Supporting Information

Discovery of Novel Schizocommunin Derivatives as Telomeric G-quadruplex Ligands that Trigger Telomere Dysfunction and the Deoxyribonucleic Acid (DNA) Damage Response

Tong Che^a, Shuo-Bin Chen^a, Jia-Li Tu^a, Bo Wang^a, Yu-Qing Wang^a, Yan Zhang^a, Jing Wang^a, Zeng-Qing Wang^a, Ze-Peng Zhang^b, Tian-Miao Ou^a, Yong Zhao^b, Jia-Heng Tan^a, *, and Zhi-Shu Huang^a, *

^a Guangdong Provincial Key Laboratory of New Drug Design and Evaluation, School of Pharmaceutical Sciences, Sun Yat-sen University, Guangzhou, Guangzhou 510006, China

^b School of Life Sciences, Sun Yat-sen University, Guangzhou 510006, People's Republic of China

Table of Contents

Table S1	Oligomers used in this study	S3
Table S2	Stabilization activity of compounds on telomeric G-quadruplex	S 3
Table S3	Stabilization activity of compound 16 on various G-quadruplexes	S4
Table S4	Equilibrium binding constants (K_D) on various G-quadruplexes	S4
Figure S1	The X-ray crystallographic analysis of spectroscopy of compound 4	S4
Figure S2	The NOE spectroscopy of compounds 35 and 36	S5
Figure S3	The melting curves of compounds 7-21 by using FRET melting assays	S 6
Figure S4	The melting curves of compounds 22-36 by using FRET melting assays	S 7
Figure S5	The melting curves of compounds 37 and the reference compound 1 by using	S 8
	FRET melting assays	
Figure S6	The melting curves of compounds by using CD melting assays	S 8
Figure S7	The SPR sensorgrams and fitting curves of compounds 11-29	S9
Figure S8	The SPR sensorgrams and fitting curves of compounds 30-35	S10
Figure S9	TRAP gel for compound 7, 16 and 30.	S10
Figure S10	Immunofluorescence images of BG4 and TRF2 foci in U2OS cells	S11
Figure S11	Multiple cells images of TRF2 and BG4 foci in SiHa cells.	S11
Figure S12	Multiple cells images of TRF2 and γ -H ₂ AX foci in SiHa cells.	S12
Figure S13	Immunofluorescence images of γ -H ₂ AX and TRF2 foci in U2OS cells	S12
Figure S14	Immunofluorescence images of BG4 and γ -H ₂ AX foci in U2OS cells	S13
Figure S15	Expression of telomeric related proteins in SiHa cells	S14
Figure S16	Compound 16 induced cell cycle arrest and apoptosis in U2OS cells	S14
Figure S17	Cell cycle analysis of compound 7	S15
Figure S18	Antiproliferation activities of compound 16	S15
Methods		S16
¹ H NMR, ¹³ C I	NMR, HRMS and HPLC spectrum of final compounds	S18-S79

Oligomer	Sequence
F21T	5'-FAM-d(GGGTTAGGGTTAGGGTTAGGG)-TAMRA-3'
F10T	5'-FAM-d(TATAGCTATA-HEG-TATAGCTATA)-TAMRA-3'
ds26	5'-GTTAGCCTAGCTTAAGCTAGGCTAAC-3'
biotin-HTG21	5'-biotin-d(GGGTTAGGGTTAGGGTTAGGG)-3'
biotin-pu22	5'-biotin-d(TGAGGGTGGGGGGGGGGGGGGGGGGGGGGAA)-3'
biotin-c-kit1	5'-biotin-d(AGGGAGGGCGCTGGGAGGAGGG)-3'
biotin-bcl-2	5'-biotin-d(GGGCGCGGGAGGAAGGGGGGGGGGGG)-3'
biotin-Hairpin	5'-biotin-d(TATAGCTATA-HEG-TATAGCTATA)-3'
HTG21	5'-GGGTTTGGGTTTGGGTTTGGG-3'
pu22	5'-TGAGGGTGGGGGGGGGGGGGGAA-3'
c-kit1	5'-AGGGAGGGCGCTGGGAGGAGGG-3'
bcl-2	5'-GGGCGCGGGAGGAAGGGGGGGGGG-3'
HRAS	5'-TCGGGTTGCGGGCGCAGGGCACGGGCG-3'
TBA	5'-GGTTGGTGTGGTTGG-3'

Table S1. Oligomers used in this study.

