

1 Background and Objectives

Background

- Current initiatives in environmental toxicology include integrating toxicity testing data from *in vitro* and alternative methods into risk assessment practices.
- Previously, *in vitro* point-of-departure (PODs) estimates suitable for use in screening-level risk assessments have been based on panels of *in vitro* assays that report a limited number of endpoints and that may not provide comprehensive coverage of all biological pathways.
- Increasing efficiency and declining cost of generating whole transcriptome profiles has made high-throughput transcriptomics (HTTr) a practical option for determining *in vitro* bioactivity thresholds and increasing coverage of biological space in chemical screening.

Objectives

- Develop a laboratory workflow for concentration-response screening of environmental chemicals using a targeted RNA-Seq human whole transcriptome assay (TempO-Seq).
- Perform concentration-response screening of a large number of chemicals (n=2,200) from the USEPA Toxicity Forecaster (ToxCast) compound library in MCF7 cells at a single time point (6 hours post-exposure).
- Evaluate the performance of the targeted RNA-Seq assay across plates and screening blocks using reference samples (i.e. commercially-available purified RNAs & laboratory generated bulk cell lysates) and treatment with reference chemicals.
- Establish a data processing pipeline for data normalization, differentially expressed gene (DEG) identification and determination of *in vitro* PODs.

2 Laboratory Workflow

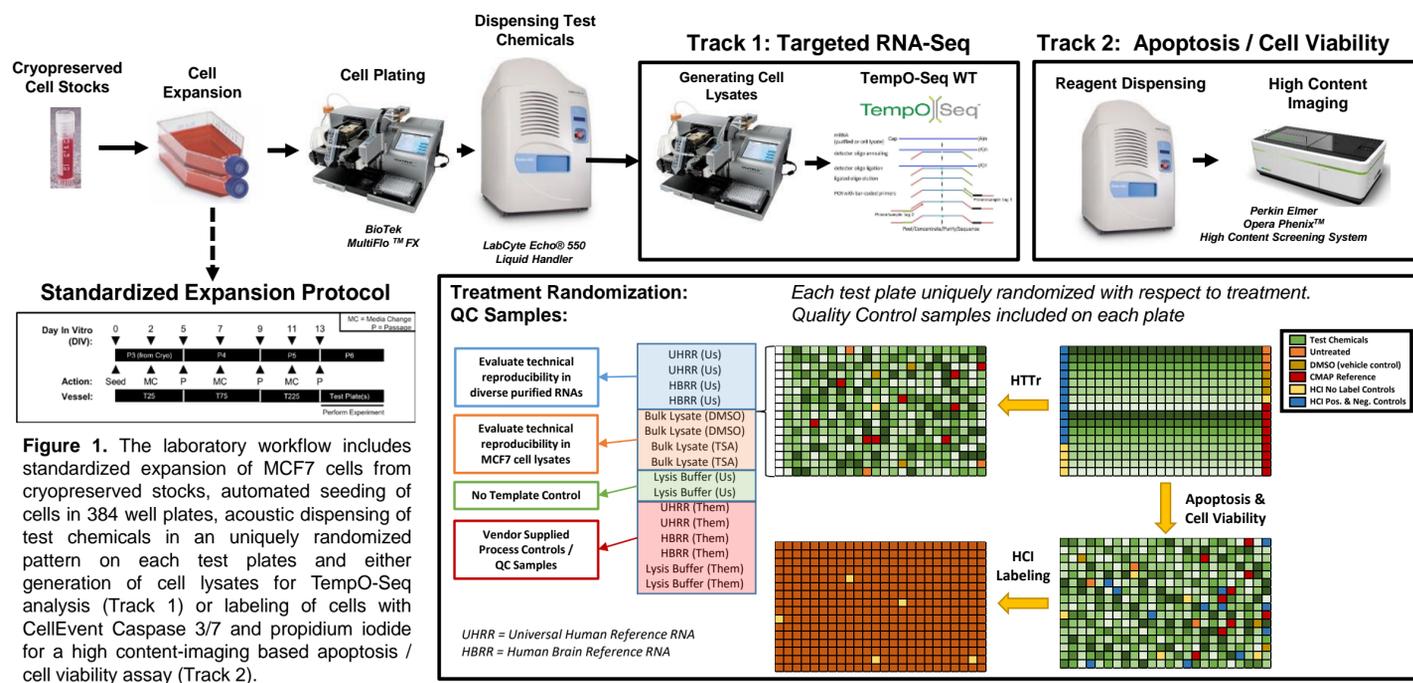
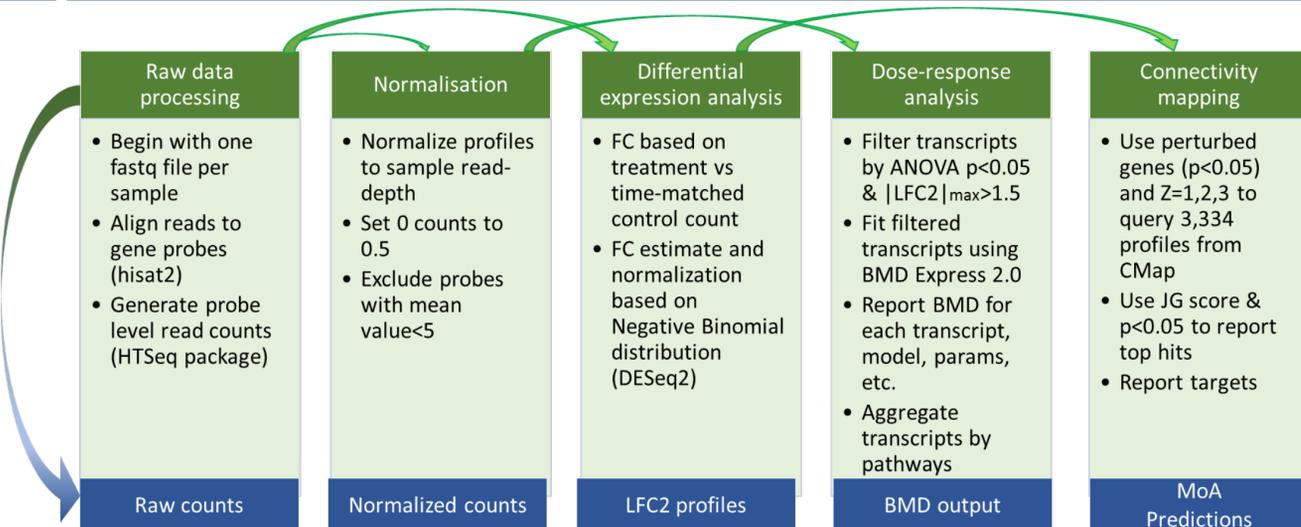


