

Supplementary Material

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Table S1. Coordinates of the sampling stations from South Bay, Livingston Island.

Station	Coordinates	
	S	W
St-1	62°38.792'	60°38.411'
St-2	62°38.346'	60°23.912'
St-3	62°39.425'	60°23.277'
St-4	62° 39.556'	60° 22.132'
St-5	62°37.718'	060°21.40'

Table S2. Ancillary data for the seawater sampling sites.

Date	Sampling station	Fluorescence (RFU)	Salinity (PSU)	Temperature (°C)	Turbidity (FTU)
08/01/2018	St-4	n.d	n.d	n.d	n.d
16/01/2018	St-4	n.d	n.d	n.d	n.d
18/01/2018	St-3	0.27	33.49	1.56	10.11
19/01/2018	St-4	0.27	33.49	1.56	10.11
20/01/2018	St-2	0.66	34.14	1.60	3.56
22/01/2018	St-4	0.40	33.30	1.42	7.84
24/01/2018	St-2	0.88	33.94	2.23	3.66
25/01/2018	St-4	0.27	33.46	1.71	5.85
27/01/2018	St-2	0.31	33.68	2.43	4.86
29/01/2018	St-4	0.58	33.49	1.87	6.04
03/02/2018	St-2	0.15	34.75	1.59	3.55
05/02/2018	St-4	0.44	33.61	1.47	6.41
08/02/2018	St-1	n.d	n.d	n.d	n.d
12/02/2018	St-2	0.18	33.67	1.76	7.11
16/02/2018	St-4	0.23	33.26	1.78	7.44
20/02/2018	St-2	0.23	33.86	2.93	3.87
23/02/2018	St-4	0.19	33.16	1.82	9.83
28/02/2018	St-5	0.28	33.63	1.14	5.44
01/03/2018	St-1	n.d	n.d	n.d	n.d
<i>Averaged by station</i>					
	St-1	n.d	n.d	n.d	n.d
	St-2	0.4±0.3	34.01±0.41	2.09±0.54	4.43±1.4
	St-3	0.27	33.49	1.56	10.11
	St-4	0.32±0.14	33.38±0.15	1.67±0.17	7.62±1.62
	St-5	0.28	33.63	1.14	5.44

Table S3. Meteorological conditions for each sampling event. The values were averaged coinciding the sampling hours (From 9 am to 11.30 am).

Date	Wind speed (m/s)	Wind direction (°)	Temperature (°C)	Minimu m temp. (°C)	Maximu m temp. (°C)	Relative humidity (%)	Precipitation (mm)	Atmospheric pressure (hPa)	Radiatio n (Wm ⁻²)
08/01/2018	1.2	250.4	1.9	1.8	2.0	88.6	0.0	1000.9	137.5
11/01/2018	2.0	203.4	2.4	2.2	2.6	95.9	0.0	980.4	86.1
16/01/2018	3.7	56.3	2.7	2.6	2.8	89.9	0.0	978.3	107.4
17/01/2018	0.7	197.3	2.3	2.2	2.5	93.2	0.0	978.8	67.4
18/01/2018	1.3	160.9	1.3	1.1	1.4	88.5	0.0	989.3	98.0
19/01/2018	1.1	156.8	3.0	2.9	3.2	90.9	0.0	978.2	6.7
20/01/2018	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
22/01/2018	3.1	0.0	1.9	1.8	2.0	92.8	0.0	973.8	29.2
24/01/2018	3.6	18.6	1.8	1.8	1.9	93.9	0.0	985.8	50.7
25/01/2018	1.4	102.5	2.7	2.6	2.8	92.5	0.0	984.5	33.8
27/01/2018	1.5	65.7	2.7	2.6	2.8	75.9	0.0	990.6	153.8
29/01/2018	4.6	53.9	5.6	5.4	5.7	74.3	0.0	979.1	136.6
03/02/2018	1.6	50.8	3.0	3.1	3.0	92.9	0.0	977.2	18.0
05/02/2018	1.0	188.3	2.4	2.3	2.6	88.4	0.0	987	126.8
06/02/2018	6.5	0.0	3.0	2.9	3.5	95.4	0.1	980.9	14.5
08/02/2018	3.9	32.7	2.5	2.4	4.2	93.0	0.0	973.4	23.9
12/02/2018	0.4	131.8	3.4	1.4	3.2	91.1	0.0	974.3	39.1
16/02/2018	4.0	33.5	4.1	3.4	4.0	88.0	0.0	986.4	21.4
20/02/2018	6.7	22	1.4	1.3	1.3	88.4	0.0	997.7	41.8
23/02/2018	1.8	74.4	3.3	3.2	3.2	94.6	0.0	992.0	22.5
28/02/2018	2.8	155.8	0.6	0.6	0.7	84.3	0.0	991.4	38
01/03/2018	1.5	52.1	3.1	3.0	3.3	91.1	0.0	970.1	19.5

