

# High-Throughput Transcriptomics (HTTr) in *In Vitro* Chemical Screening, A Sensitive Tool for Benchmark Dose Assessment

Joshua A. Harrill, USEPA National Center for Computational Toxicology (NCCT)



American Society of Pharmacology and Experimental Therapeutics  
Annual Meeting, San Diego, CA  
April 24<sup>th</sup>, 2018

# Disclaimer

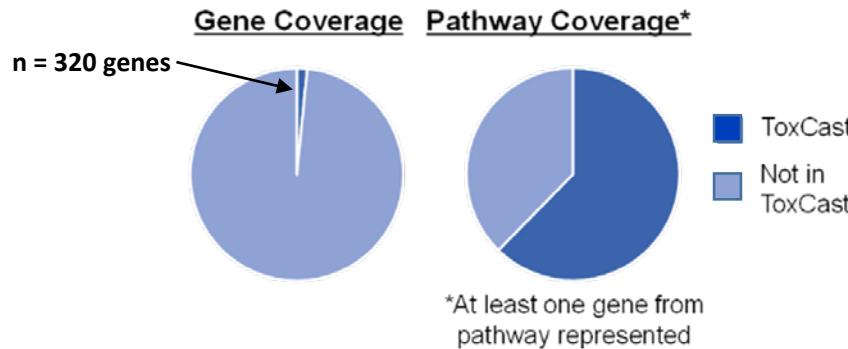
*The views expressed in this [article/presentation/poster] are those of the author(s) and do not necessarily represent the views or policies of the U.S. Environmental Protection Agency, nor does mention of trade names or products represent endorsement for use.*

# Outline

- **Background**
  - Why high-throughput transcriptomics (HTTr)?
  - TempO-Seq Technology Overveiw
- **Experimental Design & Workflows**
  - Screening format
  - Treatment Randomization
  - Quality Control Samples
- **Technical Reproducibility**
- **Applications**
  - Concentration-Response Modeling
  - Bioactivity Thresholds
  - Comparison of Structurally Related Chemicals
  - Gene Set Analysis
  - Mechanism of Action (MoA) Prediction

# Background

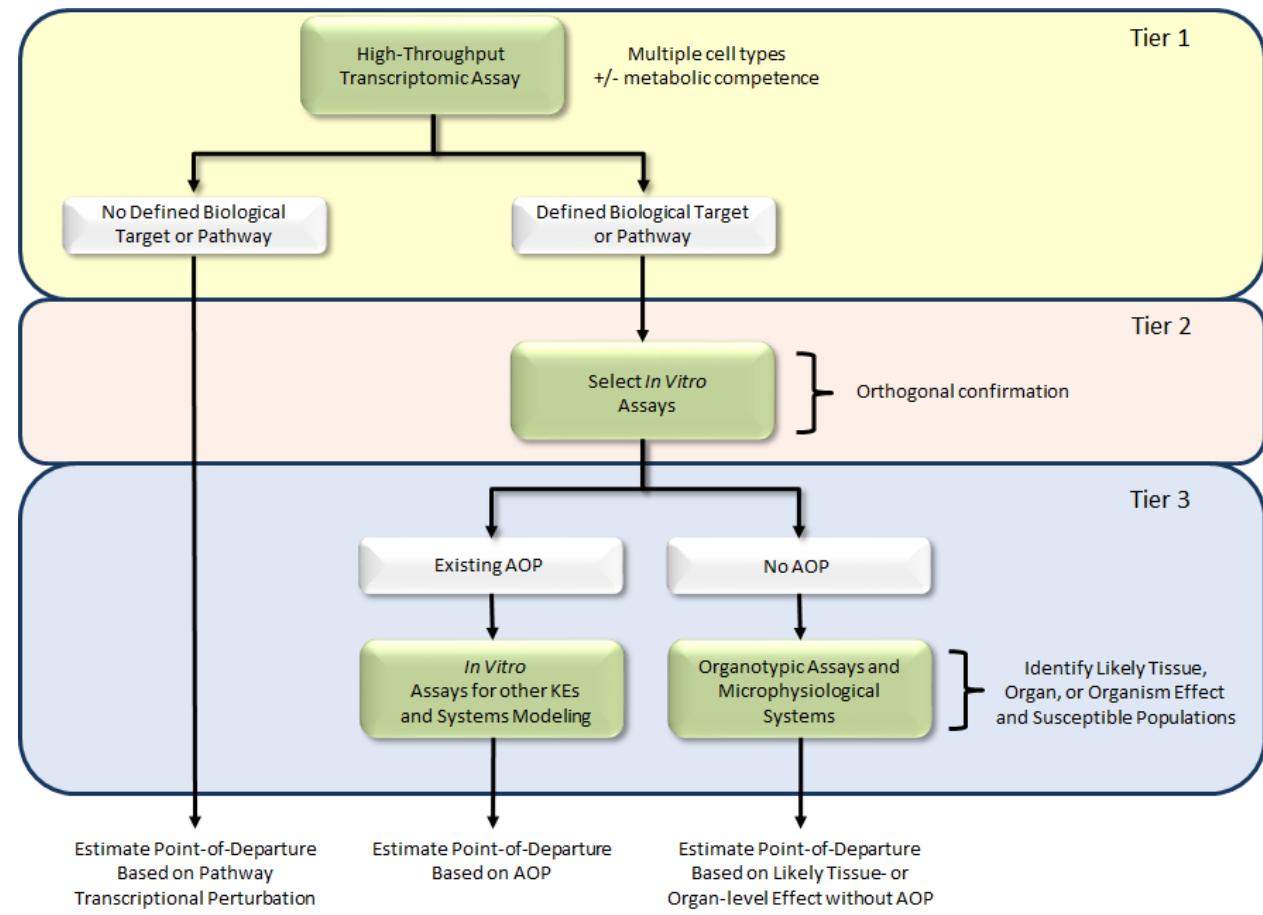
- ToxCast assays cover many genes and pathways, but do not provide complete coverage of biological space.



- USEPA Strategic Vision and Operational Roadmap:**

- Tier 1 strategy must cast the broadest net possible for capturing hazards associated with chemical exposure.
- Global gene expression provides a robust and comprehensive evaluation of chemically induced changes in biological processes.
- Increasing efficiency and declining cost of generating whole transcriptome profiles has made high-throughput transcriptomics (HTTr) a practical option for determining bioactivity thresholds in *in vitro* models.

## A strategic vision and operational road map for computational toxicology at the U.S. Environmental Protection Agency

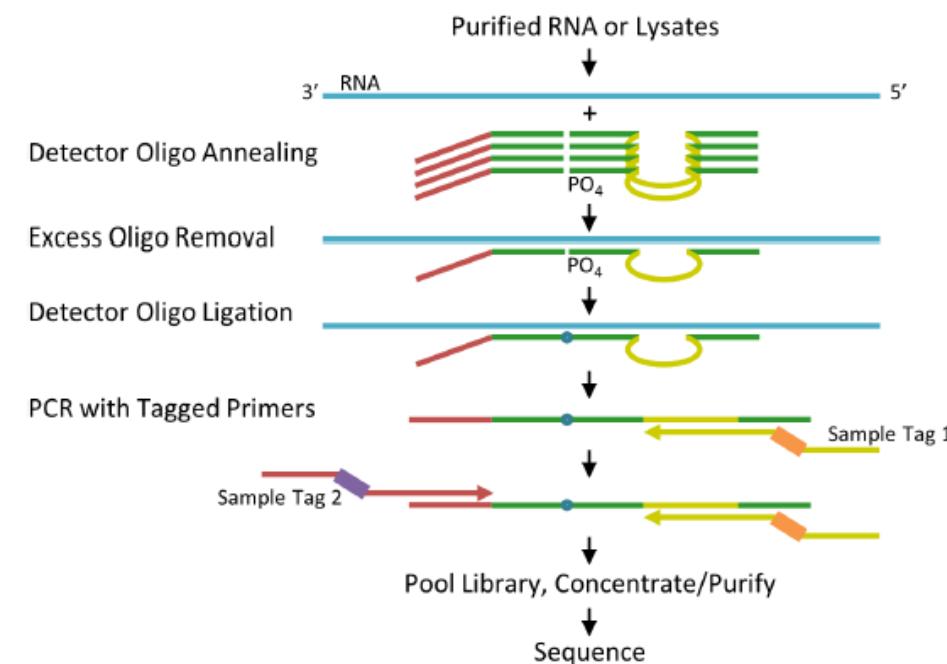


# Background

## Technology

- The **TempO-Seq** human whole transcriptome assay measures the expression of greater than 20,000 transcripts.
- Requires only picogram amounts of total RNA per sample.
- Compatible with purified RNA samples or **cell lysates**.
- Transcripts in cell lysates generated in 384-well format are barcoded according to well position and combined in a single library for sequencing using industry standard instrumentation.
- Scalable, targeted assay:
  - 1) specifically measures transcripts of interest
  - 2) ~50-bp reads for all genes
  - 3) requires less flow cell capacity than RNA-Seq
- Per sample fastq files are generated and aligned to BioSpyder sequence manifest to generate integer count tables.

## TempO-Seq Assay Illustration



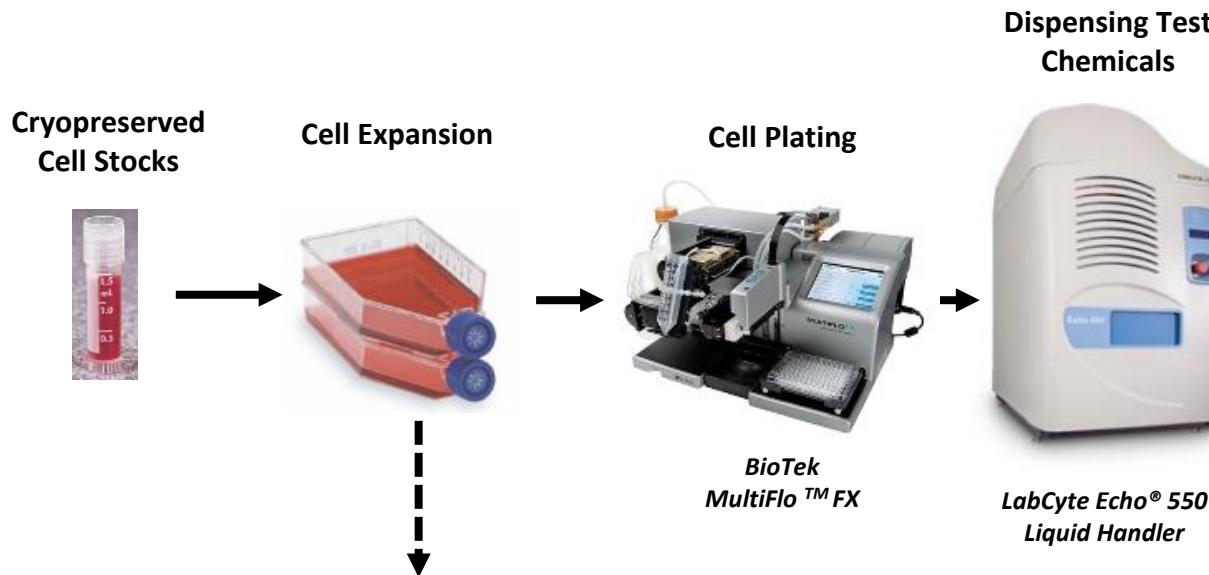
# HTTr MCF Screen: Experimental Design

Parameter	Multiplier	Notes
Cell Type(s)	1	MCF7
Culture Condition	1	DMEM + 10% HI-FBS <sup>a</sup>
Chemicals	2,112	ToxCast ph1, ph2 Nominated chemicals from e1k / ph3
Time Points:	1	6 hours
Assay Formats:	2	TempO-Seq HCl Cell Viability & Apoptosis
Concentrations:	8	3.5 log <sub>10</sub> units; semi log <sub>10</sub> spacing
Biological Replicates:	3	--

- **Total number of samples:** 54,432
- **Total number of endpoint readouts:** 1.15x10<sup>9</sup>
- **Total size of fastq files:** 32.5 to 54.4 TB

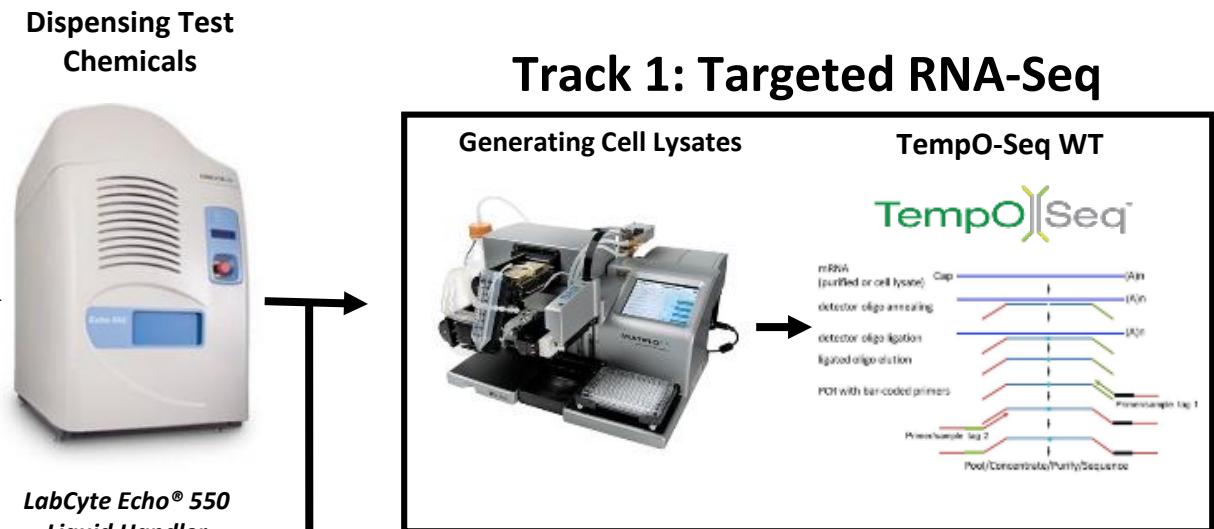
<sup>a</sup> MCF7 cells cultured in DMEM + 10% HI-FBS was selected as the test system to facilitate comparability to the Broad Institute Connectivity Map (CMAP) database (<http://portals.broadinstitute.org/cmap/>).

