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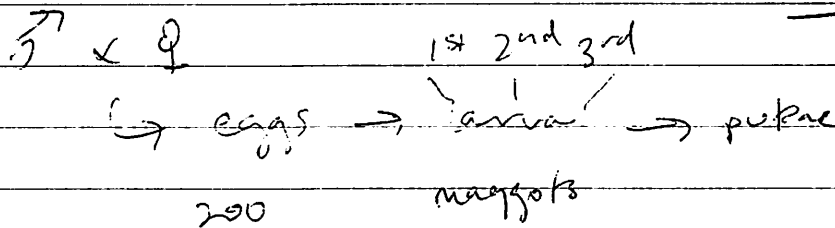
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Notes

Wellcome Trust Advanced course

- Red-book 1st ed, last pre-molecular mutant list
- shows phenotypes of mutants
- ^{easy} short life cycle, high fecundity (200+ offspring/♀)
- safe, not a pest, no allergies, easy to work with
- eukaryotic X, higher evk.
- = simple genome, allows rearrangement
- many μ , 1/3 current, maybe a mutant in every gene
- application to human biology
- cheap, Falk anecdote (garage)

Fly husbandry



- 8 hrs until receptive (not 10-12!)
- older male will "rape" 5-6 ^{12 day} virgins

- critical role of "virginity"

1) clear to look < 8 hrs

2) use only young virgins

3) hold virgins for 1-2 days, look for larval tracks
♀ will lay eggs

4) use μ stocks that show contamination

Desplan 5) μ (yrid) stocks to virgin, hsp promoter
sel-gfp embryo sorting (all ♂ die)

? 18° overnight → Scott says don't trust it?

- ♀ store sperm, requires virginity

- keep 2 copies of stocks, different places

- mites: can alter results of cross

jump from ♀ → eggs & kill eggs

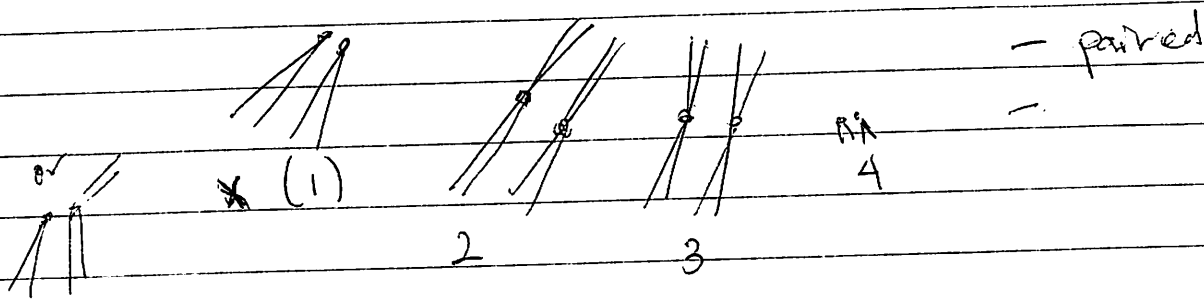
- < 21 days all vials

- 18 days, 24.5°C

- dark vials

→ y chrom
genes

- Ms. Stevens worked out karyotype of Dros.



- 13th nuc. division pair → stay paired
- or it pairs in G1 in each cell division
- 3rd instar larva acetic acid ^{stain} / orcein
- triploids are viable, fertile
- genes & chromosomes
- Mendel, Moravian monk

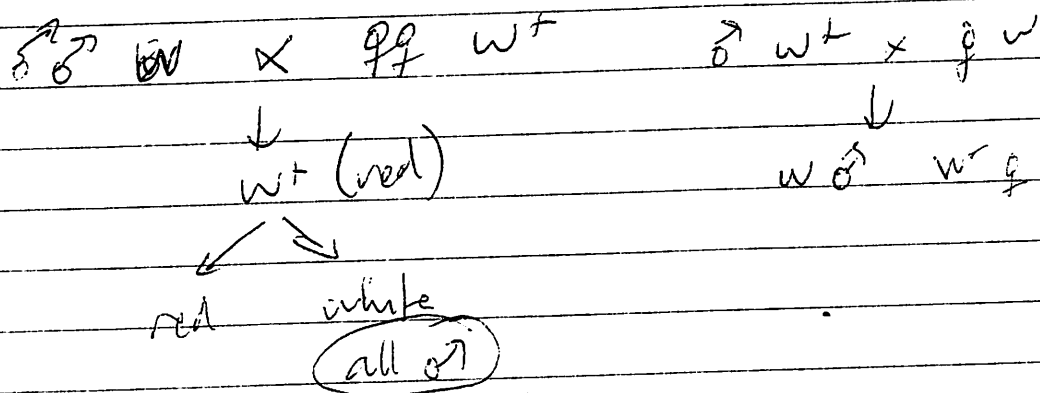
- 1) purity & consistency of gene
- 2) law of the gene (2 copies) ^{particulate inheritance}
- 3) independent assortment

- ebony follows Mendel's rules
- early μ , white didn't follow rules
- white → nomenclature

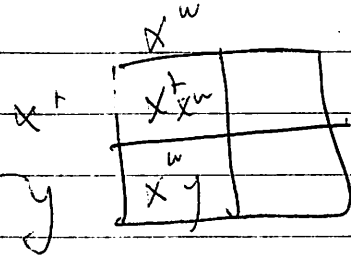
WW normal nomenclature
W⁺w Flies

had system - >1 allele, some dom/some recessive
- case sensitive

- wt doesn't mean wild, it means the stock doesn't carry μ allele



- one of the first μ didn't follow Mendel's rule
- Stevens solved riddle via cytology
- eg. color vision



Widengraf project

obly score 100 to demonstrate sex linkage

- in order to explain chromosomal theory, needed to use a violation of Mendel's law.
- how do we prove this hypothesis?
- Bridges, 48 yrs
 - greatest geneticists w/ McClintock
 - McClintock described as Greek God.
- Dean Parker did μ work for Muller's Nobel prize, irradiation/ice block, white μ do w/ 2000-3000 progeny
 - 2998 red μ / white σ
 - 1 red σ
 - 1 white μ

? would you have recognized this result as important, or contamination of parents?

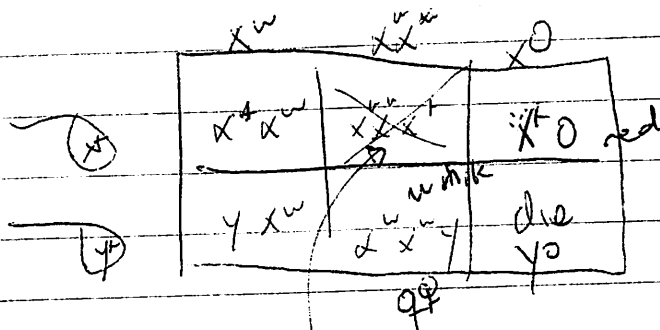
Bridges repeated expt & found repeatable frequencies.

- not parents
- red σ is sterile, not parent
 - white μ , lead to high freq of white daughters
 - didn't always come on 1st day of cross

- first has to solve XX & sex differentiation
- second, errors in meiosis
- third, XXX diploid abnormal not OK
- exception proves the rule

het seg

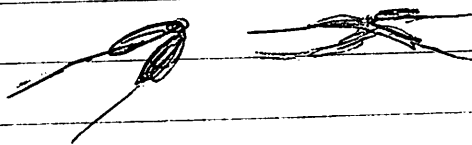
- bridges associated nondisjunction, go to same pole
- homologs pair \rightarrow recombine \rightarrow segregate
match lock move



Bridges predicted triploid X dies but

- recessive/dominant
 - most genes are limited by substrate not by enzyme
 - structural genes can be dominant since amount of product matters
- Nattie Stevens did adult male spread from oogenesis showed XXY ♀ are triploid
- proves chrom theory XXY is exception to Mendel
- begins field of cytogenetics
- identified nondisjunction which is emp for birth defects in humans (trisomy 21)

Cytology — larval brain mitotic



ev - 98-99% of genes
het - highly repet, simple seq.