Table S4. Inorganic nutrient concentrations and PFOS concentrations for the SML and SSL and for each sampling event. PFOS is taken as surrogate of ADOC and SML occurrence.

Proxy of ADOC and						
			Inorganic nutrients			
Date	Sampling Station	sea layer	N-NH ₄ ⁺	NO ₋₃ + NO ₋₂	-PO ₄ ³⁻	SML stability
			(μmolL ⁻¹)	(μmolL ⁻¹)	(μmol L ⁻¹)	PFOS (pg L ⁻¹)
08/01/2018	St-4	sml	n.d	n.d	n.d	n.d
		ssl	7.65±7.42	9.56±1.08	1.06±0.1	n.d
16/01/2018	St-4	sml	2.43	22.21	1.71	85
		ssl	10.8±9.23	19.83±2.87	2.12±0.1	32.5
18/01/2018	St-3	sml	n.d	n.d	n.d	55
		ssl	5.43	14.6	1.26	17.5
19/01/2018	St-4	sml	6.89	14.52	1.68	40
		ssl	n.d	n.d	n.d	17.5
20/01/2018	St-2	sml	1.9	27.04	1.78	15
		ssl	1.97±0.27	26.15±1.39	1.69±0.09	16.25
22/01/2018	St-4	sml	5.24	21.36	1.78	15
		ssl	20.91±20.34	19.37±2.19	1.535±0.19	8.75
24/01/2018	St-2	sml	2.98	26.39	1.52	25
		ssl	2.65±1.14	26.83±0.13	1.87±0.13	8.75
25/01/2018	St-4	sml	n.d	n.d	n.d	60
		ssl	2.84±0.0	18.31±3.41	1.42±0.14	8.75
27/01/2018	St-2	sml	n.d	n.d	n.d	230
		ssl	2.52±1.81	23.27±7.12	1.4±0.06	8.75
29/01/2018	St-4	sml	3.04	24.3	1.87	35
		ssl	3.05±0.49	20.47±3.73	1.59±0.16	10
03/02/2018	St-2	sml	13.39	19.7	1.39	30
		ssl	10.44	16.4	1.68	17.5
05/02/2018	St-4	sml	11.29	18.6	1.89	n.d
		ssl	5.08±1.69	15.32±4.31	1.46±0.11	n.d
08/02/2018	St-1	sml	1.74	27.37	1.93	15
		ssl	3.07±1.48	23.36±3.43	1.62±0.18	12.5
12/02/2018	St-2	sml	1.31	23.08	1.55	20
		ssl	13.05±0.62	20.7±1.5	1.35±0.13	10
16/02/2018	St-4	sml	2.73	19.23	1.52	35
		ssl	9.76±0.58	17.75±1.35	1.32	33.75
20/02/2018	St-2	sml	n.d	n.d	n.d	25
		ssl	6.975±6.47	20.78±2.94	1.38±0.144	7.5
23/02/2018	St-4	sml	7.6±1.84	16.92±0.76	1.40±0.20	n.d
		ssl	8.51±5.34	14.53±3.26	1.26±0.11	n.d
28/02/2018	St-5	sml	n.d	n.d	n.d	n.d
		ssl	5.32±2.34	24.25±0.63	1.39±0.01	n.d
01/03/2018	St-1	sml	n.d	n.d	n.d	70
		ssl	1.12±0.02	10.6±0.75	1.16±0.1	22.5
Averaged by station						
St-1		sml	1.72±0.02	25.4±2.28	1.75±0.2	42.5±38.8
		ssl	2.09±1.43	16.98±7.2	1.39±0.28	17.5± 7.1
St-2		sml	4.08±4.96	24.2±2.78	1.65±0.24	64±93
		ssl	5.89±5.07	22.9±4.45	1.55±0.23	12.3±4.2
St-3		sml	2.83	26.79	1.8	55
		ssl	5.43	14.61	1.26	17.5
St-4		sml	5.61±2.99	18.95±3.07	1.64±0.2	43.8±21.9
		ssl	8.57±9.33	16.9±4.32	1.47±0.31	18.5± 11.1
St-5		ssl	5.32±2.34	24.25±0.63	1.39±0.01	n.d
TOTAL AVERAGED		sml	4.3 ± 3.7	22 ± 4.1	1.7 ± 0.2*	50.3±50.8
		ssl	6.3 ± 7	19.1 ± 5.4	1.4 ± 0.3*	16.5±8.7

