

Dynein light chain DLC-1 facilitates the function of GLD-1 germline cell fate regulator in *C. elegans*

Supplemental Materials

Mary Ellenbecker, Emily Osterli, Xiaobo Wang, Nicholas Day, Ella Baumgarten, Benjamin Hickey, and
Ekaterina Voronina

Including: Supplemental Tables 1 and 2, Supplemental Note, Supplemental Figures S1-S5

Supplemental Table 1. List of strains used in this study.

Genotype	Transgene description	Strain	Reference
Transgenes: GFP::H2B::3'UTR			
<i>unc-119(ed3) axIs1649 III</i>	<i>pie-1</i> prom::GFP::H2B:: <i>cye-1</i> 3'UTR + <i>unc-119(+)</i>	JH2261	Merritt et al., 2008
<i>unc-119(ed3) III; axIs1668</i>	<i>pie-1</i> prom::GFP::H2B:: <i>spn-4</i> 3'UTR + <i>unc-119(+)</i>	JH2311	Merritt et al., 2008
<i>unc-119(ed3) III; axIs1713</i>	<i>pie-1</i> prom::GFP::H2B:: <i>mex-3</i> 3'UTR + <i>unc-119(+)</i>	JH2377	Merritt et al., 2008
<i>unc-119(ed3) III; axIs1688</i>	<i>pie-1</i> prom::GFP::H2B:: <i>mex-3</i> 3'UTR + <i>unc-119(+)</i>	JH2333	Merritt et al., 2008
<i>unc-119(ed3) III; axIs1721</i>	<i>pie-1</i> prom::GFP::H2B:: <i>puf-5</i> 3'UTR + <i>unc-119(+)</i>	JH2418	Merritt et al., 2008
<i>unc-119(ed3) III; rrrIs1</i>	<i>pie-1</i> prom::GFP::H2B:: <i>cye-1</i> 3'UTR + <i>unc-119(+)</i>	RAF1	Biedermann et al., 2009
<i>dhc-1(js121) I/hT2 I; axIs1649/hT2 III</i>	<i>pie-1</i> prom::GFP::H2B:: <i>cye-1</i> 3'UTR + <i>unc-119(+)</i>	UMT377	this study
Transgenes: ORF+3'UTR			
<i>gld-1(q485) I; mntSi19 [pME5.13;GLD-1 wt] II</i>	<i>gld-1</i> prom::GLD-1 ^{wt} ::OLLAS:: <i>gld-1</i> 3'UTR	UMT327	this study
<i>gld-1(q485) I/hT2 (I;III); mntSi18 [pME5.17; GLD-1 ndb mutant] V</i>	<i>gld-1</i> prom::GLD-1 ^{ndb} ::OLLAS:: <i>gld-1</i> 3'UTR	UMT314	this study
<i>gfp::3xflag::mex-3 gld-1(q495) I; mntSi19 II</i>	<i>gld-1</i> prom::GLD-1 ^{wt} ::OLLAS:: <i>gld-1</i> 3'UTR	UMT384	this study
<i>gfp::3xflag::mex-3 gld-1(q495) I/hT2 (I;III); mntSi18 V</i>	<i>gld-1</i> prom::GLD-1 ^{ndb} ::OLLAS:: <i>gld-1</i> 3'UTR	UMT374	this study
CRISPR tagged strains			
<i>spn-4(tn1699[spn-4::gfp::3xflag]) V</i>	-	DG4185	Tsukamoto et al., 2017
<i>dlc-1(tm3153)/qC1 III; spn-4(tn1699[spn-4::gfp::3xflag]) V</i>	-	UMT356	this study
<i>puf-5(tn1726[gfp::3xflag::puf-5]) II</i>	-	DG4215	Tsukamoto et al., 2017
<i>puf-5(tn1726[gfp::3xflag::puf-5]) II; dlc-1(tm3153) III/hT2 (I;III)</i>	-	UMT359	this study
<i>mex-3(tn1753[gfp::3xflag::mex-3]) I</i>	-	DG4269	Tsukamoto et al., 2017
<i>mex-3(tn1753[gfp::3xflag::mex-3]) dhc-1(js121) I/hT2 (I;III)</i>	-	UMT364	this study
Mutant Strains; no transgene			
<i>dlc-1(tm3153)/qC1 III</i>	-	UMT222	Wang et al., 2016
<i>gld-3(q730)/mIn1 [mIs14 dpy-10(e128)] II; him-5(e1490) V</i>	-	JK3375	Eckmann et al., 2002
<i>nos-3(q650) II</i>	-	JK2589	Kraemer et al., 1999
<i>gld-2(q497)/dpy-5(e61) unc-13(e51) I</i>	-	JK1743	Kadyk and Kimble, 1998
<i>dhc-1(or195) I</i>	-	EU828	Hamill et al., 2002
<i>gld-1(q485) I/hT2 [bli-4(e937) let-?(q782) qIs48] (I;III)</i>	-	JK3025	Francis et al., 1995a
<i>dhc-1(js121) I/hT2 (I;III); jsIs37 V</i>	-	NM2040	Koushika et al., 2004
<i>gld-2(q497) I/hT2 (I;III)</i>	-	UMT235	this study
<i>gld-3(q730)/mT1 II; dlc-1(tm3153)/mT1 III</i>	-	UMT317	this study
<i>nos-3(q650)/mT1 II; dlc-1(tm3153)/mT1 III</i>	-	UMT318	this study
<i>gld-2(q497)/hT2 I; dlc-</i>	-	UMT319	this study