60-70% ev
30-40% het

100 / 1000
3 / 1000
150 / division

polytene rep & seg decoupled

2 → 1028, no cell division

differences in degree of replication

60% of genome ~~polytene~~ ^{next} does polytenize

- nomenclature

- 2000 bands per chrom arm
100 on 4th

50 bands / division, 20 divisions / chrom

- deletion mapping link physical ^{cytological} map & genetics

- 1 gene / 1 band

exceptions < , >
are bands missed

= triploids, chrom 4 will x-over, also heat shock

- 4 does ~~not~~ x-over in ♀

- ♂ no recombination

LV Morgan ♂ w^+/y × ♀ w^+/w^+

- lost & found fly
↓
 w/w ♀ xxy

backcross to w^+/y ♂ ↑ nondisjunction 4-10%

" secondary nondisjunction "

- 1916 problem stated

- but w/ LV Morgan's fly

x^+/y - $x^w x^w y$

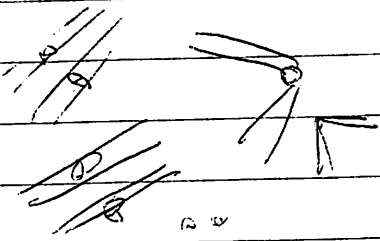
↓

♂ all red

♀ all white

100% nondisjunction

- N. Stevens solves problem thru aptogenics



attached X or
compound X

$\hat{X}X$

C(1)RM

around 1st reverse mechanism

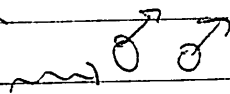
- chromosome mechanics in 1950's/60's game to create all types of compounds

- Dattell Falk

- EMS (warnings, melanomas) high efficiency

X-rays - messy

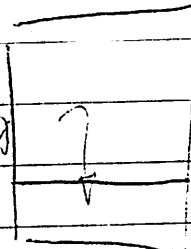
transposons - weak hypomorphs



$\hat{X}X/Y$

X^*/Y

20°



1hr
29°C

20°

those that

fly are T_{H1}

T_{H1} paralytic

UV bloom
open same color

X^{TSP}/Y

$\hat{X}X/Y$

- compound X, pure transmission $\sigma \rightarrow \sigma$ useful
for μ screens & making stocks

Balancer Muller

wanted to show X-ray cause recessive lethals

X/Y

3500 Rads

X^*

minimal effect on
fertility/viability

(550 Rads LD 50
human in 30 days
Kills immune cells)

X^*/X

how to keep μ (test each generation)

- need to keep, & keep from recombination w/ homolog

Balancer

- 1) inversions suppress recombination of recombinants
- 2) carry a strong dominant μ

2a) carry at least 1 recessive lethal μ
not true for X chrom (recess. female sterile)

~~()~~

~~m() () m~~ base (B, ac, sc)
Muller's 5 (M-5)

= people use FMT not M5

- "M-5 test" (can be used like Ames test)

$X^*Y \times M5/M5$ < - many no rec lethal

\downarrow

$X^*/M5$

$M5/Y$

cross inter se

look for round eyed σ^7

yes \rightarrow no μ

no \rightarrow ~~no~~ μ on ~~X~~ recessive lethal

- allows lethal to be kept & homozygotes to be made

\rightarrow variant of FMT with sn (q sterile)

- "new" generation of balancers w/ Δ -GFP

- use to test genotype of embryonic lethal

- Now Zuker Plan

= 2-3 th

- Zuker lines have secondary μ

- autosomal balancers suppress recovery of recombinants, X chrom suppress recomb directly

* polymorphism can prevent genetic-based gene disruption

- Monica Justice, cre-lox in mouse

22/6/06 - Mapping

- X-linked lethals can do complementation testing

- recombination mapping shuckvant (although)

Muller claimed he explained data)

old Mary map
Y chrom story
50% recombination

♀ mirror of
verus
→ sword & shield
of Mars

y w ct m f
Y

X
TMA7
ff

♂♂
X

y w ct m f dent
+ virgin

0% 1 X-over
30% 2 X-over
5% 0 X-over
41% 1 X-over

typical ♀ meiosis

some all
single
xovers

| | | | | | | |
|---|---|----|---|---|-----|-----|
| y | w | ct | m | f | 200 | 400 |
| + | + | + | + | + | 200 | |
| y | + | + | + | + | 5 | 10 |
| + | w | ct | m | f | 5 | |
| y | w | + | + | + | 45 | 90 |
| + | + | ct | m | f | 45 | |
| y | w | ct | + | + | 125 | 250 |
| + | + | + | m | f | 125 | |
| y | w | ct | m | + | 125 | 250 |
| + | + | + | + | f | 125 | |

score
sons 3

? why 3 major classes non xover f 1.2 X-over
per meiosis & only 5% non xover?

- Haldane's Rule all meiosis have one xover per
chrom

- map distance & physical distance is a lie!

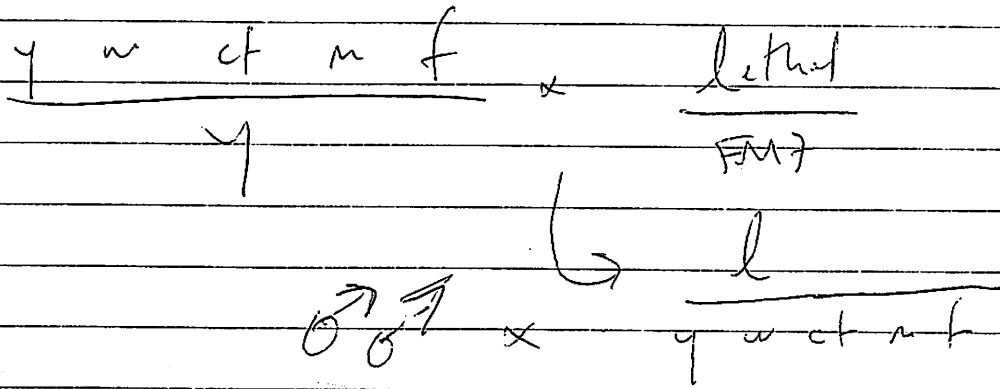
→ A! bc xover occurs at 4 strand stage

- 4 strand stage any 2 non sister chromatids (Sunder)
- genetic maps measure freq of recombination

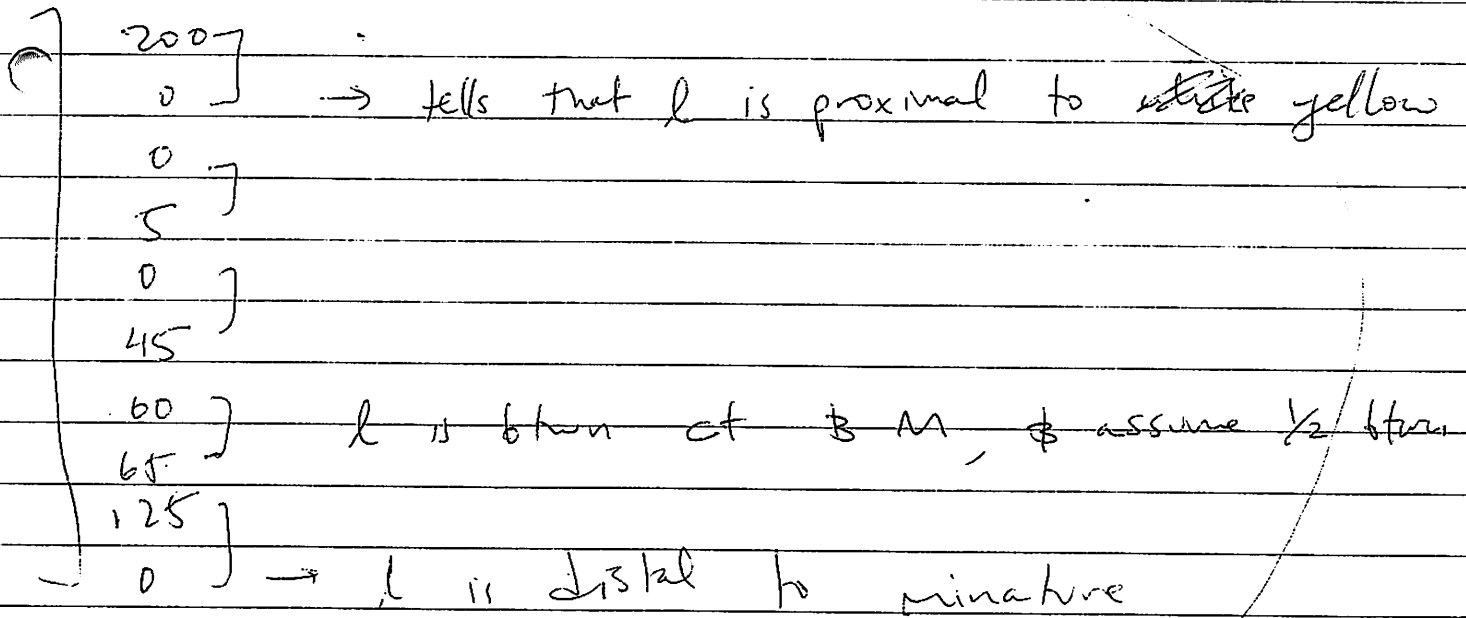


map length = recomb fraction
1 map unit = 1% recomb

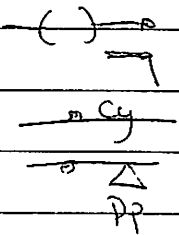
Cyo - Erwin Oster



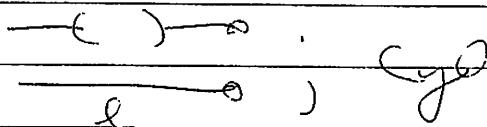
alternative strategy is move work \rightarrow df mapping ^{Bridges & Painter}
 - but no counting!



