Supplementary Information for

Asymmetric Photocatalytic C-H Functionalization of Toluene and Derivatives

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A. General Information

The NMR spectra were recorded at 400 MHz and 500 MHz for ¹H, at 101 MHz and 125.8 MHz for ¹³C, at 376 MHz for ¹⁹F and at 47 MHz for ¹¹B. The chemical shifts (δ) for ¹H and ¹³C are given in ppm relative to residual signals of the solvents (CHCl₃ @ 7.26 ppm ¹H NMR, 77.00 ppm ¹³C NMR). Coupling constants are given in Hz and are quoted to the nearest 0.5 Hz. The following abbreviations are used to indicate the multiplicity: s, singlet; d, doublet; t, triplet; q, quartet; p, pentet; m, multiplet; br s, broad signal;, app t, apparent triplet.

High-resolution mass spectra (HRMS) were obtained from the ICIQ High Resolution Mass Spectrometry Unit on MicroTOF Focus and Maxis Impact (Bruker Daltonics) with electrospray ionizationOptical rotations were measured on a Polarimeter Jasco P-1030 and are reported as follows: $[\alpha]_D$ rt (c in g per 100 mL, solvent).

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General Procedures. All reactions were set up under an argon atmosphere in oven-dried glassware using standard Schlenk techniques, unless otherwise stated. Synthesis grade solvents were used as purchased. Anhydrous solvents were taken from a commercial SPS solvent dispenser. Chromatographic purification of products was accomplished using force-flow chromatography (FC) on silica gel (35-70 mesh). For thin layer chromatography (TLC) analysis throughout this work, Merck precoated TLC plates (silica gel 60 GF₂₅₄, 0.25 mm) were used, using UV light as the visualizing agent and either phosphomolybdic acid in EtOH or basic aqueous potassium permangante (KMnO₄), and heat as developing agents. Organic solutions were concentrated under reduced pressure on a Büchi rotary evaporator (in vacuo at 40 °C, ~5 mbar).

Determination of Diastereomeric Ratio. The diastereomeric ratio was determined by ¹H NMR analysis of the crude reaction mixture through integration of diagnostic signals.

Determination of Enantiomeric Purity: HPLC analysis on chiral stationary phase was performed on an Agilient 1200 series HPLC, using a Daicel Chiralpak IC-3 column with *i*-PrOH:hexane as the eluent. UPC² analysis on chiral stationary phase was performed on an Waters Acquity instrument using a CEL1, ID3 or IE3 chiral colums. The exact conditions for the analyses are specified within the characterization section. HPLC/UPC² traces were compared to racemic samples prepared by running the reactions in the presence of a catalytic amount (20 mol%) of racemic catalyst **A**. This was prepared by mixing equimolar amounts of amine catalysts (*S*)-**A** and its enantiomer (*R*)-**A**, both synthetized according to literature procedures.^{1,2}

Materials: Commercial grade reagents and solvents were purchased at the highest commercial quality from Sigma Aldrich, Fluka, Acros Organics, Tokyo Chemical Industry, Fluorochem or Alfa Aesar and used as received, unless otherwise stated. Organic catalysts (*S*)-**A**, (*R*)-**A** and (*S*)-**B** were synthesized according to previously reported procedures.^{1,2} The majority of enals **1** and toluene derivatives are commercially available. Cinnamaldehyde **1a** was distilled prior to use. Enals **1d** and **1e** were prepared according to the literature procedure,² whereas the synthetic procedures for

compounds **1f**, **1g**, **3** and for the benzylic derivatives employed in the synthesis of products **2m** and **2q** are reported in section B of the Supporting Information.

B. Substrate Synthesis

Synthesis of (E)-3-(4-(trimethylsilyl)phenyl)acrylaldehyde (1f):



Compound **1f** was prepared according to a reported literature procedure.² An oven-dried two-necked flask, fitted with a condenser, was charged with Pd(OAc)₂ (29.0 mg, 131 µmol), K₂CO₃ (905 mg, 6.54 mmol), KCl (325 mg, 4.36 mmol), (*n*-Bu)₄NOAc (2.63 g, 8.73 mmol), (4-bromophenyl)trimethylsilane (853 µL, 4.36 mmol) and 3,3-diethoxyprop-1-ene (2.00 mL, 13.1 mmol) in anhydorus DMF (45 mL). The resulting mixture was heated to 90 °C and stirred for 18 hours. The reaction was allowed to cool to room temperature and quenched by dropwise addition of 2M *aq*. HCl solution (30 mL). The resulting mixture was stirred at room temperature for further 15 minutes. Water (100 mL) was added and the crude mixture was extracted with Et₂O (3 x 150 mL). The organic phase were collected, dried over Na₂SO₄, filtered and concentrated *in vacuo*. Purification of the crude material by column chromatography on silica gel (gradient from 5% EtOAc in hexane to 10% EtOAc in hexane) gave product **1f** (766 mg, 86% yield) as an off-white solid.

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.73 (d, J = 7.5 Hz, 1H), 7.65 – 7.43 (m, 5H), 6.76 (dd, J = 16.0, 7.5 Hz, 1H), 0.31 (s, 9H).

¹³C NMR (101 MHz, CDCl₃): δ 193.8, 152.9, 145.2, 134.2, 134.0, 128.7, 127.6, -1.3.

Synthesis of (*E*)-3-(3-(trifluoromethyl)phenyl)acrylaldehyde (1g):



Compound **1g** was prepared according to a reported literature procedure.² To a solution of (formylmethyl)triphenylphosphonium chloride (3.55 g, 10.4 mmol) in anhydrous CH_2Cl_2 (50 mL) was added Et_3N (2.18 mL, 15.6 mmol) and, successively, 3-(trifluoromethyl)benzaldehyde (1.39 mL, 10.4 mmol) dropwise. The reaction was stirred for 48 hours at room temperature and, then, concentrated under reduced pressure. Purification of the crude material by column chromatography on silica gel (gradient from 5% EtOAc in hexane to 10% EtOAc in hexane) gave product **1g** (630 mg, 30% yield) as a yellow oil. The spectroscopic properties of this compound were consistent with the data available in the literature.³

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.77 (d, J = 7.5 Hz, 1H), 7.83 (br. s, 1H), 7.78 (d, J = 7.5 Hz, 1H), 7.72 (d, J = 7.5 Hz, 1H), 7.60 (dd, J = 7.5 Hz, 1H), 7.53 (d, J = 16.0 Hz, 1H), 6.80 (dd, J = 16.0, 7.5 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃): δ 193.1, 150.3, 134.8, 131.8 (q, ${}^{2}J_{C-F}$ = 33.0 Hz), 131.2 (q, ${}^{4}J_{C-F}$ = 1.5 Hz), 130.0, 129.7, 127.6 (q, ${}^{3}J_{C-F}$ = 3.5 Hz), 125.2 (q, ${}^{3}J_{C-F}$ = 4.0 Hz), 123.6 (q, ${}^{1}J_{C-F}$ = 272.5 Hz). ¹⁹F NMR (376 MHz, CDCl₃): δ -63.09 (s, 3F).

Synthesis of (cyclopropylmethyl)benzene:



This compound was prepared according to a reported literature procedure.⁴ To a mixture of (bromomethyl)cyclopropane (1.94 mL, 20.0 mmol), CuCl₂ (54.0 mg, 0.40 mmol) and 1-phenyl-1-propyne (0.25 mL, 2.00 mmol) in anhydorus THF (25 mL) at 0 °C was added a 3M solution of phenylmagnesium bromide (10.0 mL, 30.0 mmol) in Et₂O. The mixture was stirred at 0 °C for 20 minutes, then, warmed up to room temperature and stirred overnight. The reaction was quenched by slow addition of 1M *aq*. HCl solution (25 mL) and *sat. aq*. NH₄Cl solution (25 mL). The resulting mixture was extracted with Et₂O (3 x 100 mL). The organic phase were collected, dried over Na₂SO₄ and the solvent was removed under reduced pressure to deliver the crude material. Purification by column chromatography on silica gel (hexane) provided the desired copuling product (2.38 g, 90% yield) as a colourless liquid. The spectroscopic properties of this compound were consistent with the data available in the literature.⁵

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 7.42 – 7.31 (m, 4H), 7.32 – 7.23 (m, 1H), 2.63 (d, J = 7.0 Hz, 2H), 1.14 – 1.01 (m, 1H), 0.63 – 0.58 (m, 2H), 0.38 – 0.21 (m, 2H). ¹³C NMR (101 MHz, CDCl₃): δ 142.2, 128.4, 128.3, 125.9, 40.4, 11.9, 4.7.

Synthesis of 2-(4-phenylbutyl)isoindoline-1,3-dione:



A solution of 4-phenylbutylamine (0.79 mL, 5.00 mmol), phthalic anhydride (741 mg, 5.00 mmol) and triethylamine (2.79 mL, 20.0 mmol) in toluene (50 mL) was refluxed (120 °C) with a Dean–Stark apparatus for 16 hours. The reaction mixture was allowed to cool down to room temperature and concentrated under reduced pressure. The residue was re-dissolved in EtOAc (50 mL) and washed, first, with *sat. aq.* NaHCO₃ solution (2 x 25 mL) and, then, with water (25 mL). The organic phase were combined, dried over Na₂SO₄ and the solvent was removed *in vacuo*. Purification by column chromatography on silica gel support (*gradient from* hexane *to* 20% EtOAc in hexane) gave the desired amide (1.14 g, 82% yield) as a colourless solid. The spectroscopic properties of this compound were consistent with the data available in the literature.⁶

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 7.86 (dd, J = 5.5, 3.0 Hz, 2H), 7.72 (dd, J = 5.5, 3.0 Hz, 2H), 7.35 – 7.24 (m, 2H), 7.23 – 7.13 (m, 3H), 3.74 (t, J = 7.0 Hz, 2H), 2.68 (t, J = 7.5 Hz, 2H), 1.82 – 1.62 (m, 4H).

¹³C NMR (101 MHz, CDCl₃): δ 168.4, 142.0, 133.9, 132.2, 128.4, 128.3, 125.8, 123.2, 37.8, 35.4, 28.6, 28.2.

Synthesis of 1,3-dioxoisoindolin-2-yl 4-phenylbutanoate:



This compound was prepared according to a reported literature procedure.⁷ To a stirring solution of 4-phenylbutirric acid (1.07 g, 6.50 mmol), *N*-hydroxyphthalimide (1.17 g, 7.15 mmol) and DMAP (79.0 mg, 0.65 mmol) in CH₂Cl₂ (30 mL) was added DCC (1.48 g, 7.15 mmol). The reaction was stirred at room temperature for 6 hours and then placed in a freezer at -20 °C for 2 hours observing the precipitation of the urea by-product, which was filtered and rinsed with cold Et₂O. The filtrate was concentrated under reduced pressure to provide the crude material. Purification by column chromatography on silica gel (*gradient from* hexane *to* 20% EtOAc in hexane) gave the desired ester (1.50 g, 75% yield) as a colourless solid.

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 7.96 – 7.87 (m, 2H), 7.86 – 7.77 (m, 2H), 7.37 – 7.30 (m, 2H), 7.31 – 7.20 (m, 3H), 2.80 (t, J = 7.5 Hz, 2H), 2.70 (t, J = 7.5 Hz, 2H), 2.14 (tt, J = 7.5 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃): δ 169.4, 162.0, 140.7, 134.8, 129.0, 128.6, 128.5, 126.2, 124.0, 34.6,

<u>-C NMR</u> (101 MHz, CDC1₃): 6 169.4, 162.0, 140.7, 154.8, 129.0, 128.0, 128.5, 120.2, 124.0, 54.0 30.2, 26.3.

Synthesis of methyl (S)-2-(4-isobutylphenyl)propanoate (3):



This compound was prepared according to a reported literature procedure.⁸ To a solution of (*S*)-2-(4-isobutylphenyl)propanoic acid (0.775 g, 5.00 mmol) in MeOH (50 mL) at 0 °C was added dropwise thionyl chloride (2.4 g, 20.0 mmol). Completed the addition, the reaction was warmed to room temperature and stirred overnight. The solvent was removed under reduced pressure and the resulting residue was purified by column chromatography on silica gel (*gradient from* hexane *to* 10% Et₂O in hexane) to give ester **3** (0.828 g, 90% yield) as a colourless liquid. The spectroscopic properties of this compound were consistent with the data available in the literature.⁸ $[\alpha]_D^{26} = +69.4$ (c = 0.95, CHCl₃); Lit: $[\alpha]_D^{26} = +64.6$ (c = 1.00, CHCl₃, for (*S*)-enantiomer).⁹

<u>¹H NMR</u> (400 MHz, CDCl₃): δ 7.19 (d, J = 8.0 Hz, 2H), 7.09 (d, J = 8.0 Hz, 2H), 3.70 (q, J = 7.0 Hz, 1H), 3.66 (s, 3H), 2.44 (d, J = 7.0 Hz, 2H), 1.85 (dp, J = 13.6, 7.0 Hz, 1H), 1.49 (d, J = 7.0 Hz, 3H), 0.90 (d, J = 7.0 Hz, 6H);

¹³C NMR (101 MHz, CDCl₃): δ 175.4, 140.7, 137.9, 129.5, 127.3, 52.1, 45.2, 31.9, 30.3, 22.5, 18.8.

C. Optimization Studies and Control Experiments

<u> </u>	H + toluene	catalyst (S)- A (20 mol%) TFA (40 mol%)		F ₃ C	⁻ ₃ CF₃ ≼
Ph h 1a	10 equiv.	<i>HP</i> single LED (420 nm) irradiance 45±2 mW/cm ² solvent, 35 °C, 16 h	Ph 2a		
	entry	Solvent	Yield (%) ^a	ee (%)	
	1	Cyclohexane	6	-	
	2	Benzene	28	68	
	3	Toluene	19	60	
	4	Chlorobenzene	40	68	
	5	Dichloroethane	57	83	
	6	Dichloromethane	57	83	
	7	Methyl <i>t</i> -butyl ether	-	-	
	8	Acetonitrile	-	-	
	9	Dimethylsulfoxide	-	-	
	10	Hexafluoroisopropanol	-	-	

 Table S1. Evaluation of different solvents

^{*a*} Yield of 2a determined by ¹H NMR analysis of the crude mixture using trichloroethylene as the internal standard.

