Supplemental online material

Metabolite profiling of Huaiyang Medicago polymorpha with different mowing crops

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1. Experimental

1.1 Chemicals, solvents and herbal materials

Apigenin, Apigenin-7-O-β-D-glucopyranoside, Luteolin, Isoliquiritigenin, Daidzein, Formononetin, Genkwanin, Soyasaponin Bb, β-Sitoterol were purchased from Chengdu must bio-technology co., Ltd. (Chengdu, China) α-Hederin, Tricin, Cynaroside, Chryseriol were purchased from Saipuruisi (beijing) co. (Beijing, China) The purity of the standard was no less than 98%. Oleic acid was purchased from the National Institute for Control of Pharmaceutical and Biological Products. Cultivated Medicago polymorpha (Yang Cao) in six greenhouses in November at Yangzhou (YC-201801, Figure S5), Jiangsu Province, one of the indigenous cultivating regions in China, were mowed twice with every 30 days. After vacuum dried at low temperature then grinded into superfine powder, and approximately 1 kg. HPLC grade methanol was purchased from DIKMA (America) and MS grade formic acid from Fisher Scientific. All other chemicals and solvents were of an analytical grade. Ultra-pure water (18.2MΩ) was prepared with a Milli-Q water purification system (Millipore, Bedford, MA, USA).

1.2 Standard solutions and sample preparation

The mentioned standard were prepared in methanol. Samples of the fruits of *L. lucidum* Ait from different collections were pulverized, and then the powder was sieved through 40 mesh sieves. The pulverized samples were accurately weighed (approximately 0.2 g), and ultrasonically-extracted with 1 ml methanol for 20 min. The resulting solutions were centrifuged at 4000 r/min. The extraction procedure was repeated twice. After cooling, the solutions were combined. Prior to use, all samples were filtered through a 0.22 µm membrane filter.

In the sample preparation procedure, the extraction solvent, extraction procedure, and extraction time were also optimized. Samples of 0.2 g were accurately weighed and extracted using different volumes and percentages of methanol. It was found that the sample extracted by 1 mL methanol showed the greatest

number of detectable components in Medicago polymorpha. And moreover, ultrasonic extraction was simple, reproducible and effective. The weighed powdered samples were extracted with 1 mL methanol for 15 min, 30 min, 60 min, respectively. It was found that the samples with 30, 60 extraction time gave a very similar amount of compounds, and all showed a higher level of compounds than 15 min extraction time. So a 30 min extraction method repeated twice was used throughout this study.

1.3 RRLC-TOFMS conditions

For qualitative analysis of metabolite profiling of Medicago polymorpha, an Agilent 1200 series rapid resolution liquid chromatography system was coupled to an Agilent 6520 quadrupole time-of-flight mass spectrometer An Agilent 6520 quadrupole time-of-flight mass spectrometer (Agilent Technologies, California, USA) which was equipped with an electrospray ionization (ESI) source and operated in positive ion mode and negative mode. The separation was performed on Waters ACQUITY BEH C₁₈ column (100 mm×2.1 mm, 1.7 µm). The mobile phase consisted of (A) water containing 0.1% formic acid and (B) acetonitrile containing 0.1% formic acid. The linear gradient conditions were as follows: 0-5 min, 10-50% B; 5-25 min, 50-100%; 25-30 min, 100%. The flow rate was 0.3 ml/min. The conditions of MS analysis were as follows: the nebulization gas was set to 45 psig, the drying gas was set at 10 L/min, the gas temperature of 350 °C. The capillary voltage was set to 3000 V. All MS data was acquired using reference masses to ensure mass accuracy and reproducibility. The [M+H]⁺ ion of purine at m/z 121.0510 and the $[M+H]^+$ ion of HP-0921 at m/z 922.0098 were used as the lock mass in positive ESI mode. The $[M-H]^-$ ion of purine at m/z 119.0363 and the [M-H]⁻ ion of HP-0921 at m/z 966.0007 were used as the lock mass in negative ESI mode. Data was acquired for each sample from 100 to 1500 Da. The RRLC-QTOFMS data of all determined samples were analyzed by Masshunter qualitative analysis version B. 03. 01 (Agilent, USA) to identify the potential variables for Medicago polymorpha collected. Different kinds of mobile phases, such as acetonitrile and methanol 0.05%, 0.1% and 0.2% aqueous formic acid, were tested. The best peak shape and resolution was obtained from a mixture of methanol and aqueous 0.1% formic acid solution. By an optimized gradient elution in the column, the main components were separately eluted within 30 min. The nebulization gas was set to 45 psi, the drying gas was set at 10

