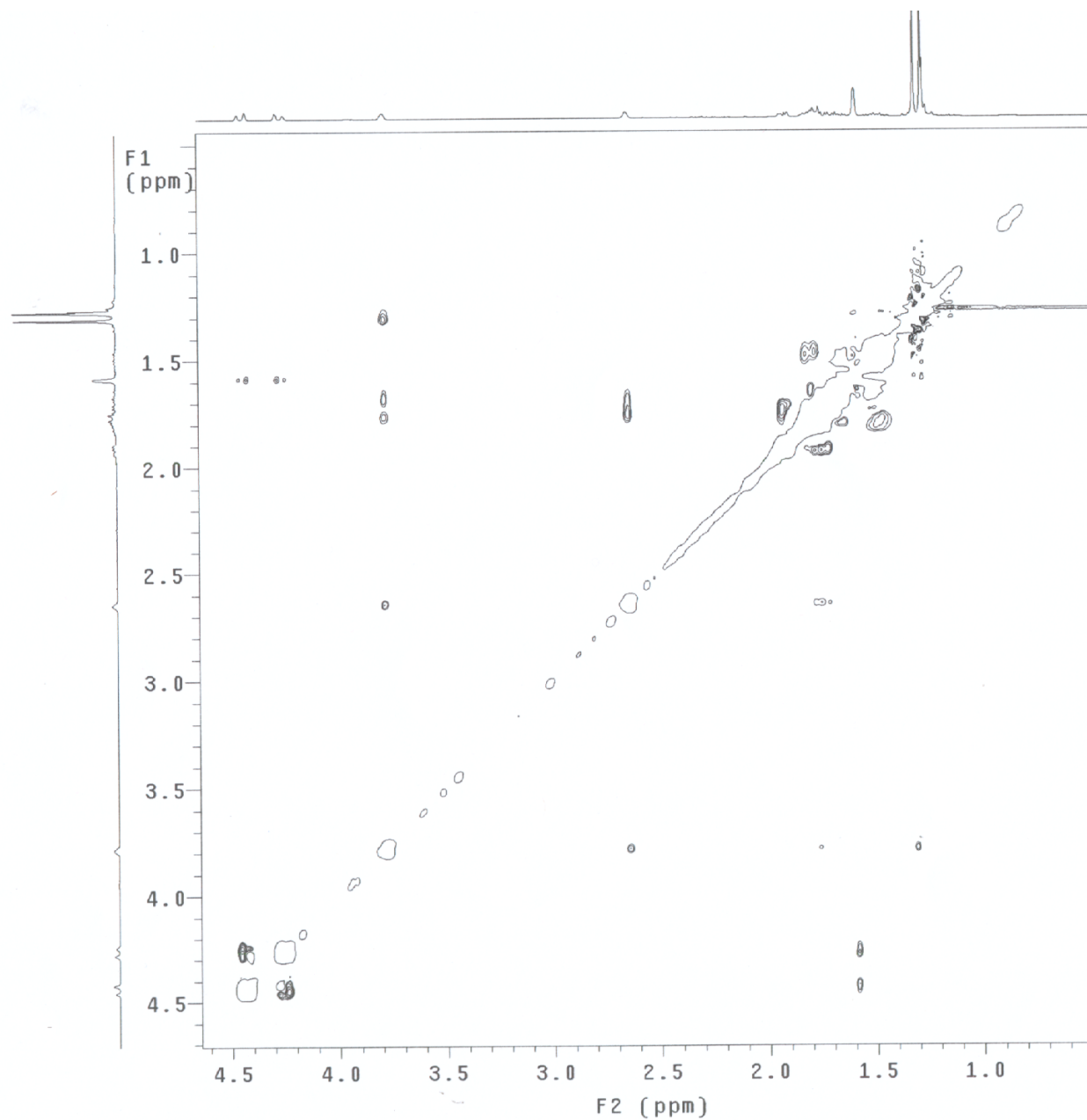
Temp. 25.0 C / 298.1 K
Operator: sioc

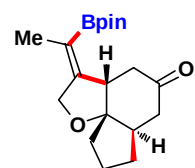
Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 4058.4 Hz
2D Width 4058.4 Hz
64 repetitions
2 x 200 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.045 sec
FT size 4096 x 4096
Total time 13 hr, 22 min

