

# High-Throughput Toxicokinetics for Rapid Risk Prioritization

*John Wambaugh*

*National Center for Computational Toxicology  
Office of Research and Development, U.S. EPA,*

**NARROWING THE IN VITRO TO IN VIVO  
TRANSLATION GAP**

**Predicting Drug Toxicity  
June 21, 2018**

The views expressed in this presentation are those of the author and  
do not necessarily reflect the views or policies of the U.S. EPA

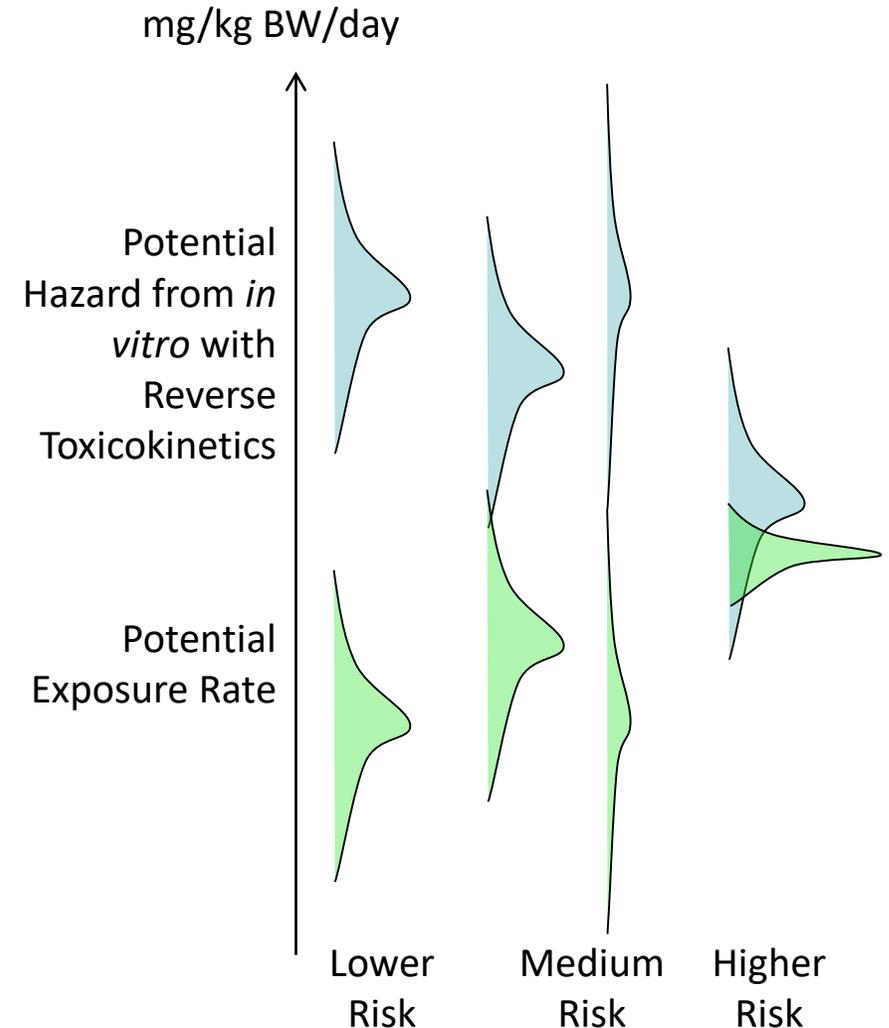
# Chemical Regulation in the United States

- Park *et al.* (2012): At least 3221 chemicals in pooled human blood samples, many appear to be exogenous
- A tapestry of laws covers the chemicals people are exposed to in the United States (Breyer, 2009)
- Different testing requirements exist for food additives, pharmaceuticals, and pesticide active ingredients (NRC, 2007)
- Most other chemicals, ranging from industrial waste to dyes to packing materials, are covered by the Toxic Substances Control Act (TSCA) and regulated by EPA
- TSCA was updated in June, 2016 and new approach methodologies (NAMs) are being considered prioritize these existing and new chemicals for testing



# Chemical Risk = Hazard + Exposure

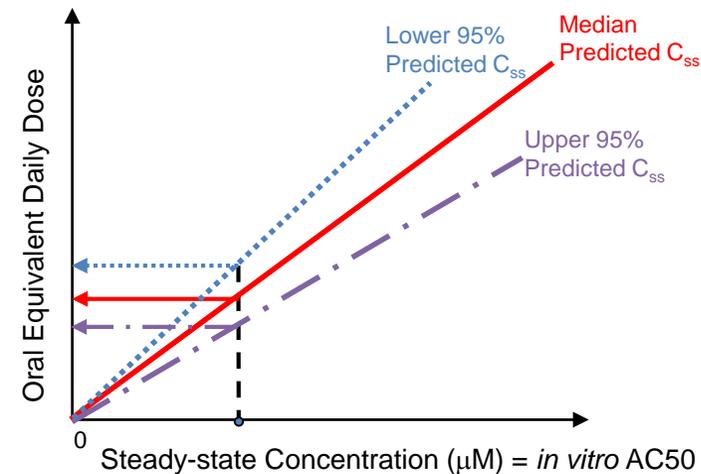
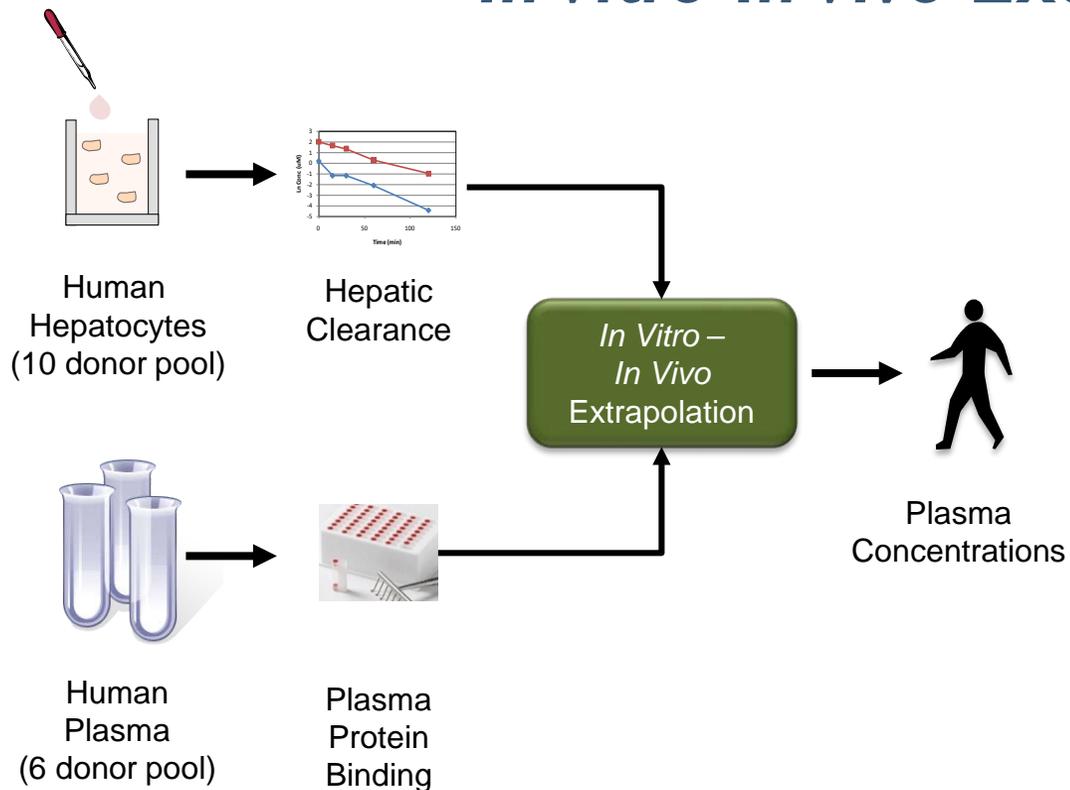
- National Research Council (1983) identified chemical risk as a function of both inherent hazard and exposure
- To address thousands of chemicals, we need to use NAMs to prioritize those chemicals most worthy of additional study
- **High throughput risk prioritization** needs:
  1. high throughput **hazard** characterization (Dix et al., 2007, Collins et al., 2008)
  2. high throughput **exposure** forecasts (Wambaugh et al., 2013, 2014)
  3. high throughput **toxicokinetics** (*i.e.*, dose-response relationship) linking hazard and exposure



Rotroff et al. (2010)

Wetmore et al. (2012, 2014, 2015)