Ph 1a	H + toluene 10 equiv. HP single LED (420 mm) irradiance 45±2 mW/cm ² CH ₂ Cl ₂ , 35 °C, 16 h	h Fundamental Factor	CF ₃ CF ₃ CF ₃ CF ₃ CF ₃
entry	Acid	Yield (%) ^a	ee (%)
1	Trifluoroacetic acid (40 mol%)	19	60
2	Diphenyl phosphate (40 mol%)	-	-
3	Trifluoromethansulfonic acid (40 mol%)	-	-
4	Methansulfonic acid (40 mol%)	34	78
5	Trichloroacetic acid (40 mol%)	7	-
6	Zn(CF ₃ CO ₂) ₂ (40 mol%)	5<	-
7	Zn(MeSO ₃) ₂ (40 mol%)	8	-
8	Zn(ClO ₄) ₂ (40 mol%)	10	-
9	Zn(BF4)2 (40 mol%)	5<	-
10	ZnBr ₂ (40 mol%)	5<	-
11	Zn(CF ₃ SO ₃) ₂ (40 mol%)	52	83
12	Zn(CF3SO3)2 (50 mol%)	57	83
13	Mg(CF ₃ SO ₃) ₂ (50 mol%)	40	80
14	Ca(CF ₃ SO ₃) ₂ (50 mol%)	39	78

Table S2. Screening of acid co-catalysts

^{*a*} Yield of 2a determined by ¹H NMR analysis of the crude mixture using trichloroethylene as the internal standard.



Table S3. Evaluation of toluene stoichiometry

^{*a*} Yield of **2a** determined by ¹H NMR analysis of the crude mixture using trichloroethylene as the internal standard.



Table S4. Evaluation of the stoichiometry for elaborated substrates

In most of the cases, such as for products **20-v** and **4**, the unreacted benzylic substrates can be completely recovered in high purity during the purification procedure *via* flash column chromatography.



Figure S1. Evaluating the ability of different Brønsted acids to promote the iminium ion formation. Characteristic ¹H NMR signals of the species involved (400 MHz, CDCl₃): 9.50 (d, J = 7.5 Hz, CHO of 1a), 8.73 (d, J = 11.0 Hz, CHN of iminium ion I), 4.57 (t, J = 8.0 Hz, C1-H of (S)-A), 4.43 (br. t, J = 14.0 Hz, C1-H of iminium ion I).² The spectrum on the left conerns the experiment performed with trifluoroacetic acid (TFA), while the spectrum on the right refers to the experiment performed with diphenyl phosphate (DPP) as acid co-catalyst. The presence of a doublet and broad triplet at 8.73 ppm and 4.43 ppm, respectively, is diagnostic of the iminium ion formation occurring. Accordingly, this study showes that DPP is not acidic enough to promote the condensation step, whereas TFA elicits the formation of iminium ion I.

Ph	о Н 1а	+ p-xylene 10 equiv. HP single LED (420 nm) irradiance 45±2 mW/cm ² CH ₂ Cl ₂ , 35 °C, 48 h	Find Find H	OTDS CF
	entry	Acid	Yield (%) ^a	ee (%)
	1	Zn(CF ₃ SO ₃) ₂ (50 mol%)	15	-
	2	Trifluoroacetic acid (40 mol%)	57	80
	3	Trifluoroacetic acid (60 mol%)	58	77
	4	Trifluoroacetic acid (80 mol%)	60	75
	5	Trifluoroacetic acid (100 mol%)	62	71
	6	Trichloroacetic acid (100 mol%)	62	83

 Table S5. Reaction conditions re-optimization for adduct 2v

^{*a*} Yield of 2v determined by ¹H NMR analysis of the crude mixture using trichloroethylene as the internal standard.



Table S6. Effect of (S)-B vs (S)-A on enantioselectivity

Table S7. Control Experiments



^{*a*} Yield of **2a** determined by ¹H NMR analysis of the crude mixture using trichloroethylene as the internal standard - no diagnostic signals for **2a** formation were detected.



Figure S2. Determination of the reaction kinetic profile. The kinetic profile was studied monitoring the progression in time of a benchmark reaction between cinnamaldehyde **1a** and toluene, under optimized catalytic conditions. The reaction mantained a constant level of enantioinduction (81-83% *ee*), while complete consumption of enal **1a** was observed after more than 40 hours. Accordingly, we decided to evaluate the scope of the methodology running each reaction for 48 hours.

D. Experimental Procedures

D.1 General Procedure for the photochemical C-H functionalization of toluene and derivatives



General Procedure: A 15 mL Schlenk tube was charged with a mixture of amine catalyst (*S*)-A (0.02 mmol, 20 mol%, 14 mg), acid co-catalst (0.05-0.1 mmol, depending on the substrate) and enal **1** (0.1 mmol, 1 equiv.) in CH₂Cl₂ (300 µL). The corresponding toluene derivative (1 mmol, 10 equiv.) was added to the reaction vessel. This was placed under an atmosphere of argon, cooled to -78 °C, degassed *via* vacuum evacuation (5 minutes), backfilled with argon and, ultimately, warmed to room temperature. The freeze-pump-thaw cycle was repeated four times, and then the Schlenk tube was sealed with Parafilm and placed into a 3D-printed plastic support mounted on an aluminium block fitted with a 420 nm high-power single LED ($\lambda = 420$ nm, irradiance = 45 mW/cm², as controlled by

an external power supply; the setup is detailed in Figure S3). This setup secured a reliable irradiation while keeping a distance of 1 cm between the reaction vessel and the light source. The reaction was stirred at ambient temperature for 48 hours. The solvent was removed *in vacuo* and the crude mixture was purified by column chromatography on silica gel to give the corresponding product 2 in the stated yield and enantiomeric purity. If not otherwise stated, full consumption of the limiting enal (1) was observed by ¹H NMR analysis on the crude mixture at the end of the reaction.

In the case of the reactions delivering products 2o-v and 4, it was possible to fully recover the unreacted starting material (~90% recovered yield), as the first eluting compound, after purification procedure *via* flash column chromatography.

N.B. The internal temperature of the reaction was detected to be ~ 35 °C, as measured with a thermometer. In all cases, when the photochemical reaction was run carefully controlling and mantaining the temperature at 35 °C by means of a chiller connected to the irradiation plate, it proceeded with analogous efficiency.



Figure S3. Detailed set-up and illumination system. The light source for illuminating the reaction vessel consisted in a 420 nm high-power single LED (OCU-440 UE420-X-T) purchased from OSA OPTO.^{*}

D.2 Characterization of Products 2



(*S*)-3,4-diphenylbutanal (2a). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), toluene (92.0 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2a (14.6 mg, 63% yield, 83% ee, average of two runs) as a colourless

oil that displayed spectroscopic data consistent with those reported previously.² The enantiomeric excess was determined to be 83% by HPLC analysis on a Daicel Chiralpak IC-3 column: 90:10 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm: $\tau_{major} = 12.4$ min, $\tau_{minor} = 13.5$ min. $[\alpha]_D^{26} = -47.5$ (c = 0.44, CHCl₃, 83% ee); Lit: $[\alpha]_D^{26} = -45.8$ (c = 0.49, CHCl₃, 88% ee for (*S*)-enantiomer).² The

^{*}For more information on the LED used, visit https://www.osa-opto.com/tl_files/osa_opto/inhalte/files/datasheets/ocl-440/440_UE420.pdf

absolute configuration for compound **2a** was determined in comparison to the data available in the literature.² Accordingly, the absolute configuration for all of the products **2** was assigned likewise. ¹<u>H NMR</u> (500 MHz, CDCl₃): δ 9.59 (t, *J* = 2.0 Hz, 1H), 7.31 – 7.14 (m, 8H), 7.08 – 7.04 (app d, *J* = 5.0 Hz 2H), 3.49 (p, *J* = 7.5 Hz, 1H), 2.96 (dd, *J* = 13.5, 7.1 Hz, 1H), 2.88 (dd, *J* = 13.5, 7.9 Hz, 1H), 2.81 – 2.70 (m, 2H); ¹³<u>C NMR</u> (125.8 MHz, CDCl₃): δ 201.7, 143.4, 139.4, 129.3, 128.7, 128.5, 127.7, 126.9, 126.5, 49.1, 43.5, 42.2.



1 mmol scale reaction. The model reaction was repeated on a more synthetically useful scale using three high power single LEDs and a 50 mL round-bottom Schlenk flask to increase the surface-area-to-volume ratio, which secured a more efficient irradiation of the reaction mixture (see pictures to the left). To a 50 mL round-bottom Schlenk flask containing aminocatalyst (*S*)-**A** (0.2 mmol, 20 mol%, 141 mg), zinc(II)

triflate (0,5 mmol, 181 mg) and cynnamaldehyde **1a** (1 mmol, 1 equiv., 132 mg) in dichloromethane (3 mL) was added toluene (10 mmol, 921 mg). The reaction vessel was placed under an atmosphere of argon, cooled to -78 °C, degassed via vacuum evacuation (5 min), backfilled with argon and, ultimately, warmed to room temperature. The freeze-pump-thaw cycle was repeated five times, and then three 420 nm high-power single LEDs ($\lambda = 420$ nm, irradiance = 45 mW/cm², as controlled by an external power supply) were placed at 1 cm distance from the sides and the bottom of the Schlenk flask. The reaction was stirred under visible light irradiation at ambient temperature (~35 °C) for 60 hours. The solvent was removed under reduced pressure and the crude mixture was purified by column chromatography on silica gel (hexane/diethyl ether 92:8) to afford product **2a** as a pale yellow oil (111 mg, 51% yield, 83% ee).



(*R*)-3,4-diphenylbutanal (*ent*-2a). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*R*)-A (14.1 mg, 20 mol%), toluene (92.0 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product *ent*-2a (14.6 mg, 63% yield, 82% ee, average of two runs) as a

colourless oil that displayed spectroscopic data consistent with those reported previously.² The enantiomeric excess was determined to be 82% by HPLC analysis on a Daicel Chiralpak IC-3 column: 90:10 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm: $\tau_{major} = 13.5$ min, $\tau_{minor} = 12.4$ min. [α]_D²⁶ = +55.3 (c = 0.067, CHCl₃, 82% ee).



(*S*)-3-(4-fluorophenyl)-4-phenylbutanal (2b). Prepared according to the general procedure using enal 1b (15.0 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), toluene (92.0 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2b (17.0 mg, 70% yield, 80% ee, average of two runs) as a colourless oil that displayed spectroscopic

data consistent with those reported previously.² The enantiomeric excess was determined to be 80% by UPC² analysis on a Daicel Chiralpak ID-3 column: gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 275$ nm: $\tau_{major} = 2.49$ min, $\tau_{minor} = 2.57$ min. $[\alpha]_D{}^{26} = -38.4$ (c = 0.71, CHCl₃, 80% ee); Lit: $[\alpha]_D{}^{26} = -45.3$ (c = 0.69, CHCl₃, 85% ee for (*S*)-enantiomer).² ¹<u>H NMR</u> (500 MHz, CDCl₃): δ 9.61 (t, *J* = 2.0 Hz, 1H), 7.25 – 7.21 (m, 2H), 7.20 – 7.15 (m, 1H), 7.12 – 7.07 (m, 2H), 7.04 – 7.01 (m, 2H), 6.96 (t, *J* = 9.0 Hz, 2H), 3.49 (p, *J* = 7.5 Hz, 1H), 2.89 (d, *J* = 7.5 Hz, 2H), 2.75 (dd, *J* = 7.5, 2.0 Hz, 2H);

¹³<u>C NMR</u> (125.8 MHz, CDCl₃): δ 201.7, 161.7 (d, ¹*J*_{*C*-*F*} = 245.0 Hz), 139.1, 139.0 (d, ⁴*J*_{*C*-*F*} = 3.0 Hz), 129.3, 129.1 (d, ³*J*_{*C*-*F*} = 8.0 Hz), 128.5, 126.6, 115.5 (d, ²*J*_{*C*-*F*} = 21.0 Hz), 49.3, 43.5, 41.3.



(*S*)-3-(4-Chlorophenyl)-4-phenylbutanal (2c). Prepared according to the general procedure using enal 1c (16.7 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), toluene (92.0 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2c (15.3 mg, 59% yield, 67% ee, average of two runs) as a colourless oil that displayed

spectroscopic data consistent with those reported previously.² The enantiomeric excess was determined to be 67% by UPC² analysis on a Daicel Chiralpak IE-3 column: gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm: $\tau_{major} = 4.2$ min, $\tau_{minor} = 4.41$ min. [α]_D²⁶ = -46.5 (c = 0.49, CHCl₃, 67% ee); Lit: [α]_D²⁶ = -56.8 (c = 0.69, CHCl₃, 86% ee for (*S*)-enantiomer).²

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.61 (t, J = 2.0 Hz, 1H), 7.32 – 7.11 (m, 5H), 7.10 – 7.00 (m, 4H), 3.48 (p, J = 7.5 Hz, 1H), 2.89 (d, J = 7.5 Hz, 2H), 2.75 (dd, J = 7.5, 2 Hz, 2H); ¹³<u>C NMR</u> (101 MHz, CDCl₃): δ 201.1, 141.8, 139.0, 132.6, 129.3, 128.9, 128.6, 126.6, 49.1, 43.3, 41.4.



(*S*)-4-phenyl-3-(p-tolyl)butanal (2d). Prepared according to the general procedure using enal 1d (14.6 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), toluene (92 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2d (11.3 mg, 48% yield, 60% ee, average of two runs) as a colourless oil that displayed

spectroscopic data consistent with those reported previously.² The enantiomeric excess was determined to be 60% by UPC² analysis on a Daicel Chiralpak ID-3 column: gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm: $\tau_{major} = 3.01$ min, $\tau_{minor} = 3.28$ min. $[\alpha]_D^{26} = -19.3$ (c = 0.25, CHCl₃, 60% ee); Lit: $[\alpha]_D^{26} = -42.5$ (c = 0.50, CHCl₃, 78% ee for (*S*)-enantiomer).²

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.57 (t, *J* = 2.0 Hz, 1H), 7.28 – 7.15 (m, 3H), 7.11 – 7.02 (m 6H), 3.45 (p, *J* = 7.5 Hz, 1H), 2.95 (dd, *J* = 13.5, 7.0 Hz, 1H), 2.85 (dd, *J* = 13.5, 8.0 Hz, 1H), 2.77 – 2.66 (m, 2H), 2.31 (s, 3H);

¹³C NMR (101 MHz, CDCl₃): δ 201.9, 140.3, 139.6, 136.4, 129.4, 128.5, 127.5, 126.5, 49.1, 43.5, 41.8, 21,2.



(*S*)-4-phenyl-3-(o-tolyl)butanal (2e). Prepared according to the general procedure using enal 1e (14.6 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), toluene (92 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2e (11.5 mg, 48% yield, 72% ee, average of two runs) as a colourless oil that displayed spectroscopic data consistent with those reported

previously.² The enantiomeric excess was determined to be 72% by UPC² analysis on a Daicel Chiralpak IE-3 column: gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm: $\tau_{major} = 3.58$ min, $\tau_{minor} = 3.75$ min. $[\alpha]_D^{26} = -26.4$ (c = 0.37, CHCl₃, 72% ee).

