Supporting Information for

Base-assisted aerobic C–H oxidation of alkylarenes with a murdochitetype oxide Mg₆MnO₈ nanoparticle catalyst

Eri Hayashi, Takatoshi Tamura, Takeshi Aihara, Keigo Kamata,* and Michikazu Hara*

Laboratory for Materials and Structures, Institute of Innovative Research, Tokyo Institute of Technology, Nagatsuta-cho 4259, Midori-ku, Yokohama 226-8503, Japan. Email address: kamata.k.ac@m.titech.ac.jp,

Experimental Section

Solvents (n-octane (Kanto Chemical), chlorobenzene (TCI), and toluene Materials. (Kanto Chemical)) were pretreated with molecular sieves (3A) that were evacuated at 250 °C for 3 h.^{S1} Reagents for catalytst synthesis and characterization $(Mn(OAc)_2 \cdot 4H_2O)$ (Kanto Chemical), $Mg(OAc)_2 \cdot 4H_2O$ (Kanto Chemical). Sr(OAc)₂·0.5H₂O (Kanto Chemical), NaMnO₄·H₂O (Sigma-Aldrich), MnSO₄·5H₂O (Kanto Chemical), DL-malic acid (Kanto Chemical), L-aspartic acid (Kanto Chemical), 5% HCl (Kanto Chemical), KI (Kanto Chemical), MgO (UBE Industries, LTD.), activated MnO₂ (Sigma-Aldrich), Mn₃O₄ (Sigma-Aldrich), MnO (Sigma-Aldrich), and Na₂S₂O₃ (1 M aqueous solution, Kanto Chemical)) and substrates and products for catalytic oxidation (TCI, Kanto Chemical, and Aldrich) were used as-received. Oxygen-¹⁸O₂ (97 atom%) was purchased from ISOTEC. SrMnO₃ and Mn₂O₃ were synthesized by amino-aided method and characterized according to procedures given in the literature.^{S2} High-surface-area β -MnO₂ nanoparticles was synthesized by a lowtemperature crystallization of low-crystallinity layer-type Mn⁴⁺ precursors and characterized according to procedures given in the literature.^{S3} DHA-*d*₄ was synthesized according to a literature procedure and recrystallized.^{S4}

Instruments. X-ray diffraction (XRD) patterns were recorded on a diffractometer (MiniFlex 600, Rigaku; Cu K α , $\lambda = 1.5405$ Å, 40 kV–15 mA) equipped with a high-speed 1-dimensional detector (D/teX Ultra, Rigaku). Diffraction data were collected in the range of $2\theta = 10-80^{\circ}$ in 0.02° steps with a scan rate of 20° min⁻¹. Fourier transform infrared (FTIR) spectra were obtained at a resolution of 4 cm⁻¹ using a spectrometer (FT/IR-6100, Jasco) equipped with an extended KBr beam-splitting device and a mercury cadmium telluride (MCT) and a triglycine sulfate (TGS) detectors.

Raman spectra were recorded on a spectrometer (NRS-3200, Jasco) with excitation at 532 nm using a green laser. Inductively coupled plasma-atomic emission spectroscopy (ICP-AES) analyses were performed with a Shimadzu ICPS-8100 spectrometer.

Nitrogen adsorption-desorption isotherms were measured at -196 °C with a surface area and porosity analyzer (micromeritics TriStar II, Shimazu). Prior to measurement, the samples were heated at 150 °C for 3 h under vacuum to remove physiosorbed water. The Brunauer-Emmett-Teller (BET) surface areas were estimated over the relative pressure (*P*/*P*₀) range of 0.05–0.30. The morphology of the samples was examined using scanning electron microscopy (SEM; S-5500, Hitachi).

Transmission electron microscopy (TEM) observations were conducted using a JEOL JEM2100F microscope operated at an accelerating voltage of 200 kV. After directly mixing Cu grids with samples, the Cu grids were collected and mounted on a stage.

X-ray photoelectron spectroscopy (XPS) analysis were performed with Shimadzu ESCA-3200 spectrometer using Mg K α radiation (1253.6 eV) at 8 kV and 30 mA. Samples were pressed into the pellet and fixed on a double-stick carbon tape. The binding energies were calibrated using the C 1s band at 284.6 eV. The spectrum was fitted and evaluated by the XPS Peak 4.1 program, whereas the background was subtracted using the Shirley function. The deconvoluted Mn 2p spectrum of Mg₆MnO₈ shows three peaks with binding energies of 641.8, 642.8, and 644.5 eV, which correspond to Mn(III), Mn(IV), and the shakeup peak, respectively.^{S5,S6} The deconvoluted O 1s spectrum of Mg₆MnO₈ shows three peaks with binding energies of 529.2, 531.2, and 533.0 eV, which correspond to lattice oxygen, adsorbed oxygen, and adsorbed molecular water, respectively.^{S5,S7}

The average oxidation states of Mn species were determined by iodometric titration using an auto titrator (Mettler Toledo, Easy Pro Titrator System). Mg6MnO8 samples (ca. 20 mg) was added to a mixed solution of 0.5 M HCl aq. (12 mL) and 2 M KI aq. (5 mL), and the resulting solution was titrated with an aqueous solution of 0.01 M Na2S₃O₃.^{S8}

Thermo gravimetry-differential thermal analysis (TG-DTA) measurement was performed with a differential thermal analyzer (Shimadzu DTG-60 differential thermal analyzer). TGA-DTA analysis was carried out at a heating rate of 10 °C min⁻¹ from r.t. to 800 °C under air flow (200 mL min⁻¹).

