## Supporting Information

## Fusaristatin C, a Cyclic Lipodepsipeptide from *Pithomyces* sp. RKDO 1698

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Screening Protocols. Experimental procedures for measuring antimicrobial activity and cytotoxicity of fusaristatin C (1)



Figure S1. Cyclic depsipeptides from *Pithomyces* and their producing species.



**Figure S2.** UPLC-HRMS analysis of *Pithomyces* sp. RKDO 1698 EtOAc extracts in addition to ELSD and UV chromatograms. (A) Mycelia extract (**1** elutes at 3.86 min in the TIC) and (B) broth extract.



Figure S3. ESI+ HRMS spectrum of 1.



**Figure S4.** <sup>1</sup>H NMR spectrum (600 MHz, DMSO- $d_6$ ) of **1**.



**Figure S5.** <sup>13</sup>C NMR spectrum (600 MHz, DMSO- $d_6$ ) of **1**.



**Figure S6.** COSY NMR spectrum (600 MHz, DMSO- $d_6$ ) of **1**.



**Figure S7.** HSQC NMR spectrum (600 MHz, DMSO- $d_6$ ) of **1**. Blue contours are positively phased (methyl and methine) and red contours are negatively phased (methylene).



**Figure S8.** HMBC NMR spectrum (600 MHz, DMSO- $d_6$ ) of **1**.



**Figure S9.** TOCSY NMR spectrum (600 MHz, DMSO- $d_6$ ) of **1**.



**Figure S10.** ROESY NMR spectrum (600 MHz, DMSO- $d_6$ ) of **1**.



**Figure S11.** 1D ROESY NMR spectrum (400 MHz, DMSO- $d_6$ ) of **1**; H-2 (2.68 ppm) was selectively irradiated. Negatively phased peaks indicate a nOe between the corresponding proton(s) and H-2.



**Figure S12.** 1D ROESY NMR spectrum (400 MHz, DMSO- $d_6$ ) of **1**; H-3 (4.91 ppm) was selectively irradiated. Negatively phased peaks indicate a nOe between the corresponding proton(s) and H-3.



Figure S13. HETLOC NMR spectrum (400 MHz, DMSO-*d*<sub>6</sub>) of 1.



Figure S14. ESI+ MS2 of 1 and select predicted fragments.



**Figure S15.** Marfey's analysis of **1**. (A) TIC and (B) XIC 358.0993  $\pm$  10 ppm (retention time window of 0-30 min) for L-FDAA-derivatized serine derived from **1** and from pure serine standards. (C) ESI+ HRMS of L-FDAA-derivatized serine, which is representative of both pure standards and **1** samples.



Figure S16. ESI- HRMS spectrum of 6.



Figure S17. <sup>1</sup>H NMR spectrum (600 MHz, CD<sub>3</sub>OD) of **6**.



Figure S18. <sup>13</sup>C NMR spectrum (600 MHz, CD<sub>3</sub>OD) of **6**.



Figure S19. COSY NMR spectrum (600 MHz, CD<sub>3</sub>OD) of 6.



**Figure S20.** HSQC NMR spectrum (600 MHz, CD<sub>3</sub>OD) of **6**. Blue contours are positively phased (methyl and methine) and red contours are negatively phased (methylene).



Figure S21. HMBC NMR spectrum (600 MHz, CD<sub>3</sub>OD) of 6.

Position	δ <sub>c</sub> , Type	δ <sub>H</sub> , ( <i>J</i> in Hz)	<sup>1</sup> H- <sup>1</sup> H COSY	НМВС
1	177.5, C			
2	47.7, CH	2.54, m (7.1)	2-CH <sub>3</sub> , 3	1, 2-CH <sub>3</sub> , 3, 4
$2-CH_3$	13.8, CH <sub>3</sub>	1.11, d (7.1) 3.66, td (7.7,	2	1, 2, 3
3	74.5, CH	3.3)	2, 4a, 4b	1, 2, 2-CH <sub>3</sub> , 5
4	32.5, CH <sub>2</sub>	1.39, m	3, 4b	2
	32.5, CH <sub>2</sub> 33.25,	1.5, m	3, 4a	2
5	CH <sub>2</sub>	1.29, m		
6 <sup>a</sup>	33.2, CH <sub>2</sub>	1.29, m		
7 <sup>a</sup>	31.3, CH <sub>2</sub>	1.29, m		
8 <sup>a</sup>	30.9, CH <sub>2</sub>	1.29, m		
9 <sup>a</sup>	30.6, CH <sub>2</sub> 38.54,	1.29, m		
10	CH₂ 38.54,	1.16, m		
	$CH_2$	1.31, m		
11	33.9, CH <sub>2</sub>	1.41, m		
11-CH <sub>3</sub>	20.1, CH <sub>3</sub>	0.88, d (6.5)	11	10, 12
12	34.1, CH <sub>2</sub>	1.37, m		
13	23.9, CH <sub>2</sub>	1.31, m		
14	14.6, CH <sub>3</sub>	0.9, t (7.2)	13	12, 13

Table S1. Tabulated NMR spectroscopic data (600 MHz, CD<sub>3</sub>OD) for HDMT (6).



**Figure S22.** Mosher's ester analysis of **6** cleaved from **1**. (A) <sup>1</sup>H NMR spectra and (B) HSQC spectra (both 600 MHz,  $CDCI_3$ ) after reaction with *R*-MTPA-CI (red) and *S*-MTPA-CI (blue). HSQC contours corresponding to H4a and H4b are enclosed within the dashed box.



**Figure S23.** Dose-response curves illustrating results of cytotoxicity assays of **1**. Growth inhibition was observed for one healthy cell line (Vero) and three cancer cell lines (HTB-26, HTB-22 and HCT-116).

Antimicrobial and Cytotoxicity Screening Protocols. Compound 1 was tested for antimicrobial activity against methicillin-resistant Staphylococcus aureus ATCC 33591 (MRSA), S. warneri ATCC 17917, vancomycin-resistant Enterococcus faecium EF379 (VRE), Pseudomonas aeruginosa ATCC 14210, Proteus vulgaris ATCC 12454, and Candida albicans ATCC 14035. All testing was carried out in triplicate according to the Clinical Laboratory Standards Institute testing standards in a 96-well plate microbroth dilution assay as previously described.<sup>20</sup> Optical density was measured using a Thermo Scientific Varioskan Flash plate reader at 600 nm, recording at time zero and then again after incubation for 22 h (37.0 °C) to determine percent growth inhibition. Testing against Mycobacterium tuberculosis ATCC25177 was carried out using a previously reported protocol.<sup>21</sup> Fusaristatin C was also tested for cytotoxicity against Vero kidney cell line from African green monkey, adult human epidermal keratinocytes (HEKa), human foreskin BJ fibroblasts, MCF7 human breast adenocarcinoma (ATCC HTB-22), human breast adenocarcinoma cells (ATCC HTB-26) and HCT-116 human colorectal carcinoma cells (ATCC CCL-247). All assays were carried out as described previously.<sup>20</sup> Fluorescence was measured using a Thermo Scientific Varioskan Flash plate reader at 560/12 excitation, 590 nm emission both at time zero and 4 h after Alamar blue addition in order to calculate IC<sub>50</sub> values.

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