

Supporting Information

Quantitative Perspective on Online Flow Reaction Profiling Using a Miniature Mass Spectrometer

Huaming Sheng^{1*}, Emily B. Corcoran^{2*}, Zachary E. X. Dance³, Joseph P. Smith³, Zhihao Lin⁴, Victoria Ordsmith⁵, Simon Hamilton³, Ping Zhuang³

¹*Analytical Science, Merck & Co., Inc., Rahway, New Jersey 07065, United States*

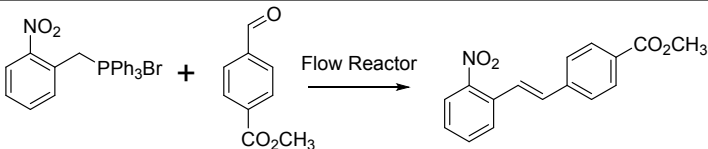
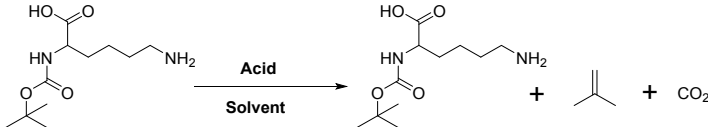
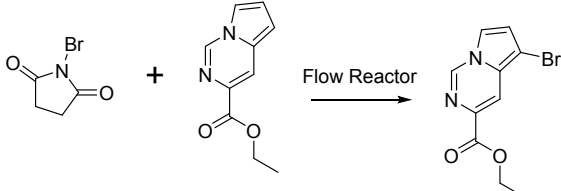
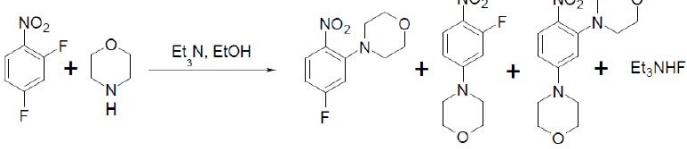
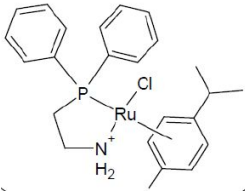
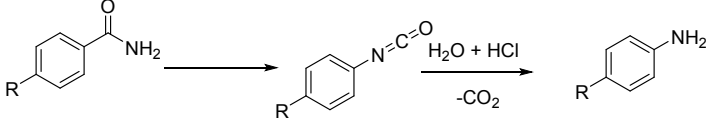
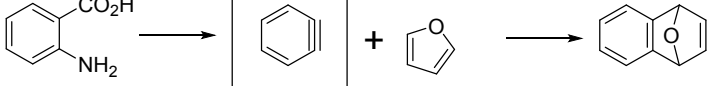
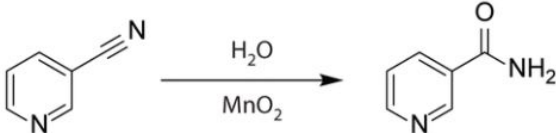
²*Small Molecule Process Research & Development, Merck & Co., Inc., Boston, Massachusetts 02115, United States*

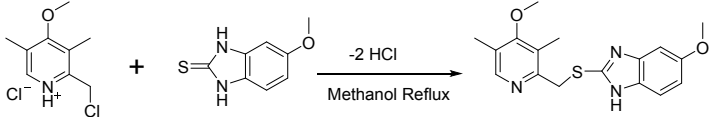
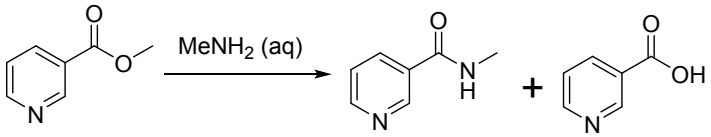
³*Analytical Research & Development, Merck & Co., Inc., Rahway, New Jersey 07065, United States*

⁴*ACDS-PAT, Merck & Co., Inc., Rahway, New Jersey 07065, United States*

⁵*Microsaic Systems plc., Woking GU21 5BX, U.K.*

Table S1. Summary of recent literature of mini-MS reaction monitoring showing the substrates, reaction types and profiling methods