L/min, the gas temperahture of 350 °C. The capillary voltage was set to 3000 V, so as to remove redundant solvent resulting from a flow rate of 0.3 mL/min for mass spectrometer. The MS/MS analysis of components in this study was performed at different collision energies ranging from 25 to 110 V.

1.4 Data processing and statistical analysis

The peak finding, peak alignment, and peak filtering of the ESI+ and ESI- raw data were carried out using Mass Profiler Professional version B.12.05 (Agilent). For data collection, the method parameters were set as follows: retention time range 0-30 min, mass range 100-1500 Da, retention time tolerance 0.01 min, mass tolerance 0.02 Da. For peak integration, the peak intensity threshold was set to 5000. No specific mass or adduct was excluded. The resulting 3-D matrix containing arbitrarily assigned peak index, retention time, and normalized peak area were further exported to a web-based analytical pipeline Metaboanalyst for multivariate statistical analysis (Xia et al. 2015). Mathematical methods, mean-centered and divided by the standard deviation of each variable, were applied to pretreat the data sets resulting from the above samples. Principal Component Analysis (PCA) was to visualize general clustering, trends and outliers among the observations. In order to select potential biomarkers worthy of preferential study in the next step, these differential metabolites were validated using Independent-Samples T-test (SPSS 17.0). *P*-values were generated for all metabolites.

References

- Chen P, Zhan PJ, Yan J. 2012. Research progress of alfalfa flavonoids and their application in poultry production. China Feed (20):33-36. (in Chinese)
- Chen QH. 2012. New methods for analysis of active compounds in tradition Chinese medicine by LC/CE-IT-MS [dissertation]. Wuhan: Wu Han University. (in Chinese)
- Chen, LH, Shan JJ, Xie T, Di LQ. 2014. Influence of Zushima combined with Gancao on dissolution of their eight components by LC-MS/MS. Chinese Trad Pat Med 36:965-969. (in Chinese)

- Chu J, Li C, Dai G, Ju W. 2015. Simultaneous determination of five components in Astragalus Injection and Astragalus Oral Liquid by LC-MS-MS. Chinese Trad Pat Med 12:2647-2651. (in Chinese)
- Fan XS. 2014. Simultaneous detection of the assay of the major constituents and ITs blood medidinal concentration in traditional Chinese medicine by UPLC-MS/MS [master's thesis]. Nanning: Guangxi University. (in Chinese)
- Han H, Li XS, Gao Y, Liu X, Yang ZY, Ma JY. 2012. Determination of B-Vitamins in black wheats by utility of UPLC-MS/MS. Chinese J Biochem Pharm 33:528-571. (in Chinese)
- He C, Li Z, Gao W, Zuo J. 2006. Progress of studies in flavonoids compounds on plants Medicago. Chinese Pharm J 41: 565-568. (in Chinese)
- Jin MC. 2007. Study on the determination of some natural products and some trace environmental organic contaminations by high-performance liquuid chromatography and ion chromatography coupled with mass spectrometry [dissertation]. Hangzhou: Zhe Jiang University. (in Chinese)

Lang X. 2008. Separation, purification of Alfalfa Saponins and their structure elucidation [master's thesis]. Nanjing: Nanjing Normal University. (in Chinese)