# Experimental Workflow

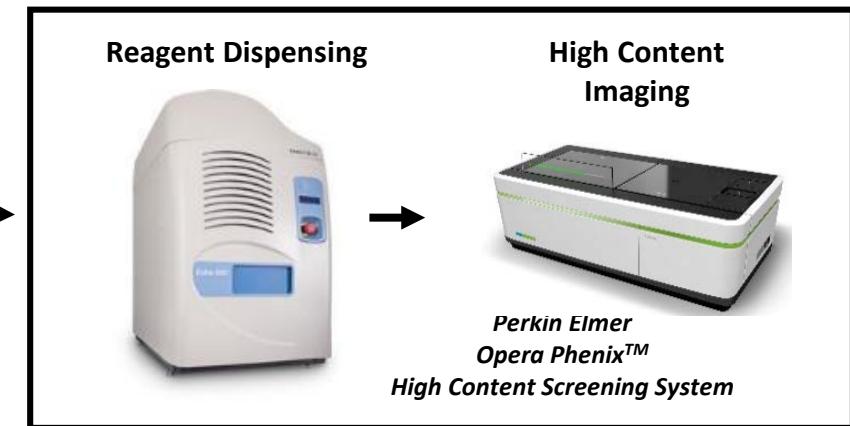


## Standardized Expansion Protocol

Day In Vitro (DIV):	0	2	5	7	9	11	13	
Action:	Seed	MC	P	MC	P	MC	P	MC = Media Change P = Passage
Vessel:	T25		T75		T225		Test Plate(s)	Perform Experiment

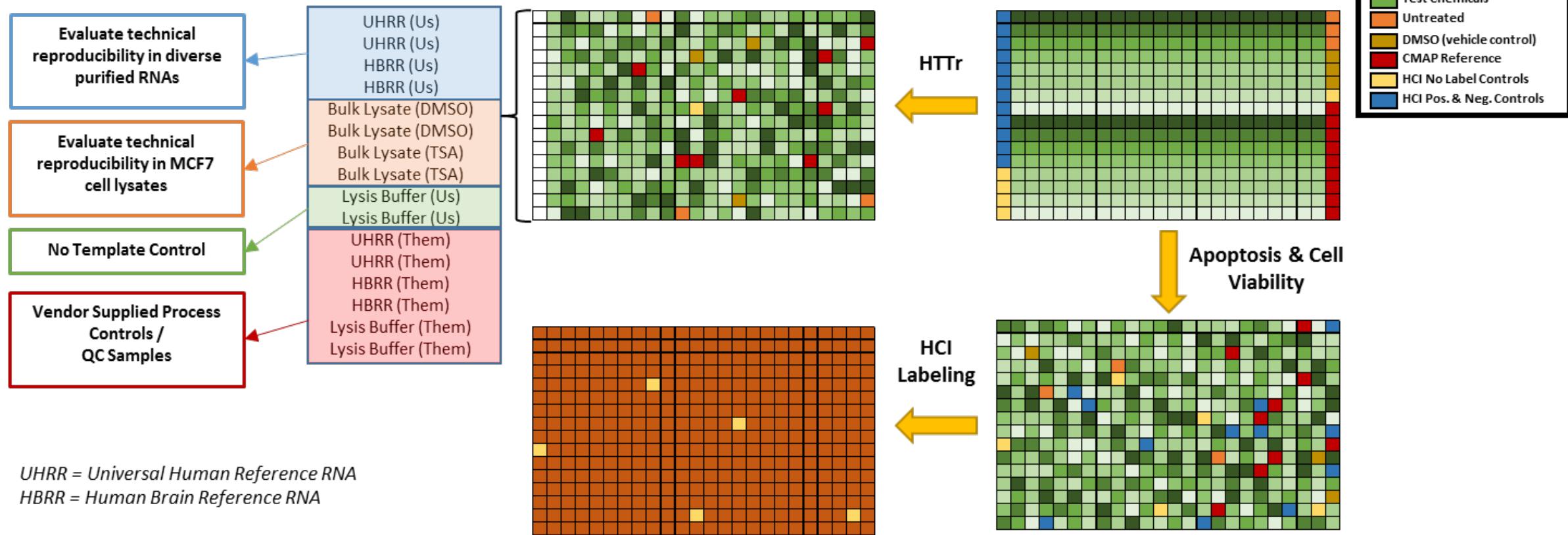


## Track 2: Apoptosis / Cell Viability

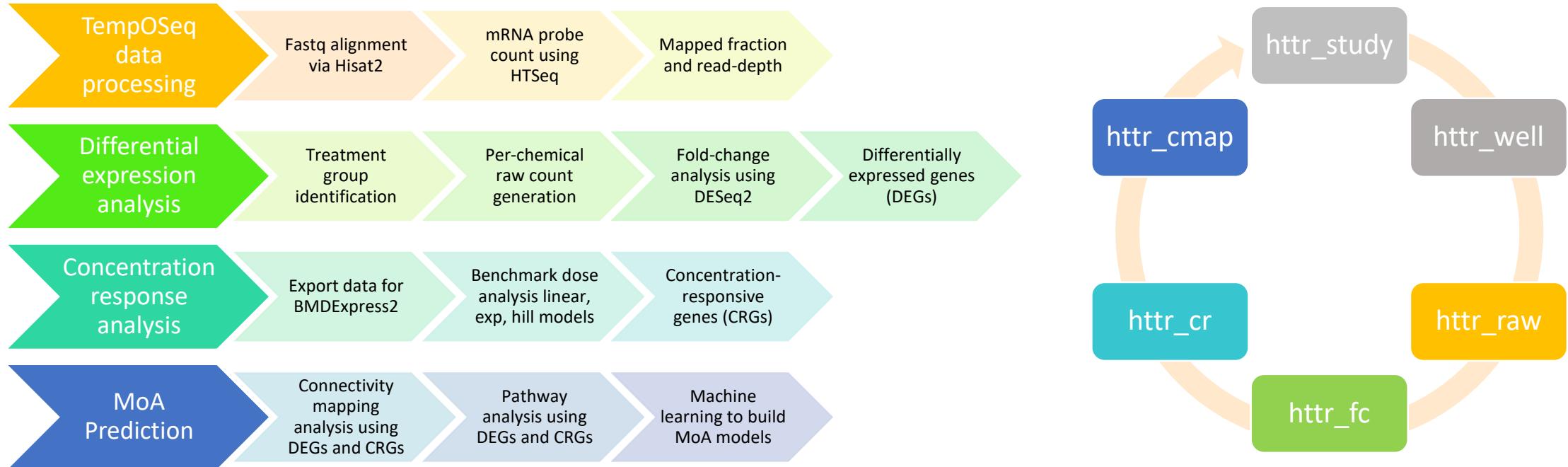


# Treatment Randomization & Quality Control Samples

**Treatment Randomization:** *Each test plate uniquely randomized with respect to treatment.*  
**QC Samples:** *Quality Control samples included on each plate*



# HTTr Analysis Pipeline & Infrastructure



## Python & R analysis pipeline

<http://bitbucket.zn.epa.gov/projects/HTTR/repos/httr-wf-dev>

*Imran Shan  
Josh Harrill  
Woodrow Setzer  
Richard Judson  
Derik Haggard*

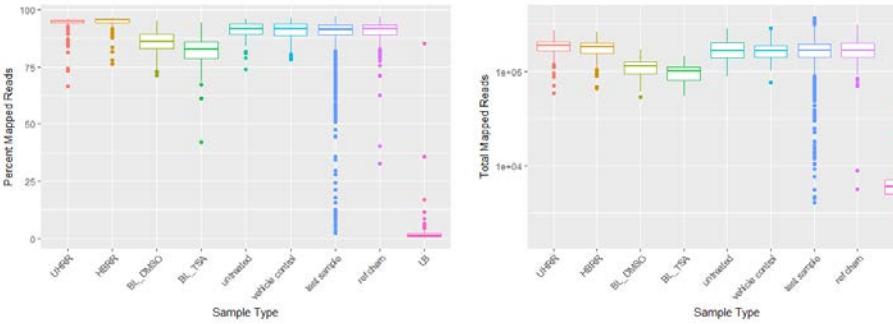
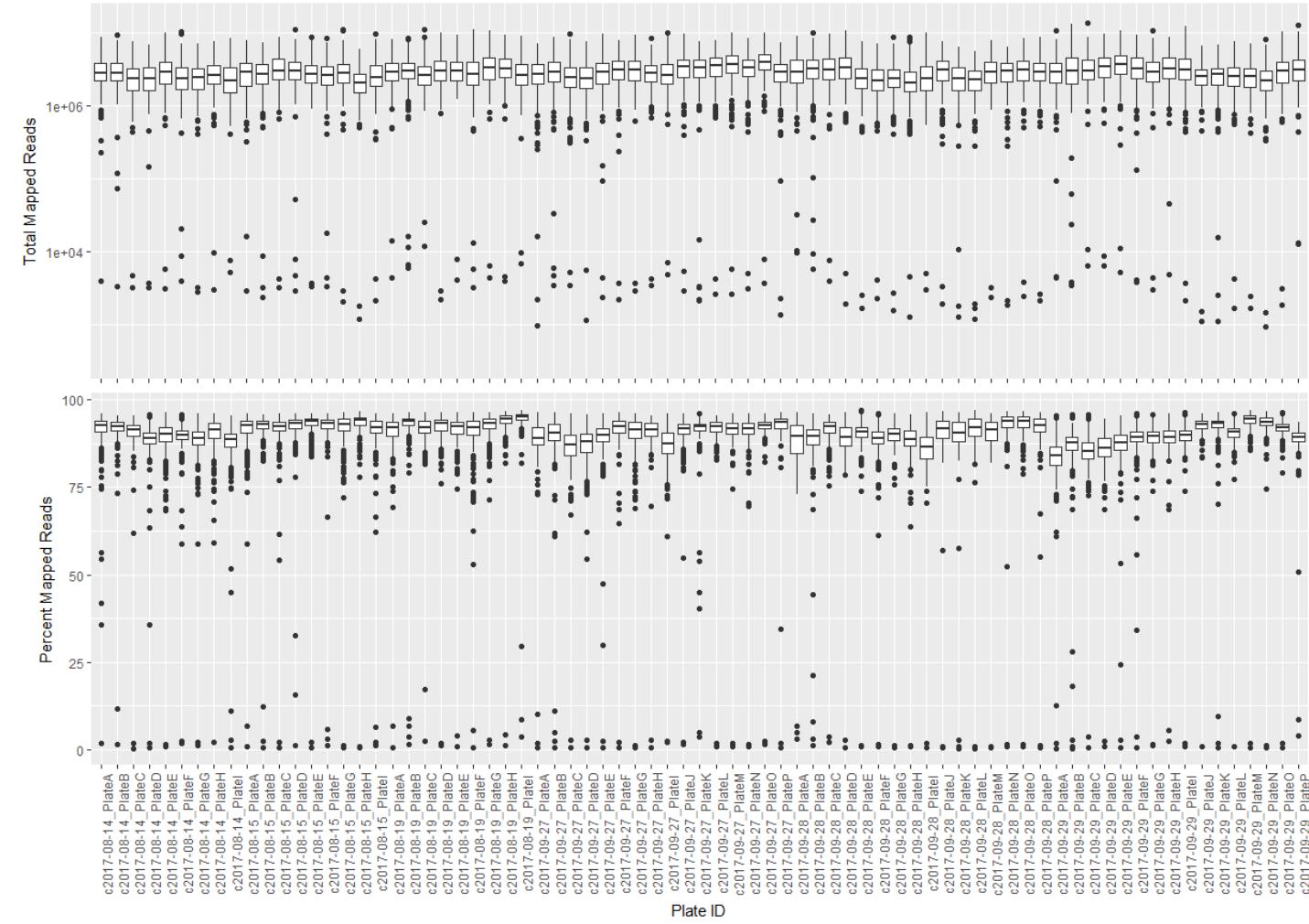
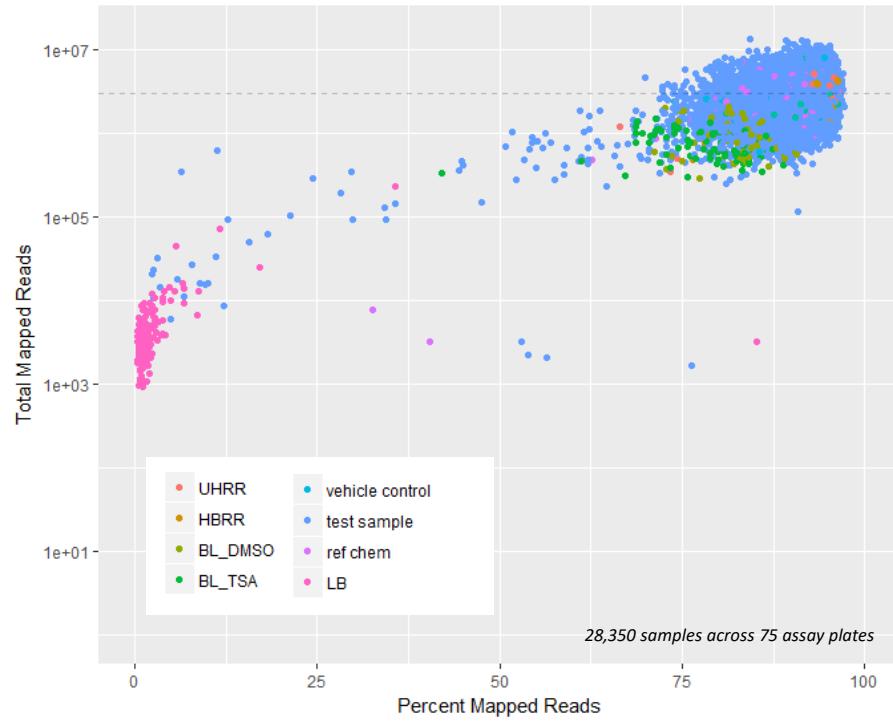
## MongoDB