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.56 (t, J = 2.0 Hz, 1H), 7.25 – 7.14 (m, 5H), 7.12 – 7.07 (m, 2H), 7.04 (d, J = 8.0 Hz, 2H) 3.76 (p, J = 7.5 Hz, 1H), 2.91 (dd, J = 13.5, 7.0 Hz, 1H), 2.85 – 2.70 (m, 3H), 2.21 (s, 3H);

¹³<u>C NMR</u> (101 MHz, CDCl₃): δ 201.7, 141.6, 139.5, 136.0, 130.6, 129.3, 128.4, 126.51, 126.0, 49.0, 43.2, 36.9, 19,7.



(S)-4-phenyl-3-(4-(trimethylsilyl)phenyl)butanal (2f). Prepared according to the general procedure using enal 1f (20.4 mg, 0.1 mmol), aminocatalyst (S)-A (14.1 mg, 20 mol%), toluene (92 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2f (14.8 mg, 50% yield, 70% ee, average of two runs) as an off-white solid. The

enantiomeric excess was determined to be 70% by UPC² analysis on a Daicel Chiralpak IE-3 column: gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 2 mL/min, $\lambda = 210$ nm: $\tau_{major} = 3.97$ min, $\tau_{minor} = 4.79$ min. $[\alpha]_D^{26} = -19.8$ (c = 0.14, CHCl₃, 70% ee).

 $\frac{1 \text{H NMR}}{7.24 - 7.18} (400 \text{ MHz, CDCl}_3): \delta 9.59 (t, J = 2.0 \text{ Hz}, 1\text{H}), 7.47 (d, J = 8.0 \text{ Hz}, 2\text{H}), 7.31 - 7.25 (m, 2\text{H}), 7.24 - 7.18 (m, 3\text{H}), 7.15 - 7.10 (m, 2\text{H}), 3.51 (tt, J = 8.5, 6.5 \text{ Hz}, 1\text{H}), 3.03 (dd, J = 13.5, 6.5 \text{ Hz}, 1\text{H}), 2.88 (dd, J = 13.5, 8.5 \text{ Hz}, 1\text{H}), 2.82 - 2.69 (m, 2\text{H}), 0.28 (s, 9\text{H}).$

¹³<u>C NMR</u> (101 MHz, CDCl₃): δ 201.6, 143.9, 139.3, 138.7, 133.7, 129.2, 128.4, 126.9, 126.4, 48.7, 43.2, 41.9, -1.10.

HRMS (ESI) Exact mass calculated for C₁₉H₂₄NaOSi [M+Na]⁺: 319.1489, found: 319.1488.



(S)-4-phenyl-3-(3-(trifluoromethyl)phenyl)butanal (2g). Prepared according to the general procedure using enal 1g (20.0 mg, 0.1 mmol), aminocatalyst (S)-A (14.1 mg, 20 mol%), toluene (92 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2g (9.6 mg, 33% yield, 84% ee, average of two runs) as a pale yellow oil. The

enantiomeric excess was determined, upon sodium borohydride reduction of the isolated aldehyde to

afford the corresponding alcohol, to be 84% by UPC² analysis on a Daicel Chiralpak IE-3 column: gradient CO₂/MeOH from 100% CO₂ to 80:20 over 4 minutes, curve 6, flow rate 2 mL/min, $\lambda = 215$ nm: $\tau_{major} = 3.97$ min, $\tau_{minor} = 4.06$ min. [α]_D²⁶ = -25.8 (c = 0.21, CHCl₃, 84% ee).

 $\frac{1}{H}$ NMR (400 MHz, CDCl₃): δ 9.64 (t, *J* = 1.5 Hz, 1H), 7.48 (d, *J* = 7.5 Hz, 1H), 7.44 – 7.37 (m, 2H), 7.34 (d, *J* = 8.0 Hz, 1H), 7.28 – 7.17 (m, 3H), 7.07 – 7.01 (m, 2H), 3.60 (tt, *J* = 7.5 Hz, 1H), 2.94 (d, *J* = 7.5 Hz, 2H), 2.83 (dd, *J* = 7.5, 1.5 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃): δ 200.5, 144.3, 138.6, 131.13 (q, ${}^{4}J_{C-F} = 1.0$ Hz), 130.9 (q, ${}^{2}J_{C-F} = 32.0$ Hz), 129.2, 129.0, 128.4, 126.6, 124.2 (q, ${}^{3}J_{C-F} = 4.0$ Hz), 124.1 (q, ${}^{1}J_{C-F} = 272.5$ Hz), 123.6 (q, ${}^{3}J_{C-F} = 4.0$ Hz), 48.7, 43.0, 41.6.

¹⁹F NMR (376 MHz, CDCl₃): δ -62.70 (s, 3F).

<u>HRMS (ESI)</u> Exact mass calculated for $C_{18}H_{19}F_3NaO_2$ [M+CH₃OH+Na]⁺: 347.1229, found: 347.1245.



(*S*)-4-(4-fluorophenyl)-3-phenylbutanal (2h). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 4-fluorotoluene (110 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2h (16.2

mg, 67% yield, 80% ee, average of two runs) as a pale yellow oil that displayed spectroscopic data consistent with those reported previously.² The enantiomeric excess was determined, upon sodium borohydride reduction of the isolated aldehyde to afford the corresponding alcohol, to be 80% by HPLC analysis on a Daicel Chiralpak IC-3 column: 90:10 hexane/*i*-PrOH, flow rate 0.5 mL/min, $\lambda = 215$ nm. $\tau_{Major} = 16.0$ min, $\tau_{minor} = 15.1$ min. $[\alpha]_D^{26} = -49.1$ (c = 0.18, CHCl₃); Lit: $[\alpha]_D^{26} = -46.0$ (c = 0.63, CHCl₃, 92% ee for (*S*)-enantiomer).²

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.65 (t, J = 2.0 Hz, 1H), 7.33 – 7.26 (m, 2H), 7.26 – 7.19 (m, 1H), 7.18 – 7.11 (m, 2H), 7.03 – 6.96 (m, 2H), 6.96 – 6.88 (m, 2H), 3.47 (tt, J = 7.5 Hz, 1H), 2.92 (d, J = 7.5 Hz, 2H), 2.85 – 2.72 (m, 2H).

¹³C NMR (101 MHz, CDCl₃): δ 201.3, 161.5 (d, ¹*J*_{*C*-*F*} = 244.5 Hz), 142.8, 134.9 (d, ⁴*J*_{*C*-*F*} = 3.5 Hz), 130.6 (d, ³*J*_{*C*-*F*} = 8.0 Hz), 128.6, 127.5, 126.8, 115.1 (d, ²*J*_{*C*-*F*} = 21.0 Hz), 49.0, 42.4, 42.0. ¹⁹F NMR (376 MHz, CDCl₃): δ -116.92 (tt, ³*J*_{*H*-*F*} = 8.5 Hz, ⁴*J*_{*H*-*F*} = 5.5 Hz, 1F).



(*S*)-3-phenyl-4-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-yl)phenyl)butanal (2i). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 4,4,5,5-tetramethyl-2-(*p*tolyl)-1,3,2-dioxaborolane (218 mg, 1 mmol), zinc(II) triflate (18.2 mg, 0.05 mmol) and dichloromethane (300 μL). The crude mixture was purified by flash

column chromatography (hextane/diethyl ether 92:8) to afford product **2i** (19.0 mg, 54% yield, 80% ee, average of two runs) as a pale yellow oil. The enantiomeric excess was determined to be 80% by UPC² analysis on a Daicel Chiralpak IE-3 column: gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 2 mL/min, $\lambda = 229$ nm: $\tau_{major} = 1.74$ min, $\tau_{minor} = 2.00$ min. [α]_D²⁶ = -29.7 (c= 0.37, CHCl₃, 80% ee).

 $\frac{^{1}\text{H NMR}}{^{7.22} - 7.13} (\text{m}, 3\text{H}), 7.06 (\text{d}, J = 7.9 \text{ Hz}, 2\text{H}), 7.67 (\text{d}, J = 7.9 \text{ Hz}, 1\text{H}), 7.30 - 7.24 (\text{m}, 3\text{H}), 7.22 - 7.13 (\text{m}, 3\text{H}), 7.06 (\text{d}, J = 7.9 \text{ Hz}, 2\text{H}), 3.50 (\text{p}, J = 7.5 \text{ Hz}, 1\text{H}), 2.93 (\text{qd}, J = 13.5, 7.5 \text{ Hz}, 2\text{H}), 2.82 - 2.67 (\text{m}, 2\text{H}), 1.33 (\text{s}, 1\text{H});$

¹³C NMR (101 MHz, CDCl₃) δ 201.6, 143.2, 142.7, 135.0, 128.8, 128.8, 128.7, 127.7, 126.9, 83.9, 49.1, 43.6, 42.0, 29.9, 25.0; ¹¹B NMR (47 MHz, CDCl₃) δ 30.8; HRMS (ESI) Exact mass calculated for C₂₂H₂₇NaO₃B [M+Na]⁺: 372.1982, found: 372.1973.

H Ph SiMe₃

(*S*)-3-phenyl-4-(4-(trimethylsilyl)phenyl)butanal (2j). Prepared according to the general procedure using enal **1a** (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), trimethyl(*p*-tolyl)silane (164 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl

ether 92:8) to afford product **2j** (16.5 mg, 56% yield, 80% ee, average of two runs) as a colourless oil. The enantiomeric excess was determined, upon sodium borohydride reduction of the isolated aldehyde to afford the corresponding alcohol, to be 80% by HPLC analysis on a Daicel Chiralpak IC-3 column: 90:10 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm: $\tau_{Major} = 10.0$ min, $\tau_{minor} = 11.0$ min. $[\alpha]_D^{26} = -40.1$ (c = 0.67, CHCl₃, 80% ee).

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.58 (t, J = 2.0 Hz, 1H), 7.40 (d, J = 7.9 Hz, 2H), 7.32 – 7.27 (m, 2H), 7.24 – 7.17 (m, 3H), 7.07 (d, J = 7.9 Hz, 2H), 3.55-3.44 (m, 1H), 2.99 (dd, J = 13.5, 6.7 Hz, 1H), 2.85 (dd, J = 13.6, 8.3 Hz, 1H), 2.80 – 2.67 (m, 2H), 0.24 (s, 9H);

¹³C NMR (101 MHz, CDCl₃): δ 201.8, 143.5, 140.0, 138.3, 133.6, 128.8 (2 signals), 127.7, 126.9, 49.0, 43.4, 42.0, -0.9.

HRMS (ESI) Exact mass calculated for C₁₉H₂₄NaOSi [M+Na]⁺: 319.1484, found: 319.1489.



(S)-4-methyl-3,4-diphenylpentanal (2k). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (S)-A (14.1 mg, 20 mol%), cumene (120 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2k (9.6 mg, 38% yield,

53% ee, average of two runs) as a pale yellow oil that displayed spectroscopic data consistent with those reported previously.² The enantiomeric excess was determined to be 53% by HPLC analysis on a Daicel Chiralpak IC-3 column: 90:10 hexane/*i*-PrOH, flow rate 1.0 mL/min, $\lambda = 215$ nm: $\tau_{Major} = 8.4$ min, $\tau_{minor} = 12.2$ min. $[\alpha]_D^{26} = -35.2$ (c = 0.11, CHCl₃, 53% ee); Lit: $[\alpha]_D^{26} = -33.0$ (c = 0.37, CHCl₃, 71% ee for (*S*)-enantiomer).²

 $\frac{^{1}\text{H NMR}}{^{4}\text{H NMR}}$ (400 MHz, CDCl₃): δ 9.34 (dd, J = 3.0, 1.5 Hz, 1H), 7.41 – 7.31 (m, 4H), 7.31 – 7.21 (m, 4H), 7.15 – 7.09 (m, 2H), 3.53 (dd, J = 11.0, 4.0 Hz, 1H), 2.81 (ddd, J = 16.5, 11.0, 3.0 Hz, 1H), 2.46 (ddd, J = 16.5, 4.0, 1.5 Hz, 1H), 1.33 (s, 3H), 1.24 (s, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 202.0, 147.9, 140.2, 130.0, 128.3, 128.0, 127.0, 126.6, 126.3, 51.2, 45.0, 41.2, 28.8, 23.2.



(3*R*)-3,4-diphenylpentanal (2l). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), ethylbenzene (106 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1.5:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column chromatography (hexane/diethyl ether 92:8) to

afford product 21 (15.9 mg, 67% yield, average of two runs) as an off-white solid that displayed

spectroscopic data consistent with those reported previously.² The enantiomeric excess of both diastereoisomers was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, by HPLC analysis on a Daicel Chiralpak IC-3 column, 97:3 hexane/*i*-PrOH, flow rate 1.2 mL/min, $\lambda = 215$ nm. *Major diastereomer:* $\tau_{Major} = 24.9$ min, $\tau_{minor} = 20.4$ min. *Minor diastereomer:* $\tau_{Major} = 18.0$ min, $\tau_{minor} = 19.1$ min. $[\alpha]_D^{26} = -19.4$ (c = 0.087, CHCl₃, 1.5:1 *d.r.*, 80% *eemajor*, 77% *eeminor*); Lit: $[\alpha]_D^{26} = -40.4$ (c = 0.42, CHCl₃, 1.2:1 *d.r.*, 92% *eemajor*, 92% *eeminor* for (3*R*)-enantiomer).²

Spectroscopic data for major diastereoisomer:

 $\frac{1}{\text{H NMR}} (400 \text{ MHz, CDCl}_3) \delta 9.38 (dd, J = 2.5, 1.5 \text{ Hz}, 1\text{H}), 7.43 - 7.12 (m, 8\text{H}), 7.05 - 6.97 (m, 2\text{H}), 3.33 (td, J = 10.5, 4.5 \text{ Hz}, 1\text{H}), 2.92 (dq, J = 10.5, 7.0 \text{ Hz}, 1\text{H}, A), 2.65 (ddd, J = 16.5, 10.5, 2.5 \text{ Hz}, 1\text{H}), 2.49 (ddd, J = 16.5, 4.5, 1.5 \text{ Hz}, 1\text{H}), 1.07 (d, J = 7.0 \text{ Hz}, 3\text{H}).$

¹³C NMR (125.8 MHz, CDCl₃) δ 201.6, 145.1, 142.6, 128.7, 128.7, 128.1, 127.6, 126.8, 126.7, 49.0, 47.5, 46.0, 20.5.

Spectroscopic data for minor diastereoisomer:

¹<u>H NMR</u> (400 MHz, CDCl₃) δ 9.60 (t, *J* = 2.0 Hz, 1H), 7.43 – 7.12 (m, 8H), 7.05 – 6.97 (m, 2H), 3.49 (td, *J* = 7.5, 7.5 Hz, 1H), 3.09 (dq, *J* = 7.0, 7.0 Hz, 1H), 2.83 (dd, *J* = 7.5, 2.0 Hz, 2H), 1.31 (d, *J* = 7.0 Hz, 3H).

