Supplementary Material



Figure S1. Primer design for the PSR assay. (A), Nucleotide sequence of the *rscA* gene (part) and locations of the primers are underlined. (B), Primer sequences targeting *rscA*.



Figure S2. Detection and confirmation of PSR products. (**A**), The biosensor applied for visual detection of PSR products. (**B**), Colorimetric indicator (VDR) applied for visual determination of PSR amplification products. Biosensor/Tube 1, positive amplification of PSR method (*K. pneumoniae*, ATCC BAA-2146); Biosensor/Tube 2, negative amplification of PSR method (*L. monocytogenes*); Biosensor/Tube 3, negative amplification of PSR assay (*S. flexneri*); Biosensor/Tube 4, blank control (DW).



Figure S3. Optimal reaction temperature for PSR primer set. PSR reactions were monitored by real-time measurement of turbidity (LA-320c), and the threshold value was 0.1. The turbidity of >0.1 was regarded as positive amplification. Eight kinetic graphs were observed at different temperatures (60° C- 67° C, 1° C intervals) with target templates at the level of 1 pg per tube. Signal 1, positive amplification of PSR assay (*K. pneumoniae*, ATCC BAA-2146); Signal 2, negative amplification of PSR assay (*L. monocytogenes*); Signal 3, negative amplification of PSR assay (*S. flexneri*); Signal 4, blank control (DW).



Figure S4. Sensitivity of ATSU-PSR for detecting *K. pneumoniae* in sputum samples.

Biosensor (A)/Tubes (B) 1-8 represented the cell levels of 11000 CFU per reaction (~ 550000 CFU/mL), 1100 CFU per reaction (~ 55000 CFU/mL), 110 CFU per reaction (~ 5500 CFU/mL), 111 CFU per reaction (~ 550 CFU/mL), 1.1 CFU per reaction (~ 55 CFU/mL), 0.11 CFU per reaction (~ 5.5 CFU/mL), 0.011 CFU per reaction (~ 0.55 CFU/mL), negative control (non-contaminated sputum samples). The cell levels of 11000 CFU, 1100 CFU, 110 CFU and 11 CFU per reaction produced the positive amplifications.