Comm 1		$\Delta T_{\rm m}$ (°C)	
Compd.	FRET melting ^a	CD melting ^b	
7	8.4	3	
8	11.8	11	
9	10.0	8	
16	23.5	20	
22	9.0	7	
27	23.1	20	
30	24.6	21	

Table S2. The stabilization activity of compounds on telomeric G-quadruplex.

 ${}^{a}\Delta T_{\rm m} = T_{\rm m}$ (DNA + ligand) - $T_{\rm m}$ (DNA). $\Delta T_{\rm m}$ values for the incubation of 0.2 μ M F21T with and without 1.0 μ M of the compound in Tris-HCl buffer (10 mM, pH 7.4) containing 60 mM KCl. In the absence of a ligand, the $T_{\rm m}$ values of annealed F21T is 60 °C. ${}^{b}\Delta T_{\rm m}$ values for incubation of 2 μ M HTG21 with and without 10 μ M compound in Tris-HCl buffer (10 mM, pH 7.4) containing 60 mM KCl. In the absence of ligand, $T_{\rm m}$ values of annealed HTG21 is 67 °C.

	$T_{ m m}$	(°C) ^a	
DNA —	16 (0 µM)	16 (10 µM)	$\Delta I_{\rm m}$ (C)
HTG21	67	87	20
pu22	82	95	13
c-kit1	64	78	14
bcl-2	87	95	8
HRAS	62	76	14
TBA	54	60	6

Table S3. The stabilization activity of compound 16 on various G-quadruplexes.

^a Melting temperature (T_m) of DNA samples with and without compounds in 10 mM Tris-HCl (2.5 mM KCl for pu22, c-kit1 and bcl-2; 60 mM KCl for HTG21, HRAS and TBA) buffer detected by circular dichroism spectroscopy.

Compound	DNA	<i>K</i> _D (M)
16	HTG21	8.1×10 ⁻⁶
	pu22	1.2×10 ⁻⁵
	c-kit1	7.8×10 ⁻⁶
	bcl-2	1.1×10 ⁻⁵

Table S4. Equilibrium binding constants (*K*_D) on various G-quadruplexes.



Figure S1. The X-ray crystallographic analysis of spectroscopy of compound 4. (reference: Uehata, K.; Kimura, N.; Hasegawa, K.; Arai, S.; Nishida, M.; Hosoe, T.; Kawai, K.; Nishida, A. Total synthesis of schizocommunin and revision of its structure. *J. Nat. Prod.* 2013, *76*, 2034-2039.)



Figure S2. The NOE spectroscopy of compounds 35 (A) and 36 (B).



Figure S3. The melting curves of compounds $7 \sim 21$ by using FRET melting assays.



Figure S4. The melting curves of compounds $22 \sim 36$ by using FRET melting assays.



Figure S5. The melting curves of compounds 37 and the reference compound 1 by using FRET melting assays.



Figure S6. The melting curves of compounds 7, 8, 9, 16, 22, 27, 30 by using CD melting assays.



Figure S7. The SPR sensorgrams and fitting curves of compounds (A. compound 11. B. compound 16. C. compound 17. D. compound 18. E. compound 19. F. compound 26. G. compound 27. H. compound 28. I. compound 29).



Figure S8. The SPR sensorgrams and fitting curves of compounds (A. compound 30. B. compound 31. C. compound 32. D. compound 33. E. compound 34. F. compound 35).



Figure S9. TRAP gel for compounds 7, 16 and 30.



Figure S10. Compound **16** stabilized and induced the formation of telomeric G-quadruplexes in U2OS cells. (A) Representative immunofluorescence images of TRF2 (red) and BG4 (green) foci in U2OS cells (B) Quantification of the number of BG4 foci per nucleus in U2OS cells. (C) Quantification of the co-localization of BG4 and TRF2 per nucleus in U2OS cells.



Figure S11. Multiple cells immunofluorescence images of TRF2 (green) and BG4 (red) foci in SiHa cells.



Figure S12. Multiple cells immunofluorescence images of γ -H2AX (green) and TRF2 (red) foci in SiHa cells.