- Liu SY, Zhang XP, Shang ZP, Wang F, Zhang XX, Zhang JY, Lu JQ. 2016. Rapid characterization of chemical constituents and rats metabolites of Kudiezi injection by UHPLC-LTQ-Orbitrap. Zhongguo Zhong Yao Za Zhi 41:2235-2244.
- Liu W. 2010. Study on extraction and separation of flavonoids with estrogenic activity from pigeon pea leaves and antitumor activity [dissertation]. Harbin: Northeast Forestry University. (in Chinese)
- Liu Z, Ouyang H, Li ZF, Li Y, Feng YL, He MZ, Yang SL. 2016. Pharmacokinetics and tissue distribution of α-hederin sodium salt in rats. Zhongguo Zhong Yao Za Zhi 41:2543-2548. (in Chinese)
- Meng YH, Huang HS, Yu HP, He MZ, Sun YB, Feng YL, Yang SL. 2013. Identification of main chemical constituents in extracts of Akebiae Fructus by HPLC-ESI-MS. Zhongcaoyao: yue kan 44:1562-1567. (in Chinese)
- Peng BA, Gao YG, Wang CZ, Wang YH, Zhang XX, Fan K. 2010. Effect of alfalfa on the reproduction of sows. Pratac Sci 27:106-111. (in Chinese)
- Shi WT. 2007. Separation and determination of saponin components in soybean germs [master's thesis]. Shanghai: Shanghai Jiao Tong University. (in Chinese)
- Wang SC, Zhang GG, Mi WZ. 2009. Advances in the study on the chemical constituents and pharmacological activities of Medicago sativa L. J Shenyang Pharm Univ 26:243-248. (in Chinese)

- Wang Y, Yuan J, Xiao J, Cai Q, Wang X, An R, Ma Y. 2012. Pharmacokinetics of active ingredients of Radix Glycyrrhizae in rats with oral administration of Banxia Xiexin decoction and its different compatibilities. Chinese J Pharm Anal 32:1331-1338. (in Chinese)
- Xia J, Sinelnikov IV, Han B, Wishart DS. 2015. MetaboAnalyst 3.0 making metabolomics more meaningful. Nucleic Acids Res 43:W251-W257.
- Xiong X, Pan Y, Zhang T, Yang M, Tan QZ, Xu RC, Zhang DK, Han L. 2017. Consistency evaluation for commercially Xiaojin pills using determination of multi-component contents by HPLC-MS/MS method coupled with chemometrics. Zhongcaoyao: yue kan 48: 2189-2196. (in Chinese)
- Xu M, Liu C, Lv W, Mi S, Wang N. 2014. Determination of genkwanin in rat plasma by using LC-MS/MS and pharmacokinetic study of genkwanin. Traditional Chinese Drug Res Clin Pharmacol 25 (1):51-54. (in Chinese)
- Yan XY, Yin XJ, Wang GK, Hong JL, Lin BB, Qin MJ. 2012. Chemical Constituents of Medicago polymorpha. Chinese Pharm J 47:415-418. (in Chinese)

Yang BL. 2004. Study on the chemical components in alfalfa seed. Grassland Turf (03):33-35. (in Chinese)

- Yang GY, Hu P, Li L, Zhang CN. 2016. Content determination of biochanin A in bile from rats model with single-pass perfusion by UPLC-MS/MS. J Hubei Univ Med 35:111-4+122. (in Chinese)
- Yu S, Zhang L, Shan MQ, Qian Y, Ding AW. 2016. Simultaneous determination of eight organic acids in Chaenomelis Fructus by UFLC-MS. Zhongcaoyao: yue kan 47: 2465-2469. (in Chinese)
- Yuan M. 2016. Isolation of galuteolin from Elsholtiza Bodinieri Vaniot and its biological [master's thesis]. Shanghai: Shanghai Ocean University. (in Chinese)

Zhao HF, Zhao Q. 2013. Study on species, distribution and chemical Composition of domestic alfalfa. China Dairy Cattle (14):53-58. (in Chinese)

Zheng L, Yin L, Xu L, Wang Y. 2015. Pharmacokinetic study of apigenin, 3-hydroxy-genkwanin and genkwanin in rats after oral administration of extraction of fephne genkwa sieb.et zucc by LC-MS/MS. Chinese J Modern Appl Pharm 32:54-59. (in Chinese)

Zhu JM, Li N, Zhang YJ, Li XD, Wang CZ. 2009. The research progress of alfalfa flavonoids. Pratac Sci 26:156-162. (in Chinese)