$\times Df/y; cyo/Dp \times l/FM7$



$l/Df \rightarrow \text{♀♀ } cyo$



- balancers makes this possible

- SNP mapping, induces lethal on known chrom

- 300 piggy, all non-lethal
- single fly prep, seq center
- large # of loci

- complementation test set of new μ lethal mapping minimal set

min. class
Dg centromere

- Robert did F1 dominant on /y/p screens

Chromosome Aberrations

Inversions

a b c d e f g h
a b g f e d c h

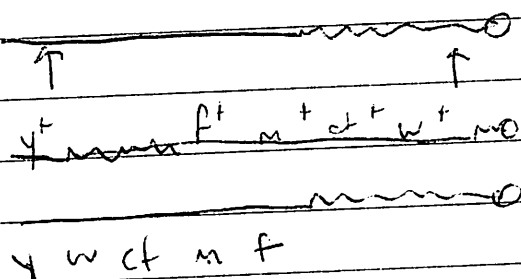
- nothing lost, nothing gained
- pericentric includes centromere
- paracentric ~~not~~ centromere not in inversion
- size also imp't, large $> 3/4$ arm 15-20 # units
small < 10 units $< 1/2$ chrom arm

- balancers have combination

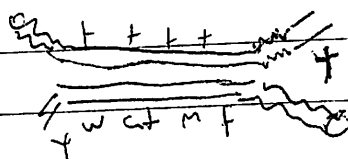
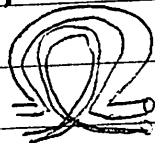
x - 1 large, > 2 small para
auto combine per. + > 2 para

large para

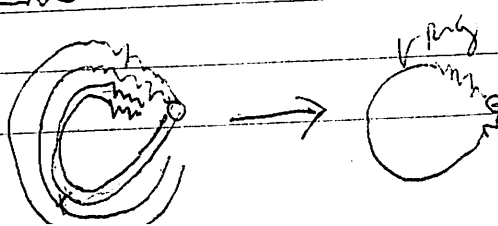
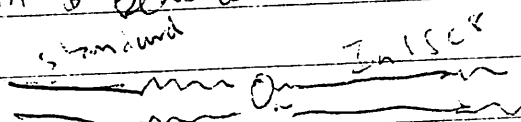
In(1) sc⁸



inversion loop



- no single recombinants, why? acentric/dicentric
- same fertility
- maybe just xovers don't occur?
no, double xovers can be seen
- Anderson & Beadle



- processes that ^{single x-over} (recomb) does occur in inversion that
- freq of ring recovery says single xovers occur at normal freq 66%
- ♂ meiosis, checkpoints, apoptosis if all chrom are not paired

- ♀ meiosis, doesn't have checkpoints since only a few eggs produce (what abt hsh egg ~ stem)

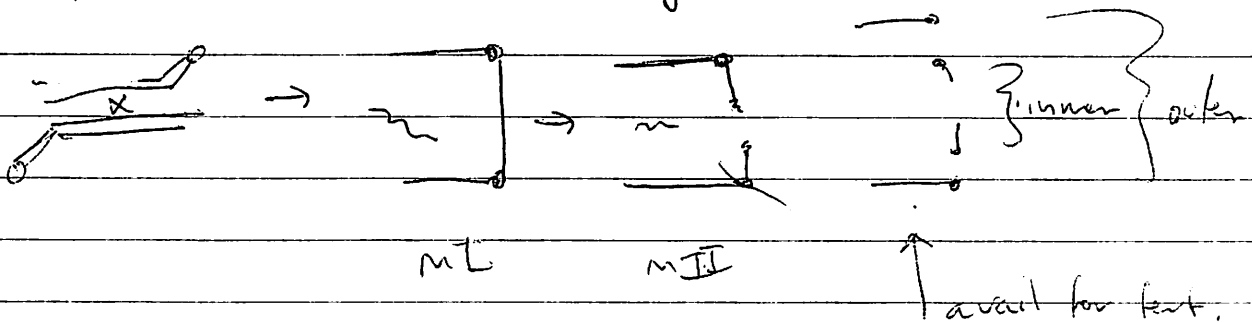
- Sturtevant & Beadle 1936

- Drs Dayle mei I mei II all 4 products in a row, only innermost nucleus is available for fertilization.

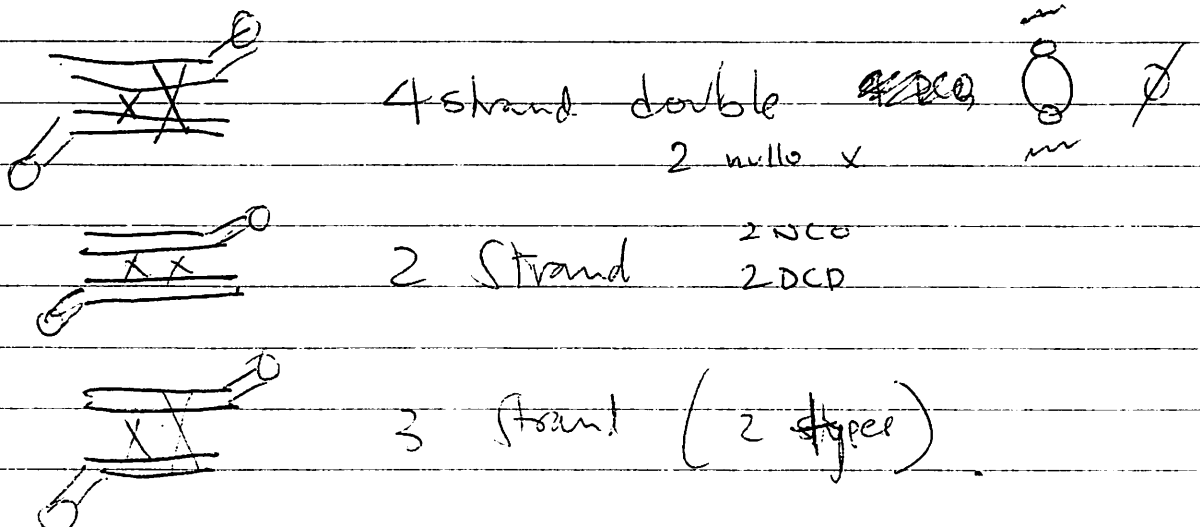
- 500x more force on cent than required to move it

- 500x more force than needed to break phosphodiester bond

⇒ break dicentric bridge



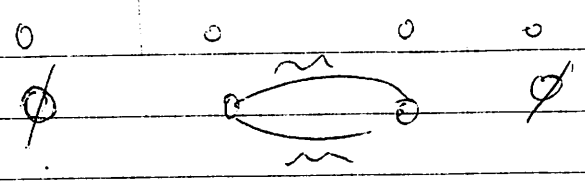
double xover



4 strand
 3
 2
 1
 0

outermost
 fertilizable

4 strand

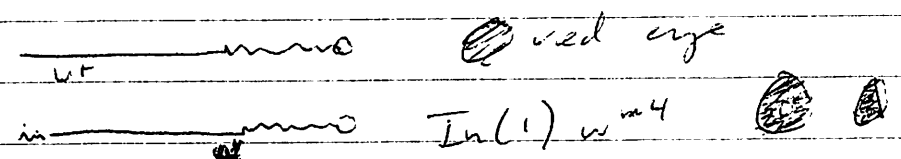


DCO NCO

DCO NCO DCO NCO

- only ad has spermatation was double budge
- predicted ratio of patriclinous ♂ : doubles
non-disjunction
- Isabel stone (Texas) did X's before St & Red
 b didn't know about cell biology of fert.
 showed predicted ratio but didn't explain result.
 (similar to La Franklin story)
- large inversion suppress recovery of x over
 do not suppress ~~recovery~~ (autosomal balancer)
 except around break points
- Muller, "ever-sporting displacements"