`mongodb://pb.epa.gov/httr_v1`  
`mongodb://pb.epa.gov/cmap_v2`

*NCCT Investigators  
NCCT IT*

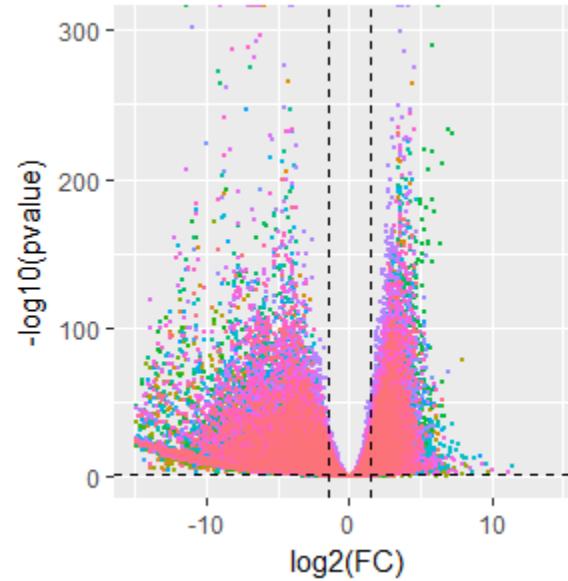
# Technical Reproducibility

# Reproducibility of Read Depths and Mapping Rates

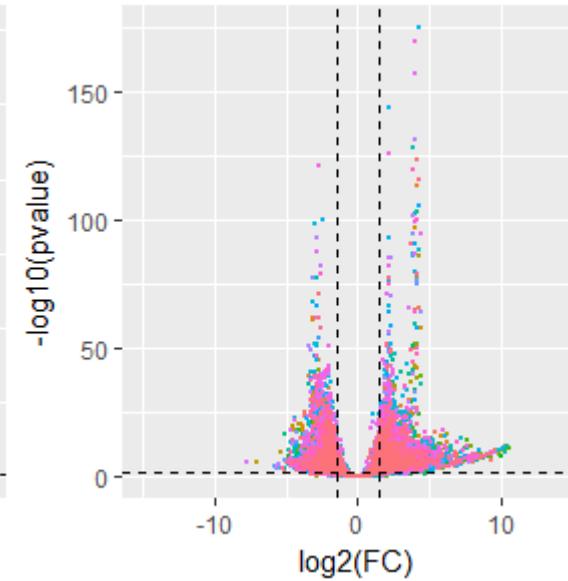


# Reproducibility of $\log_2(\text{FC})$ Estimates

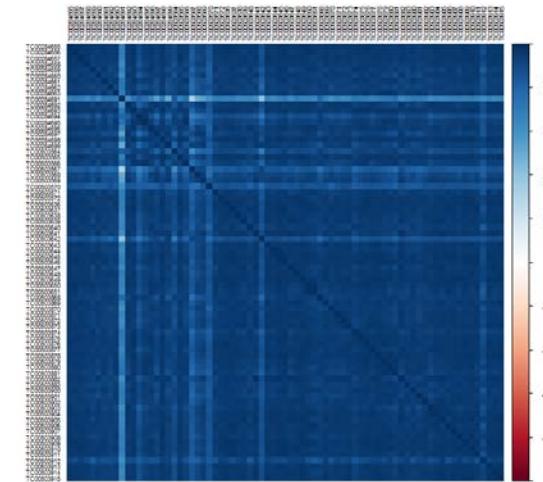
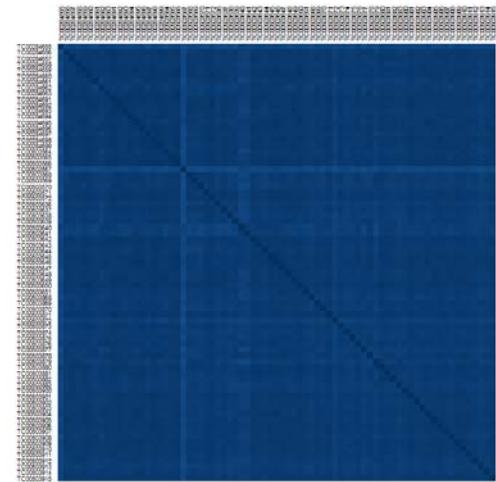
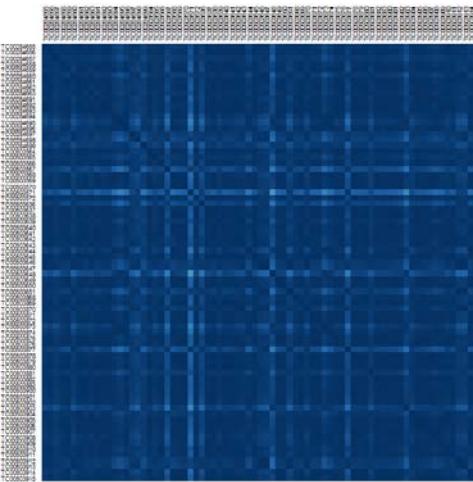
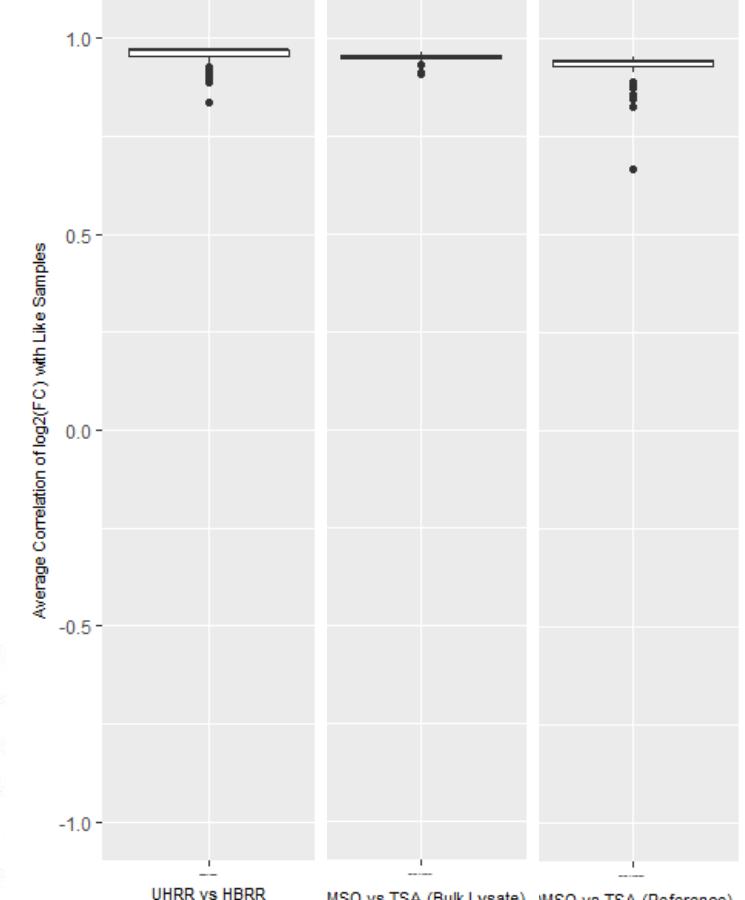
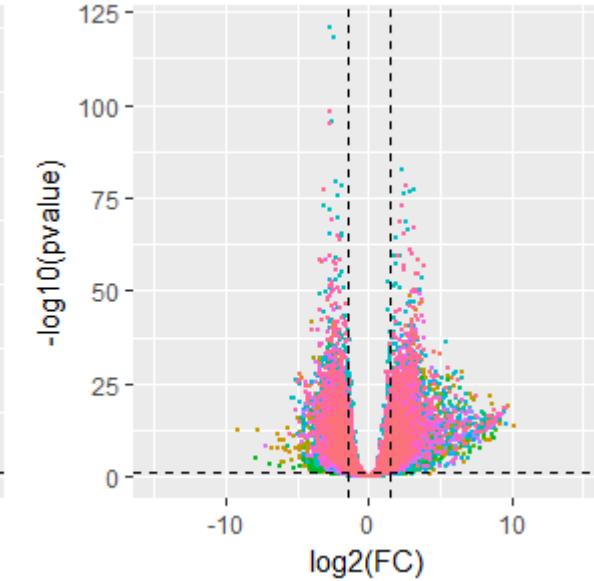
**UHRR v HBRR**