¹³C NMR (125.8 MHz, CDCl₃) δ 202.0, 143.8, 141.5, 128.5, 128.1 (2 *signals*), 128.0, 126.6, 126.3, 46.6, 46.1, 45.0, 18.0.



(*3R*)-4-cyclopropyl-3,4-diphenylbutanal (2m). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), (cyclopropylmethyl)benzene (132 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column

chromatography (hexane/diethyl ether 92:8) to afford product **2m** (10.3 mg, 39% yield, average of two runs) as an off-white solid. The enantiomeric excess of both diastereoisomers was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, by UPC² analysis on a Daicel Chiralpak CEL-1 column: gradient CO₂/*i*-PrOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm. *Diastereomer A:* $\tau_{Major} = 4.07$ min, $\tau_{minor} = 3.96$ min. *Diastereomer B:* $\tau_{Major} = 4.42$ min, $\tau_{minor} = 4.13$ min. [α]_D²⁶ = -15.4 (c = 0.15, CHCl₃, 1:1 *d.r.*, 82% *ee_A*, 85% *ee_B*).

Spectroscopic data for diastereoisomer A:

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.69 (t, J = 2.0 Hz, 1H), 7.38 – 7.11 (m, 10H), 3.71 (ddd, J = 9.0, 8.0, 6.0 Hz, 1H), 3.10 (ddd, J = 17.0, 6.0, 2.0 Hz, 1H), 2.84 (ddd, J = 17.0, 9.0, 2.0 Hz, 1H), 2.01 (dd, J = 10.5, 8.0 Hz, 1H), 1.07 (dddd, J = 13.0, 10.5, 8.0, 5.0 Hz, 1H), 0.76 (dddd, J = 9.0, 7.5, 6.0, 4.5 Hz, 1H), 0.44 (dddd, J = 9.0, 8.0, 5.5, 4.5 Hz, 1H), 0.34 (ddd, J = 10.5, 9.0, 5.0 Hz, 1H), 0.03 (ddd, J = 9.0, 5.5, 4.5 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃, aromatic C-H signals were not assigned and reported for both diastereoisomers): δ 201.7, 142.8, 141.5, 128.6 (C_{Ar}-H), 128.5 (2 signals, C_{Ar}-H), 128.4 (C_{Ar}-H), 128.3 (C_{Ar}-H), 128.2 (C_{Ar}-H), 128.0 (C_{Ar}-H), 127.8 (C_{Ar}-H), 56.9, 47.4, 46.3, 14.9, 7.7, 1.0. Spectroscopic data for diastereoisomer B:

 $\frac{1}{11}$ NMR (400 MHz, CDCl₃): δ 9.46 (dd, J = 2.5, 1.5 Hz, 1H), 7.38 – 7.11 (m, 6H), 7.00 – 6.96 (m, 2H), 6.95 – 6.92 (m, 2H), 3.62 (ddd, J = 9.5, 9.5, 5.0 Hz, 1H), 2.75 (ddd, J = 17.0, 9.5, 2.5 Hz, 1H),

2.61 (ddd, *J* = 17.0, 5.0, 1.5 Hz, 1H), 2.09 (dd, *J* = 9.5, 9.5 Hz, 1H), 0.98 – 0.88 (m, 1H), 0.31 – 0.21 (m, 2H), -0.08 – -0.15 (m, 1H), -0.21 – -0.27 (m, 1H).

¹³C NMR (101 MHz, CDCl₃, aromatic C-H signals were not assigned and reported for both diastereoisomers): δ 202.1, 143.4, 142.3, 128.6 (C_{Ar}-H), 128.5 (2 signals, C_{Ar}-H), 128.4 (C_{Ar}-H), 128.3 (C_{Ar}-H), 128.2 (C_{Ar}-H), 128.0 (C_{Ar}-H), 127.8 (C_{Ar}-H), 56.7, 48.2, 46.7, 15.3, 7.2, 2.8. HRMS (ESI) Exact mass calculated for C₁₉H₂₀NaO [M+Na]⁺: 287.1406, found: 287.1403.



(*3R*)-3,4-diphenylhept-6-enal (2n). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 4-phenyl-1-butene (66.1 mg, 0.5 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and CH₂Cl₂ (300 μ L). The d.r. was determined to be 2.5:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column chromatography (hexane/diethyl

ether 92:8) to afford product **2n** (6.8 mg, 26% yield, average of two runs) as a colourless solid that displayed spectroscopic data consistent with those reported previously.² The enantiomeric excess was determined by HPLC analysis on a Daicel Chiralpak IC-3 column: 95:5 hexane/*i*-PrOH, flow rate 0.5 mL/min, $\lambda = 215$ nm. *Major diastereomer*: $\tau_{Major} = 19.9$ min, $\tau_{minor} = 26.4$ min. *Minor diastereomer*: $\tau_{Major} = 15.9$ min, $\tau_{minor} = 16.7$ min. $[\alpha]_D{}^{26} = -1.3$ (c = 0.11, CHCl₃, 4:1 *d.r.*, 86% *ee_{major}*, 79% *ee_{minor}*); Lit: $[\alpha]_D{}^{26} = -1.7$ (c = 0.48, CHCl₃, 1.5:1 *d.r.*, 92% *ee_{major}*, 90% *ee_{minor}* for (3*R*)-enantiomer).²

Spectroscopic data for major diastereoisomer:

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.36 (dd, J = 2.5, 1.5 Hz, 1H), 7.41 – 7.16 (m, 10H), 5.43 (ddt, J = 17.0, 10.5, 7.0 Hz, 1H), 4.84 – 4.69 (m, 2H), 3.42 (td, J = 10.5, 4.5 Hz, 1H), 2.93 – 2.75 (m, 1H), 2.62 (ddd, J = 17.0, 10.5, 2.5 Hz, 1H), 2.57 – 2.37 (m, 1H), 2.27 – 2.14 (m, 2H).

¹³C NMR (101 MHz, CDCl₃): δ 201.5, 142.7, 142.4, 136.5, 128.9, 128.8, 128.6, 128.3, 127.1, 127.0, 116.2, 52.1, 49.0, 46.3, 38.5.

Spectroscopic data for minor diastereoisomer:

<u>¹H NMR</u> (400 MHz, CDCl₃): δ 9.64 (t, J = 2.0 Hz, 1H), 7.41 – 7.16 (m, 8H), 6.95 – 6.85 (m, 2H), 5.64 (ddt, J = 17.0, 10.0, 6.5 Hz, 1H), 5.08 – 4.92 (m, 2H), 3.61 (dt, J = 9.0, 6.0 Hz, 1H), 3.01 (dt, J = 9.6, 6.0 Hz, 1H), 2.93 – 2.75 (m, 2H), 2.57 – 2.37 (m, 2H).

¹³<u>C NMR</u> (101 MHz, CDCl₃): δ 202.0, 142.8, 142.4, 136.8, 129.4, 129.1, 128.1, 127.9, 126.8, 126.6, 116.7, 50.6, 47.2, 44.5, 37.0.



(*3R*)-3,4,5-triphenylpentanal (20). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 1,2-diphenylethane (91 mg, 0.5 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1.8:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column chromatography (hexane/diethyl

ether 97:3) to afford product **20** (16.7 mg, 53% yield, average of two runs) as an off-white solid. The enantiomeric excess of both diastereoisomers was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, by UPC² analysis on a Daicel Chiralpak CEL-1 column: gradient CO₂/MeOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 210$ nm. *Major diastereomer:* $\tau_{Major} = 4.13$ min, $\tau_{minor} = 4.00$ min. *Minor diastereomer:* $\tau_{Major} = 4.41$ min, $\tau_{minor} = 4.27$ min. [α]_D²⁶ = -20.4 (c = 0.11, CHCl₃, 1.8:1 *d.r.*, 85% *ee_{major}*, 83% *ee_{minor}*).

Spectroscopic data for major diastereoisomer:

 $\frac{1 \text{H NMR}}{1400 \text{ MHz}, \text{CDCl}_3): \delta 9.39 \text{ (dd}, J = 2.5, 1.5 \text{ Hz}, 1\text{H}), 7.46 - 6.82 \text{ (m, 13H)}, 6.70 \text{ (dd}, J = 7.5, 2.0 \text{ Hz}, 2\text{H}), 3.54 \text{ (td}, J = 10.5, 4.5 \text{ Hz}, 1\text{H}), 3.15 - 2.99 \text{ (m, 1H)}, 2.97 - 2.87 \text{ (m, 1H)}, 2.87 - 2.74 \text{ (m, 1H)}, 2.71 - 2.54 \text{ (m, 1H)}, 2.49 \text{ (ddd}, J = 17.0, 4.5, 1.5 \text{ Hz}, 1\text{H}).$

¹³C NMR (101 MHz, CDCl₃): δ 201.3, 142.8, 142.0, 140.2, 128.9, 128.8, 128.6, 128.5, 128.2, 127.8, 127.1, 126.8, 125.6, 54.5, 49.0, 46.2, 40.7.

Spectroscopic data for **minor diastereoisomer**:

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.62 (t, J = 2.0 Hz, 1H), 7.46 – 6.82 (m, 15H), 3.65 (dt, J = 9.0, 6.0 Hz, 1H), 3.30 (dt, J = 9.0, 6.0 Hz, 1H), 3.15 – 2.99 (m, 1H), 2.87 – 2.74 (m, 1H), 2.71 – 2.54 (m, 2H). ¹³<u>C NMR</u> (101 MHz, CDCl₃): δ 201.8, 140.5, 140.4, 140.0, 129.3, 129.1, 129.0, 128.2, 128.0, 127.7, 126.7, 126.5, 125.9, 52.2, 47.4, 44.3, 38.9.

HRMS (ESI) Exact mass calculated for C₂₄H₂₆NaO₂ [M+CH₃OH+Na]⁺: 369.1825, found: 369.1841.



(4*R*)-6-oxo-3,4-diphenylhexyl acetate (2p). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 3-phenylpropyl acetate (89 mg, 0.5 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1.6:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column

chromatography (*gradient from* hexane/diethyl ether 90:10 *to* hexane/diethyl ether 50:50) to afford the *major diastereoisomer* of product **2p** (10.4 mg) as a pale yellow oil. Continued elution delivered the *minor diastereoisomer* of **2p** (6.9 mg) as a yellow oil (55% overall yield, average of two runs). The enantiomeric excess of both diastereoisomers was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, by HPLC analysis on a Daicel Chiralpak IC-3 column: 90:10 hexane/*i*-PrOH, flow rate 1.2 mL/min, $\lambda = 215$ nm. *Major diastereomer:* $\tau_{Major} = 26.4$ min, $\tau_{minor} = 32.0$ min. *Minor diastereomer:* $\tau_{Major} = 26.7$ min, $\tau_{minor} = 24.8$ min.

Characterization for major diastereoisomer:

 $[\alpha]_D^{26} = -8.8 \ (c = 0.15, CHCl_3, 83\% \ ee_{major}).$

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.35 (dd, J = 2.5, 1.5 Hz, 1H), 7.41 – 7.33 (m, 4H), 7.32 – 7.22 (m, 6H), 3.78 (ddd, J = 11.0, 7.0, 5.5 Hz, 1H), 3.64 (dt, J = 11.0, 7.5 Hz, 1H), 3.39 (ddd, J = 10.5, 10.5, 4.5 Hz, 1H), 2.89 (ddd, J = 10.5, 10.5, 5.0 Hz, 1H), 2.62 (ddd, J = 17.0, 10.5, 2.5 Hz, 1H), 2.44 (ddd, J = 17.0, 4.5, 1.5 Hz, 1H), 1.90 (s, 3H), 1.82 – 1.70 (m, 2H).

¹³C NMR (101 MHz, CDCl₃): δ 201.1, 170.8, 142.3, 141.8, 129.0, 128.9, 128.2, 128.1, 127.2, 127.1, 62.8, 49.0, 48.7, 46.6, 32.8, 20.8.

<u>HRMS (ESI)</u> Exact mass calculated for $C_{20}H_{22}NaO_3$ [M+Na]⁺: 333.1461, found: 333.1461.

Characterization for **minor diastereoisomer**:

 $[\alpha]_D^{26} = +6.2 \ (c = 0.26, CHCl_3, 78\% \ ee_{minor}).$

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.64 (t, J = 2.0 Hz, 1H), 7.24 – 7.14 (m, 6H), 6.95 – 6.91 (m, 2H), 6.91 – 6.86 (m, 2H), 3.96 (ddd, J = 11.0, 7.5, 5.0 Hz, 1H), 3.81 (dt, J = 11.0, 7.5 Hz, 1H), 3.55 (ddd, J = 9.0, 6.5, 6.5 Hz, 1H), 3.03 (ddd, J = 11.0, 6.5, 4.0 Hz, 1H), 2.88 (ddd, J = 17.0, 6.5, 2.0 Hz, 1H), 2.79 (ddd, J = 17.0, 9.0, 2.0 Hz, 1H), 2.19 – 2.06 (m, 1H), 2.05 – 1.90 (m, 4H).

¹³C NMR (101 MHz, CDCl₃): δ 201.5, 170.9, 140.5, 139.9, 128.9, 128.8, 128.1, 128.0, 126.8, 126.8, 62.9, 47.5, 46.9, 45.1, 31.6, 20.9.

HRMS (ESI) Exact mass calculated for C₂₀H₂₂NaO₃ [M+Na]⁺: 333.1461, found: 333.1473.



(*R*)-6-chloro-3,4-diphenylhexanal (2q). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), (3-chloropropyl)benzene (77 mg, 0.5 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1.4:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column

chromatography (hexane/diethyl ether 97:3) to afford product **2q** (14.5 mg, 51% yield, average of two runs) as an off-white solid. The enantiomeric excess of both diastereoisomers was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, by UPC² analysis on a Daicel Chiralpak CEL-1 column: isocratic CO₂/*i*-90:10, flow rate 3 mL/min, $\lambda = 220$ nm. *Major diastereomer*: $\tau_{Major} = 3.89$ min, $\tau_{minor} = 3.01$ min. *Minor diastereomer*: $\tau_{Major} = 3.36$ min, $\tau_{minor} = 2.82$ min. [α]_D²⁶ = -64.7 (c = 0.43, CHCl₃, 1.4:1 *d.r.*, 77% *eemajor*, 76% *eeminor*). *Spectroscopic data for major diastereoisomer*:

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.63 (t, *J* = 2.0 Hz, 1H), 7.42 – 7.35 (m, 2H), 7.33 – 7.25 (m, 5H), 7.24 – 7.15 (m, 2H), 6.92 (ddd, *J* = 14.0, 8.0, 2.0 Hz, 1H), 3.47 – 3.37 (m, 1H), 3.30 – 3.13 (m, 1H), 3.12 – 2.97 (m, 2H), 2.64 (ddd, *J* = 17.0, 10.0, 2.5 Hz, 1H), 2.44 (ddd, *J* = 17.0, 4.5, 1.5 Hz, 1H), 1.93 – 1.77 (m, 2H).