<i>1(tm3153)/hT2 III</i>			
<i>dhc-1(or195) I; gld-3(q730)/mIn1 II</i>	-	UMT325	this study
<i>ttTi5605(mos) II; unc-119(ed3) III; oxEx1578[eftp::GFP+Cbr-unc-119]</i>	-	EG6699	Frøkjær-Jensen et al., 2008
<i>unc119(ed3) III; oxTi365 V</i>	-	EG8082	Frøkjær-Jensen et al., 2014
<i>gld-1(q485)/hT2 I; dlc-1(tm3153)/hT2 III; him-8(tm611) IV</i>	-	UMT350	this study
<i>gld-1(op236) I</i>	-	TG34	Schumacher et al., 2005
<i>gld-1(op236)/hT2 I; dlc-1(tm3153)/hT2 III</i>	-	UMT383	this study
<i>dlc-1(tm3153) unc-32(e189) glp-1(q46) III/hT2 (I; III)</i>	-	UMT387	this study
<i>gld-3(q730)/mT1 II; dlc-1(tm3153) unc-32(e189) glp-1(q46)/mT1 III</i>	-	UMT388	this study
<i>gld-2(q497)/hT2 I; dlc-1(tm3153) unc-32(e189) glp-1(q46)/hT2 III</i>	-	UMT390	this study

Supplemental Table 2. Analysis of tagged protein or reporter transgene repression in meiotic pachytene region of germline.

Genotype	% derepressed	<i>n</i>
<i>spn-4::gfp::3xflag V</i>	0	64
<i>dlc-1(tm3153) III; spn-4::gfp::3xflag V</i>	0	34
<i>gfp::3xflag::puf-5 II</i>	0	51
<i>gfp::3xflag::puf-5 II; dlc-1(tm3153) III</i>	0	22
<i>axIs1649 (pie-1 prom::GFP::H2B::cye-1 3'UTR) III</i>	0	30
<i>dhc-1(js121) I; axIs1649 (pie-1 prom::GFP::H2B::cye-1 3'UTR) III</i>	0	18
<i>gfp::3xflag::mex-3 I</i>	0	35
<i>gfp::3xflag::mex-3 dhc-1(js121) I</i>	0	22
<i>gfp::3xflag::mex-3 gld-1(q485) I; gld-1::ollas(wt) II</i>	0	43
<i>gfp::3xflag::mex-3 gld-1(q485) I; gld-1::ollas(ndb) V</i>	0	35

n = number of germlines scored for repression of gene expression in meiotic phase cells. Scoring was 1 day after L4 stage.

Supplemental Note

Dissected gonads from both single heterozygous *gld-1(-)/hT2* and double heterozygous *gld-1(-)/hT2; dlc-1(-)/hT2* animals resemble wild type germlines where proliferative cells are restricted to the distal end of the gonad (Fig. S1A and C). Homozygous *gld-1(-)* mutant germline exhibits proximal tumor phenotype where germ cells enter meiosis but exit meiotic prophase early and continue to proliferate (Francis et al., 1995b Francis et al., 1995a Jones et al., 1996) (Fig. S1B). Homozygous *gld-1(-); dlc-1(-)* double mutant germline exhibits same tumor phenotype as *gld-1(-)*, however, some germ cell nuclei appear larger which is reminiscent of *dlc-1(-)* mutant phenotype (Fig. S1D).