Figure S13. Compound **16** induced telomeric DNA damage in U2OS cells. (A) Representative immunofluorescence images of γ -H2AX (green) and TRF2 (red) foci in U2OS cells (B) Quantification of the number of γ -H2AX foci per nucleus in U2OS cells. (C) Quantification of the co-localization of γ -H2AX and TRF2 per nucleus in U2OS cells.



Figure S14. (A) Representative immunofluorescence images of γ -H2AX (green) and BG4 (red) foci in SiHa cells treated with 2 μ M 7, 2 μ M 16 or 0.1% DMSO (control) for 24 h. The nuclei were stained with DAPI (blue). (B) The quantification of γ H2AX foci number per nucleus. (C) The quantification of co-localization of BG4 and γ H2AX per nucleus.



Figure S15. Expression of telomeric related proteins in SiHa cells. Cells were treated with 1 μ M 16 or 0.1% DMSO for 48 h.



Figure S16. Compound **16** induced cell cycle arrest and apoptosis in U2OS cells. (A) Cell cycle analysis after propidium iodide (PI) staining after a 12-h treatment with **16** or 0.1% DMSO in U2OS cells. (B) The percentage of U2OS cells in different phases of the cell cycle, analyzed by EXPO32 ADC software. (C) Apoptosis evaluation of U2OS cells after a 96-h treatment with **16**. The cells were collected and stained with Annexin V-FITC and PI. (D) The percentage of apoptosis in U2OS cells, analyzed by EXPO32 ADC software.



Figure S17. Cell cycle analysis of compound 7. (A) Cell cycle analysis after PI staining after a 12-h treatment with 7 or 0.1% DMSO in SiHa cells. (B) The percentage of SiHa cells in different phases of the cell cycle, analyzed by EXPO32 ADC software.



Figure S18. Antiproliferation activities of compound **16**. (A) Colony formation of SiHa cells in the presence of **16**. Cells were plated in a six wells dish with increasing concentrations of **16** for 12 days and stained with crystal violet before counting. (B) Quantification of colony numbers, the value of group without compound was set as 100%. (C) RTCA analysis of SiHa cells with **16** treatment. The growth curves of cells were obtained from the RTCA software 1.1.2.

Methods

CD melting assay. CD studies were performed on a Chirascan circular dichroism spectrophotometer (Applied Photophysics, UK). A quartz cuvette with a 4 mm path length was used to record the spectra over a wavelength range of 230-330 nm with a 1nm bandwidth, 1 nm step size and a time of 0.5 s per point. A CD melting assay was performed at a fxed concentration of G-quadruplex (2 μ M), either with or without a fixed concentration (10 μ M) of compounds in Tris-HCl buffer (10 mM, pH 7.4) with 60 mM KCl. The data were recorded at intervals of 2.5 °C over the range 25-95 °C with a heating rate of 1°C/min. A buffer baseline was collected in the same cuvette and was subtracted from the sample spectra. The final analysis of the data was conducted using Origin 9.0 (OriginLab Corp.)

TRAP-LIG Assay. The ability of schizocommunin derivatives to inhibit telomerase in a cell-free system was assessed with the TRAP-LIG assay following previously published procedures.⁴⁷ Protein extracts from exponentially growing SiHa cells were used. Briefly, 0.1 µg of TS forward primer (5'-AATCCGTCGAGCAGAGTT-3') was elongated by telomerase (500 ng protein extract) in TRAP buffer (20 mM Tris-HCl [pH 8.3], 68 mM KCl, 1.5 mM MgCl₂, 1 mM EGTA, and 0.05% Tween 20) containing 125 µM dNTPs and 0.05 µg BSA. The mix was added to tubes containing freshly prepared ligand at various concentrations and to a negative control containing no ligand. The initial elongation step was carried out for 30 min at 30 °C, followed by 94 °C for 5 min and a final maintenance of the mixture at 10 °C. Then the ligands were removed by phenol extraction and ethanol precipitation. The purified extended samples were then subject to PCR amplification. For this, a second PCR master mix was prepared consisting of 1 µM ACX reverse primer (5'-GCGCGG[CTTACC]3CTAACC-3'), 0.1 µg TS forward primer (5'-AATCCGTCGAGCAGAGTT-3'), TRAP buffer, 5 µg BSA, 0.5 mM dNTPs, and 2 units of Taq polymerase. A 10 µL aliquot of the master mix was added to the purified telomerase extended samples and amplified for 35 cycles of 94 °C for 30 s, of 61 °C for 1 min, and of 72 °C for 1 min. Samples were separated on a 12% PAGE and visualized with EB. TelIC50 values were then calculated from the optical density quantitated from the AlphaEaseFC software.