NOE correlation

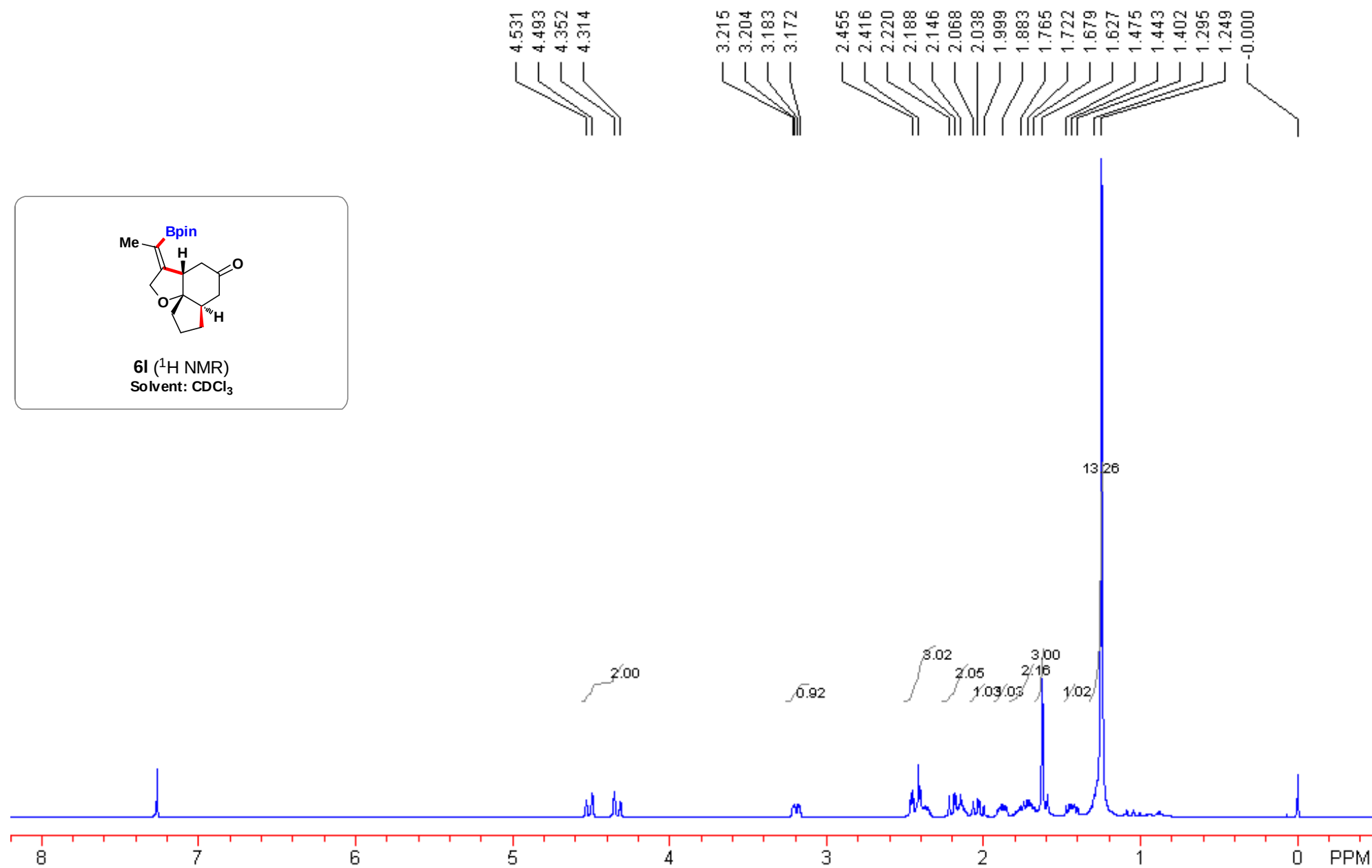


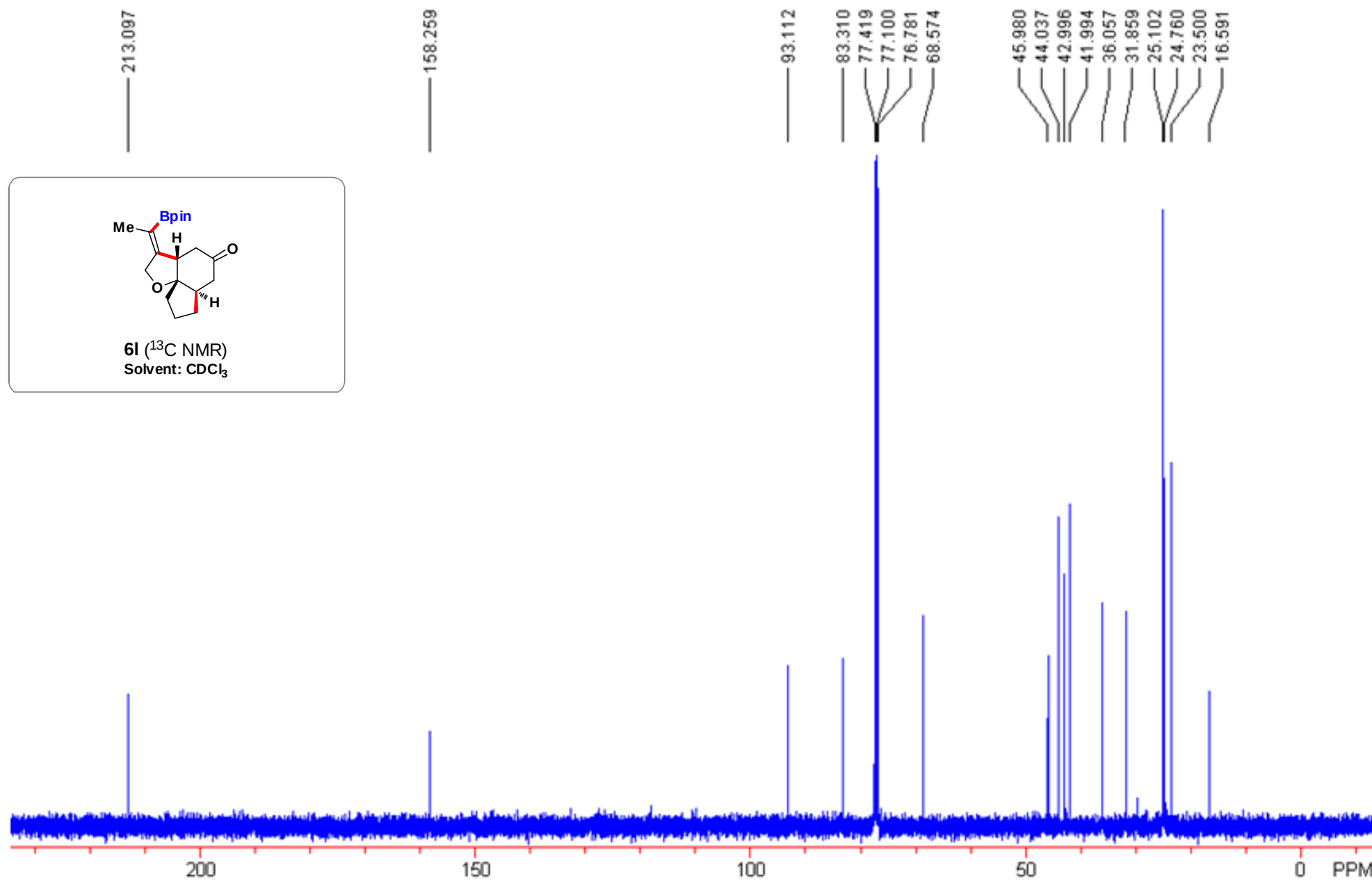
5l (NOSEY)
Solvent: CDCl₃

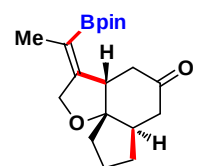




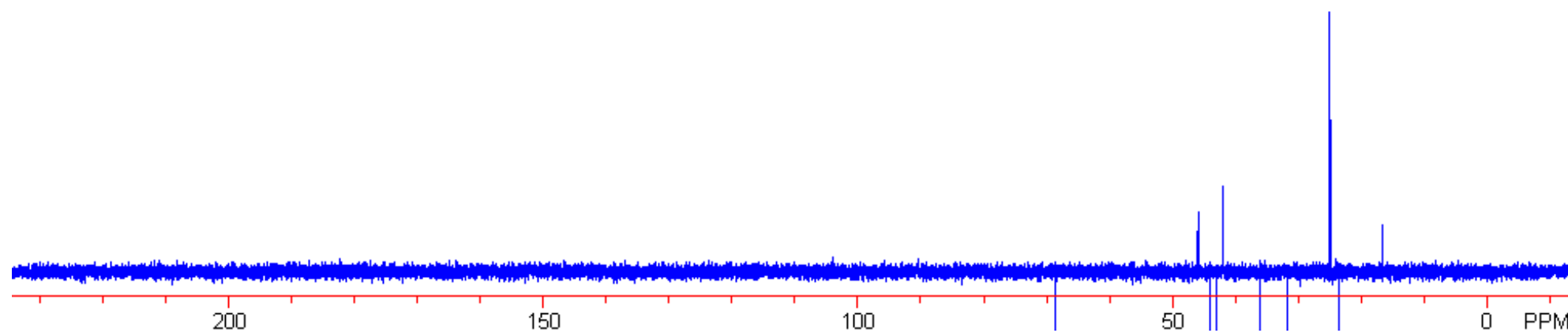
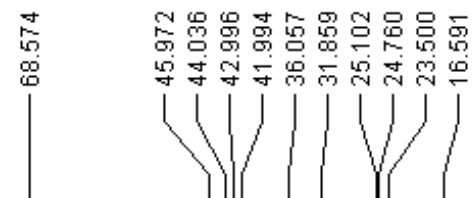
6I (^1H NMR)
Solvent: CDCl_3







6l (DEPT, $\theta = 135^\circ$)
Solvent: CDCl_3



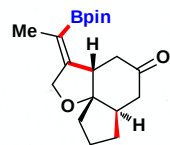