# High-Throughput Toxicokinetics (HTTK) for *In Vitro-In Vivo* Extrapolation (IVIVE)

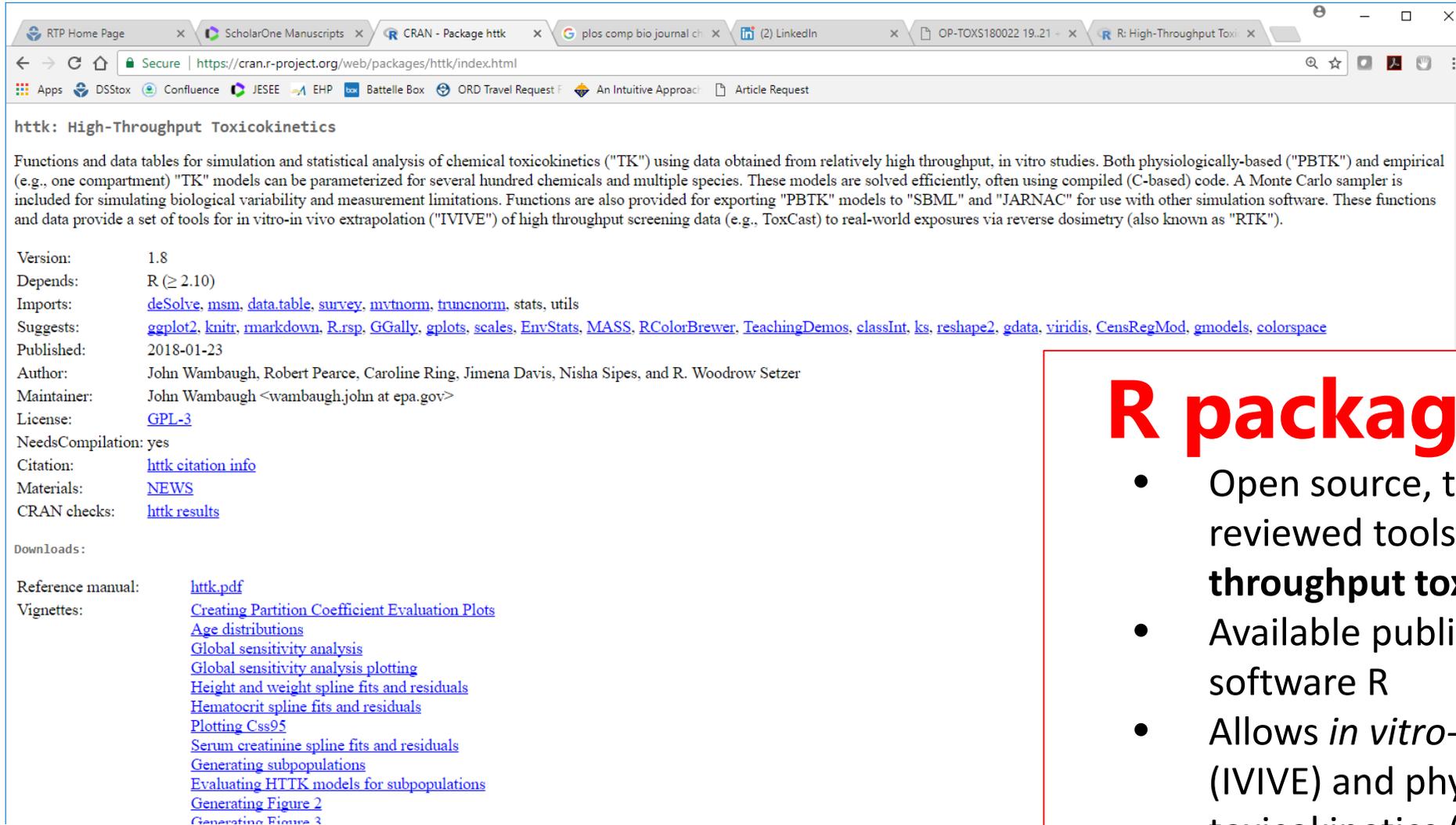


Rotroff et al. (2010) 35 chemicals  
Wetmore et al. (2012) +204 chemicals  
Wetmore et al. (2015) +163 chemicals  
Wambaugh et al. (in prep.) + ~300 chemicals

- **Most chemicals do not have TK data** – we use *in vitro* HTTK methods adapted from pharma to fill gaps
- In drug development, HTTK methods allow IVIVE to estimate therapeutic doses for clinical studies – predicted concentrations are typically on the order of values measured in clinical trials (Wang, 2010)

# Open Source Tools and Data for HTTK

<https://CRAN.R-project.org/package=httk>



The screenshot shows the CRAN package page for 'httk'. The browser tabs include 'RTP Home Page', 'ScholarOne Manuscripts', 'CRAN - Package httk', 'plos comp bio journal ch', '(2) LinkedIn', 'OP-TOXS180022 19.21', and 'R: High-Throughput Tox'. The address bar shows the URL 'https://cran.r-project.org/web/packages/httk/index.html'. The page content includes the package name 'httk: High-Throughput Toxicokinetics', a description of its functions, and various metadata fields.

**httk: High-Throughput Toxicokinetics**

Functions and data tables for simulation and statistical analysis of chemical toxicokinetics ("TK") using data obtained from relatively high throughput, in vitro studies. Both physiologically-based ("PBTk") and empirical (e.g., one compartment) "TK" models can be parameterized for several hundred chemicals and multiple species. These models are solved efficiently, often using compiled (C-based) code. A Monte Carlo sampler is included for simulating biological variability and measurement limitations. Functions are also provided for exporting "PBTk" models to "SBML" and "JARNAC" for use with other simulation software. These functions and data provide a set of tools for in vitro-in vivo extrapolation ("IVIVE") of high throughput screening data (e.g., ToxCast) to real-world exposures via reverse dosimetry (also known as "RTK").

Version: 1.8  
Depends: R (≥ 2.10)  
Imports: [deSolve](#), [msm](#), [data.table](#), [survey](#), [mvtnorm](#), [truncnorm](#), stats, utils  
Suggests: [ggplot2](#), [knitr](#), [rmarkdown](#), [R.rsp](#), [GGally](#), [gplots](#), [scales](#), [EnvStats](#), [MASS](#), [RColorBrewer](#), [TeachingDemos](#), [classInt](#), [ks](#), [reshape2](#), [gdata](#), [viridis](#), [CensRegMod](#), [gmodels](#), [colorspace](#)  
Published: 2018-01-23  
Author: John Wambaugh, Robert Pearce, Caroline Ring, Jimena Davis, Nisha Sipes, and R. Woodrow Setzer  
Maintainer: John Wambaugh <wambaugh.john at epa.gov>  
License: [GPL-3](#)  
NeedsCompilation: yes  
Citation: [httk citation info](#)  
Materials: [NEWS](#)  
CRAN checks: [httk results](#)

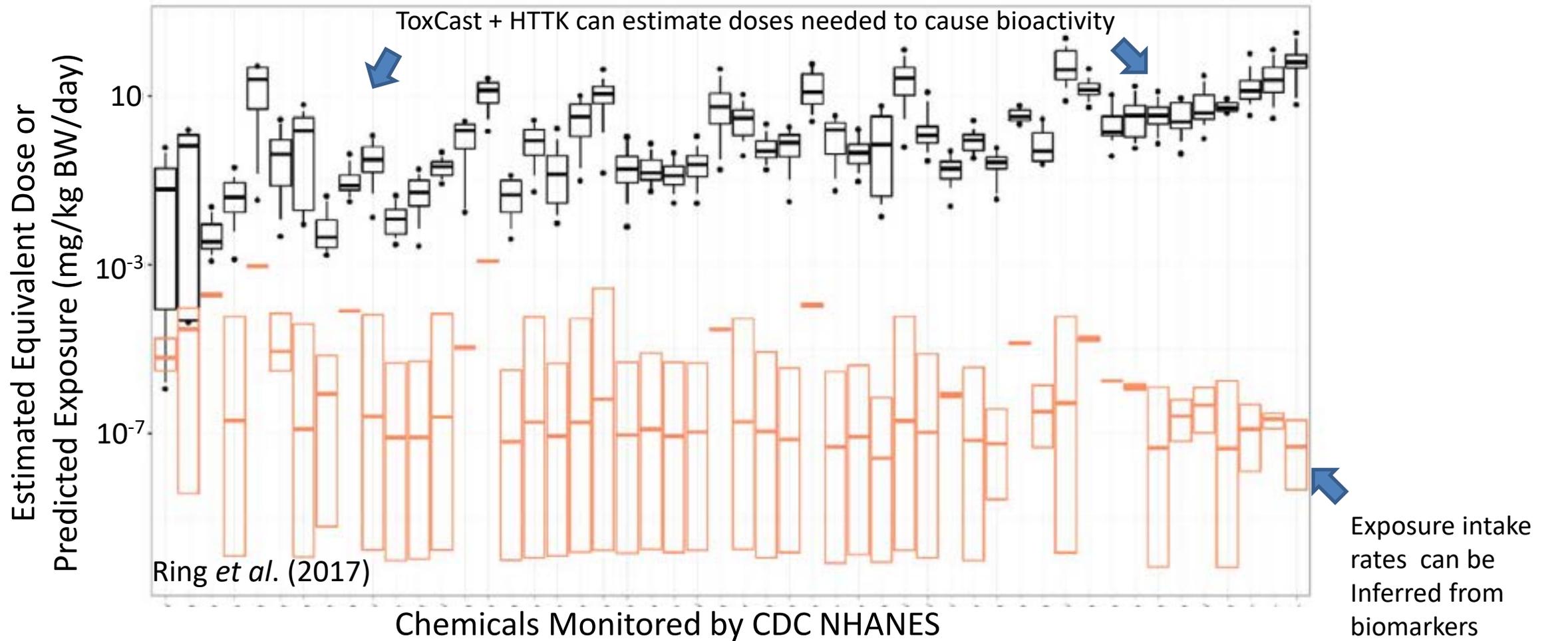
Downloads:

Reference manual: [httk.pdf](#)  
Vignettes: [Creating Partition Coefficient Evaluation Plots](#), [Age distributions](#), [Global sensitivity analysis](#), [Global sensitivity analysis plotting](#), [Height and weight spline fits and residuals](#), [Hematocrit spline fits and residuals](#), [Plotting C<sub>ss95</sub>](#), [Serum creatinine spline fits and residuals](#), [Generating subpopulations](#), [Evaluating HTTK models for subpopulations](#), [Generating Figure 2](#), [Generating Figure 3](#)

## R package "httk"

- Open source, transparent, and peer-reviewed tools and data for **high throughput toxicokinetics (httk)**
- Available publicly for free statistical software R
- Allows *in vitro-in vivo* extrapolation (IVIVE) and physiologically-base toxicokinetics (PBTk)

# High Throughput Risk Prioritization



National Health and Nutrition Examination Survey (NHANES) is an ongoing survey that covers ~10,000 people every two years

Most NHANES chemicals do not have traditional PK models (Strope et al., 2018)

# Variability

Different crayons have different colors, and none of them are the “average” color



# Variability

Different crayons have different colors, and none of them are the “average” color



# Population simulator for HTTK



Correlated Monte Carlo sampling of physiological model parameters built into R “httk” package (Pearce et al., 2017):

*Sample* NHANES biometrics for actual individuals:

- Sex
- Race/ethnicity
- Age
- Height
- Weight
- Serum creatinine

# Population simulator for HTTK



Correlated Monte Carlo sampling of physiological model parameters built into R “httk” package (Pearce et al., 2017):

*Sample* NHANES biometrics for actual individuals:

Sex  
Race/ethnicity  
Age  
Height  
Weight  
Serum creatinine



Regression equations from literature (McNally *et al.*, 2014)  
(+ residual marginal variability)

(Similar approach used in SimCYP [Jamei et al. 2009], GastroPlus, PopGen [McNally et al. 2014], P3M [Price et al. 2003], physB [Bosgra et al. 2012], etc.)

