# Highly efficient synthesis of DNA-binding polyamides using a convergent fragmentbased approach

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# Contents

1.0	Abbre	viations	4
2.0	Traditi	ional Solid Phase Synthesis of Polyamides	5
3.0	Conve	rgent Synthesis Approach of this Study	6
4.0	Solutio	on Phase Tetramer Synthesis	7
5.0	Experi	imental Section	7
5.1	Gen	neral	7
5.2	Solı	ution phase synthesis of polyamide tetramers	8
5.3	Mar	nual solid phase synthesis protocol	33
5.	3.1	General preparation of resin and coupling protocol for solid phase synthesis of polyamides	33
5.	3.2	General cleavage protocol and isolation of polyamide	33

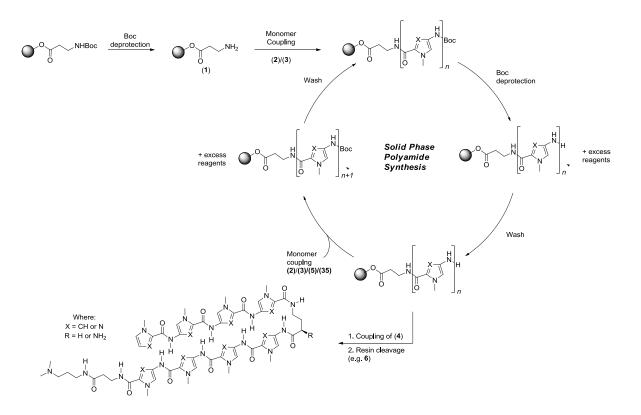
5.3.	HPLC analysis and purification of polyamides	
5.4	General activation protocols for polyamide synthesis	
5.4.	Activation with HATU	
5.4.	Activation with DCC/HOAt	
5.4.	Activation with BTC	
5.5	Synthesis of polyamides 17a-19a through solid phase synthesis	
5.5.	5.1 Synthesis of polyamide (17a)	
5.5.	5.2 Synthesis of polyamide (18a)	
5.5.	5.3 Synthesis of polyamide (19a)	
5.5.	Characterisation of Polyamides 17a-19a	
5.6	Synthesis of polyamides using a half-half coupling protocol	
5.6.	5.1 Preparation of resin bound tetramer	
5.6.	5.2 Synthesis of polyamide (17b)	50
5.6.	5.3 Synthesis of polyamide (18b)	51
5.6.	5.4 Synthesis of polyamide (19b)	
5.7	Characterisation of half-half polyamides	53
5.7.	7.1 Crude HPLC traces	53
5.7.	Analytical HPLC traces of polyamides purified by semi-preparative HPLC	56
5.7.	HRMS data of crude polyamides 17b-19b	59
5.8 Synthesis of polyamides using a hydrazine linker		
5.8.	Resin Preparation	
5.8.	S.2 Synthesis of polyamides using the PAM-hydrazine hybrid resin	64

5.	8.3	Cleavage of polyamides using the hydrazine resin	. 69
5.9	HP	LC analysis and purification of polyamides prepared on the hydrazine hybrid resin	. 72
5.	9.1	Crude HPLC traces of polyamides (21-24)	. 72
5.	9.2	HPLC traces of polyamides (21-24) after semi-preparative HPLC purification	. 76
5.	9.3	Crude HRMS data of polyamides (21-24)	. 80
5.10	Pre	paration of polyamides using the half-half coupling method on the hydrazine resin	. 84
5.	10.1	Preparation of resin-bound tetramer (16)	. 84
5.	10.2	Synthesis of (20)	. 85
5.11	Cha	aracterisation of (20)	. 85
6.0	NMR	Spectra	. 88
7.0	Refer	ences	129

#### 1.0 Abbreviations

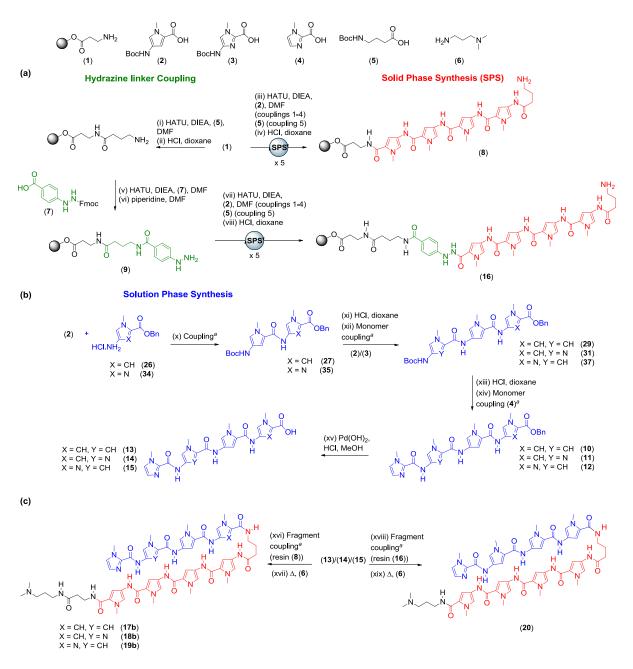
BTC, bis(trichloromethyl)carbonate; DCC, N,N'-dicyclohexylcarbodiimide; DCM, dichloromethane; DCU, dicyclohexylurea; DIEA, Ndiisopropylethylamine; DMF, N,N'-dimethylformamide; Fmoc-D-Dab(Boc)-OH, N-α-(9-fluorenylmethyloxycarbonyl)-N-γ-t-butyloxycarbonyl-D-2,4diaminobutyric acid; GABA, γ-aminobutyric acid; HATU, 2-(1H-7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyl uronium hexafluorophosphate; HBA, 4-(2-Fmoc-hydrazino)benzoic acid; HOAt, Hydroxy-7-azabenzotriazole; TFA, trifluoroacetic acid; THF, tetrahydrofuran; TPW, TFA:phenol:water (92.5 mL: 5 g: 2.5 mL).

# 2.0 Traditional Solid Phase Synthesis of Polyamides



Scheme S1 (Schematic representation of solid phase polyamide synthesis using existing Boc-based methodology. Stepwise Boc-deprotection of resin-bound amine and coupling of the next monomer requires a total of 18 steps to form a standard 8 ring hairpin polyamide.)

# 3.0 Convergent Synthesis Approach of this Study



Scheme S2: Schematic representation of our new convergent synthetic approach. (a) Solid phase synthesis of C-terminal tetramers (8 and 16, shown in red). (b) Solution phase synthesis of N-terminal tetramers (13-15, shown in blue). (c) Ligation of the two respective "halves" (13-15) and (8 and 16) and cleavage from the resin afforded full length PAs.

#### 4.0 Solution Phase Tetramer Synthesis

Entry	Compound Number	Tetramer	Yield (%) <sup>a</sup>	Purity (%) <sup>b</sup>	
1	13	●	73	89	
2	14	●-●-○-○-<С	80	95	
3	15	●-O-O-●-{ <sub>H</sub>	59	93	

Table 1: Isolated yields of pyrrole/imidazole acid tetramers 13-15 prepared by solution-phase synthesis.

<sup>a</sup>Isolated yield after 7 steps.<sup>b</sup> Purity determined by analytical RP-HPLC at 310 nm. Imidazole units are denoted as solid circles, and pyrrole units as open circles.

#### 5.0 Experimental Section

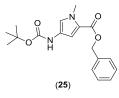
#### 5.1 General

All reagents were either HPLC or peptide synthesis grade and used as supplied. DIEA, HATU and TFA were purchased from Alfa Aesar. BTC, DCC, DCM, DMF, HBA and THF were purchased from Sigma Aldrich. GABA was obtained from NovaBioChem. Fmoc-D-Dab(Boc)-OH was purchased from Fluorochem. Boc-β-Ala PAM resin (0.26 mmol/G loading) was purchased from Peptides International. 1-Methyl-1H-imidazole-2-carboxylic acid was purchased from Maybridge Chemicals.

Boc-Py-OH and Boc-Im-OH were prepared according to the literature.<sup>1</sup>

Analytical and semi-preparative RP-HPLC were performed at room temperature on an ULTIMAT 3000 Instrument (DIONEX). UV absorbance was measured using a photodiode array detector at 310nm. A Phenomenex Gemini-NX 5µm C18 110Å packed column with dimensions 150 x 4.60 mm was used for analytical HPLC. For semi-preparative HPLC, a Phenomenex Gemini- NX 5u C18 110Å AXIA Packed column with dimensions 250 x 21.20 mm was used. LC-MS was performed using a Xevo QTof mass spectrometer (Waters) coupled to an Acquity LC system (Waters) using an Acquity UPLC BEH C18 column (2.1 x 50 mm, Waters).

# 5.2 Solution phase synthesis of polyamide tetramers Benzyl 4-((tert-butoxycarbonyl)amino)-1-methyl-1H-pyrrole-2-carboxylate (25)



To a solution of 4-((tert-butoxycarbonyl)amino)-1-methyl-1H-pyrrole-2-carboxylic acid (1.00 g; 4.16 mmol) in ethanol/water (3:1 v/v, 20 mL) was added a solution of caesium carbonate (0.678 g; 2.08 mmol) in water (5 mL). The reaction mixture was stirred at room temperature for 2 hours. The solution was then filtered and the solvent removed under vacuum, then distilled azeotropically to dryness with ethanol to give the caesium salt of (2) as an off-white/pale yellow solid. This was then dissolved in dry DMF (10 mL) and benzyl bromide (0.718 g; 4.2 mmol) was added dropwise at room temperature with constant stirring. The reaction was then heated to 40°C and stirred overnight. The reaction was quenched by the addition of ice water

(45 mL). The crude product was isolated by centrifugation and decanting of the supernatant. Any remaining water was removed azeotropically with ethanol to yield (25) as an off-white solid (0.736 g; 54 %) which required no further purification.

<sup>1</sup>H NMR (300 MHz; CDCl<sub>3</sub>) δ 1.49 (s, 9 H), 3.86 (s, 3 H), 5.25 (s, 2 H), 6.38 (s, 1 H), 6.70 (d, *J* = 2.1 Hz, 1 H), 7.08 (s, 1 H), 7.28-7.42 (m, 5 H).

<sup>13</sup>C NMR (75 MHz; CDCl<sub>3</sub>) δ 28.4 (3C), 36.7, 65.5, 80.2, 108.3, 119.8, 120.1, 122.2, 128.0 (3C), 128.5 (2C), 136.4, 153.3, 160.8.

HRMS (ESI +ve mode) 331.1658 calculated for  $C_{18}H_{23}N_2O_4^+$  [M+H]<sup>+</sup> found 331.1660.

Data in agreement to literature.<sup>21</sup>

Benzyl 4-amino-1-methyl-1H-pyrrole-2-carboxylate hydrochloride (26):



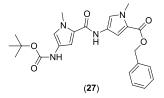
To a solid sample of (**25**) (0.200 g; 0.6 mmol) was added 4 M HCl in dioxane (2 mL; 8 mmol). The mixture was stirred at room temperature for 2 hours and reaction was confirmed complete by analytical RPHPLC. The product was precipitated with hexane then isolated by centrifugation and removal of the supernatant. The crude material was washed twice more with hexane, then suspended in water (5 mL) and lyophilised to dryness, giving the product as an off-white solid (0.160 g; quantitative), which was of sufficient purity to carry forward.

<sup>1</sup>H NMR (300 MHz;  $d_4$ -methanol)  $\delta$  3.94 (s, 3 H), 5.28 (s, 2 H), 6.94 (d, J = 2.1 Hz, 1 H), 7.18 (d, J = 2.0 Hz, 1 H), 7.28-7.45 (m, 5 H).

<sup>13</sup>C NMR (75 MHz; *d*<sub>4</sub>-methanol) δ 37.5, 67.1, 112.5, 114.5, 123.6, 124.7, 129.3 (3C), 129.6 (2C), 137.6, 161.5.

HRMS (ESI +ve mode) 231.1134 calculated for [M+H]<sup>+</sup>, found 231.1125.

Benzyl 4-(4-((tert-butoxycarbonyl)amino)-1-methyl-1H-pyrrole-2-carboxamido)-1-methyl-1H-pyrrole-2-carboxylate (27):



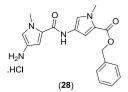
To a solution of Boc-Py-OH (2) (0.087 g; 0.365 mmol) and HATU (0.139 g; 0.365mmol) in DMF (5 mL; peptide grade) was added DIEA (500  $\mu$ L; 3 mmol). The mixture was stirred at room temperature for 30 minutes. This was then added to a separate solution of (26) (0.100 g; 0.375 mmol) and DIEA (500  $\mu$ L; 3 mmol) in DMF (5 mL; peptide grade). The mixture was stirred for 2 hours at room temperature and the reaction was shown to be complete by analytical RPHPLC. The product was precipitated by the addition of acidified water (pH 3; 40 mL) and isolated by centrifugation and removal of the supernatant. The resulting solid was washed twice more with acidified water, then suspended in water (8 mL) and lyophilised to dryness. The product was collected as an off-white solid (0.162 mg; 98 %) and was of sufficient purity to continue.

<sup>1</sup>H NMR (300 MHz; *d*<sub>6</sub>-DMSO) δ 1.44 (s, 9 H), 3.79 (s, 3 H), 3.83 (s, 3 H), 5.23 (s, 2 H), 6.82 (s, 1 H), 6.88 (s, 1 H), 6.95 (s, 1 H), 7.28-7.44 (m, 5 H), 7.46 (s, 1 H), 9.09 (s, 1 H), 9.82 (s, 1 H).

<sup>13</sup>C NMR (75 MHz; *d*<sub>6</sub>-DMSO) δ 28.2 (3C), 36.1, 36.2, 64.8, 78.3, 103.8, 108.6, 117.2, 118.4, 121.0, 122.4, 122.5, 123.1, 127.8 (2C), 128.0 (2C), 128.5, 136.6, 152.8, 158.4, 160.1.

HRMS (ESI +ve mode) 453.2138 calculated for  $C_{24}H_{29}N_4O_5^+$  [M+H]<sup>+</sup>, found 453.2120.

#### Benzyl 4-(4-amino-1-methyl-1H-pyrrole-2-carboxamido)-1-methyl-1H-pyrrole-2-carboxylate hydrochloride (28):



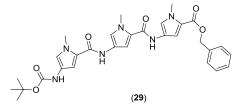
To a solid sample of (27) (0.150 g; 0.33 mmol) was added 4 M HCl in dioxane (4 mL; 16 mmol). The mixture was stirred for 2 hours at room temperature. Reaction completion was confirmed by analytical RPHPLC. The product was precipitated by addition of hexane (40 mL) and isolated by centrifugation and removal of the supernatant. The product was washed twice more with hexane, then suspended in water (5 mL) and lyophilised to dryness, yielding (28) as red solid (0.127 g; 99 %).

<sup>1</sup>H NMR (500 MHz; *d*<sub>6</sub>-DMSO) δ 3.86 (s, 3 H), 3.89 (s, 3 H), 5.25 (s, 2 H), 6.99 (d, *J* = 1.9 Hz, 1 H), 7.03 (d, *J* = 1.9 Hz, 1 H), 7.12 (d, *J* = 1.8 Hz, 1 H), 7.33-7.45 (m, 5 H), 7.50 (d, *J* = 1.9 Hz, 1 H), 10.14 (s, 1 H), 10.19 (s, 2 H).

<sup>13</sup>C NMR (75 MHz; *d*<sub>6</sub>-DMSO) δ 36.2, 36.5, 64.9, 107.2, 108.6, 113.0, 118.5, 121.0, 121.7, 122.6, 124.5, 127.8 (2C), 127.9, 128.5 (2C), 136.5, 157.6, 160.0.

HRMS (ESI +ve mode) 353.1614 calculated for  $C_{19}H_{21}N_4O_3^+$  [M+H]+ found 353.1609.

Benzyl 4-(4-((tert-butoxycarbonyl)amino)-1-methyl-1H-pyrrole-2-carboxamido)-1-methyl-1H-pyrrole-2-carboxylate (29):



To a solution of Boc-Py-OH (**2**) (0.048 g; 0.2 mmol) and HATU (0.076 g; 0.2 mmol) in DMF (peptide grade; 4 mL) was added DIEA (200 µL; 1.2 mmol). The mixture was stirred at room temperature for 30 minutes. A separate solution of (**28**) (0.080 g; 0.21mmol) with DIEA (100 µL; 0.57 mmol) in DMF (peptide grade; 3 mL) was then added and the reaction stirred at room temperature overnight. Reaction was confirmed complete by analytical RPHPLC as indicated by the disappearance of starting material. The product was precipitated by addition of acidified water (pH 3; 40 mL) and isolated by centrifugation and removal of the supernatant. The crude material was washed three times by suspension in 0.25 M HCl followed by centrifugation to remove the supernatant. The residual solid was suspended in water and lyophilised to dryness, giving the product as a pale brown solid (0.104 g; 90 %), which was used without further purification.

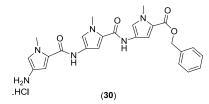
<sup>1</sup>H NMR (400 MHz; *d*<sub>6</sub>-DMSO) δ 1.44 (s, 9H, *t*-Bu), 3.80 (s, 3H, NC*H*<sub>3</sub>), 3.83 (s, 3H, NC*H*<sub>3</sub>), 3.84 (s, 3H, NC*H*<sub>3</sub>), 5.24 (s, 2H, C*H*<sub>2</sub>Ph), 6.82 (s, 1H), 6.88 (s, 1H), 6.96 (d, *J* = 2 Hz, 1 H), 7.06 (d, *J* = 1.6 Hz, 1H), 7.20 (d, *J* = 1.6 Hz, 1H), 7.30-7.45 (m, 5H, benzyl protons), 7.47 (d, *J* = 1.6 Hz, 1 H), 9.06 (s, 1H), 9.84 (s, 1H), 9.88 (s, 1H).

<sup>13</sup>C NMR (100 MHz; *d*<sub>6</sub>-DMSO) δ 28.2 (3C), 36.0 (2C), 36.1, 64.8, 78.4, 103.8, 104.8, 108.6, 117.0, 118.4, 118.5, 120.9, 122.3 (2C), 122.4, 122.8, 123.1, 127.8 (2C), 127.9, 128.4 (2C), 136.6, 152.9, 158.4, 158.5, 160.1.

HRMS (ESI +ve mode) 575.2618 calculated for  $C_{30}H_{35}N_6O_6^+$  [M+H]<sup>+</sup>, found 575.2623.

 Benzyl
 4-(4-(4-amino-1-methyl-1H-pyrrole-2-carboxamido)-1-methyl-1H-pyrrole-2-carboxylate

 hydrochloride (30):



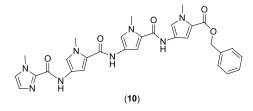
To a solid sample of (**29**) (0.2 g; 0.35 mmol) was added 4M HCl in 1,4-dioxane (10 ml; 40 mmol). The mixture was stirred at room temperature for 3 hours. Complete reaction was confirmed by TLC as indicated by the absence of starting material. The product was precipitated by addition of ether (30 mL) and isolated by centrifugation and removal of the supernatant. The crude mixture was washed twice more by sonication in ether, then centrifugation and removal of the supernatant. The resulting solid was suspended in water (15 mL) and lyophilised to dryness, giving the pure product as a red solid (0.179 g; quantitative).

<sup>1</sup>H NMR (300 MHz; *d*<sub>6</sub>-DMSO) δ 3.83 (s, 6H, 2 x NMe), 3.88 (s, 3H, NMe), 5.23 (s, 2H, C*H*<sub>2</sub>Ph), 6.96 (s, 1H), 7.07(s, 1H), 7.10 (s, 1H), 7.24 (s, 1H), 7.31-7.45 (m, 5H, benzyl protons), 7.47 (s, 1H), 9.94 (s, 1H), 10.11 (s, *br*, 4H).

<sup>13</sup>C NMR (100 MHz; *d*<sub>6</sub>-DMSO) δ 36.1, 36.2, 36.6, 64.8, 104.8, 107.2, 108.6, 113.0, 118.4, 118.7, 121.0, 121.6, 121.8, 122.5, 123.0, 124.8, 127.8 (2C), 127.9, 128.5 (2C), 136.5, 157.7, 158.4, 160.1.