Temperature-programmed desorption (TPD), H₂ temperature-programmed reduction (H₂-TPR), and CO₂ temperature-programmed desorption (CO₂-TPD) profile were measured on a BEL Japan BELCAT-A equipped with thermal conductivity (TCD)

and mass (Belmass) detectors. A 50 mg amount of sample for TPD and H₂-TPR was placed in a quartz cell and then heated at rate of 10 °C min⁻¹ from 50 °C to 800 °C under He flow (30 mL min⁻¹) or at rate of 10 °C min⁻¹ from 50 °C to 600 °C under 5% H₂/Ar flow (50 mL min⁻¹), respectively. Each catalyst (50 mg) for CO₂-TPD was pretreated under a He flow at 250 °C for 1 h. CO₂ adsorption was carried out over 1 h with 10% CO₂/He at 100 °C followed by purging with He for 30 min. The temperature was linearly increased from 100–250 °C (10 °C min⁻¹).

Liquid-phase catalytic oxidation was performed with an organic synthesizer (ALHB-80&DTC-200HZ-3000, Techno Applications), a liquid phase organic synthesizer (CHEMIST PLAZACP-1000, Sibata), or a block bath shaker (MyBL-100CS, AS ONE). The isolation of products was performed with a single channel automated flash chromatography system (Smart Flash EPCLC AI-580S, Yamazen). Gas chromatography (GC) analyses were performed on a Shimadzu GC-2025 equipped with a stabilwax capillary column (internal diameter = 0.25 mm, length = 30 m) or GC-2014 equipped with an InertCap 17 capillary column (internal diameter = 0.25 mm, length = 30 m) and a flame ionization detector. Mass spectra were recorded on a spectrometer (GCMS-QP2010 SE, Shimadzu) equipped with an InertCap 17MS capillary column (internal diameter = 0.25 mm, length = 30 m) at an ionization voltage of 70 eV.

Synthesis of murdochite-type oxide Mg₆MnO₈ nanoparticles (Mg₆MnO₈-MA) by sol-gel method using malic acid. A low-density amorphous precursor (i.e., an amorphous malate salt with the metal cations linked by carboxylate anions) decomposes starting from 400 °C and the formation of the ramified aggregate of crystalline nanoparticles is observed at an appropriate calcination temperature.^{S2a} In our previous investigation on the sol-gel methods for the synthesis of SrMnO₃ nanoparticles using various carboxylic acids, we found that malic acid and/or aspartic acid were more effective than typical citric acid and that the formation of an amorphous precursor by using dicarboxylic acid such as malic acid and aspartic acid with a low carbon content would be important in the present synthesis method. We separately confirmed the formation of similar Mg6MnO8 nanoparticles using aspartic acid instead of malic acid. Mg6MnO8-MA was synthesized as follows: Mg(OAc)2·4H2O (9.01 g, 42.0 mmol), Mn(OAc)2·4H2O (6.44 g, 7.0 mmol), and DL-malic acid (10.48 g, 73.5 mmol) were dissolved in water (200 mL). The pale pink solution was evaporated to dryness at 70 °C. The resulting white powder was dried at 190 °C for 1 h to give a pale-yellow powder, which is referred to as the precursor. This precursor was calcined at 550 °C for 5 h to give an analytically pure Mg₆MnO₈-MA. Yield: 2.23 g (97%). Elemental analysis:

calcd (%) for Mg₆MnO₈: Mg 44.4, Mn 16.7; found: Mg 38.0, Mn 14.7.

Procedure for oxidation with O_2 as the sole oxidant catalyzed by Mg₆MnO₈-MA. The catalytic oxidation of alkylarenes was conducted in a 20 mL Schlenk-type glass tube containing a magnetic stir bar. Typical procedures are as follows: Mg₆MnO₈-MA (0.1 g) was evacuated at 200 °C for 1 h in a Schlenk tube flowed by cooling down to room temperature. Then, **1a** (0.5 mmol), *n*-octane (2 mL), and an internal standard (naphthalene) were charged into the Schlenk tube under Ar atmosphere. After the replacement of Ar gas with O₂ gas (1 atm), the reaction solution was heated at 50 °C and periodically analyzed using GC. The products are known and were identified by comparison of their GC retention times, MS spectrum, and ¹H and ¹³C NMR signals with the literature data. After the reaction was completed, the catalyst was recovered by filtration. The recovered Mg₆MnO₈ was washed with *n*-octane and then calcined at 400 °C for 1.5 h before recycling. In some cases, the products were isolated using a flash chromatography separation system with a silica gel column.