# Population simulator for HTTK



Correlated Monte Carlo sampling of physiological model parameters built into R “httk” package (Pearce et al., 2017):

Sample NHANES biometrics for actual individuals:

- Sex
- Race/ethnicity
- Age
- Height
- Weight
- Serum creatinine

Predict physiological quantities

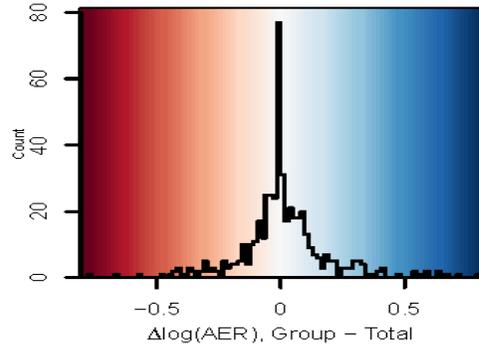
- Tissue masses
- Tissue blood flows
- GFR (kidney function)
- Hepatocellularity

Regression equations from literature (McNally *et al.*, 2014) (+ residual marginal variability)

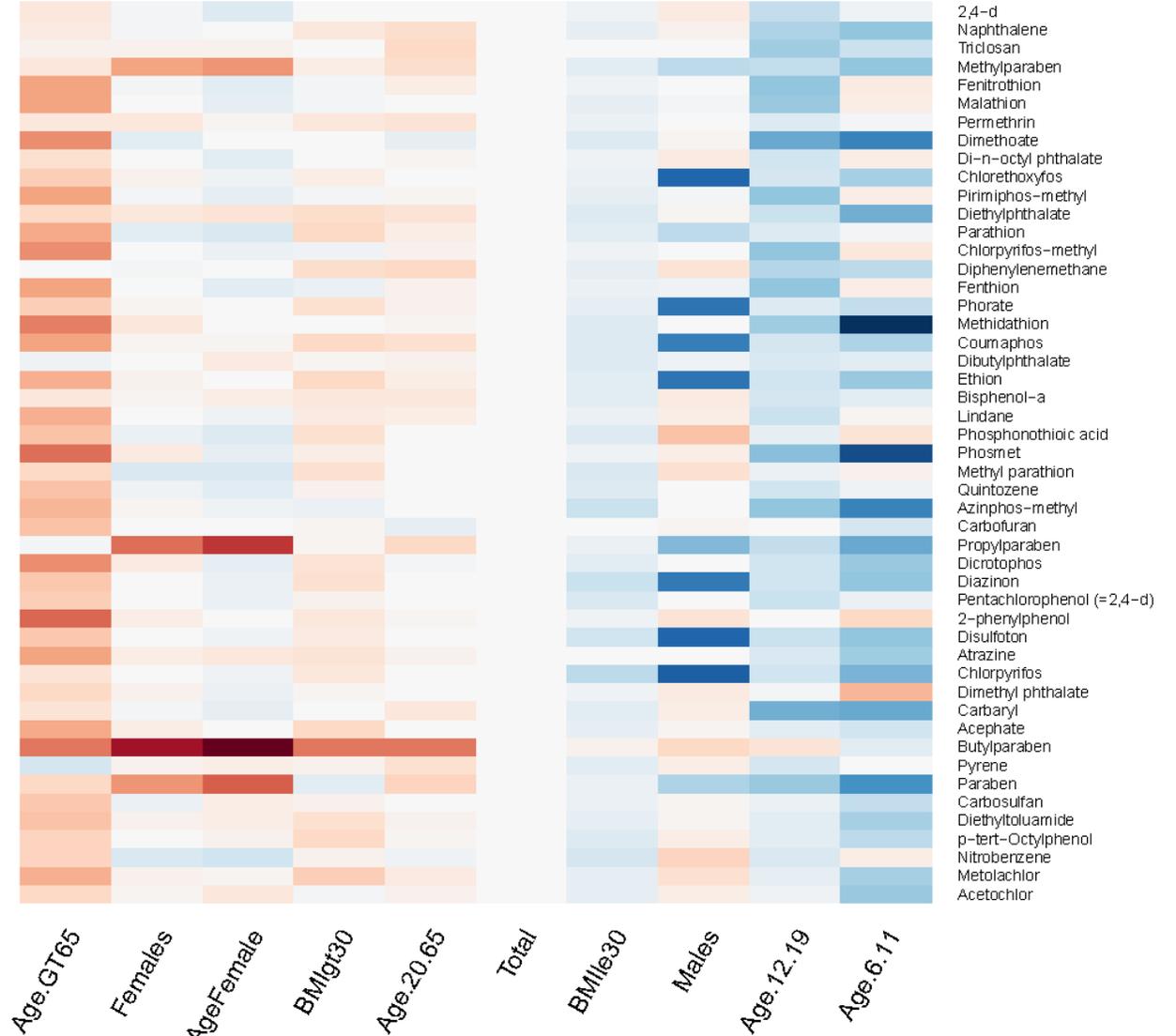
(Similar approach used in SimCYP [Jamei et al. 2009], GastroPlus, PopGen [McNally et al. 2014], P3M [Price et al. 2003], physB [Bosgra et al. 2012], etc.)

# Life-stage and Demographic Specific Predictions

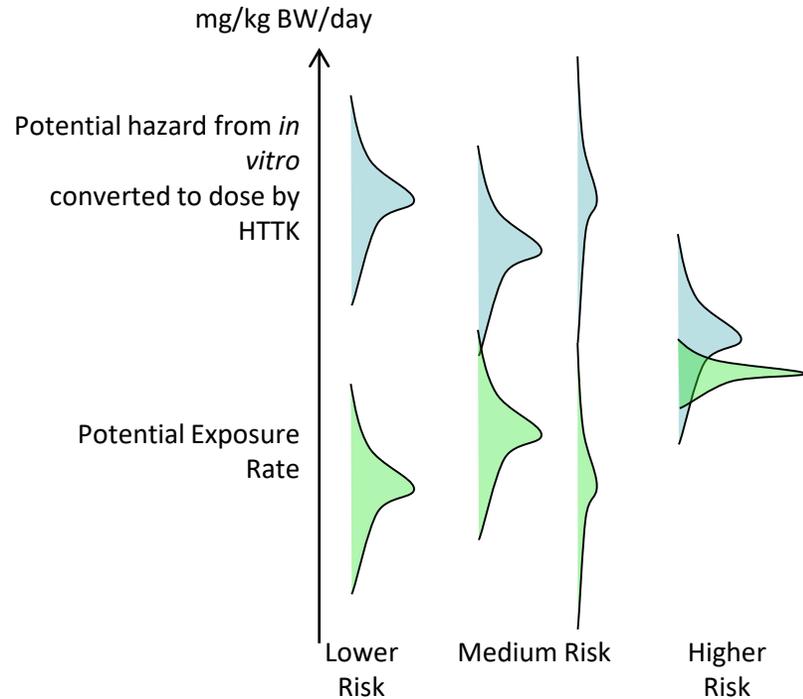
- We use HTTK to calculate margin between bioactivity and exposure for specific populations



## Change in Activity : Exposure Ratio



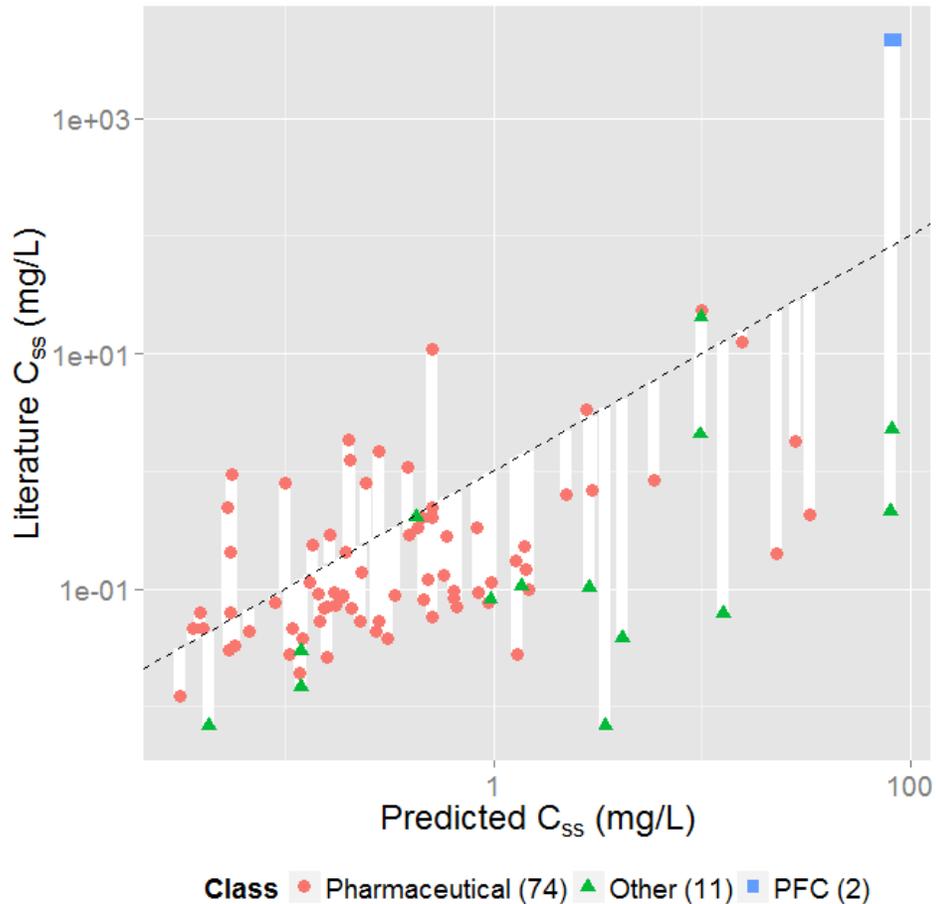
NHANES Chemicals



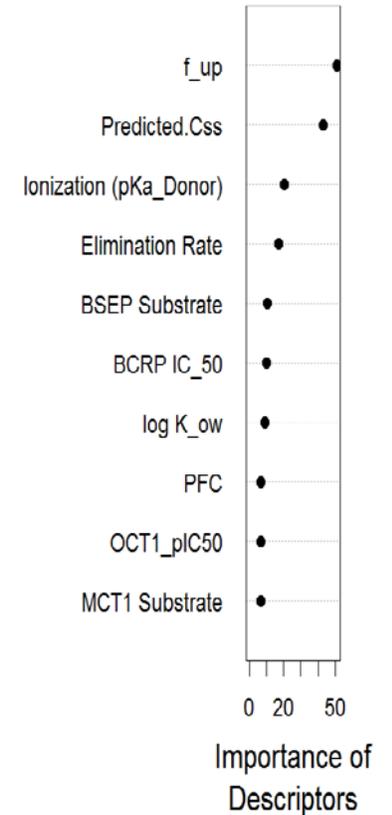
NHANES Demographic Groups

Ring et al. (2017)