HRMS (ESI +ve mode) 475.2094 calculated for  $C_{25}H_{27}N_6O_4^+$  [M+H]<sup>+</sup>, found 475.2098.

Benzyl 1-methyl-4-(1-methyl-4-(1-methyl-1H-imidazole-2-carboxamido)-1H-pyrrole-2-carboxamido)-1H-pyrrole-2-carboxylate (10):



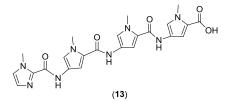
To a solution of (4) (0.095 g; 0.75 mmol) with HOAt (0.102 g; 0.75 mmol) in DMF (10 mL) was added DCC (0.146 g; 0.71 mmol) and the mixture was stirred at room temperature for 2 hours. The solid DCU was then removed by filtration and the filtrate added to a stirred solution of (**30**) (0.394 g; 0.77 mmol) and DIEA (700  $\mu$ L; 4 mmol) in DMF (peptide grade; 10 mL) and the reaction was stirred at room temperature overnight. Complete reaction was confirmed by analytical RPHPLC. The product was precipitated with acidified water (pH 3; 80 mL) and isolated by centrifugation and removal of the supernatant. The product was washed by adding portions of aqueous HCl (0.25 M; 30 mL) and agitating before centrifugation and removal of the supernatant. The resulting solid was suspended in water (5 mL) and lyophilised to dryness, giving the product as an off-white solid (0.380 g; 92 %).

<sup>1</sup>H NMR (400 MHz; *d*<sub>6</sub>-DMSO) δ 3.91 (s, 3 H), 3.91 (s, 3 H), 3.92 (s, 3 H), 4.06 (s, 3 H), 5.31 (s, 2 H), 7.03 (d, *J* = 1.9 Hz, 1 H), 7.14 (d, *J* = 1.8 Hz, 1 H), 7.20 (d, *J* = 0.8 Hz, 1 H), 7.22 (d, *J* = 1.8 Hz, 1 H), 7.29 (d, *J* = 1.7 Hz, 1 H), 7.36 (d, *J* = 1.7 Hz, 1 H), 7.4-7.52 (m, 6 H), 7.54 (d, *J* = 1.9 Hz, 1 H), 9.97 (s, 1 H), 10.02 (s, 1 H), 10.58 (s, 1 H).

<sup>13</sup>C NMR (125 MHz; *d*<sub>6</sub>-DMSO) δ 35.2, 36.1 (3C), 64.8, 104.8, 104.9, 108.6, 118.4, 118.6, 118.7, 121.0, 121.2, 122.2, 122.4, 123.1 (2C), 126.0, 126.3, 127.8 (2C), 127.9, 128.5 (2C), 136.6, 138.5, 155.3, 158.4, 158.4, 160.1.

HRMS (ESI +ve mode) 583.2417 calculated for  $C_{30}H_{31}N_8O_5^+$  [M+H]<sup>+</sup> found 583.2419.

1-methyl-4-(1-methyl-4-(1-methyl-1H-imidazole-2-carboxamido)-1H-pyrrole-2-carboxamido)-1H-pyrrole-2-carboxylic acid (13):



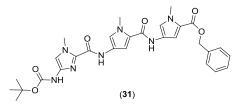
To a solution of (10) (0.095 g; 0.16 mmol) in methanol (60 mL) was added palladium hydroxide on carbon (0.029 g; 20 wt. % loading) and aqueous HCl (0.5 M; 0.55 mL; 1.7 equivalents). The mixture was shaken at 50 PSI pressure of hydrogen for 5 hours, and deemed complete by analytical RPHPLC as indicated by the consumption of starting material. The solution was filtered through Celite and methanol removed under vacuum and the resulting residue was suspended in water (2 mL) and lyophilised to dryness, giving the product as an off-white solid (0.079 g; 99 %), which was used without any further purification.

<sup>1</sup>H NMR (400 MHz; *d*<sub>6</sub>-DMSO) δ 3.83 (s, 3 H), 3.85 (s, 3 H), 3.87 (s, 3 H), 4.00 (s, 3 H), 6.85 (d, *J* = 1.6 Hz, 1 H), 7.07 (d, *J* = 1.6 Hz, 1 H), 7.15 (s, 1 H), 7.16 (d, *J* = 1.6 Hz, 1 H), 7.24 (d, *J* = 1.6 Hz, 1 H), 7.31 (d, *J* = 1.6 Hz, 1 H), 7.42 (d, *J* = 1.6 Hz, 1 H), 7.46 (s, 1 H), 9.89 (s, 1 H), 9.96 (s, 1 H), 10.53 (s, 1 H).

<sup>13</sup>C NMR (125 MHz; *d*<sub>6</sub>-DMSO) δ 35.2, 36.0, 36.1 (2C), 104.8, 104.9, 108.4, 118.5, 118.7, 119.5, 120.2, 121.2, 122.2, 122.6, 122.7, 123.1, 125.9, 126.3, 138.5, 155.3, 158.4 (2C), 161.9.

HRMS (ESI +ve mode) 493.1948 calculated for  $C_{23}H_{25}N_8O_5^+$  [M+H]<sup>+</sup> found 493.1939.

Benzyl 4-(4-((tert-butoxycarbonyl)amino)-1-methyl-1H-imidazole-2-carboxamido)-1-methyl-1H-pyrrole-2-carboxylate (31):



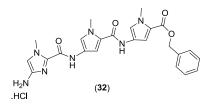
To a solution of Boc-Im-OH (**3**) (0.096 g; 0.39 mmol) and HOAt (0.054 g; 0.39 mmol) in DMF (peptide grade; 2.5 mL) was added DCC (0.078 g; 0.378 mmol). The mixture was stirred for 2 hours at room temperature. The solid DCU was removed by filtration and the filtrate added to a second solution of (**28**) (0.155 g; 0.40 mmol) in DMF (peptide grade; 2.5 mL) with DIEA (320  $\mu$ L; 1.8 mmol). The reaction was stirred for 5 hours at room temperature. Complete reaction was confirmed by analytical RPHPLC and the product precipitated with acidified water (pH 3; 35 mL) and collected by centrifugation and removal of the supernatant. The crude material was washed twice more with acidified water, then suspended in water and lyophilised to dryness. The crude material was isolated as a light brown solid (0.198 g; 91 %) and was of sufficient purity. An analytical sample was acquired by silica gel column chromatography (50 % EtOAc in hexane) as a pale orange solid.

<sup>1</sup>H NMR (300 MHz; *d*<sub>6</sub>-DMSO) δ 1.45 (s, 9 H), 3.85 (s, 3 H), 3.85 (s, 3 H), 3.94 (s, 3 H), 5.25 (s, 2 H), 6.97 (d, *J* = 1.7 Hz, 1 H), 7.16 (d, *J* = 1.7 Hz, 1 H), 7.21 (s, 1 H), 7.27 (d, 1.5 Hz, 1 H), 7.30-7.46 (m, 5 H), 7.49 (d, *J* = 1.5 Hz, 1 H), 9.33 (s, 1 H), 9.92 (s, 1 H), 9.97 (s, 1 H).

<sup>13</sup>C NMR (75 MHz; *d*<sub>6</sub>-DMSO) δ 28.1 (3C), 34.9, 36.2 (2C), 64.9, 79.0, 104.9, 108.6, 113.6, 118.4, 118.8, 121.0, 121.3, 122.7, 123.0, 127.8 (2C), 128.0, 128.5 (2C), 134.1, 136.4, 136.6, 153.0, 155.8, 158.4, 160.1.

HRMS (ESI +ve mode) 576.2571 calculated for  $C_{29}H_{34}N_7O_6^+$  [M+H]<sup>+</sup> found 576.2598.

Benzyl 4-(4-(4-amino-1-methyl-1H-imidazole-2-carboxamido)-1-methyl-1H-pyrrole-2-carboxamido)-1-methyl-1H-pyrrole-2-carboxylate hydrochloride (32):



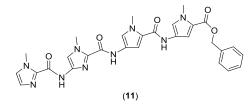
To a solid sample of (**31**) (0.248 g; 0.35 mmol) was added 4M HCl in 1,4-dioxane (5 ml; 20 mmol). The mixture was stirred at room temperature overnight. The crude product was precipitated with the addition of hexane (40 mL) and isolated by centrifugation and removal of the supernatant. The crude material was washed twice more with hexane before being suspended in water (5 mL) and lyophilised to dryness, giving the product as a pale red solid (0.217 g; 98 %).

<sup>1</sup>H NMR (500 MHz; *d*<sub>6</sub>-DMSO) δ 3.86 (s, 3 H), 3.86 (s, 3 H), 3.97 (s, 3H), 5.26 (s, 2 H), 6.98 (d, *J* = 2.0 Hz, 1 H), 7.12 (s, 1 H), 7.18 (d, *J* = 1.9 Hz, 1 H), 7.29 (d, *J* = 1.8 Hz, 1 H), 7.34-7.46 (m, 5 H), 7.49 (d, *J* = 1.9 Hz, 1 H), 9.95 (s, 1 H), 10.31 (s, 1 H).

<sup>13</sup>C NMR (125 MHz; *d*<sub>6</sub>-DMSO) δ 35.3, 36.1, 36.2, 64.8, 105.0, 108.6, 114.0, 118.4, 119.0, 121.0, 121.2, 122.7, 123.0, 127.8 (2C), 127.9, 128.5 (2C), 134.3, 135.3, 136.6, 155.1, 158.3, 160.1.

HRMS (ESI+) 476.2046 calculated for C<sub>24</sub>H<sub>26</sub>N<sub>7</sub>O<sub>4</sub><sup>+</sup> [M+H]<sup>+</sup> found 476.2048.

Benzyl 1-methyl-4-(1-methyl-4-(1-methyl-1H-imidazole-2-carboxamido)-1H-imidazole-2-carboxamido)-1H-pyrrole-2-carboxylate (11):



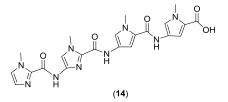
To a solution of (4) (0.025 g; 0.20 mmol) with HOAt (0.027 g; 0.20 mmol) in DMF (2 mL) was added DCC (0.039 g; 0.19 mmol). The mixture was shaken for 2 hours at room temperature. The solid DCU was then removed by filtration and the filtrate added to a stirred solution of (32) (0.120 g; 0.23 mmol) and DIEA (500  $\mu$ L; 3 mmol) in DMF (peptide grade; 2 mL). The mixture was stirred at room temperature for two hours. The reaction was confirmed complete by analytical RPHPLC. The product was precipitated by the addition of acidified water (pH 3; 40 mL) and collected by centrifugation and removal of the supernatant. The crude material was washed twice more by addition of acidified water and centrifugation and removal of the supernatant. The resulting solid was suspended in water (5 mL) and lyophilised to dryness, giving (11) as a brown solid (0.100 g; 90 %) which was used without further purification.

<sup>1</sup>H NMR (500 MHz; *d*<sub>6</sub>-DMSO) δ 3.86 (s, 3 H), 3.86 (s, 3 H) 4.02 (s, 6 H), 5.26 (s, 2 H), 6.99 (d, *J* = 1.9 Hz, 1 H), 7.09 (d, *J* = 1.0 Hz, 1 H), 7.18 (d, *J* = 1.8 Hz, 1 H), 7.29 (d, *J* = 1.8 Hz, 1 H), 7.34-7.46 (m, 5 H), 7.46 (d, *J* = 0.7 Hz, 1 H), 7.50 (d, *J* = 1.9 Hz, 1 H), 7.58 (s, 1 H), 9.76 (s, 1 H), 9.96 (s, 1 H), 10.38 (s, 1 H).

<sup>13</sup>C NMR (75 MHz; *d*<sub>6</sub>-DMSO) δ 35.1, 35.2, 36.1 (2C), 64.8, 104.9, 108.6, 114.0, 118.4, 118.8, 121.0, 121.4, 122.7, 123.0, 127.1, 127.7, 127.8 (2C), 127.9, 128.5 (2C), 134.4, 134.6, 136.6, 137.8, 155.5, 155.7, 158.4, 160.1.

HRMS (ESI+) 584.2370 calculated for  $C_{29}H_{30}N_9O_5^+$  [M+H]<sup>+</sup> found 584.2393.

1-methyl-4-(1-methyl-4-(1-methyl-1H-imidazole-2-carboxamido)-1H-imidazole-2-carboxamido)-1H-pyrrole-2-carboxamido)-1H-pyrrole-2-carboxylic acid (14):



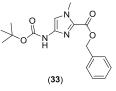
To a solution of (11) (0.140 g; 0.24 mmol) in methanol (60 mL) was added palladium hydroxide on carbon (0.040 g; 20 wt. % loading) and aqueous HCl (1 M; 1 mL; 4.2 equivalents). The mixture was shaken at 50 PSI pressure of hydrogen overnight and confirmed complete by analytical RPHPLC. The solution was filtered through Celite and methanol removed under vacuum. The resulting solid was suspended in water (2 mL) and lyophilised to dryness, giving the product as an off-white solid (0.111 g; 94 %), which was used without any further purification.

<sup>1</sup>H NMR (300 MHz;  $d_6$ -DMSO)  $\delta$  3.81 (s, 3H), 3.84 (s, 3 H), 4.00 (s, 6 H), 6.84 (d, J = 1.9 Hz, 1 H), 7.10 (s, 1 H), 7.15 (d, J = 1.7 Hz, 1 H), 7.26 (d, J = 1.5 Hz, 1 H), 7.41 (d, J = 1.8 Hz, 1 H), 7.46 (s, 1 H), 7.57 (s, 1 H), 9.82 (s, 1 H), 9.92 (s, 1 H), 10.36 (s, 1 H).

<sup>13</sup>C NMR (125 MHz; *d*<sub>6</sub>-DMSO) δ 35.1, 35.2, 36.1 (2C), 104.8, 108.4, 114.1, 118.7, 119.5, 120.3, 121.4, 122.6, 122.9, 127.1, 127.4, 134.4, 134.6, 137.7, 155.5, 155.6, 158.4, 161.9.

HRMS (ESI +ve mode) 494.1900 calculated for  $C_{22}H_{24}N_9O_5^+$  [M+H]<sup>+</sup> found 494.1893.

#### Benzyl 4-((tert-butoxycarbonyl)amino)-1-methyl-1H-imidazole-2-carboxylate (33):



To a stirred solution of Boc-Im-OH (**3**) (0.350 g; 1.45 mmol) in ethanol/water (3:1 v/v, 7 mL) was added a solution of caesium carbonate (0.236 g; 0.725 mmol) in water (2 mL) drop-wise. The reaction mixture was then stirred at room temperature for 2 hours. The reaction mixture was then filtered and the solvent removed under vacuum, then distilled azeotropically to dryness with ethanol, giving the caesium salt of (**3**) as a pale yellow solid. The crude solid was then dissolved in DMF (3.5 mL; dry) and benzyl bromide (180  $\mu$ L; 1.5 mmol) was added dropwise at room temperature with constant stirring. The temperature was raised to 40 °C and stirred overnight. The reaction was quenched by the addition of ice water (45 mL) and the crude solid isolated by centrifugation and removal of the supernatant. Any remaining water was removed azeotropically with ethanol to yield the crude product as a brown solid (0.401 g; 84 %). An analytical sample was obtained as a pale orange crystalline solid by re-crystallisation from ethanol/hexane.

<sup>1</sup>H NMR (300 MHz; CDCl<sub>3</sub>) δ 1.47 (s, 9 H), 3.92 (s, 3 H), 5.31 (s, 2 H), 7.24 (s, 1H), 7.27-7.40 (m, 5 H), 7.52 (s, 1 H).

<sup>13</sup>C NMR (75 MHz; CDCl<sub>3</sub>) δ 28.2 (3C), 35.9, 66.8, 80.9, 113.4, 128.3 (2C), 128.5 (2C), 131.1, 135.4, 138.0, 152.7, 158.4, 162.7.

HRMS (ESI +ve mode) 332.1610 calculated for  $C_{17}H_{22}N_3O_4^+$  [M+H]<sup>+</sup> found 332.1610.

Benzyl 4-amino-1-methyl-1H-imidazole-2-carboxylate hydrochloride (34):



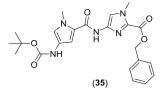
To a solid sample of compound (**33**) (0.200 g; 0.6 mmol) was added 4M HCl in dioxane (2 mL; 8 mmol). The reaction mixture was stirred at room temperature for 2 hours. The reaction was deemed complete by RPHPLC analysis. The product was precipitated by addition of hexane and isolated by centrifugation and removal of the supernatant. The product was washed twice more with hexane, then suspended in water (2 mL) and lyophilised to dryness, yielding (**34**) as an orange solid (0.162 g; quantitative).

<sup>1</sup>H NMR (300 MHz; *d*<sub>4</sub>-Methanol) δ 4.06 (s, 3H), 5.44 (s, 2 H), 6.96 (s, 1 H), 7.36-7.52 (m, 5 H).

<sup>13</sup>C NMR (75 MHz; *d*<sub>4</sub>-Methanol) δ 37.6, 69.4, 111.1, 127.5, 129.8 (2C), 130.0 (3C), 136.2, 140.7, 155.5.

HRMS (ESI +ve mode) 232.1086 calculated for  $C_{12}H_{14}N_3O_2^+$  [M+H]<sup>+</sup> found 232.1085.

Benzyl 4-(4-((tert-butoxycarbonyl)amino)-1-methyl-1H-pyrrole-2-carboxamido)-1-methyl-1H-imidazole-2-carboxylate (35):



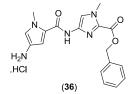
To a solution of (2) (0.284 g; 1.18 mmol) in DMF (5mL; peptide grade) was added DCC (0.227 g; 1.1 mmol) and HOAt (0.161 g; 1.18 mmol). The reaction mixture was stirred at room temperature for 2 hours. The solid DCU was removed by filtration and the filtrate added to a solution of (34) (0.210 g; 0.79 mmol) with DIEA (2.7 mL; 15.5 mmol) in DMF (5 mL; peptide grade). The reaction was heated in a microwave at 60 °C for 1 hour. The crude product was precipitated by the addition of acidified water (pH 3; 50 mL) and isolated by centrifugation and decanting of the supernatant. The solid material was washed twice with portions of acidified water before being suspended in water (5 mL) and lyophilised to dryness, to yield the crude material as a yellow/brown solid. The product was purified by silica gel chromatography (30 % ethyl acetate in hexane) to give the pure material as a pale yellow solid (0.241 g; 68%).

<sup>1</sup>H NMR (300 MHz; *d*<sub>6</sub>-DMSO) δ 1.43 (s, 9 H), 3.80 (s, 3 H), 3.94 (s, 3 H), 5.29 (s, 2 H), 6.92 (s, 1 H), 6.98 (s, 1 H), 7.32-7.50 (m, 5 H), 7.66 (s, 1 H), 9.04 (s, 1 H), 10.63 (s, 1 H).

<sup>13</sup>C NMR (75 MHz; *d*<sub>6</sub>-DMSO) δ 28.1 (3C), 35.4, 36.2, 66.1, 78.2, 105.1, 115.6, 118.2, 121.7, 122.3, 128.3, 128.5 (2C), 128.6 (2C), 130.5, 135.7, 137.9, 152.8, 158.2, 158.6.

HRMS (ESI +ve mode) 454.2090 calculated for  $C_{23}H_{28}N_5O_5^+$  [M+H]<sup>+</sup> found 454.2089.

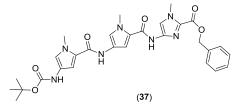
#### Benzyl 4-(4-amino-1-methyl-1H-pyrrole-2-carboxamido)-1-methyl-1H-imidazole-2-carboxylate hydrochloride (36):



To a solid sample of compound (**35**) (0.200 g; 0.18 mmol) was added 4 M HCl in dioxane (4 mL; 16 mmol). The mixture was stirred at room temperature for 2 hours, after which time the formation of a cloudy white precipitate was observed. The product was collected by addition of hexane and isolated by centrifugation and removal of the supernatant. The crude material was washed twice more with hexane. The resulting solid was suspended in water (4 mL) and lyophilised, giving (**36**) as a very pale yellow solid (0.171.8 g; quantitative).