Colony formation assay. SiHa cells were seeded on 6-well plates (300/well) and treated with compound **16** or DMSO. DMEM was replaced and different concentrations of **16** were added every 2 days. Cells were fixed with methanol and dyed with crystal violet after cultured 12 days. The pictures were taken using a x40 magnification using a FL Auto Imaging System EVOS microscope (Life Technologies).

Real-Time Cellular Activity Assay. Compound-mediated cytotoxicity was monitored using a real-time cellular analysis (RTCA) multiplate instrument, xCELLigence RTCA DP instrument (Roche Applied Science, Indianapolis, IN), which was placed in a humidified incubator at 37 °C and 5% CO2. This system monitors cellular events (cell viability, cell number, cell morphology, and degree of adhesion) in real time measuring electrical impedance across interdigitated microelectrodes integrated on the bottom of tissue culture E-Plates 16-well plates (Roche Applied Science, Indianapolis, IN). The evaluation of cellular events was performed on SiHa cells. For time-dependent cell response profiling, 50 µL of media was added to 16-well electric plates (E-Plates) to obtain background readings followed by the addition of $100 \ \mu L$ of cell suspension at the density of 20,000 cells/mL. After seeding, cells were allowed to settle for 20 min at room temperature before being inserted into the xCELLigence RTCA DP instrument. After 24 h, the cells were treated compounds or DMSO at various concentrations. The cells were monitored every 2 min for duration of 2 h after compounds addition to capture the short-term response and for every 15 min from 2 h after compound addition until the end of the treatment to capture the long-term response. Results were plotted using the RTCA software 1.1.2 (Roche Applied Science, Indianapolis, IN). The data expressed in cell index (CI) units was exported to Microsoft Excel software (Microsoft, Redmond, WA) for mathematical analysis and normalization.



¹H NMR, ¹³C NMR, HRMS and HPLC spectrum of final compounds



HRMR spectrum of 7



HPLC analysis of 7



S20





Peak#	Ret. Time	Area	Height	Area %	Height %
1	6.943	4759939	412739	4.174	9.358
2	8.715	109278265	3997675	95.826	90.642
Total		114038204	4410413	100.000	100.000

HPLC analysis of **8**



S22





DA Ch1 25	54nm 4nm		I Cak I	able	
Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.425	9313964	1182302	96.008	97.621
2	10.555	387291	28808	3.992	2.379
Total		9701255	1211110	100.000	100.000

HPLC analysis of **9**





HRMR spectrum of 10



DA Ch1 25	4nm 4nm					
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	5.136	35991	4035	0.196	0.652	
2	5.675	79626	6078	0.433	0.983	
3	6.003	29897	2034	0.163	0.329	
4	9.368	14481	102	0.079	0.016	
5	9.741	18113646	594061	98.506	96.044	
6	15.223	114810	12221	0.624	1.976	
Total		18388450	618532	100.000	100.000	

HPLC analysis of **10**



¹³C NMR spectrum of **11**



HRMR spectrum of 11



DA Chi 25	Anm Anm		Peal	Table	
Peak#	Ret. Time	Area	Height	Area %	Height %
1	7.043	6992016	476396	95.065	96.952
2	10.231	146996	4287	1.999	0.872
3	13.503	91924	5473	1.250	1.114
4	16.150	124020	5217	1.686	1.062
Total		7354955	491372	100.000	100.000

HPLC analysis of 11



S28





PDA Ch1 254nm 4nm						
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	4.663	955157	101179	2.270	2.633	
2	4.928	41119741	3741450	97.730	97.367	
Tota	1	42074898	3842629	100.000	100.000	

HPLC analysis of 12







HPLC analysis of 13





HRMR spectrum of 14



PDA Ch1 25	4nm 4nm		Pea	kTable	
Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.654	66759	7759	0.466	0.545
2	5.030	14102248	1410423	98.522	99.095
3	6.255	144761	5118	1.011	0.360
Total		14313769	1423299	100.000	100.000