A larger-scale (10 mmol scale) production of 2a. The larger-scale catalytic oxidation of 1a into 2a was conducted in a 200 mL Schlenk-type glass tube containing a magnetic stir bar. Mg6MnO8-MA (0.5 g) was evacuated at 200 °C for 3 h in a Schlenk tube flowed by cooling down to room temperature. Then, 1a (10 mmol) and *n*-octane (40 mL) were charged into the Schlenk tube under Ar atmosphere. After the replacement of Ar gas with O₂ gas (1 atm), the reaction solution was heated and stirred at 70 °C for 10 h to give 2a in 88% GC yield. After cooling the reactor at room temperature, catalyst was removed by filtration, and then solvents were removed by evaporation. The analytically pure 2a (1.49 g (83% yield), \geq 99% purity by ¹H NMR) was isolated using a flash chromatography separation system with a silica gel column (AcOEt:hexane = 0:10 to 10:0 eluents). The amounts of surface Mn species of MgMnO8-MA were estimated to be 490 µmol g⁻¹ based on the BET surface area (104 m² g⁻¹) assuming that the (100) plane is a possible surface structure. The TOF value of MgMnO8-MA was estimated to be 54 h⁻¹ under the reaction conditions of eq 1.

Procedure for competitive oxidation of 1a and 9-fluorenol with O₂ as the sole oxidant catalyzed by Mg₆MnO₈-MA. The competitive oxidation of 1a and 9-fluorenol was conducted in a 20 mL Schlenk-type glass tube containing a magnetic stir bar. The procedures are as follows: Mg₆MnO₈-MA (0.1 g) was evacuated at 200 °C for 1 h in a Schlenk tube flowed by cooling down to room temperature. Then, 1a (0.25 mmol), 9-

fluorenol (0.25 mmol), chlorobenzene (5 mL), and an internal standard (naphthalene) were charged into the Schlenk tube under Ar atmosphere. After the replacement of Ar gas with O₂ gas (1 atm), the reaction solution was heated at 50 °C and periodically analyzed using GC. The reaction rates for the oxidation of 9-fluorenol to **2a** and **1a** to **2a** were 3.39 and 1.78×10^{-1} mM min⁻¹, respectively.

¹⁸O-Labeling Experiments. The oxidation of 1a with ¹⁸O₂ was carried out according to the literature procedures.^{S2} Before the reaction, the catalyst was evacuated at 200 °C for 1 h and *n*-octane was degassed by freeze-pump-thaw cycling. The ¹⁸O-Labeling experiment was carried out in a schlenk tube containing a magnetic stir bar. Catalyst (0.1 g), 1a (0.5 mmol), *n*-octane (2 mL), and an internal standard (naphthalene) were charged into the schlenk tube under Ar atmosphere, followed by the evacuation and the introduction of 97% ¹⁸O₂ (1 atm). The reaction solution was heated at 50 °C and periodically analyzed using GC and GC-MS. The ¹⁸O content in 2a was determined by using the ratio of the peak intensity at m/z = 182 to the sum of the peak intensities at m/z= 180 and 182.

Procedure for IR measurements. The basicity on Mg₆MnO₈-MA, SrMnO₃, and β -MnO₂ catalysts were estimated from FT-IR measurements for chloroform-adsorbed samples at 25 °C.^{S9} Samples were pressed into self-supporting disks (10 mm diameter, 7 mg and 7.8 mg for Mg₆MnO₈-MA and SrMnO₃, respectively; 20 mm diameter, 21 mg for β -MnO₂) and placed in an IR cell attached to a closed glass-circulation system. Prior to chloroform adsorption, the sample was dehydrated by heating at 200 °C for 1 h under vacuum. A dehydrated sample was exposed to chloroform vapor, and Figure 5c shows IR spectra for Mg₆MnO₈-MA, SrMnO₃, and β -MnO₂ at gas-phase chloroform pressures of 1.1×10^{-1} , 1.1×10^{-1} , and 1.2×10^{-1} kPa, respectively.

Data of products

9-Fluorenone (2a): ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 7.58-7.55$ (m, 2H), 7.40– 7.35 (m, 4H), 7.23–7.17 (m, 2H). ¹³C{¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 193.7$, 144.3, 134.6, 134.0, 128.9, 124.1, 120.2. MS (EI): *m/z* (%): 181 (15), 180 (*[M]*+, 100), 153 (6), 152 (44), 151 (21), 150 (13), 126 (7), 76 (15), 75 (6), 63 (7). **2-Fluoro-9-fluorenone (2b)**^{S10}: ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = \delta$ 7.64 (d, J = 7.3 Hz, 1H), 7.50–7.45 (m, 3H), 7.33 (dd, J = 7.6 Hz, J = 2.3 Hz, 1H), 7.29–7.26 (m, 1H), 7.15 (td, J = 8.5 Hz, J = 2.4 Hz, 1H); ¹³C{¹H} (CDCl₃, 100MHz, TMS): $\delta = 192.6$, 163.8 (d, J = 248.3 Hz), 144.1, 140.3, 136.5 (d, J = 7.2 Hz), 135.2, 134.5, 128.9, 124.8, 121.7 (d, J = 7.9 Hz), 121.0 (d, J = 23.1 Hz), 120.2, 112.1 (d, J = 23.5 Hz); ¹⁹F{¹H} (CDCl₃, 376.4 MHz): $\delta = -112.7$. MS (EI): m/z (%): 199 ($[M+1]^+$, 15), 198 ($[M]^+$, 100), 171 (5), 170 (41), 169 (15), 168 (9), 150 (6), 144 (7), 99 (7), 85 (21), 75 (5), 72 (6)