<sup>1</sup>H NMR (300 MHz; *d*<sub>6</sub>-DMSO) δ 3.87 (s, 3 H), 3.94 (s, 3 H), 5.29 (s, 2 H), 7.14 (d, *J* = 1.5 Hz, 1 H), 7.18 (d, *J* = 1.7 Hz, 1 H), 7.32-7.48 (m, 5 H), 7.69 (s, 1 H), 10.19 (s, 3 H), 10.88 (s, 1 H).

<sup>13</sup>C NMR (75 MHz;  $d_6$ -DMSO)  $\delta$  35.5, 36.8, 66.2, 108.3, 113.2, 115.7, 122.5, 123.6, 128.3, 128.5 (2C), 128.6 (2C), 130.5, 135.6, 137.3, 157.9, 158.0. HRMS (ESI+) 354.1566 calculated for C<sub>18</sub>H<sub>20</sub>N<sub>5</sub>O<sub>3</sub><sup>+</sup> [M+H]<sup>+</sup> found 354.1564. Benzyl 4-(4-((tert-butoxycarbonyl)amino)-1-methyl-1H-pyrrole-2-carboxamido)-1-methyl-1Himidazole-2-carboxylate (37):



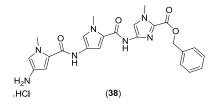
To a solution of (2) (0.035 g; 0.145 mmol) and HATU (0.055 g; 0.145 mmol) in DMF (1 mL; peptide grade) was added DIEA (400  $\mu$ L; 2.3 mmol). The reaction mixture was stirred at room temperature for 30 minutes. The activated ester was added to a stirred solution of (36) (0.058 g; 0.15 mmol) in DMF (1 mL; peptide grade) and the mixture stirred at room temperature for 2 hours. Reaction was deemed complete by analytical RPHPLC as indicated by absence of starting material, and the product precipitated by the addition of acidified water (pH 3; 35 mL) and isolated by centrifugation and decanting off the supernatant. The solid material was washed twice more with portions of acidified water before being suspended in water (5 mL) and lyophilised to dryness, giving the crude product as a light brown solid (0.078 g; 94 %) which was used without further purification. An analytical sample was acquired by silica gel column chromatography (40 % EtOAc in hexane) as an off-white powder.

<sup>1</sup>H NMR (300 MHz; *d*<sub>6</sub>-DMSO) δ 1.44 (s, 9 H), 3.79 (s, 3 H), 3.83 (s, 3 H), 3.94 (s, 3 H), 5.30 (s, 2 H), 6.83 (s, 1 H), 6.88 (s, 1 H), 7.07 (d, *J* = 1.5 Hz, 1 H), 7.32 (d, *J* = 1.4 Hz, 1 H), 7.34-7.47 (m, 5 H), 7.68 (s, 1 H), 9.07 (s, 1 H), 9.84 (s, 1 H), 10.69 (s, 1 H).

<sup>13</sup>C NMR (75 MHz; *d*<sub>6</sub>-DMSO) δ 28.2 (3C), 35.5, 36.0, 36.2, 66.1, 78.2, 103.9, 105.8, 115.7, 117.0, 119.7, 121.6, 122.2, 122.3, 122.7, 128.3, 128.5 (2C), 128.6 (2C), 130.5, 135.7, 137.9, 152.8, 158.2, 158.5, 158.7.

HRMS (ESI +ve mode) 576.2571 calculated for  $C_{29}H_{34}N_7O_6^+$  [M+H]<sup>+</sup> found 576.2598.

Benzyl 4-(4-(4-amino-1-methyl-1H-pyrrole-2-carboxamido)-1-methyl-1H-pyrrole-2-carboxamido)-1-methyl-1H-imidazole-2-carboxylate hydrochloride (38):



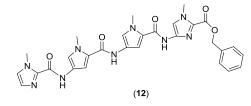
To a solid sample of (**37**) (0.200 g; 0.35 mmol) was added 4 M HCl in dioxane (4 mL; 16 mmol) and the mixture was stirred at room temperature for 2 hours. Reaction was confirmed to be complete by analytical RPHPLC as indicated by absence of starting material. The product was precipitated with hexane then isolated by centrifugation and removal of the supernatant. The crude material was washed twice more with hexane, then suspended in water (5 mL) and lyophilised to dryness, giving the product as an off-white solid (0.179 g; quantitative).

<sup>1</sup>H NMR (400 MHz;  $d_6$ -DMSO)  $\delta$  3.90 (s, 3 H), 3.93 (s, 3 H), 4.00 (s, 3 H), 5.35 (s, 2 H), 7.01 (d, J = 1.9 Hz, 1 H), 7.13 (d, J = 2.5 Hz, 1 H), 7.14 (d, J = 2.1 Hz, 1 H), 7.39 (d, J = 1.6 Hz, 1 H), 7.41-7.52 (m, 5 H), 7.73 (s, 1 H), 9.88 (s br, 2 H), 10.08 (s, 1 H), 10.75 (s, 1 H).

<sup>13</sup>C NMR (75 MHz; *d*<sub>6</sub>-DMSO) δ 35.5, 36.3, 36.6, 66.1, 105.7, 107.0, 112.9, 115.7, 119.8, 121.5, 121.6, 121.8, 124.9, 128.3, 128.5 (2C), 128.6 (2C), 130.5, 135.7, 137.9, 157.7, 158.2, 158.6.

HRMS (ESI +ve mode) 476.2046 calculated for C<sub>24</sub>H<sub>26</sub>N<sub>7</sub>O<sub>4</sub><sup>+</sup> [M+H]<sup>+</sup>, found 476.2058.

Benzyl 1-methyl-4-(1-methyl-4-(1-methyl-1H-imidazole-2-carboxamido)-1H-pyrrole-2-carboxamido)-1H-pyrrole-2-carboxamido)-1H-imidazole-2-carboxylate (12):



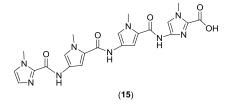
To a solution of (4) (0.053 g; 0.42 mmol) and HOAt (0.057 g; 0.42 mmol) in DMF (dry; 6 mL) was added DCC (0.085 g; 0.41 mmol) and the reaction mixture was stirred at room temperature for 2 hours. The solid DCU was then removed by filtration and the filtrate added to a stirred solution of (38) (0.220 g; 0.43 mmol) and DIEA (0.75 mL; 4.3 mmol) in DMF (dry; 6 mL). The reaction mixture was stirred at room temperature overnight. The reaction was confirmed to be complete by analytical RPHPLC as indicated by the absence of starting material. The crude product was precipitated by the addition of acidified water (pH 3; 40 mL) and was collected by centrifugation and removal of the supernatant. The crude solid was washed twice more with acidified water before being suspended in water (5 mL) and lyophilised to dryness, giving the product as a pale brown powder (0.225 g; 94 %) which was used without further purification. An analytical specimen was obtained by semi-preparative HPLC purification as a fluffy, off-white solid.

<sup>1</sup>H NMR (300 MHz; *d*<sub>6</sub>-DMSO) δ 3.89 (s, 6 H), 4.00 (s, 3 H), 4.05 (s, 3 H), 5.35 (s, 2 H), 7.14 (d, *J* = 1.7 Hz, 1 H), 7.20-7.21 (m, 2 H), 7.35 (d, *J* = 1.6 Hz, 1 H), 7.40 (d, *J* = 1.7 Hz, 1 H), 7.42-7.51 (m, 6 H), 7.74 (s, 1 H), 10.01 (s, 1 H), 10.60 (s, 1 H), 10.76 (s, 1 H).

<sup>13</sup>C NMR (125 MHz; *d*<sub>6</sub>-DMSO) δ 35.3, 35.5, 36.1, 36.3, 66.1, 104.9, 105.8, 115.7, 118.7, 119.8, 121.1, 121.7, 122.1, 123.1, 125.4, 126.2, 128.3, 128.5 (2C), 128.6 (2C), 130.5, 135.7, 137.9, 138.3, 154.8, 158.2, 158.5, 158.7.

HRMS (ESI +ve mode) 584.2370 calculated for  $C_{29}H_{30}N_9O_5^+$  [M+H]<sup>+</sup>, found 584.2398.

1-methyl-4-(1-methyl-4-(1-methyl-1H-imidazole-2-carboxamido)-1H-pyrrole-2-carboxamido)-1H-pyrrole-2-carboxamido)-1H-imidazole-2-carboxylic acid (15):



To a solution of (12) (0.100 g; 0.17 mmol) in methanol (60 mL) was added palladium hydroxide on carbon (0.030 g; 20 wt. % loading) and aqueous HCl (1 M; 0.3 mL; 1.7 equivalents). The mixture was shaken at 50 PSI pressure of hydrogen for 5 hours. Analytical HPLC showed the reaction stalled at approximately 50 % conversion. A further addition of palladium hydroxide on carbon (0.030 g; 20 wt. % loading) and aqueous HCl (1M; 0.5 mL; 2.9 equivalents) was made and the pressure returned to 50 psi and the reaction left overnight. The solution was filtered through Celite and methanol removed under vacuum to give a solid which was suspended in water (2 mL) and lyophilised to dryness, giving the product as a yellow/brown solid (0.084 g; 99

%), which was used without any further purification. Compound (15) was found to readily decarboxylate when stored at RT, however the compound was stable when stored at -20 °C after lyophilisation.

<sup>1</sup>H NMR (500 MHz;  $d_6$ -DMSO)  $\delta$  3.86 (s, 3 H), 3.87 (s, 3 H), 3.94 (s, 3 H), 4.01 (s, 3 H), 7.07 (d, J = 0.8 Hz, 1 H), 7.11 (d, J = 1.8 Hz, 1 H), 7.18 (d, J = 1.8 Hz, 1 H), 7.31 (d, J = 1.7 Hz, 1 H), 7.37 (d, J = 1.6 Hz, 1 H), 7.41 (s, 1 H), 7.63 (s, 1 H), 9.98 (s, 1 H), 10.46 (s, 1 H), 10.65 (s, 1 H).

<sup>13</sup>C NMR (125 MHz; *d*<sub>6</sub>-DMSO) δ 35.1, 35.5, 36.1, 36.2, 105.0, 105.7, 115.1, 118.6, 119.6, 121.3, 121.8, 122.1, 123.0, 126.3, 126.7, 131.7, 137.4, 138.7, 155.9, 158.5, 158.7, 160.0.

HRMS (ESI +ve mode) 494.1900 calculated for  $C_{22}H_{24}N_9O_5^+$  [M+H]<sup>+</sup>, found 494.1876.

#### 5.3 Manual solid phase synthesis protocol

#### 5.3.1 General preparation of resin and coupling protocol for solid phase synthesis of polyamides

Boc- $\beta$ -Alanine-PAM resin (100 mg; 0.026 mmol) was placed in a fritted glass reaction vessel, washed with DMF (5 x 1 min) and then drained. The Boc group was removed using a TFA:phenol:water mixture (92.5:5:2.5) twice (1 x 1 min, and 1 x 5 min) with shaking. The resin was washed with DMF (4 x 1 min), followed by another washing step (DMF for DCC\HOAt and HATU couplings, THF for BTC couplings) for 1 min before the vessel was drained. The activated monomer was added (followed by DIEA if required) and the reaction was mixed resulting in the formation of a slurry. The couplings were conducted for the allotted time (dependant on sequence and coupling method), before being drained and washed with DMF (4 x 1 min).

To ensure complete deprotection of the amine, the resin bound material was treated with 3 consecutive treatments of TPW mixture. The timings for these treatments are dependent upon which monomer the Boc group is located and are as follows:

Boc-pyrrole, GABA and Fmoc-D-Dab(Boc)-OH building blocks – 1 minute, 3 minutes, 3 minutes respectively.

Boc-imidazole building blocks – 1 minute, 3 minutes, 20 minutes for successive rounds of coupling.

#### 5.3.2 General cleavage protocol and isolation of polyamide

Polyamides were cleaved from the resin by placing them into a sealed container with the appropriate cleavage amine and heating to 55 °C for 16 hours. The resin was removed by filtration through cotton wool and washed with 1 mL of DCM. The crude material was precipitated by addition of cold diethyl ether and isolated by centrifugation and decanting off the supernatent. The polyamide was washed twice more by dissolving in DCM (1 ml) and reprecipitating with diethyl ether to yield the crude material.

### 5.3.3 HPLC analysis and purification of polyamides

Analytical HPLC retention times were acquired using 0.1 % TFA in water as buffer A and 0.1 % TFA in acetonitrile as buffer B with a flow rate of 0.5 mL/minute, starting at 15 % B and holding for 5 minutes before increasing to 60 % B over 20 minutes. Unless otherwise stated, polyamides were purified by semi-preparative HPLC with a gradient starting at 17.5 % B and holding for 5 minutes before increasing to 30 % B over 30 minutes and a constant flow rate of 10.6 mL/minute.

#### 5.4 General activation protocols for polyamide synthesis

#### 5.4.1 Activation with HATU

General procedure for 100 mg of  $\beta$ -ala-PAM resin (0.26 meq/g): Monomer (4.0 equivalents; 0.104 mmol) and HATU (3.6 equivalents; 0.3744 mmol; 36 mg) were dissolved in dry DMF (1 mL). Upon addition of DIEA (150  $\mu$ L; 0.9 mmol) the solution was left to activate for 2 minutes, before being added to the resin.

#### 5.4.2 Activation with DCC/HOAt

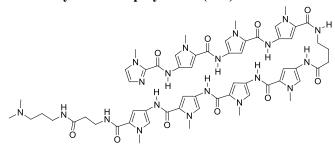
General procedure for 100 mg of  $\beta$ -ala-PAM resin (0.26 meq/g): Monomer (4.0 equivalents; 0.104 mmol) was added to DCC (19.31 mg; 0.0936 mmol; 3.6 equivalents) and HOAt (13.45 mg; 0.0988 mmol; 3.8 equivalents) in dry DMF (1 mL) and activated for 2 hours with constant shaking. The DCU that precipitates is filtered off and the solution added to resin followed by addition of DIEA (150  $\mu$ L; 0.9 mmol).

## 5.4.3 Activation with BTC

General procedure for 100 mg of  $\beta$ -ala-PAM resin (0.26 meq/g): To a solution of monomer (4.0 equivalents; 0.104 mmol) and BTC (9.6 mg; 0.0324 mmol; 1.2 equivalents) in dry THF (1 mL) was added 2,3,5-collidine (50 µL; 0.38 mmol). This resulted in the formation of a white emulsion, which was allowed to mix for 2 minutes. The activated monomer was then added to the resin with DIEA (150 µL; 0.9mmol).<sup>3</sup>

#### 5.5 Synthesis of polyamides 17a-19a through solid phase synthesis

5.5.1 Synthesis of polyamide (17a)



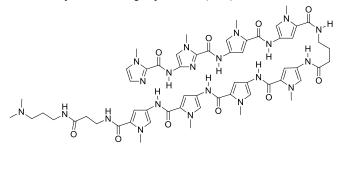
(17a)

Polyamide (**17a**) was synthesised on 50 mg of Boc-β-ala-PAM resin as shown in Scheme S3 using the following reagents and times: (i) TPW; (ii) BocPyOH, HATU, DIEA, 40 minutes; (vi) BocPyOH, HATU, DIEA, 40 minutes; (vii) TPW; (vii) BocPyOH, HATU, DIEA, 40 minutes; (vii) TPW; (viii) BocPyOH, HATU, DIEA, 60 minutes; (ix) TPW; (x) GABA, HATU, DIEA, 60 minutes; (xi) TPW; (xii) BocPyOH, HATU, DIEA, 60 minutes; (xiii) TPW; (xiv) BocPyOH, HATU, DIEA, 90 minutes; (xv) TPW; (xvi) BocPyOH, HATU, DIEA, 120 minutes; (xvii) TPW; (xviii) ImOH, DCC, HOAt, DIEA, 90 minutes. This was cleaved from the resin under standard cleavage conditions (See section 2.3.2, page S22) to give the crude material in 68 % purity. After purification by semi-preparative HPLC the pure material was isolated as a fluffy white solid (1.9 mg; 12 % recovery).

Analytical HPLC  $t_r = 14.4$  minutes.

HRMS (ESI +ve mode) 1221.5819 calculated for  $C_{59}H_{73}N_{20}O_{10}^+$  [M+H]<sup>+</sup>, found 1221.5776.

### 5.5.2 Synthesis of polyamide (18a)



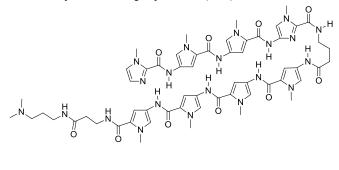
(18a)

Polyamide (**18a**) was synthesised on 50 mg of Boc-β-ala-PAM resin as shown in Scheme S3 using the following reagents and times: (i) TPW; (ii) BocPyOH, HATU, DIEA, 40 minutes; (vi) DocPyOH, HATU, DIEA, 40 minutes; (vii) TPW; (vii) BocPyOH, HATU, DIEA, 40 minutes; (vii) TPW; (viii) BocPyOH, HATU, DIEA, 60 minutes; (ix) TPW; (x) GABA, HATU, DIEA, 60 minutes; (xi) TPW; (xii) BocPyOH, HATU, DIEA, 60 minutes; (xiii) TPW; (xiv) BocPyOH, HATU, DIEA, 90 minutes; (xv) TPW; (xvi) BocImOH, DCC, HOAt, DIEA, 120 minutes; (xvii) TPW; (xviii) ImOH, DCC, HOAt, DIEA, 90 minutes. This was cleaved from the resin under standard cleavage conditions (See section 2.3.2, page S22) to give the crude material in 76 % purity. After purification by semi-preparative HPLC the pure material was isolated as a fluffy white solid (1.6 mg; 10 % recovery).

Analytical HPLC  $t_r = 14.2$  minutes.

HRMS (ESI +ve mode) 1222.5771 calculated for  $C_{59}H_{73}N_{20}O_{10}^+$  [M+H]<sup>+</sup>, found 1222.5826.

### 5.5.3 Synthesis of polyamide (19a)

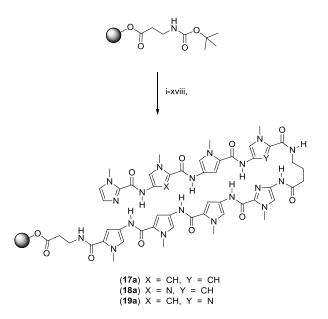




Polyamide (**19a**) was synthesised on 50 mg of Boc-β-ala-PAM resin as shown in Scheme S3 using the following reagents and times: (i) TPW; (ii) BocPyOH, HATU, DIEA, 40 minutes; (iii) TPW; (iv) BocPyOH, HATU, DIEA, 40 minutes; (v) TPW; (vi) BocPyOH, HATU, DIEA, 40 minutes; (vii) TPW; (viii) BocPyOH, HATU, DIEA, 60 minutes; (ix) TPW; (x) GABA, HATU, DIEA, 60 minutes; (xi) TPW; (xii) BocImOH, DCC, HOAt, DIEA, 60 minutes; (xiii) TPW; (xiv) BocPyOH, BTC/Collidine, DIEA, 90 minutes; (xv) TPW; (xvi) BocPyOH, HATU, DIEA, 120 minutes; (xvii) TPW; (xviii) ImOH, DCC, HOAt, DIEA, 90 minutes. This was cleaved from the resin under standard cleavage conditions to give the crude material in 78 % purity. After purification by semi-preparative HPLC the pure material was isolated as a fluffy white solid (1.7 mg; 11 % recovery).

Analytical HPLC  $t_r = 14.6$  minutes.

HRMS (ESI +ve mode) 1222.5771 calculated for  $C_{59}H_{73}N_{20}O_{10}^+$  [M+H]<sup>+</sup>, found 1221.5746.



Scheme S3: Synthesis of PAs 17a-19a. Steps (i)-(xviii) are stated in the respective polyamide experimental.

## 5.5.4 Characterisation of Polyamides 17a-19a

## 5.5.4.1 Crude HPLC data

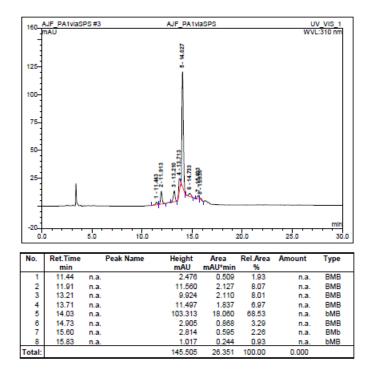


Figure S1: Crude HPL chromatogram of polyamide (17a).