Supplemental Figures

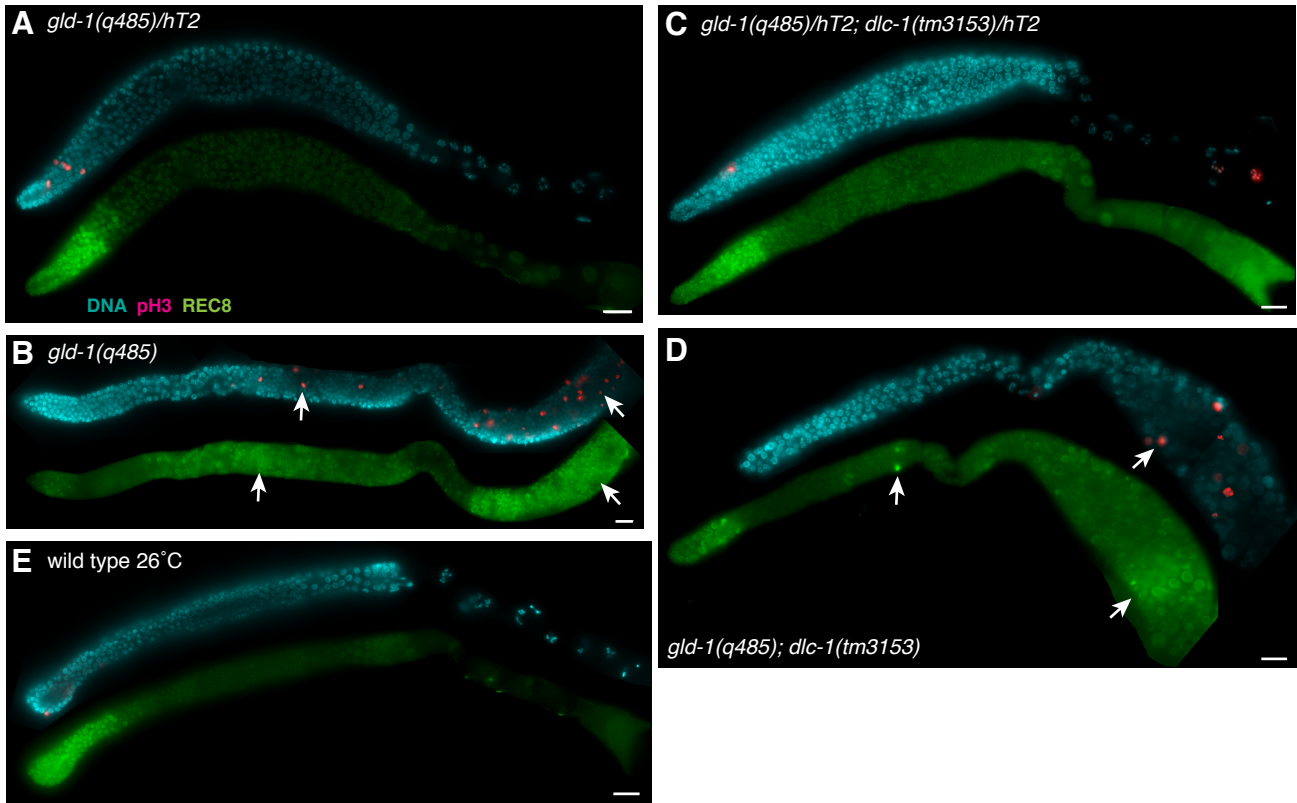


Figure S1. Germline phenotypes of heterozygous and homozygous *gld-1(-)* and *gld-1(-); dlc-1(-)* mutant alleles as well as of wild type control cultured at 25°C. Dissected gonads were stained for the M-phase marker pH3 (pink), stem and progenitor cells marker REC-8 (green), and DNA (blue). (A) Germline from a heterozygous mutant animal *gld-1(q485)/hT2*. (B) Dissected gonad from single mutant animal *gld-1(q485)* (null allele). (C) Dissected gonad from double heterozygous animal *gld-1(q485)/hT2; dlc-1(tm3153)/hT2*. (D) Dissected gonad from double mutant animal *gld-1(q485); dlc-1(q485)*. White arrows indicate aberrant cell proliferation (tumor). (E) Wild type control cultured at 26°C. Scale bars: 10 μm.