Ionization Source	MS Type	Substrates and Reactions	Reaction Profiling Method	Reference
Chip-ESI	Mini-MS	 <p>Wittig Reaction</p>	Monitoring SM and Product	[1]
Chip-ESI	Mini-MS	 <p>Hydrolysis Reaction</p>	Monitoring SM and Product	[2]
Chip-ESI	Mini-MS	 <p>Bromination Reaction</p>	Monitoring SM and Product	[3]
Chip-ESI	Mini-MS	 <p>S_NAr Reaction</p>	Monitoring SM and Product	[4]
Chip-ESI	Mini-MS	 <p>Catalytical Conversion of Ethanol into biofuel</p>	Only Monitoring Catalyst	[5]
Chip-ESI	Mini-MS	 <p>Hofmann Rearrangement</p>	Monitoring SM, Intermediate and Product	[6]
Chip-ESI	Mini-MS	 <p>Diels-Alder Reaction</p>	Monitoring SM, Intermediate and Product	[7]
ESI	Mini-MS		Monitoring SM and Product	[8]

		Cyano-hydrolysis		
ESI	Mini-MS	 <p>SN2 Reaction</p>	Monitoring SM and Product	[9]
ESI	Mini-MS	 <p>Ester Hydrolysis</p>	Monitoring SM and Product	[10]