**DMSO vs TSA (Bulk Lysate)**



**DMSO vs TSA (Treatment)**



# Concentration-Response Modeling / Bioactivity Thresholds

# Benchmark Dose Modeling



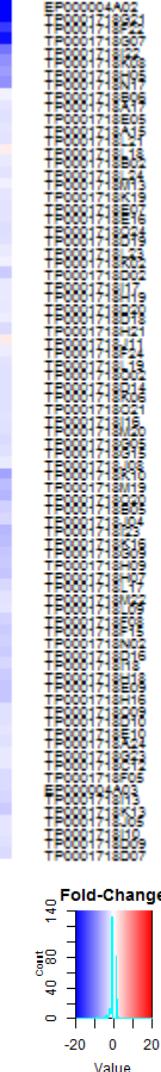
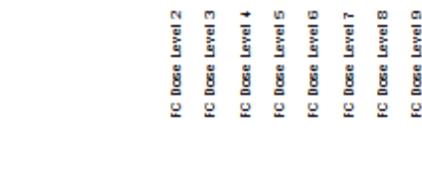
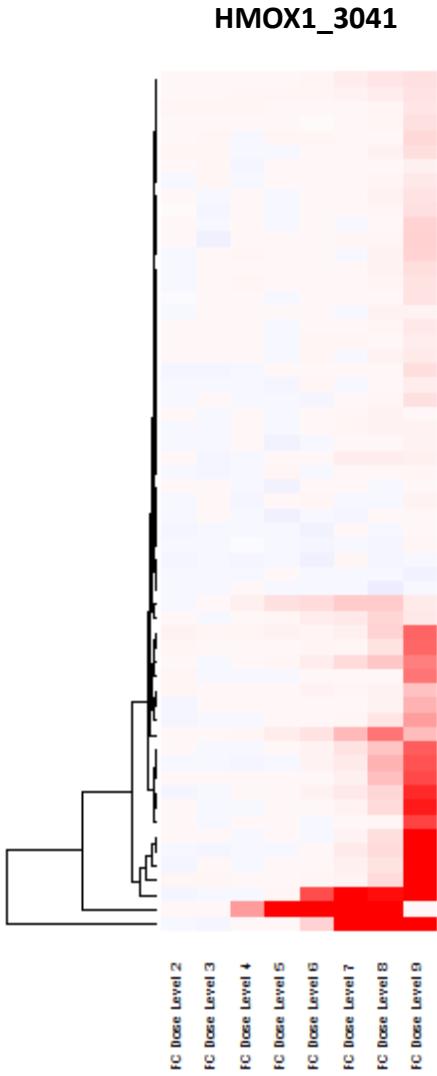
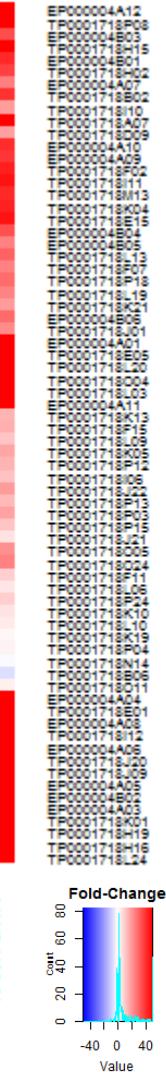
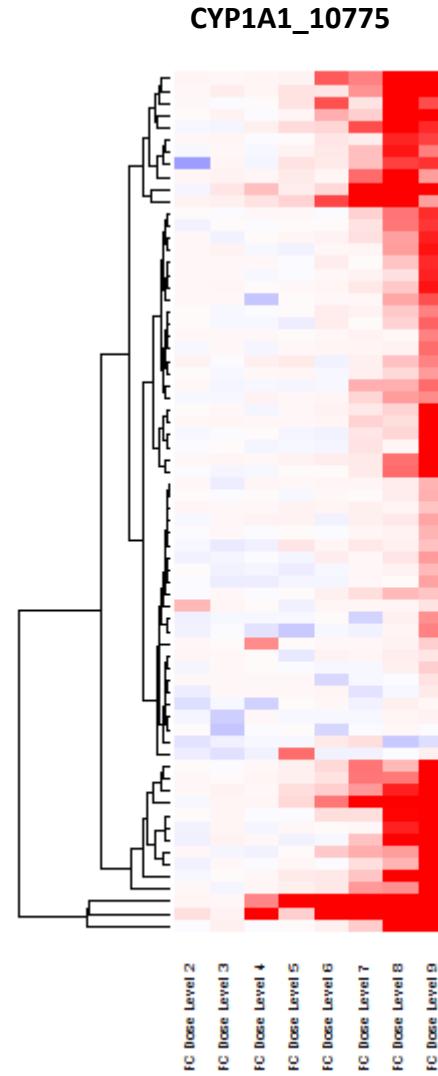
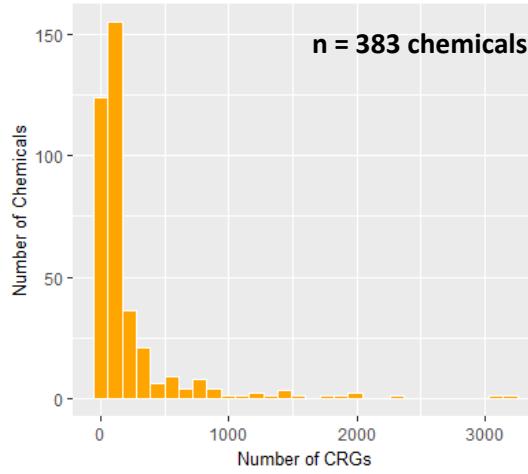
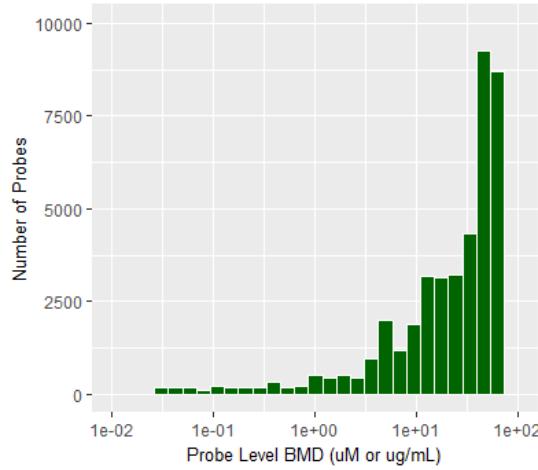
Parameter	Criteria <sup>a</sup>
Pre-filter:	ANOVA ( $p_{\text{raw}} < 0.05$ & $ FC  \geq 2$ )
Models	Hill, Exponential 2, <i>poly2</i> , <i>power</i> , <i>linear</i>
BMR Factor:	1.349 (10 %)
Best Model Selection:	Lowest AIC
Hill Model Flagging <sup>b</sup> :	'k' < 1/3 Lowest Positive Dose Retain Flagged Models
Pathway Analysis:	Genes with $\text{BMD} \leq \text{Highest Dose} \geq 3$ $\geq 1\%$ Gene Set Coverage
Gene Set Collections <sup>c</sup> :	MSigDB_C2 MSigDB_H Reactome BioPlanet KEGG

<sup>a</sup> Exploratory analysis – modeling criteria not finalized

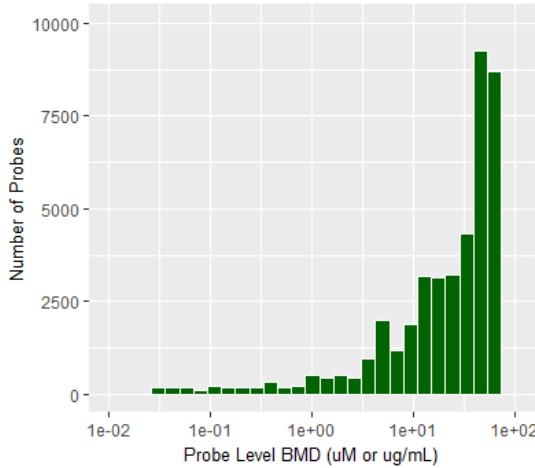
<sup>c</sup> Gene Set Collections:

- **MSigDB\_C2**: Curated gene sets from online pathway databases, publications and knowledge of domain experts (n = 4738).
- **MSigDB\_H**: Coherently expressed signatures derived by aggregating many MSigDB gene sets to represent well-defined biological states or processes (n = 50).
- **Reactome**: Open-source, curated and peer reviewed pathway database with hierarchical pathway relationships in specific domains of biology. (n = 1764). Some pathways included in MSigDB\_C2.
- **BioPlanet** (n = 1700): Curated pathway set developed by National Toxicology Program.

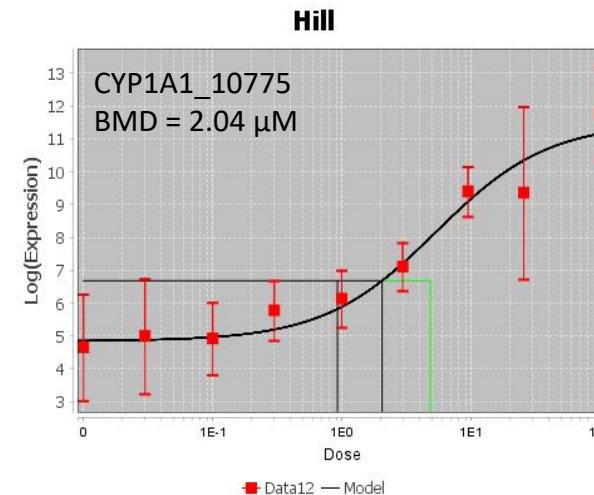
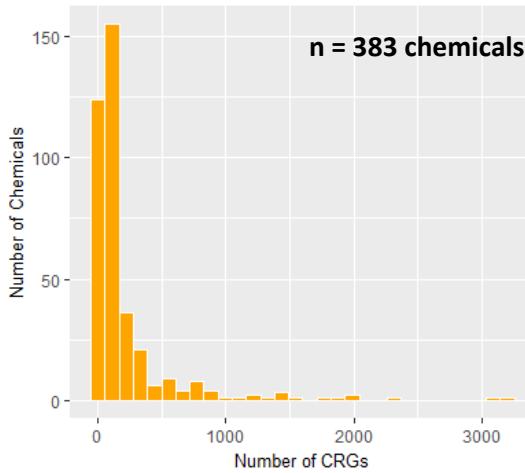
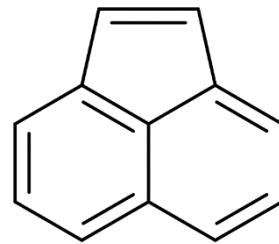
# Benchmark Dose Modeling Summary



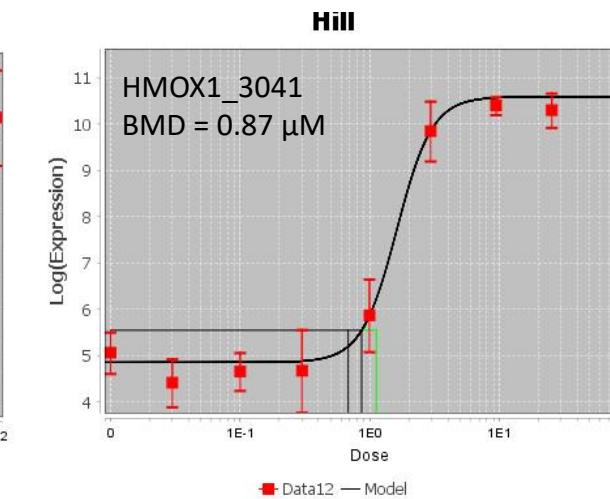
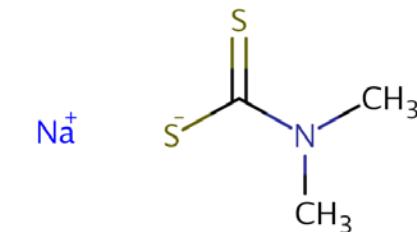
# Benchmark Dose Modeling Summary



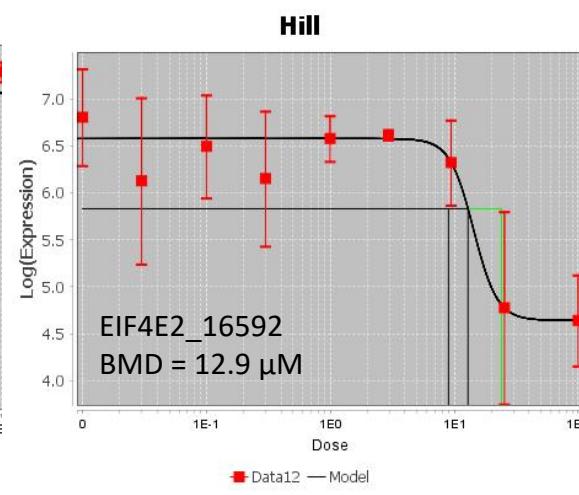
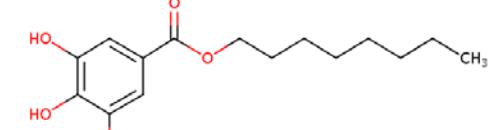
Acenaphthylene  
208-96-8 | DTXSID3023845



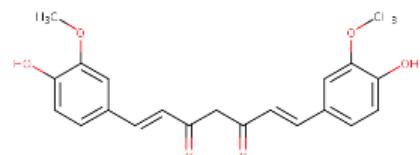
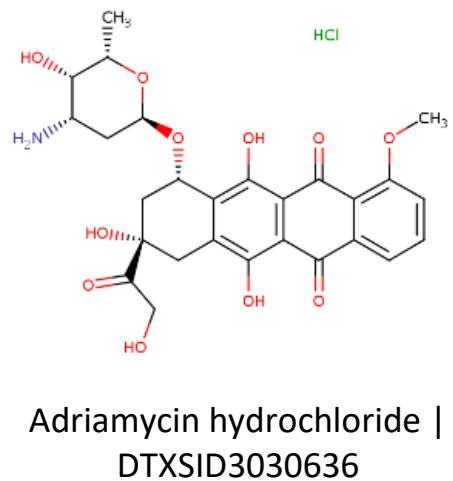
Sodium  
dimethyldithiocarbamate  
128-04-1 | DTXSID6027050



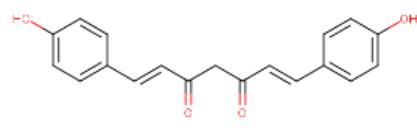
Octyl gallate  
1034-01-1 | DTXSID4040713



