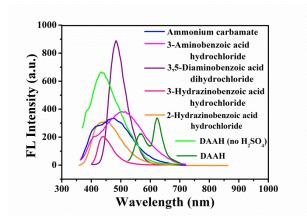
1	Supporting Information
2	Modification-Free Fabricating Ratiometric Nanoprobe Based on
3	Dual Emissive Carbon Dots for Nitrite Determination in Food
4	Samples
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- 17
- 18 Figure S1. Fluorescence spectra of the CDs prepared through using different carbon
- 19 sources and H_2SO_4 for CDs.
- 20

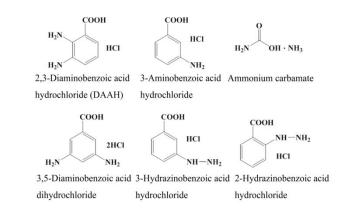


Figure S2. Structural formulas of DAAH, 3-aminobenzoicacid hydrochloride,
ammounium carbamate, 3,5-diaminobenzoic acid dihydrochloride,
3-hydrazinobenzoic acid hydrochloride, and 2-hydrazinobenzoic acid hydrochloride,
respectively.

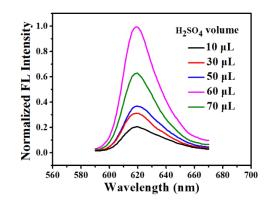
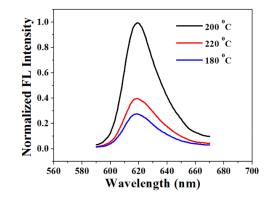


Figure S3. Effect of different volume of H_2SO_4 (98%) on the fluorescence intensity



30



31

32 Figure S4. Effect of different carbonization temperature on the fluorescence intensity

of the RYDE CDs.

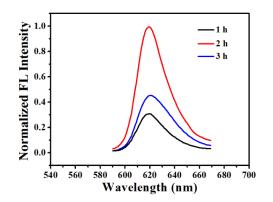


Figure S5. Effect of different carbonization time on the fluorescence intensity of the



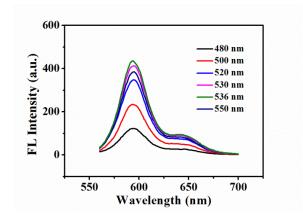
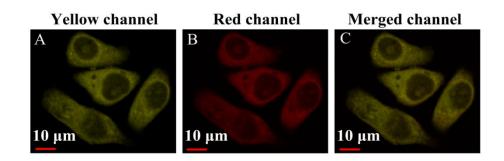


Figure S6. Fluorescence spectra scanned under different excitation wavelength of the

⁴¹ RODE CDs.

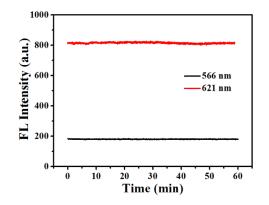


44 Figure S7. Confocal microscopy fluorescence images of HeLa cells treated with the

45 RYDE CDs (208 μ g/mL) in yellow channel, red channel and the merged image of the

46 two channels.

47



48

49 Figure S8. Fluorescence intensity variation of the RYDE CDs as a function of

50 illumination time.

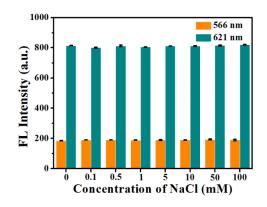
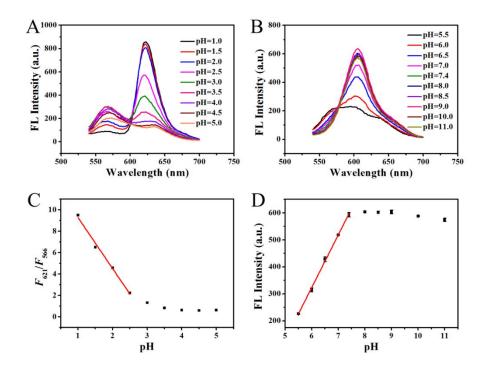




Figure S9. Fluorescence intensity variation of the RYDE CDs as a function of NaCl
concentrations. Three replicate measurements were completed for each point.



56

Figure S10. Fluorescence spectra of the RYDE CDs probe under (A) acid and (B) alkaline pH conditions. Effect of pH of buffer solution on the (C) fluorescence intensity ratio (F_{621}/F_{566}) and the (D) fluorescence intensity centered at 605 nm of the RYDE CDs solution.

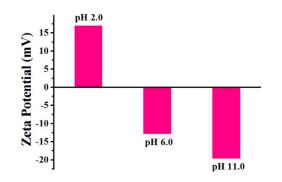
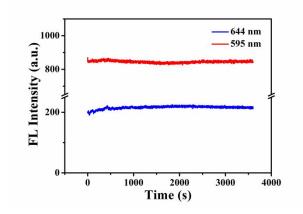


Figure S11. The zeta potential histogram of RYDE CDs under different pHconditions.



64

65 Figure S12. Fluorescence intensity variation of the RODE CDs as a function of

66	11	luminat	tion	time.

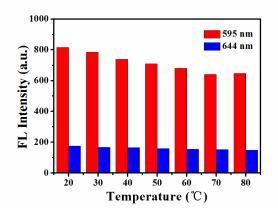


Figure S13. Fluorescence intensity variation of the RODE CDs as a function oftemperature.