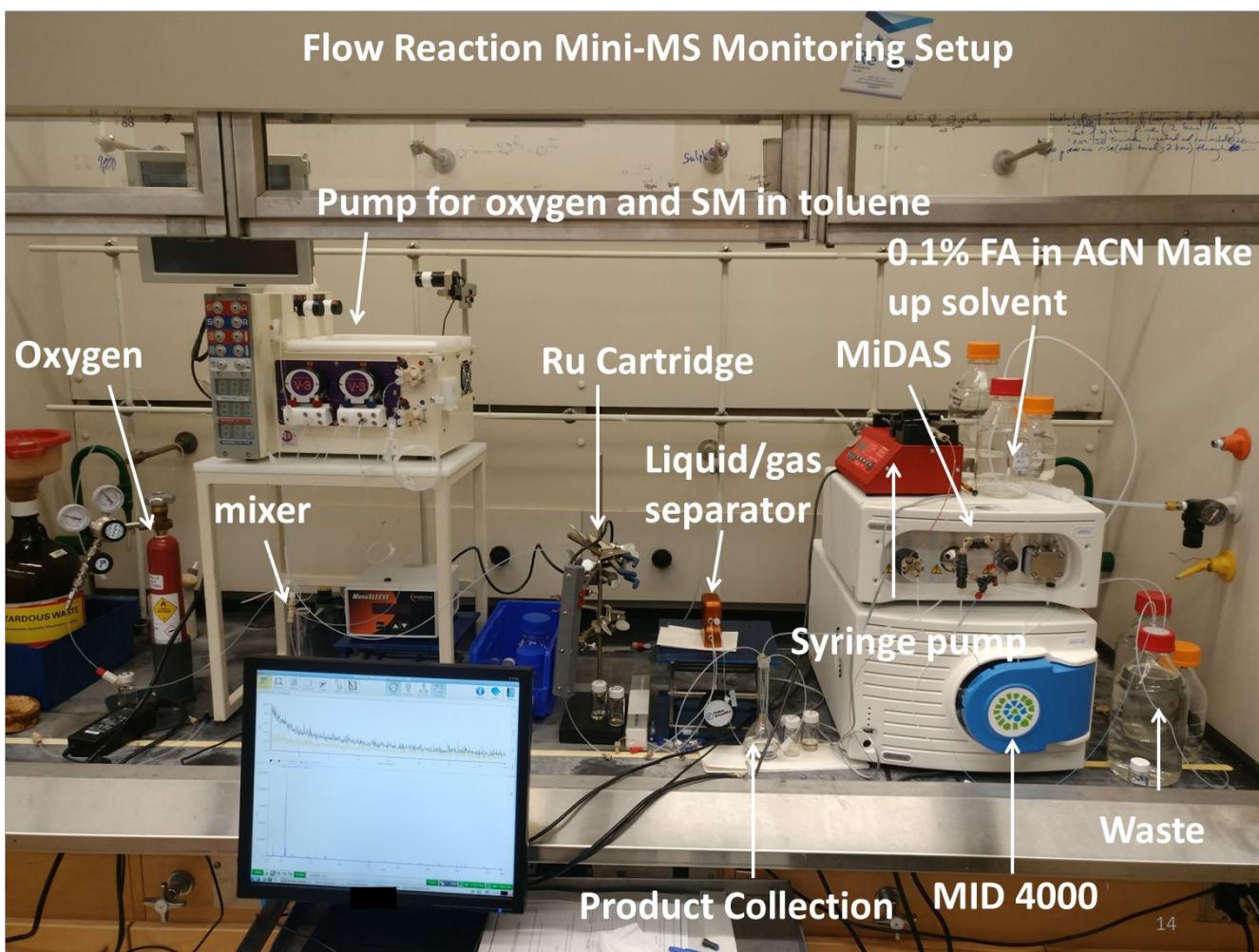


Figure S1. The flow reaction setup with Microsaic miniature MS 4000 MiD and online dilutor MiDAS.

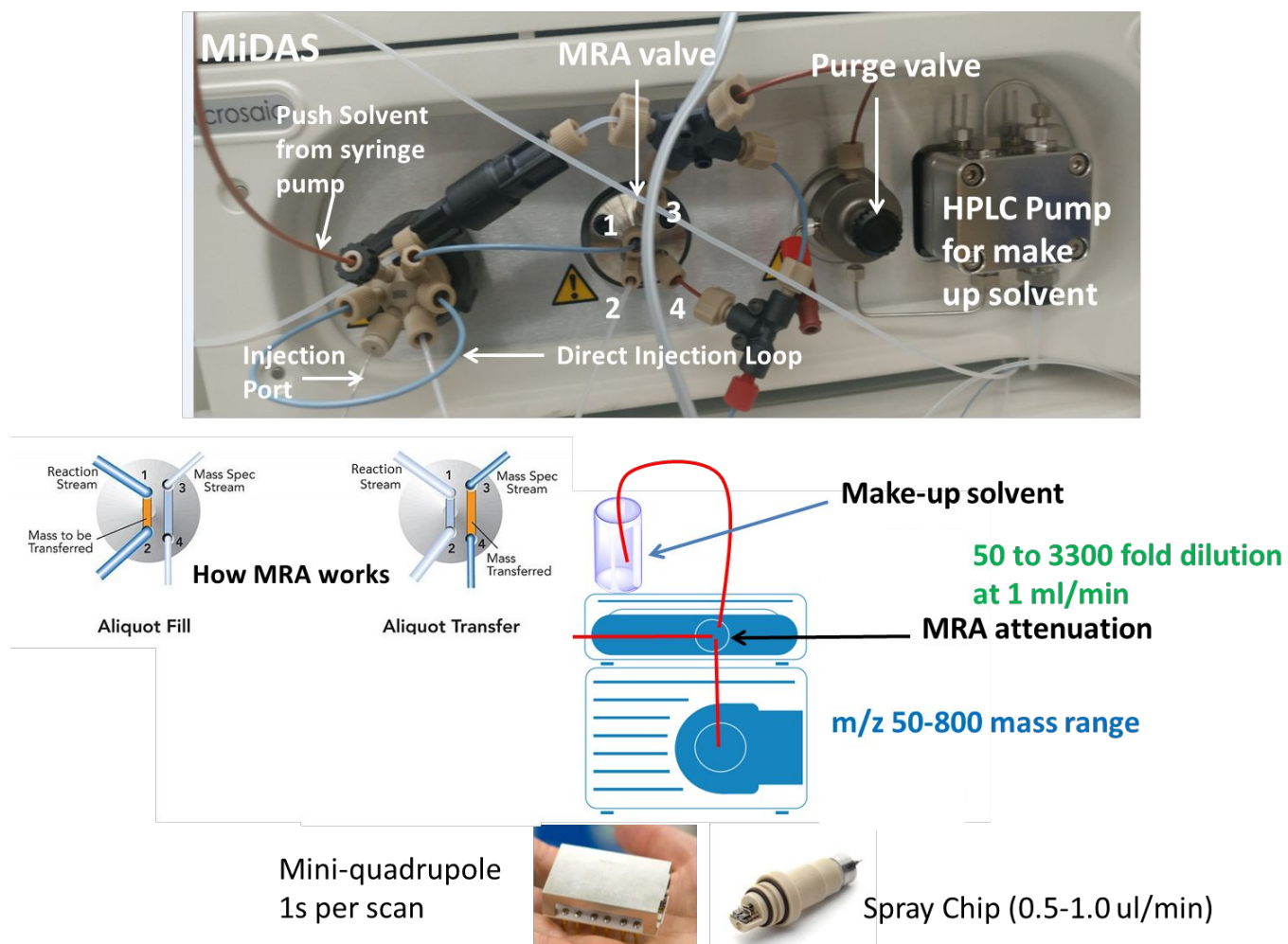
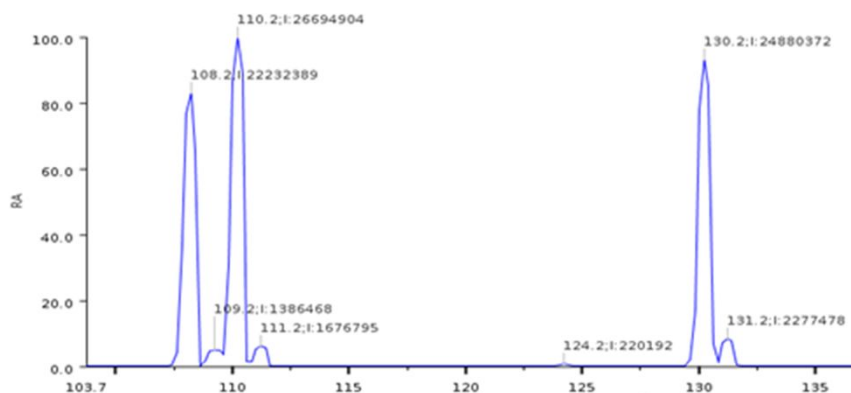