# Using *in vivo* Data to Evaluate RTK

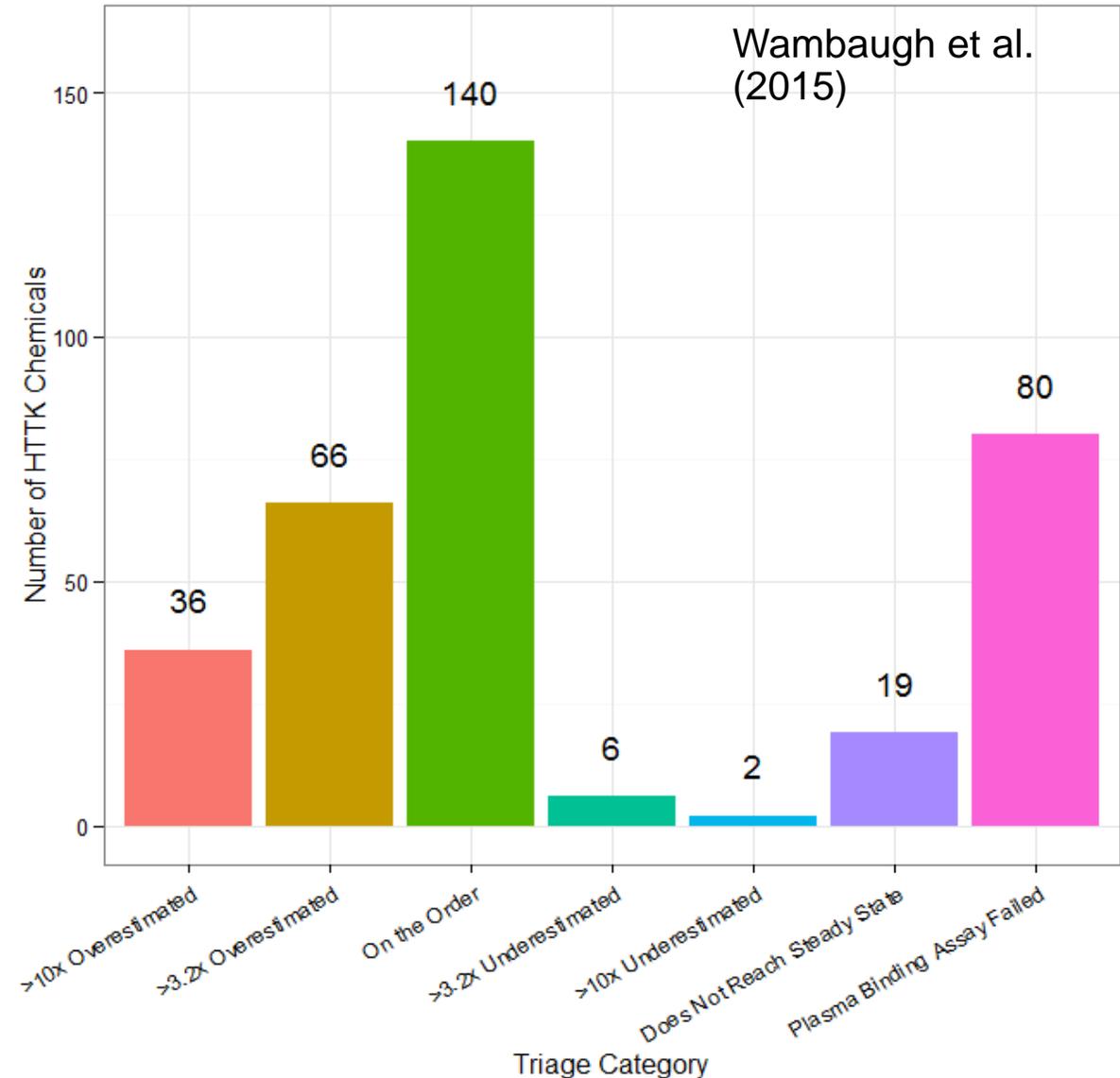


- When we compare the  $C_{ss}$  predicted from *in vitro* HTTK with *in vivo*  $C_{ss}$  values determined from the literature we find limited correlation ( $R^2 \sim 0.34$ )
- The dashed line indicates the identity (perfect predictor) line:
  - Over-predict for 65
  - Under-predict for 22
- The white lines indicate the discrepancy between measured and predicted values (the residual)

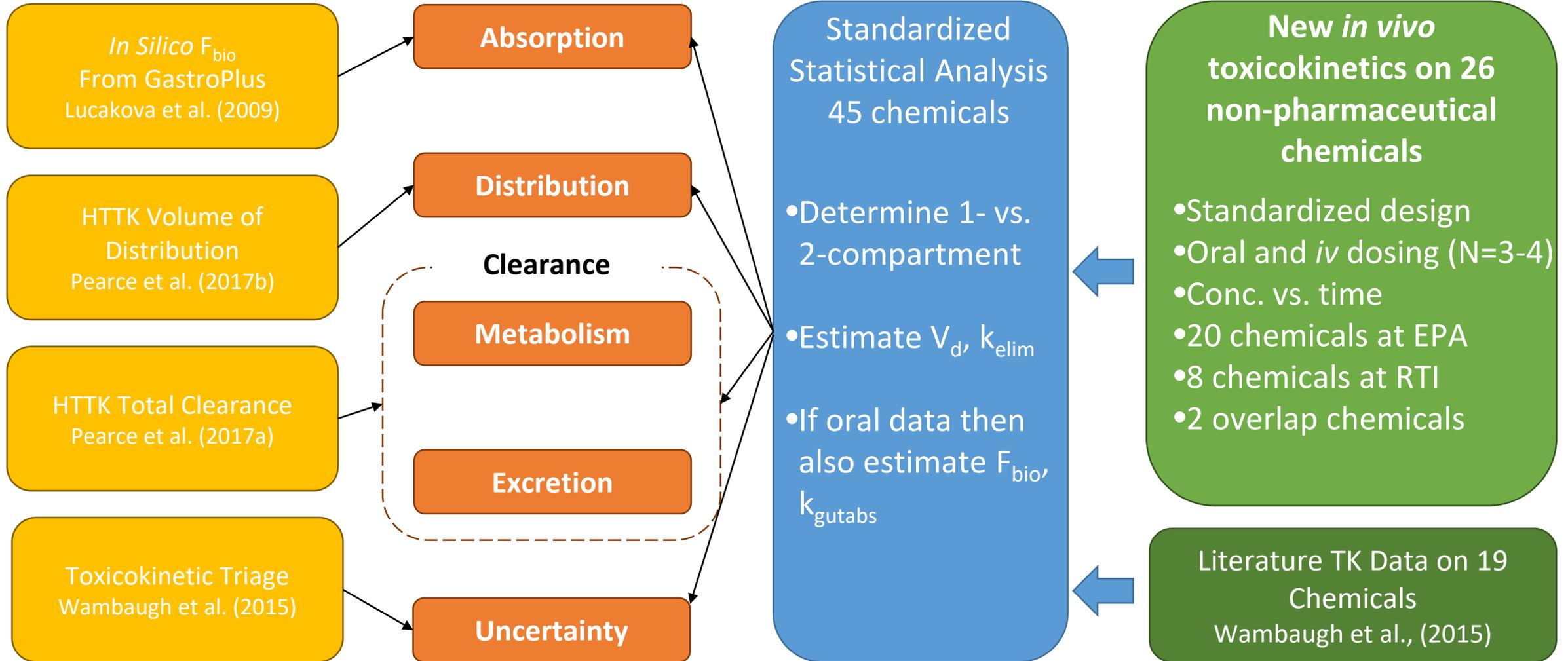


# Toxicokinetic Triage

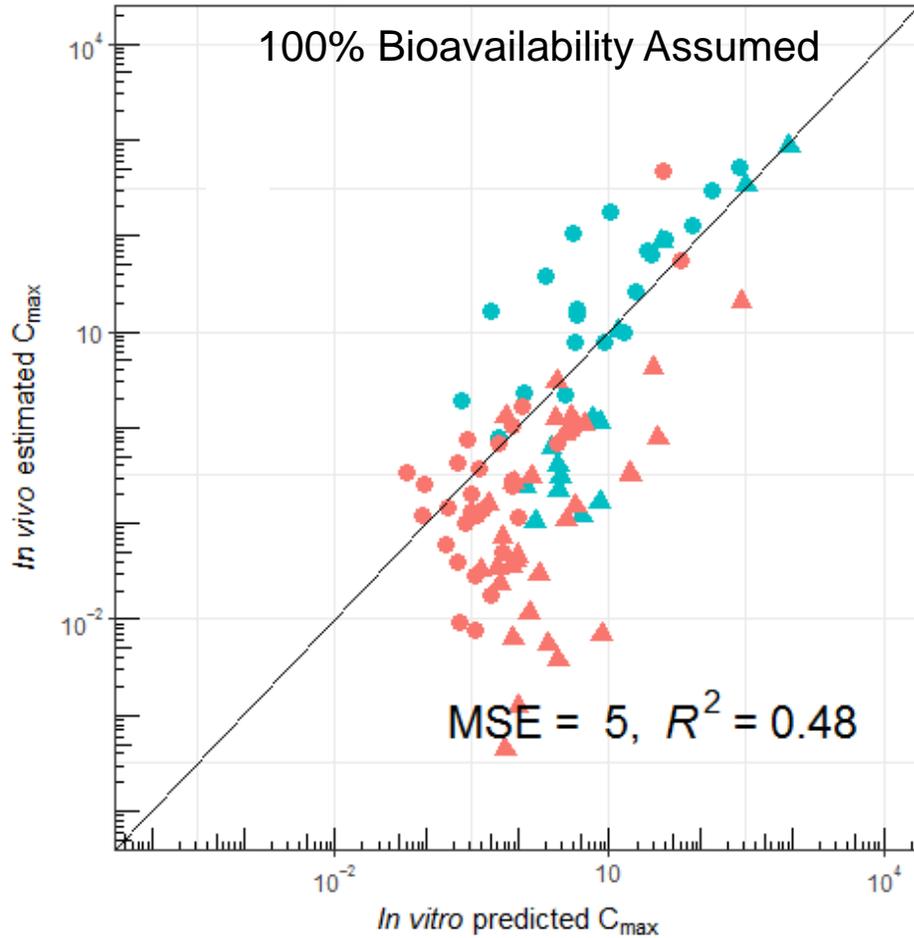
- Through comparison to *in vivo* data, a cross-validated (random forest) predictor of success or failure of HTTK has been constructed
- Add categories for chemicals that do not reach steady-state or for which plasma binding assay fails
- All chemicals can be placed into one of seven confidence categories
- Plurality of chemicals end up in the “on the order” bin (within a factor of 3.2x) which is consistent with Wang (2010)



