

Mutations in the *fabI* promoter associate to other mechanisms of triclosan resistance in *Staphylococcus aureus*

Grandgirard D¹, Furi L^{2,3}, Cuisa M^{1,2}, Baldassarri L⁴, Knight DR⁵, Morrissey J⁶, Morrissey J⁶, and Oggioni M^{2,3}

¹Neuroinfection Laboratory, Institute for Infectious Diseases, University of Bern, Switzerland; ²La Microbiologia Molecolare e Biotecnologia, Dip. Biologia Molecolare, Università di Siena, Italy; ³Department of Genetics, University of Leicester, United Kingdom; ⁴Istituto Superiore di Sanità, Rome, Italy; ⁵The University of Western Australia, Nedlands, Western Australia; ⁶Immunology, University of Leicester, United Kingdom; ⁷Center of Laboratory Medicine, Institute of Clinical Chemistry, Inselspital, University of Bern, Switzerland; ⁸Swiss Agency for Research in Innovation, Federal Office for Culture Protection (COP), Switzerland.

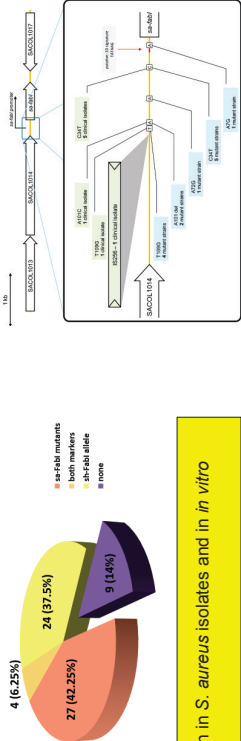
BACKGROUND: The biocide triclosan is a chlorinated bis-phenol with a broad spectrum antimicrobial activity. Triclosan, unlike other biocides, at low concentrations has a single intracellular target identified so far: the enoyl-acyl carrier protein (ACP) reductase enzyme (*FabI*), involved in bacterial fatty acid biosynthesis. From a collection of 1600 *S. aureus* isolates, we have previously shown that 64 strains demonstrated an increased MBC towards triclosan. Mutations in *fabI* gene locus and heterodiploidy for *fabI* have been shown to confer resistance in *S. aureus*. Still, reduced susceptibility in 14% of these strains could not be attributed to one of these mechanisms.

AIM: The present study investigates the molecular nature of *fabI* overexpression in *S. aureus* isolates and in *in vitro* selected mutants with reduced susceptibility to triclosan.

METHODS: Promoter analysis was performed on 38 triclosan resistant clinical strains and 23 laboratory mutants selected after multipassage exposure to triclosan. The promoter region was amplified by PCR and sequenced. Furthermore, 7 laboratory mutants and their 3 corresponding parental strains were subjected to all genome sequencing. The gene expression profile of 4 clinical strains and 4 laboratory mutants with mutations in the promoter regions was compared to their wild type, triclosan susceptible isogenic or prototypical strains. For this purpose we used a self-designed custom array containing probes targeting genes extracted from 2 different *S. aureus* genomes and a variety of genes of plasmid origin.

RESULTS

Strain	RNA228 derived				ATCC35968 derived				Comment
	POS*	MUT	POS	MUT	POS	MUT	POS	MUT	
208002 T-A									
208027 C-A									
208092 T-C									
919997 C-T									
920008 C-C									
2461395 C-A									
2737373 T-G									



Some of the mutations identified in the clinical isolates were detected also in the series of laboratory mutants, indicating that in this case *in vitro* model mimics selective pressure in the field.

The comparisons between *in vitro* selected mutants and their isogenic wild type strains identified *fabI* as one of the few significantly up-regulated genes. A similar comparison between triclosan resistant clinical isolates containing mutations in the *fabI* promoter and sensitive prototypical strains also demonstrated a significant up-regulation of the *fabI* gene. In all 4 clinical isolates, 37 genes were commonly up-regulated and 6 down-regulated (see following table). Furthermore, one clinical strain showed a particularly high level of *fabI* gene expression and associated triclosan resistance. This strain was characterized by the insertion of an IS256 element in the promoter region of *fabI*.