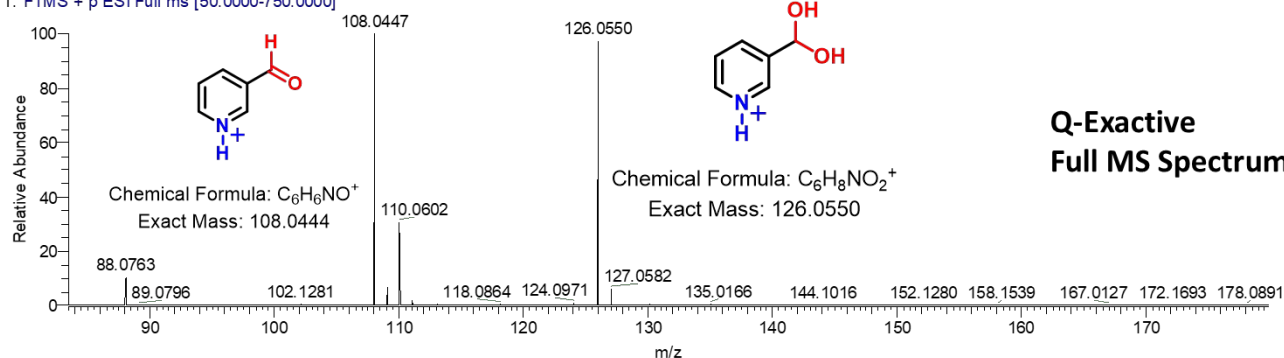
Figure S2. The setup of Microsaic miniature MS 4000 MiD and online dilutor MiDAS.



No hydrolysis product was observed when using 0.1%FA in ACN as makeup solvent

MID-4000
Full MS Spectrum

QE17MS1052 #233-280 RT: 1.27-1.52 AV: 48 NL: 1.47E8
T: FTMS + p ESI Full ms [50.0000-750.0000]



Q-Exactive
Full MS Spectrum

Aldehyde Hydrolysis Under Aqueous HPLC Conditions

Figure S3. The MS spectrum of aldehyde SM with m/z 108 showed significant on-column hydrolysis product m/z 126 when analyzed with reverse phase LCMS; Whereas no aldehyde hydrate was observed in -4000 MiD when using 0.1% formic acid in 100% ACN as the makeup solvent in MiDAS.