2-Bromo-9-fluorenone (2c)^{S10}: ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 7.75$ (d, J = 1.3 Hz, 1H), 7.65 (d, J = 7.4 Hz, 1H), 7.60 (dd, J = 7.9 Hz, J = 1.7 Hz, 1H), 7.52–7.47 (m, 2H), 7.38 (d, J = 7.9 Hz, 1H), 7.34–7.28 (m, 1H); ¹³C{¹H} (CDCl₃, 100 MHz, TMS): $\delta = 192.4$, 143.9, 143.2, 137.2, 136.0, 135.2, 133.9, 129.6, 127.7, 124.8, 123.1, 121.8, 120.6. MS (EI): m/z (%): 261 ($[M+2]^+$, 14), 260 ($[M+1]^+$,95), 259 ($[M]^+$, 15), 258 ($[M-1]^+$, 100), 232 (5), 230 (6), 179 (7), 152 (12), 151 (93), 150 (49), 149 (5), 125 (8), 116 (11), 115 (11), 99 (7), 98 (7), 90 (7), 77 (5), 76 (9), 75 (42), 74 (10), 62 (9).

2,7-Dibromo-9-fluorenone (2d)^{S11}: ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 7.77$ (d, J = 1.2 Hz, 2H), 7.62 (dd, J = 7.9 Hz, 1.6 Hz, 2H), 7.38 (d, J = 7.9 Hz, 2H); ¹³C {1H} (CDCl₃, 100 MHz, TMS): $\delta = 191.0$, 142.5, 137.6, 135.5, 128.0, 123.5, 122.0. MS (EI): m/z (%): 341 ($[M+3]^+$, 7), 340 ($[M+2]^+$, 49), 339 ($[M+1]^+$, 15), 338 ($[M]^+$, 100), 337 ($[M-1]^+$, 8), 336 ($[M-2]^+$, 52), 231 (23), 229 (24), 169 (5), 155 (8), 150 (75), 151 (10), 149 (10), 124 (5), 123 (6), 115 (10), 114 (10), 100 (5), 99 (8), 98 (10), 76 (6), 75 (44), 74 (18), 62 (10).

11-*H***-Benzo[***a***]fluoren-11-one (2e)^{S12}: ¹H NMR (400 MHz, CDCl₃, TMS): \delta = 8.93 (d, J=8.5Hz, 1H), 7.95 (d, J = 8.2Hz, 1H), 7.75 (d, J=8.3Hz, 1H), 7.62–7.54 (m, 3H), 7.46–7.39 (m, 3H), 7.24 (t, J=7.2Hz, 1H); ¹³C{¹H} (CDCl₃, 100 MHz, TMS): \delta = 195.4, 146.3, 144.0, 136.0, 134.8, 134.6, 134.3, 130.3, 129.5, 129.4, 128.6, 127.0, 126.5, 124.5, 123.9, 120.1, 118.2. MS (EI):** *m/z* **(%): 231 (***[M+1]***⁺, 19), 230 (***[M]***⁺, 100), 202 (23), 201 (13), 200 (19), 115 (7), 100 (16), 101 (24), 88 (14), 87 (6).**

Xanthone (2f): ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 8.34$ (dd, J = 7.6 and 1.6 Hz, 2H), 7.74–7.70 (m, 2H), 7.50–7.48 (m, 2H), 7.39–7.35 (m, 2H); ¹³C{¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 177.5$, 156.5, 135.1, 127.1, 124.2, 122.2, 118.3. MS (EI): m/z (%): 197 (14), 196 ($[M]^+$, 100), 169 (6), 168 (47), 140 (9), 139 (46), 84 (9), 69 (6), 63 (8).

Thioxanthone (2g)^{S13}: ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 8.62$ (d, J = 8.0 Hz, 2H), 7.63–7.56 (m, 4H), 7.48 (td, J = 7.4 Hz, J = 1.0 Hz, 2H); ¹³C {¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 180.1$, 137.4, 132.4, 130.0 129.5, 126.3 (d, J = 32). MS (EI): m/z (%): 214 ($[M+2]^+$, 6), 213 ($[M+1]^+$, 15), 212 ($[M]^+$, 100), 185 (8), 184 (60), 183 (8), 152 (16), 139 (26), 108 (7), 106 (6), 92 (18), 91 (7), 82 (5), 79 (14), 69 (10), 63 (5).