Figure 1. The laboratory workflow includes standardized expansion of MCF7 cells from cryopreserved stocks, automated seeding of cells in 384 well plates, acoustic dispensing of test chemicals in a uniquely randomized pattern on each test plates and either generation of cell lysates for TempO-Seq analysis (Track 1) or labeling of cells with CellEvent Caspase 3/7 and propidium iodide for a high content-imaging based apoptosis / cell viability assay (Track 2).

3 HTTr Data Analysis Pipeline



5 POD Determination

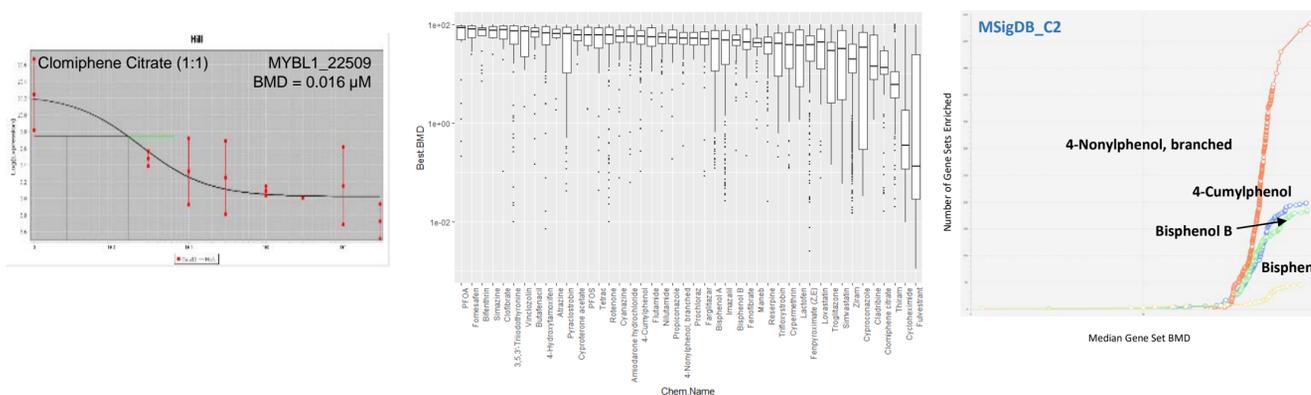


Figure 3. Benchmark Dose Modeling with BMDExpress2.0. The BMDExpress2.0 transcriptomic concentration-response modeling tool was used to fit concentration-response curves for the respective test chemicals to each probe in the TempO-Seq whole transcriptome assay. (A) An example concentration-response curve for an estrogen-responsive gene (MYBL1) following treatment of MCF7 cells with clomiphene citrate, an estrogen receptor antagonist. Benchmark doses were defined as concentrations which produce a >10% change in a response. (B) The distribution of probe level BMDs for 44 example chemicals. Heterogeneity in the potency and distribution of probe-level BMDs were observed across chemicals. (C) Probes with significant concentration-responses were input into a gene set enrichment analysis using the Molecular Signatures C2 curated gene set database. The accumulation plot illustrates the number of enriched gene sets (y-axis) and the respective probe-level median BMD within each gene set (x-axis) for a group of estrogen receptor agonists. Difference in potency were observed at the pathway level.