	·	0		[M+H] ⁺ m/z			$[M+Na]^+$ m/z			
ta	Assigned identity	Molecular	Mean	Theoretical	Mass	Mean	Theoretical	Mass	Product	References
۲R	Assigned identity	formula	measured	extract mass	Accuracy	measured	extract mass	Accuracy	ion	References
			mass (Da)	(Da)	(ppm)	mass (Da)	(Da)	(ppm)		
2.75	Kaempferol- 3-robinobioside*	$C_{27}H_{30}O_{15}$	595.1664	595.1657	0.05	-	617.1477	-	578, 449, 287	He et al. (2006); Lang (2008)
2.86	Riboflavin*	$C_{17}H_{20}N_4O_6$	377.1459	377.1456	-0.9	-	399.1275	-	243, 172	Peng et al. (2010); Han et al. (2012)
2.99	Apigenin-7-0- [β-D-GluA-O-β-D-GluA]*	$C_{27}H_{26}O_{17}$	623.1253	623.1243	-1.65	-	645.1062	-	607, 447, 271	He et al. (2006); Lang (2008); Yan et al. (2012); Liu et al. (2016)
3.59	Cynaroside* [#]	$C_{21}H_{20}O_{11}$	449.1068	449.1078	2.4	-	471.0898	-	287	Wang et al. (2009); Zhao HF and Zhao Q (2013); Yuan (2016)
4.03	Apigenin-7-O- β-D-glucopyranoside* [#]	$C_{21}H_{20}O_{10}$	433.1133	433.1129	-0.87	-	455.0949	-	271	Yin and Qin (2008); Yan et al. (2012)
4.69	Daidzein* [#]	$C_{15}H_{10}O_4$	255.0651	255.0652	0.34	-	277.0471	-	181, 137	Lang (2008); Yan et al. (2012); Zhao HF and Zhao Q (2013); Fan (2014)
4.87	Luteolin* [#]	$C_{15}H_{10}O_6$	287.0551	287.0550	-0.3	-	309.0370	-	153	Lang (2008); Yin and Qin (2008); Zhu et al. (2009); Chen et al. (2012); Yan et al. (2012); Zhao HF and Zhao Q (2013); Zheng et al. (2015) Liu et al. (2016); Yuan (2016)
5.43	Apigenin ^{*#}	$C_{15}H_{10}O_5$	271.0599	271.0601	0.74	-	293.0420	-	153	Lang (2008); Yin and Qin (2008); Zhu et al. (2009); Chen et al. (2012); Yan et al. (2012); Zhao HF and Zhao Q (2013); Zheng et al. (2015) Liu et al. (2016); Yuan (2016)

Table S1. Components identified using ESI + from the Medicago Polymorpha Linnin the first mowing crops.

				[M+H] ⁺ m/z			[M+Na] ⁺ m/z			
ta	Assigned	Molecular	Mean	Theoretical	Mass	Mean	Theoretical	Mass	Product ion	References
•	identity	formula	measured	extract mass	Accuracy	measured	extract mass	Accuracy	ribudeetion	hererenees
			mass (Da)	(Da)	(ppm)	mass (Da)	(Da)	(ppm)		
										Lang (2008); Wang et al. (2009);
5.60	Chryseriol ^{*#}	$C_{16}H_{12}O_{6}$	301.0704	301.0707	0.88	-	323.0526	-	155	Zhu et al. (2009); Chen et al.
										(2012); Liu et al. (2016);
										Yin and Qin (2008); Wang et al.
5.96	Isoliquiritigenin [#]	$C_{15}H_{12}O_4$	257.0809	257.0808	-0.25	-	279.0628	-	240	(2012); Yan et al. (2012); Chen
										et al. (2014)
										Lang (2008); Zhu et al. (2009);
6.25	Formononetin ^{*#}	$C_{16}H_{12}O_4$	269 0805	269 0808	1.25	_	291.0628	-	197,252,	Chen et al. (2012); Zhao HF and
0.25	ronnononeun	01011204	20010000	20310000			291.0020		238,221	Zhao Q (2013); Chu et al.
										(2015); Yang et al. (2016)
	Sovasaponin Bb								798, 636,	He et al. (2005); Jin (2007); Shi
6.89	★#	$C_{48}H_{78}O_{18}$	943.5269	943.5261	-0.86	965.512	965.5080	-4.2	599, 520,	(2007); Lang (2008); Tava et al.
									441	(2011)
7.05	Genkwanin★#	C16H12O5	285.0776	285.0757	-6.51	-	307.0577	-	254, 251,	He et al. (2006); Xu et al.
		-10 12 - 5							189	(2014); Zheng et al. (2015)
18.94	Palmitic acid 🖈	$C_{16}H_{32}O_2$	257.2471	257.2475	1.59	279.2299	279.2295	-1.75	240	Yang (2004); Wang et al. (2009)
19.55	Oleic acid★	$C_{18}H_{34}O_2$	283.2631	283.2632	0.2	305.2441	305.2451	3.55	266	Yang (2004)

Table S1. Components identified using ESI + from the Medicago Polymorpha Linnin the first mowing crops (continued).