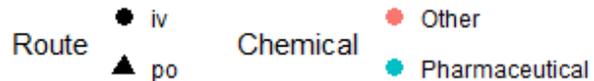
# New Data for Evaluation



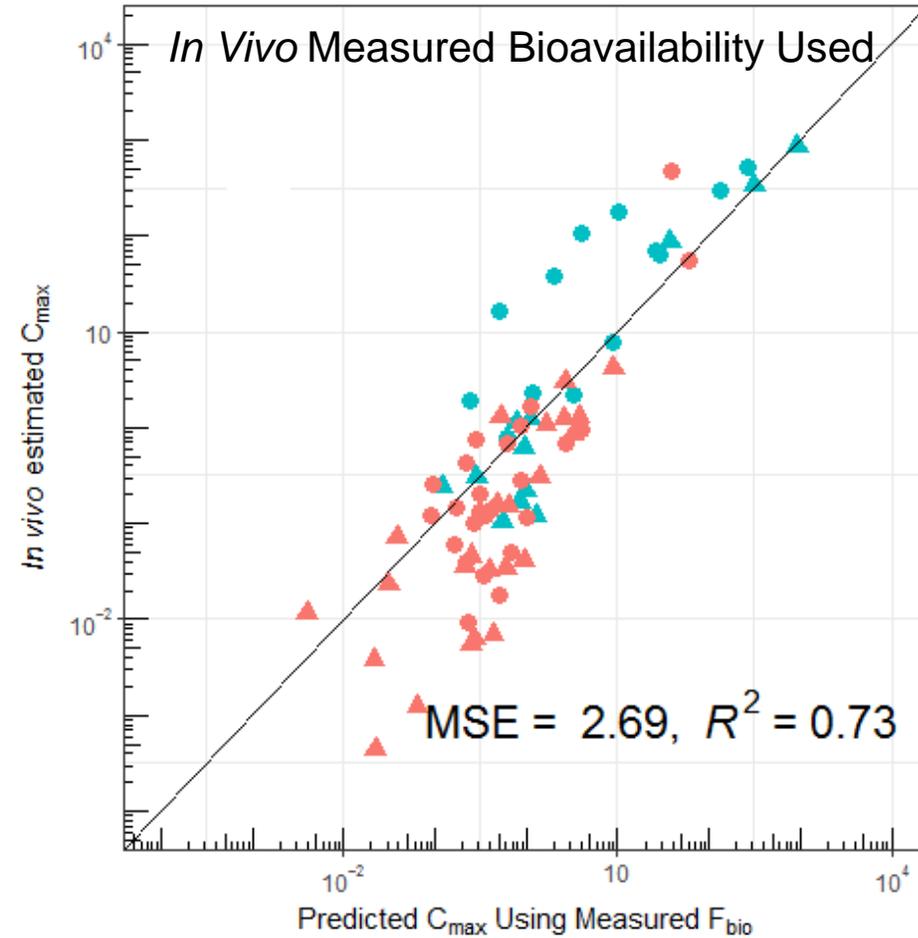
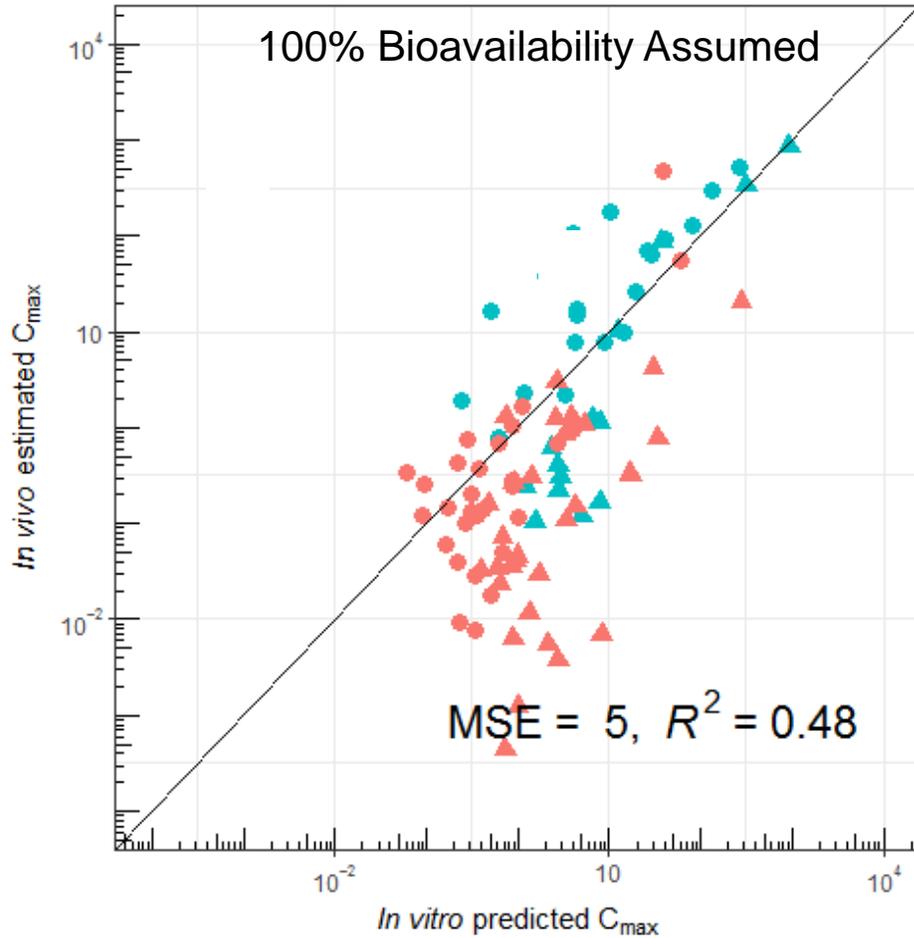
# Impact of Oral Bioavailability



We evaluate HTTK by comparing predictions with observations for as many chemicals as possible



# Impact of Oral Bioavailability

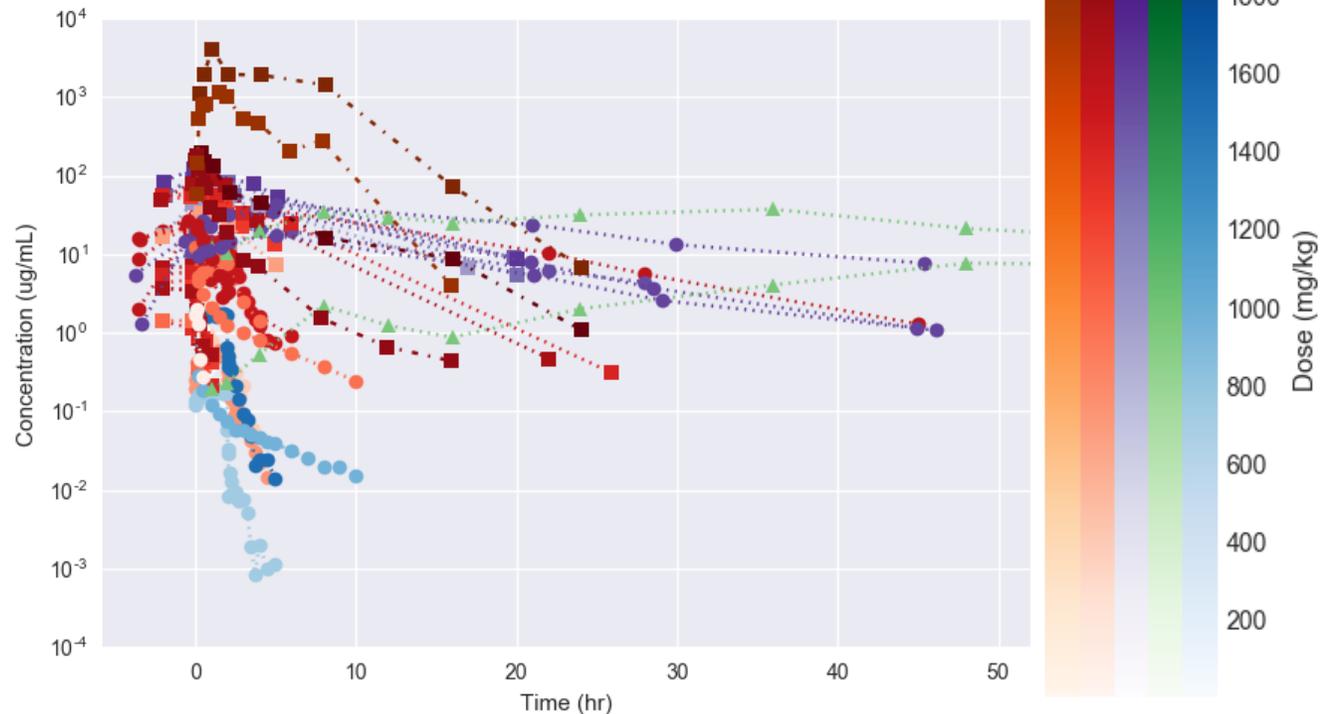


Route ● iv      Chemical ● Other  
           ▲ po                ● Pharmaceutical

Route ● iv      Chemical ● Other  
           ▲ po                ● Pharmaceutical

# In Vivo TK Database

- EPA is developing a public database of concentration vs. time data for building, calibrating, and evaluating TK models
- Curation and development ongoing, but to date includes:
  - 175 analytes (EPA, National Toxicology Program, literature)
  - Routes: Intravenous, dermal, oral, sub-cutaneous, and inhalation exposure
  - Species: dog, frog, human, monkey, mouse, rabbit, rat
  - Media: plasma, as well as adipose, bile, blood, brain, digestive tract, exhaled air, heart, kidney, liver, lung, muscle, pancreas, serum, skin, spleen, testes, thymus, urine
  - Multiple studies per chemical
- Database will be made available through web interface and through the “httk” R package
- Standardized, open source curve fitting software *invivoPKfit* used to calibrate models to all data:
  - <https://github.com/USEPA/CompTox-ExpoCast-invivoPKfit>