Anthraquinone (2h): ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 8.34-8.30$ (m, 4H), 7.83–7.79 (m, 44H); ¹³C{1H} NMR (100 MHz, CDCl₃, TMS): $\delta = 183.3$, 134.3, 133.7, 127.4. MS (EI): m/z (%): 209 ($[M+1]^+$, 15), 208 ($[M]^+$, 100), 207 ($[M-1]^+$, 15), 181 (12), 180 (80), 179 (6), 153 (9), 152 (74), 151 (37), 150 (19), 126 (11), 90 (9), 77 (9), 76 (53), 75 (19), 74 (10), 63 (14), 51 (5), 50 (16),

Dibenzosuberone (2i)^{S14}: ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 8.01$ (d, J = 7.8 Hz, 2H), 7.42 (t, J = 7.4 Hz, 2H), 7.33 (t, J = 7.4 Hz, 2H), 7.22 (d, J = 7.5 Hz, 2H), 3.21 (s, 4H); ¹³C{¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 195.7$, 142.1, 138.8, 132.5, 130.7, 129.4, 126.8, 35.1. MS (EI): m/z (%): 209 ($[M+1]^+$, 16), 208 ($[M]^+$, 100), 207 ($[M-1]^+$, 38), 193 (10), 181 (9), 180 (57), 179 (65), 178 (53), 177 (9), 176 (13), 166 (6), 165 (39), 152 (19), 151 (9), 115 (5), 103 (8), 102 (5), 90 (13), 89 (44), 88 (10), 83 (5), 77 (12) 76 (24), 75 (6), 63 (12), 51 (8), 39 (6).

Benzophenone (2j): ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 7.82-7.79$ (m, 4H), 7.61–7.57 (m, 2H), 7.51–7.46 (m, 4H). ¹³C{¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 197.0$, 138.0, 132.7, 130.4, 128.6. MS (EI): m/z (%): 183 (7), 182 ([M]+, 47), 181 ([M-1]⁺, 8), 106 (8), 105 (100), 78 (18), 77 (56), 76 (5), 51(18), 50 (7).

2-Benzoylpyridine (**2k**)^{**S**15}: ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 8.72$ (d, J = 4.6 Hz, 1H), 8.08–8.03 (m, 3H), 7.90 (t, J = 7.7 Hz, 1H), 7.59 (t, J = 7.4 Hz, 1H), 7.50–7.46 (m, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 193.9$, 155.4, 148.7, 137.1, 136.5, 133.0, 131.1, 128.3, 126.2, 124.7. MS (EI): m/z (%): 183 ($[M]^+$, 18), 182 ($[M-1]^+$, 42), 156 (10), 155 (83), 154 (28), 127 (7), 106 (7), 105 (86), 78 (17), 77 (100), 76 (6), 52 (7), 51 (39), 50 (10).

3-Benzoylpyridine (21)^{S16}: ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 9.00$ (s, 1H), 8.82 (d, J = 4.8 Hz, 1H), 8.12 (d, J = 7.8 Hz, 1H), 7.82 (d, J = 7.8 Hz, 2H), 7.64 (t, J = 7.3 Hz, 1H), 7.52 (t, J = 7.6 Hz, 2H), 7.45 (dd, J = 7.5, 4.9 Hz, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 195.0$, 153.0, 151.1, 137.3, 137.0, 133.4, 133.3, 130.1 128.8, 123.5.

MS (EI): *m/z* (%): 184 (*[M+1]*⁺, 10), 183 (*[M]*⁺, 73), 182 (*[M-1]*⁺, 28), 154 (6), 106 (26), 105 (100), 78 (33), 77 (73), 51 (37), 50 (10).

4-Benzoylpyridine (2m)^{S17}: ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 8.81$ (d, J = 5.8 Hz, 2H), 7.82 (d, J = 7.3 Hz, 2H), 7.65 (t, J = 7.4 Hz, 1H), 7.58 (d, J = 5.8 Hz, 2H), 7.52 (t, J = 7.6 Hz, 2H); ¹³C{¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 195.2$, 150.5, 144.6, 136.1, 133.6, 130.3, 128.8, 123.0. MS (EI): m/z (%): 184 ($[M+1]^+$, 5), 183 ($[M]^+$, 35), 106 (13), 105 (100), 78 (14), 77 (63), 51 (29), 50 (7).

Benzil (2n): ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 7.98$ (d, J = 7.8 Hz, 4H), 7.66 (t, J = 7.4 Hz, 2H), 7.51 (t, J = 7.7 Hz, 4H); ¹³C{¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 194.7$, 135.0, 133.3, 130.0, 129.2. MS (EI): m/z (%): 211 ($[M+1]^+$, 0.4), 106 (11), 105 (100), 77 (60), 51 (19).

Indole (20): ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 8.08$ (br, 1H), 7.64 (d, J = 7.8, 1H), 7.38 (d, J = 8.1 Hz, 1H), 7.21–7.10 (m, 3H), 6.55 (s, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 136.0$, 128.1, 124.2, 122.2, 120.9, 120.0, 111.1, 102.8. MS (EI): m/z (%): 117 ($[M]^+$, 100), 90 (46), 89 (32), 59 (12), 63 (11), 118 (10), 116 (8), 39 (5).