			[M-H] ⁻ m/z							
t-	Assigned identity	Molecular	Mean	Theoretical	Mass	Product ion	References			
۲R	Assigned identity	formula	measured extract A		Accuracy	FIGULETION	Neterences			
			mass(Da)	mass(Da)	(ppm)					
2.75	Kaempferol- 3-robinobioside*	$C_{27}H_{30}O_{15}$	593.1500	593.1512	2.01	285	He et al. (2006); Lang (2008)			
2.86	Riboflavin*	$C_{17}H_{20}N_4O_6$	375.1299	375.1310	2.95	358	Peng et al. (2010); Han et al. (2012)			
2.99	Apigenin-7-0- [β-D-GluA-O-β-D-GluA]*	$C_{27}H_{26}O_{17}$	621.1070	621.1097	0.04	445,552	He et al. (2006); Lang (2008); Yan et al. (2012); Liu et al. (2016)			
3.59	Cynaroside* [#]	$C_{21}H_{20}O_{11}$	447.0925	447.0933	1.75	285, 431	Wang et al. (2009); Zhao HF and Zhao Q (2013); Yuan (2016)			
4.03	Apigenin-7-O- β-D-glucopyranoside ^{★#}	$C_{21}H_{20}O_{10}$	431.0972	431.0984	2.71	269, 225, 201, 183				
4.69	Daidzein* [#]	$C_{15}H_{10}O_4$	253.0495	253.0506	4.46	209, 185, 167	Lang (2008); Yan et al. (2012); Zhao HF and Zhao Q (2013); Fan (2014)			
4.87	Luteolin* [#]	$C_{15}H_{10}O_6$	285.0405	285.0405	-0.13	241, 199, 217,175,133	Lang (2008); Yin and Qin (2008); Zhu et al. (2009); Liu (2010); Chen et al. (2012); Yan et al. (2012); Zhao HF and Zhao Q (2013); Fan (2014); Liu et al. (2016)			
5.43	Apigenin* [#]	$C_{15}H_{10}O_5$	269.0458	269.0455	-0.94	117,225, 201, 183	Lang (2008); Yin and Qin (2008); Zhu et al. (2009); Chen et al. (2012); Yan et al. (2012); Zhao HF and Zhao Q (2013); Zheng et al. (2015); Liu et al. (2016); Yuan (2016)			
5.60	Chryseriol* [#]	$C_{16}H_{12}O_{6}$	299.0542	299.0561	6.37	268, 225, 201, 183	Lang (2008); Wang et al. (2009); Zhu et al. (2009); Chen et al. (2012); Liu et al. (2016)			
5.96	Isoliquiritigenin [#]	$C_{15}H_{12}O_4$	255.0664	255.0663	-0.46	135,119	Yin and Qin (2008); Wang et al. (2012); Yan et al. (2012); Chen et al. (2014)			
6.25	Formononetin ^{*#}	$C_{16}H_{12}O_4$	267.0659	267.0663	1.43	252	Lang (2008); Zhu et al. (2009); Chen et al. (2012); Zhao HF and Zhao Q (2013); Chu et al. (2015); Yang et al. (2016)			
6.89	Soyasaponin Bb ^{*#}	$C_{48}H_{78}O_{18}$	941.5156	941.5115	-4.31	880,796,734,616,598	He et al. (2005); Jin (2007); Shi (2007); Lang (2008); Tava et al. (2011)			
7.05	Genkwanin ^{*#}	$C_{16}H_{12}O_5$	283.0585	283.0612	9.49	116, 268	He et al. (2006); Xu et al. (2014); Zhang et al. (2015)			
18.94	Palmitic acid *	$C_{16}H_{32}O_2$	255.2327	255.2330	0.99	238	Yang (2004); Wang et al. (2009)			
19.55	Oleic acid *	$C_{18}H_{34}O_2$	281.2476	281.2486	3.56	264	Yang (2004)			

Table S2. Components identified using ESI - from the Medicago Polymorpha Linnin the first mowing crops.