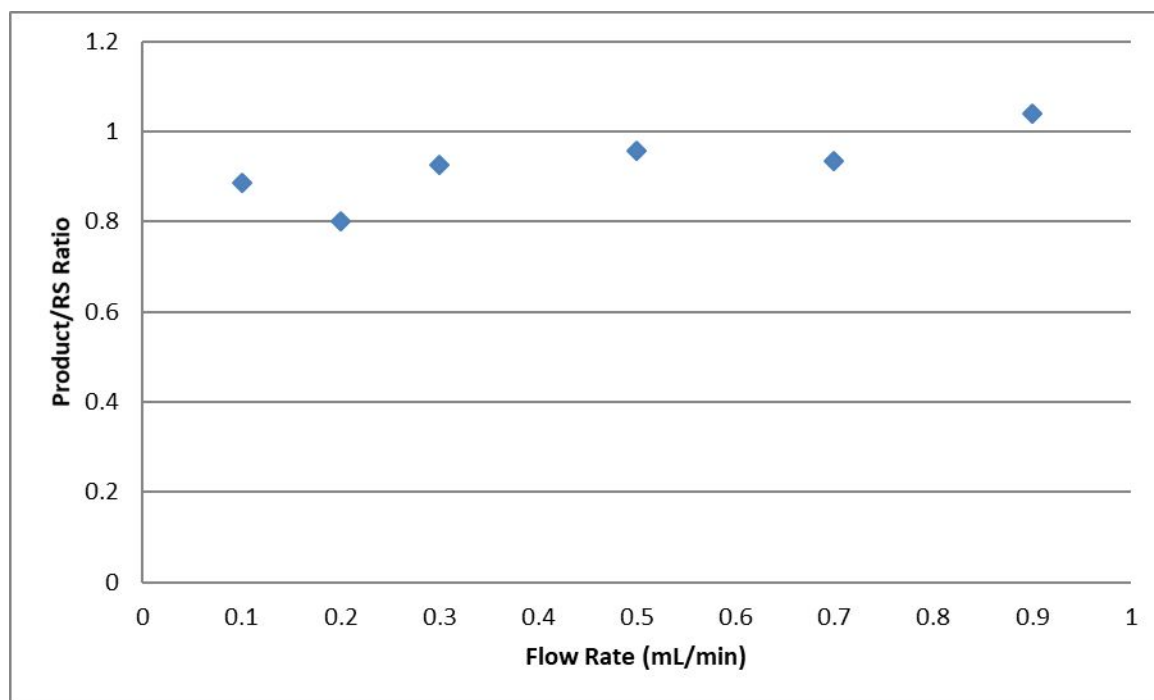


Figure S4. The effect of reaction flow rate on the product/RS ratio was studied in the external RS approach (Figure S4). The solution of product standard (0.2 mol/L in acetonitrile) was introduced directly into the mini-MS module through a syringe pump at varied flow rate from 0.1 to 0.9 mL/min. The attenuation (2000X) and makeup flow rate (1 mL/min) were maintained constant. The results showed that the product/RS ratio changed 25% from 0.8 to 1.0 with the reaction flow rate increased from 0.1 mL/min to 0.9 mL/min. This indicated that the flow rate has some impact on the quantitation result.