Table S5. Subset of taxa, previously reported as bacterial ADOC degraders (AD) (Takahashi *et al.*, 2013; Lozada *et al.*, 2014), detected in our dataset from the maritime Antarctica. As these taxa have also been reported to exudate biosurfactants, the type of surfactants and citation is also given. In bold, marine biosurfactant-producing bacterial strains.

Label	Phylum/Class	Genus	Biosurfactant-producing bacteria	
Actinobacteria AD	Phylum: Actinobacteria	<i>Mycobacterium</i>	Polymeric surfactants	Vijayakumar and Saravanan, 2015
		<i>Nocardioide</i>		
		<i>Rhodococcus</i>	Exopolysaccharides/complex surface active polymer producer	Peng <i>et al.</i> , 2007
		<i>Arthrobacter</i>	Glycolipid surface active molecules	Schulz <i>et al.</i> , 1991
		<i>Gordonia</i>	Extracellular bioemulsifiers, and cell-bound glycolipid biosurfactants (isolated from hydrocarbon polluted site)	Franzetti <i>et al.</i> , 2009
		<i>Microbacterium</i>	Biosurfactant (long chain fatty acid lipid type) (isolated from Amazonas river)	Silva Lima <i>et al.</i> , 2017
		<i>Dietzia</i>	Rhamnolipid	Wang <i>et al.</i> , 2014
		<i>Micrococcus</i>	Biosurfactant (isolated from waste water sludge)	Yilmaz <i>et al.</i> , 2009
		<i>Sulfitobacter</i>		
		<i>Sphingopyxis</i>	Biosurfactant (Contaminated soil)	Saisa-ard <i>et al.</i> , 2013
Alphaproteobacteria AD	Phylum: Proteobacteria Class: Alphaproteobacteria	<i>Sphingomonas</i>	Polymeric surfactant	Santos <i>et al.</i> , 2016
		<i>Roseovarius</i>	Biosurfactant	Antoniou <i>et al.</i> , 2015
		<i>Roseobacter</i>		
		<i>Sphingobium</i>	Biourfactant (Contaminated soil)	Saisa-ard <i>et al.</i> , 2013
		<i>Tropicibacter</i>		
		<i>Novosphingobium</i>		
		<i>Jannaschia</i>		
		<i>Ochrobactrum</i>	Biosurfactant	Zarinviarsagh