		U	$[M+H]^+ m/z$ $[M+Na]^+ m/z$							
+	Accigned identity	Molecular	Mean	Theoretical	Mass	Mean	Theoretical	Mass	Product	Poforoncoc
۲R	Assigned identity	formula	measured	extract mass	Accuracy	measured	extract mass	Accuracy	ion	References
			mass (Da)	(Da)	(ppm)	mass(Da)	(Da)	(ppm)		
2.75	Kaempferol-3-robinobioside*	$C_{27}H_{30}O_{15}$	595.1661	595.1657	-0.59	-	617.1477	-	578, 449, 287	He et al. (2006); Lang (2008); Liu et al. (2016)
2.85	Riboflavin*	$C_{17}H_{20}N_4O_6$	377.1459	377.1456	-0.9	-	399.1275	-	243, 172	Peng et al. (2010); Han et al. (2012)
2.98	Apigenin-7*-0-[β-D-GluA-O-β-D-GluA]*	$C_{27}H_{26}O_{17}$	623.1242	623.1243	0.12	-	645.1062	-	607, 447, 271	He et al. (2006); Lang (2008); Yan et al. (2012); Liu et al. (2016)
4.01	Apigenin-7-O-β-D-glucopyranoside* [#]	$C_{21}H_{20}O_{10}$	433.1123	433.1129	1.44	-	455.0949	-	271	Yin and Qin (2008); Yan et al. (2012)
4.69	Daidzein* [#]	$C_{15}H_{10}O_4$	255.0653	255.0652	-0.45	-	277.0471	-	181, 137	Lang (2008); Yan et al. (2012); Zhao HF and Zhao Q (2013); Fan (2014)
4.87	Luteolin* [#]	$C_{15}H_{10}O_{6}$	287.0551	287.055	-0.3	-	309.037	-	153	Lang (2008); Yin and Qin (2008); Zhu et al. (2009); Liu (2010); Chen et al. (2012); Yan et al. (2012); Zhao HF and Zhao Q (2013); Fan (2014); Liu et al. (2016);
5.32	Hederacoside I *	$C_{35}H_{86}O_{22}$	1075.5695	1075.5684	-1.07	1097.549	1097.5503	1.21	1058.5	Tava et al. (2011); Meng et al. (2013)
5.44	Apigenin ^{*#}	$C_{15}H_{10}O_5$	271.0615	271.0601	-5.18	-	293.042	-	153	Lang (2008); Yin and Qin (2008); Zhu et al. (2009); Chen et al. (2012); Yan et al. (2012); Zhao HF and Zhao Q (2013); Zheng et al. (2015) Liu et al. (2016); Yuan (2016)

Table S3. Components identified using ESI+ from the Medicago Polymorpha Linn in the second mowing crops.

 Table S3. Components identified using ESI+ from the Medicago Polymorpha Linn in the second mowing crops (continued).