¹³C NMR (101 MHz, CDCl₃): δ 201.2, 142.3, 141.4, 129.2, 129.1, 128.2, 128.2, 127.5, 127.3, 49.4, 48.8, 46.6, 43.2, 36.9.

Spectroscopic data for minor diastereoisomer:

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.33 (dd, J = 2.5, 1.5 Hz, 1H), 7.42 – 7.35 (m, 2H), 7.33 – 7.25 (m, 5H), 7.24 – 7.15 (m, 2H), 6.92 (ddd, J = 14.0, 8.0, 2.0 Hz, 1H), 3.60 – 3.51 (m, 1H), 3.47 – 3.37 (m, 1H), 3.30 – 3.13 (m, 2H), 2.95 – 2.74 (m, 2H), 2.28 – 2.06 (m, 2H).

¹³C NMR (101 MHz, CDCl₃): δ 201.6, 140.7, 139.6, 129.1, 128.9, 128.4, 128.3, 127.0, 126.9, 48.1, 47.3, 46.4, 36.0, 29.8.

<u>HRMS (ESI)</u> Exact mass calculated for $C_{19}H_{23}CINaO_2$ [M+CH₃OH+Na]⁺: 341.1279, found: 341.1268.



(*R*)-6-bromo-3,4-diphenylhexanal (2r). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), (3-bromopropyl)benzene (100 mg, 0.5 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1.5:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column

chromatography (hexane/diethyl ether 97:3) to afford product **2r** (13.5 mg, 41% yield, average of two runs) as an off-white solid. The enantiomeric excess of both diastereoisomers was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, by UPC² analysis on a Daicel Chiralpak CEL-1 column: isocratic CO₂/*i*-PrOH 90:10, flow rate 3 mL/min, $\lambda = 205$ nm. *Major diastereomer:* $\tau_{Major} = 4.69$ min, $\tau_{minor} = 3.59$ min. *Minor diastereomer:* $\tau_{Major} = 4.02$ min, $\tau_{minor} = 3.38$ min. [α]_D²⁶ = -38.8 (c = 0.49, CHCl₃, 1.5:1 *d.r.*, 80% *eemajor*, 74% *eeminor*).

Spectroscopic data for major diastereoisomer:

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.35 (dd, J = 2.5, 1.5 Hz, 1H), 7.43 – 7.34 (m, 2H), 7.34 – 7.25 (m, 5H), 7.24 – 7.13 (m, 2H), 6.92 (ddd, J = 12.0, 8.0, 2.0 Hz, 1H), 3.42 (td, J = 10.5, 4.5 Hz, 1H), 3.20 – 2.98 (m, 2H), 2.95 – 2.76 (m, 1H), 2.64 (ddd, J = 17.0, 10.0, 2.5 Hz, 1H), 2.44 (ddd, J = 17.0, 4.5, 1.5 Hz, 1H), 2.03 – 1.83 (m, 2H).

1³C NMR (101 MHz, CDCl₃): δ 201.2, 142.2, 141.3, 129.2, 129.0, 128.2, 128.2, 127.5, 127.3, 49.8, 49.4, 46.3, 36.9, 32.1.

Spectroscopic data for minor diastereoisomer:

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.65 (t, J = 2.0 Hz, 1H), 7.43 – 7.34 (m, 2H), 7.34 – 7.25 (m, 5H), 7.24 – 7.13 (m, 2H), 6.92 (ddd, J = 12.0, 8.0, 2.0 Hz, 1H), 3.60 – 3.51 (m, 1H), 3.34 – 3.26 (m, 1H), 3.20 – 2.98 (m, 2H), 2.95 – 2.76 (m, 2H), 2.35 – 2.15 (m, 2H).

¹³C NMR (101 MHz, CDCl₃): δ 201.5, 140.7, 139.4, 129.1, 128.9, 128.4, 128.3, 127.0, 126.9, 49.4, 47.3, 45.0, 36.1, 31.9.

<u>HRMS (ESI)</u> Exact mass calculated for $C_{19}H_{23}BrNaO_2$ [M+CH₃OH+Na]⁺: 385.0774, found: 385.0768.



(*R*)-7-(1,3-dioxoisoindolin-2-yl)-3,4-diphenylheptanal (2s). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 2-(4-phenylbutyl)isoindoline-1,3-dione (140 mg, 0.5 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and

dichloromethane (300 µL). The d.r. was determined to be 1.6:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column chromatography on silica gel support (*gradient from* hexane/diethyl ether 95:5 *to* hexane/diethyl ether 70:30) to afford the *major diastereoisomer* of product **2s** (15.0 mg) as an off-white solid. Continued elution delivered the *minor diastereoisomer* of **2s** (8.7 mg) as a pale yellow solid (57% overall yield, average of two runs). The enantiomeric excess for both diastereoisomer was determined by UPC² analysis on a Daicel Chiralpak IC-3 column: gradient CO₂/*i*-PrOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 210$ nm: *Major diastereomer*: $\tau_{major} = 3.92$ min, $\tau_{minor} = 4.04$ min; *Minor diastereomer*: $\tau_{major} = 4.13$ min, $\tau_{minor} = 4.23$ min.

Characterization for major diastereoisomer:

 $[\alpha]_D^{26} = -191.0$ (c = 0.18, CHCl₃, 83% *ee_{major}*).

 $\frac{1 \text{H NMR}}{(400 \text{ MHz, CDCl}_3): \delta 9.33 \text{ (dd, } J = 2.5, 1.5 \text{ Hz}, 1\text{H}), 7.78 \text{ (dd, } J = 5.5, 3.0 \text{ Hz}, 2\text{H}), 7.70 \text{ (dd, } J = 5.5, 3.0 \text{ Hz}, 2\text{H}), 7.34 \text{ (ddd, } J = 7.0, 7.0, 1.0 \text{ Hz}, 2\text{H}), 7.28 - 7.20 \text{ (m, 7H)}, 7.16 - 7.08 \text{ (m, 1H)}, 3.52 - 3.39 \text{ (m, 2H)}, 3.34 \text{ (ddd, } J = 10.5, 10.0, 4.5 \text{ Hz}, 1\text{H}), 2.79 \text{ (ddd, } J = 10.5, 10.5, 2.5 \text{ Hz}, 1\text{H}), 2.59 \text{ (ddd, } J = 17.0, 10.0, 2.5 \text{ Hz}, 1\text{H}), 2.41 \text{ (ddd, } J = 17.0, 4.5, 1.5 \text{ Hz}, 1\text{H}), 1.54 - 1.43 \text{ (m, 2H)}, 1.41 - 1.33 \text{ (m, 2H)}.$

¹³C NMR (101 MHz, CDCl₃): δ 201.4, 168.2, 142.5, 142.3, 133.8, 132.0, 128.8, 128.6, 128.2, 128.0, 127.0, 126.8, 123.1, 51.2, 49.0, 46.6, 37.4, 30.7, 26.1.

HRMS (ESI) Exact mass calculated for C₂₇H₂₅NNaO₃ [M+Na]⁺: 434.1727, found: 434.1716.

Characterization for **minor diastereoisomer**:

 $[\alpha]_D^{26} = +62.7 \ (c = 0.21, CHCl_3, 80\% \ ee_{minor}).$

 $\frac{^{1}\text{H NMR}}{^{1}\text{H NMR}} (400 \text{ MHz, CDCl}_{3}): \delta 9.61 (t, J = 2.0 \text{ Hz}, 1\text{H}), 7.84 (dd, J = 5.5, 3.0 \text{ Hz}, 2\text{H}), 7.72 (dd, J = 5.5, 3.0 \text{ Hz}, 2\text{H}), 7.21 - 7.10 (m, 6\text{H}), 6.94 - 6.83 (m, 4\text{H}), 3.70 - 3.56 (m, 2\text{H}), 3.50 (ddd, J = 9.0, 6.5, 6.0 \text{ Hz}, 1\text{H}), 2.94 (ddd, J = 11.0, 6.5, 4.5 \text{ Hz}, 1\text{H}), 2.86 (ddd, J = 17.0, 6.0, 2.0 \text{ Hz}, 1\text{H}), 2.77 (ddd, J = 17.0, 9.0, 2.0 \text{ Hz}, 1\text{H}), 1.85 - 1.75 (m, 1\text{H}), 1.75 - 1.65 (m, 1\text{H}), 1.58 - 1.47 (m, 2\text{H}).$

¹³C NMR (101 MHz, CDCl₃): δ 201.9, 168.4, 140.8, 140.7, 133.9, 132.1, 128.9, 128.8, 128.0 (2 *signals*), 126.6, 126.5, 123.2, 50.5, 47.0, 45.4, 37.7, 29.6, 26.7.

HRMS (ESI) Exact mass calculated for C₂₇H₂₅NNaO₃ [M+Na]⁺: 434.1727, found: 434.1722.



1,3-dioxoisoindolin-2-yl (*5R*)-**7-oxo-4,5-diphenylheptanoate** (**2t**). Prepared according to the general procedure using enal **1a** (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-**A** (14.1 mg, 20 mol%), 1,3-dioxoisoindolin-2-yl 4-phenylbutanoate (155 mg, 0.5 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be

1.5:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column chromatography on *neutral* silica gel support in order to prevent hydrolization of the product (*gradient from* hexane/diethyl ether 90:10 *to* hexane/diethyl ether 40:60) to afford the *major diastereoisomer* of product **2t** (14.3 mg) as an off-white solid. Continued elution delivered the *minor diastereoisomer* of **2t** (8.3 mg) as an amber wax (51% overall yield, average of two runs). The enantiomeric excess for both diastereoisomer was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding diols, by UPC² analysis on a Daicel Chiralpak CEL-1 column: gradient CO₂/EtOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 215$ nm: *Major diastereomer*: $\tau_{major} = 4.43$ min, $\tau_{minor} = 4.29$ min; *Minor diastereomer*: $\tau_{major} = 4.43$ min, $\tau_{minor} = 4.34$ min.

Characterization for major diastereoisomer:

 $[\alpha]_D^{26} = -11.2$ (c = 0.12, CHCl₃, 80% *ee_{major}*).

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.32 (dd, J = 2.5, 1.5 Hz, 1H), 7.86 (dd, J = 5.5, 3.0 Hz, 2H), 7.77 (dd, J = 5.5, 3.0 Hz, 2H), 7.38 (dd, J = 8.5, 6.5 Hz, 2H), 7.34 – 7.26 (m, 7H), 7.24 – 7.20 (m, 1H), 3.38 (td, J = 11.0, 4.5 Hz, 1H), 2.95 (td, J = 11.0, 3.5 Hz, 1H), 2.65 – 2.58 (m, 2H), 2.42 (ddd, J = 17.0, 4.5, 1.5 Hz, 1H), 2.34 (ddd, J = 16.5, 8.0, 5.0 Hz, 1H), 2.25 – 2.18 (m, 1H), 1.94 – 1.84 (m, 1H), 1.81 – 1.72 (m, 1H).

¹³<u>C NMR</u> (101 MHz, CDCl₃): δ 201.5, 169.5, 162.3, 142.4, 141.7, 135.1, 129.5, 129.3, 128.7, 128.4, 127.9, 127.5, 125.9, 124.3, 50.6, 49.7, 47.0, 29.4, 29.2.

<u>HRMS (ESI)</u> Exact mass calculated for $C_{27}H_{23}NNaO_5$ [M+Na]⁺: 464.1468, found: 464.1481.

Characterization for **minor diastereoisomer**:

 $[\alpha]_D^{26} = +10.2 \ (c = 0.09, CHCl_3, 83\% \ ee_{minor}).$

<u>¹H NMR</u> (400 MHz, CDCl₃): δ 9.62 (t, J = 2.0 Hz, 1H), 7.88 (dd, J = 5.5, 3.0 Hz, 2H), 7.78 (dd, J = 5.5, 3.0 Hz, 2H), 7.21 – 7.09 (m, 6H), 6.95 – 6.88 (m, 4H), 3.57 – 3.51 (m, 1H), 3.03 (ddd, J = 11.0, 7.0, 3.5 Hz, 1H), 2.93 (ddd, J = 17.0, 6.0, 2.0 Hz, 1H), 2.82 (ddd, J = 17.0, 9.0, 2.0 Hz, 1H), 2.54 – 2.45 (m, 1H), 2.41 – 2.25 (m, 3H), 2.08 – 1.98 (m, 1H).

¹³C NMR (101 MHz, CDCl₃): δ 201.5, 169.4, 161.9, 140.7, 139.6, 134.8, 128.9 (2 *signals*), 128.7, 128.3, 128.1, 126.9, 126.7, 124.0, 49.9, 47.3, 45.2, 29.3, 28.1.

HRMS (ESI) Exact mass calculated for C₂₇H₂₃NNaO₅ [M+Na]⁺: 464.1468, found: 464.1464.



methyl (4*R*)-6-oxo-3,4-diphenylhexanoate (2u). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), methyl 3-phenylpropionate (82 mg, 0.5 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1.1:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column

chromatography (*gradient from* hexane/diethyl ether 95:5 *to* hexane/diethyl ether 75:25) to afford the *major diastereoisomer* of product 2u (7.3 mg) as a colourless solid. Continued elution delivered the *minor diastereoisomer* of 2u (5.9 mg) as a colourless oil (45% overall yield, average of two runs). The enantiomeric excess of both diastereoisomers was determined by HPLC analysis on a Daicel

Chiralpak IC-3 column: 80:20 hexane/*i*-PrOH, flow rate 1.0 mL/min, $\lambda = 215$ nm. *Major diastereomer*: $\tau_{Major} = 11.1$ min, $\tau_{minor} = 14.3$ min. *Minor diastereomer*: $\tau_{Major} = 16.0$ min, $\tau_{minor} = 14.4$ min.

Characterization for major diastereoisomer:

 $[\alpha]_D^{26} = +18.3 (c = 0.04, CHCl_3, 82\% ee).$

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.35 (dd, J = 2.5, 1.5 Hz, 1H), 7.41 – 7.24 (m, 10H), 3.46 – 3.31 (m, 2H), 3.40 (s, 3H), 2.67 (ddd, J = 17.0, 9.5, 2.5 Hz, 1H), 2.55 – 2.38 (m, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 200.8, 172.3, 141.7, 141.6, 129.0, 128.8, 128.2, 128.1, 127.3 (2 *signals*), 51.3, 48.7, 47.9, 45.9, 39.5.

HRMS (ESI) Exact mass calculated for C₁₉H₂₀NaO₃ [M+Na]⁺: 319.1305, found: 319.1307.

