

SUPPORTING INFORMATION

Evaluating Heterogeneous Binding Avidities of Populations of Ligand-Functionalized Multivalent Nanoparticles

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Figure S1. Potentiometric titration of G5 PAMAM dendrimers with surface acylation shows that approximately 80% of the surface is converted from amines to acyl groups (G5-Ac₈₀-NH₂).

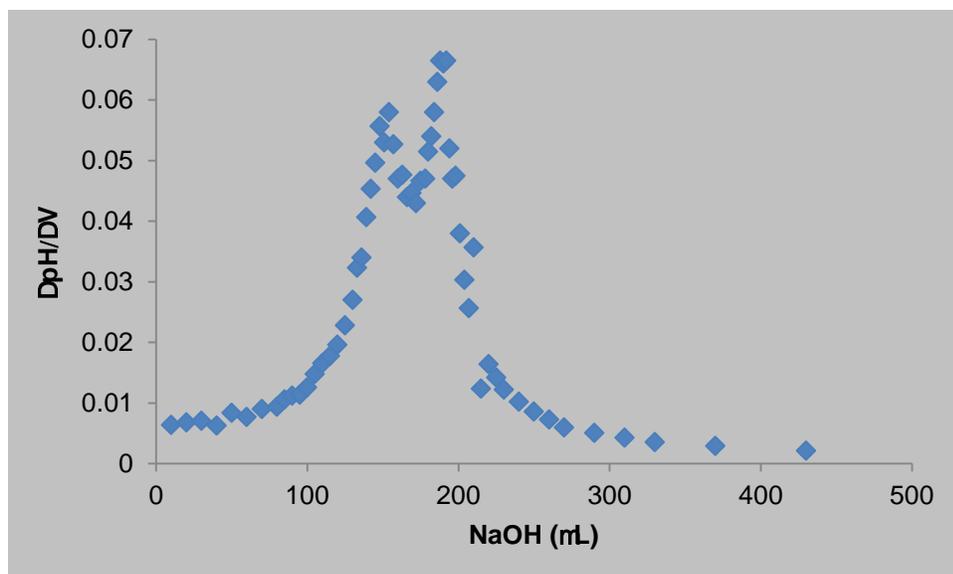
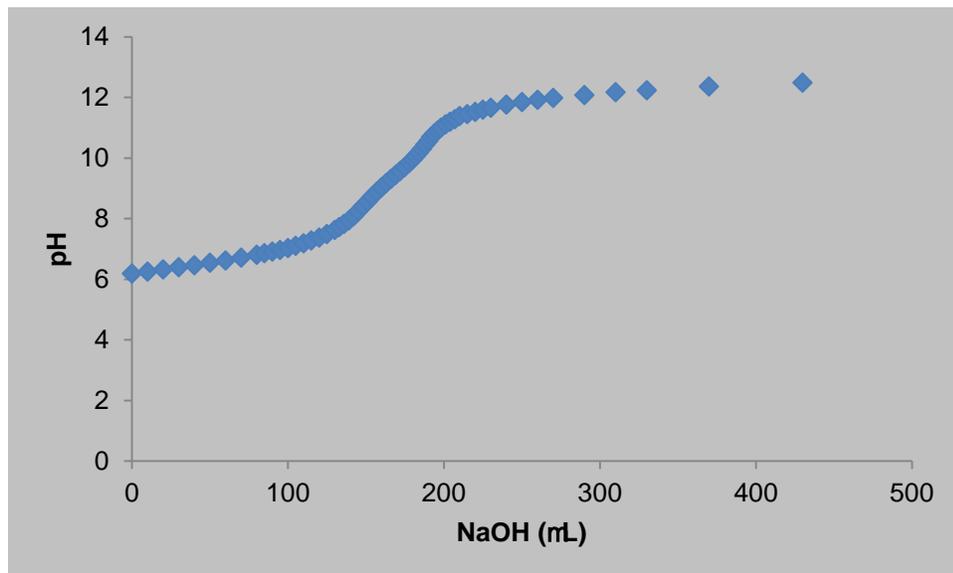
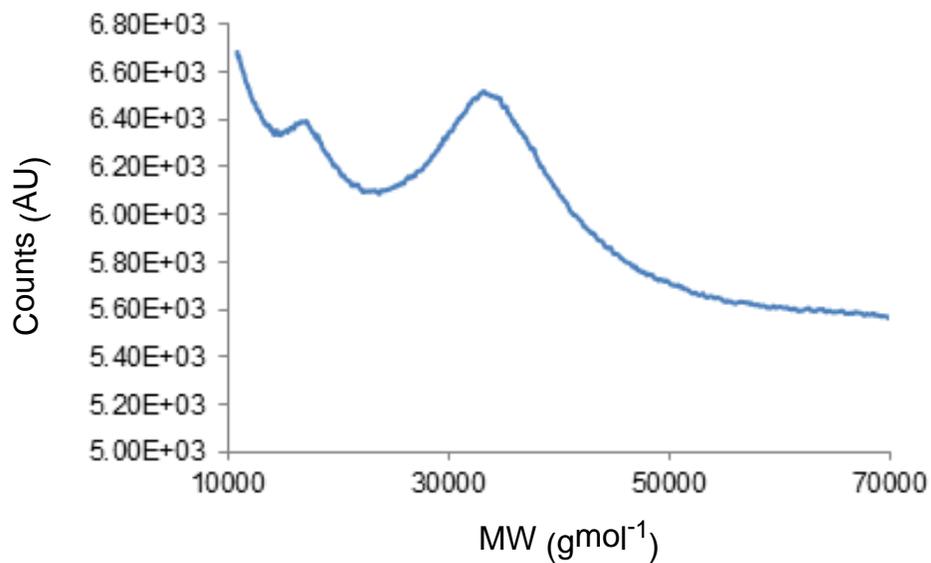