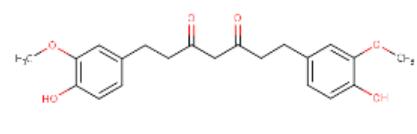
# Case Study Chemicals



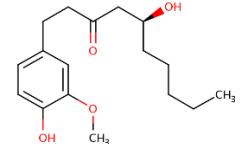
Curcumin |  
DTXSID8031077



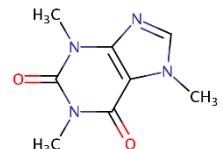
Bisdemethoxycurcumin |  
DTXSID00872663



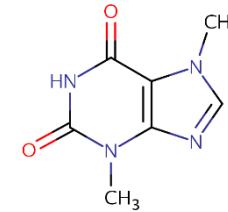
Tetrahydrocurcumin |  
DTXSID30865801



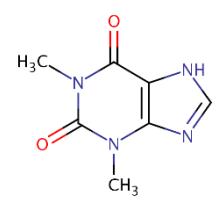
(6)-Gingerol |  
DTXSID3041035



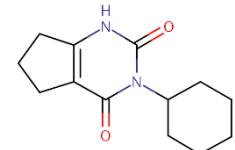
Caffeine |  
DTXSID0020232



Theobromine |  
DTXSID9026132

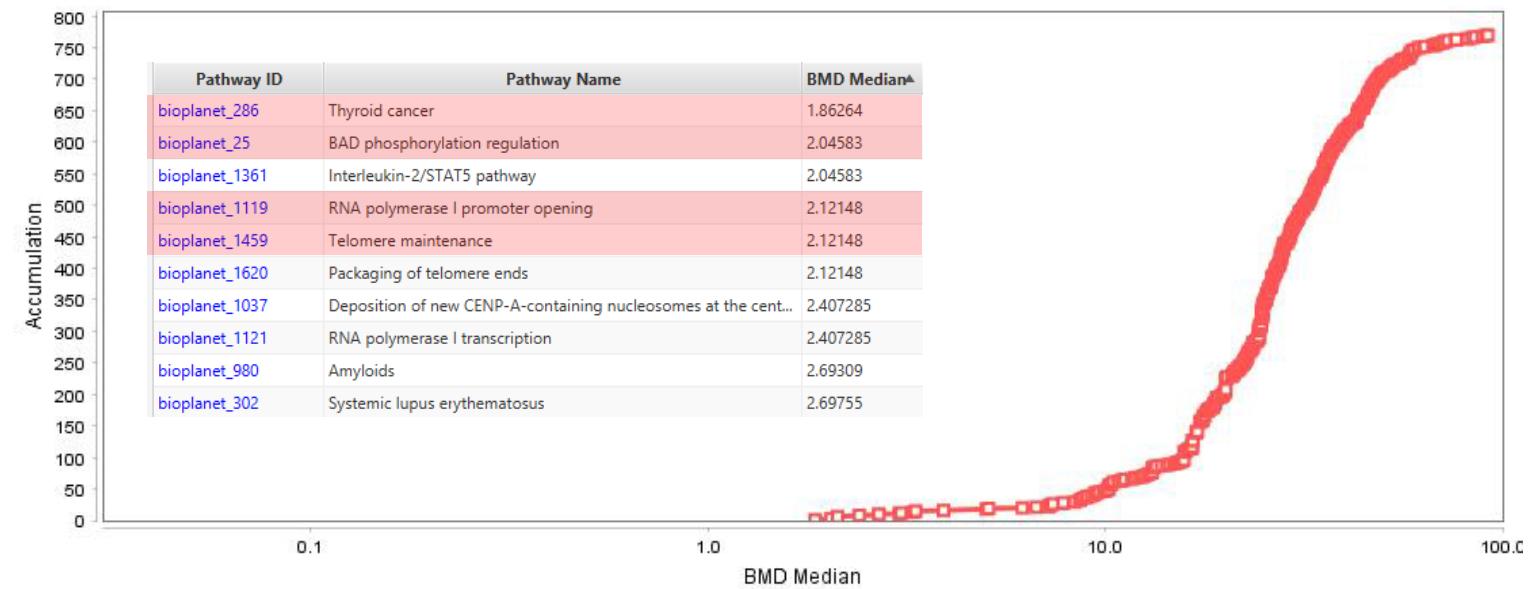
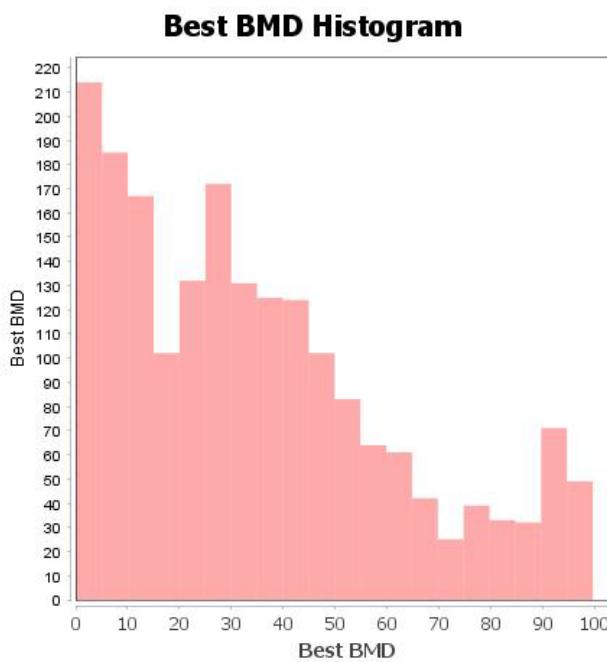
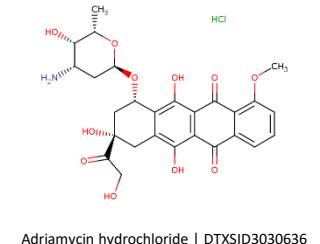
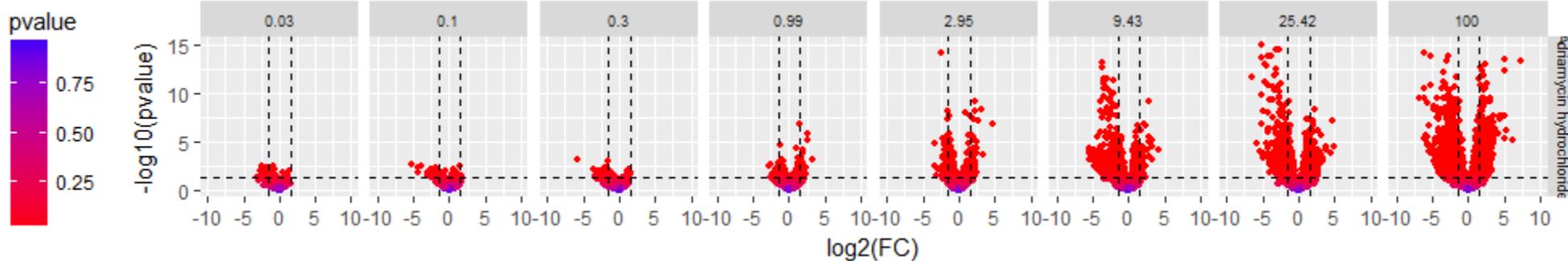


Theophylline |  
DTXSID5021336



Lenacil |  
DTXSID9042093

# Doxorubicin (aka Adriamycin hydrochloride)

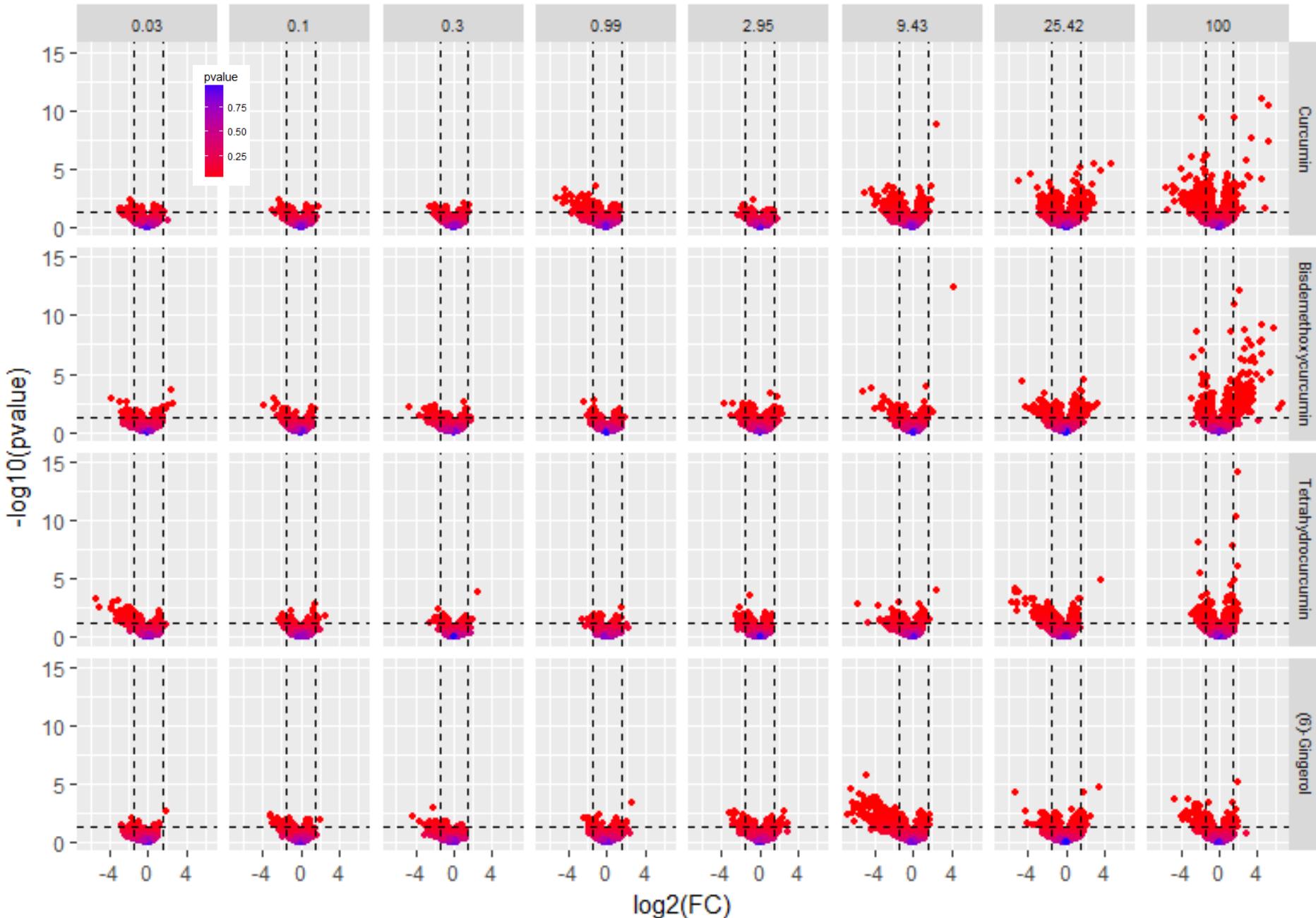


[Int J Mol Cell Med.](#) 2015 Spring;4(2):94-102.

Elevation of cAMP Levels Inhibits Doxorubicin-Induced Apoptosis in Pre-B ALL NALM-6 Cells Through Induction of BAD Phosphorylation and Inhibition of P53 Accumulation.

Fatemi A<sup>1</sup>, Kazemi A<sup>1</sup>, Kashiri M<sup>1</sup>, Safa M<sup>2</sup>.

# Evaluating Structurally Related Chemicals


CC(O)c1ccc(cc1)C=CC2=C(C=C(C=C2)C(=O)C(=O)c3ccc(O)c(O)c3)C(=O)C(=O)c4ccc(O)c(O)c4

Curcumin |  
DTXSID8031077

CC(O)c1ccc(cc1)C=CC2=C(C=C(C=C2)C(=O)C(=O)c3ccc(O)c(O)c3)C=O

Bisdemethoxycurcumin |  
DTXSID00872663

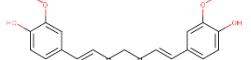
CC(O)c1ccc(cc1)C=CC2=C(C=C(C=C2)C(=O)C(=O)C3=CC=CC=C3)C(=O)C(=O)c4ccc(O)c(O)c4

Tetrahydrocurcumin |  
DTXSID30865801

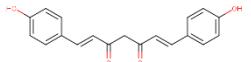
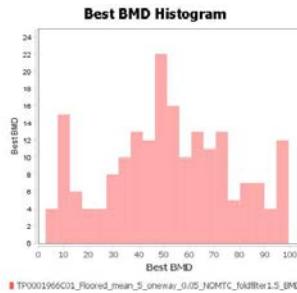
CC(O)C1=CC=CC=C1C2=CC=CC=C2C(=O)C3=CC=CC=C3

(6)-Gingerol |  
DTXSID3041035

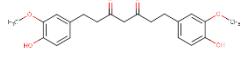
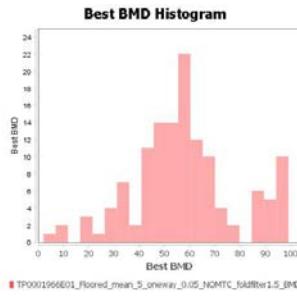
# Gene Set Analysis Using BMD Modeling Results



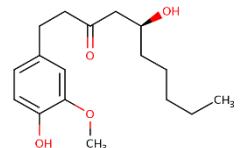
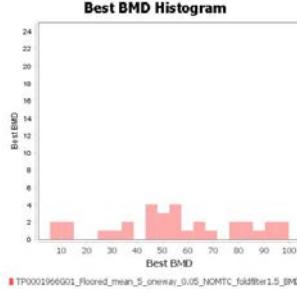
Curcumin | DTXSID8031077