HPLC analysis of **14**



S34





HPLC analysis of 15




HRMR spectrum of 16



DA Ch1 254nm 4nm					
Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.651	14071	2026	0.146	0.201
2	4.976	9641428	1004540	99.854	99.799
Total		9655499	1006566	100.000	100.000

HPLC analysis of **16**





HPLC analysis of 17







HPLC analysis of 18





HRMR spectrum of 19



HPLC analysis of 19





HRMR spectrum of 20



DA Ch1 25	4nm 4nm		100	ik l'abie	
Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.689	28606	3281	0.497	0.589
2	5.043	5708453	553530	99.166	99.283
3	7.242	19397	715	0.337	0.128
Total		5756456	557526	100.000	100.000

HPLC analysis of **20**





HRMR spectrum of 21



DA Ch1 25	254nm 4nm					
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	5.403	135067	13620	1.443	2.049	
2	6.043	213676	16215	2.282	2.440	
3	7.442	9014609	634837	96.275	95.511	
Total	1000	9363352	664672	100.000	100.000	

HPLC analysis of **21**





HRMR spectrum of 22



DA Ch1 25	4nm 4nm		Peak	Table	
Peak#	Ret. Time	Area	Height	Area %	Height %
1	9.427	6889193	607130	95.474	97.275
2	15.206	326595	17007	4.526	2.725
Total		7215788	624137	100.000	100.000

HPLC analysis of **22**





HRMR spectrum of 23



HPLC analysis of 23

Total





HRMR spectrum of 24



HPLC analysis of 24



S54





HPLC analysis of 25







DA Ch1 25	4nm 4nm		Pea	kTable	
Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.916	6353684	675894	98.852	99.239
2	5.584	33310	2501	0.518	0.367
3	6.851	7094	732	0.110	0.107
4	9.446	33356	1947	0.519	0.286
Total		6427445	681074	100.000	100.000

HPLC analysis of **26**







HPLC analysis of 27





HRMR spectrum of 28



DA Ch1 25	54nm 4nm					
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	4.728	48308	6041	1.209	1.425	
2	5.045	3939325	416190	98.574	98.156	
3	9.653	8693	1777	0.218	0.419	
Total		3996325	424008	100.000	100.000	

HPLC analysis of **28**







DA Ch1 25	4nm 4nm	kTable			
Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.807	143057	16346	1.417	1.785
2	5.065	9885045	895604	97.910	97.782
3	7.315	67956	3968	0.673	0.433
Total		10096059	915918	100.000	100.000

HPLC analysis of **29**



S64



HRMR spectrum of 30



DA Ch1 25	4nm 4nm		Peak	Table	
Peak#	Ret. Time	Area	Height	Area %	Height %
1	4.622	32466	6324	0.169	0.333
2	4.826	19181353	1891340	99.831	99.667
Total		19213819	1897663	100.000	100.000

HPLC analysis of **30**



S66



HRMR spectrum of 31



HPLC analysis of **31**

3279898

Total







DA Chi 25	Anm Anm	PeakTable				
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	4.697	56410	5248	0.887	0.825	
2	5.009	6223062	623804	97.868	98.076	
3	5.658	9136	928	0.144	0.146	
4	6.075	8020	864	0.126	0.136	
5	7.166	28159	2068	0.443	0.325	
6	9.673	33849	3132	0.532	0.492	
Total		6358636	636044	100.000	100.000	

HPLC analysis of **32**





HRMR spectrum of 33



HPLC analysis of **33**




HRMR spectrum of 34



DA Ch1 25	A Ch1 254nm 4nm					
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	4.689	28606	3281	0.497	0.589	
2	5.043	5708453	553530	99.166	99.283	
3	7.242	19397	715	0.337	0.128	
Total		5756456	557526	100.000	100.000	

HPLC analysis of **34**





HRMR spectrum of 35



HPLC analysis of 35





HRMR spectrum of 36



DA Ch1 25	4nm 4nm	and the second second				
Peak#	Ret. Time	Area	Height	Area %	Height %	
1	4.864	62872902	3991166	99.234	99.711	
2	6.394	485643	11553	0.766	0.289	
Total		63358545	4002719	100.000	100.000	

HPLC analysis of **36**









HPLC analysis of **37**