Phthalide (2p)^{S13}: ¹H NMR (400 MHz, CDCl₃, TMS): $\delta = 7.93$ (d, J = 7.7 Hz, 1H), 7.69 (t, J = 7.5 Hz, 1H), 7.56–7.49 (m, 2H), 5.33 (s, 2H); ¹³C{¹H} NMR (100 MHz, CDCl₃, TMS): $\delta = 171.1$, 146.7, 134.1, 129.2, 126.0, 122.2, 69.7. MS (EI): m/z (%): 134 ($[M]^+$, 32), 133 ($[M-1]^+$, 12), 106 (8), 105 (100), 77 (44), 76 (10), 51 (15), 50 (12).

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Entry	Catalyst/additive	Oxidant	Solvent	Temp. (°C)	Time (h)	Yield (%)	Reaction rate ^{<i>a</i>} (mmol $g_{cat}^{-1} h^{-1}$)	Ref.
			Mn-based catalyst					
	SrMnO ₃ (aspartic acid-aided method)	O ₂ (1 atm)	<i>n</i> -octane	60	24	96	0.2	S2(<i>a</i>)
	SrMnO ₃ (polymerize complex method)	O ₂ (1 atm)	<i>n</i> -octane	80	12	93	0.4	S18
	MnO ₂ @wool	air (1 atm)	o-xylene	reflux	9	99	1.0	S19
	MnO ₂ /porous chitosan	air	<i>p</i> -xylene	100	4	95	5.9	S20
	Mo-MnO ₂	O ₂ (5 atm)	o-dichlorobenzene	150	6	90	1.5	S21
	Ni-MnO _x	O ₂ (1 atm)	<i>n</i> -octane	80	1	99	9.9	S22
	$Mn_{0.5}Ce_{0.5}O_x$	O ₂ (10 atm)	CH ₃ CN	110	4	82	6.8	S23
d	Mn-MOF-74	$O_2 (5 \text{ mL min}^{-1})$	<i>m</i> -dichlorobenzene	135	9	24	19.6	S24
е	Ni ₂ Mn-LDH	O ₂ (1 atm)	dodecane	120	3.5	81	1.2	S25
0 ^f	OMS-2	air (1 atm)	isooctane	reflux	4	99	5.0	S26
1	LaMnO ₃	O ₂ (1 atm)	heptane	80	24	96	0.1	S27
2 ^g	MnAlPO-5-meso	O ₂ (5 atm)	CH ₃ CN	120	5	24	1.0	S28
			metal-based catalyst	1				
3 ^g	FeAlPO-5-meso	O ₂ (5 atm)	CH ₃ CN	120	5	15	0.6	S28
4^h	CuTSPc@3D-(N)GFs	air (1 atm)	H ₂ O	r.t.	12	96	16.0	S29
5	Cu-doped Co ₃ O ₄	O ₂ (10 atm)	cyclohexane	90	4	99	5.0	S30
6 ^{<i>i</i>}	NCC-PC	air (1 atm, bubbling)	H ₂ O	r.t.	10	88	0.3	S31
7	Co-N-C nanocomposite	O ₂ (1 atm, balloon)	DMSO	140	20	81	25.3	S32
8 ^j	CrCoFeO4@G–GO	air (1 atm)	o-xylene	reflux	4	96	2.0	S33
9^k	Co–NHAp	air	<i>p</i> -xylene	100	8	85	0.6	S34
0^l	CrMCM-41	O_2 (1 atm)	_	95	24	23	2.9	S35

Table S1. Additive-free Heterogeneous Catalyst Systems for the Oxidation of Fluorene (1a) with O₂ as the sole oxidant

21	BaFeO _{3-δ}	O ₂ (1 atm)	<i>n</i> -octane	80	96	59	0.03	S2(c)
22^{m}	NiAl HT	$O_2 (5 \text{ mL min}^{-1})$	_	135	5	58	5.8	S36
23 ^{<i>n</i>}	CNB _{0.15}	O ₂ (10 atm)	CH ₃ CN	130	24	45	0.4	S37
24 ^o	AuNPs@resin	O ₂ (1 atm)	CH ₃ CN	80	20	30	0.1	S38
25	Ru ³⁺ -substituted silicotungstate	O ₂ (1 atm)	isobutyl acetate	110	96	10	0.7	S39
26	Ru(OH) _x /Al ₂ O ₃	O ₂ (1 atm)	o-dichlorobenzene	170	4	93	0.9	S40
27	Ir/TiO ₂	O ₂ (1 atm)	Mesitylene	150	50	62	0.2	S41
			carbon-based catalys	st				
28^p	g-C ₃ N ₄	O ₂ (1 atm, balloon)	H_2O	r.t. + blue LED	24	62	0.3	S13
29	activated carbon (Shirasagi KL)	O ₂ (?)	<i>m</i> -xylene	120	24	83	0.2	S42