Characterization for minor diastereoisomer:

 $[\alpha]_D^{26} = +18.3 (c = 0.07, CHCl_3, 82\% ee).$

 $\frac{1}{\text{H NMR}}$ (400 MHz, CDCl₃): δ 9.61 (t, *J* = 2.0 Hz, 1H), 7.26 – 7.16 (m, 6H), 6.95 – 6.87 (m, 4H), 3.66 – 3.59 (m, 1H), 3.59 (s, 3H), 3.56 – 3.51 (m, 1H), 2.83 – 2.79 (m, 2H), 2.76 (dd, *J* = 16.0, 6.5 Hz, 1H), 2.65 (dd, *J* = 16.0, 8.5 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃): δ 201.3, 172.5, 139.9, 139.7, 128.9, 128.7, 128.1, 128.0, 127.0, 126.9, 51.7, 46.5, 46.5, 44.2, 37.4.

HRMS (ESI) Exact mass calculated for C₁₉H₂₀NaO₃ [M+Na]⁺: 319.1305, found: 319.1295.



(*R*)-3-(6-methoxy-3-oxo-2,3-dihydro-1H-inden-1-yl)-3-phenylpropanal (2v) Prepared according to the general procedure using cinnamaldehyde (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 5-methoxy-2,3-dihydro-1H-inden-1-one (162 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1.9:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column

chromatography (*gradient from* hexane/diethyl ether 90:10 *to* hexane/diethyl ether 80:20, two consecutive purifications) to afford the *minor diastereoisomer* of product **2v** (7.6 mg) as an off-white solid. Continued elution delivered the *major diastereoisomer* of **2v** (9..4 mg) as an off-white solid (58% overall yield, average of two runs). The enantiomeric excess was determined UPC² analysis on a Daicel Chiralpak IC-3 column: gradient CO₂/*i*-PrOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm. *Major diastereomer*: $\tau_{major} = 4.67$ min, $\tau_{minor} = 4.82$ min. *Minor diastereomer*: $\tau_{major} = 4.58$ min, $\tau_{minor} = 4.42$ min. UPC² (Daicel Chiralpak IC-3 column, 20 °C, gradient CO₂/*i*-PrOH from 100% CO₂ to 60:40 wrate 3 mL/min, $\lambda = 220$ nm).

Characterization for major diastereoisomer:

 $[\alpha]_D^{26} = -5.41$ (c = 0.21 CHCl₃, 91% *ee_{major}*).

¹<u>H NMR</u> (500 MHz, CDCl₃) δ 9.70 (dd, J = 2.0, 1.5 Hz, 1H), 7.65 (d, J = 8.5 Hz, 1H), 7.31 – 7.17 (m, 5H), 6.87 (dd, J = 8.5, 2.0 Hz, 1H), 6.74 (d, J = 2.0 Hz, 1H), 3.83 (s, 3H), 3.62 – 3.69 (m, 1H), 3.56 (ddd, J = 17.5, 5.5, 1.5 Hz, 1H), 3.04 – 2.91 (m, 3H), 2.69 (d, J = 13.5 Hz, 1H);

¹³C NMR (128 MHz, CDCl₃) δ 205.1, 200.1, 165.5, 156.0, 141.9, 130.0, 128.7, 128.2, 127.0, 125.7, 115.5, 109.4, 55.6, 51.2, 48.2, 41.0, 29.7;

Characterization data for minor diastereoisomer

 $[\alpha]_D^{26} = -23.85 \ (c = 0.08, CHCl_3, 86\% \ ee_{minor}).$

¹<u>H NMR</u> (500 MHz, CDCl₃) δ 9.71 (dd, J = 2.5, 1.5 Hz, 1H), 7.66 (d, J = 8.5 Hz, 1H), 7.30 – 7.14 (m, 5H), 6.87 (dd, J = 8.5, 2.5 Hz, 1H), 6.79 (bs, 1H), 3.97 (ddd, J = 9.5, 6.0, 4.0 Hz, 1H), 3.85 (s, 3H), 3.08 (qd, J = 8.0, 4.5 Hz, 2H), 3.03 – 2.87 (m, 3H);

¹³C NMR (128 MHz, CDCl₃) δ 205.4, 201.5, 165.6, 156.6, 141.2, 130.8, 128.8, 128.2, 127.2, 125.7, 115.7, 109.6, 55.8, 51.2, 44.8, 40.1, 29.8;

HRMS (ESI) Exact mass calculated for C₁₉H₁₈NaO₃ [M+ Na]⁺: 317.1148, found: 317.1148.

(3*R*)-3-(2,3-dihydro-1H-inden-1-yl)-3-phenylpropanal (2w). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), indane (118 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1.1:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column chromatography (hexane/diethyl ether 95:5) to afford

product **2w** (23.7 mg, 91% yield, 1.1:1 mixture of diastereoisomers, average of two runs) as a pale yellow oil. The enantiomeric excess of both diastereoisomers was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, by HPLC analysis on a Daicel Chiralpak IC-3 column, 98.5:1.5 hexane/*i*-PrOH, flow rate 1.2 mL/min, T = 30 °C, $\lambda = 215$ nm. *Major diastereomer:* $\tau_{minor} = 45.9$ min, $\tau_{major} = 62.2$ min. *Minor diastereomer:* $\tau_{major} = 43.7$ min, $\tau_{minor} = 52.1$ min. [α]_D²⁶ = -28.8 (c = 0.80, CHCl₃, 1.1:1 *d.r.*, 80% *eemajor*, 77% *eeminor*). *The spectroscopic data for compound* **2w** *were not assigned and are reported as a mixture of diastereoisomers:*

<u>¹H NMR</u> (400 MHz, CDCl₃, isolated as a 1.1:1 mixture of diastereoisomers) δ 9.60 (dd, J = 2.5, 1.7 Hz, 1H), 9.57 (t, J = 2.1 Hz, 1H), 7.33 – 7.11 (m, 16H), 7.07 (app t, J = 7.2 Hz, 1H), 6.88 (d, J = 7.5 Hz, 1H), 3.65 – 3.58 (m, 1H), 3.53 (q, J = 6.4 Hz, 1H), 3.43 – 3.35 (m, 2H), 2.96 – 2.83 (m, 3H), 2.80 – 2.60 (m, 5H), 2.20 – 2.11 (m, 1H), 2.10 – 1.99 (m, 1H), 1.97 – 1.88 (m, 1H), 1.87 – 1.79 (m, 1H); <u>¹³C NMR</u> (101 MHz, CDCl₃) δ 201.9, 201.8, 144.9, 144.8, 144.7, 144.3, 142.4, 142.0, 128.5, 128.4, 128.3, 128.2, 127.0, 126.9, 126.8, 126.7, 126.0, 125.9, 125.3, 124.8, 124.6, 124.5, 50.4, 47.6, 46.1, 43.8, 43.6, 31.2, 30,7, 29.3, 29.1;

HRMS (ESI) Exact mass calculated for C₁₉H₂₂NaO₂ [M+CH₃OH+Na]⁺: 305.1512, found: 305.1516.



(3*R*)-3-(2,3-dihydro-1H-inden-1-yl)-3-(3-methoxyphenyl)propanal (2x). Prepared according to the general procedure using enal 1u (16.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), indane (118 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1.4:1 by ¹H NMR analysis of the crude

mixture, was purified by flash column chromatography (hexane/diethyl ether 95:5) to afford product **2x** (18.0 mg, 64% yield, 1.4:1 mixture of diastereoisomers, average of two runs) as a pale yellow oil. The enantiomeric excess of both diastereoisomers was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, by UPC² analysis on a Daicel Chiralpak ID-3 column: gradient CO₂/*i*PrOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 273$ nm: *Minor diastereomer:* $\tau_{major} = 3.35$ min, $\tau_{minor} = 3.48$ min; *Major diastereomer:* $\tau_{major} = 3.40$ min, $\tau_{minor} = 3.67$ min. [α]_D²⁶ = -24.0 (c = 0.55, CHCl₃, 1.4:1 *d.r.*, 86% *ee_{Major}*, 82% *ee_{minor}*).

Spectroscopic data for major diastereoisomer:

¹<u>H NMR</u> (400 MHz, CDCl₃) δ 9.60 (dd, *J* = 2.5, 1.5 Hz, 1H, *A*), 7.30 – 7.25 (m, 1H), 7.23 – 7.12 (m, 3H,), 7.11 – 7.04 (m, 1H), 6.81 – 6.73 (m, 2H), 6.66 (t, *J* = 2.0 Hz, 1H), 3.74 (s, 3H), 3.63 – 3.47 (m, 1H), 3.41 – 3.32 (m, 1H), 2.93 – 2.60 (m, 4H), 2.23 – 1.99 (m, 1H), 1.97 – 1.90 (m, 1H); *Spectroscopic data for minor diastereoisomer*:

¹<u>H NMR</u> (400 MHz, CDCl₃) δ 9.58 (t, J = 2.0 Hz, 1H), 7.30 – 7.25 (m, 1H), 7.23 – 7.12 (m, 3H), 7.11 – 6.93 (d, 7.07 J = 7.5 Hz, 1H), 6.81 – 6.73 (m, 2H), 6.62 (t, J = 2.0 Hz, 1H), 3.72 (s, 3H), 3.63 – 3.47 (m, 1H), 3.41 – 3.32 (m, 1H), 2.93 – 2.60 (m, 4H), 2.23 – 1.99 (m, 1H), 1.97 – 1.90 (m, 1H); *The signals relative to* ¹³C NMR analysis were not assigned and are reported as a mixture of

The signals relative to ¹³C NMR analysis were not assigned and are reported as a mixture of diastereoisomers:

¹³C NMR (101 MHz, CDCl₃) δ 202.0, 202.0, 159.8, 159.6, 145.1, 145.0, 144.9, 144.4, 144.1, 143.7, 129.6, 129.5, 127.2, 127.1, 126.2, 126.0, 125.4, 125.0, 124.7, 124.7, 120.8, 120.6, 114.4, 114.1, 112.3, 112.1, 55.3, 50.4, 47.7, 46.2, 44.0, 43.7, 31.7, 30.9, 29.3.

HRMS (ESI) Exact mass calculated for C₁₉H₂₀NaO₂ [M+Na]⁺: 303.1356, found: 303.1368.



(S)-3-phenyl-4-(*p*-tolyl)butanal (2y). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (S)-A (14.1 mg, 20 mol%), *p*-xylene (134 mg, 1 mmol), trichloroacetic acid (16.3 mg, 100 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2y (15.0

mg, 63% yield, 83% ee, average of two runs) as a colourless oil that displayed spectroscopic data consistent with those reported previously.² The enantiomeric excess was determined to be 83% by HPLC analysis on a Daicel Chiralpak IC-3 column: 96:4 hexane/*i*-PrOH, flow rate 0.8 mL/min, $\lambda = 215$ nm: $\tau_{major} = 12.1$ min, $\tau_{minor} = 13.1$ min. $[\alpha]_D^{26} = -17.3$ (c = 0.45, CHCl₃, 83% ee); Lit: $[\alpha]_D^{26} = -50.8$ (c = 0.69, CHCl₃, 91% ee for (*S*)-enantiomer).²

¹<u>H NMR</u> (500 MHz, CDCl₃): δ 9.58 (t, *J* = 2.0 Hz, 1H), 7.31–7.26 (m, 2H), 7.04 (d, *J* = 8.0 Hz, 2H), 6.95 (d, *J* = 8.0 Hz, 2H), 3.46 (p, 7.04 (d, *J* = 7.5 Hz, 1H) 2.94 (dd, *J* = 13.5, 7.0 Hz, 1H), 2.83 (dd, *J* = 13.5, 8.0 Hz, 1H), 2.78 – 2.68 (m, 2H), 2.29 (s, 3H);

¹³C NMR (125.8 MHz, CDCl₃): δ 201.8, 143.5, 136.3, 136.0, 129.2, 129.2, 128.7, 127.7, 126.8, 49.1, 43.0, 42.2, 21.2.



(S)-3-phenyl-4-(*o*-tolyl)butanal (2z). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (S)-A (14.1 mg, 20 mol%), *o*-xylene (134 mg, 1 mmol), trichloroacetic acid (16.3 mg, 100 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 2z (11.0 mg, 47%)

yield, 77% ee, average of two runs) as a colourless oil that displayed spectroscopic data consistent with those reported previously.² The enantiomeric excess was determined to be 77% by HPLC analysis on a Daicel Chiralpak IC-3 column: 90:10 hexane/*i*-PrOH, flow rate 0.7 mL/min, $\lambda = 215$ nm: $\tau_{major} = 10.5$ min, $\tau_{minor} = 11.9$ min. $[\alpha]_D^{26} = -40.7$ (c = 0.23, CHCl₃, 77% ee); Lit: $[\alpha]_D^{26} = -22.2$ (c = 0.59, CHCl₃, 90% ee for (*S*)-enantiomer).²

 $\frac{1 \text{H NMR}}{14} (400 \text{ MHz}, \text{CDCl}_3): \delta 9.57 \text{ (t, } J = 1.9 \text{ Hz}, 1\text{H}), 7.32-7.25 \text{ (m, 2H)}, 7.23-7.03 \text{ (m, 6H)}, 6.97 \text{ (d, } J = 7.0 \text{ Hz}, 1\text{H}), 6.78 \text{ (s, 1H)}, 3.47 \text{ (p, } J = 7.4 \text{ Hz}, 1\text{H}), 2.97 - 2.86 \text{ (m, 2H)}, 2.85 - 2.72 \text{ (m, 2H)}, 2.26 \text{ (s, 3H)};$

¹³<u>C NMR</u> (101 MHz, CDCl₃): δ 201.7, 143.6, 137.7, 136.5, 130.6, 130.3, 128.8, 127.6, 126.9, 126.7, 125.9, 49.0, 41.0, 19.6.



(*S*)-4-(4-(tert-butyl)phenyl)-3-phenylbutanal (2aa). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 1-(*t*-butyl)-4-methylbenzene (148 mg, 1 mmol), trichloroacetic acid (16.3 mg, 100 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl

ether 92:8) to afford product **2aa** (15.0 mg, 53% yield, 77% ee, average of two runs) as a colourless oil. The enantiomeric excess was determined, upon sodium borohydride reduction of the isolated aldehyde to afford the corresponding alcohol, to be 77% by HPLC analysis on a Daicel Chiralpak IC-3 column: 90:10 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm: $\tau_{Major} = 10.9$ min, $\tau_{minor} = 12.2$ min. $[\alpha]_D^{26} = -33.4$ (c = 0.63, CHCl₃, 78% ee).

¹<u>H NMR</u> (500 MHz, CDCl₃): δ 9.57 (t, J = 2.0 Hz, 1H), 7.33–7.22 (m, 4H), 7.24–7.18 (m, 3H), 7.02 (d, J = 8.2 Hz, 2H), 3.54 – 3.47 (m, 1H), 3.00 (dd, J = 13.6, 6.4 Hz, 1H), 2.85 (dd, J = 13.6, 8.6 Hz, 1H) 2.81 – 2.69 (m, 2H), 1.32 (s, 9H);

¹³C NMR (125.8 MHz, CDCl₃): δ 202.0, 149.4, 143.7, 136.3, 129.0, 128.8, 127.6, 126.9, 125.4, 48.9, 42.9, 42.1, 34.5, 31.5.