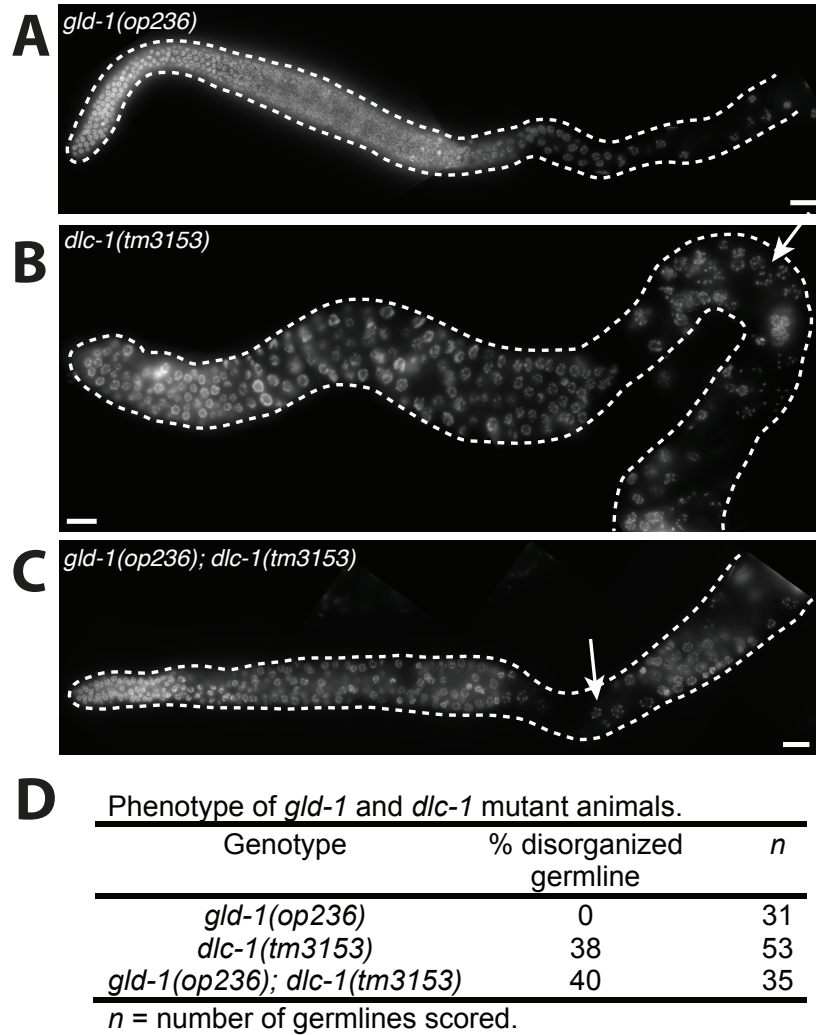


Figure S2. No synthetic interactions between *dlc-1* null allele and a weak *gld-1(op236)* mutant. Germlines in A-C were dissected and chromatin was stained by DAPI. (A) *gld-1(op236)* germlines are superficially normal at 20°C. (B) A subset of *dlc-1(tm3153)* germlines at 20°C exhibit patches of pachytene cells unable to progress to diplotene (white arrow). (C) *gld-1(op236); dlc-1(tm3153)* germlines at 20°C showing disorganized germline phenotypes with diplotene cells interspersed with cells unable to progress from pachytene. (D) Quantification of disorganized germline phenotypes in the indicated genetic backgrounds at 20°C, 1 day after L4 stage.