			[M+H] ⁺ m/z			[M+Na] ⁺ n	n/z			
+	Assigned identity	Molecular	Mean	Theoretical	Mass	Mean	Theoretical	Mass	Product	Poforoncoc
LR	Assigned identity	formula	measured	extract mass	Accuracy	measured	extract mass	Accuracy	ion	References
			mass (Da)	(Da)	(ppm)	mass(Da)	(Da)	(ppm)		
5.60	Chryseriol ^{*#}	$C_{16}H_{12}O_6$	301.0706	301.0707	0.22	-	323.0526	-	155	Lang (2008); Wang, et al. (2009); Zhu et al. (2009); Chen et al. (2012); Liu et al. (2016)
5.97	Isoliquiritigenin [#]	$C_{15}H_{12}O_4$	257.081	257.0808	-0.64	-	279.0628	-	240	Yin and Qin (2008); Wang et al. (2012); Yan et al. (2012); Chen et al. (2014)
6.25	Formononetin ^{*#}	C ₁₆ H ₁₂ O ₄	269.081	269.0808	-0.61	-	291.0628	-	197,252, 238,221	Lang (2008); Zhu et al. (2009); Chen et al. (2012); Chu et al. (2015); Yang et al. (2016); Zhao HF and Zhao Q (2013)
6.90	Soyasaponin Bb [*]	$C_{48}H_{78}O_{18}$	943.5261	943.5261	-0.01	965.508	965.508	0.04	798, 636, 599, 520, 441	He et al. (2005); Jin (2007); Shi (2007); Lang (2008); Tave et al. (2011)
7.09	Genkwanin ^{*#}	$C_{16}H_{12}O_5$	285.075	285.0757	2.64	-	307.0577	-	254, 251, 189	He et al. (2006); Xu et al. (2014); Zheng et al. (2015)
7.97	α-Hederin ^{*#}	$C_{41}H_{66}O_{12}$	751.4625	751.4627	0.27	773.4452	773.4446	-0.73	734.4	Tava et al. (2011); Meng et al. (2013); Liu et al. (2016)
15.58	Oleanolic acid ^{*#}	$C_{30}H_{48}O_3$	457.364	457.3676	7.96	479.3443	479.3496	11.54	439, 411, 393	Kinjo et al. (1994); Chen (2012); Yu et al. (2016); Xiong et al. (2017)
19.01	Palmitic acid [*]	$C_{16}H_{32}O_2$	257.2476	257.2475	-0.36	279.2302	279.2295	1.13	240	Yang (2004); Wang et al. (2009)
19.51	Oleic acid [*]	$C_{18}H_{34}O_2$	283.2629	283.2632	0.91	305.2445	305.2451	2.13	266	Yang (2004)

			[M-H] m/z				
ta	Assigned identity	Molecular	Mean	Theoretical	Mass	Product ion	References
٩K	Assigned dentity	formula	measured extract mass		Accuracy	rioduction	hererences
			mass (Da)	(Da)	(ppm)		
2.75	Kaempferol-3-robinobioside*	$C_{27}H_{30}O_{15}$	595.1661	595.1657	-0.59	285	He et al. (2006); Lang (2008); Peng et al. (2010); Han et al. (2012)
2.85	Riboflavin*	$C_{17}H_{20}N_4O_6$	377.1459	377.1456	-0.9	358	Peng et al. (2010); Han et al. (2012)
2.98	Apigenin-7*-0-[β-D-GluA-O-β-D-GluA]*	$C_{27}H_{26}O_{17}$	623.1242	623.1243	0.12	445,552	He et al. (2006); Lang (2008); Yan et al. (2012); Liu et al. (2016)
4.01	Apigenin-7-O-β-D-glucopyranoside* [#]	$C_{21}H_{20}O_{10}$	433.1123	433.1129	1.44	269, 225, 201, 183	Yin and Qin (2008); Yan et al. (2012)
4.69	Daidzein* [#]	$C_{15}H_{10}O_4$	255.0653	255.0652	-0.45	209, 185, 167	Lang (2008); Yan et al. (2012); Zhao HF and Zhao Q (2013); Fan (2014)
4.87	Luteolin ^{*#}	$C_{15}H_{10}O_6$	287.0551	287.055	-0.3	241, 199, 217,175,133	Lang (2008); Yin and Qin (2008); Zhu et al. (2009); Liu (2010); Chen et al. (2012); Yan et al. (2012); Zhao HF and Zhao Q (2013); Fan (2014); Liu et al. (2016)
5.32	Hederacoside I *	$C_{35}H_{86}O_{22}$	1075.5695	1075.5684	-1.07	927,765,603,471	Tava et al. (2011); Meng et al. (2013)
5.44	Apigenin ^{*#}	C ₁₅ H ₁₀ O ₅	271.0615	271.0601	-5.18	117,225, 201, 183	Lang (2008); Yin and Qin (2008); Zhu et al. (2009); Chen et al. (2012); Yan et al. (2012); Zhao HF and Zhao Q (2013); Zheng et al. (2015); Liu et al. (2016); Yuan (2016)
5.60	Chryseriol ^{*#}	$C_{16}H_{12}O_{6}$	301.0706	301.0707	0.22	268, 225, 201, 183	Lang (2008); Wang et al. (2009); Zhu et al. (2009); Chen et al. (2012); Liu et al. (2016)
5.97	Isoliquiritigenin [#]	$C_{15}H_{12}O_4$	257.081	257.0808	-0.64	135,119	Yin and Qin (2008); Wang et al. (2012); Yan et al. (2012); Chen et al. (2014)
6.25	Formononetin ^{*#}	$C_{16}H_{12}O_4$	269.081	269.0808	-0.61	252	Lang (2008); Zhu et al. (2009); Chen et al. (2012); Zhao HF and Zhao Q (2013); Chu et al. (2015); Yang et al. (2016)
6.90	Soyasaponin Bb [*]	$C_{48}H_{78}O_{18}$	943.5261	943.5261	-0.01	880,796,734,616,598	He et al. (2005); Jin (2007); Shi (2007); Lang (2008); Tava et al. (2011)
7.09	Genkwanin ^{*#}	$C_{16}H_{12}O_5$	285.075	285.0757	2.64	116, 268	He et al. (2006); Xu et al. (2014); Zheng et al. (2015)
*represe	ented these metabolites were significant chan	ged in different	mowing crops.	# represented t	hese metabo	lites were identified by r	eference compounds.

