## Supporting Information

## Sheathless Acoustic Fluorescence Activated Cell Sorting (aFACS) with High Cell Viability

Peixian Li <sup>1</sup>, Minhui Liang <sup>1</sup>, Xiaoguang Lu <sup>1</sup>, Joycelyn Jia Ming Chow <sup>1</sup>, Chrishan J.A.

Ramachandra <sup>2,3</sup> and Ye Ai \*,1

<sup>1</sup> Pillar of Engineering Product Development, Singapore University of Technology and Design, Singapore 487372, Singapore

<sup>2</sup> National Heart Research Institute Singapore, National Heart Centre Singapore, Singapore 169609, Singapore

<sup>3</sup> Cardiovascular and Metabolic Disorders Program, Duke-National University of Singapore Medical School, Singapore 169857, Singapore

\* Corresponding author. Email: aiye@sutd.edu.sg; Tel: (+65) 6499 4553

## **Supplementary Figures**

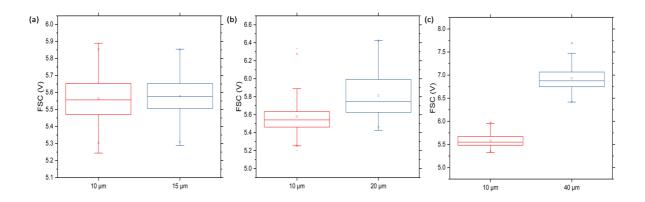


Figure S1 Forward scattering (FSC) measurement of particles with different sizes. (a) FSC signals between 10 and 15  $\mu$ m polystyrene particles without significant difference. (b) FSC signals between 10 and 20  $\mu$ m polystyrene particles with statistically significant difference. (c) FSC signals between 10 and 40  $\mu$ m polystyrene particles with statistically significant difference.

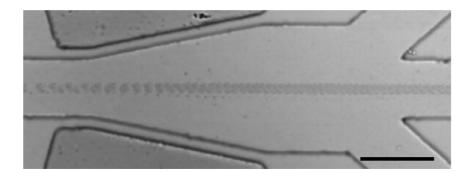


Figure S2 Superimposed image with 40 frames over 30 ms to show elasto-inertial focusing for cell alignment at a flow rate of 20  $\mu$ L/min. The scale bar is 100  $\mu$ m.

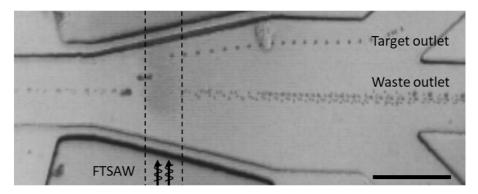


Figure S3 Superimposed image with 40 frames over 30 ms to demonstrate 3  $\mu m$  fluorescent polystyrene particle sorting. The scale bar is 100  $\mu m$ .

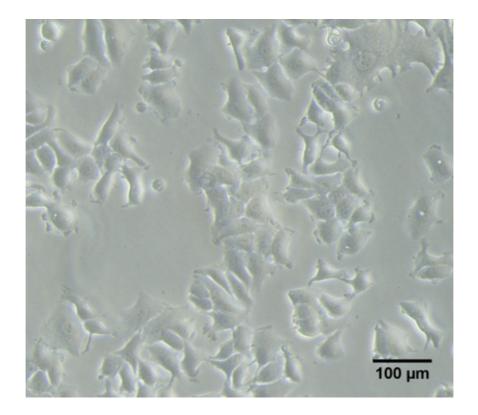


Figure S4 A representative microscopic image of sorted MCF-7 cells after culture for 24 hours.