HRMS (ESI) Exact mass calculated for C₂₁H₂₈NaO₂ [M+CH₃OH+Na]⁺: 335.1980, found: 335.1982.



(*S*)-4-(3,5-di-tert-butylphenyl)-3-phenylbutanal (2ab). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 1,3-di-*t*-butyl-5-methylbenzene (204 mg, 1 mmol), trichloroacetic acid (16.3 mg, 100 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl

ether 92:8) to afford product **2ab** (20.0 mg, 59% yield, 74% ee, average of two runs) as a colourless oil. The enantiomeric excess was determined to be 74% by HPLC analysis on a Daicel Chiralpak IC-3 column: 98:2 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm: $\tau_{Major} = 10.9$ min, $\tau_{minor} = 12.2$ min. $[\alpha]_D^{26} = -19.1$ (c = 0.73, CHCl₃, 74% ee).

¹<u>H NMR</u> (500 MHz, CDCl₃): δ 9.59 (t, J = 2.0 Hz, 1H), 7.30–7.25 (m, 2H), 7.23–7.17 (m, 2H), 7.14 (d, J = 6.8 Hz, 2H), 6.82 (d, J = 1.8 Hz, 2H), 3.45 (p, J = 7.4 Hz, 1H), 2.95 (dd, J = 13.4, 7.2 Hz, 1H), 2.87 (dd, J = 13.4, 7.6 Hz, 1H) 2.81 – 2.72 (m, 2H), 1.26 (s, 18H);

¹³C NMR (125.8 MHz, CDCl₃): δ 202.0, 150.7, 143.5, 138.2, 128.7, 127.8, 126.8, 123.7, 120.2, 49.0, 44.1, 42.4, 34.8, 31.5.

HRMS (ESI) Exact mass calculated for C₂₄H₃₂NaO [M+Na]⁺: 359.2348, found: 359.2345.



(S)-4-(4-(tert-butyl)-2-methylphenyl)-3-phenylbutanal and (S)-4-(5-(tert-butyl)-2-methylphenyl)-3-phenylbutanal (2ac). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (S)-A (14.1 mg, 20 mol%), 4-(t-butyl)-1,2-dimethylbenzene (134 mg, 1 mmol), trichloroacetic acid (16.3 mg, 100 mol%) and dichloromethane (300 μ L). The

regioisomeric ratio was determined to be 6.5:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product **2ac** (18.0 mg, 60% yield, average of two runs) as a colourless oil. The enantiomeric excess was determined, upon

sodium borohydride reduction of the isolated aldehyde to afford the corresponding alcohol, to be 73% for the major regioisomer and 74% for the minor by UPC² analysis on CEL1 chiral column: 95:5 CO₂/EtOH, curve 6, flow rate 3 mL/min, $\lambda = 203$ nm. *Minor regioisomer:* $\tau_{minor} = 1.51$ min, $\tau_{major} = 1.85$ min; *major regioisomer:* $\tau_{minor} = 1.63$ min, $\tau_{major} = 1.97$ min. $[\alpha]_D^{26} = -15.6$ (c = 0.25, CHCl₃, 6.5:1 *regioisomeric ratio*, 73% *eemajor*, 74% *eeminor*).

Spectroscopic data for major regioisomer (2ac-p):

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.55 (t, *J* = 2.0 Hz, 1H), 7.33–7.26 (m, 2H), 7.24–7.18 (m, 3H), 7.16–7.04 (m, 2H), 6.90 (d, *J* = 8.0 Hz, 1H), 3.50 – 3.41 (m, 1H), 2.94 (dd, *J* = 14.0, 6.0 Hz, 1H), 2.90 – 2.60 (m, 3H), 2.28 (s, 3H), 1.29 (s, 9H);

¹³C NMR (101 MHz, CDCl₃): δ 202.0, 149.5, 143.9, 143.6, 135.9, 134.6, 129.9, 128.8, 127.5, 126.9, 122.9, 48.8, 40.9, 40.6, 34,4, 31.5, 19.9.

Spectroscopic data for minor regioisomer (2ac-m):

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.60 (t, *J* = 2.0 Hz, 1H), 7.33–7.26 (m, 2H), 7.24–7.18 (m, 3H), 7.16–7.04 (m, 2H), 6.82 (d, *J* = 2.0 Hz, 1H), 3.50–3.41 (m, 1H), 2.94 (dd, *J* = 14.0, 6.0 Hz, 1H), 2.90–2.60 (m, 3H), 2.27 (s, 3H), 1.19 (s, 9H);

¹³C NMR (101 MHz, CDCl₃): δ 201.8, 148.6, 137.0, 133.3, 130.2, 128.7, 127.7, 126.8, 123.4, 49.2, 41.4, 40.9, 34.2, 31.4, 19.0.

HRMS (ESI) Exact mass calculated for C₂₂H₃₀NaO₂ [M+CH₃OH+Na]⁺: 349.2154, found: 349.2138.



(*S*)-3-phenyl-4-(2,4,5-trimethylphenyl)butanal (2ad). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 1,2,4,5-tetramethylbenzene (134 mg, 1 mmol), trichloroacetic acid (16.3 mg, 100 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography (hexane/diethyl

ether 92:8) to afford product **2ad** (18.0 mg, 68% yield, 82% ee, average of two runs) as a colourless oil. The enantiomeric excess was determined by HPLC analysis on a Daicel Chiralpak IC-3 column: 90:10 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm: $\tau_{Major} = 11.1$ min, $\tau_{minor} = 12.0$ min. [α]_D²⁶ = -32.7 (c = 0.50, CHCl₃, 82% ee).

¹<u>H NMR</u> (500 MHz, CDCl₃): δ 9.54 (t, *J* = 2.0 Hz, 1H), 7.32–7.27 (m, 2H), 7.23 – 7.18 (m, 3H), 6.90 (s, 1H), 6.78 (s, 1H), 3.47 – 3.40 (m, 1H), 2.89 (dd, *J* = 13.7, 6.2 Hz, 1H), 2.82 – 2.71 (m, 3H), 2.19 (s, 6H), 2.17 (s, 6H);

¹³C NMR (125.8 MHz, CDCl₃): δ 202.0, 144.0, 134.9, 134.7, 133.9, 133.6, 131.9, 131.6, 128.8, 127.5, 126.8, 48.8, 41.2, 40.6, 19.3, 19.0.

HRMS (ESI) Exact mass calculated for C₂₀H₂₆NaO₂ [M+CH₃OH+Na]⁺: 321.1828, found: 321.1825.



(S)-4-(2,3,4,5,6-pentamethylphenyl)-3-phenylbutanal (2ae). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (S)-A (14.1 mg, 20 mol%), hexamethylbenzene (162 mg, 1 mmol), trichloroacetic acid (16.3 mg, 100 mol%) and dichloromethane (300 μ L). The crude mixture was purified by flash column chromatography

(hexane/diethyl ether 92:8) to afford product **2ae** (22.2 mg, 75% yield, 82% ee, average of two runs) as off-white solid. The enantiomeric excess was determined by HPLC analysis on a Daicel Chiralpak IC-3 column: 90:10 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm: $\tau_{major} = 11.1$ min, $\tau_{minor} = 14.7$ min. $[\alpha]_D^{26} = -23.6$ (c = 0.50, CHCl₃, 82% ee).

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.47 (dd, *J* = 2.5, 1.5 Hz, 1H), 7.35–7.30 (m, 2H), 7.30–7.20 (m, 3H), 3.42 (tt, *J* = 10.0, 5.5 Hz 1H), 3.11 (dd, *J* = 14.5, 9.5 Hz, 1H), 3.00 (dd, *J* = 14.5, 5.5 Hz, 1H), 2.86 (ddd, *J* = 16.5, 10.0, 2.5 Hz, 1H), (ddd, *J* = 16.5, 5.0, 2.0 Hz, 1H), 2.26 (s, 3H), 2.23 (s, 12H); ¹³<u>C NMR</u> (101 MHz, CDCl₃): δ 202.2, 144.1, 133.4, 133.3, 132.9, 132.4, 128.8, 127.4, 126.9, 47.9, 41.3, 37.9, 17.3, 17.1, 17.1.

HRMS (ESI) Exact mass calculated for C₂₂H₃₀NaO₂ [M+CH₃OH+Na]⁺: 349.2138, found: 349.2139



methyl (*S*)-2-(4-((*R*)-6-methyl-1-oxo-3-phenylheptan-4-yl)phenyl) propanoate (4). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 3 (110 mg, 0.5 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The crude mixture, which was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford product 4 (34.5 mg,

50% yield) as a colorless oil. Due two the overlap of diagnostic signals in the ¹H NMR spectrum, the diastereomeric ratio of the product mixture was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, *via* UPC² analysis to be 1:1.3:8.2:10. $[\alpha]_D^{26} = 42.4$ (c = 0.99, CHCl₃, 1:1.3:8.2:10 *d.r*).

The spectroscopic data for compound 4 were not assigned and are reported as a mixture of diastereoisomers:

<u>¹H NMR</u> (400 MHz, CDCl₃, *isolated as a 1.2:1 mixture of diastereoisomers*): δ 9.64 (t, J = 2.0, Hz, 1H), 9.41 (dd, J = 2.5, 1.0 Hz, 1H), 7.35–7.28 (m, 2H), 7.26–7.19 (m, 4H), 7.14–7.08 (m, 6H), 7.04 (d, J = 8.0 Hz, 2H), 6.89–6.84 (m, 2H), 6.67 (d, J = 8.0 Hz, 2H), 3.85 (dt, J = 8.5, 6.0 Hz, 1H), 3.76 – 3.61 (m, 9H), 2.84 – 2.74 (m, 2H), 2.67 (ddd, J = 16.5, 8.5, 2.0 Hz, 1H), 2.61 – 2.52 (m, 2H), 2.44 (ddd, J = 17.0, 4.0, 1.5 Hz, 1H), 2.04 – 1.93 (m, 1H), 1.69 – 1.59 (m, 1H), 1.51 (d, J = 7.0 Hz, 3H), 1.44 (d, J = 7.0 Hz, 3H), 1.13 (d, J = 6.5 Hz, 3H), 0.74 (d, J = 7.0 Hz, 3H), 0.71 (d, J = 6.5 Hz, 2H), 0.65 (d, J = 7.0 Hz, 3H);

¹³C NMR (101 MHz, CDCl₃): δ 202.3, 202.1, 175.3, 175.3, 143.0, 140.8, 138.9, 138.4, 138.3, 138.2, 130.7, 129.3, 128.9, 128.2, 127.9, 127.2, 126.9, 126.6, 126.4, 57.6, 57.0, 52.2, 52.1, 49.5, 48.4, 45.1, 45.1, 42.3, 40.6, 28.8, 28.4, 22.3, 20.1, 18.8, 16.6;

HRMS (ESI) Exact mass calculated for C₂₃H₂₈NaO₃ [M+Na]⁺: 375.1923, found: 375.1931.

To gain further insight about the C- ζ selectivity for the reaction between **1a** and **3**, the recovered **3** after the photochemical protocol was submitted to HPLC analysis to determine the degree of its racemization. An analogous level of enantiopurity compared to the un-submitted starting material was inferred. The lack of racemization indicates that radical formation is not occurring at C- α position of **3**.



Figure S4. HPLC analysis of an authentic sample of rac-3, (S)-3 and the recovered starting material after the reaction to deliver 4.

We investigated the recativity of two substrates that, individually, could mimic both substitution patterns of (S)-ibuprofen methyl ester **3**. While methyl 2-phenylpropanoate remained completely unreacted, *iso*-butylbenzene (a secondary-carbon containing substrate) provided the corresponding product in good yield and enantioselectivity. The lack of reactivity of the former substrate is particularly informative since it mimics the tertiary carbon and the electronics of (S)-ibuprofen methyl ester **3**. Overall, these results indicate the difficulty to form a tertiary radical using the PCET mechanism operative in our system.





(*R*)-5-methyl-3,4-diphenylhexanal (2af). Prepared according to the general procedure using enal 1a (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-A (14.1 mg, 20 mol%), 1,2-diphenylethane (134 mg, 1 mmol), zinc(II) triflate (18.2 mg, 50 mol%) and dichloromethane (300 μ L). The d.r. was determined to be 1.2:1 by ¹H NMR analysis of the crude mixture, which was purified by flash column chromatography

(hexane/diethyl ether 97:3) to afford product **2af** (17 mg, 64% yield, average of two runs) as a paleyellow oil. The enantiomeric excess of both diastereoisomers was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, by UPC² analysis on a Daicel Chiralpak IB column: gradient CO₂/EtOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 210$ nm. *Major diastereomer:* $\tau_{Major} = 3.17$ min, $\tau_{minor} =$ 2.71 min. *Minor diastereomer:* $\tau_{Major} = 3.51$ min, $\tau_{minor} = 2.95$ min. [α]_D²⁶ = +53.5 (c = 0.57, CHCl₃, 1.2:1 *d.r.*, 74% *ee_{major}*, 75% *ee_{minor}*).

Spectroscopic data for major diastereoisomer:

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.66 (t, J = 2.0 Hz, 1H), 7.35 – 7.29 (m, 2H), 7.29 – 7.20 (m, 2H), 7.19 – 7.10 (m, 4H), 6.76 – 6.71 (m, 2H), 3.87 (dt, J = 9.0, 6.5 Hz, 1H), 2.87 – 2.76 (m, 1H), 2.69 (ddd, J = 17.0, 9.0, 2.0 Hz, 1H), 2.63 – 2.54 (m, 1H), 2.03 (dp, J = 8.0, 7.0 Hz, 1H), 1.13 (d, J = 6.5 Hz, 3H), 0.76 (d, J = 7.0 Hz, 3H).

Spectroscopic data for minor diastereoisomer:

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.42 (dd, J = 2.5, 1.5 Hz, 1H), 7.35 – 7.29 (m, 2H), 7.29 – 7.20 (m, 2H), 7.19 – 7.10 (m, 4H), 6.92 – 6.86 (m, 2H), 3.73 (td, J = 11.0, 4.0 Hz, 1H), 2.87 – 2.76 (m, 1H), 2.63 – 2.54 (m, 1H), 2.46 (ddd, J = 17.0, 4.0, 1.5 Hz, 1H), 1.67 (pd, J = 7.0, 4.0 Hz, 1H), 0.74 (d, J = 6.5 Hz, 3H), 0.67 (d, J = 7.0 Hz, 3H).