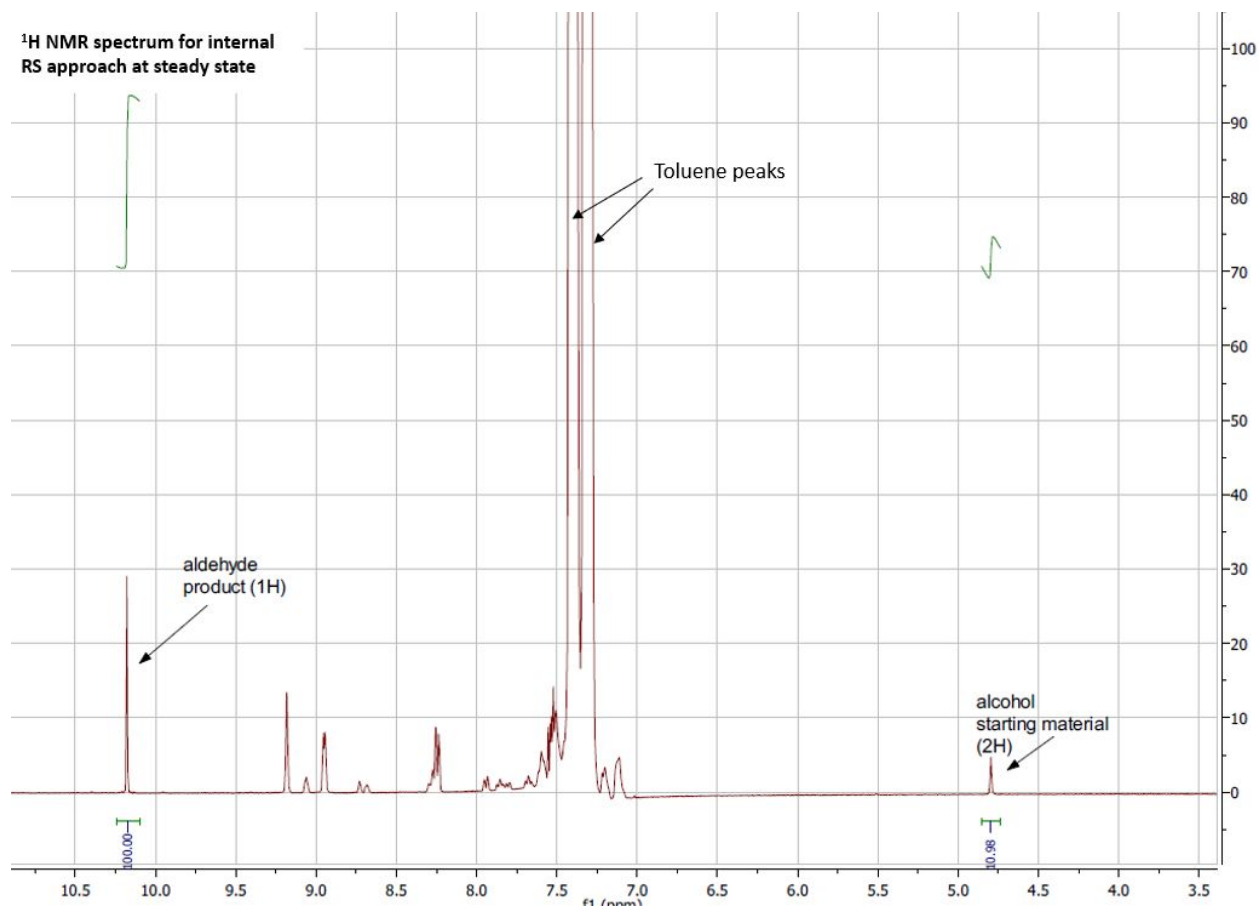


Figure S5. ¹H NMR spectrum of reaction stream sampled at steady state for the internal reference standard approach. Peaks corresponding to quinoline reference standard not visible due to peak overlap and low concentrations.