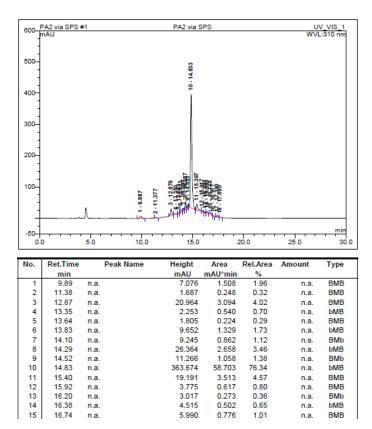


Figure S2: Crude HPL chromatogram of polyamide (18a).

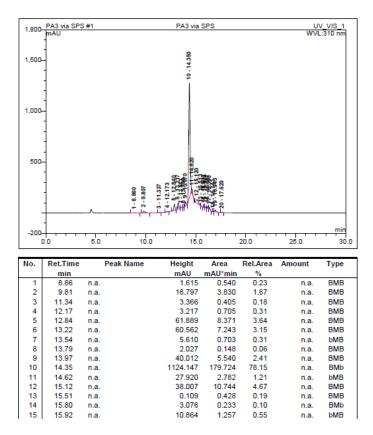


Figure S3: Crude HPL chromatogram of polyamide (19a).

5.5.4.2 Analytical HPLC traces of polyamides purified by semi-preparative HPLC

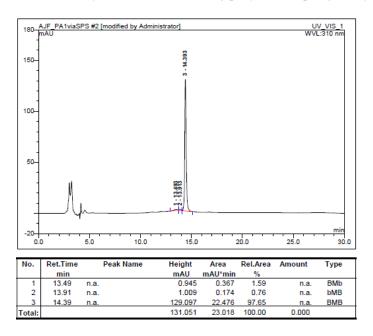


Figure S4: HPL chromatogram of polyamide (17a).

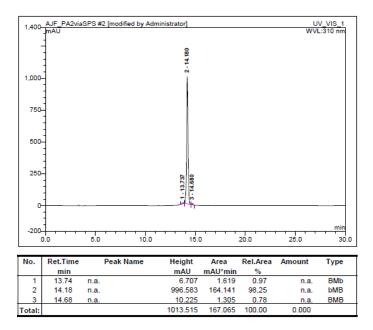


Figure S5: HPL chromatogram of polyamide (18a).

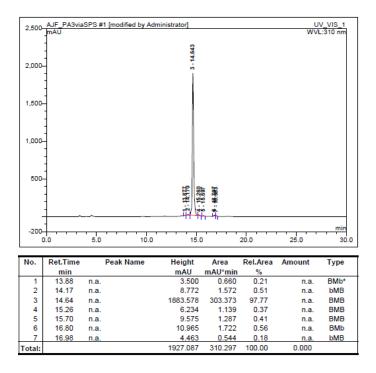


Figure 6: HPL chromatogram of polyamide (19a).

# 5.5.4.3 HRMS data of polyamides 17a-19a purified by semi-preparative HPLC.

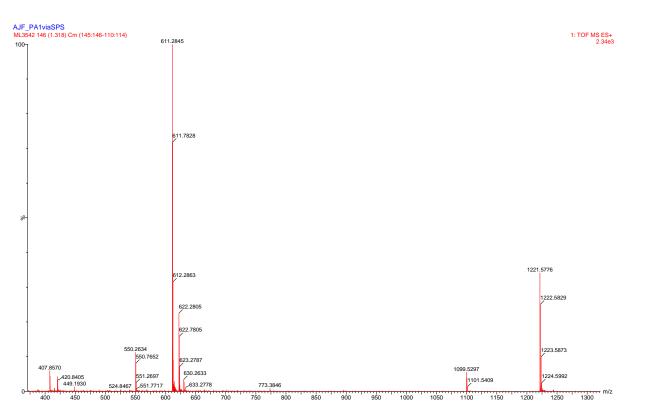
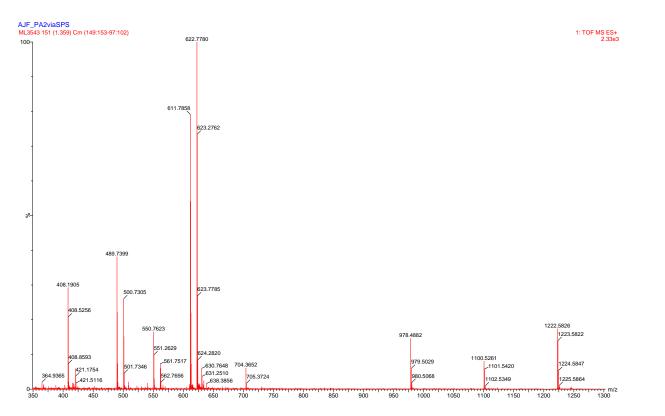
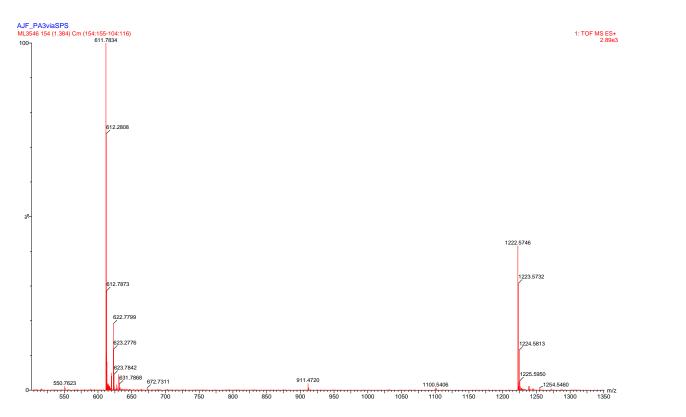


Figure S7: HRMS of polyamide (17a). Key peaks shown are [M+H]<sup>+</sup> at 1221.5776, [M+2H]<sup>2+</sup> at 611.2845, [M+H+Na]<sup>2+</sup> at 622.2805.



**Figure S8:** HRMS of polyamide (**18a**). Key peaks shown are [M+H]<sup>+</sup> at 1222.5826, [M+2H]<sup>2+</sup> at 611.7858, [M+H+Na]<sup>2+</sup> at 622.7780.

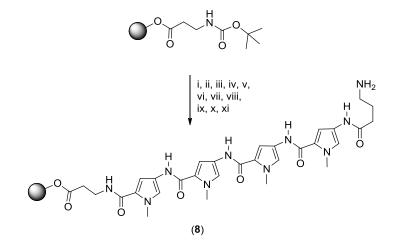


**Figure S9:** HRMS of polyamide (**19a**). Key peaks shown are [M+H]<sup>+</sup> at 1222.5746, [M+2H]<sup>2+</sup> at 611.7834, [M+H+Na]<sup>2+</sup> at 622.7799.

## 5.6 Synthesis of polyamides using a half-half coupling protocol

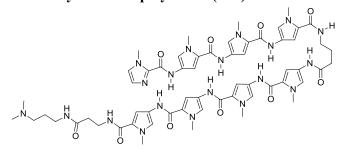
## 5.6.1 Preparation of resin bound tetramer

The first four pyrrole units were coupled to the resin using the protocols described above. Reaction times and conditions are shown in Scheme S4.



**Scheme S4:** Synthesis of polyamide tetramer on Boc- $\beta$ -Ala PAM resin. (i) TPW; (ii) BocPyOH, HATU, DIEA, 40 minutes; (iii) TPW; (iv) BocPyOH, HATU, DIEA, 40 minutes; (v) TPW; (vi) BocPyOH, HATU, DIEA, 40 minutes; (vi) TPW; (viii) BocPyOH, HATU, DIEA, 60 minutes; (ix) TPW; (x) GABA, HATU, DIEA. 60 minutes: (xi) TPW.

5.6.2 Synthesis of polyamide (17b)



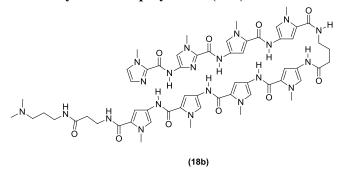
(17b)

Resin-bound tetramer (8) (20 mg; 5.2  $\mu$ mol) was swollen in DMF for 2 hours. The resin was then drained and washed three times with DMF. The tetramer (13) (10 mg; 20.3  $\mu$ mol) was pre-activated with HATU (7 mg; 18.4  $\mu$ mol) and DIEA (50  $\mu$ L; 0.3 mmol) in DMF (600  $\mu$ L) for 30 minutes before being added to the resin, and then shaken for 4 hours. The reaction mixture was then drained from the vessel and the resin washed three times with DMF, then DCM, then twice with methanol before being flushed with nitrogen for 5 minutes. (17b) was cleaved from the resin with  $N^1$ , $N^1$ -dimethylpropane-1,3-diamine (500  $\mu$ L) under standard cleavage conditions with the crude material being 90 % pure by HPLC at 310 nm (Figure S10). This was purified by semipreparative HPLC to give the product as a fluffy white solid (1.9 mg; 30 % recovery).

Analytical HPLC  $t_r = 14.8$  minutes.

HRMS (ESI +ve mode) 1221.5819 calculated for  $C_{59}H_{73}N_{20}O_{10}^+$  [M+H]<sup>+</sup>, found 1221.5808.

## 5.6.3 Synthesis of polyamide (18b)

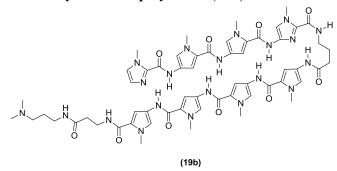


Resin-bound tetramer (8) (20 mg; 5.2 µmol) was swollen in DMF for 2 hours. The resin was then drained and washed three times with DMF. The tetramer (14) (10 mg; 20.3 µmol) was pre-activated with HATU (7 mg; 18.4 µmol) and DIEA (50 µL; 0.3mmol) in DMF (600 µL) for 30 minutes before being added to the resin, and then shaken for 4 hours. The reaction mixture was then drained from the vessel and the resin washed three times with DMF, then DCM, then twice with methanol before being flushed with nitrogen for 5 minutes. Crude (18b) was cleaved from the resin using  $N^1$ , $N^1$ -dimethylpropane-1,3-diamine (500 µL) under standard cleavage conditions and the resulting crude product was found to be 88 % pure by HPLC at 310 nm (Figure S11). This was purified by semi-preparative HPLC to give the product as a fluffy white solid (1.9 mg; 30 % recovery).

Analytical HPLC  $t_r = 14.4$  minutes.

HRMS (ESI +ve mode) 1222.5771 calculated for  $C_{58}H_{72}N_{21}O_{10}^+$  [M+H]<sup>+</sup>, found 1222.5734.

## 5.6.4 Synthesis of polyamide (19b)



Resin-bound tetramer (8) (20 mg; 5.2 µmol) was swollen in DMF for 2 hours. The resin was then drained and washed three times with DMF. The tetramer (15) (15 mg; 30.4 µmol) was pre-activated with DCC (5 mg; 24.2 µmol) and HOAt (4 mg; 30 µmol) in DMF (600 µL) for 2 hours before being filtered through a cotton wool plug and added to the resin with DIEA (50 µL; 0.3 mmol), and then shaken for 4 hours. The reaction mixture was then drained from the vessel and the resin washed three times with DMF, then DCM, then twice with methanol before being flushed with nitrogen for 5 minutes. Crude (19b) was cleaved from the resin with  $N^1$ , $N^1$ -dimethylpropane-1,3-diamine (500 µL) under standard cleavage conditions and the resulting crude product was found to be 85 % pure by HPLC at 310 nm (Figure S12). This was purified by semi-preparative HPLC to give the product as a fluffy white solid (1.7 mg; 28 % recovery).

Analytical HPLC  $t_r = 14.8$  minutes.

HRMS (ESI +ve mode) 1222.5771 calculated for  $C_{58}H_{72}N_{21}O_{10}^+$  [M+H]<sup>+</sup>, found 1222.5726.

## 5.7 Characterisation of half-half polyamides

# 5.7.1 Crude HPLC traces

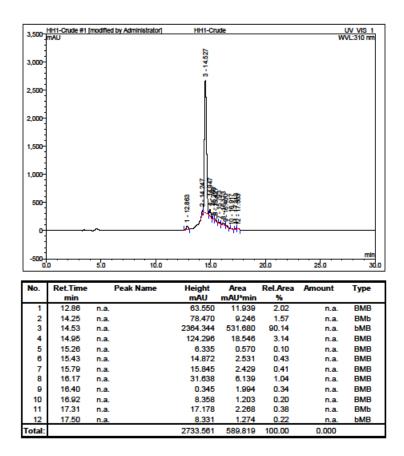


Figure S10: Crude HPLC trace of (17b).

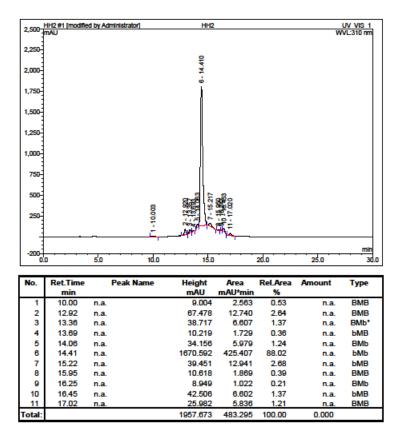


Figure S11: Crude HPLC trace of (18b).

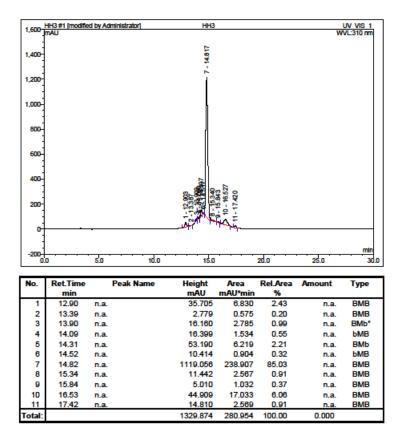


Figure S12: Crude HPLC trace of (19b).

# 5.7.2 Analytical HPLC traces of polyamides purified by semi-preparative HPLC

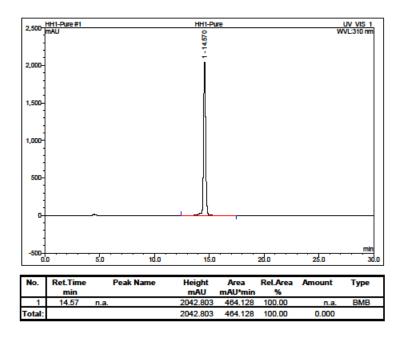


Figure S13: Pure HPLC trace of (17b).

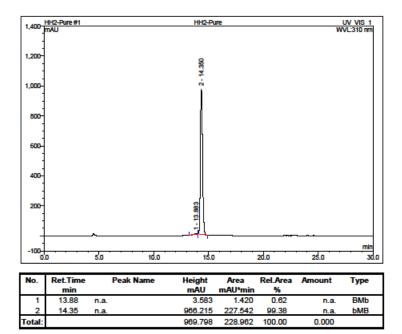
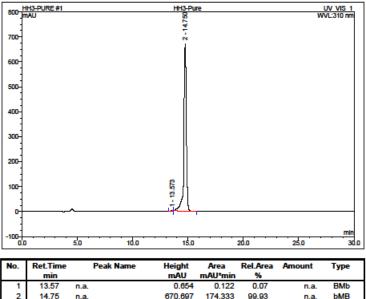


Figure S14: Pure HPLC trace of (18b).



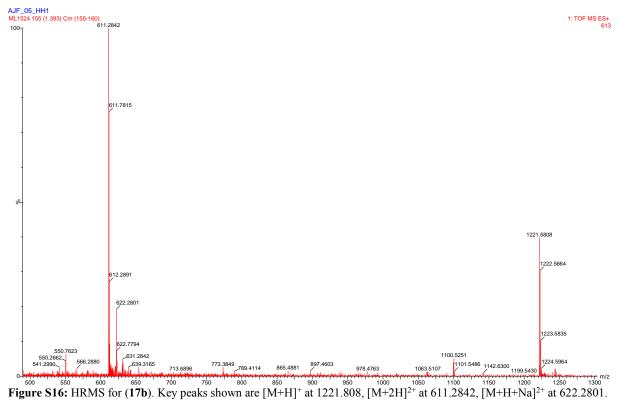
 1
 13.57
 n.a.
 0.654
 0.122
 0.07
 n.a.

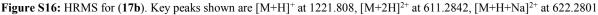
 2
 14.75
 n.a.
 670.697
 174.333
 99.93
 n.a.

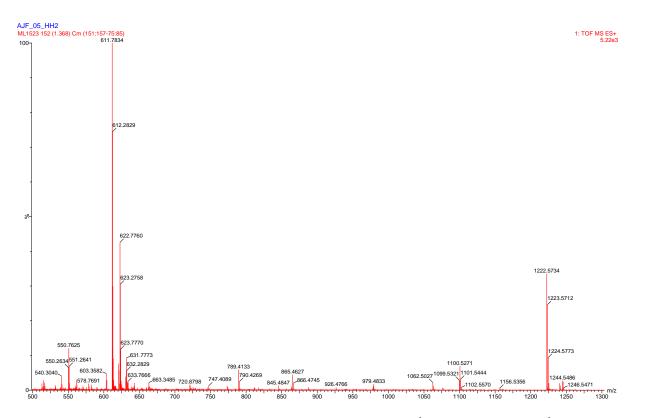
 Total:
 671.351
 174.455
 100.00
 0.000

Figure S15: Pure HPLC trace of (19b).

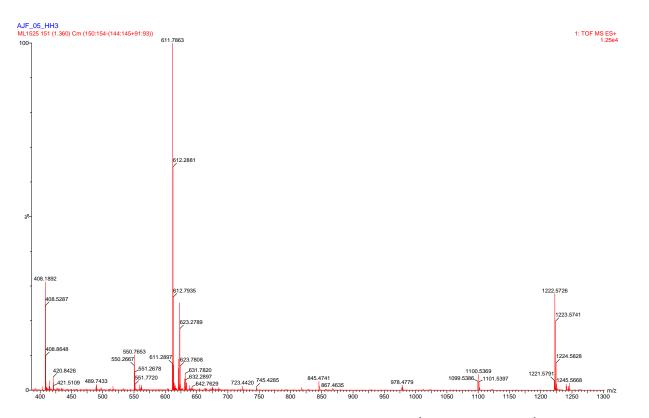
# 5.7.3 HRMS data of crude polyamides 17b-19b.







**Figure S17:** HRMS of (**18b**). Key peaks shown are [M+H]<sup>+</sup> at 1222.5734, [M+2H]<sup>2+</sup> at 611.7834, [M+H+Na]<sup>2+</sup> at 622.7760.

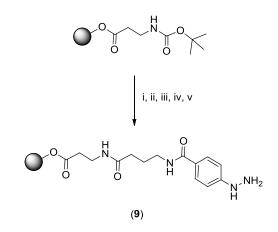


**Figure S18:** HRMS of (**19b**). Key peaks shown are [M+H]<sup>+</sup> at 1222.5726, [M+2H]<sup>2+</sup> at 611.7863, [M+3H]<sup>3+</sup> at 408.1892.

# 5.8 Synthesis of polyamides using a hydrazine linker

## 5.8.1 Resin Preparation

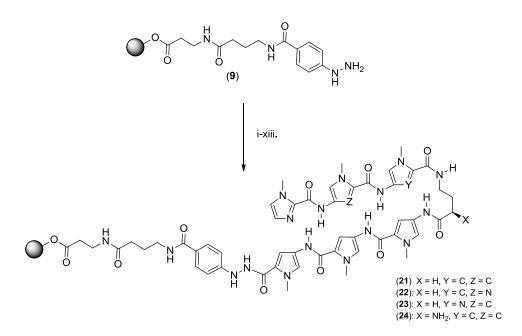
The preparation of Boc- $\beta$ -ala PAM resin is outlined in Scheme S5. The resin bound amine was deprotected through treatment with TPW solution for 1 minute, then 5 minutes with fresh reagent, after which GABA (4.0 equivalents) was preactivated with HATU (3.6 equivalents) and DIEA (9 equivalents) in DMF (dry; 1 mL) for 2 minutes before being added to the resin and shaken for 40 minutes at room temperature. The resin was then drained and washed successively with DMF (3x) and DCM (3x) before being swollen in DMF for 5 minutes. The amine was deprotected by treatment with TPW for 1 minute, then 2 x 3 minutes (fresh reagent each time), before being washed with DMF (4x) and dry DMF. The HBA monomer (4.0 equivalents) was preactivated with HATU (3.6 equivalents) and DIEA (9.0 equivalents) in DMF (dry; 1 mL) for 5 minutes, then was added to resin and shaken for 40 minutes. Once drained, the resin was washed again with DMF (4x).