Figure S2. MALDI-TOF MS spectra of (a) G5-(oligo)₆ (MW= 34500) (b) G5-(oligo)_{3.1} (MW= 32200) (c) G5-(oligo)_{1.7} (MW = 32400).

(a)



(b)

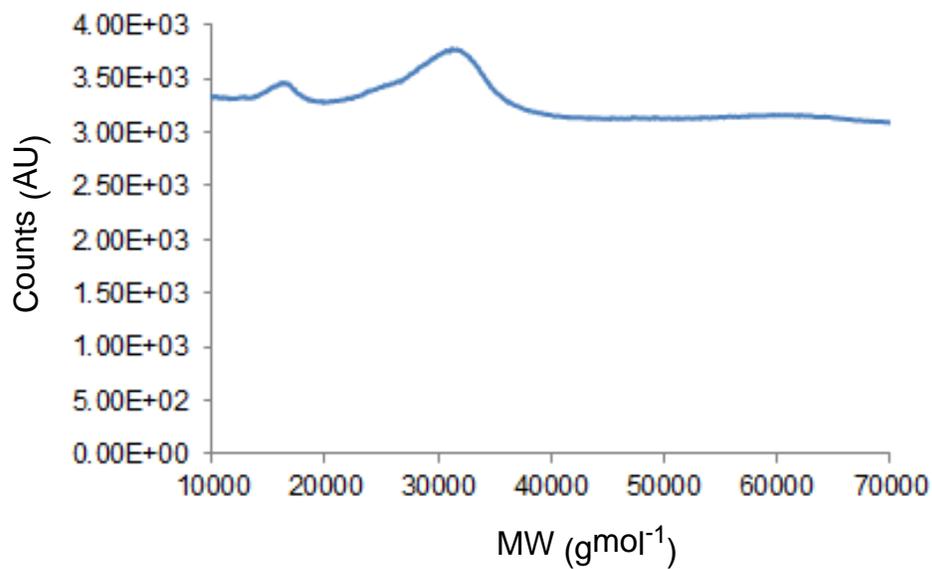


Figure S3. UV-Vis spectra of G5-(oligo)₆ (blue); G5-(oligo)_{3.1} (red) (G5-oligo)_{1.7} (green), measured at 0.05 mg/mL. Peak at absorption max of DNA (260 nM) clearly shows difference in the number of oligonucleotides per dendrimer.