The signals relative to ${}^{13}C$ NMR analysis were not assigned and are reported as a mixture of diastereoisomers:

¹³C NMR (125.8 MHz, CDCl₃) & 202.3, 202.0, 143.1, 141.0, 139.5, 139.3, 130.6, 129.2, 128.9, 128.2, 128.1, 127.9, 127.4, 126.9, 126.8, 126.6, 126.3, 57.9, 57.4, 49.4, 48.5, 42.3, 40.6, 28.7, 28.4, 22.3, 19.8, 16.7.

D.3 Unreactive Substrates



Figure S5. Survey of unreactive or scarcely reactive substrates.

E. Mechanistic Insights

E.1. Site-selectivity study

	1a + Me H _B	catalyst (S)- A (20) acid co-catalyst (x <i>HP</i> single LED (42 irradiance 45±2 m\ CH ₂ Cl ₂ , 35 °C, 1	mol%) <u>mol%)</u> H 0 nm) N/cm ² Pr 6 h	Me 6A	Me H Ph 6B	Et
entry	Acid	Yield $(\%)^a$	6A : 6B ^b	dr 6A °	ee 6A (%) (major/minor)	ee 6B (%)
1	$Zn(CF_3SO_3)_2$ (50 mol%)	63	1:6	1.7:1	76/75	77
2	TFA (100 mol%)	65	1:3.3	1.6:1	71/70	74
3	TCA (100 mol%)	62	1:3	1.5:1	73/77	71
4	DCA (100 mol%)	50	1:2.6	1.4:1	69/67	70
5	MCA (100 mol%)	-	-	-	-	-

Table S8. Evaluating the influence of the strenght of the conjugate base on the regioselectivity

^{*a*} Overall isolated yields of mixtures of regioisomers **6A** and **6B**. ^b Regioisomeric ratio measured by ¹H NMR analysis of the crude mixture. ^{*c*} Diastereoisomeric ratio measured by ¹H NMR analysis of the crude mixture. TFA: trifluoroacetic acid; TCA: trichloroacetic acid; DCA: dichloroacetic acid; MCA: monochloroacetic acid.



(*3R*)-**3-phenyl-4-(p-tolyl)pentanal** (**6A**) *and* (*S*)-**4-(4-ethylphenyl)-3-phenylbutanal** (**6B**). Prepared according to the general procedure using enal **1a** (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-**A** (14.1 mg, 20 mol%), methyl 4-ethyltoluene (120 mg, 1 mmol), zinc(II) triflate (18.2 mg,

50 mol%) and dichloromethane (300 µL). The crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford products **6** [16.0 mg, 63% yield, 6:1 mixture of regioisomers **6A** (1.7:1 d.r., 76% *ee_{major}*, 75% *ee_{minor}*) and **6B** (77% *ee*)] as a pale yellow oil. The enantiomeric excess of both regioisomers, and corresponding diastereoisomers for **6A**, was determined, upon sodium borohydride reduction of the isolated aldehydes to afford the corresponding alcohols, by UPC² analysis on a Daicel Chiralpak CEL-1 column: CO₂/MeOH 95:5, flow rate 3 mL/min, $\lambda = 210$ nm. For compound **6A**, *Major diastereomer:* $\tau_{Major} = 3.27$ min, $\tau_{minor} = 2.59$ min. *Minor diastereomer:* $\tau_{Major} = 3.47$ min, $\tau_{minor} = 3.02$ min. For compound **6B**: $\tau_{Major} = 4.46$ min, $\tau_{minor} = 3.70$ min.

Spectroscopic data for compound 6A, major diastereoisomer:

¹<u>H NMR</u> (400 MHz, CDCl₃): δ 9.47 (dd, J = 2.5, 1.5 Hz, 1H), 7.48 – 7.23 (m, 7H), 7.16 – 7.10 (m, 2H), 3.40 (ddd, J = 10.0, 10.0, 4.5 Hz, 1H), 3.02 – 2.93 (m, 1H), 2.76 – 2.69 (m, 1H), 2.60 (ddd, J = 16.5, 4.5, 1.5 Hz, 1H), 2.47 (s, 3H), 1.15 (d, J = 7.0 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 201.8, 142.9, 142.2, 136.4, 129.5, 128.8, 128.3, 127.6, 126.9, 49.2, 47.7, 45.8, 21.2, 20.8.

Spectroscopic data for compound 6A, minor diastereoisomer:

 $\frac{1}{H}$ NMR (400 MHz, CDCl₃): δ 9.68 (t, J = 2.0 Hz, 1H), 7.48 – 7.23 (m, 7H), 7.03 – 6.99 (m, 2H), 3.64 – 3.57 (m, 1H), 3.17 (qd, J = 7.0, 7.0 Hz, 1H), 2.93 – 2.89 (m, 2H), 2.41 (s, 3H), 1.38 (d, J = 7.0 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 202.3, 141.7, 140.8, 135.9, 128.8, 128.7, 128.2, 128.1, 126.7, 46.6, 46.1, 44.6, 21.1, 18.0.

Spectroscopic data for compound **6B**:

 $\frac{1 \text{H NMR}}{7.14 - 7.10} (\text{m, 2H}), 3.61 - 3.55 (\text{m, 1H}), 3.08 (\text{dd}, J = 13.5, 6.5 \text{ Hz}, 1\text{H}), 3.01 - 2.93 (\text{m, 1H}), 2.88 - 2.84 (\text{m, 2H}), 2.76 - 2.69 (\text{m, 2H}), 1.34 (\text{t}, J = 7.5 \text{ Hz}, 3\text{H}).$

¹³C NMR (101 MHz, CDCl₃): δ 201.9, 143.6, 142.4, 136.5, 129.3, 128.7, 128.0, 127.7, 126.8, 49.0, 43.1, 42.2, 28.6, 15.7.

HRMS (ESI) Exact mass calculated for C₁₉H₂₄NaO₂ [M+CH₃OH+Na]⁺: 307.1669, found: 307.1665.

E.2 Determination of the Kinetic Isotope Effect (KIE)

E.2.1. Intermolecular competition experiment

A 15 mL Schlenk tube was charged with enal **1a** (13.2 mg, 0.1 mmol), aminocatalyst (*S*)-**A** (14.1 mg, 20 mol%), zinc(II) triflate (18.2 mg, 50 mol%), and an equimolar solution of toluene (46.5 mg, 5 mmol) and toluene- d_8 (50 mg, 5 mmol) in dichloromethane (300 µL). The Schlenk tube was placed under an atmosphere of argon, cooled to – 78 °C, degassed *via* vacuum evacuation (5 min), backfilled with argon, and warmed to room temperature. The freeze-pump-thaw cycle was repeated five times, and then the Schlenk tube was sealed with Parafilm and placed into a 3D-printed plastic support mounted on an aluminium block fitted with a 420 nm high-power single LED (λ = 420 nm, irradiance = 45 mW/cm², as controlled by an external power supply). The reaction was stirred at ambient temperature (~35 °C) for 48 hours. The solvent was removed *in vacuo* and the crude mixture was purified by flash column chromatography (hexane/diethyl ether 92:8) to afford a mixture of product **2a** and **2a**-*d*. Integration of diagnostic peaks for **2a** in the ¹H NMR spectrum (specifically multiplets at 2.96 and 2.88, respectively) showed a deuterium incorporation at C**4** of 0.46D (over 2H, see figure below). Consequently, the magnitude of the kinetic isotope effect relative to the intermolecular competition was determined to be 3.3.



Figure S6. ¹H NMR analysis for the intermolecular competition experiment.

E.2.2. Parallel experiments



Two 5 mL argon-purged glass vials were charged with zinc(II) triflate (18.2 mg, 50 mol%), 300 μ L of a stock solution of enal **1a** (13.2 mg, 0.1 mmol) and aminocatalyst (*S*)-**A** (14.1 mg, 20 mol%) in dichloromethane and either toluene (92.0 mg, 1 mmol) or toluene-*d*₈ (118 mg, 1

mmol). The vials were further flushed with argon, sealed with Parafilm, and then simoltaneously placed into two 3D-printed plastic supports mounted on an aluminium block fitted with a 420 nm high-power single LED ($\lambda = 420$ nm, irradiance = 45 mW/cm², as controlled by an external power supply, see figure above). The reactions were stirred under visible light irradiation at ambient temperature for the time stated for each experiment. Trichloroethylene (9 µL, 0.1 mmol) was added as the internal standard to the crude mixture and an aliquot of the resulting solution was taken to be directly analysed by ¹H NMR. The magnitude of the kinetic isotope effect, calculated plotting the kinetic profile of the two reactions (with toluene and toluene-*d*₈, respectively), was determined to be 2.00.



F. References

- 1. Verrier, C.; Alandini, N.; Pezzetta, C.; Moliterno, M.; Buzzetti, L.; Hepburn, H. B.; Vega-Peñaloza,
- A.; Silvi, M.; Melchiorre, P. ACS Catal. 2018, 8, 1062.
- 2. Silvi, M.; Verrier, C.; Rey, Y. P.; Buzzetti, L.; Melchiorre, P. Nat. Chem. 2017, 9, 868.
- 3. He, M.; Bode, J. W. Org. Lett. 2005, 7, 3131.
- 4. Terao, J.; Todo, H.; Begum, S. A.; Kuniyasu, H.; Kambe, N. Angew. Chem., Int. Ed. 2007, 46, 2086.
- 5. Kumar, G. G. K. S. N.; Laali, K. K. Org. Biomol. Chem. 2012, 10, 7347.
- 6. Li, X.; Che, X.; Chen, G.-H.; Zhang, J.; Yan, J.-L.; Zhang, Y.-F.; Zhang, L.-S.; Hsu, C.-P.; Gao,
- Y. Q.; Shi, Z.-J. Org. Lett. 2016, 18, 1234.
- 7. Huang, L.; Olivares, A. M.; Weix, D. J. Angew. Chem., Int. Ed. 2017, 56, 11901.
- 8. Jiang, M. Y.; Dolphin, D. J. Am. Chem. Soc. 2008, 130, 4236.
- 9. Piccolo, O.; Spreafico, F.; Visentin, G.; Valoti, E. J. Org. Chem. 1987, 52, 10.

G. NMR Spectra



¹H NMR, 400 MHz, CDCl₃



¹³C NMR, 101 MHz, CDCI₃






















12.26-

1.5 1.0 0.5 0.0 -0





















 $^{13}\mathrm{C}$ NMR, 101 MHz, CDCI_3





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^{13}\text{C} NMR, 101 MHz, CDCl_3
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¹³C NMR, 101 MHz, CDCl₃



¹H NMR, 400 MHz, CDCl₃



¹H NMR, 500 MHz, CDCl₃




























S73











S77



H. HPLC and UPC² Traces







Conditions: UPC² (Daicel Chiralpak ID-3 column, 20 °C, gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 275$ nm)

Conditions: UPC² (Daicel Chiralpak IE-3 column, 20 °C, gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm)





Conditions: UPC² (Daicel Chiralpak ID-3 column, 20 °C, gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm)

Conditions: UPC² (Daicel Chiralpak IE-3 column, 20 °C, gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm)





Conditions: UPC² (Daicel Chiralpak IE-3 column, 20 °C, gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 2 mL/min, $\lambda = 210$ nm)

Conditions: UPC² (Daicel Chiralpak IE-3 column, 20 °C, gradient CO₂/MeOH from 100% CO₂ to 80:20 over 4 minutes, curve 6, flow rate 2 mL/min, $\lambda = 215$ nm)



Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 90:10 hexane/*i*-PrOH, flow rate 0.5 mL/min, $\lambda = 215$ nm)



Conditions: UPC² (Daicel Chiralpak IE-3 column, 20 °C, gradient CO₂/MeCN from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 229$ nm)





Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 90:10 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm)







Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 97:3 hexane/*i*-PrOH, flow rate 1.2 mL/min, $\lambda = 215$ nm)







Conditions: UPC² (Daicel Chiralpak CEL-1 column, 20 °C, gradient CO₂/*i*-PrOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm)



Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 95:5 hexane/*i*-PrOH, flow rate 0.5 mL/min, $\lambda = 215$ nm)



Conditions: UPC² (Daicel Chiralpak CEL-1 column, 20 °C, gradient CO₂/MeOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 210$ nm)

Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 90:10 hexane/*i*-PrOH, flow rate 1.2 mL/min, $\lambda = 215$ nm)





Conditions: UPC² (Daicel Chiralpak CEL-1 column, 20 °C, isocratic CO₂/*i*-PrOH 90:10 flow rate 3 mL/min, $\lambda = 220$ nm)



Conditions: UPC² (Daicel Chiralpak CEL-1 column, 20 °C, isocratic CO₂/*i*-PrOH 90:10 flow rate 3 mL/min, $\lambda = 205$ nm)



Conditions: UPC² (Daicel Chiralpak IC-3 column, 20 °C, gradient CO₂/*i*-PrOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 210$ nm)



Conditions: UPC² (Daicel Chiralpak IC-3 column, 20 °C, gradient CO₂/*i*-PrOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 210$ nm)



Conditions: UPC² (Daicel Chiralpak CEL-1 column, 20 °C, gradient CO₂/EtOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 215$ nm)



Conditions: UPC² (Daicel Chiralpak CEL-1 column, 20 °C, gradient CO₂/EtOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 215$ nm)

Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 80:20 hexane/*i*-PrOH, flow rate 1.0 mL/min, $\lambda = 215$ nm)





Conditions: UPC² (Daicel Chiralpak IC-3 column, 20 °C, gradient CO₂/*i*-PrOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm)



Conditions: HPLC (Daicel Chiralpak IC-3 column, 30 °C, 98.5:1.5 hexane/*i*-PrOH, flow rate 1.2 mL/min, $\lambda = 215$ nm)

Conditions: UPC² (Daicel Chiralpak ID-3 column, 20 °C, gradient CO₂/*i*-PrOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 273$ nm)



Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 96:4 hexane/*i*-PrOH, flow rate 0.8 mL/min, $\lambda = 215$ nm)







Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 90:10 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm)



Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 98:2 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm)





Conditions: UPC² (Daicel Chiralpak CEL1 column, 20 °C, isocratic CO₂/EtOH 95:5, flow rate 3 mL/min, $\lambda = 203$ nm)



Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 90:10 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm)

Conditions: HPLC (Daicel Chiralpak IC-3 column, 20 °C, 90:10 hexane/*i*-PrOH, flow rate 0.6 mL/min, $\lambda = 215$ nm)





Conditions: UPC² (Daicel Chiralpak CEL1 column, 20 °C, isocratic CO₂/MeCN 93.5:6.5, flow rate 3 mL/min, $\lambda = 205$ nm)

Conditions: UPC² (Daicel Chiralpak IB column, 20 °C, isocratic CO₂/EtOH 95:5, flow rate 3 mL/min, $\lambda = 201$ nm)





Conditions: UPC² (Daicel Chiralpak CEL-1 column, 20 °C, isocratic CO₂/MeOH 95:5, flow rate 3 mL/min, $\lambda = 210$ nm)