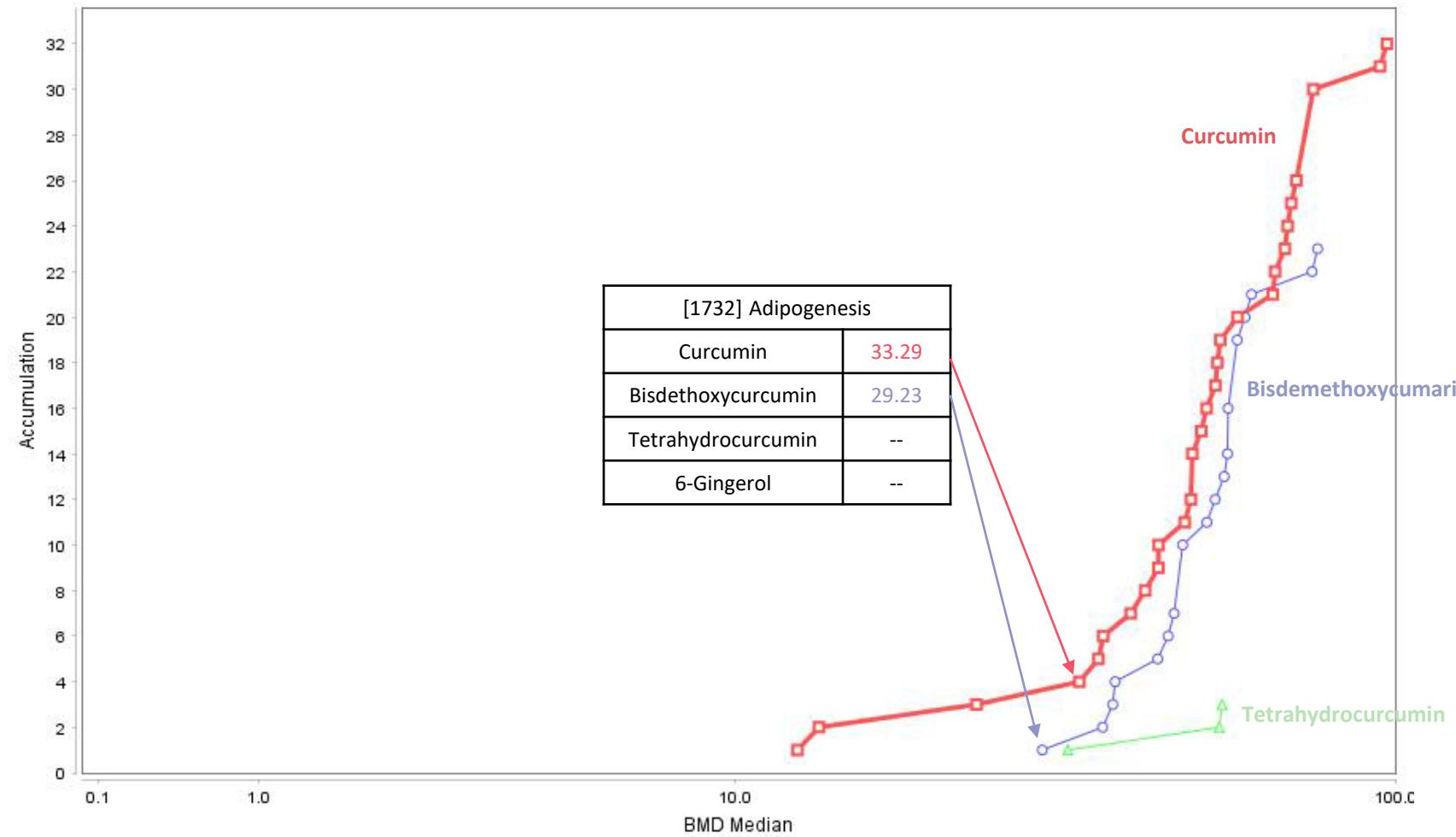
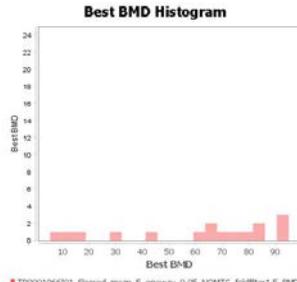
Bisdemethoxycurcumin | DTXSID00872663



Tetrahydrocurcumin | DTXSID30865801



(6)-Gingerol | DTXSID3041035



[Toxicol Appl Pharmacol.](#) 2017 Aug 15;329:158-164. doi: 10.1016/j.taap.2017.05.036.

**Curcumin inhibits adipogenesis induced by benzyl butyl phthalate in 3T3-L1 cells.**

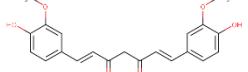
Sakuma S<sup>1</sup>, Sumida M<sup>2</sup>, Endoh Y<sup>2</sup>, Kurita A<sup>2</sup>, Yamaguchi A<sup>2</sup>, Watanabe T<sup>2</sup>, Kohda T<sup>2</sup>, Tsukiyama Y<sup>2</sup>, Fujimoto Y<sup>3</sup>.

[J Agric Food Chem.](#) 2016 Feb 3;64(4):821-30. doi: 10.1021/acs.jafc.5b05577. Epub 2016 Jan 25.

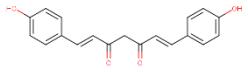
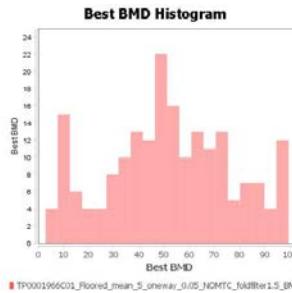
**Bisdemethoxycurcumin Inhibits Adipogenesis in 3T3-L1 Preadipocytes and Suppresses Obesity in High-Fat Diet-Fed C57BL/6 Mice.**

Lai CS<sup>1,2</sup>, Chen YY<sup>1</sup>, Lee PS<sup>1</sup>, Kalyanam N<sup>3</sup>, Ho CT<sup>4</sup>, Liou WS<sup>5</sup>, Yu RC<sup>1</sup>, Pan MH<sup>1,6,7,8</sup>.

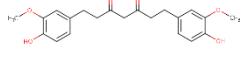
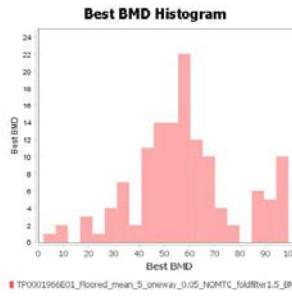
# Gene Set Analysis Using BMD Modeling Results



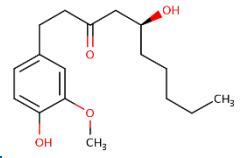
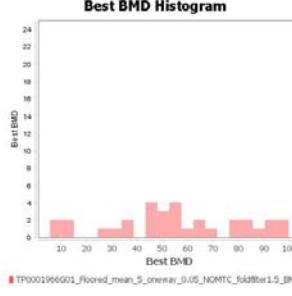
Curcumin | DTXSID8031077



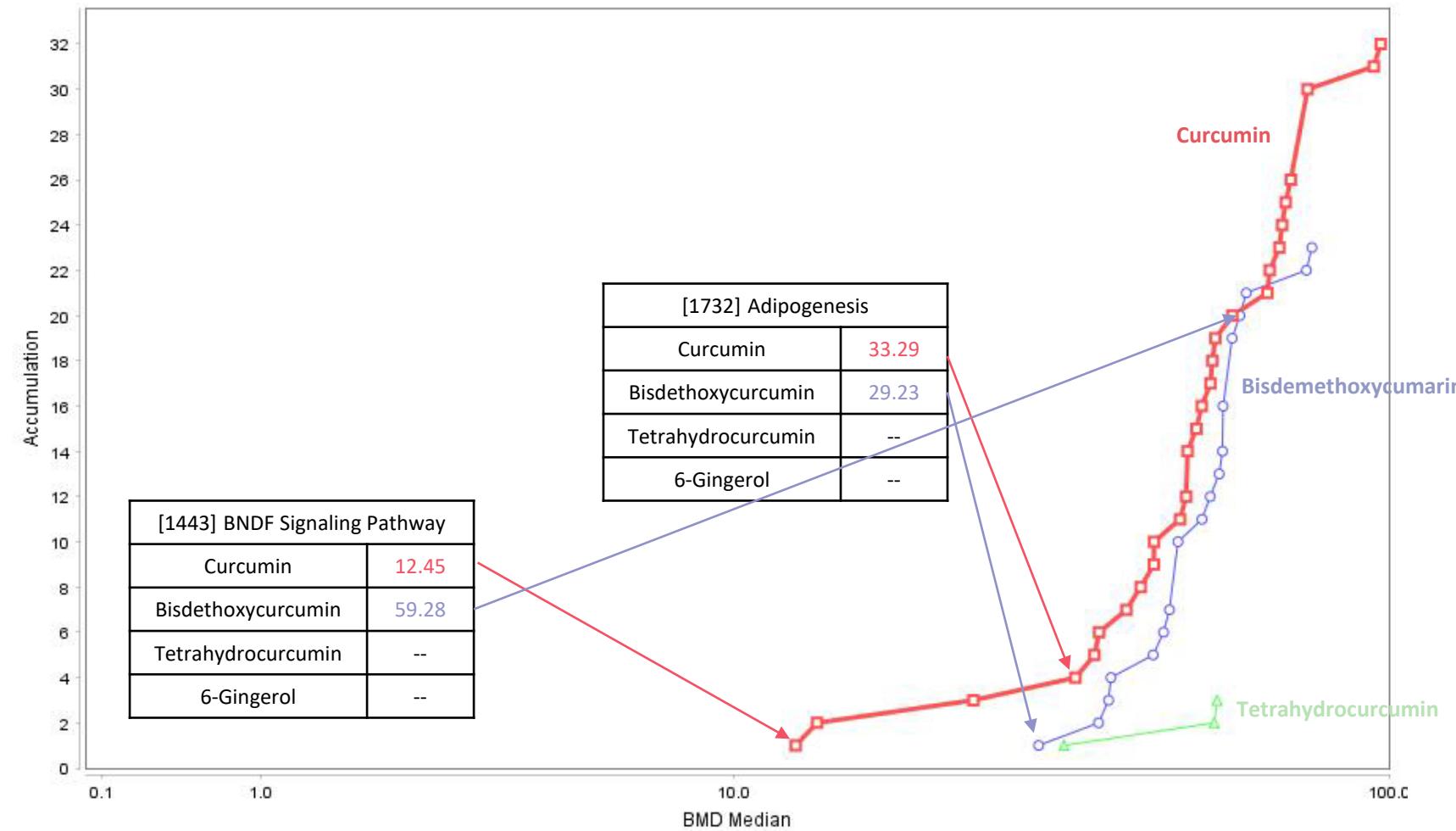
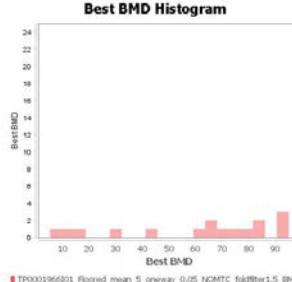
Bisdemethoxycurcumin | DTXSID00872663



Tetrahydrocurcumin | DTXSID30865801



(6)-Gingerol | DTXSID3041035



[Neuropeptides](#). 2016 Apr;56:25-31. doi: 10.1016/j.npep.2015.11.003. Epub 2015 Nov 11.