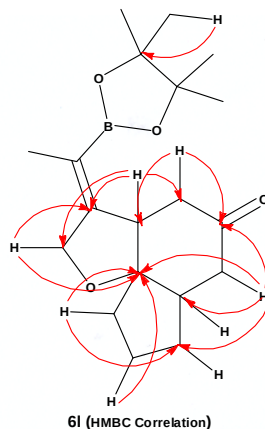
2012576yf02-22-01

Sample Name:
2012576yf02-22-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-22-01_20130410_01
FidFile: gHMBCAD_01

Pulse Sequence: gHMBCAD
Solvent: cdcl3
Data collected on: Apr 10 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3720.2 Hz
2D Width 24118.2 Hz
16 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.010 sec
FT size 2048 x 2048
Total time 2 hr, 49 min



6I (HMBC)
Solvent: CDCl₃

F2
(ppm)

0.5
1.0
1.5
2.0
2.5
3.0
3.5
4.0
4.5
5.0

200 180 160 140 120 100 80 60 40 20 0

F1 (ppm)

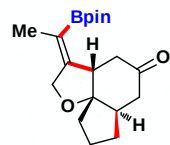
2012576yf02-22-01

Sample Name:
2012576yf02-22-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-22-01_20130410_01
FidFile: gHMBCAD_01