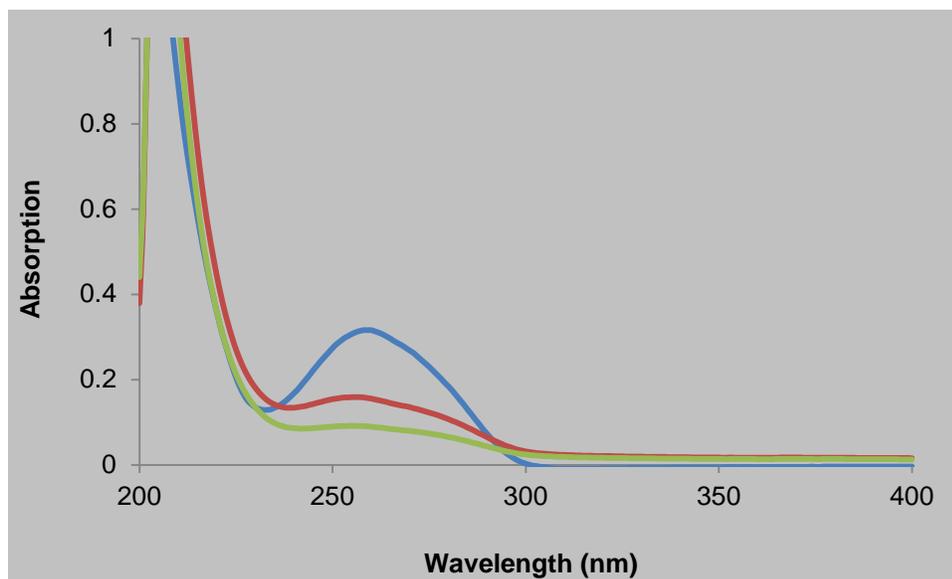
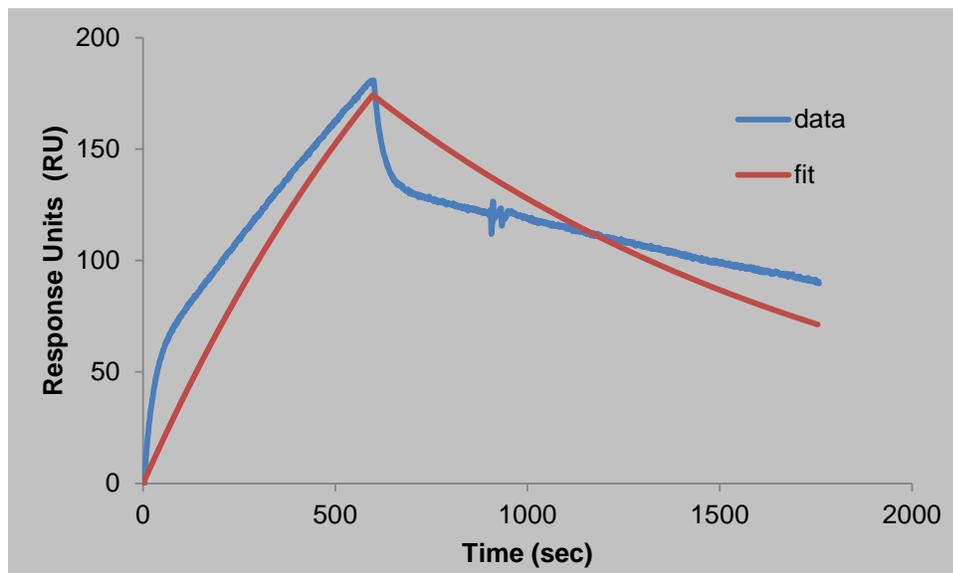


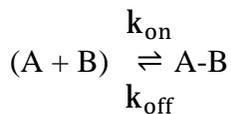
Figure S4. Binding curve of G5-(oligo)₆ and a representative fit of the curve data using a 1:1 Langmuir model. The concentration of G5-(oligo)₆ was 62.5 nM. ($c^2 = 369$)



Initial rate analysis coupled with SPR analysis

Ligand-receptor interactions are typically described with 1:1 Langmuir kinetic model (shown as Equation 1, below). In this equation, A and B are the ligand and receptor, where k_{on} , and k_{off} represent the association and dissociation rate constants of the interaction. The ratio of these constants, K_D , the dissociation rate constant, has units of concentration and is conventionally used as an indicator of binding strength. Kinetic data from optical biosensor analyses can be described as a first-order differential equation (Equation 2) and its integrated form can be employed to fit kinetic parameters. (shown in Equation 3) In this analysis R represents the response unit (RU) from the sensor surface at the time of measurement, R_{max} is the maximum RU of the receptor-covered surface and C is the concentration of the ligand. Equation 4 is derived by the differentiation of Equation 3 with respect to time (t), At $t=0$, Equation 4 can be simplified to yield Equation 5. These transformations document that the initial slope of the RU (at $t=0$) is positively associated with the concentration of the ligand because in this situation R_{max} and C are constants.

Equation 1



Equation 2

$$\frac{dR}{dt} = R_{max} * k_{on} * (R_{max} - R) - k_{off} * R$$

Equation 3

$$R = \frac{R_{\max} * C}{k_{\text{on}} * C + k_{\text{off}}} * (1 - e^{(-k_{\text{on}} * C + k_{\text{off}})t})$$

Equation 4

$$\frac{dR}{dt} = \frac{R_{\max} * C}{K_D + C} * (k_{\text{on}} * C + k_{\text{off}}) * e^{(-k_{\text{on}} * C + k_{\text{off}})t}$$

Equation 5

$$\left. \frac{dR}{dt} \right|_0 = R_{\max} * k_{\text{on}} * C$$