Scheme S5: Preparation of Boc-β-ala PAM resin. (i) TPW; (ii) GABA, HATU, DIEA; (iii) TPW; (iv) HBA, HATU, DIEA; (v) 20 % piperidine in DMF (V/V).

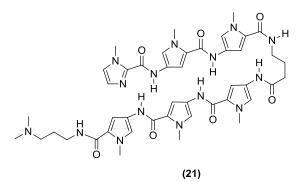
## 5.8.2 Synthesis of polyamides using the PAM-hydrazine hybrid resin

Polyamides were synthesised using the same activation strategies described earlier in Section 5.4. The initial Fmoc deprotection was carried out by two treatments of shaking with a 20 % piperidine in DMF solution (v/v) for 2 x 10 minutes (fresh reagent each time). The resin was then washed with DMF (4x), then dry DMF. Polyamides (**21-24**) were then synthesised according to Scheme S6.



Scheme S6: Synthesis of PAs 21-24. Steps (i)-(xiii) are stated in the respective polyamide experimental.

5.8.2.1 Synthesis of (21)

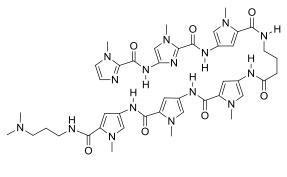


Polyamide (**21**) was synthesised on 50 mg of resin (**9**) as shown in Scheme S6 using the following reagents and times: (i) BocPyOH, HATU, DIEA, 40 minutes; (ii) TPW; (iii) BocPyOH, HATU, DIEA, 40 minutes; (iv) TPW; (v) BocPyOH, HATU, DIEA, 40 minutes; (vi) TPW; (vii) GABA, HATU, DIEA, 60 minutes; (viii) TPW; (ix) BocPyOH, HATU, DIEA, 60 minutes; (x) TPW; (xi) BocPyOH, HATU, DIEA, 90 minutes; (xii) TPW; (xiii) ImOH, DCC/HOAt, DIEA, 60 minutes. This was cleaved from the resin under standard cleavage conditions to give the crude material in 68 % purity. After purification by semi-preparative HPLC the pure material was isolated as a fluffy white solid (1.3 mg; 11 % recovery).

Analytical HPLC  $t_r = 13.6$  minutes.

HRMS (ESI+) 906.4487 calculated for  $C_{44}H_{56}N_{15}O_7^+$ ,  $[M+H]^+$ , found 906.4487.

## 5.8.2.2 Synthesis of (22)



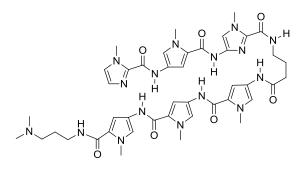
(22)

Polyamide (**22**) was synthesised on 50 mg of resin (**9**) (Scheme S6) using the following reagents and times: (i) BocPyOH, HATU, DIEA, 40 minutes; (ii) TPW; (iii) BocPyOH, HATU, DIEA, 40 minutes; (iv) TPW; (v) BocPyOH, HATU, DIEA, 40 minutes; (vi) TPW; (vii) GABA, HATU, DIEA, 60 minutes; (viii) TPW; (ix) BocPyOH, HATU, DIEA, 60 minutes; (x) TPW; (xi) BocImOH, DCC/HOAt, DIEA, 60 minutes; (xii) TPW; (xiii) ImOH, DCC/HOAt, DIEA, 60 minutes. This was cleaved under standard cleavage conditions to give the crude material in 70 % purity by HPLC at 310 nm. After purification by semi-preparative HPLC the pure material was isolated as a fluffy white solid (1.3 mg; 11 % recovery).

Analytical HPLC  $t_r = 14.0$  minutes.

HRMS (ESI+) 907.4440 calculated for C<sub>43</sub>H<sub>55</sub>N<sub>16</sub>O<sub>7</sub><sup>+</sup>, [M+H]<sup>+</sup>, found 907.4456.

5.8.2.3 Synthesis of (23)



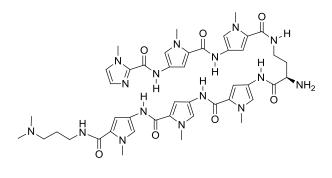
(23)

Polyamide (23) was synthesised on 50 mg of resin (9) (Scheme S6) using the following reagents and times: (i) BocPyOH, HATU, DIEA, 40 minutes; (ii) TPW; (iii) BocPyOH, HATU, DIEA, 40 minutes; (iv) TPW; (vi) BocPyOH, HATU, DIEA, 40 minutes; (vi) TPW; (vii) GABA, HATU, DIEA, 60 minutes; (viii) TPW; (ix) BocImOH, DCC/HOAt, DIEA, 60 minutes; (x) TPW; (xi) BocPyOH, BTC/collidine, DIEA, 90 minutes; (xii) TPW; (xiii) ImOH, DCC/HOAt, DIEA, 60 minutes; of the crude material in 74 % purity by HPLC at 310 nm. After purification by semi-preparative HPLC the pure material was isolated as a fluffy white solid (1.4 mg; 12 % recovery).

Analytical HPLC  $t_r = 13.3$  minutes.

HRMS (ESI +ve mode) 907.4440 calculated for  $C_{43}H_{55}N_{16}O_7^+$ ,  $[M+H]^+$ , found 907.4449.

## 5.8.2.4 Synthesis of (24)



(24)

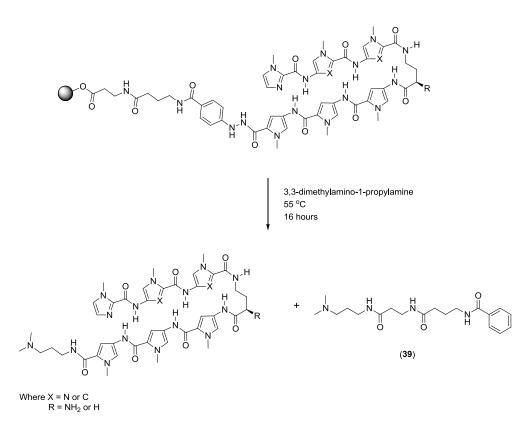
Polyamide (**24**) was synthesised on 50 mg of resin (**9**) (Scheme S6) using the following reagents and times: (i) BocPyOH, HATU, DIEA, 40 minutes; (ii) TPW; (iii) BocPyOH, HATU, DIEA, 40 minutes; (iv) TPW; (v) BocPyOH, HATU, DIEA, 40 minutes; (vi) TPW; (vii) Fmoc-D-Dab(Boc)-OH, HATU, DIEA, 60 minutes; (viii) TPW; (ix) BocPyOH, HATU, DIEA, 60 minutes; (x) TPW; (xi) BocPyOH, HATU, DIEA, 90 minutes; (xii) TPW; (xiii) ImOH, DCC/HOAt, DIEA, 60 minutes. This was cleaved under standard cleavage conditions to give the crude material in 79 % purity by HPLC at 310 nm. After purification by semi-preparative HPLC the pure material was isolated as a fluffy white solid (1.6 mg; 13 % recovery).

Analytical HPLC  $t_r = 12.1$  minutes.

HRMS (ESI +ve mode) 921.4596 calculated for  $C_{44}H_{57}N_{16}O_7^+$ ,  $[M+H]^+$ , found 921.4561.

## 5.8.3 Cleavage of polyamides using the hydrazine resin

Polyamides (**21-24**) were cleaved from the resin under standard cleavage conditions and found to yield only the shortened polyamide through cleavage at the hydrazide (Scheme S7). The tail was also found to cleave from the resin under these conditions to give compound (**39**) shown in Scheme S7 which was characterised by HRMS (363.2396 calculated for  $C_{19}H_{31}N_4O_3^+$  [M+H]<sup>+</sup>, found 363.2400 - Figure S19). This confirmed that oxidation of the hydrazinyl group prior to resin cleavage was not essential in order to obtain the shortened polyamide.



Scheme S7: Illustration of products resulting from cleavage of hydrazine resin.

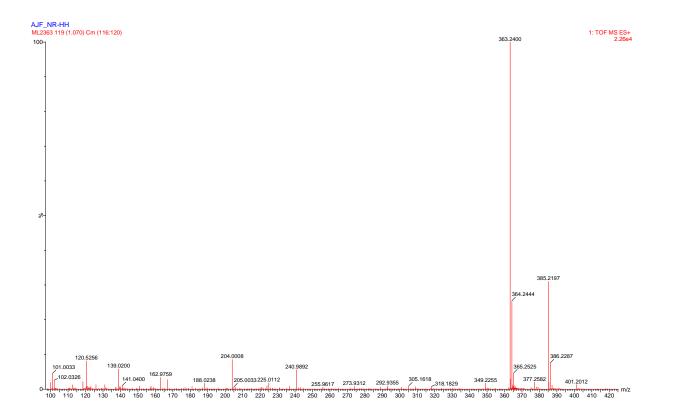
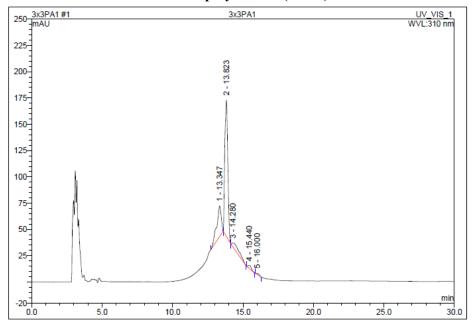


Figure S19: MS of cleavage by-product (39). Key peaks shown are  $[M+H]^+$  at 363.2400,  $[M+Na]^+$  at 385.2197.

# 5.9 HPLC analysis and purification of polyamides prepared on the hydrazine hybrid resin



# 5.9.1 Crude HPLC traces of polyamides (21-24)

No.	Ret.Time	Peak Name	Height	Area	Rel.Area	Amount	Туре
	min		mAU	mAU*min	%		
1	13.35	n.a.	28.905	10.746	22.69	n.a.	BMb
2	13.82	n.a.	129.914	32.087	67.76	n.a.	bMB
3	14.28	n.a.	3.731	3.384	7.15	n.a.	BMB
4	15.44	n.a.	2.958	0.804	1.70	n.a.	BMB
5	16.00	n.a.	1.237	0.335	0.71	n.a.	BMB
Total:			166.746	47.356	100.00	0.000	

Figure S20: Crude HPLC trace of (21).

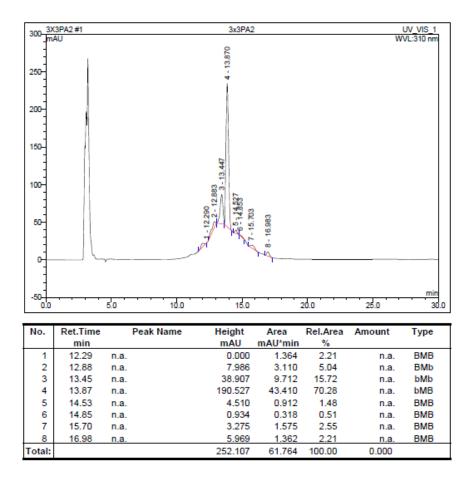


Figure S21: Crude HPLC trace of (22).

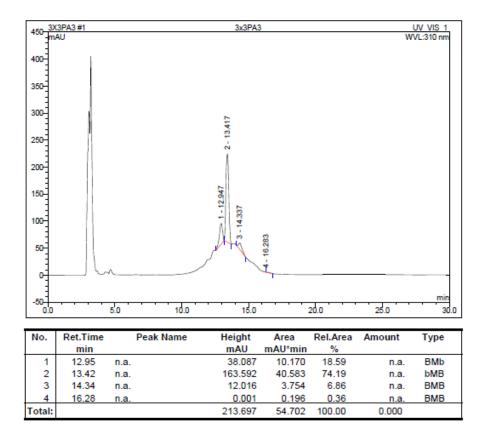


Figure S22: Crude HPLC trace of (23).

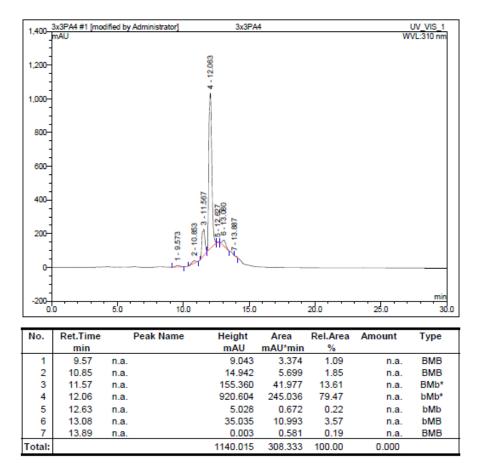
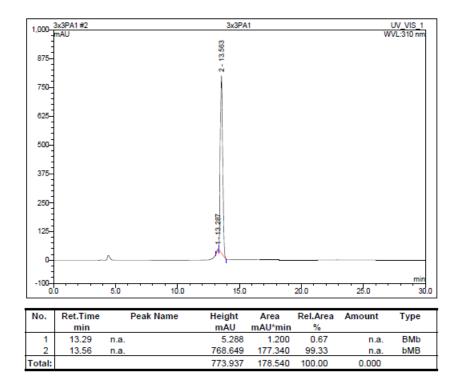


Figure S23: Crude HPLC trace of (24).



5.9.2 HPLC traces of polyamides (21-24) after semi-preparative HPLC purification

Figure S24: Pure HPLC trace of (21).

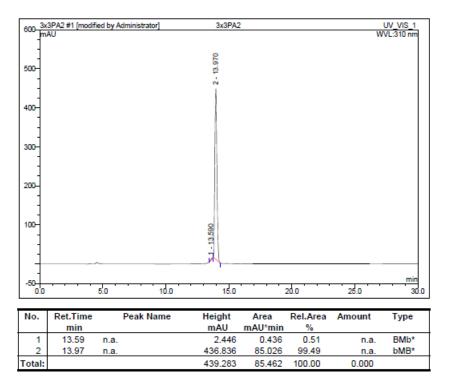


Figure S25: Pure HPLC trace of (22).

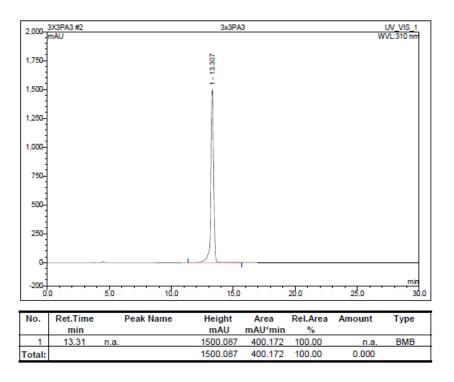


Figure S26: Pure HPLC trace of (23).

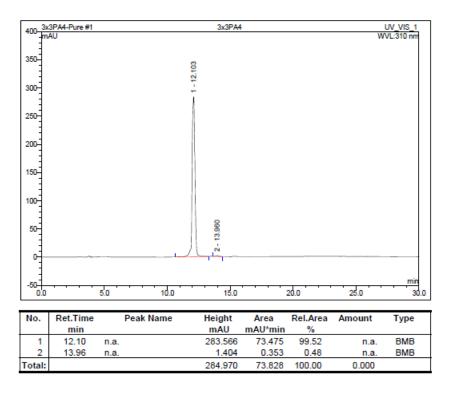
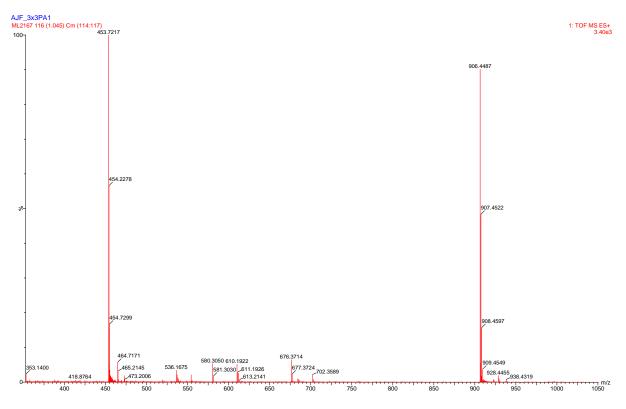
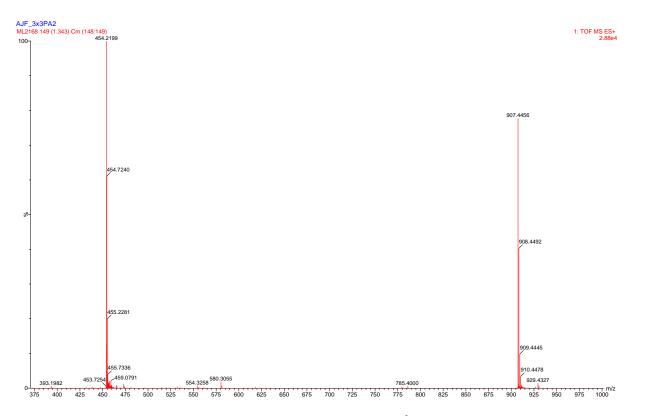


Figure S27: Pure HPLC trace of (24).

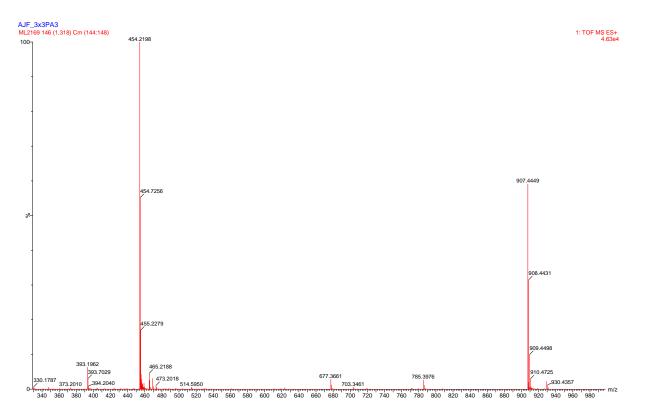
# 5.9.3 Crude HRMS data of polyamides (21-24)



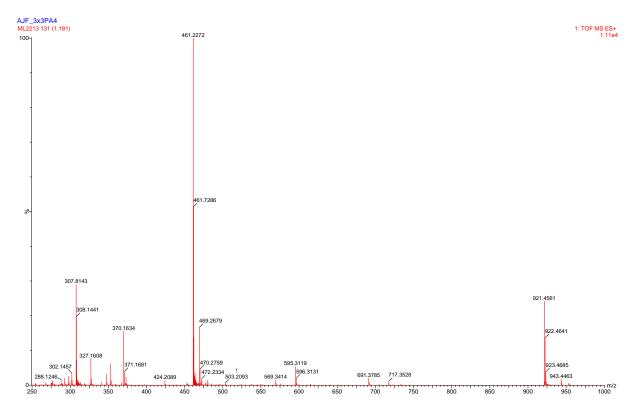
**Figure S28:** HRMS of (**21**). Key peaks shown are [M+H]<sup>+</sup> at 906.4487, [M+2H]<sup>2+</sup> at 453.7217.



**Figure S29:** HRMS of (**22**). Key peaks shown are [M+H]<sup>+</sup> at 907.4456, [M+2H]<sup>2+</sup> at 454.2199.



**Figure S30:** HRMS of (**23**). Key peaks shown are [M+H]<sup>+</sup> at 907.4449, [M+2H]<sup>2+</sup> at 454.2198.

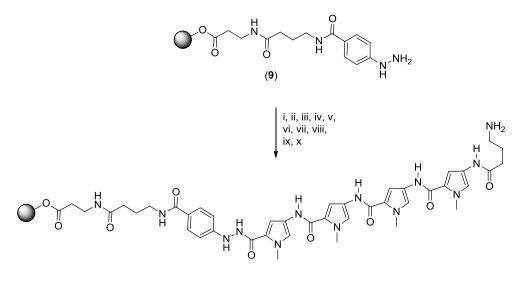


**Figure S31:** HRMS of (**24**). Key peaks shown are [M+H]<sup>+</sup> at 921.4561, [M+2H]<sup>2+</sup> at 461.2272, [M+3H]<sup>3+</sup> at 307.8143.