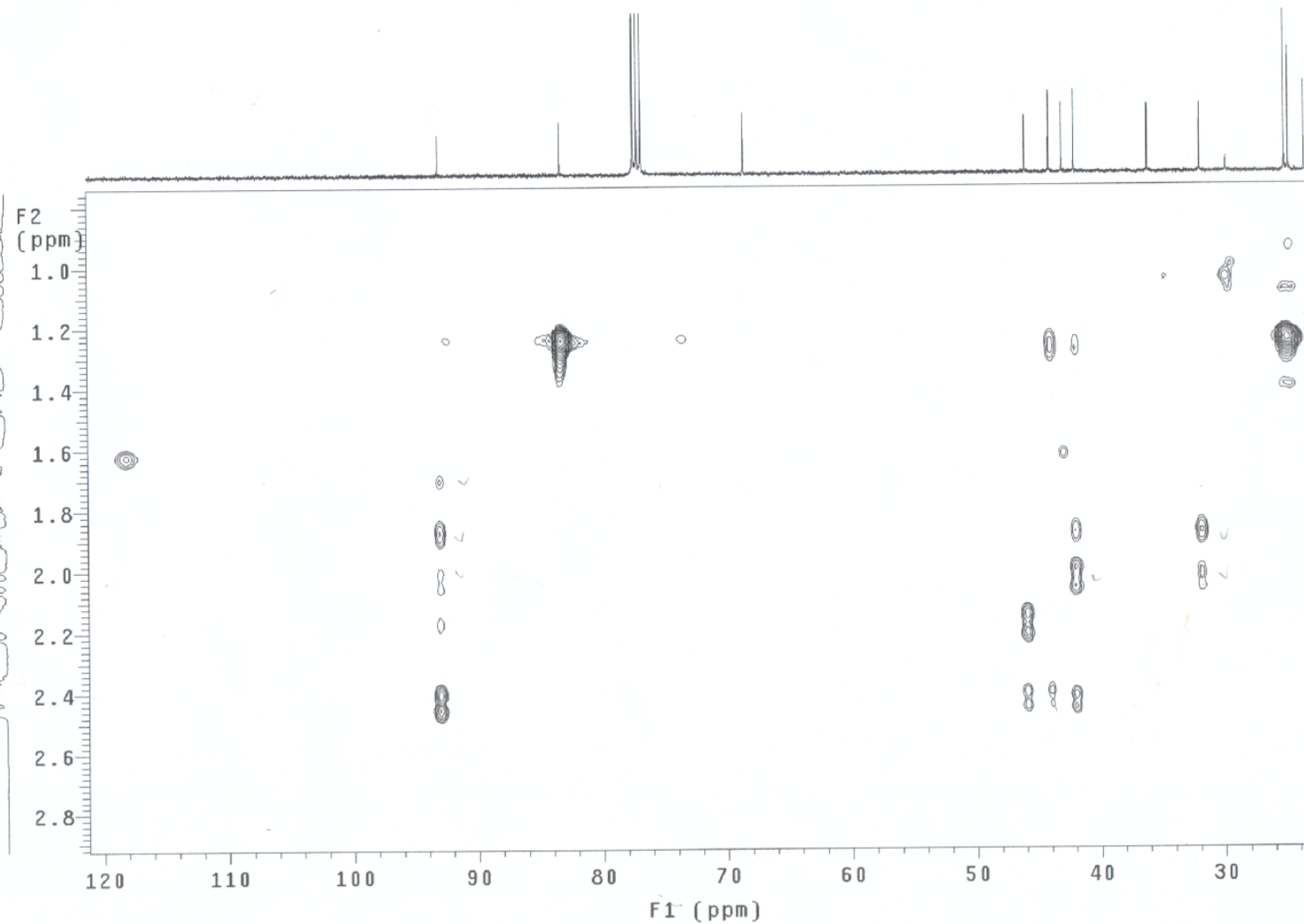
Pulse Sequence: gHMBCAD
Solvent: cdcl3
Data collected on: Apr 10 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3720.2 Hz
2D Width 24118.2 Hz
16 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Sq. sine bell 0.075 sec
F1 DATA PROCESSING
Gauss apodization 0.010 sec
FT size 2048 x 2048
Total time 2 hr, 49 min



6I (HMBC)
Solvent: CDCl₃



```
Sample Name:
  2012576yf02-22-01
Data Collected on:
  Agilent-NMR-vnmrs400
Archive directory:
  /home/sioc/date
Sample directory:
  2012576yf02-22-01_20130410_01
FidFile: gHSQCAD_01
```