# Uncertainty

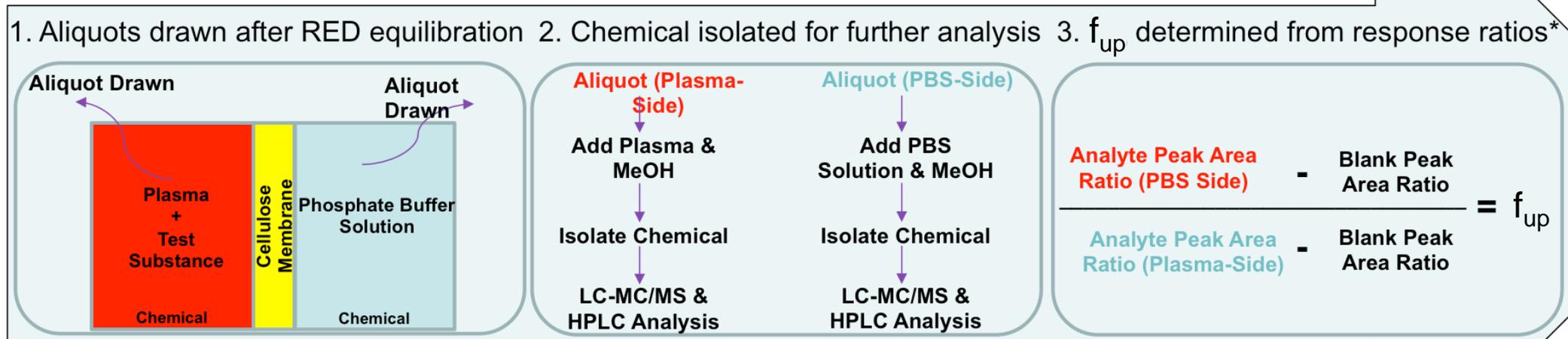
Until I open the box, I don't know what colors I have...

...especially if my five-year-old has been around.



# Analytical Chemistry is an HTTK Bottleneck

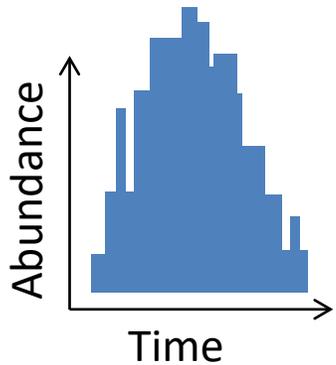
- Need to develop a chemical-specific method for quantitating amount of chemical *in vitro*
  - This is very different from HTS where same readout (e.g., bioluminescence) can be used for most chemicals
- In Wetmore et al. (2012), the rapid equilibrium dialysis (RED) assay (Waters et al. 2008) failed for fraction unbound in plasma ( $f_{up}$ ) 38% of the chemicals.



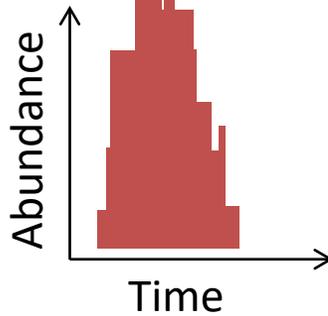
# New HTTK Measurements and Uncertainty Analysis

The HTTK *in vitro* assays need to measure differences in chemical concentration

Internal Standard



Chemical Peak

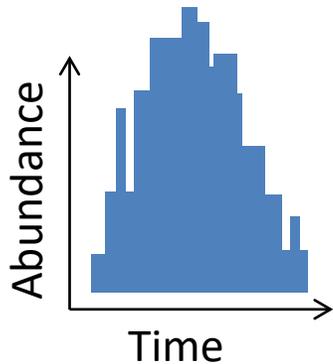


- Area of the internal standard (ITSD) at a known, fixed concentration fluctuates with time
- Find a peak that corresponds to chemical of interest, and then follow the ratio  $R$  of the chemical peak to the ITSD

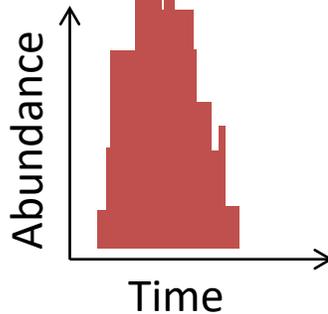
# New HTTK Measurements and Uncertainty Analysis

The HTTK *in vitro* assays need to measure differences in chemical concentration

Internal Standard

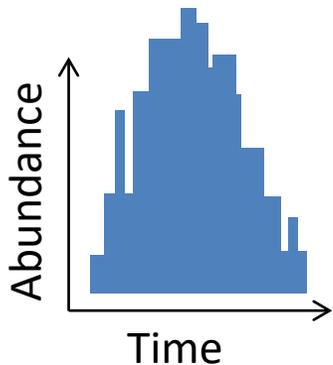


Chemical Peak

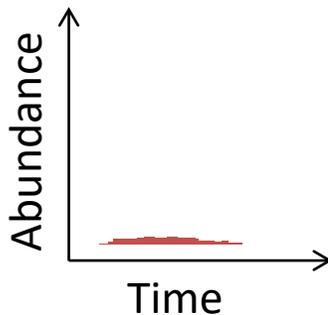


- Area of the internal standard (ITSD) at a known, fixed concentration fluctuates with time
- Find a peak that corresponds to chemical of interest, and then follow the ratio  $R$  of the chemical peak to the ITSD
- For new measurements HTTK (>200 compounds to data) performed by Cyprotex, we have modified RED protocol to use a titration of plasma protein (10%, 30%, 100%) of physiological concentration

Internal Standard

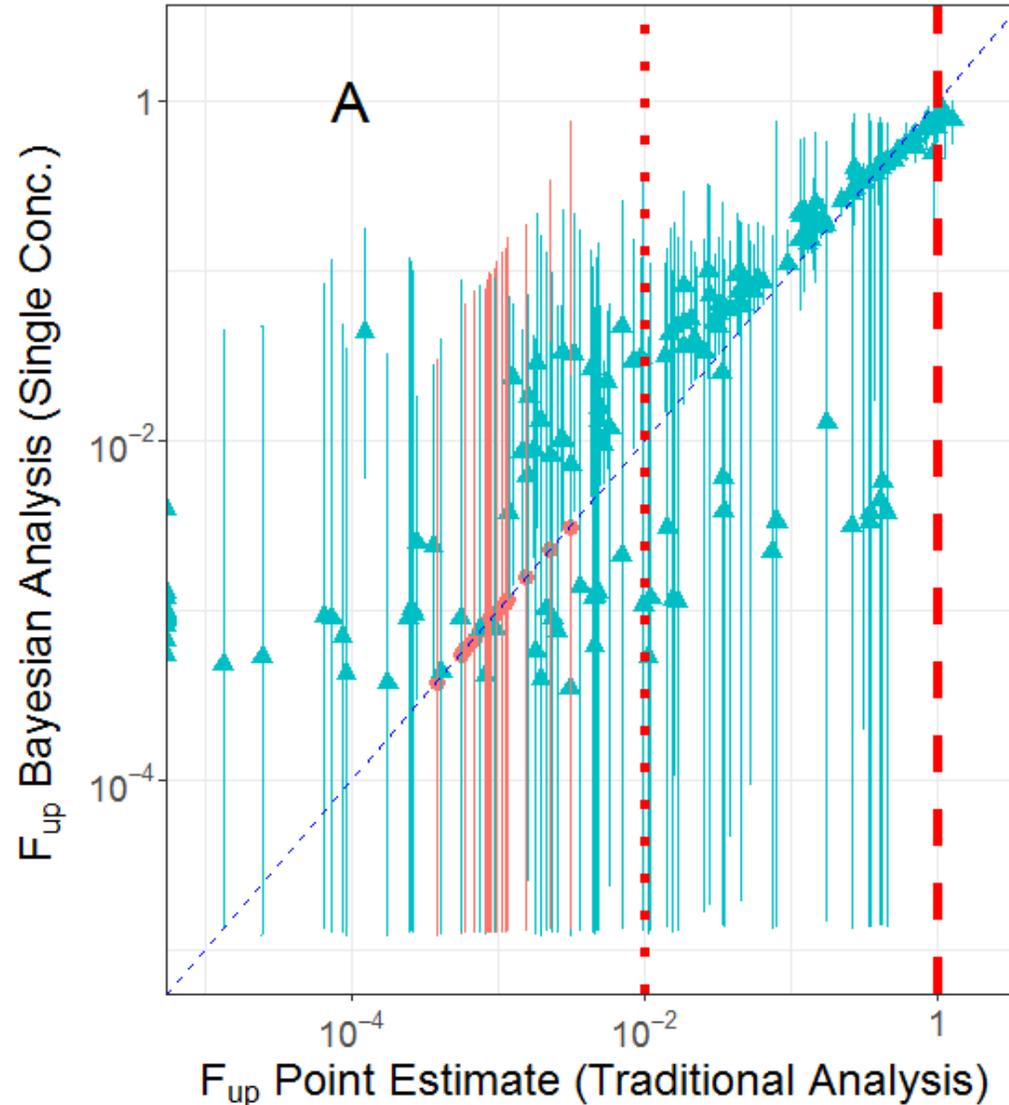


Chemical Peak



- Keeps chemical concentration in the same range
- Analyzed data in Bayesian framework that included a model for analytical chemistry
- Bayesian approach gives a credible interval (range of values that would be consistent with the data) – quantitative uncertainty