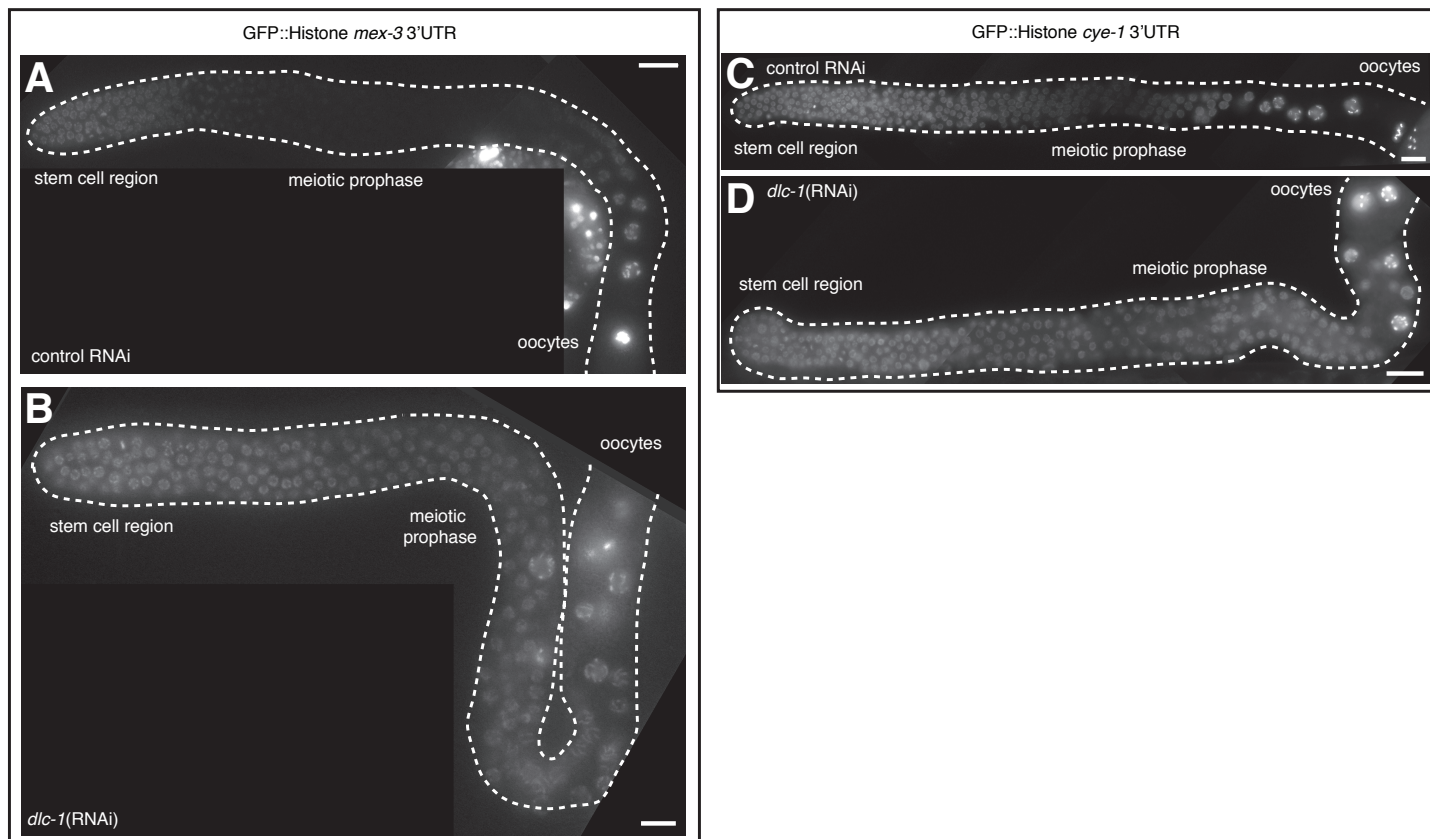


Figure S3. DLC-1 facilitates regulation of *mex-3* and *cye-1* GLD-1 target mRNAs. Fluorescent micrographs of dissected gonads expressing GFP-tagged Histone H2B under control of *mex-3* 3'UTR after either (A) control or (B) *dlc-1*(RNAi). Fluorescent micrographs of dissected gonads expressing GFP-tagged Histone H2B under control of *cye-1* 3'UTR after either (C) control or (D) *dlc-1*(RNAi). This *cye-1* transgenic reporter strain was generated in a different laboratory than the other transgenic reporters used in this study, however, it yielded similar expression pattern and results obtained using the *cye-1* transgenic reporter strain generated by Merritt et. al. (Biedermann et al., 2009, Merritt et al., 2008) (Fig. 5A). Scale bars: 10 μ m.