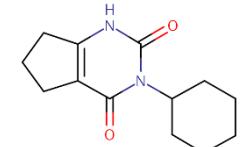
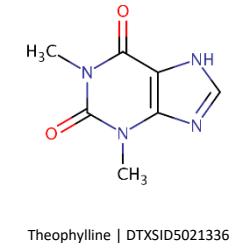
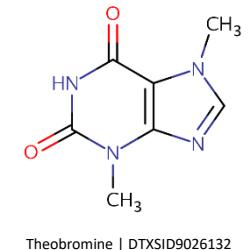
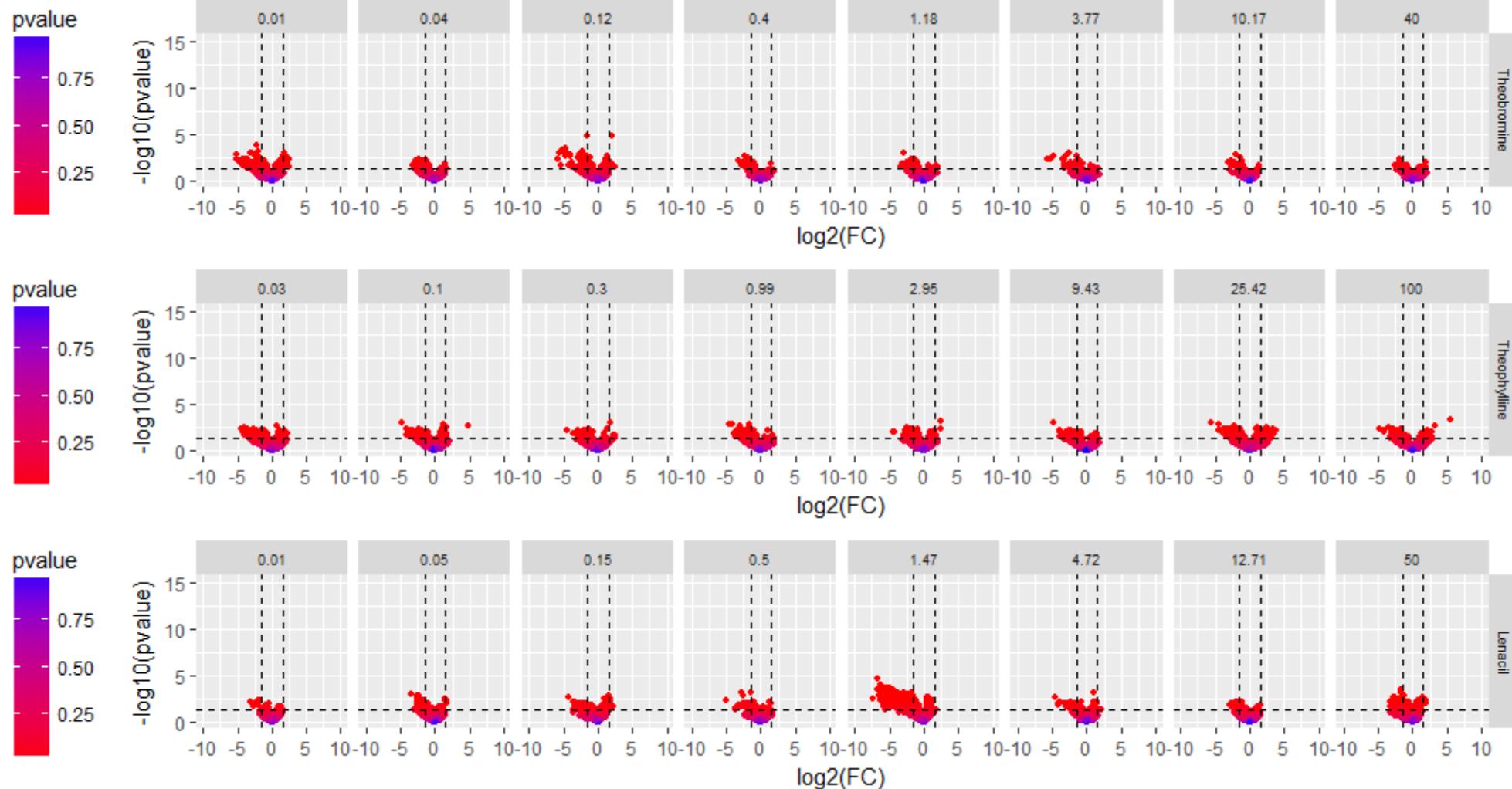
Effect of curcumin on serum brain-derived neurotrophic factor levels in women with premenstrual syndrome: A randomized, double-blind, placebo-controlled trial. [Fanaei H<sup>1</sup>](#), [Khayat S<sup>2</sup>](#), [Kasaeian A<sup>3</sup>](#), [Javadimehr M<sup>4</sup>](#).

[Biomed Pharmacother](#). 2017 Mar;87:721-740. doi: 10.1016/j.biopha.2016.12.020. Epub 2017 Jan 14.

Curcumin confers neuroprotection against alcohol-induced hippocampal neurodegeneration via CREB-BDNF pathway in rats.

[Motaghinejad M<sup>1</sup>](#), [Motevalian M<sup>2</sup>](#), [Fatima S<sup>3</sup>](#), [Hashemi H<sup>1</sup>](#), [Gholami M<sup>4</sup>](#).

# Caffeine Analogs



- Very few dose-responsive genes
- No significantly enriched pathways using BMDe standard workflow

# Alternative Approach for Gene Set Analysis

## Step 1: Calculate Response

- A gene set is a list / bag of genes
- Under one condition (chemical x dose) calculate “gene set response” separately for genes in the set and out of the set:

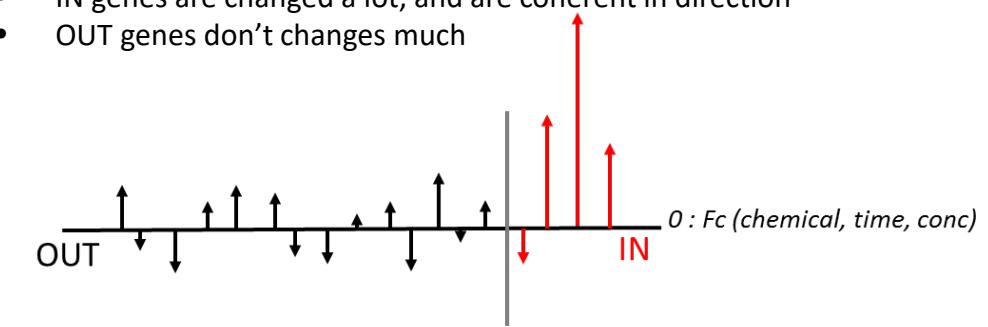
$$M = \sum_{i=1}^{ngene} \frac{fc_i}{sc_i^2} \sqrt{\sum_{i=1}^{ngene} \frac{1}{sc_i^2}}$$

$$R = M_{in} - M_{out}$$

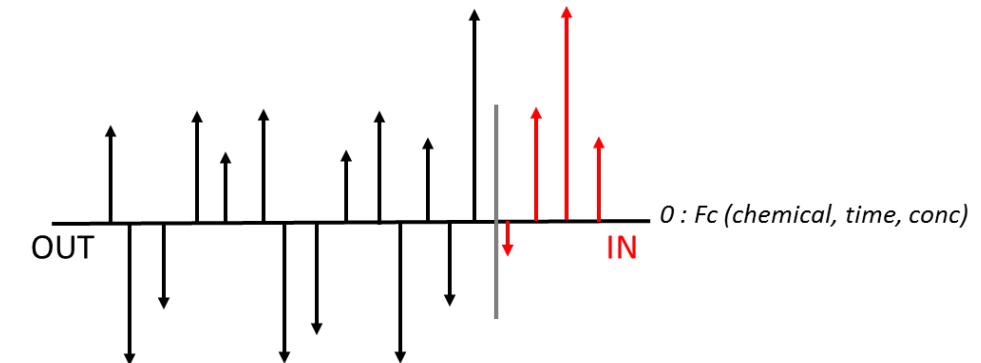
## Step 2: CR Modeling

- For each chemical, fit using tcplFit
  - Constant, Hill , Gain-Loss methods
  - BMAD(pathway) = MAD of response for the pathway across the two lowest concentrations across all chemicals and times
- Hitcall:
  - tcplFit calls a hit
  - Top > 3\*BMAD

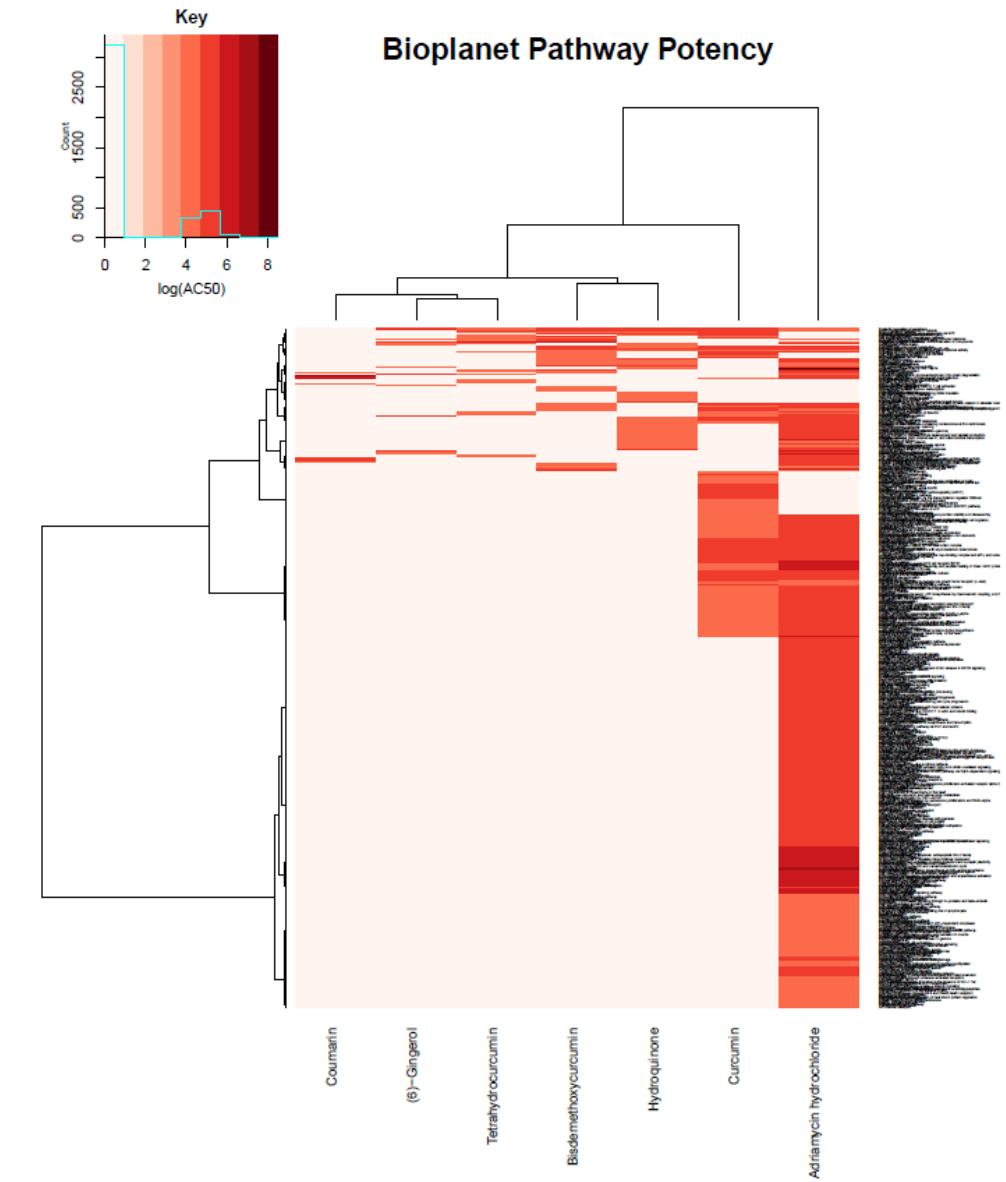
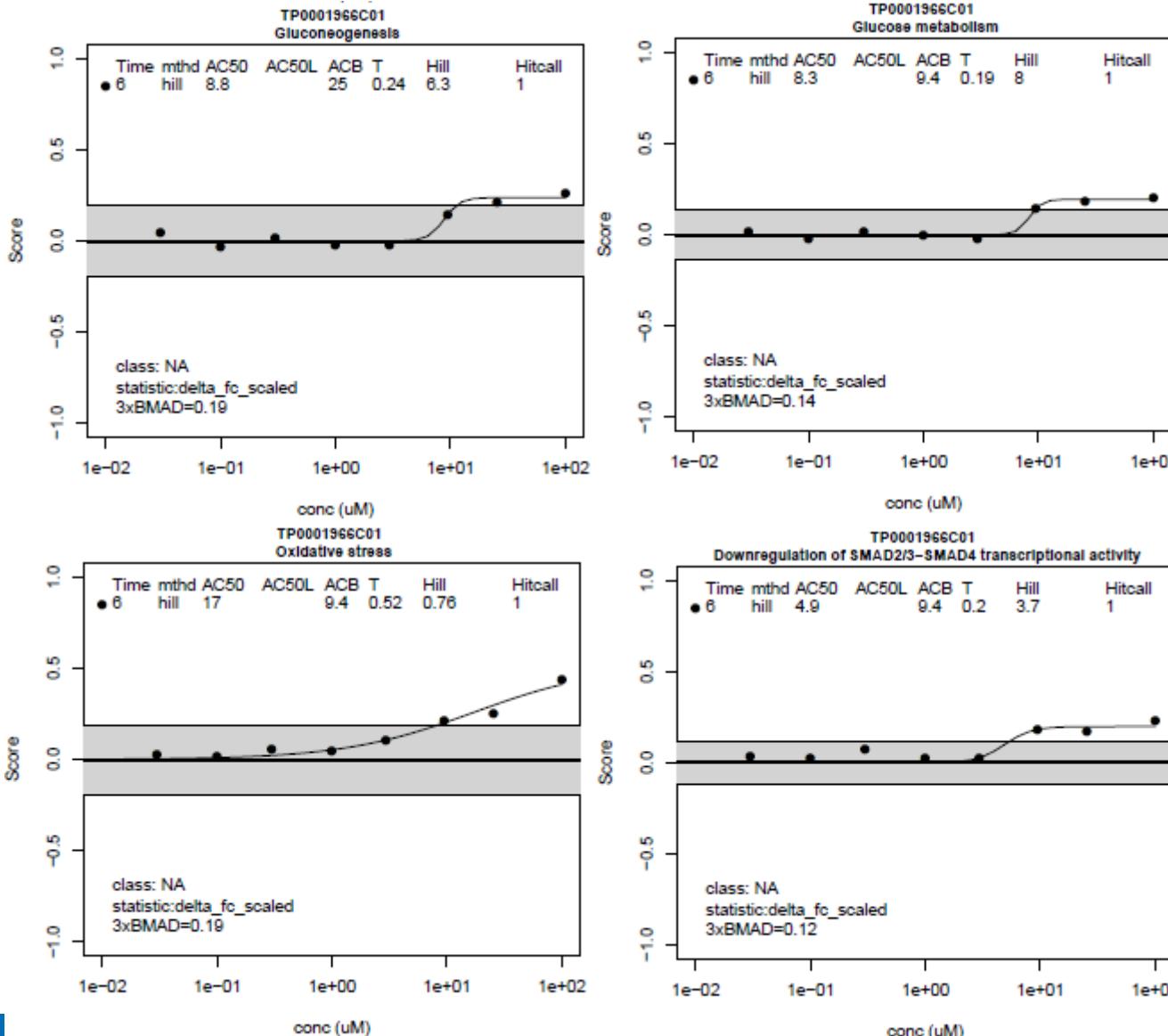
- IN genes are changed a lot, and are coherent in direction
- OUT genes don't changes much



