## Supplemental Material to:

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# Efficient engineering of a bacteriophage genome using the type I-E CRISPR-Cas system

## 2014; 11(1) http://dx.doi.org/10.4161/rna.27766

www.landesbioscience.com/journals/rnabiology/article/27766/

#### SUPPLEMENTARY INFORMATION

#### **Plasmid Construction**

Plasmids pAnti-1.7 (Table S1) and pAnti-4.3 (Table S1) encode spacers against genes 1.7 and 4.3 of bacteriophage T7, respectively, in addition to other non-relevant spacers (spacers EcoRI, K12 sp3, BamHI, K12 sp5, NsiI, and K12 sp7 on plasmid pWUR477<sup>1</sup>). The plasmids were constructed by PCR amplification of pWUR477 (Table S1) using primers DS24F + DS24R for pAnti-1.7, and primers RK77F + RK77R for pAnti-4.3. The amplified DNAs were phosphorylated and self-ligated to yield pAnti-1.7 and pAnti-4.3. Plasmids pBAD-1.7<sub>60</sub> and pUC-4.3<sub>60</sub> (Table S1) encode 60 bp of the sequences flanking genes 1.7 and 4.3, respectively. These two plasmids were constructed by PCR amplification of pBAD283 using primers DS20F + RK76R for pBAD-1.7<sub>60</sub> and pUC19 using primers RK82F + RK82R for pUC-4.3<sub>60</sub>. The amplified DNAs were phosphorylated and self-ligated to yield pBAD-1.7<sub>60</sub> and pUC-4.3<sub>60</sub>.

#### **Preparation of Recombinant Lysate**

*Escherichia coli* NEB5 $\alpha$  harboring pBAD-1.7<sub>60</sub>, pBAD18 (control), pUC-4.3<sub>60</sub>, or pUC19 (control) plasmids were aerated overnight in LB supplemented with 100 µg/ml ampicillin at 37 °C. Each overnight culture was diluted in fresh LB supplemented with 100 µg/ml ampicillin at 37 °C and aerated until the OD<sub>600</sub> reached 1. The culture was then infected with the WT-T7 phage at a multiplicity of infection of 0.1. The infected bacteria were aerated at 37 °C until the culture became clear, which was indicative of a successful phage infection.

#### **Selection of Recombinant Phage**

Phages were plated on BL21AI *E. coli*, with all bacteria harboring plasmids encoding *cas3* and *cascade* as well as different spacers. *E. coli* strains used were DS7045, a strain targeting a protospacer encoded on gene 1.7 (Table S1); RK6617, a strain targeting a protospacer encoded on gene 4.3 (Table S1); and RK6161, a strain encoding control spacers (Table S1). Bacteria were aerated overnight in LB supplemented with 50 µg/ml

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kanamycin, 35 μg/ml chloramphenicol, and 50 μg/ml streptomycin. Each overnight culture was diluted in fresh LB, supplemented with 25  $\mu$ g/ml kanamycin, 17.5  $\mu$ g/ml chloramphenicol, and 25  $\mu$ g/ml streptomycin at 37 °C and aerated until the OD<sub>600</sub> reached 1. Thereafter 0.2% L-arabinose and 0.1 mM isopropyl β-D-1thiogalactopyranoside (IPTG) were added, and the cells were aerated at 37 °C for an additional hour. A 100  $\mu$ l suspension of induced bacteria, 10  $\mu$ l of undiluted phage lysate, and  $10^{-1}$  and  $10^{-2}$  dilutions of the phage lysate in LB medium supplemented with 0.7% agar, 0.2% L-arabinose, and 0.1 mM IPTG were plated onto LB plates, supplemented with 50  $\mu$ g/ml kanamycin, 35  $\mu$ g/ml chloramphenicol, and 50  $\mu$ g/ml streptomycin, and incubated overnight at room temperature. Plaques were then counted, and their size measured. Efficiency of plating was calculated by dividing the number of plaques obtained on *E. coli* encoding targeting spacers by the plaques obtained on E. coli encoding control spacers. Plaques larger than 0.4 cm growing on the targeting bacteria lawns were picked. The plaques served as a DNA template for a PCR using primers 101F + 67R (Table S1) for determining gene 1.7 deletion and primers 114F + RK58R (Table S1) for determining gene 4.3 deletion. Plaques were streaked three times on the induced targeting bacteria for isolation, and deletions were confirmed by DNA sequencing.