Inversions
 first
 led to obs
 of PEV

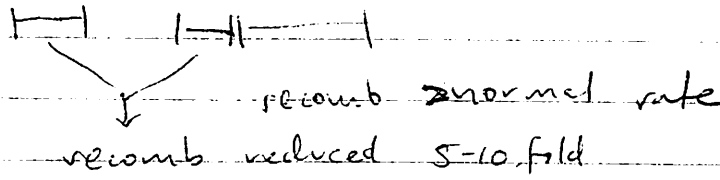
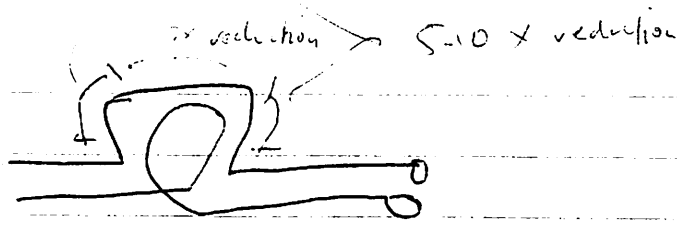


- heterochromatic spreading, reset in germline
- PEV, w^+ is normal, epigenetic effect
- many large inversions variate for w, y, etc.
- inv (sup heterochromatin / PEV)
- inversions are viable \Rightarrow genome order not imp.
except for box etc.

small
 pins

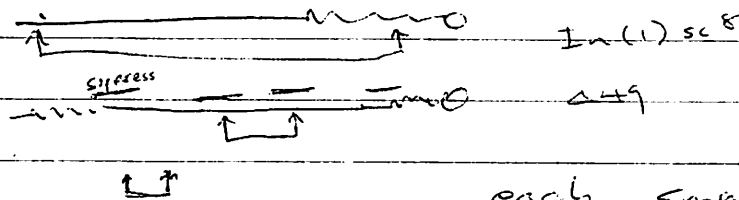
In(1)Δ49 4F-12A in middle of X
 - where most of exchange normally occurs

fast transmits dicentric
"centromere strength"

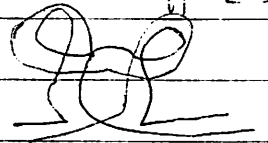


- why do recomb frequencies change, when they don't lead to acentric/dicentric
- heterozygosity for breakpoint suppress exchange directly since bivalent cannot synapse in small inv

FM7 used In(1)Sc8



each small inversion supp 2-3
of units



- FM7 pairs in Mickey mouse fashion
- topology of DNA in region of breakpoint is similar to X-over
- recall interference, possibly inversion breakpoints cause interference \rightarrow suppress exchange
- correlation of species w/ inversions & interference
- inversion explain balancers to effect of balancers

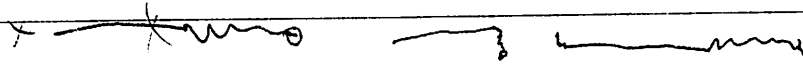
pericentric inversions

not acc/dic

- xover inside peri \rightarrow dup/del but has 4 cen.
- peri in nature have μ that suppress xover
- how much aneuploidy can Dros tolerate?
del < 1 numbered unit
60 regions that are haploinsufficient

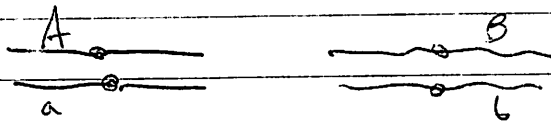
- dup $\leq 2-3$ numbered units
- most pericentric both breaks in electron
- don't use peri for dup \rightarrow use PRF

Translocations

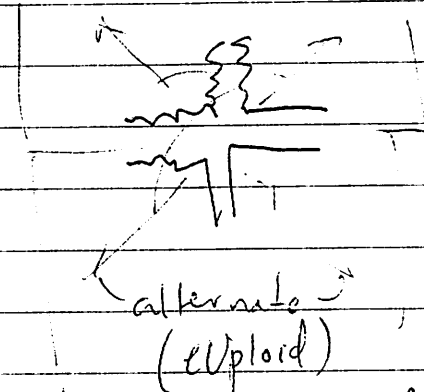


4 \times \rightarrow part of carried by 4th centromere

- strong dominant sup. of exchange (like small pane)
- σ , create ψ -linkage (only in σ) of suppress^A



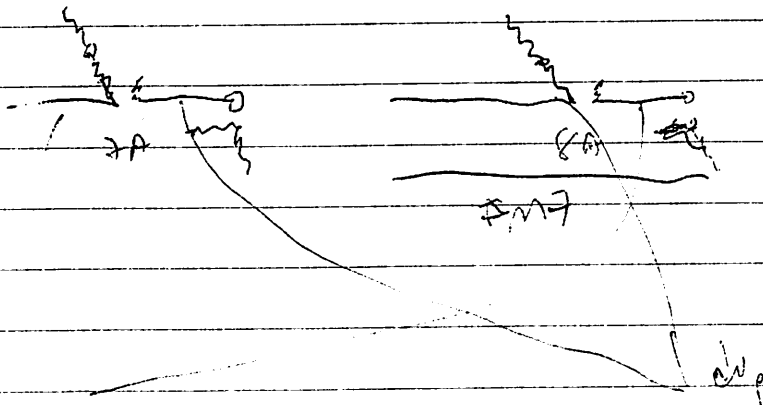
seem to be linked



adjacent (aneuploid)

alternate (euploid)

- 1st aberration pale Attenberg (look in Muller student of Morgan)
- segmental aneuploidy (Lindley & Sandler 1968) transposed & abt which regions can dup/del
- σ meiosis: no synapsis, no recombination, no gene conversion



Cross 2 to each other to see which can perm (mut) \rightarrow dup

del

Telomeres

- early obs that deletions always retained last few distal bands
- ^{McGraw}Parker & Muller indep. disc. no terminal del
- only way to get 'nd of bands was to build a ring
- Parker thought they found a term. del, but had 2L term. bands.
- Muller published 1st using his own
- telomeres discovered in Dros. but don't have typical telomere repeats.
- are telomeres necessary for fixation
- μ 2 flies, can create terminal del. chrom will shorten every generation capture tel from another chrom or de novo

Centromere

- primary constrictions
- Bridges cross - thought cen near yellow
- long runs of satellite
- epigenetic state, histone modification
- only single centromere
- minimal chromosome size? $\frac{1}{20}$ th of 4th
- maximum size in σ but not φ
- 8 min mitosis, 7 min S phase and blast 80-100 minutes (cycle 13-14)
- 11E constriction, Hoka muscle genes

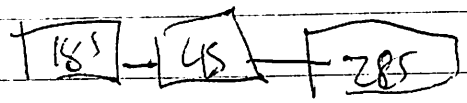


NO is more lightly staining
nuclear organizer

18S rRNA
28S

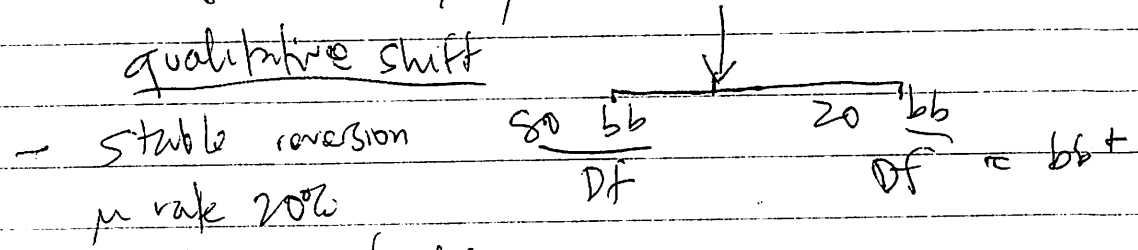
dip
defect
in
Dros

- X & short Y have Nucleolus Organizers
- equal to bobbed loci (bridges) 1938
- 220 rDNA repeats (160 is min)
- bobbed are partial deletions
- deduced that it was a duplicate
- $x^{bobbed} / y^{bobbed} \rightarrow w.t.$
- < 120 copies strong bobbed phenotype
- why bristles, bristle requires a lot of protein