## 5.10 Preparation of polyamides using the half-half coupling method on the hydrazine resin

## 5.10.1 Preparation of resin-bound tetramer (16)

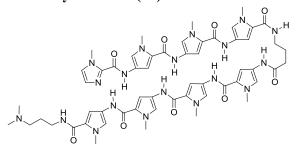
The resin bound tetramer was synthesised as in Scheme S8, utilising a hydrazine resin (Scheme S5).



(16)

Scheme S8: Synthesis of polyamide tetramer on new resin. (i) BocPyOH, HATU, DIEA, 40 minutes; (ii) TPW; (iii) BocPyOH, HATU, DIEA, 40 minutes; (iv) TPW; (v) BocPyOH, HATU, DIEA, 40 minutes; (vi) TPW; (vii) BocPyOH, HATU, DIEA, 60 minutes; (viii) TPW; (ix) GABA, HATU, DIEA, 60 minutes; (x) TPW.

#### 5.10.2 Synthesis of (20)



Resin-bound tetramer (16) (20 mg; 5.2  $\mu$ mol, Scheme S8) was swollen in DMF for 2 hours. The resin was then drained and washed three times with DMF. The tetramer (13) (10 mg; 20.3  $\mu$ mol) was pre-activated with HATU (7 mg; 18.4  $\mu$ mol) and DIEA (50  $\mu$ L; 0.3mmol) in DMF (600  $\mu$ L) for 30 minutes before being added to the resin, and then shaken for 4 hours. The reaction mixture was then drained from the vessel and the resin washed three times with DMF, then DCM, then twice with methanol before being flushed with nitrogen for 5 minutes. Polyamide (20) was cleaved from the resin with  $N^1$ , $N^1$ -dimethylpropane-1,3-diamine (500  $\mu$ L) under standard cleavage conditions and the resulting crude product was found to be 87 % pure by HPLC at 310 nm. This was purified by semi-preparative HPLC to give the product as a fluffy white solid (1.2 mg; 20 % recovery).

Analytical HPLC  $t_r = 14.7$  minutes.

HRMS (ESI+) 1150.5447 calculated for C<sub>56</sub>H<sub>68</sub>N<sub>19</sub>O<sub>9</sub><sup>+</sup> [M+H]<sup>+</sup>, found 1150.5410.

#### 5.11 Characterisation of (20)

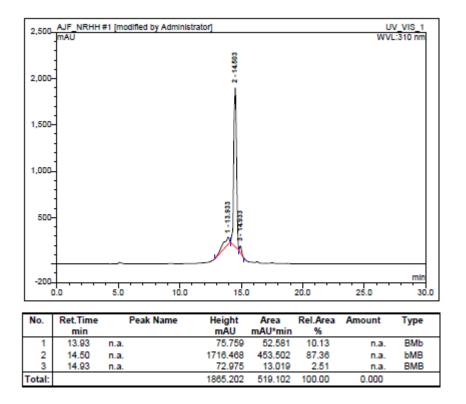


Figure S32: Crude HPLC trace of (20).

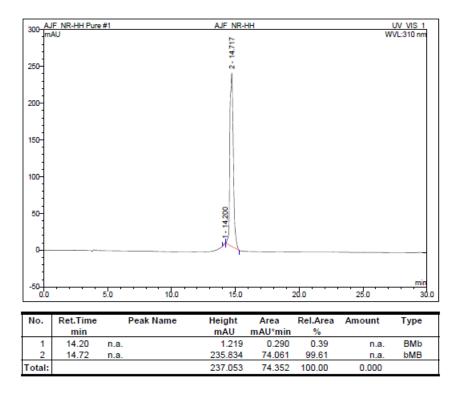
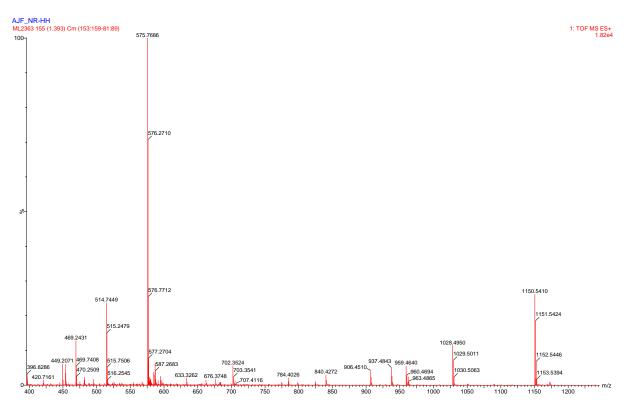
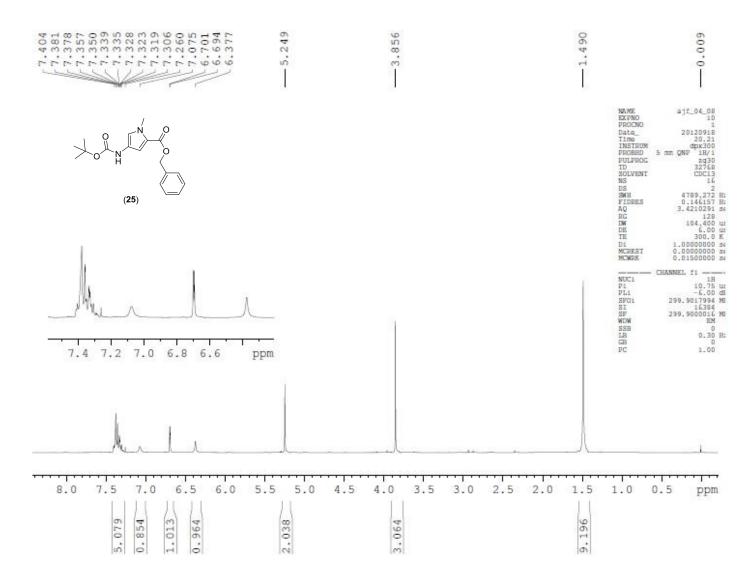


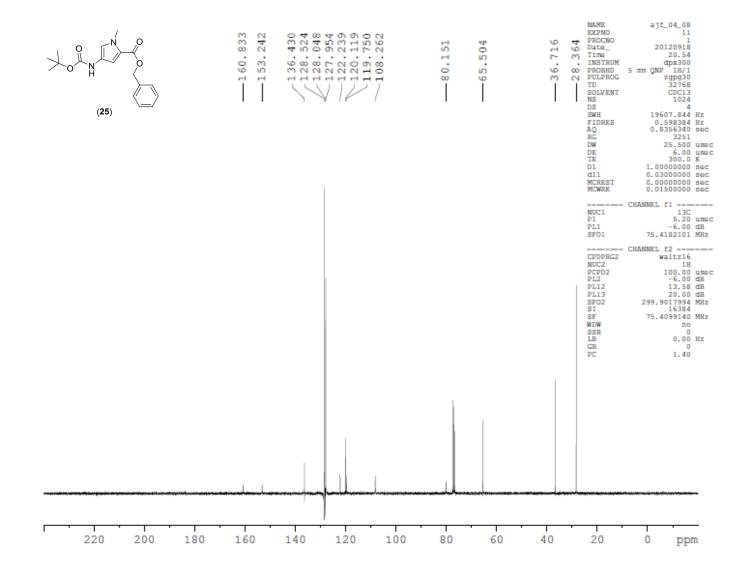
Figure S33: Pure HPLC trace of (20).

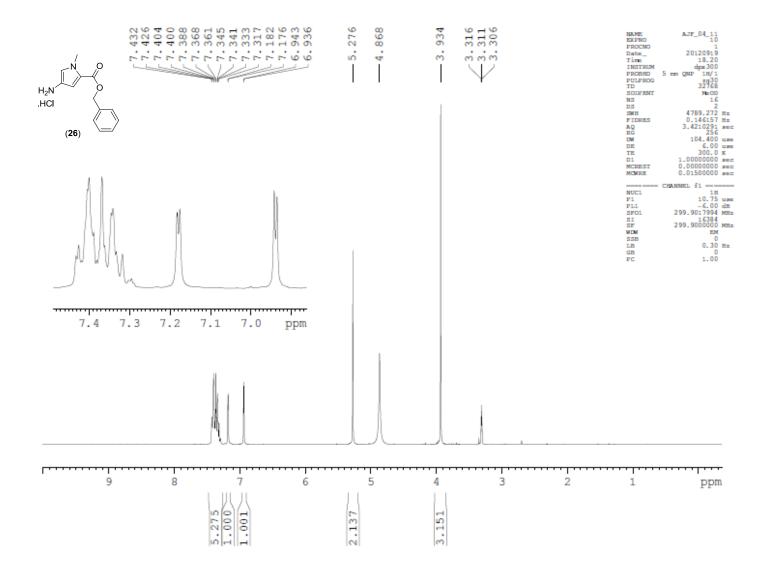


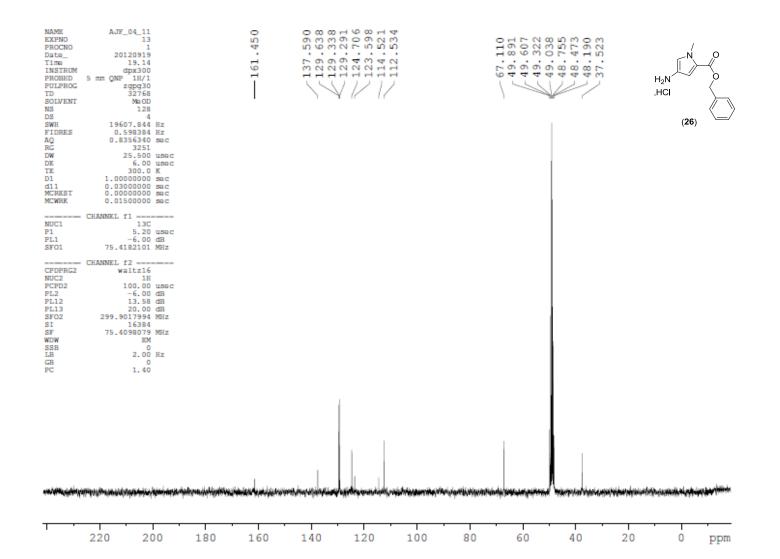
**Figure S34:** HRMS of crude (**20**). Key peaks shown are [M+H]<sup>+</sup> at 1150.5410, [M+2H]<sup>2+</sup> at 575.7686.