				<i>et al.</i> , 2017
				Domingues <i>et al.</i> , 2020
		<i>Thalassospira</i>	Biosurfactant	Antoniou <i>et al.</i> , 2015
		<i>Kordiimonas</i>		
Bacilli AD	Phylum: Firmicutes Class: Bacilli	<i>Bacillus</i>	Exopolysaccharide, complex surface active polymer/polymeric surfactants	Das <i>et al.</i> , 2008
Betaproteobacteria AD	Phylum: Proteobacteria Class: Betaproteobacteria	<i>Acidovorax</i>		
		<i>Comamonas</i>	Biosurfactant (From agricultural soil)	Sun <i>et al.</i> , 2013
		<i>Polaromonas</i>		
		<i>Ralstonia</i>	Biosurfactant (from Petroleum waste)	Plaza <i>et al.</i> , 2007
		<i>Delftia</i>	glycolipids Surfactant (from Oilfield water injection)	Lenchi <i>et al.</i> , 2020
		<i>Alcaligenes</i>	Biosurfactant (from Petroleum waste)	Plaza <i>et al.</i> , 2007
Cytophagia AD	Phylum: Bacteroidetes Class: Cytophagia	<i>Cytophaga</i>		
Deltaproteobacteria AD	Phylum: Proteobacteria Class: Deltaproteobacteria	<i>Desulfatiferula</i>		
		<i>Desulfococcus</i>		
		<i>Desulfatibacillum</i>		
		<i>Desulfobacterium</i>		
Flavobacteriia AD	Phylum: Bacteroidetes Class: Flavobacteriia	<i>Flavobacterium</i>	Biosurfactant (From contaminated arid soils)	Bodour <i>et al.</i> , 2003
Gammaproteobacteria AD	Phylum: Proteobacteria Class: Gammaproteobacteria	<i>Pseudoalteromonas</i>	Exopolysaccharides/complex Surface active polymer Polymeric biosurfactant/bioemulsifier, lipopeptides, glycolipids and particulate surfactant	Roca <i>et al.</i> , 2016
		<i>Pseudomonas</i>		Wittgens <i>et al.</i> , 2017
		<i>Colwellia</i>	Biosurfactant	Mapelli <i>et al.</i> , 2017
		<i>Cycloclasticus</i>	Biosurfactant	Mapelli <i>et al.</i> , 2017
		<i>Alkanindiges</i>		
		<i>Acinetobacter</i>	Polymeric biosurfactant/bioemulsifier	Rosenberg <i>et al.</i> , 1988
		<i>Oleispira</i>		
		<i>Shewanella</i>	Biosurfactant	Antoniou <i>et al.</i> , 2015
		<i>Halomonas</i>	Polymeric biosurfactant/bioemulsifier,	Gutiérrez <i>et al.</i> , 2007

	glycolipid surface active molecules and Exopolysacharides (complex Surface active polymer)	
<i>Vibrio</i>	Exopolysacharides (complex Surface active polymer)	Satpute <i>et al.</i> , 2010
<i>Oleibacter</i>		
<i>Alteromonas</i>	Biosurfactant	Satpute <i>et al.</i> , 2010
<i>Microbulbifer</i>		
<i>Neptunomonas</i>		
<i>Thalassolituus</i>		
<i>Marinobacter</i>	Polymeric biosurfactant/bioemulsifier	Domingues et al 2020
<i>Alcanivorax</i>	Glycolipid surface active molecules (rhamnolipid and sophorolipid)	Abraham <i>et al.</i> , 1998
All AD	*Includes all listed above	

Figure S1. Nutrient and PFOS concentrations in the SML and SSL during the austral summer. Gray background indicates those days with SML disruption. Average wind speed and solar radiation measurements are also shown.