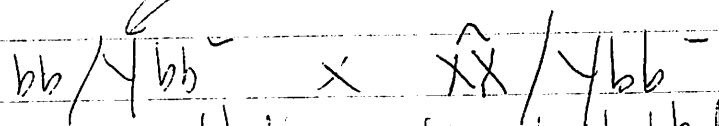
- Ritossa who found that rDNA = bobbed
- Tartoff $bb / y^{bb-} \times DF^{(bb)} / Bal$

qualitative shift



- stable reversion
- μ rate 20%
- called magnification
- doesn't happen w/ bb / y^{bb+}

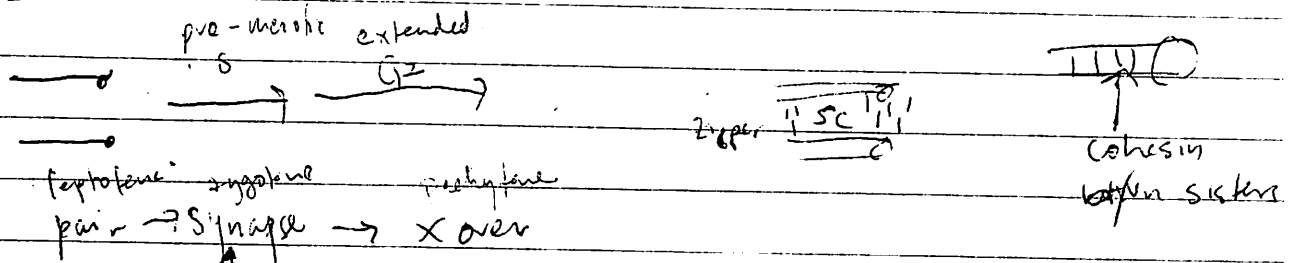
- Ritoss $bb / y^{bb-} \times \hat{X}X / y^{bb-}$



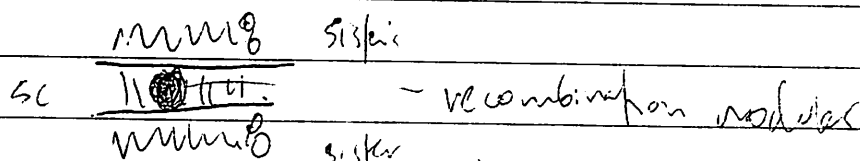
- observes quantitative shift in bobbed phen (less strong)
- in unpaired regions, see unequal pairing as if trying to pair w/ self
- unequal x-overs
- Ritossa proposed alternative model involving epigenetics

- resolution by Hawley, independently repeat exp.

Meiosis



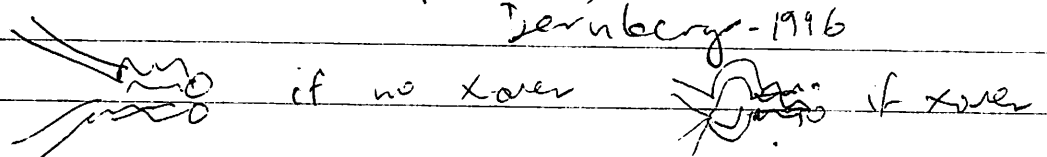
- (SC) - no SC in Dros σ^7
SC hold chrom in place until X over
- Down Syndrome / Klinefelters (XXY) nondisjunction
Lc of failure to X over
- backup system to handle no X over on 4th
- programmed double strand breaks, WDS, ~~DSB~~
- DSB occur before synapsis (synapsis will occur w/o DSB)
but resolution occurs after synapsis



- difficult to find unpaired chromosomes
- Dros may have evolved pairing dependent gene expression mechanisms \rightarrow transvection
- now cohesive forces \rightarrow repulsive forces
- diplotene & diakinesis

SC breaks down & now cohesion to remain hold

- in Dros σ^7 , het holds ^{all chrom} together
het does not repel, only each other.



- 4th all synapsed since mostly het
- Bruce Niklas explained few of X over

- ϕ nuclear envelope dissolves, a centrosome

-? how do ϕ build bipolar spindle, up until metaphase I centrosomes present, then disappear

- chrom organize spindle from inside out

- then freeze 1.5 days, pass through arrest

then anaphase I & fertilization

- C3G 1922 1st meiotic μ ,

no Xovers in C3G

C3G is transverse filament protein

- claret nondisjunctional, short-term (1st in 1 son)

2 genes 350bp apart, deletions of this

region are very common

- Equational, Jack Schultz, lost

- Sandler & Lindsley, wt. μ in bio process

meiotic mutants, mostly a failure mei-332

- trigger to anaphase \rightarrow dissolve cohesion

- Charlotte Averbach, English, EMS

- Ed Lewis showed ~~EMS~~ EMS as μ in Dros DSB

- Baker & Carpenter, 200 X chrom, 9 mei mutants, EMS

- Hawley P-screen, 10 yrs ago

FRT 3R, 2L

Recomb defective meiotic mutants

$E_0 = 5\%$

$E_1 = 60\%$

$E_2 = 33\%$

$E_3 = 2\%$

Wernstein Harvard 1930s

4th chrom $E_0 = 100\%$

X balancers $E_0 = 100\%$

X nondis 1/1000 \Rightarrow other mechanism

- will recomb defective

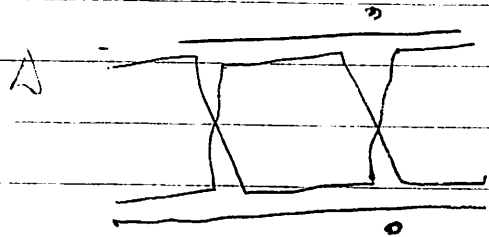
C3G, mei-68 (sp11), mei-P22 , mei-F9

- trichromosomal effect but balances on 2 ↑ x_{over} or x
- recomb defective

- 2 classes:

- $mer-9, mus312$ + 1) lower freq, but otherwise normal
- + 2) preconditioned mutants.

affect final resolution steps in recombination.



to one or both junction

Szostak model

Terry Orr - Weaver did work

- chromatids are intertwined so how do we separate chromosomes.

- 2 single strand nicks on both = GC rich
- 3 nicks at •

⇒ recombination

- Δ 2) defined as if residual exchange is normal
- no interf, points respond to modifiers of exch
- (largest class 3/4)

no idea how the work

- chap

segregation defective ψ , next in seminar

Hanley Seminar:

- central dogma of meiosis

pair → exchange → disjunction

- largest killer of humans is failures in meiosis

No embryo has Turners (1X) 40% spent abort

- % nondisjunction \ll freq of non-exchange

- most nondisj. are non x_{over}

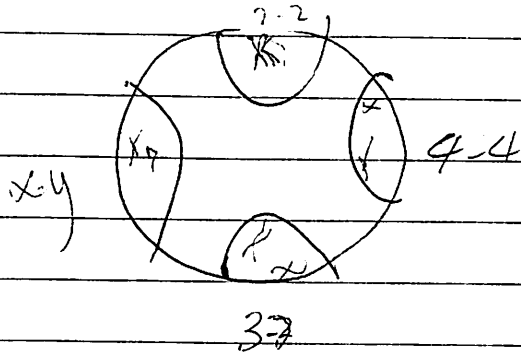
- no exchange on 4 but segregates

- pair $\begin{cases} \text{exchange} \\ \text{non-exchange} \\ \text{achiasmatic} \end{cases} \rightarrow \text{disjunction}$ "revised view"

"distribution system"

map have 1 less and add chromosome

- ? what happens w/ DSB & SC in ♂



go "go to your room"
chrom not locked
together

- mutants that specifically affect certain parts
not observed for ♀ meiosis
- most μ only affect ♂ or ♀, but not both
(except dub)
- Rulb: ♂ & ♀ meiosis differs
- ? does ^{above} affect TE life history?

- Diptera, Hymenoptera, Cole, Siphonaptera (fleas)
- Diptera, 1 of 5 major homometabolous
 - insects/arths, grow by molting, discontinuous
 - cockroach - hanni
 - metamorphosis in homometabolous morph &
ecol. diff b/w larva & adult

1) egg/embryo, - no growth only gas exchange vitelline mem. chorion

2) larva, 3-4 instars, growth

feeding ceases in last instar (minor peak) (ecdysone)

ecdysone, major peaks → molting

ecdysone receptor + USP

Wigglesworth
founded insect physiology

- 3) pupariation, crawl out of food, contract "tan" skin
form pro-pupa, last molt into pupa inside
puparium, juvenile hormone + ecdysone → molting
↓
molac. bot of it is poorly understood
? hormone receptor?