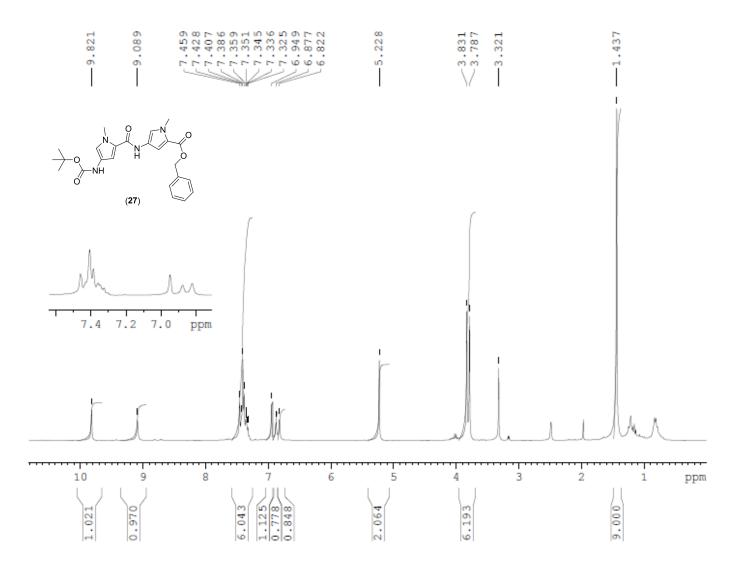
6.0 NMR Spectra

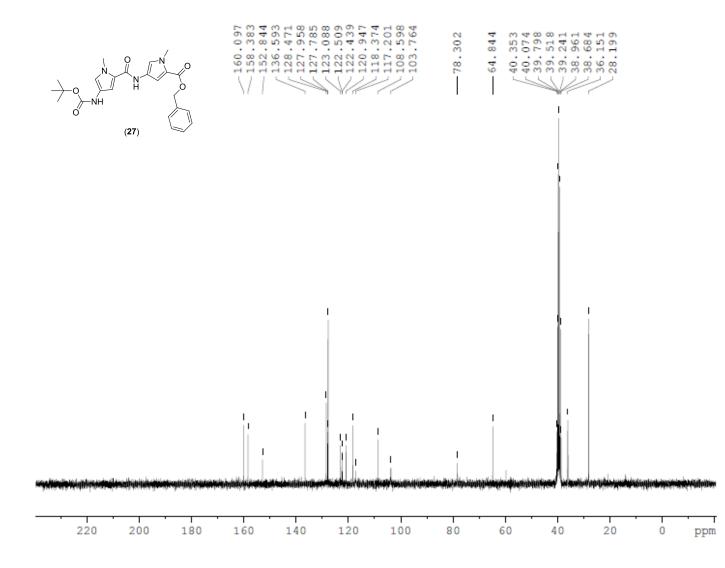


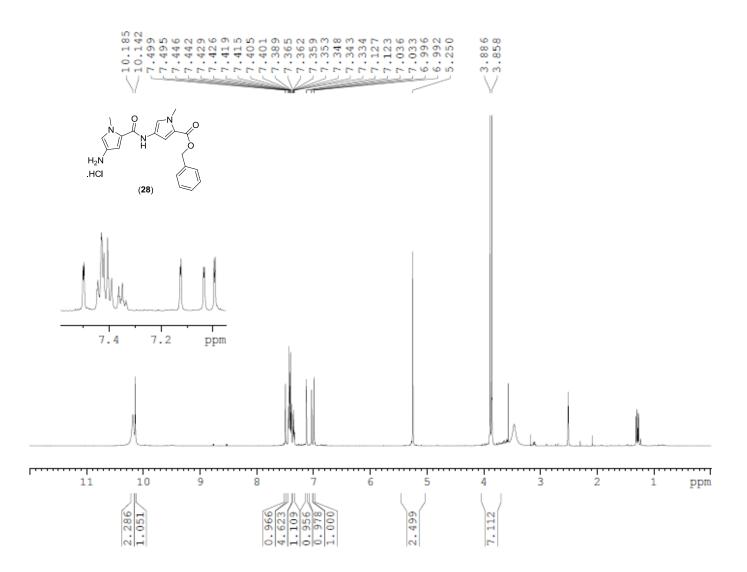


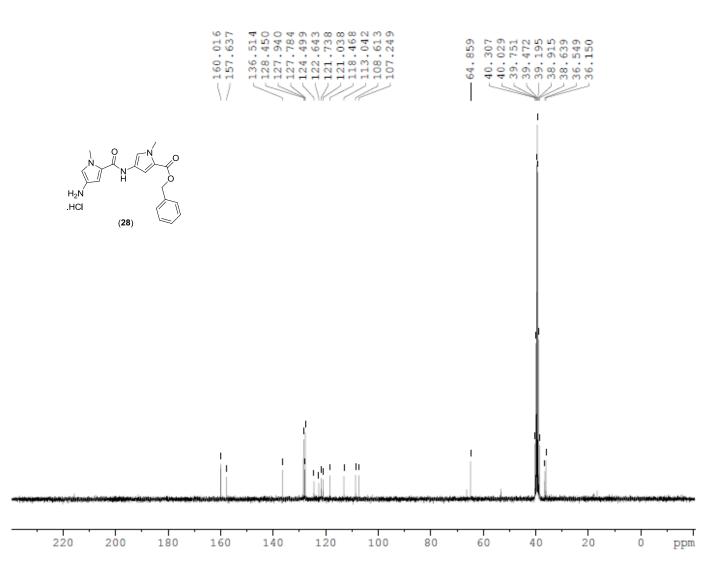


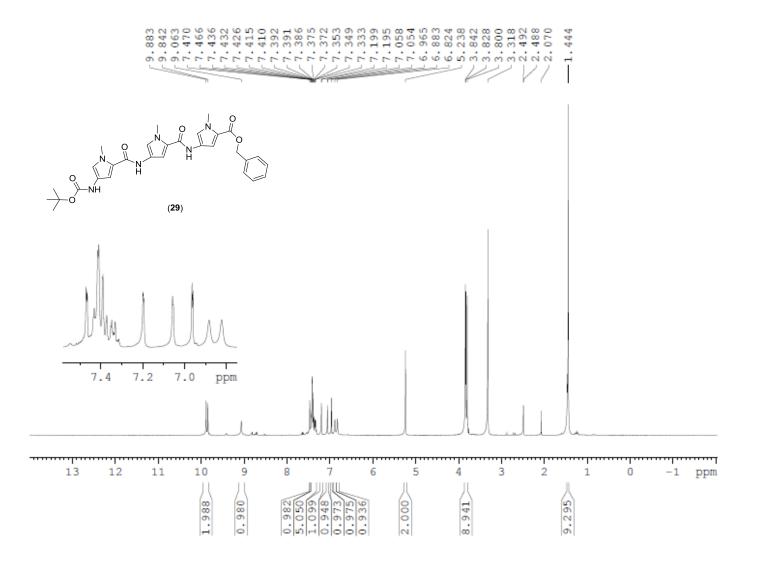


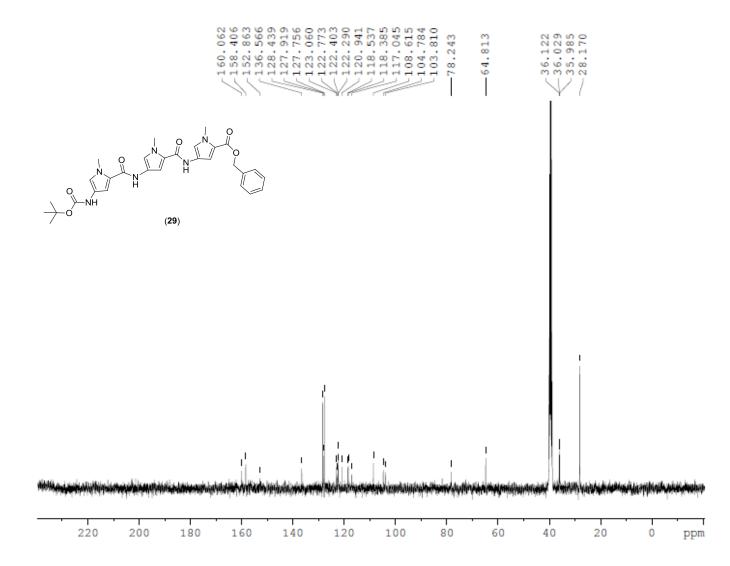


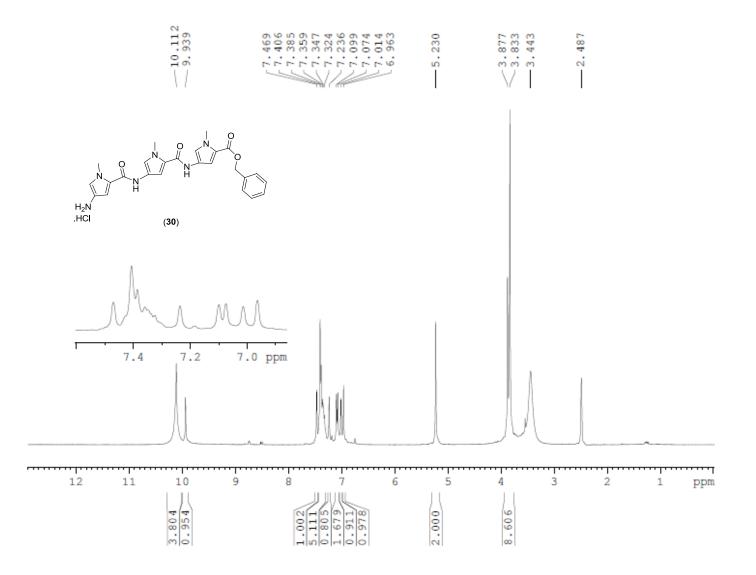


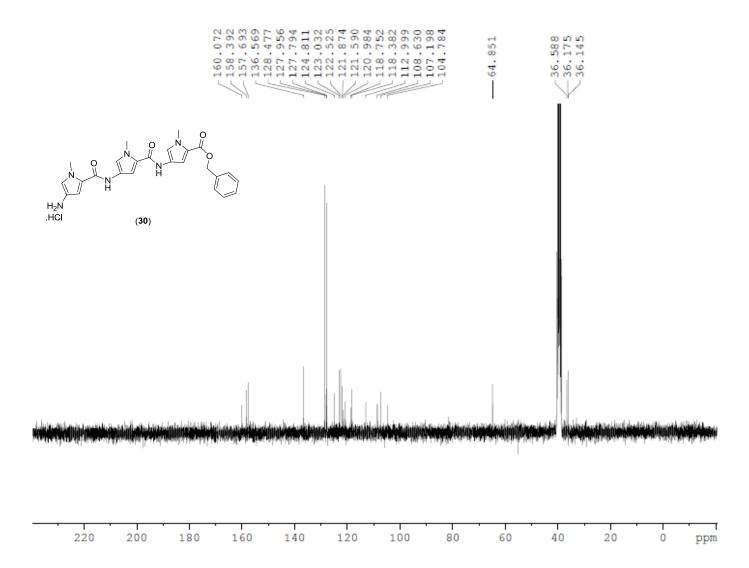


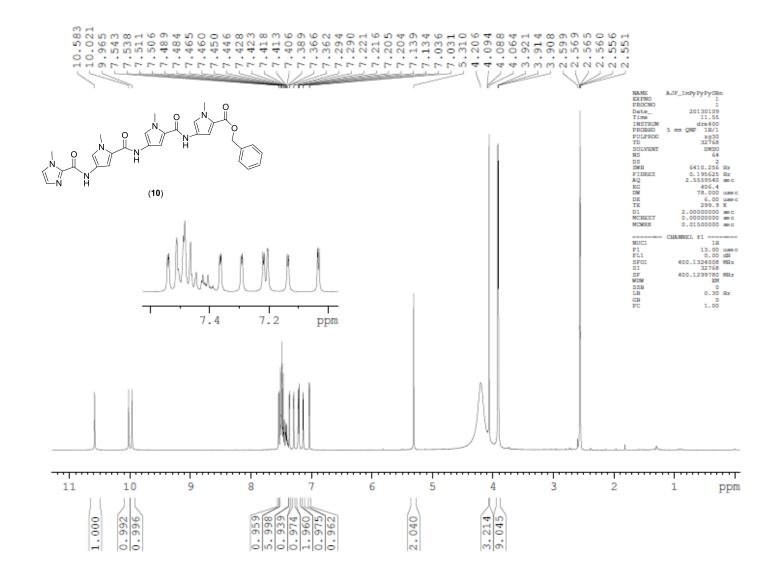


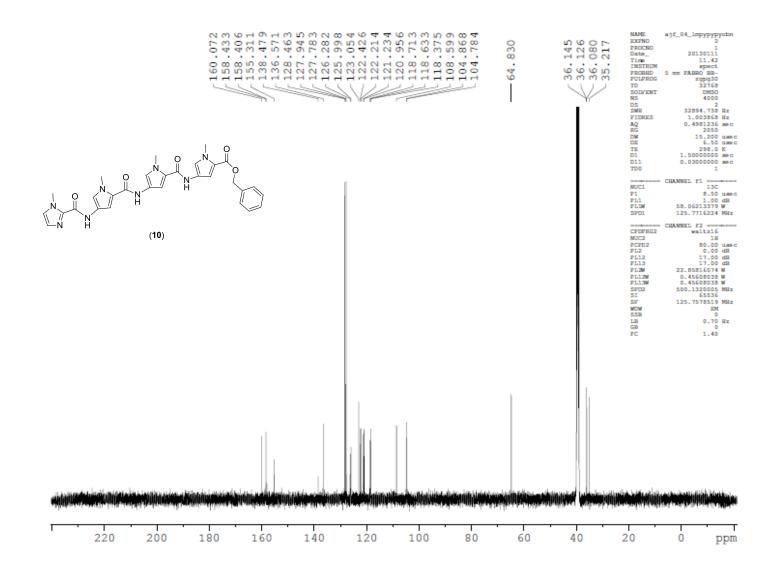


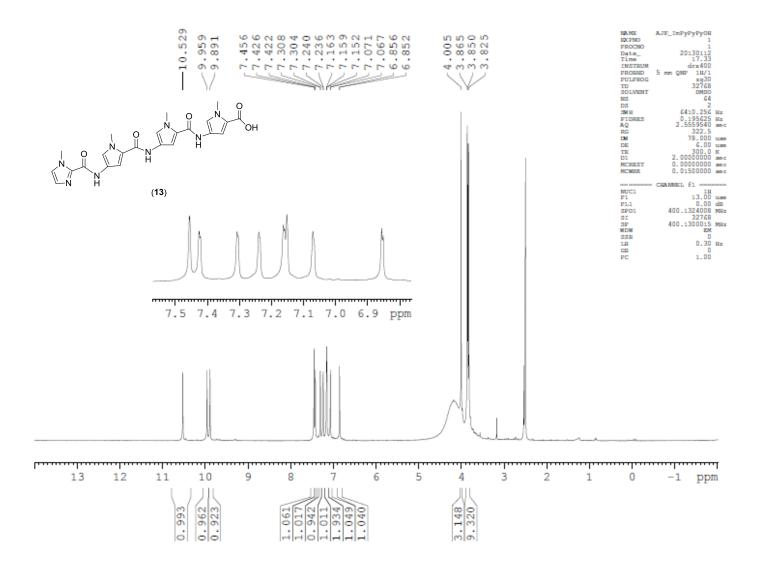


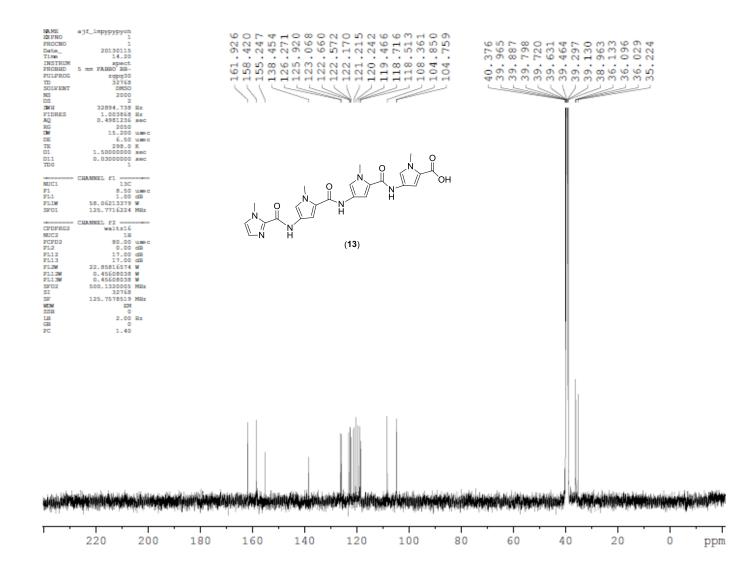


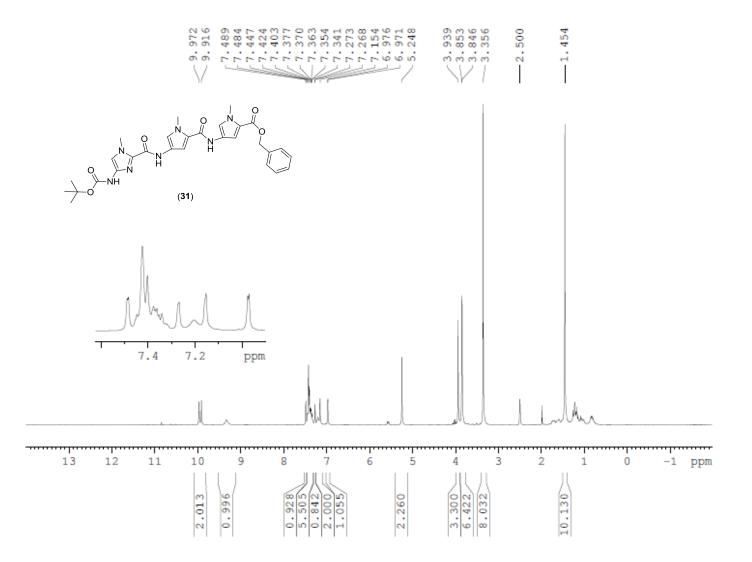


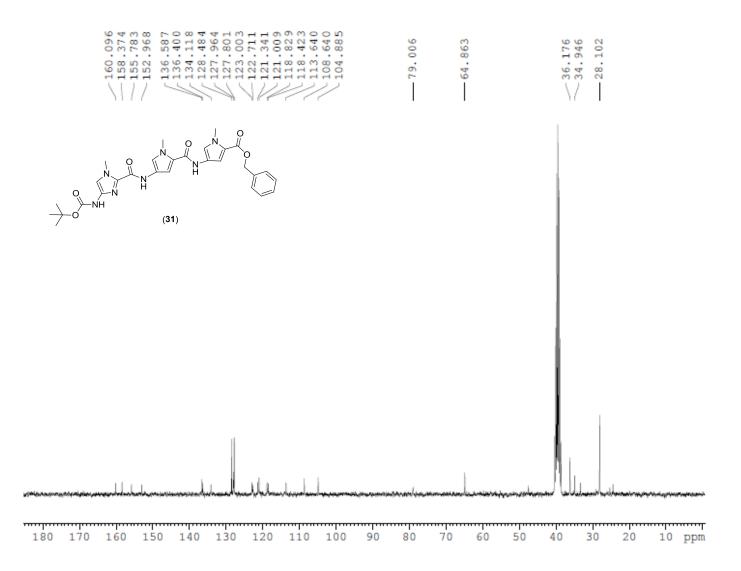


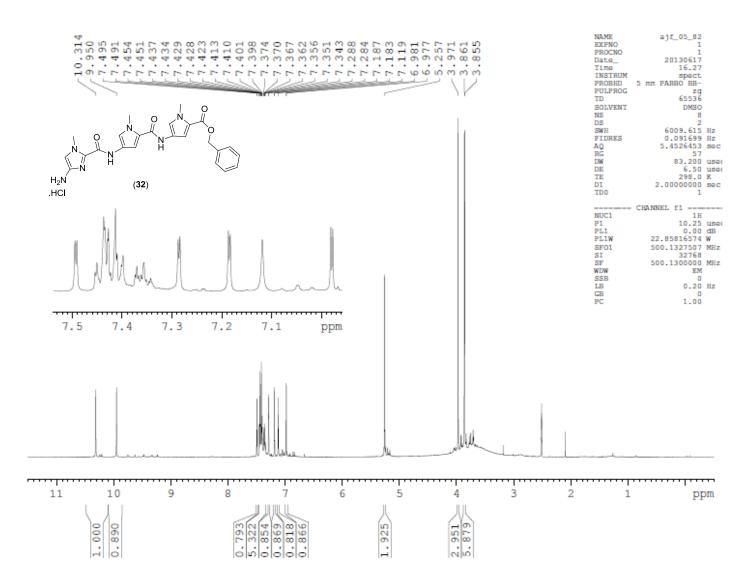


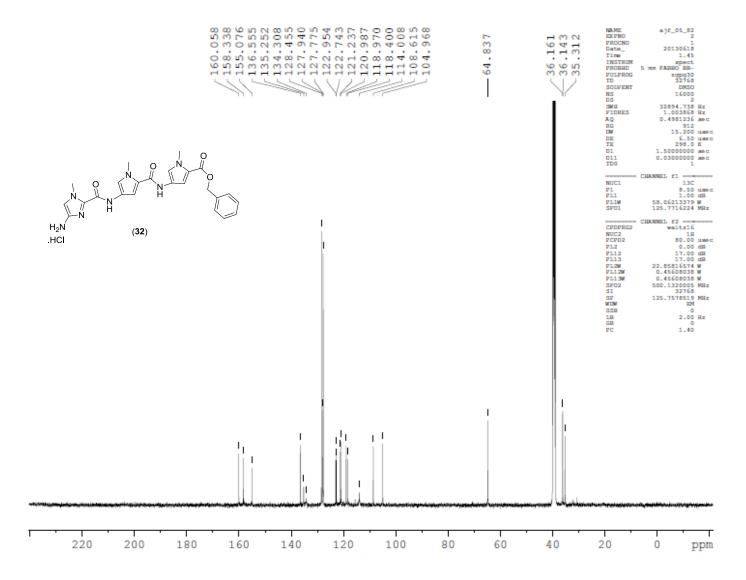


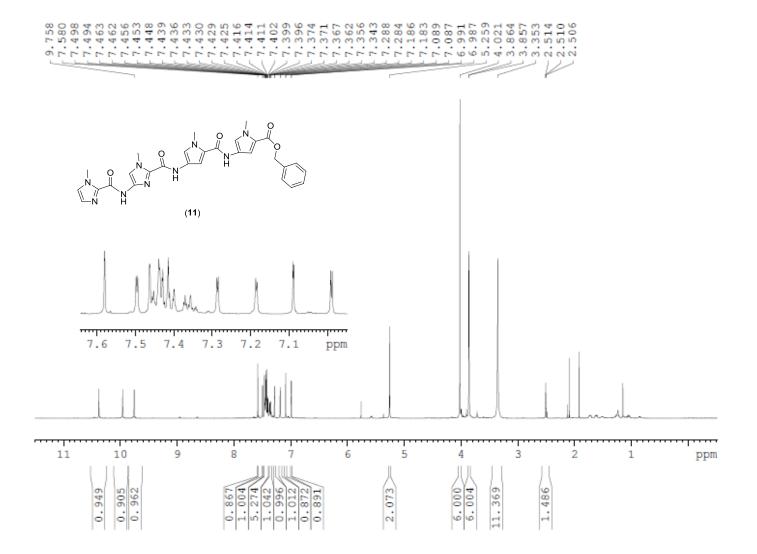


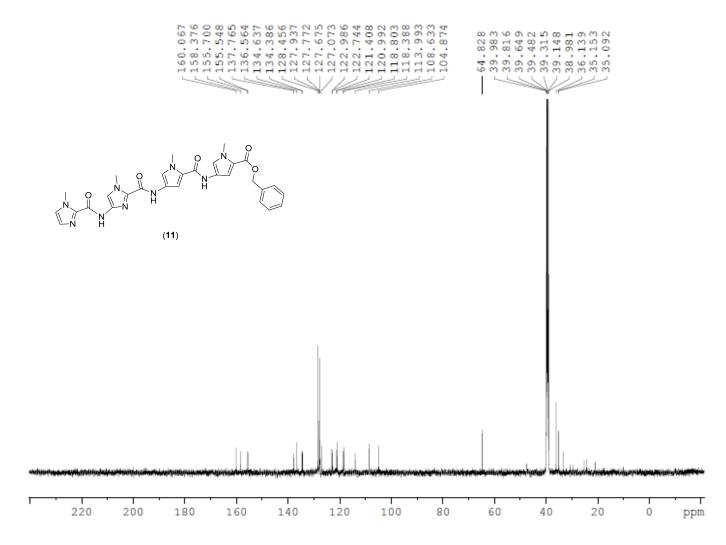


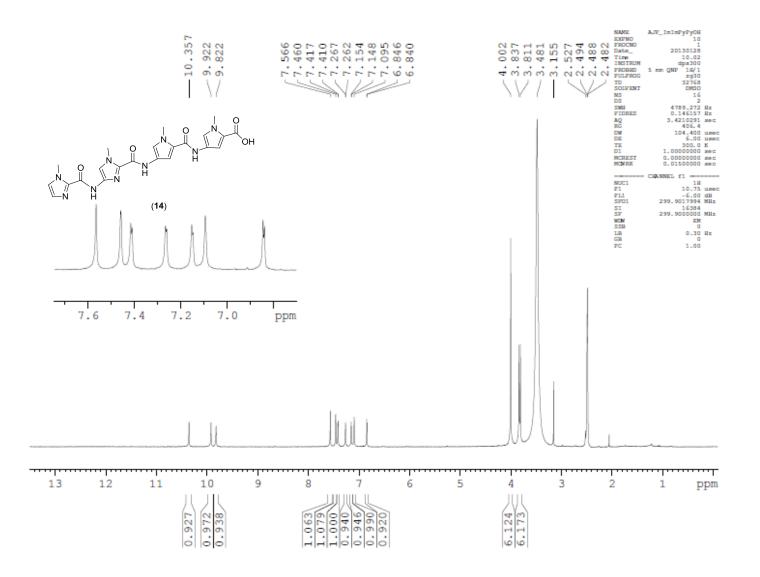


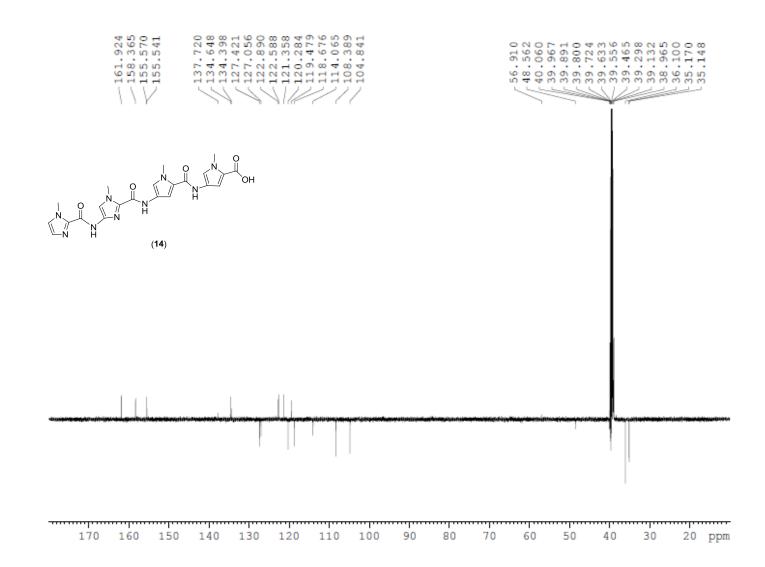


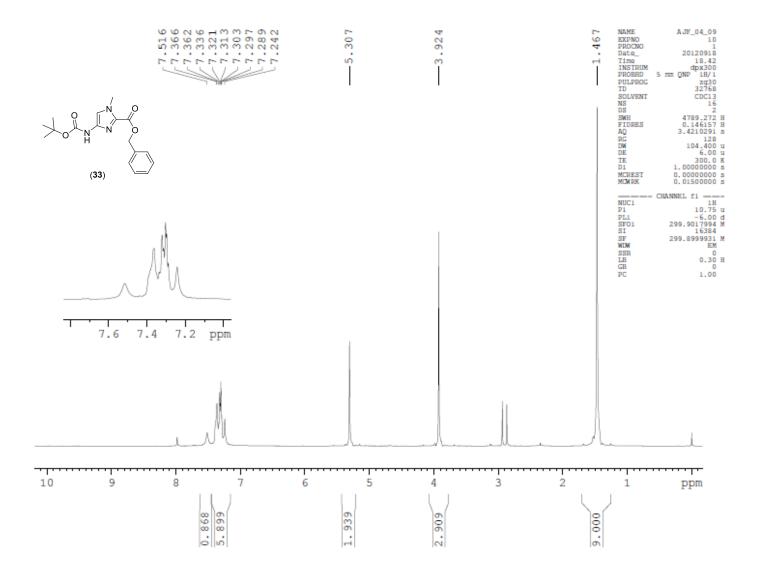