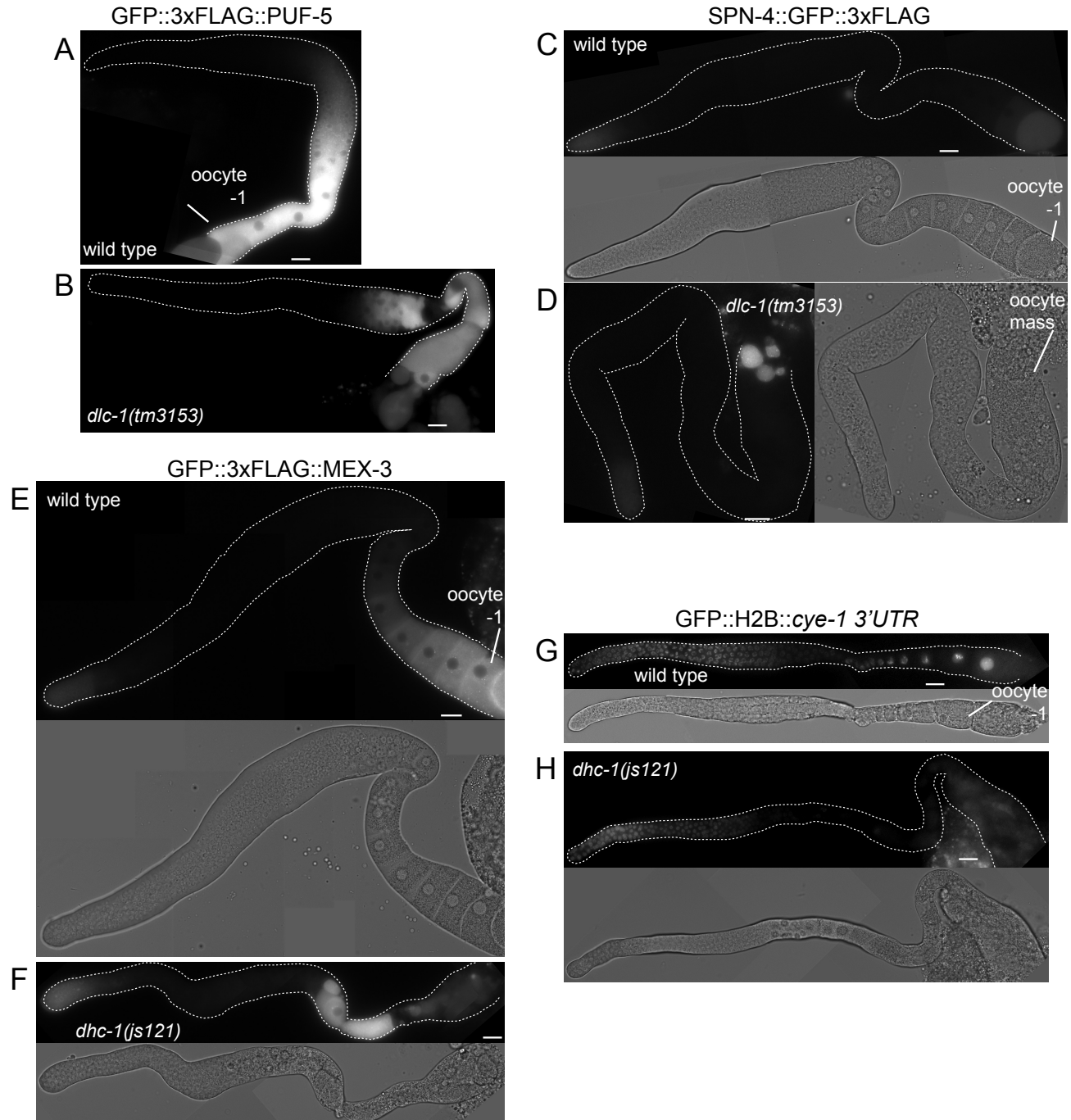


Figure S4. Regulation of *puf-5* and *spn-4* is independent of DLC-1, and regulation of *mex-3* and *cye-1* is independent of dynein motor. Fluorescent micrograph of dissected gonad expressing GFP::3xFLAG::PUF-5 in wild type (A) and *dlc-1(tm3153)* (B) genetic background. Fluorescent and brightfield micrographs of dissected gonads expressing SPN-4::GFP::3xFLAG in wild type (C) and *dlc-1(tm3153)* (D) genetic background. Fluorescent and brightfield micrographs of dissected gonads expressing GFP::3xFLAG::MEX-3 in wild type (E) and *dhc-1(js121)* (F) mutant background. GFP::H2B::*cye-1* 3'UTR in wild type (G) and *dhc-1(js121)* (H) mutant background. Scale bars: 10 μ m.