^{*a*}Reaction rate per gram (mmol h⁻¹ g⁻¹) = (**2a** (mmol)) / time (h) × catalyst (g). ^{*b*}The reaction rate per gram could not be estimated because of no information about the amount of catalyst. ^{*c*}The reaction rate per gram could not be estimated because of no information about the amount of **1a**. ^{*d*}MOF = metal organic frameworks. ^{*e*}LDH = layered double hydroxide. ^{*f*}OMS-2 = Cryptomelane-type manganese oxide. ^{*g*}ALPO = aluminophosphate. ^{*h*}TSPc = tetrasulfophthalocyanine, 3D-(N)GFs = three-dimensional nitrogen-doped graphene-based frameworks. ^{*i*}NCC-PC = nanocrystalline cellulose-anionic copper tetrasulfonate phthalocyanine. ^{*j*}CrCoFeO4@G–GO = Cr-substituted cobalt ferrite nanoparticles supported on guanidine-grafted graphene oxide nanosheets. ^{*k*}NHAp = natural hydroxyapatite. ^{*i*}MCM-41= Mobil crystalline material 41. ^{*m*}HT = hydrotalcite. ^{*n*}CNB = boron-doped polymeric carbon nitride. ^{*o*}NPs = nanoparticles. ^{*p*}g-C₃N₄ = graphitic carbon nitride.

Entry	Synthetic method	Reagents	Surface area $(m^2 g^{-1})$	Application	Ref.
1	sol-gel method (400 °C, 2 h \rightarrow 700 °C, 6 h)	$\frac{Mg(OAc)_2 \cdot 4H_2O}{Mn(OAc)_2 \cdot 4H_2O}$	15	catalyst (N ₂ O decomposition)	S43
2	sol-gel method (400 °C, 2 h, $O_2 \rightarrow 700$ °C, 6 h, O_2)	$\begin{array}{c} Mg(OAc)_2 \cdot 4H_2O, \\ Mn(OAc)_2 \cdot 4H_2O \end{array}$	15	catalyst (CO oxidation)	S44
3	sol-gel method (250 °C, 2 h \rightarrow 750 °C, 12 h)	$\begin{array}{c} Mg(OAc)_2 \cdot 4H_2O, \\ Mn(OAc)_2 \cdot 4H_2O \end{array}$	_	_	S45
4	sol-gel method (500–800 °C, 6 h, O ₂)	$Mg(OAc)_2 \cdot 4H_2O$, acetic acid, $Mn(OAc)_2 \cdot 4H_2O$	_	-	S46
5	impregnation method (1000 °C, ? h)	Mn(NO ₃) ₂ , MgO	_	catalyst (N ₂ O decomposition)	S47
6	incipient wetness impregnation method (450 °C, 3 h \rightarrow 900 °C, 8 h)	Mn(NO ₃) ₂ , MgO	4	catalyst (oxidative dehydrogenation of ethane)	S48
7	incipient wetness impregnation method (450 °C, 3 h \rightarrow 900 °C, 8 h)	Mn(NO ₃) ₂ , MgO	7	catalyst (selective hydrogen combustion)	S49
8	incipient wetness impregnation method (800 °C, 4 h)	$Mn(NO_3)_2 \cdot 4H_2O, MgO$	_	catalyst (oxidative coupling of methane)	S50
9	incipient wetness impregnation method (600 °C, 3 h)	$Mn(NO_3)_2 \cdot 6H_2O, MgOH$	_		S51
10	solid phase method (900 °C, 12 h)	MgO, MnO ₂	_	Mg-ion batteries	S52
11	solid phase method (900 °C, 12 h)	MgO, MnO ₂	2	CO ₂ capture	S53
12	solid phase method	MgO, MnO ₂	_	oxygen sensor	S54
13	solid phase method (1100, 5 h, O ₂)	Mn(CO ₃), MgO	_	_	S55
14	coprecipitation method (700 or 800 °C, few hours)	$Mg(OAc)_2 \cdot 4H_2O, Mn(OAc)_2 \cdot 4H_2O$	_	_	S 56

Table S2.Surface area, preparation method, and regent of reported Mg6MnO8

Table S3. Comparison of surface physicochemical properties of manganese oxides						
	Mg6MnO8-MA	SrMnO ₃ ^b	β -MnO ₂ ^c			
Oxidation states of surface Mn species ^a	3.7	3.6	3.5			
Amounts of adsorbed oxygen species $(\%)^a$	33	38	29			
$S_{\rm BET}~({ m m}^2~{ m g}^{-1})$	104	47	106			

Table S3. Comparison of surface physicochemical properties of manganese oxides

^{*a*}Estimated by XPS analysis. ^{*b*}Data from ref. S2(*a*). ^{*c*}Data from ref. S3(*b*).

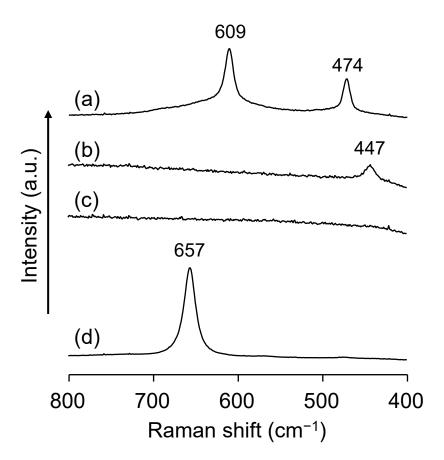


Figure S1. Raman spectra for (a) Mg6MnO8-MA, (b) MgO, (c) activated MnO2 (Aldrich), and (d) Mn2O3.