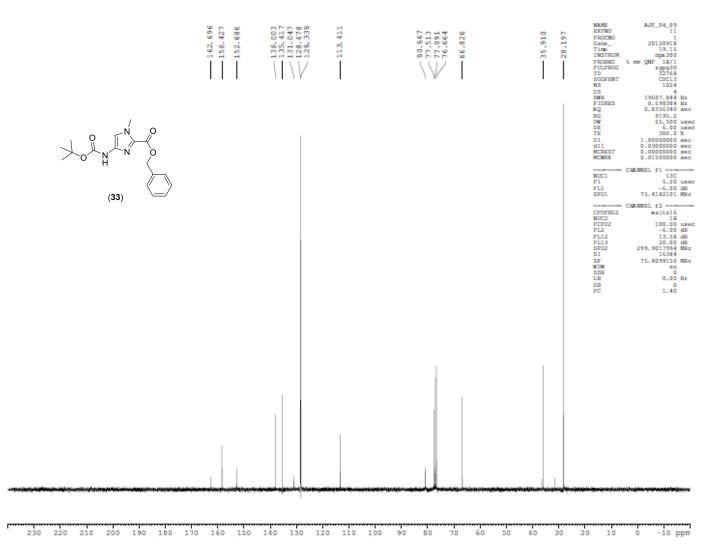


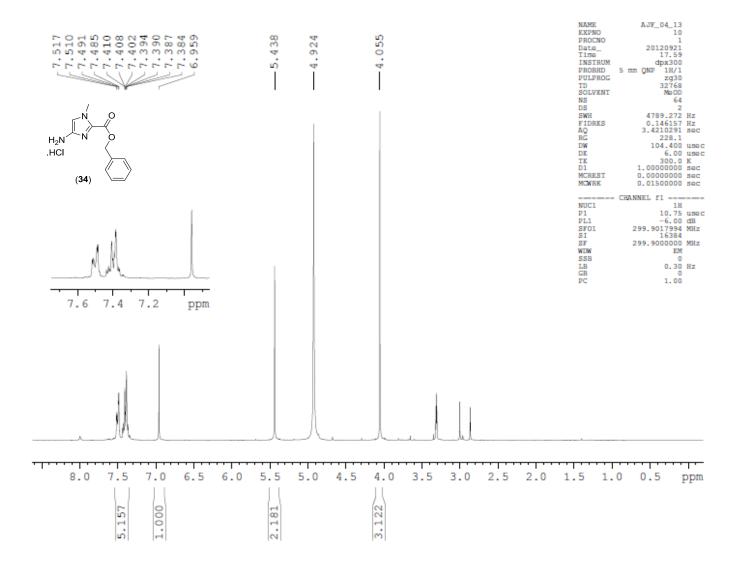


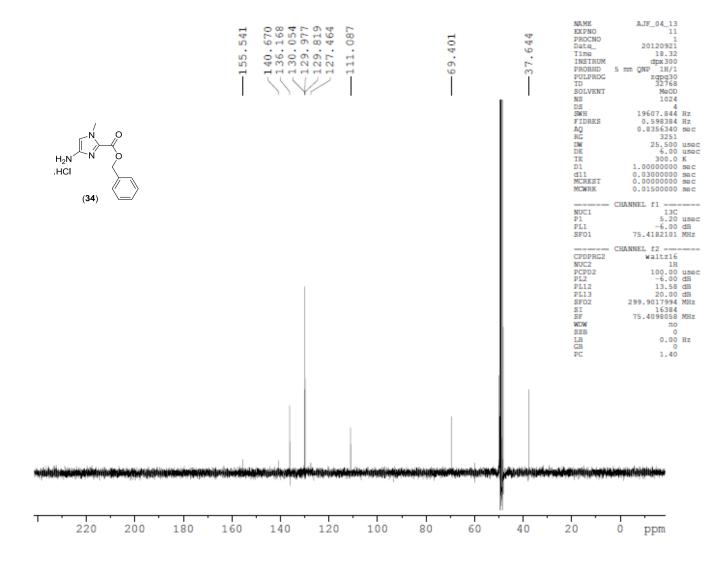


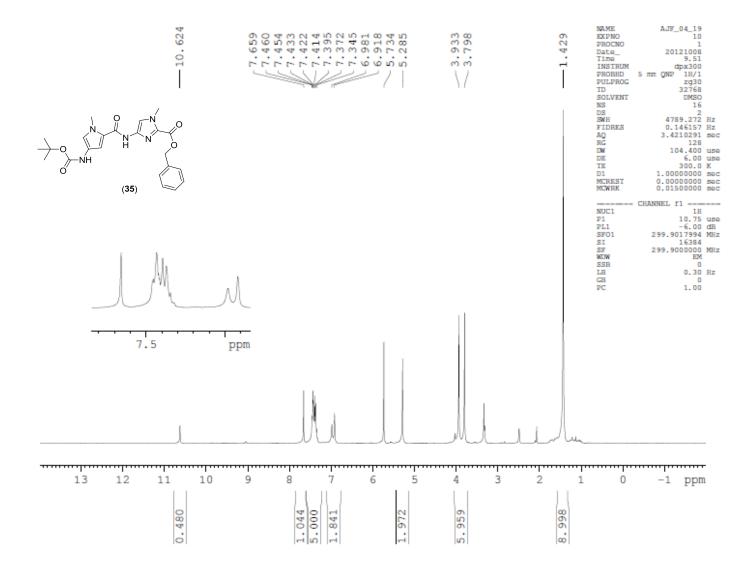


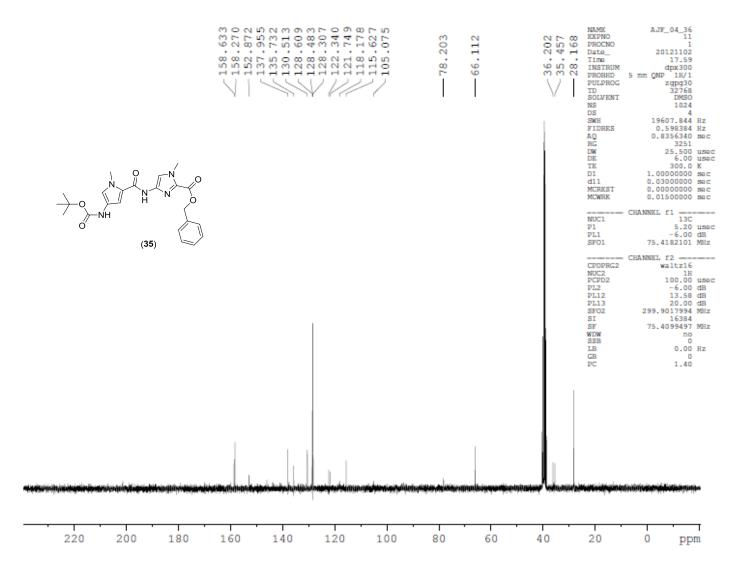


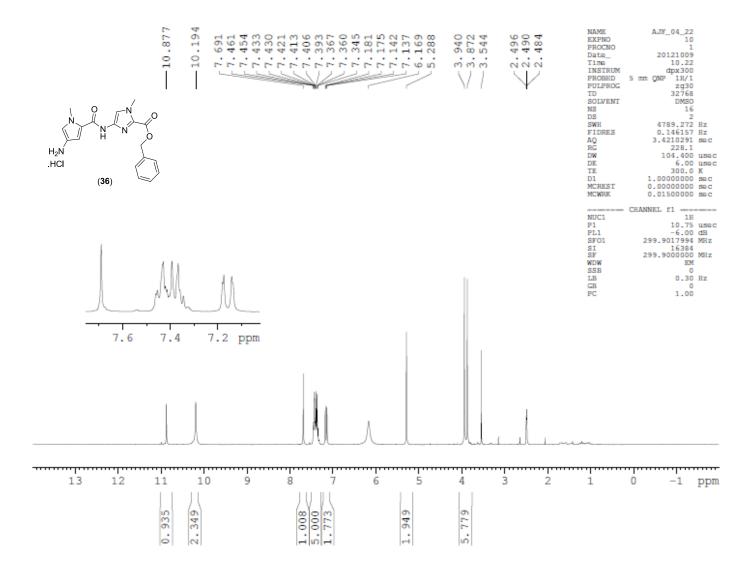


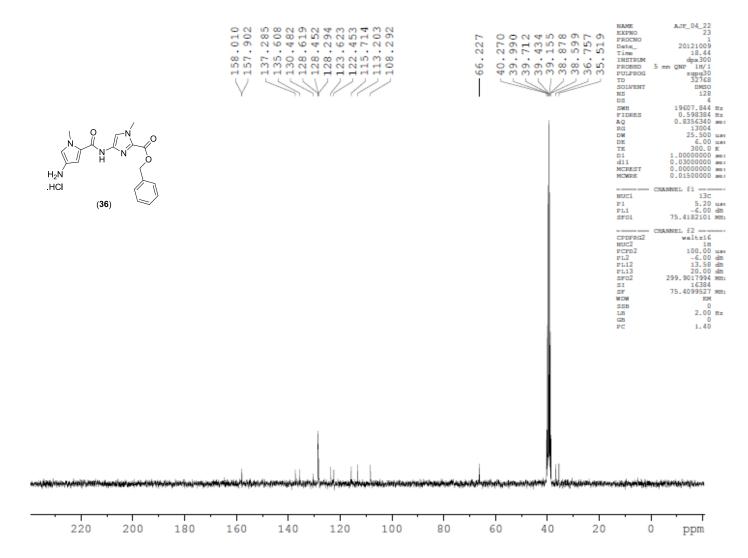


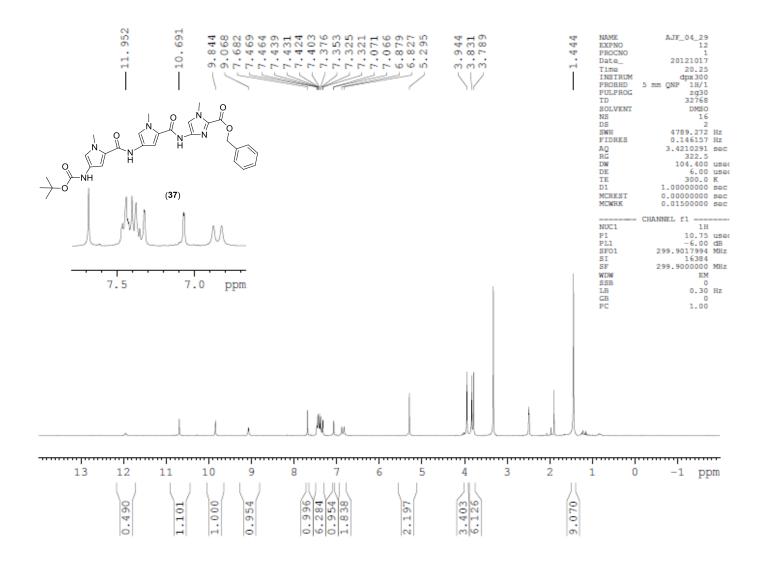


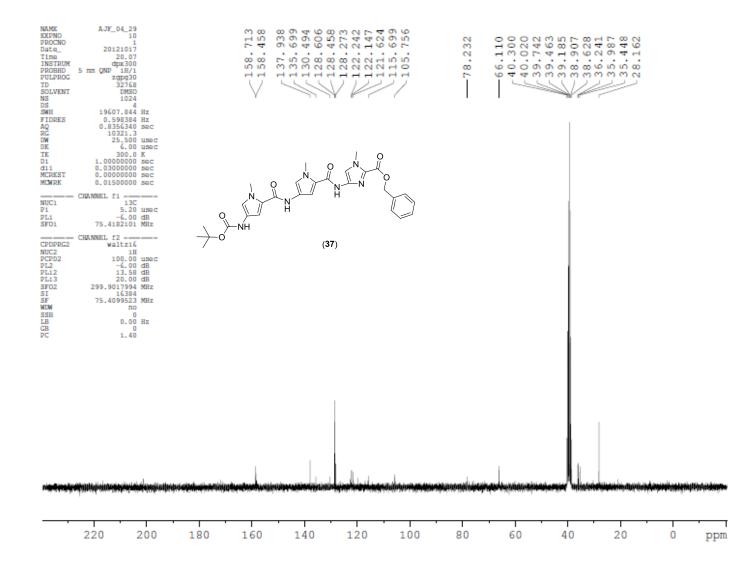


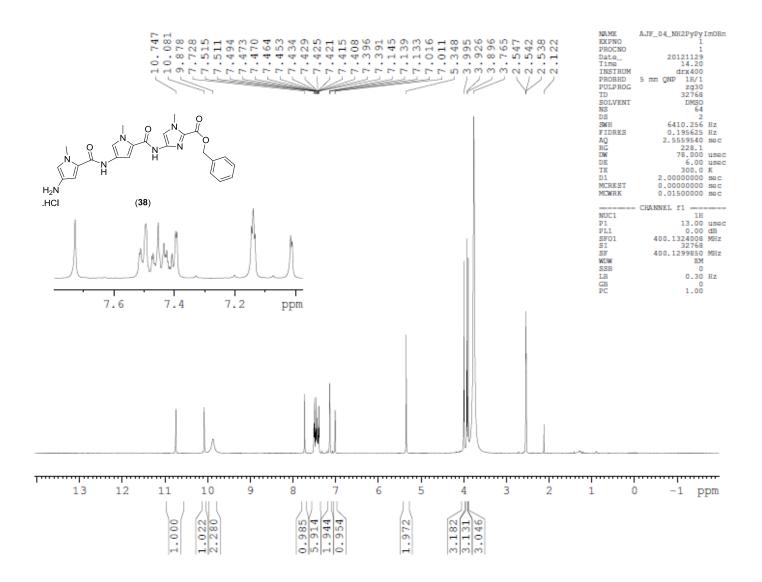


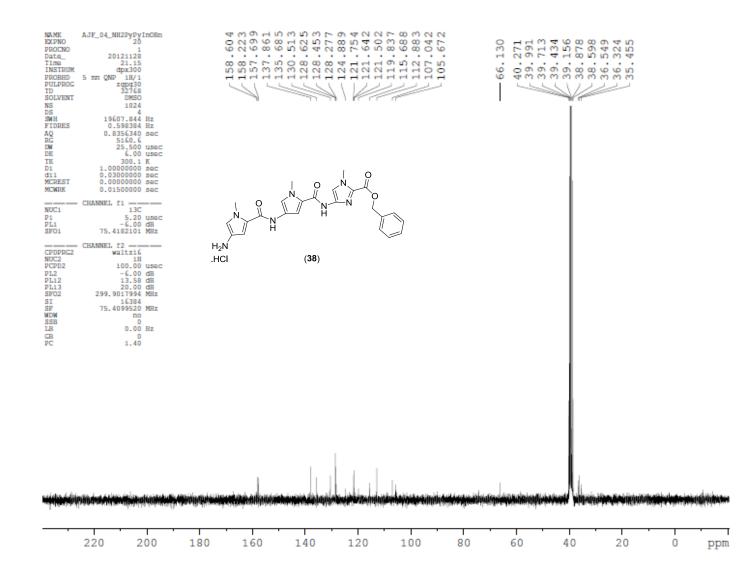


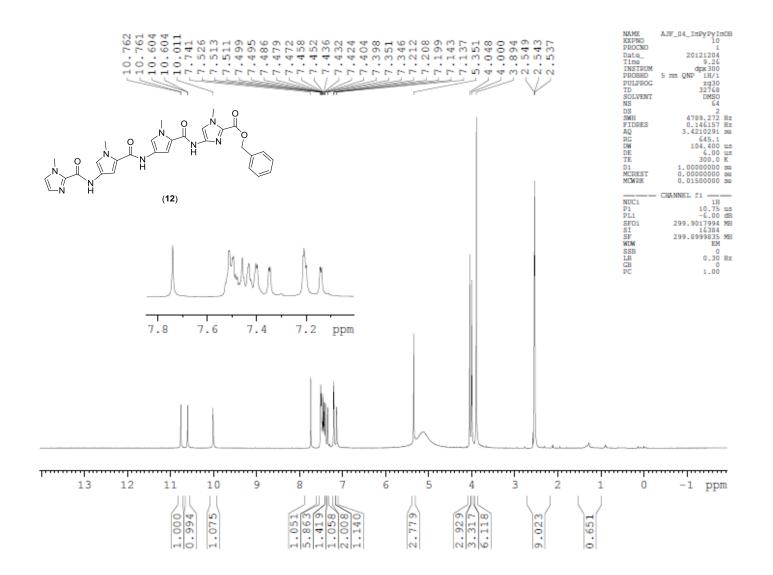


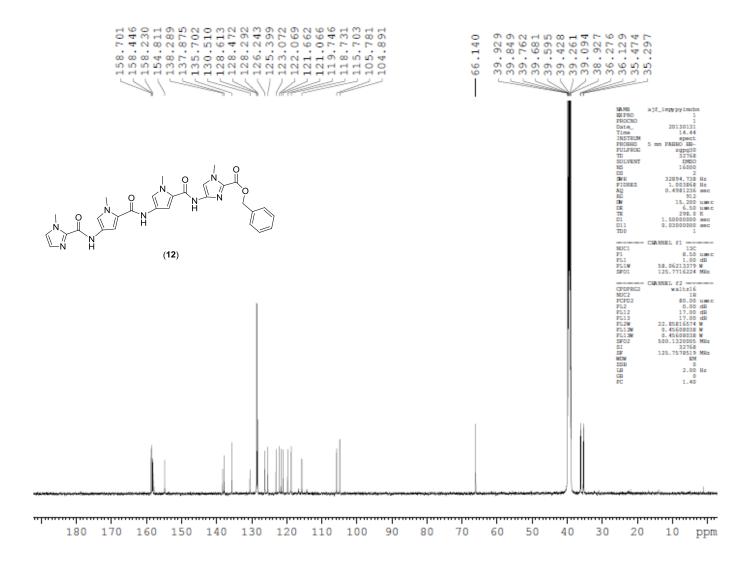


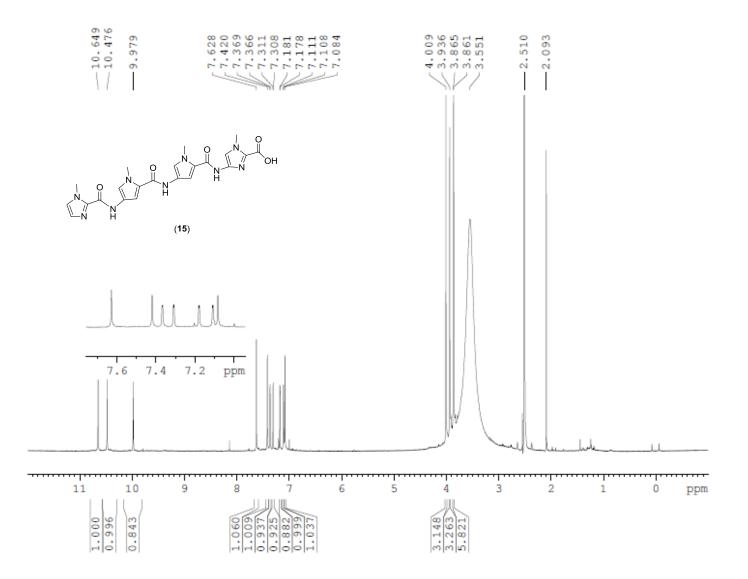


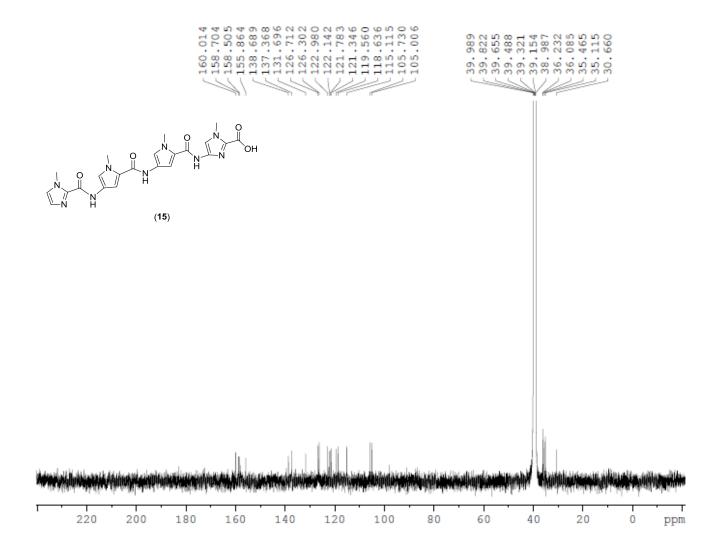












## 7.0 References

- 1. E. E. Baird and P. B. Dervan, J. Am. Chem. Soc., 1996, 118, 6141–6146.
- 2. L. Grehn and U. Ragnarsson, J. Org. Chem., 1981, 46, 3492–3497.
- 3. W. Su, S. J. Gray, R. Dondi, and G. A. Burley, *Org. Lett.*, 2009, **11**, 3910–3.