Table S4. Components identified using ESI- from the Medicago Polymorpha Linn in the second mowing crops.

 Table S4. Components identified using ESI- from the Medicago Polymorpha Linn in the second mowing crops (continued).

t _R	Assigned identity	Molecular	[M-H] ⁻ m/z			Droduct ion	Poforoncos
	Assigned identity	formula	Mean	Theoretical	Mass	Production	References

			measured	extract mass	Accuracy		
			mass (Da)	(Da)	(ppm)		
7.97	α-Hederin ^{*#}	$C_{41}H_{66}O_{12}$	751.4625	751.4627	0.27	705, 603, 471	Tava et al. (2011); Meng et al. (2013); Liu et al. (2016)
15.58	Oleanolic acid ^{*#}	$C_{30}H_{48}O_3$	457.364	457.3676	7.96	407, 391,363, 295	Kinjo et al. (1994); Yu et al. (2016); Xiong et al. (2017)
19.01	Palmitic acid [*]	$C_{16}H_{32}O_2$	257.2476	257.2475	-0.36	238	Yang (2004); Wang et al. (2009)
19.51	Oleic acid [*]	$C_{18}H_{34}O_2$	283.2629	283.2632	0.91	264	Yang (2004)



Figure S1 The BPI chromatogram of Huaiyang Medicago polymorpha with the first mowing crops (A ESI+), (B ESI-), the second mowing crop (C ESI+), (D ESI-)



Figure S2 Mass spectra of luteolin in the first mowing Medicago polymorpha compared to luteolin standard. A: EIC of luteolin standard in positive ion mode; B: mass spectra from luteolin standard in positive ion mode; C: mass spectra of luteolin in the first mowing Medicago polymorpha in positive ion mode; D: MS/MS of luteolin in the first mowing Medicago polymorpha in positive ion mode. E: EIC of luteolin



Figure S3 Mass spectra of soyasaponin Bb in the first mowing Medicago polymorpha compared to soyasaponin Bb standard. A: EIC of

soyasaponin Bb standard in positive ion mode; B: mass spectra from soyasaponin Bb standard in positive ion mode; C: mass spectra of soyasaponin Bb in the first mowing Medicago polymorpha in positive ion mode; D: MS/MS spectra of soyasaponin Bb in the first mowing Medicago polymorpha in positive ion mode. E: EIC of soyasaponin Bb standard in negative ion mode; F: mass spectra from soyasaponin Bb standard in negative ion mode; G: mass spectra of soyasaponin Bb in the first mowing Medicago polymorpha in negative ion mode; H: MS/MS spectra of soyasaponin Bb in the first mowing Medicago polymorpha in negative ion mode; H: MS/MS spectra of soyasaponin Bb in the first mowing Medicago polymorpha in negative ion mode; H: MS/MS spectra of soyasaponin Bb in the first mowing Medicago polymorpha in negative ion mode.



Figure S4 The different levels of identified compounds in two mowing by ESI+ and ESI-



 $Figure \ S5 \ {\rm The \ appearance \ and \ sample \ number \ of \ Medicago \ polymorpha.}$