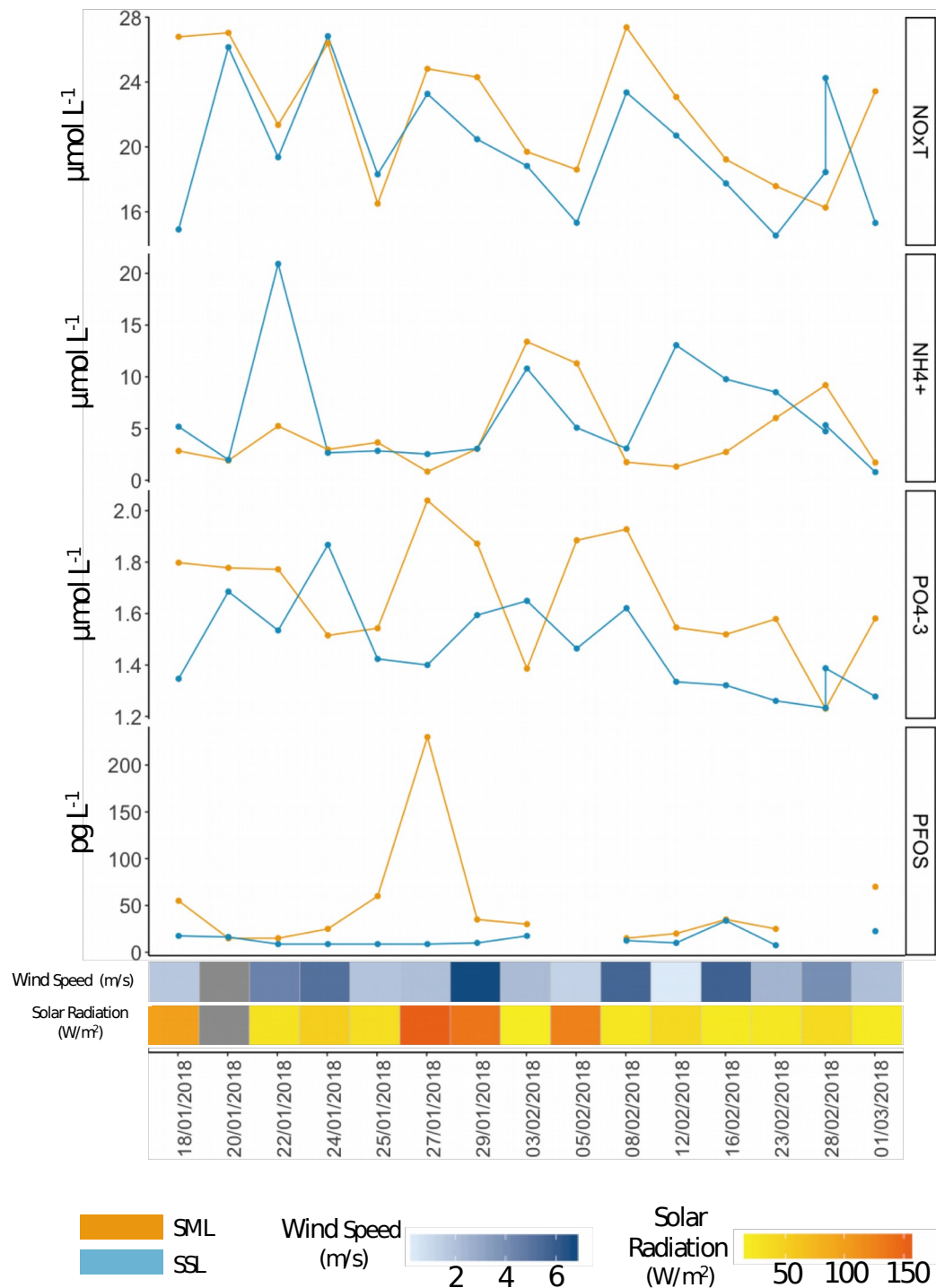


Figure S2. Relative abundances of the dominant taxonomical groups at the SML and in SSL for each sampling station.