Protein product	Gene locus TW20	Gene locus MU50	Fold change	Protein product	Gene locus TW20	Gene locus MU50	Fold change
DNA gyrase subunit A	SATW20_00060 / gyrA	SAW006 / gyrA	2.65	Spolysaccharide complement inhibitor SCIN	SATW20_19360 / scin	SAV042 / scin	65.23
Putative flavohemoglobin	SATW20_03420	SAW040	17.18	Uncharacterized protein	SATW20_20150	SAV0332	3.11
Uncharacterized protein	SATW20_04160	SAW048	2.49	UDP-N-acetylglucosamine-1-carboxymethyltransferase	SATW20_23360 / murA1	SAV009	8.59
Xanthine phosphoribosyltransferase	SATW20_04540 / xprT	SAW038 / xprT	9.9	Transcription termination factor rho	SATW20_23590 / rho	SAV021 / rho	3.78
Putative arylalanine (N-acetylurea)-alanine amidase	SATW20_05330	SAW065	8.04	Uncharacterized protein	SATW20_23220	SAV0238	5.52
Uncharacterized protein	SATW20_07380	SAW063	8.31	Xanthine (uracil) permease family protein	SATW20_23870	SAV0253	3.62
Fructose 1-phosphate kinase	SATW20_07740	SAW069 / frp	3.41	lysopaglin resistance protein A	SATW20_24670	SAV0235	2.79
PTS transport system, fructose-specific	SATW20_07750 / fruA	SAW070 / fruA	8.11	Putative lipoprotein	SATW20_25000	SAV0268	8.9
IMCComponent	SATW20_08060 / rpi1	SAW073 / rpi1	2.75	Uncharacterized protein	SATW20_25130	SAV0283	4.44
Ribonucleoside-diphosphate reductase	SATW20_09350 / rdt	SAW0935 / rdt	4.13	Putative nitrite transporter	SATW20_25330	SAV0403	3.72
Thioesterase superfamily protein	SATW20_09840	SAW0944	4.29	Uncharacterized protein	SATW20_25340	SAV0404	5.28
Nitric oxide synthase reductase (NADH)	SATW20_10000 / nsi	SAV011 / nsi	15.81	ABC transporter ATP-binding protein	SATW20_25420	SAV0412	15.01
Sodium alanine symporter family protein	SATW20_13570	SAV036 / spt	2.88	ABC transporter permease	SATW20_25430	SAV0413	7.39
Phosphatidylglycerol lysoATPase	SATW20_15600	SAV037	5.44	ABC transporter valine-binding lipoprotein	SATW20_25440	SAV0414	3.15
UPF0585 protein	SATW20_18860 / cypA	SAV086 / cypA	4.23	Immunodominant staphylococcal antigen B	SATW20_27160 / iab	SAV0368 / iab	3.28
Signal transduction protein TRAP	SATW20_18290 / trp	SAV035 / trp	14.65	Down-regulated genes			
Uncharacterized protein	SATW20_18470	SAV053	2.8	Uncharacterized protein	SATW20_27810	SAV0801	0.15
Nitric oxide synthase oxygenase	SATW20_19090	SAV034	4.94	Putative phage infection protein	SATW20_27830	SAV0843	0.37
Uncharacterized protein	SATW20_19310 / nb	SAV034	16.6	Protease-bisphosphate adenosine can 1	SATW20_27850 / fab	SAV066	0.03
Phage protein	SATW20_19350 / nb	SAV034	5.32	Putative hydrolase	SATW20_28020	SAV021	0.23
	SATW20_19350		10.12	Putative transmembrane protein ompB	SATW20_28400	SAV0515	0.06
					SATW20_35360	SAV0515	0.36

CONCLUSION: We have shown that, C34T, T109G and A101C mutations in the *fabI* promoter region confer *fabI* up-regulation both in clinical isolates and/or laboratory mutants. The insertion of IS256 may further enhance promoter activity. This is the first report on genetic evidence linking promoter mutations and up-regulated expression of the *fabI* gene.

Contact: Denis Grandgirard, Ph.D., Neuroinfection Laboratory, Institute for Infectious Diseases, University of Bern, Friedbühlstrasse 51, 3010 Bern, Switzerland, denis.grandgirard@fki.unibe.ch.