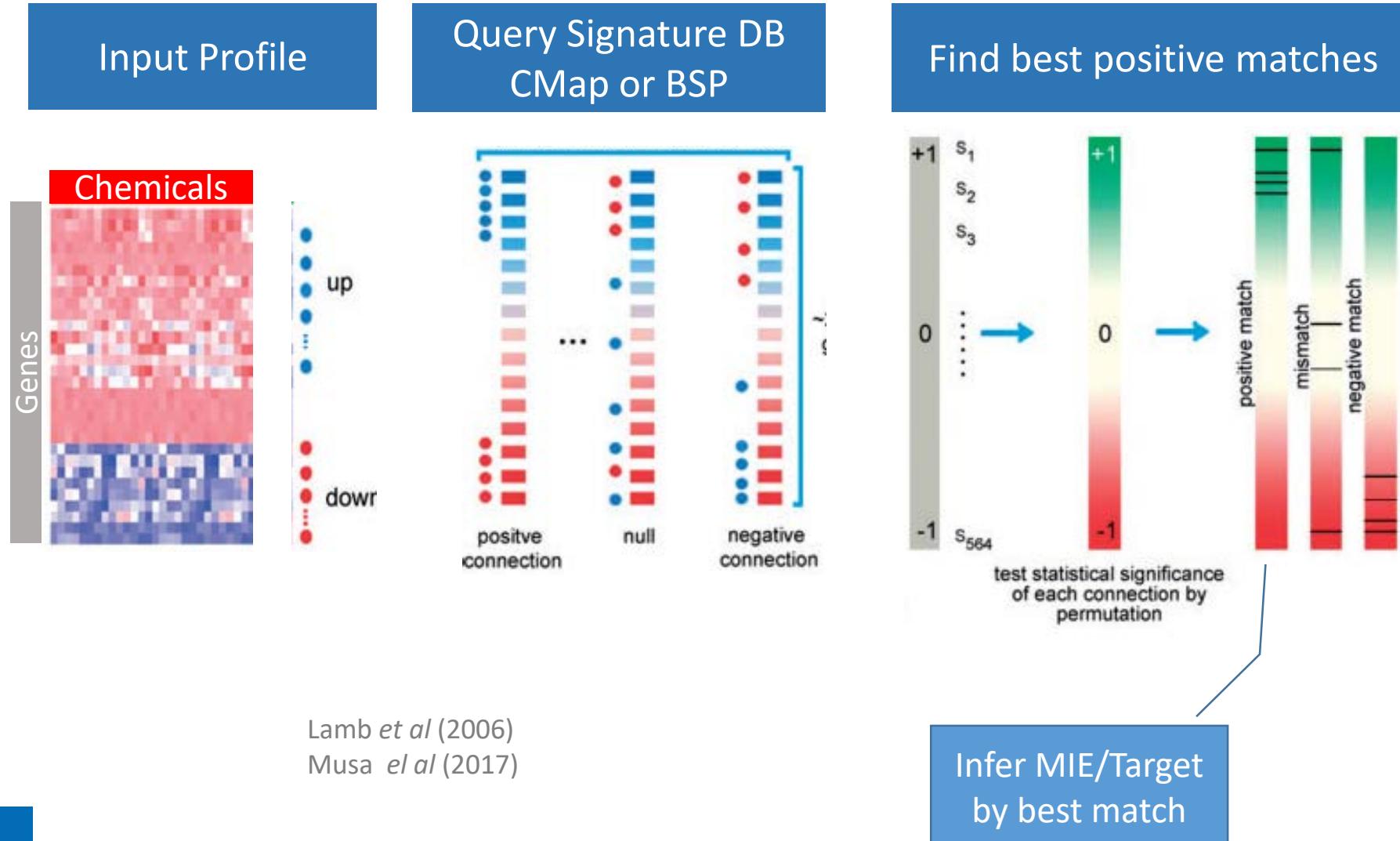
- IN genes are changed a lot, and are coherent in direction
- OUT genes change a lot but are not coherent (mean  $\sim 0$ )



# Comparing Activities Across Chemicals



# MOA by Connectivity Mapping



## Issues

- Translating DEG/CRG to signature
- Many measures of similarity
- Only as good as reference chemical MoA annotation
- Highly sensitive but not very specific
- Chemicals that cause global perturbations “hit” all MoAs – how do we distinguish signal from noise ?

# Various Approaches to Connectivity Mapping

**DRG:** Differentially Expressed Gene

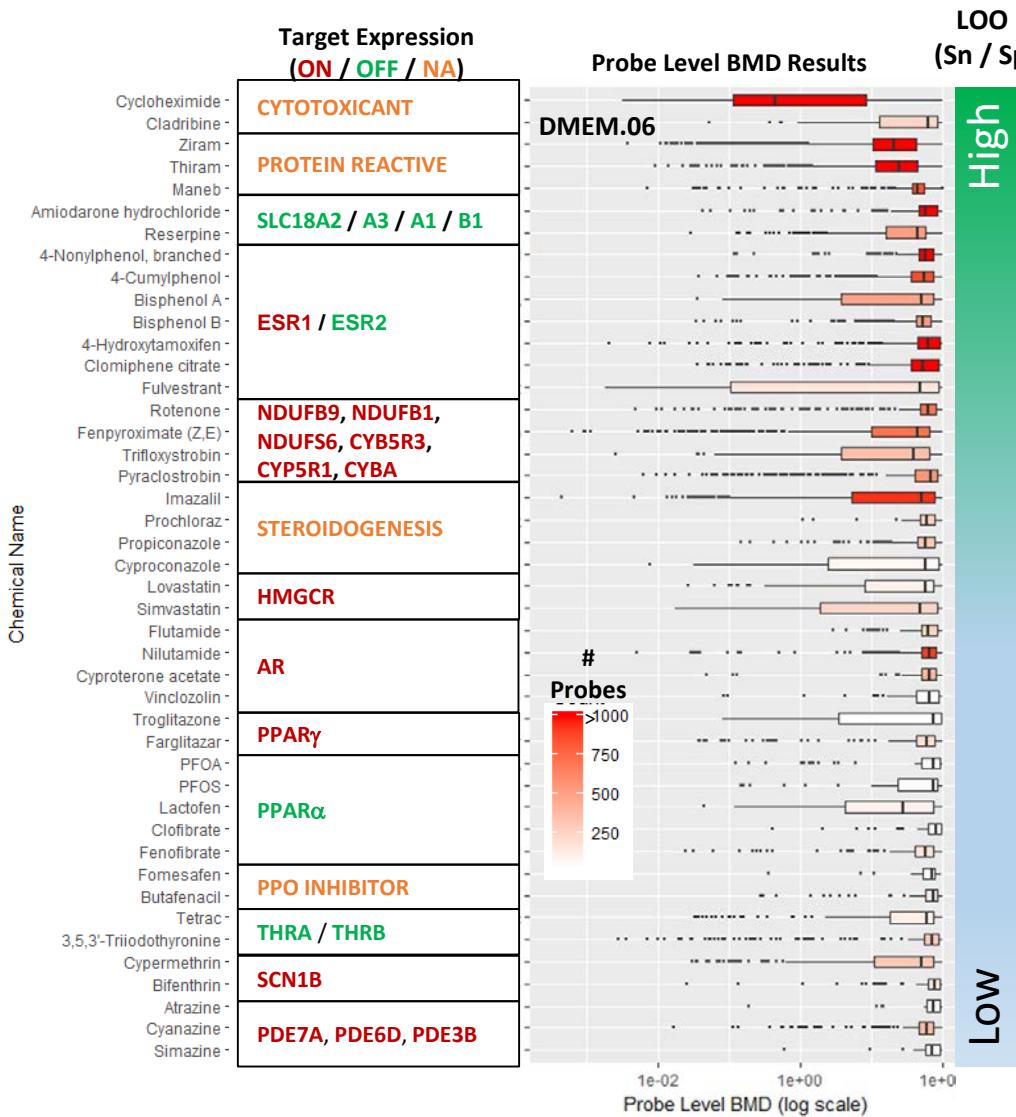
→ Based on single-conc. comparison to control.

**CRG:** Concentration-Responsive Gene

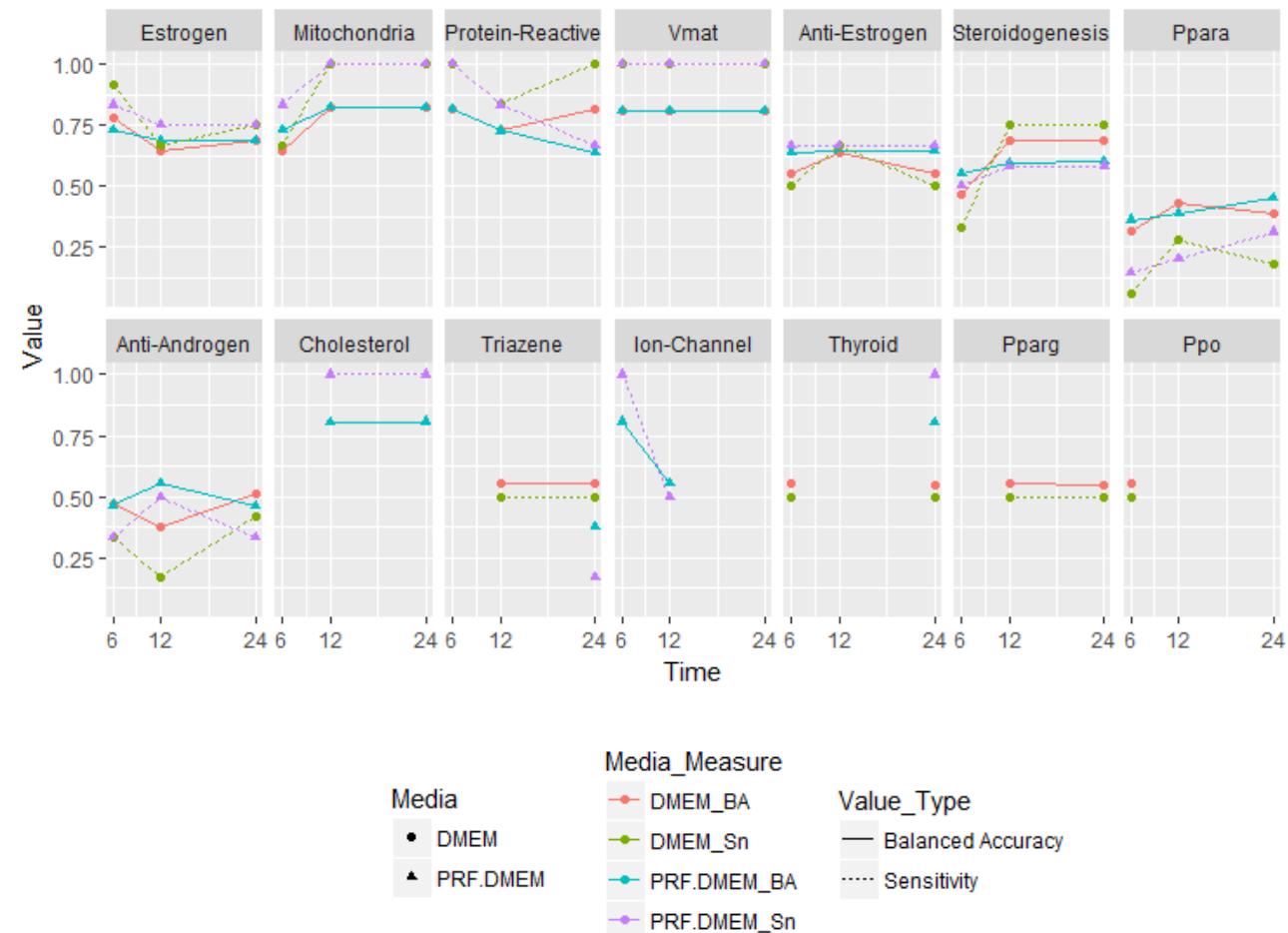
→ Based on ANOVA or conc.-response modeling.



# Connectivity Mapping for MoA Prediction



## Sensitivity and Balanced Accuracy of MoA Prediction Using CRGs and Leave-One-Out Cross-Validation



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