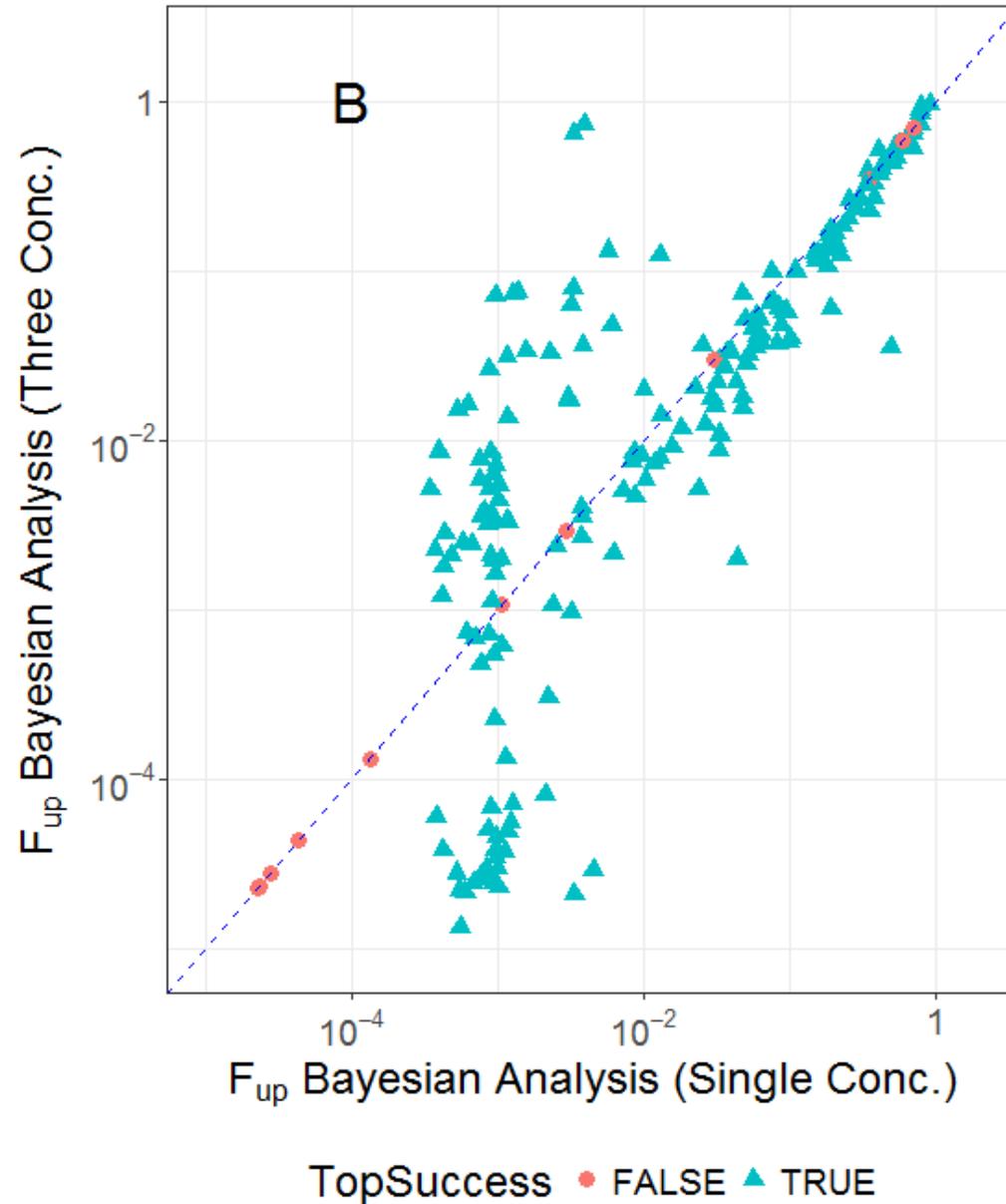
# Results of Bayesian Analysis for PPB



- Previous method allowed values above and below zero, those observations now increase measurement error estimate
- Medians from Bayesian analysis correlate with point estimates from previous method
- Larger values track with each other better (Wetmore *et al.* 2012) average LOD was ~1%)

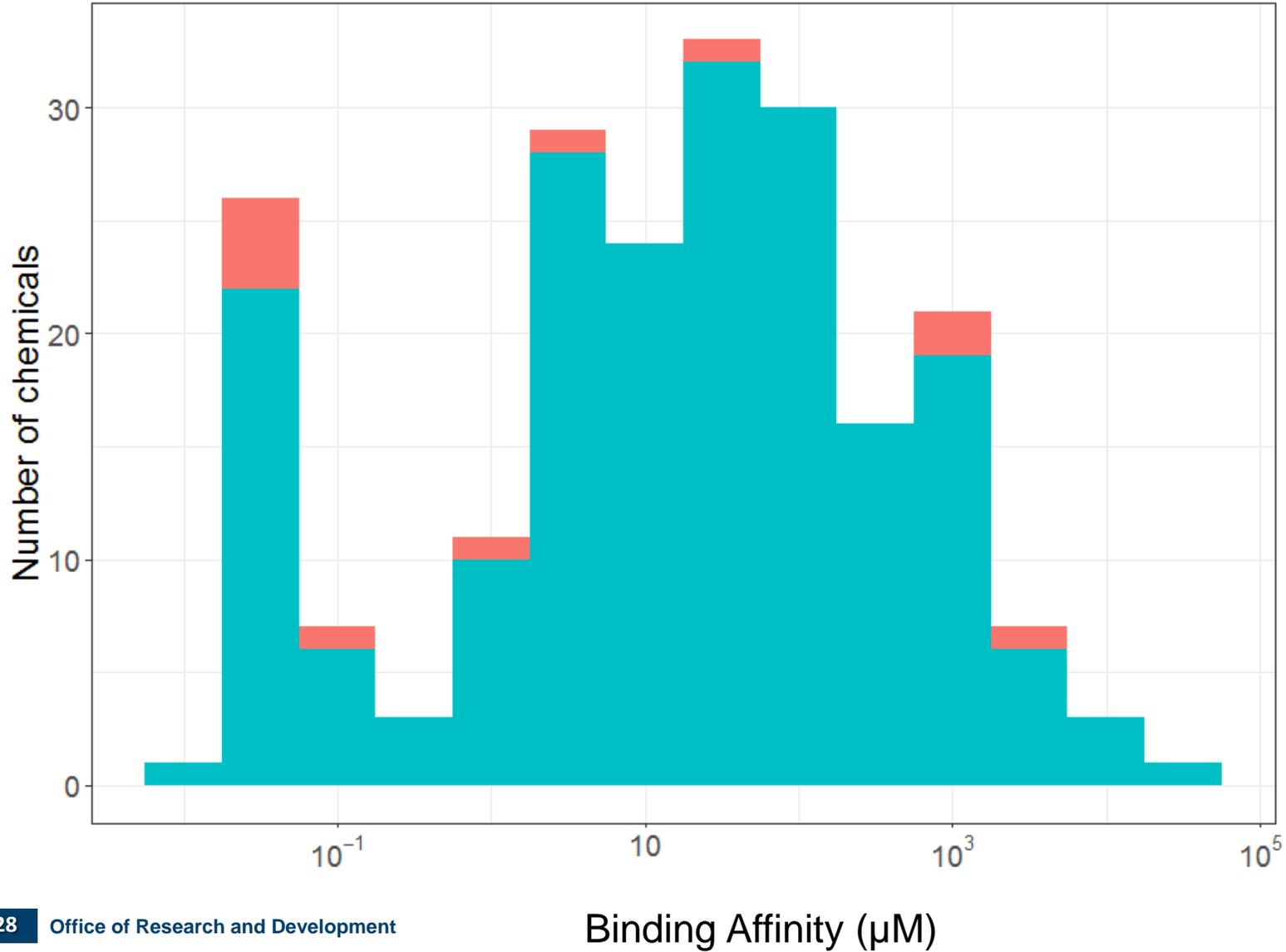
Point Estimate > 0 ● FALSE ▲ TRUE

## Results for Plasma Protein Titration



- Analysis of rapid equilibrium dialysis performed at 100%, 30%, and 10% of physiologic protein concentration
- Seven chemicals that had no measurement at 100% concentration now have a value
- Generally correlate, especially for higher  $F_{up}$ 's