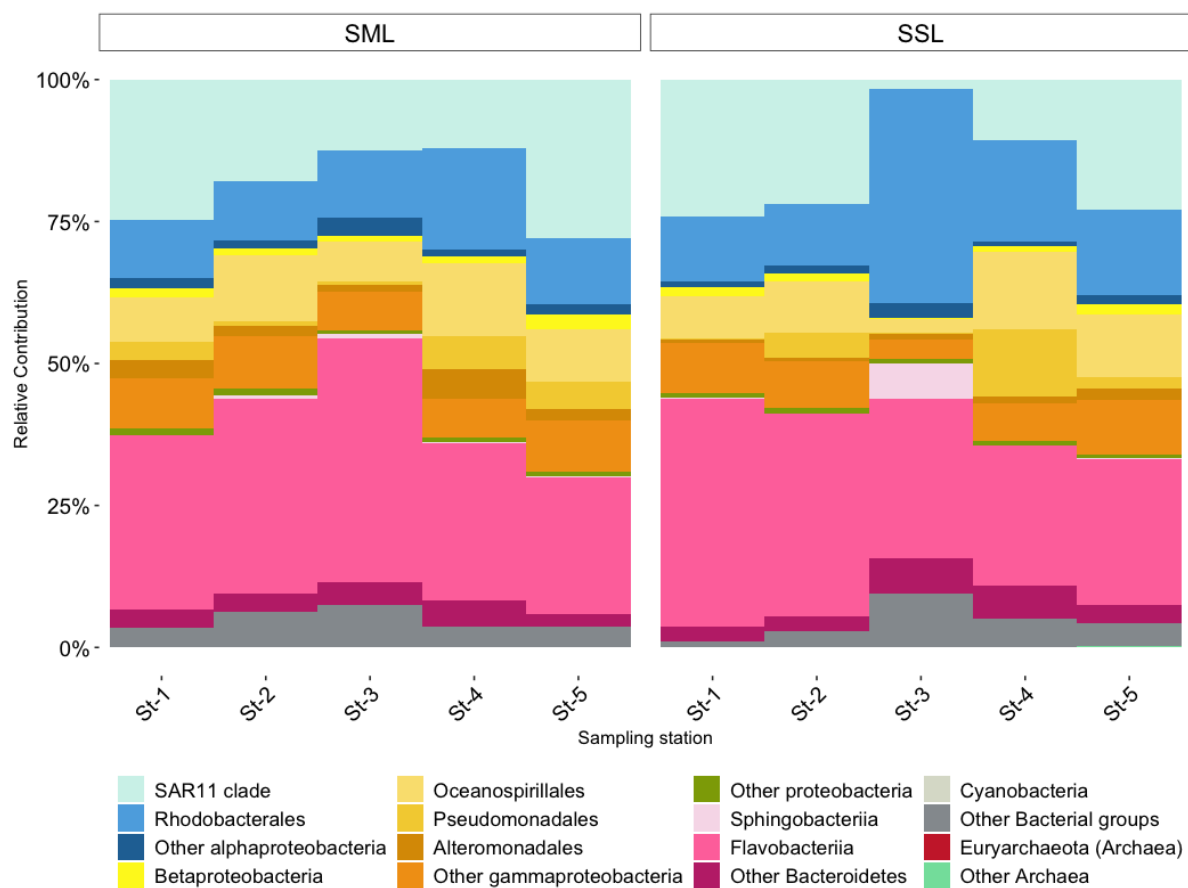


Figure S3. Relative abundance of the dominant taxonomical groups in the SML and SSL during the austral summer. Average wind speed and solar radiation measurements are also shown.