2/11/17

prothymic gland

- endocrine & JH synth in ring gland, around esoph

- insects ring gland environment steroid

- P450s involved in steroid biosynth.

mt have no whole (es. *Shannonia* etc.)

- JH in corpora allata

neuropeptides - brain hormones control synth of ecdysone

- light & metab → trigger

- eclosion hormone

- hirsuticon, tanning of adult cuticle

- Bridges 1920s introduced yeast as culture medium

- filter feeders, eat yeast & bacteria as larvae

note * grow in micro-org rich environments, fermenting substrates

- sugar → alcohol ↑ 12-13% alcohol in a yeast fermentation

- ancestral Adh → very efficient alcohol dehydrogenase

short-term protein

- made by fat body, highly abundant

- not orthologous to mam. Adh

- some Dros. have secondarily lost Adh, specialized to non fermenting substrates

- flower or fungal feeders, leaf miners, aquatic carnivorous (Africa), gills of land crabs

- diet by using ferment. substrate

- repleta (*Chydor*, *nojavensis*) rotting cacti

- specific relationship fly-cacti, rot pockets

- 3800 *Drosophilid* species (could be 2x)

- restricted geo. distr., exceptions like mel

- 800-1000 only in Hawaii endemic

pesticides in sugar plantations) risks predating wasps

- endemic to specific islands of Hawaii

- morphological innovations

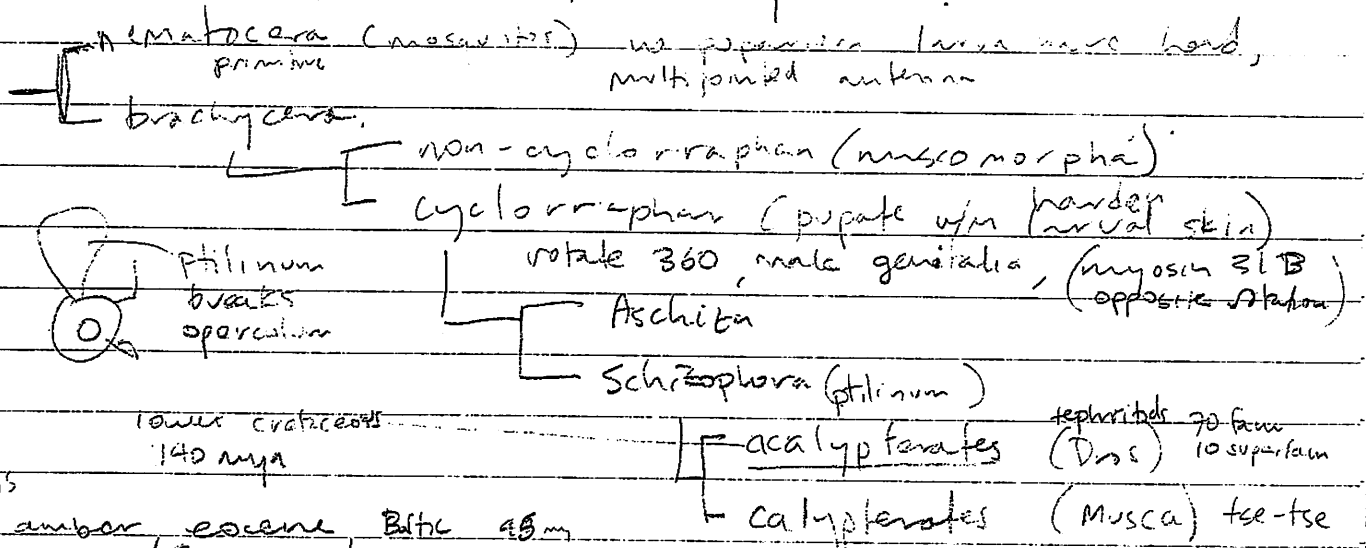
- little work in evo-devo exploiting these model.
- lek behavior, (first described in grouse) ^{Rambles}
→ strong sex sel.

* most species display & mate on breeding site

- cosmopolitan flies, (*P. mel*), synanthropic (free living)
- 6-7 other cosmo. or pan-tropical
- ecologically versatile, opportunistic feeders
- newly cosmopolitan
- buzzard, pan-tropical, opuntia (prickly pear)

* Diptera

- sister to mecoptera (stone-flies) or siphonoptera (fleas)
- first fossils, triassic, 245 mya



Grimaldi & Evolution of Insects

= most acaly. exploit flowering plants, co-evolve w/ angiosperm

- radiation of angiosperm, cretaceous
- 75-80 mya origin of family (late cretaceous)
- Hennig dipterist, invented cladistics
- no acalyptrates in NZ, endemic so evolved same split of NZ from Gondwanaland 80 mya (unless loss on NZ)

- fleshy fruits arise in panicle after K-T

AS
habitat → fossils of morphology of birds

- yeast have 2 classes of Adh

1) fermentation

2) metabolism

* - Steve Farmer? which is ancestral? ②

fermentation is a derived characteristic
needs sugary substrate

- last date 75 mya

- >40, <100 age of family

- Drosophilidae (sister unclear) one of Ephydriidae


superfamily

└ Steganines, non-fermenting, non-fermenting, Amniota

└ Drosophilinae, fermenting, detoxify GSH

- Drosophila, Cacoecus, European, artificial nest
parasitic

Drosophilinae

Zaprionus  white stripes, Africa, pan-trop

Drosophila

(Chymomyza - 1st desc b, short, boxing behavior)

Drosophila filament, discontinuous band

Sophophora 2 chorion, abdominal pigmented band

obscura - holarctic

saltans - neotropical

williston - origin of pigment

melanogaster - oriental, Africa, 180 species, 12 subgroups

montium, largest, 90, oriental + Afr.

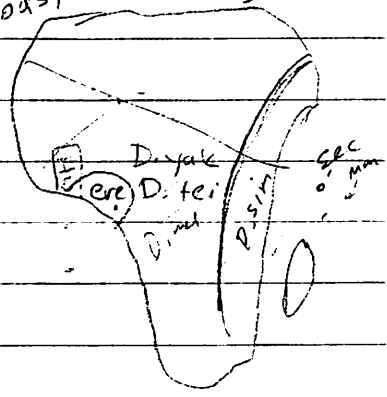
mel, African

anan, oriental

tak, oriental

suzukii, oriental

- ^{german} Mesager 1834 *metanogaster*, 1906 Bussy East Castle 1908 Magn
- Quackebush sent sp. "D mel" → *D. similans* 1919
 - distinguished by shape of genital arch
 - not as strongly tolerant to EtOH
 - hybrids in wild Lipari Islands
- Burla 1954 *D. yakube*, Ivory Coast (savanna)
- Tsacas, David, Lachaise *terssieri* (forest)
- *erecta*, pandanus fruit
- *irena*, top of mountain in Cameroon
- *sechellia* → *movinda* shrub fruit
- *mauritia* } island endemics
- *Saotomea*, yellow, look like Y



- *movinda*: not native to sechelles, Indonesian 6-7 AD
- *mariners*, smells like roquefort, octanoic/betanoic acids, (ethal to *D. mel*)
- highest species diversity in W. Africa
- rift valley, continues to Red Sea, very dry
 - E of rift S → M W of rift N → S
- rift is 2 mya
- *mel* left W. Af > 2 times
 - through non-desertified Saharan region
 - 5 mya
 - recent large trade
- 7 strains differentiated
- 20 mya Af → Eurasia, previous lethys differentiation in birds/mammals
- Dros can migrate long distance
- Hawaii
 - midway → Hawaiian islands
 - ← NW
 - *Scaptomyza*
- Guppy - origin of fish name studied long dist dispersal