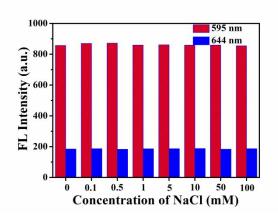
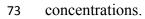
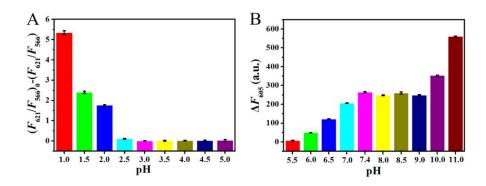




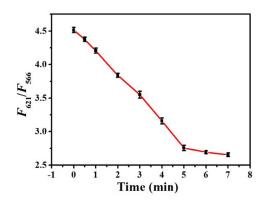
Figure S14. Fluorescence intensity variation of the RODE CDs as a function of NaCl





75

Figure S15. Effect of pH of buffer solution on $(F_{621}/F_{566})_0$ - (F_{621}/F_{566}) and ΔF_{605} . ($F_{621}/F_{566})_0$ and (F_{621}/F_{566}) were the fluorescence intensity of the RYDE CDs solution in the absence and presence of 50 μ M nitrite, ΔF_{605} was the difference of the fluorescence intensity centered at 605 nm in the absence and presence of 50 μ M nitrite, respectively. Three replicate measurements were completed for each point.



81

Figure S16. Effect of response time on F_{621}/F_{566} . F_{621} and F_{566} were the fluorescence intensity of the RYDE CDs solution at 621 nm and 566 nm in the presence of 50 μ M nitrite, respectively. Three replicate measurements were completed for each point.

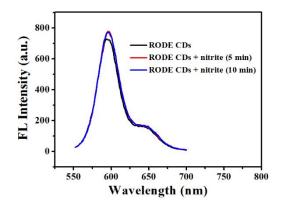


Figure S17. Fluorescence spectra of RODE CDs in ethanol solution in the absence
and presence (5 min or even 10 min) of 3.0 mM nitrite.

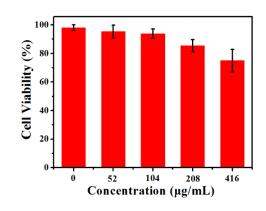


Figure S18. Cell viability of the HeLa cells under different concentrations (0, 52, 104, 208, 416 µg/mL) of the RYDE CDs. Three replicate measurements were completed
for each point.

Synthetic raw material	Preparation temperature (°C)	Preparation time (h)	Kind of CDs	Excitation wavelength (nm)	Emission wavelength (nm)	QY	Ref.
ascorbic acid, ethylene glycol	160/180	2	one	365	435/538	not mentioned	1
<i>m</i> -phenylenediamine, sulfuric acid	200	10	one	300	360/520	43%* (460 nm excitation)	2
<i>m</i> -aminophenol, oxalic acid	180	12	one	380	430/510	5.40% (380 nm excitation)	3
o-phenylenediamine, phosphoric acid	200	24	one	380	440/640	14.88%	4
2,5-diaminotoluene sulfate	150	12	one	380	525/603	9.00% (380 nm excitation)	5
DAAH	200	2	two	520 535	566/621 595/644	8% 18%	this work

Table S1. Comparison of the properties of the RYDE CDs with other CDs.

96 *: The QY was measured under 460 nm excitation wavelength, rather than 300 nm. When excited at 460 nm, the

97 CDs might be single emissive.

98

99 Table S2. Comparison of different kinds of CDs probes for the detection of nitrite.

Type of probe	Detection wavelength (nm)	Linear range (µM)	Limit of detection (nM)	Ref.	
CDs-neutral red	520	0-4.34	0.518	6	
N,P-CDs	530	0.01-0.09	3.3	7	
N-CDs	390	15-1110	13500	8	
N,P-GQDs	470	0.005-0.03	2.5	9	
N-CDs	417	0-1000	1000	10	
RYDE CDs	566/621	0.1-100	31.61	this work	

T (K)	Equation	R ²	$K_{\rm sv}$ (L·mol ⁻¹)	K_{q} (L·mol ⁻¹ ·s ⁻¹)
288	$(F_{621}/F_{566})_0/(F_{621}/F_{566})$ =0.9963+1.274×10 ⁻² [NO ₂ ⁻]	0.992	1.274×10 ⁴	8.85×10 ¹²
303	$(F_{621}/F_{566})_0/(F_{621}/F_{566})$ =0.9922+1.124×10 ⁻² [NO ₂ ⁻]	0.993	1.124×10 ⁴	7.81×10 ¹²
313	$(F_{621}/F_{566})_0/(F_{621}/F_{566})$ =0.9863+1.014×10 ⁻² [NO ₂ ⁻]	0.993	1.014×10 ⁴	7.04×10 ¹²

Table S3. The quenching parameters in Stern-Volmer equation.

105 Quantum yield (QY) measurements.

106 QY of the obtained RYDE CDs and RODE CDs was determined by the method 107 mentioned in our previous work.¹¹ The absolute fluorescence quantum yield can be 108 simply represented in the equation below:

109
$$QY = \frac{\int L_{\text{emission}}}{\int E_{\text{solvent}} - \int E_{\text{sample}}}$$
(1)

110 where QY was the absolute quantum yield, $L_{emission}$ was the fluorescence (FL) 111 emission spectrum of the sample, collected using the sphere; E_{sample} was the spectrum 112 of the light used to excite the sample, collected using the sphere; $E_{solvent}$ was the 113 spectrum of the light used for excitation with only the solvent in the sphere, collected 114 using the sphere. The solvent for RYDE CDs and RODE CDs were deionized water 115 and ethanol, respectively.

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