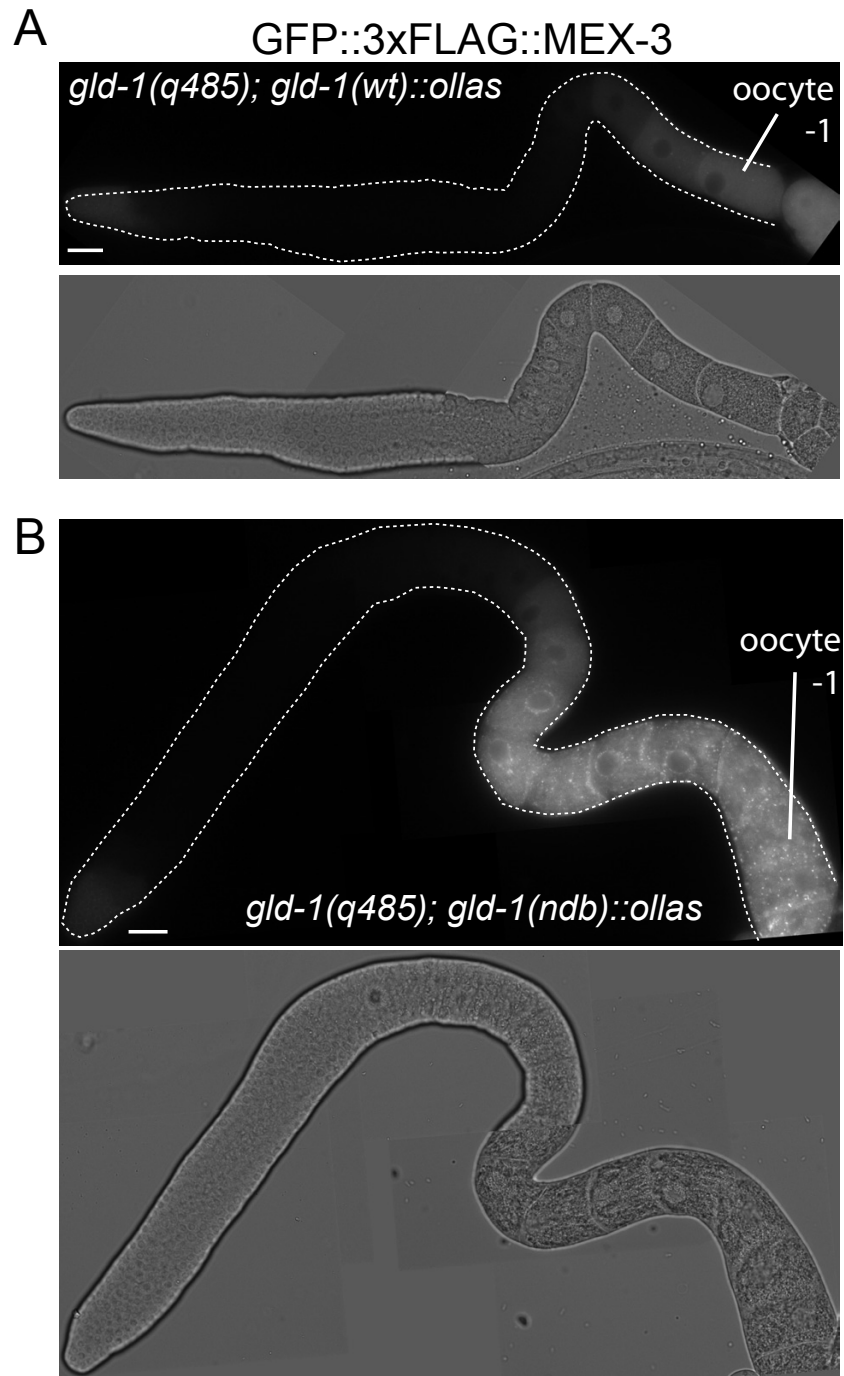


Figure S5. DLC-1 binding GLD-1 is not required to regulate MEX-3 expression *in vivo*. Fluorescent and brightfield micrographs of dissected gonads expressing GFP::3XFLAG::MEX-3 in (A) *gld-1(q485); gld-1^{wt}::ollas* and (B) *gld-1(q485); gld-1^{ndb}::ollas* genetic background. Scale bars: 10 μ m.