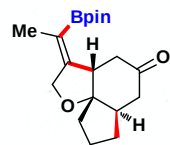
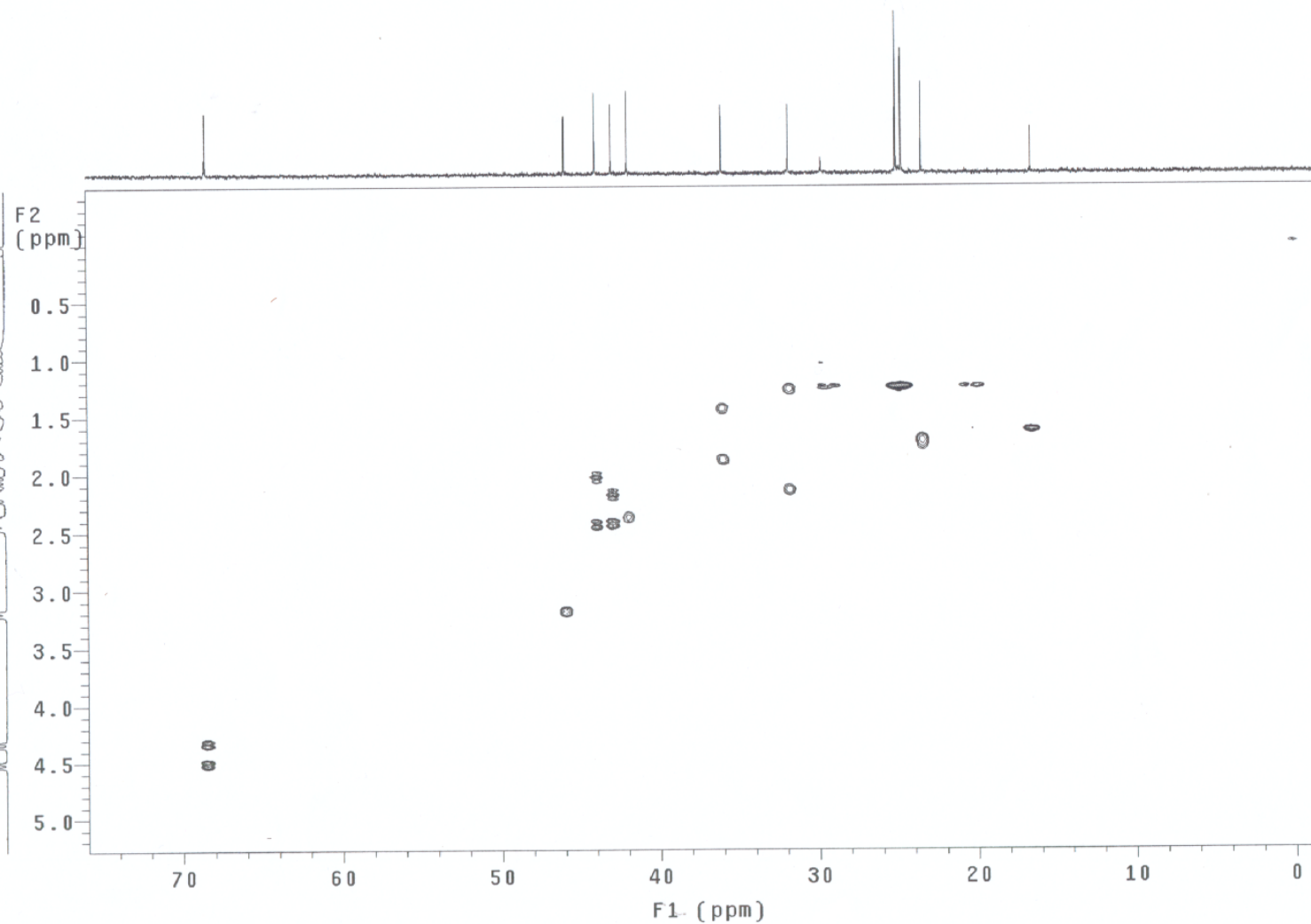
Pulse Sequence: gHSQCAD
Solvent: cdcl3
Data collected on: Apr 10 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

```

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3720.2 Hz
2D Width 20100.5 Hz
8 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DECOUPLE C13, 100.5036548 MHz
Power 37 dB
on during acquisition
off during delay
W40_ATB3 modulated
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.012 sec
FT size 2048 x 2048
Total time 1 hr, 22 min

```



6l (HSQC)
Solvent: CDCl_3

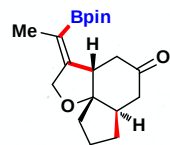
2012576yf02-22-01

Sample Name:
2012576yf02-22-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-22-01_20130410_01
FidFile: gHSQCAD_01