Pollard paper
1/1/1933

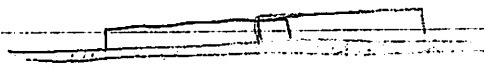
- 1933, Painter discovered polytenes

Wasserman

- 1935, bading pattern was colinear w/ genetic map

- Short collecting pseudo, poly for inversions

- loops form bc of somatic pairing

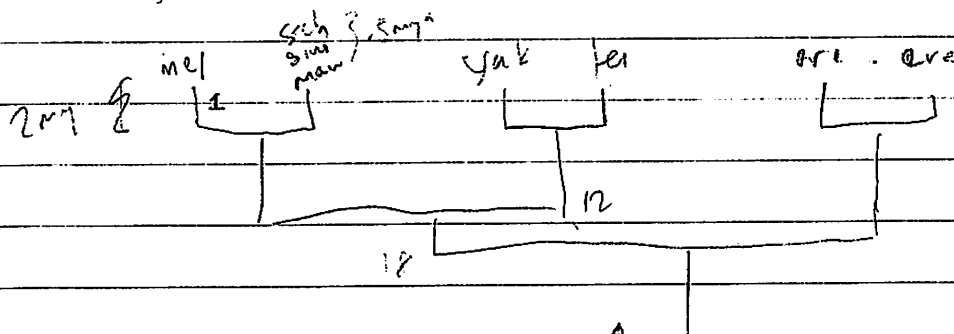


Short had intuition that overlapping inversions could be used to make syngenes

- repleta Wasserman

hawaiian picture wing Carson

melanogaster Ash & Lammie



- use tok to polarize this tree

- Short w/ sim collected visibles, mel-sim hyb

- interspecific complementation test

- genetic maps are same, except 3R

Plunket & Short 1926 all mapping, no poly

- 1st evidence of inv. diff btwn species

Patau - aneuploidy in humans trisomy 13

Horton

mel ♀ x sim ♂

mel ♂ x sim ♀

behaviour

easy cross

hard cross

♀ sterile A

♂ sterile A

♂ inv 13

♀ inv 13

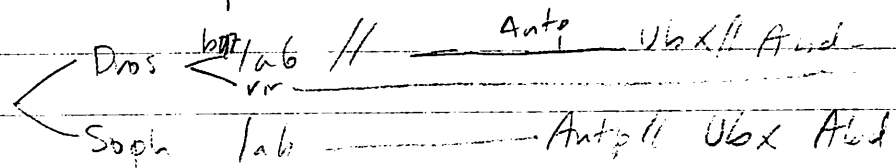
⇒ need sim x, can show long altered x

- a genetic sterile

- why sterile ♂ in hybrids? mechanism
- temperature sensitive
- Matamoras 1970s, Lhr, dominant
 - rescues ♂ lethality (but still sterile)
 - final genetic proof that lethality is under genetic control.
- Hutter, tried to find Lhr in D. mel
 - ukrainian strain
 - Hmr, hybrid ♂ rescue
- Dp Hmr lethal in hybrids
- HMR is a Myb, 20% divergence

Barbara Negró

- Ranz 2001 23-599 Kb ($\bar{x} = 188$)
6.3 ± 1 breakpoints/Mb
- breaks at random
- what about gene complexes?
- HDX, in mel broken into ANT & Ubx
- Anopheles complex intact



- whole protein genes are still near pb in bz
- pb break at 3' end, no 3' enhancers
- pb change in expr pattern but not related to split
- each Hox is
- A, cran TEs in complex but no cran Hox in complex
- estimate rate of expected rate from whole chrom
- compare to observed
- all genes near a break or transposon
- no evidence of constraint in loop

~ across metazoan: splits in rapidly developing species

Scott Hawley

learn
teach
write

X^+/Y

\times

X^w/X^w

\downarrow

$1/2000$

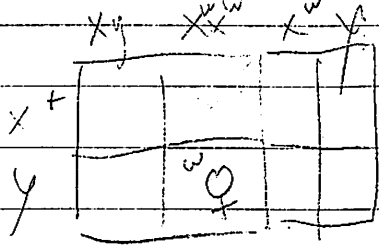
white ♀

$X^w X^w$

\times

X^+/Y

\downarrow



- 50-100 fold \uparrow in rate of disjunction "secondary nondisjunction" (doesn't mean at meiosis II)

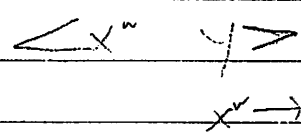
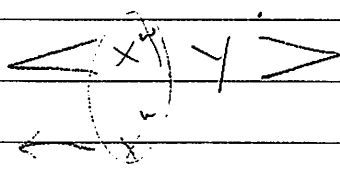
- Bridges couldn't explain, but proposed a competitive model
 $XXY \rightarrow XY$ or XX (presence of $2X^w$)

- ~450 cases 2^o nondisj. involved non-exchange chrom. 95+% used marked chromosomes

- can't pair, can't X-over, or vice versa

- Eo,

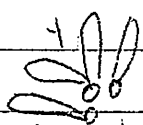
early pairing



predicts 50% nondisjunction if no exchange

- alternative model - trivalent - Cooper

late pairing



miranda-like

only after no X-over

- Grell, = availability

no association until 2EB

- site shape, 1mb. will orient 2 acrocentrics

- XXY associated in germlinum (early prophase)

- all models are wrong

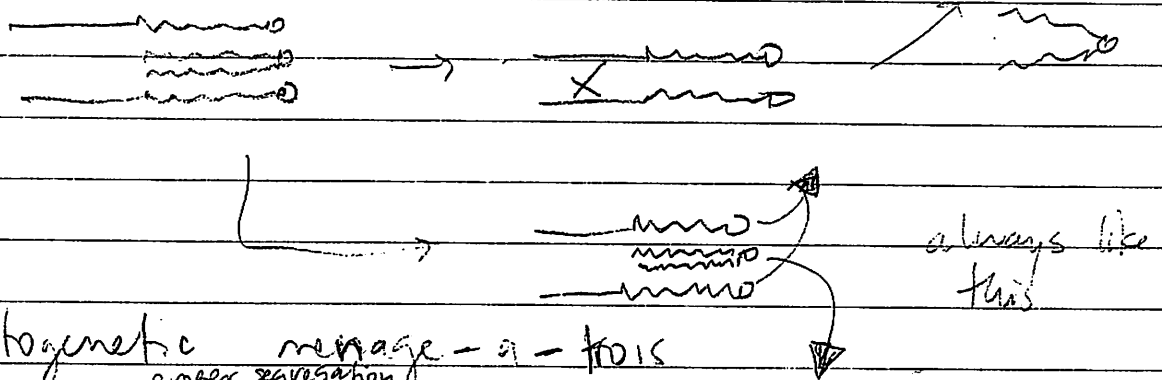
begin paired

if X over, X-Y association breaks down

if no X-over, maintain association (h)

late

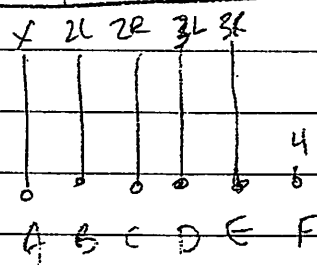
- 2nd round is form of trivalents
- x-over, lock hom. cent. together
- homeolog - ^{divergent} homolog from another species Dean



- cytogenetic message - a - tris
- explains proper segregation Robertsonian Fusion

M. Ashburner

Chromosomal Evolution

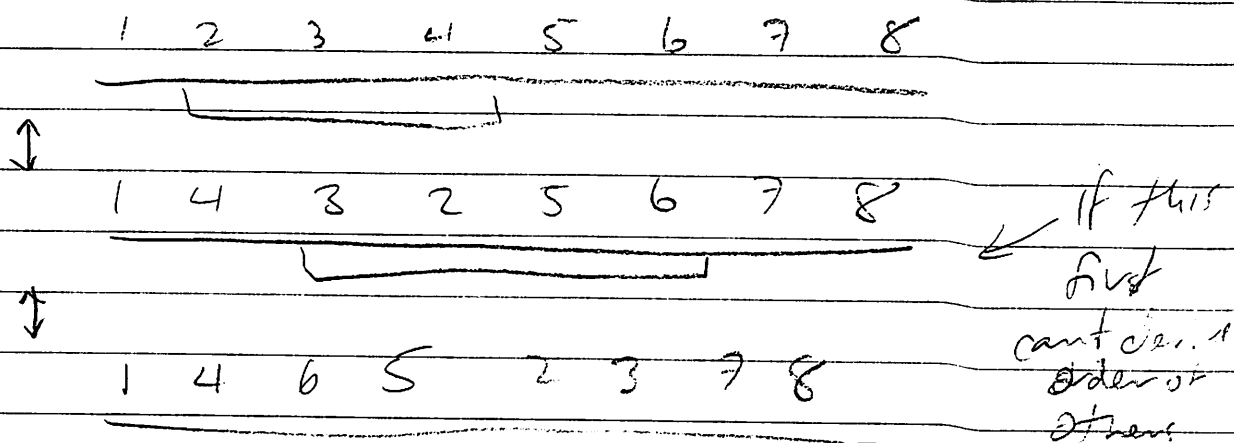


- no interference in manihana (Scott)