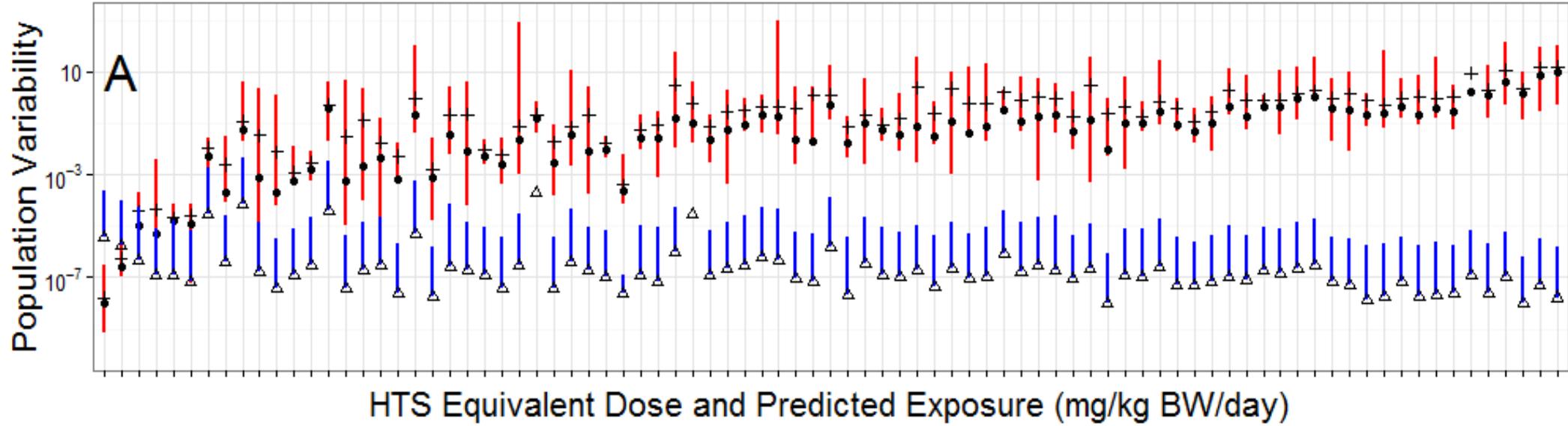
# Estimate of Protein Binding Affinity



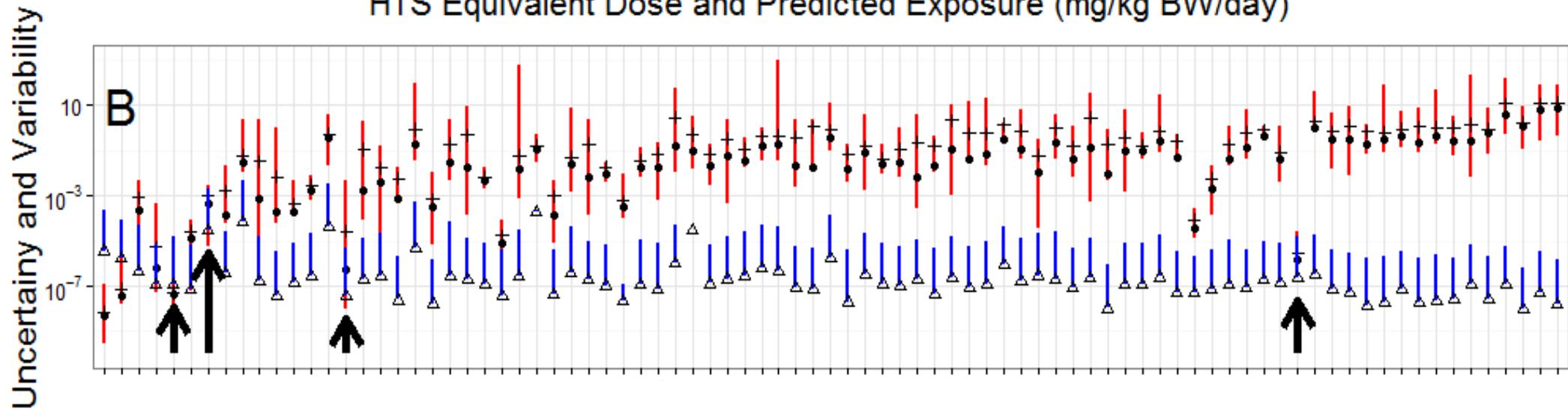
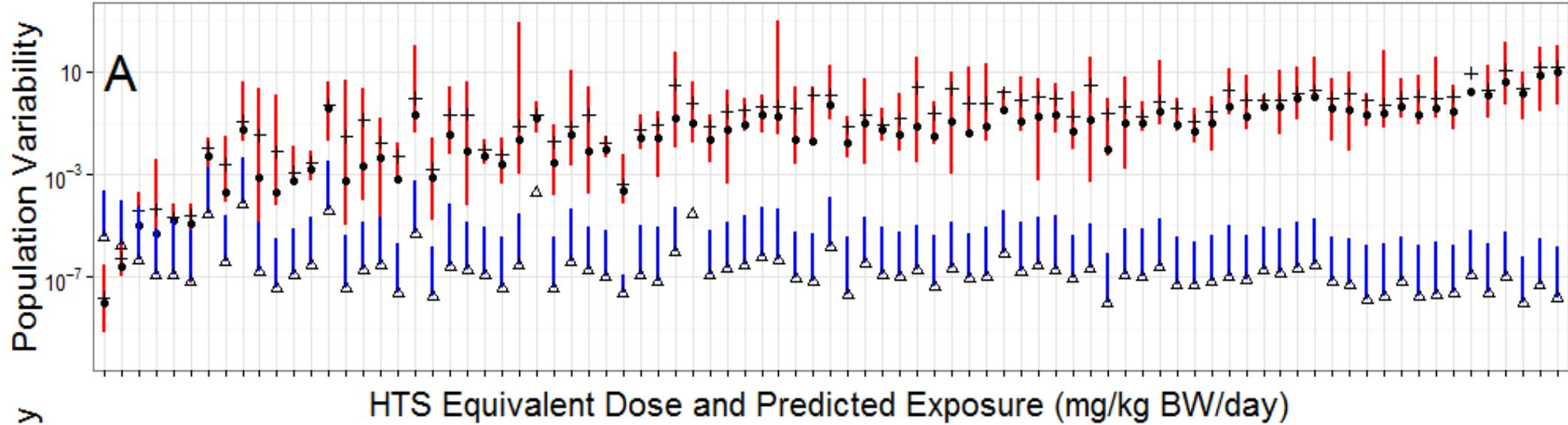
A major benefit of three protein titration protocol is that you get an estimate of binding affinity for each chemical (some are very uncertain)

Measurable at Top Conc. ■ FALSE ■ TRUE

# Relative Contribution of Uncertainty and Variability



# Relative Contribution of Uncertainty and Variability



Four chemicals where uncertainty in HTTK data changes ratio of activity to exposure enough to cause overlap

# Conclusions

- HTTK allows dosimetric adjustment of high-throughput screening (HTS) data across thousands of chemicals.
- Assessments of the impact of uncertainty and variability on these TK values and subsequent predictions are needed to guide data interpretation and provide overall confidence in the new approach methodologies
- New, chemical-specific *in vitro* experiments have been conducted by Cyprotex, using a revised protocol for measuring protein binding
- Bayesian methods were developed to provide chemical-specific uncertainty estimates for two *in vitro* TK parameters: plasma protein binding ( $f_{up}$ ) and intrinsic hepatic clearance ( $Cl_{int}$ ),
- Overall, variability contributed more significantly to  $C_{ss}$  estimations of the 95<sup>th</sup> percentile
- All EPA HTTK data and models are made publically available upon publication through the R “httk” package (Pearce et al., 2017)

# Acknowledgements



Derek Angus      Chris Strock  
Roger Dinallo      Maria Bacolod

## Rapid Exposure and Dosimetry (RED) Project

## Collaborators

**Arnot Research and Consulting**  
Jon Arnot  
Johnny Westgate  
**Battelle Memorial Institute**  
Anne Louise Sumner  
Anne Gregg  
**Chemical Computing Group**  
Rocky Goldsmith  
**National Institute for Environmental Health Sciences (NIEHS) National Toxicology Program**  
Mike Devito  
Steve Ferguson  
Nisha Sipes  
**Netherlands Organisation for Applied Scientific Research (TNO)**  
Sieto Bosgra  
**Research Triangle Institute**  
Timothy Fennell  
**ScitoVation**  
Harvey Clewell  
Kamel Mansouri  
Chantel Nicolas  
**Silent Spring Institute**  
Robin Dodson  
**Southwest Research Institute**  
Alice Yau  
Kristin Favela  
**Summit Toxicology**  
Lesa Aylward  
**Tox Strategies**  
Caroline Ring  
**University of California, Davis**  
Deborah Bennett  
Hyeong-Moo Shin  
**University of Michigan**  
Olivier Jolliet  
**University of North Carolina, Chapel Hill**  
Alex Tropsha

### NCCT

Chris Grulke  
Greg Honda\*  
Richard Judson  
Andrew McEachran\*  
Robert Pearce\*  
Ann Richard  
Risa Sayre\*  
Woody Setzer  
Rusty Thomas  
**John Wambaugh**  
Antony Williams

### NRMRL

Yirui Liang\*  
Xiaoyu Liu  
**NHEERL**  
Linda Adams  
Christopher Ecklund  
Marina Evans  
Mike Hughes  
Jane Ellen Simmons

**\*Trainees**

### NERL

Cody Addington\*  
Craig Barber  
Namdi Brandon\*  
Peter Egeghy  
Hongtai Huang\*  
Brandall Ingle\*  
**Kristin Isaacs**  
Ashley Jackson\*  
Charles Lowe\*  
Dawn Mills\*  
Seth Newton

Katherine Phillips  
Paul Price  
Jeanette Reyes\*  
Randolph Singh\*  
Jon Sobus  
John Streicher\*  
Mark Strynar  
Mike Tornero-Velez  
Elin Ulrich  
Dan Vallero  
Barbara Wetmore

### Lead CSS Matrix Interfaces:

John Kenneke (NERL)  
John Cowden (NCCT)

# References

- Breyer, Stephen. Breaking the vicious circle: Toward effective risk regulation. Harvard University Press, 2009
- Bosgra, Sieto, et al. "An improved model to predict physiologically based model parameters and their inter-individual variability from anthropometry." *Critical reviews in toxicology* 42.9 (2012): 751-767.
- Collins FS, Gray GM, Bucher JR. Transforming environmental health protection. *Science*. 2008;319:906–907. [PMC free article] [PubMed]
- Dix DJ, Houck KA, Martin M, Richard AM, Setzer RW, Kavlock RJ. The ToxCast program for prioritizing toxicity testing of environmental chemicals. *Toxicol Sci*. 2007;95:5–12
- Jamei, et al. "The Simcyp® population-based ADME simulator." *Expert opinion on drug metabolism & toxicology* 2009b;5:211-223
- McNally, et al., "PopGen: a virtual human population generator." *Toxicology* \*2014)
- National Research Council. (1983). Risk Assessment in the Federal Government: Managing the Process Working Papers. National Academies Press.
- Park, Youngja H., et al. "High-performance metabolic profiling of plasma from seven mammalian species for simultaneous environmental chemical surveillance and bioeffect monitoring." *Toxicology* 295.1 (2012): 47-55.
- Pearce, Robert, et al. "httk: R Package for High-Throughput Toxicokinetics." *Journal of Statistical Software*, (2017)
- Price, Paul S., et al. "Modeling interindividual variation in physiological factors used in PBPK models of humans." *Critical Reviews in Toxicology* 33.5 (2003): 469-503.
- Ring, Caroline L., et al. "Identifying populations sensitive to environmental chemicals by simulating toxicokinetic variability." *Environment International* 106 (2017): 105-118.
- Rotroff, Daniel M., et al. "Incorporating human dosimetry and exposure into high-throughput in vitro toxicity screening." *Toxicological Sciences* 117.2 (2010): 348-358.
- Strobe, Cory L., et al. "High-throughput in-silico prediction of ionization equilibria for pharmacokinetic modeling." *Science of science & technology* 47.15 (2013): 8479-The Total Environment 615 (2018): 150-160.
- Wambaugh, John F., et al. "High-throughput models for exposure-based chemical prioritization in the ExpoCast project." *Environmental* 848.
- Wambaugh, John F., et al. "High Throughput Heuristics for Prioritizing Human Exposure to Environmental Chemicals." *Environmental science & technology* (2014).
- Wetmore, Barbara A., et al. "Integration of dosimetry, exposure and high-throughput screening data in chemical toxicity assessment." *Tox. Sciences* (2012)
- Wetmore, Barbara A., et al. "Incorporating population variability and susceptible subpopulations into dosimetry for high-throughput toxicity testing." *Toxicological Sciences* 142.1 (2014): 210-224.
- Wetmore, Barbara A., et al. "Incorporating high-throughput exposure predictions with dosimetry-adjusted in vitro bioactivity to inform chemical toxicity testing." *Toxicological Sciences* 148.1 (2015): 121-136.
- Waters, Nigel J., et al. "Validation of a rapid equilibrium dialysis approach for the measurement of plasma protein binding." *Journal of pharmaceutical sciences* 97.10 (2008): 4586-4595