Bacteria/plasmids/ phages/ oligonucleotides	Description/sequence	Source or reference
Bacterial strains		
BL21-AI	F <sup>-</sup> ompT hsdSB(rB <sup>-</sup> , mB <sup>-</sup> ) gal dcm araB::T7RNAP-tetA, tet <sup>r</sup>	Invitrogen
NEB5a	$F^-$ , $\phi 80 lac Z\Delta M15\Delta (lac ZYA-arg F)$ , U169, $deoR$ , $recA1$ , $endA1$ ,	New England Biolabs
	hsdR17 ( $r_k^-$ , $m_k^+$ ), gal <sup>-</sup> , phoA, supE44, $\lambda^-$ , thi-1, gyrA96, relA1	
RK6161	BL21AI/pWUR397/pWUR400/pWUR477	1
DS7045	BL21AI/pWUR397/pWUR400/pAnti-1.7	This study
RK6617	BL21AI/pWUR397/pWUR400/pAnti-4.3	This study
Plasmids		
pWUR397	cas3 under T7 promoter, Kan <sup>R</sup>	1
pWUR400	cascade genes under T7 promoter, Str <sup>R</sup>	1
pWUR477	pACYCDuet-1 (Novagen) cloned with control spacers under T7	1
	promoter, cam <sup>r</sup>	
pAnti-1.7	pWUR477 cloned with anti-1.7 spacer	This study
pAnti-4.3	pWUR477 cloned with anti-4.3 spacer	This study
pBAD18	Arabinose-inducible expression vector, amp <sup>r</sup>	2
pBAD283	pBAD18 cloned with T7 gp1.7 ORF	Laboratory collection
pUC19	Cloning vector, ampicillin <sup>r</sup>	3
pBAD-1.7 <sub>60</sub>	pBAD18 cloned with a segment composed of 60 bp flanking gene	This study
	1.7 from each side	
pUC-4.3 <sub>60</sub>	puC19 cloned with a segment composed of 60 bp flanking gene 4.3	This study
	from each side	
Phages		
WT-T7	Wild-type bacteriophage T7	Laboratory collection
Τ7Δ1.7	Bacteriophage T7 deleted in 1.7 gene	This study
Τ7Δ4.3	Bacteriophage T7 deleted in 4.3 gene	This study
Oligonucleotides	5′→3′	
DS20F	CGAAAGCAGACTTCTATAAA	
DS24F	TCCGAGCGATGGACCCGAGTTCCCCGCGCCAGCGGGGATA AACCGCAGCCGAAGCCAAAGAATTC	
DS24R	TTTCACCTAAGGTTATCGGTTTATCCCCGCTGGCGCGGGGA ACTCTCTAAAAGTATACATTTGTT	

### Table S1. Bacterial strains, phages, plasmids and oligonucleotides used in this study

RK58R	TAAGCAGCTGTGCTTGGATAACTTTAGGGTCACGCATGGT AATATCTCCTATAGTGAGTCGAATAGTATTCATGAAGGCC
RK76R	TGGCGGGTTTTCTTTTCCC
RK77F	TGGTTCGTATGTACAAGAGTTCCCCGCGCCAGCGGGGATA AACCGCAGCCGAAGCCAAAGAATTC
RK77R	GCAGTTGGCCTAACTTCGGTTTATCCCCGCTGGCGCGGGG AACTCTCTAAAAGTATACATTTGTT
RK82F	TCAAACAGGAGAAACCATTATGTCTAACGTAGCTGAAACT ATCCGTCTATCCGATACAGCCGCGGGGAGAGGGCGGTTTGC
RK82R	GGTGTGTCTCCTTTAGTGAGTCGTATTAGAATGGGACTCTC CAGCGAAACTTCTCGTAGTCGTTGGCCGATTCATTAATG
67R	GCACCACTCTGAGCAAGATG
101F	TTCCTTTGGCACCGAGCGTC
114F	CGGATGGCTTGAACCATCAA

#### Table S2. Mutations in phages selected on hosts encoding anti gene 1.7 spacers

Mutation <sup>b</sup>	
∆8403-8620	
∆ <b>8211-8697</b>	
∆ <b>8211-8697</b>	
∆8403-8620	
∆8403-8620	
∆ <b>8211-8697</b>	
∆ <b>8211-8697</b>	
∆ <b>8211-8697</b>	

<sup>a</sup> Random large (>4 mm) T7 phage plaques isolated from a single representative experiment

<sup>b</sup> Gene 1.7 coordinates: 8166-8756. Protospacer coordinates: 8423-8455 (NCBI reference sequence NC\_001604.1); Bolded coordinates represent the specifically engineered recombinants

#### References

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