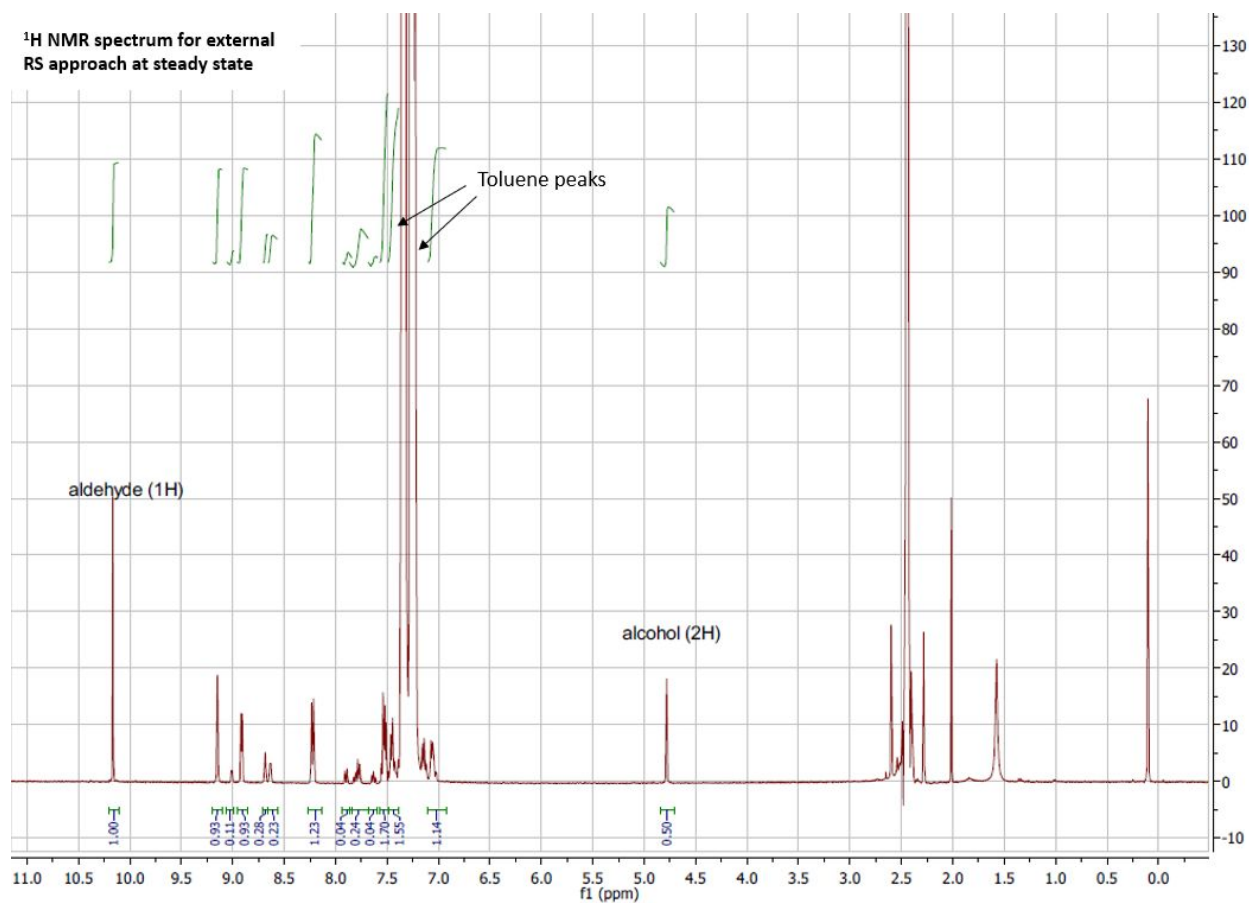


Figure S6. ¹H NMR spectrum of reaction stream sampled at steady state for the external reference standard approach.

Supporting Information References:

1. <http://www.microsaic.com/wp-content/uploads/2018/03/App-note-Online-flow-RxM-of-Wittig-reaction-using-a-deployable-MS.pdf>
2. <https://www.abreg.com/wp-content/uploads/2018/05/Microsaic-4000-MiD-and-MiDas-Batch-monitoring-of-deprotection-chemistry.pdf>
3. https://ichrom.com/wp-content/uploads/2019/09/19_App-note-Bromination-of-pypy-by-real-time-reaction-monitoring.pdf
4. https://ichrom.com/wp-content/uploads/2019/09/17_App-note-Multivariate-analysis-of-a-SNAr-by-on-line-reaction-monitoring-using-miniaturized-MS.pdf
5. https://ichrom.com/wp-content/uploads/2019/09/23_On-line-batch-reaction-monitoring-1.pdf
6. Bristow, T. W. T.; Ray, A. D.; O’Kearney-McMullan, A.; Lim, L.; McCullough, B.; Zammataro, A. On-line Monitoring of Continuous Flow Chemical Synthesis Using a Portable, Small Footprint Mass Spectrometer. *J. Am. Soc. Mass Spectrom.* **2014**, 25, 1794–1802.
7. Browne, D. L.; Wright, S.; Deadman, B. J.; Dunnage, S.; Baxendale, I. R.; Turner, R. M.; Ley, S. V. Continuous Flow Reaction Monitoring Using an On-line Miniature Mass Spectrometer, *Rapid Commun. Mass Spectrom.* **2012**, 26, 1999–2010.

8. Fitzpatrick, D. E.; Battilocchio, C.; Ley, S. V. A Novel Internet-based Reaction Monitoring, Control and Autonomous Self-optimization Platform for Chemical Synthesis. *Org. Process Res. Dev.* **2016**, *20*, 386–394.
9. Blanz A, Bristow TWT, Coombes SR, Corry T, Nunn M, Ray AD. Coupling and Optimisation of Online Nuclear Magnetic Resonance Spectroscopy and Mass Spectrometry for Process Monitoring to Cover the Broad Range of Process Concentration. *Magn Reson Chem.* **2016**, *55*, 274–282.
10. Holmes, N.; Akien, G. R.; Savage, R. J. D.; Stanetty, C.; Baxendale, I. R.; Blacker, A. J.; Taylor, B. A.; Woodward, R. L.; Meadows, R. E.; Bourne, R. A. Online Quantitative Mass Spectrometry for the Rapid Adaptive Optimisation of Automated Flow Reactors. *React. Chem. Eng.* **2016**, *1*, 96–100.