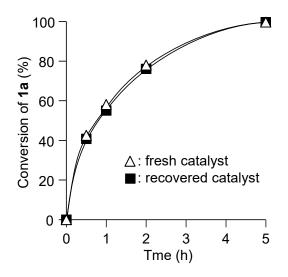


Figure S2. Time course for the oxidation of **1a** with O₂ catalyzed by fresh and recovered Mg₆MnO₈-MA. Reaction conditions: catalyst (0.1 g), **1a** (0.5 mmol), *n*-octane (2 mL), pO_2 (1 atm), 50 °C.

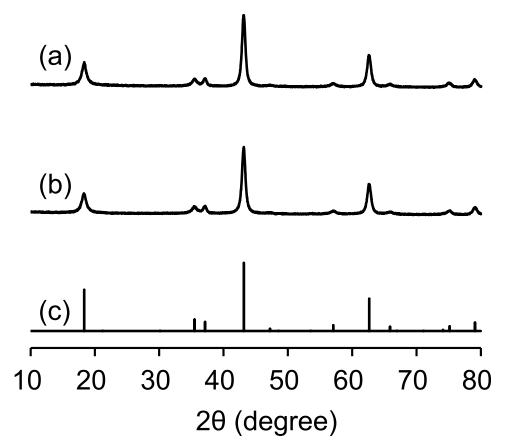


Figure S3. XRD patterns for (a) fresh Mg₆MnO₈-MA, (b) recovered Mg₆MnO₈-MA after the catalytic oxidation of **1a**, and (c) Mg₆MnO₈ (JCPDS 01-073-2156). The reaction conditions are the same as those in Figure 4b.

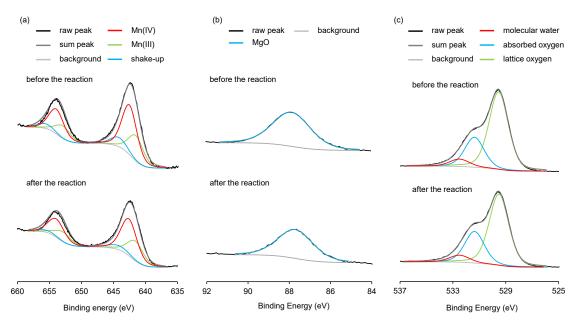


Figure S4. XPS (a) Mn 2p, (b) Mg 2s, (c) O 1s spectra of fresh Mg₆MnO₈-MA and recovered Mg₆MnO₈-MA after the catalytic oxidation of 1a. The reaction conditions are the same as those in Figure 4b.

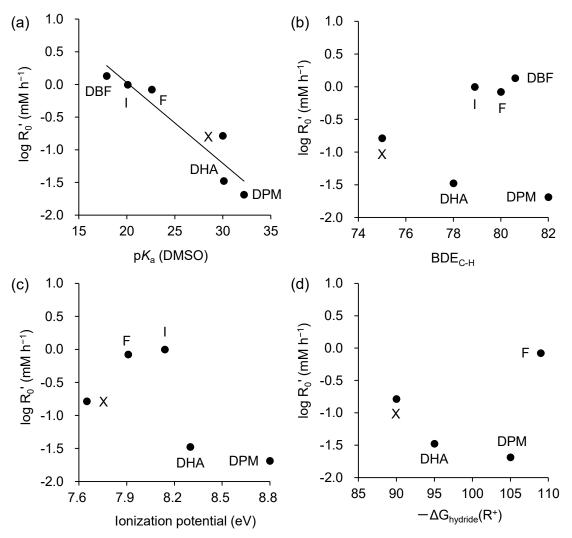


Figure S5. Log R_0 ' for the oxidations of alkylarenes (xanthene (X), 9,10dihydroanthracene (DHA), indene (I), diphenylmethane (DPM), 2,7-Dibromo-9fluorenone (DBF) and fluorene (F)) as a function of (a) pK_a , (b) homolytic bond dissociation energy (c) ionization potential, and (d) heterolytic bond dissociation energy. Reaction conditions were as follows: Alkylarene (0.3 mmol), Mg₆MnO₈ (30 mg), chlorobenzene (4 mL), 50 °C, pO_2 (1 atm). The initial rates (-d[substrate]/dt, R₀) were determined from the slopes of reaction profiles ([substrate]o-[substrate]t vs. time plots) at low conversions (<20%) of the substrates. Data of pK_a are taken from references S57, S58, and S59. BDE is the free energy of R-H homolytic bond disassociation. Data of BDE are taken from refs. S57, S58, and S59. $-\Delta G_{hydride}(R^+)$ is the free energy of R-H heterolytic bond disassociation. Data of $-\Delta G_{hydride}(R^+)$ are from refs. S60 and S61. Date of ionization potential are from NIST at http://webbook.nist.gov/chemistry.