4 HTTr Assay Performance

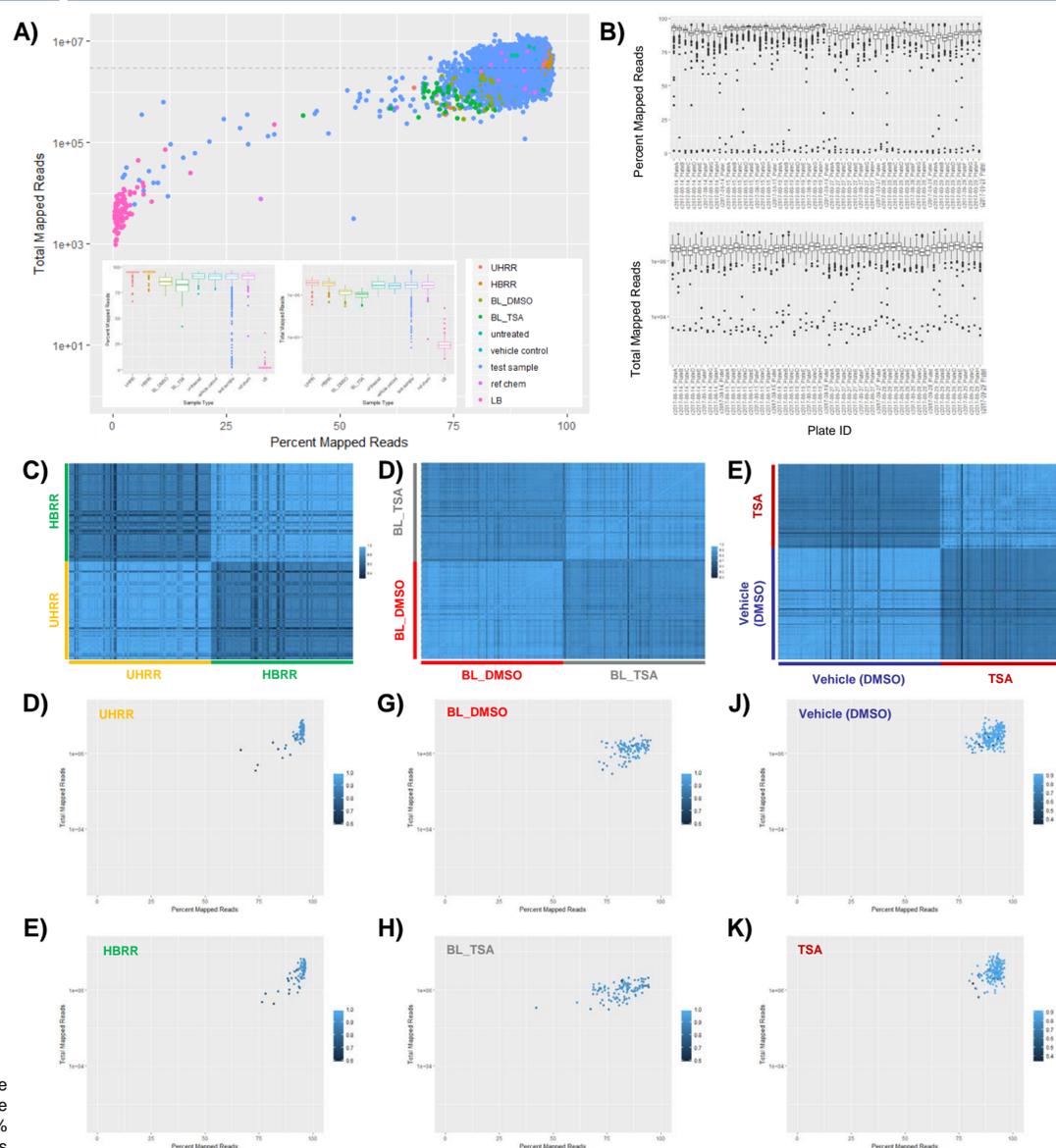


Figure 2. Summary of results from n=54 TempO-Seq assay plates. A majority of samples had total mapped reads greater than 1×10^6 (mean $\sim 3 \times 10^6$) and percent mapped reads > 75% (mean $\sim 90\%$) (A). The distribution of total mapped reads and percent mapped reads were consistent across assay plates (B). Correlations of like QC samples across plates was high (>0.9) in most cases. Paired QC samples (i.e. UHRR vs. HBRR, BL_DMSO vs BL_TSA, DMSO vs TSA) showed distinct differences in the strength of correlations (C-D). Poor correlations with like samples observed in a minority of QC samples had no apparent relationship with total mapped reads or percent mapped reads (D-K).