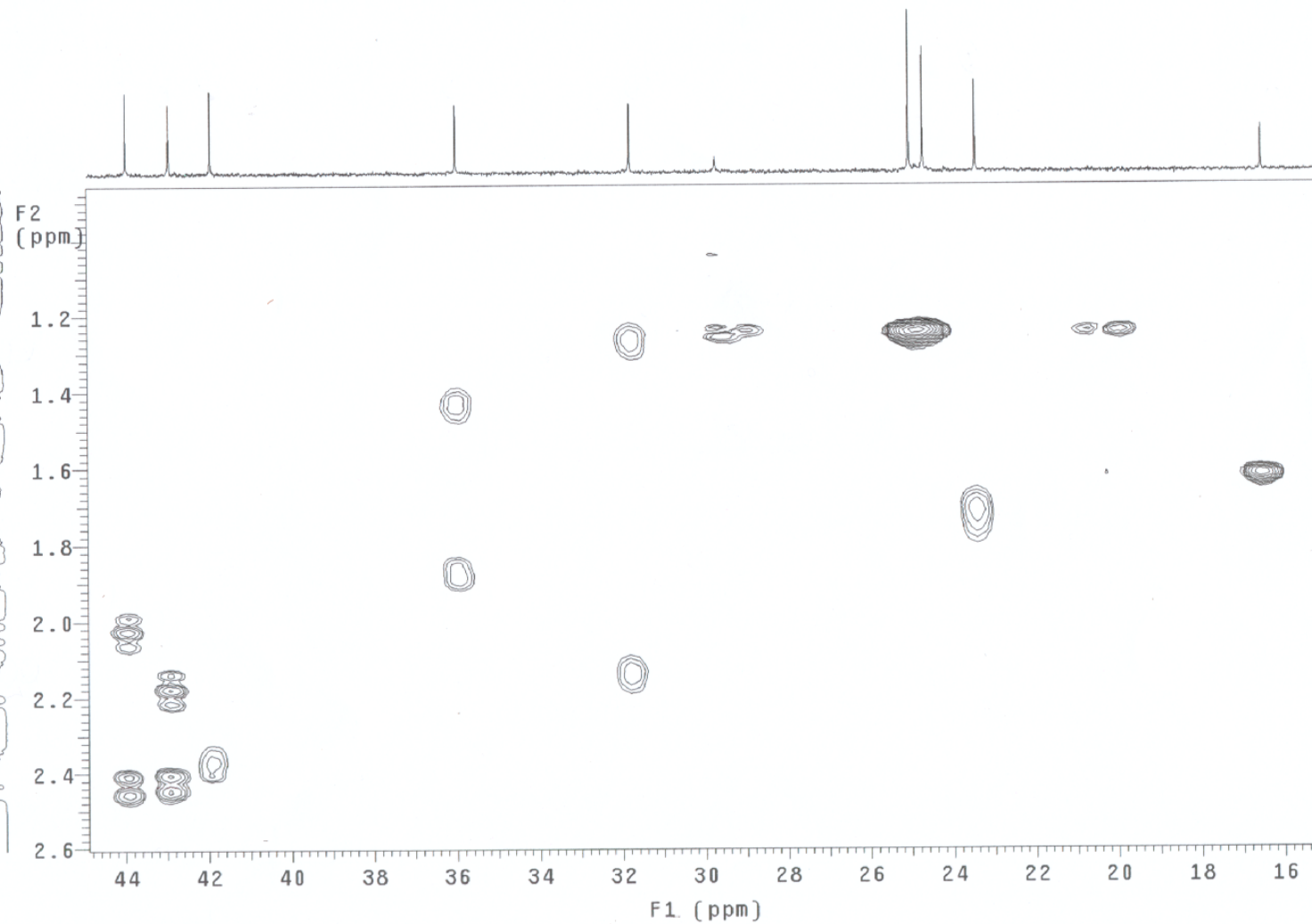
Pulse Sequence: gHSQCAD
Solvent: cdcl3
Data collected on: Apr 10 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3720.2 Hz
2D Width 20100.5 Hz
8 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DECOUPLE C13, 100.5036548 MHz
Power 37 dB
on during acquisition
off during delay
W40_ATB3 modulated
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.012 sec
FT size 2048 x 2048
Total time 1 hr, 22 min



6I (HSQC)
Solvent: CDCl₃



2012576yf02-22-01

Sample Name:
2012576yf02-22-01
Data Collected on:
Agilent-NMR-vnmrs400
Archive directory:
/home/sioc/date
Sample directory:
2012576yf02-22-01_20130410_01
FidFile: NOESY_01

Pulse Sequence: NOESY
Solvent: cdc13
Data collected on: Apr 10 2013

Temp. 25.0 C / 298.1 K
Operator: sioc

Relax. delay 1.000 sec
Acq. time 0.150 sec
Width 3720.2 Hz
2D Width 3720.2 Hz
32 repetitions
2 x 256 increments
OBSERVE H1, 399.6600928 MHz
DATA PROCESSING
Gauss apodization 0.069 sec
F1 DATA PROCESSING
Gauss apodization 0.037 sec
FT size 2048 x 2048
Total time 8 hr, 36 min