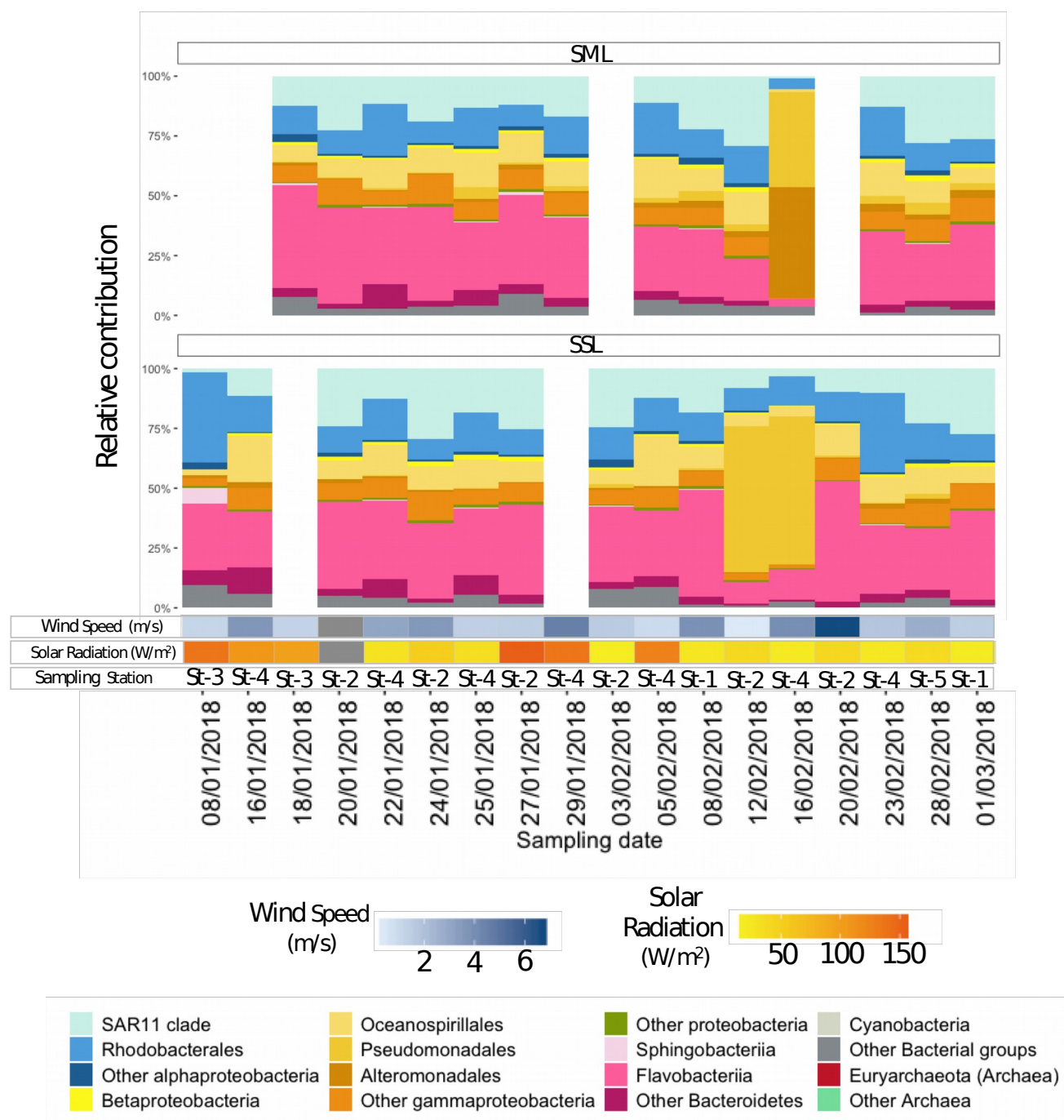
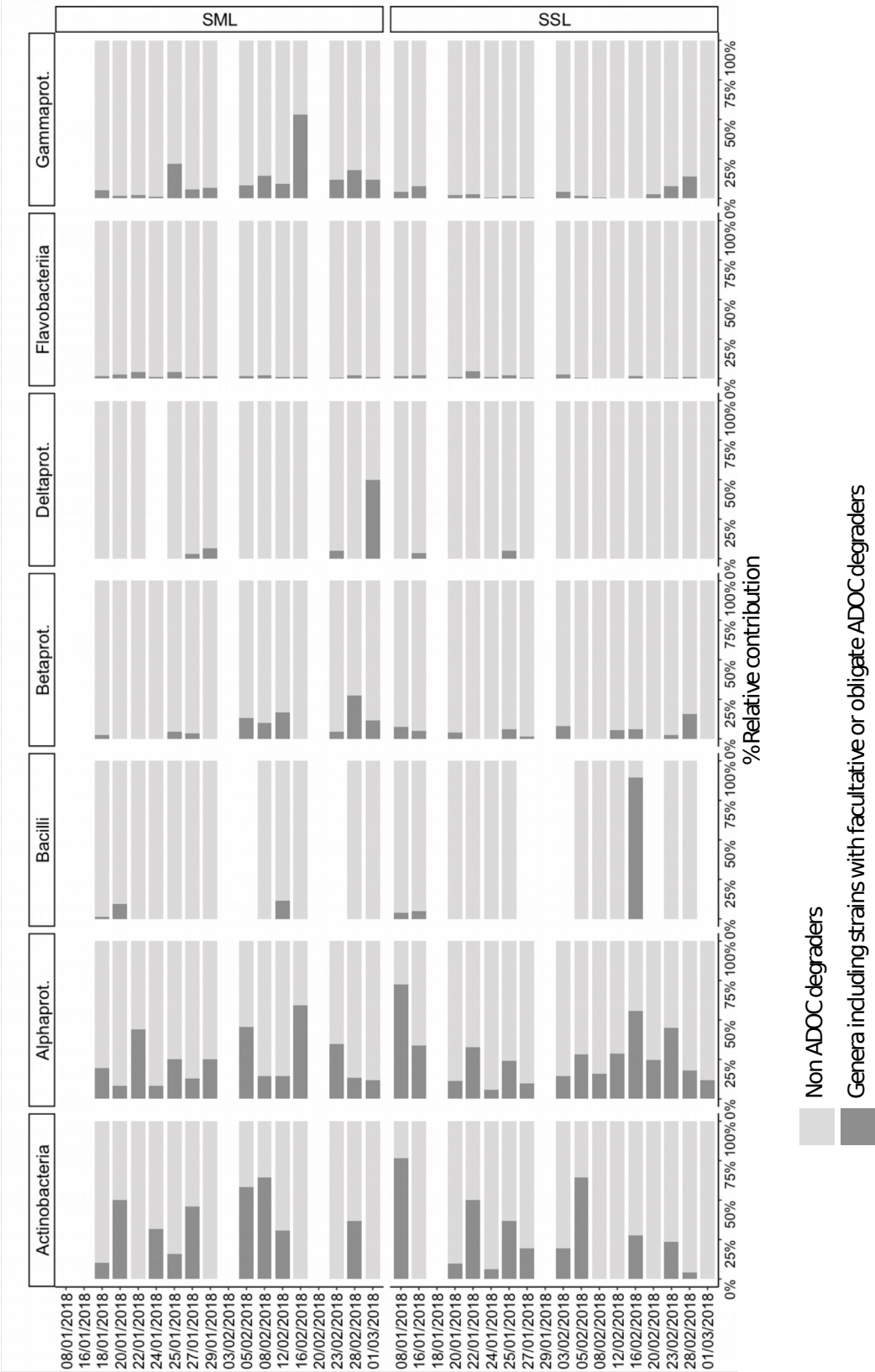


Figure S4. ADOC degraders relative abundance for each layer during the austral summer and corresponding taxonomical class.



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