Supplemental References

- BIEDERMANN, B., J. WRIGHT, M. SENFTEN, I. KALCHHAUSER, G. SARATHY, M. H. LEE, and R. CIOSK, 2009 Translational repression of cyclin E prevents precocious mitosis and embryonic gene activation during *C. elegans* meiosis. *Dev Cell* **17**: 355-364.
- ECKMANN, C. R., B. KRAEMER, M. WICKENS, and J. KIMBLE, 2002 GLD-3, a bicaudal-C homolog that inhibits FBF to control germline sex determination in *C. elegans*. *Dev Cell* **3**: 697-710.
- FRANCIS, R., M. K. BARTON, J. KIMBLE, and T. SCHEDL, 1995a *gld-1*, a tumor suppressor gene required for oocyte development in *Caenorhabditis elegans*. *Genetics* **139**: 579-606.
- FRANCIS, R., E. MAINE, and T. SCHEDL, 1995b Analysis of the multiple roles of *gld-1* in germline development: interactions with the sex determination cascade and the *glp-1* signaling pathway. *Genetics* **139**: 607-630.
- FRØKJAER-JENSEN, C., M. W. DAVIS, C. E. HOPKINS, B. J. NEWMAN, J. M. THUMMEL, S. P. OLESEN, M. GRUNNET, and E. M. JORGENSEN, 2008 Single-copy insertion of transgenes in *Caenorhabditis elegans*. *Nat Genet* **40**: 1375-1383.
- FRØKJÆR-JENSEN, C., M. W. DAVIS, M. SAROV, J. TAYLOR, S. FLIBOTTE, M. LABELLA, A. POZNIAKOVSKY, D. G. MOERMAN, and E. M. JORGENSEN, 2014 Random and targeted transgene insertion in *Caenorhabditis elegans* using a modified *Mos1* transposon. *Nat Methods* **11**: 529-534.
- HAMILL, D. R., A. F. SEVERSON, J. C. CARTER, and B. BOWERMAN, 2002 Centrosome maturation and mitotic spindle assembly in *C. elegans* require SPD-5, a protein with multiple coiled-coil domains. *Dev Cell* **3**: 673-684.
- JONES, A. R., R. FRANCIS, and T. SCHEDL, 1996 GLD-1, a cytoplasmic protein essential for oocyte differentiation, shows stage- and sex-specific expression during *Caenorhabditis elegans* germline development. *Dev Biol* **180**: 165-183.
- KADYK, L. C., and J. KIMBLE, 1998 Genetic regulation of entry into meiosis in *Caenorhabditis elegans*. *Development* **125**: 1803-1813.
- KOUSHIKA, S. P., A. M. SCHAEFER, R. VINCENT, J. H. WILLIS, B. BOWERMAN, and M. L. NONET, 2004 Mutations in *Caenorhabditis elegans* cytoplasmic dynein components reveal specificity of neuronal retrograde cargo. *J Neurosci* **24**: 3907-3916.
- KRAEMER, B., S. CRITTENDEN, M. GALLEGOS, G. MOULDER, R. BARSTEAD, J. KIMBLE, and M. WICKENS, 1999 NANOS-3 and FBF proteins physically interact to control the sperm-oocyte switch in *Caenorhabditis elegans*. *Curr Biol* **9**: 1009-1018.
- MERRITT, C., D. RASOLOSON, D. KO, and G. SEYDOUX, 2008 3' UTRs are the primary regulators of gene expression in the *C. elegans* germline. *Curr Biol* **18**: 1476-1482.
- SCHUMACHER, B., M. HANAZAWA, M. H. LEE, S. NAYAK, K. VOLKMANN, E. R. HOFMANN, R. HOFMANN, M. HENGARTNER, T. SCHEDL, and A. GARTNER, 2005 Translational repression of *C. elegans* p53 by GLD-1 regulates DNA damage-induced apoptosis. *Cell* **120**: 357-368.
- TSUKAMOTO, T., M. D. GEARHART, C. A. SPIKE, G. HUELIGAS-MORALES, M. MEWS, P. R. BOAG, T. H. BEILHARZ, and D. GREENSTEIN, 2017 LIN-41 and OMA Ribonucleoprotein Complexes Mediate a Translational Repression-to-Activation Switch Controlling Oocyte Meiotic Maturation and the Oocyte-to-Embryo Transition in. *Genetics* **206**: 2007-2039.
- WANG, X., J. R. OLSON, D. RASOLOSON, M. ELLENBECKER, J. BAILEY, and E. VORONINA, 2016 Dynein light chain DLC-1 promotes localization and function of the PUF protein FBF-2 in germline progenitor cells. *Development* **143**: 4643-4653.