- D. pse 3x map of D. mel

- 1932, Ithaca, Muller proposed that fusion/fission
- Robertson, first described in grasshoppers (attached X's)
- tobacco mouse
- Novitski (thesis) & Sturtevant, conservation of genetic mutations (1942^{ish})
- paracentric inversions
- Sturt found natural x-over suppressors "C chromosomes"
- proposed on genetic grounds "inversion"
- Sturt & Beadle 1936 mechanism
- by then, polyenes available
- para \Rightarrow peri (rare) robusta
- mel 5 common cosmopolitan, autosomal, para
- freq high \sim 20%, all simple

- nearly all are simple in mel
- in psl, complex overlapping inversions
- overlapping inversions \rightarrow order



- C element, chrom 3, all on one arm
- not atypical for one element to be more affected than others (but see obscura)
- closely related species differ by fixed inversions
 - " level of poly inversion
- poly sampling in psl by Dob.
- phylogenetics in obs & willi by Dob
- Wasserman & Carson culmination
- assumptions: single event in a lineage
 - never been rigorously tested
- no effect of X over is consequence of recomb^{out} of
- pericentric autosynaptic derivatives, derived from single X over in peri
- Charles Metz, mitotic chrom evolution
- Michael White's review was peak of this trend
 - VK \rightarrow Texas \rightarrow Australia (communist, McCarthy)
- 106/183 species \sim 57% seg poly
- 22K - 56K inversions fixed in genus
- dogma, ectopic exchange
 - but, 2j paralog inversion

- diff - mel/sim
- invert dup (detectable)

- evidence from the lab that TEs
- mel/sim 3R long, 0 short
- junction regions have inverted duplications
- comes in 2R P
- 24 transpositions, 21 replicative / retrotransposition
- (jingwei) retrogenes are rare
- 55 breaks / 29 inversions
breakpoint shaming
- 0.015 ~ 0.02 breaks / Mb / My
- w/ inverted dups 20/29 del
- no dup - close breaks
 - break at 1 end
 - lost secondary events
- TE effects not strong
- 28:1 on yakuba lineage

Databases & Resources - MA

Dutch Book 1925

- Morgan, Sturt, Bridges
- 3 yrs before X-ray induction
- all spontaneous
- visibles, some lethals
- N was first lethal to be studied in development
- 1928 Columbia → Caltech
- 1932 DJS, Bridges

Bridges & Beekun 1942

- published after Bridges listed by mutant allele

Lindsley & Grell 1948

- O. bridge
- "Red Book"

Lindsley & Fjinn 1992

- last data 1989

listed by gene

2. dist of Syngenes for gene!
nature "perfect" ...

- 1997 Fly meeting idea - launch Flybase

Harold Merriman ^{UCLA} Ash

- 1992 first grant Merriman Galbraith, Karl Ax

- 1996-1998 formal collab w/ BDGP

- original plan was L12 in a database
- electronic files

- Muller late 1930s → 1942 bibliography

- Hasegawa ^{Student of Muller} took over biblio, at Indiana

~ 100,000 journal papers

- maintain by "curation" 4-6 curators

- triage, 15 top journals

100 others scanned

- 60,000 volumes, 20,000 pdfs

- rare Russian papers translated under CIA/Israel program

- Oct 1993 first web browser

- Karffman web presence & images

M. Hanes - stock lists

- names of genes, priority rule (follow Zohar 1991)

- dpp, Notch, warty

- Df, anista

- releva 13,000 10% method to index genes

- IDs in assembly meaning

- Protein coding genes

- μ alleles & sequence

N. d. pl

- sequence

CG

- μ alleles

curved

- status uncertain

lost stocks

- sybase back end

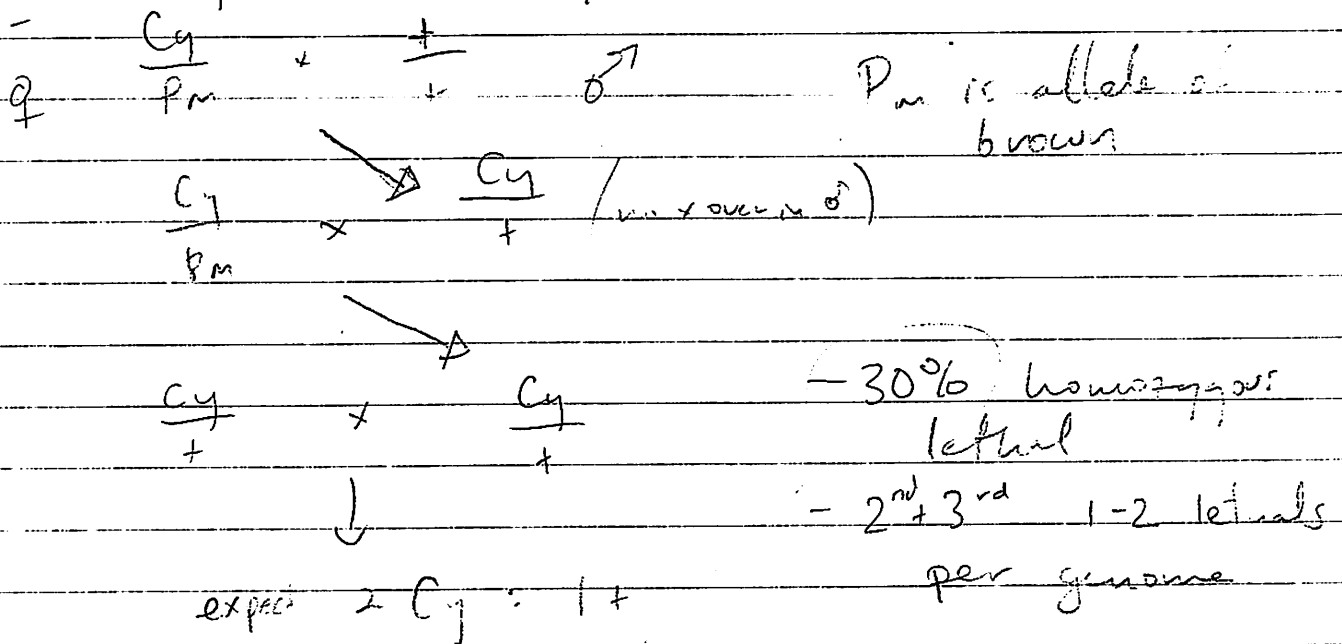
- chado, postgresql, postgenome

- structured controlled vocabularies

- G.M.O.D

P. Charlesworth

- cardinal is a natural polymorphism
- = inversion poly.
- relatively few poly in morph traits
- typically quantitative variation
- C.V. of most metric traits 5-10%
- heritability is 20%-80%
- "concealed variability" recessives shown by inbreeding
 - reduced performance (viability)
 - increased variability
- cloning of chrom by inversion



- non-lethal reduction in viability is ~20%
- freq of steriles is lower, non-sterile are reduced & fert.
- lethals are typically non-allelic (2% allele)

\Rightarrow rare in any given gene

? what % of fly genes μ to lethal?

25-30% in Adh region (vital)

\Rightarrow affects estimated # of genes using lethals

- Sved, competition, showed isogenic is ~15% fitness
- end of SD's classical - Muller, rate del variants balanced - Dob, study of inversions

all genome
 43 incorrect
 4 NE of
 non-syn
 L \rightarrow 2R \rightarrow

- Wright & Dob "wrote" a paper on neutral theory but not ever published
- wrong: Dob saw fluctuations in freq caps every year
- Dob's insistence in het adv caused some damage in relations w/ Wright

- molecular genetics

- unbiased

- Lewontin (Dob student) & Hickey (Texas school)

- prot. slightly neg, starch (Harris) neg (L & H)

- molec. phenotypes, Mendelian

= Dob was right abt 0% of poly loci

\uparrow 40% of loci w/ $>10\%$ alleles

13% of loci are het. (many \downarrow \rightarrow \uparrow)

heterozygosity

gene diversity

- refuted the classical view of low poly

- biased, only $1/3$ of aa can be detected

no insoluble ^{mem} or rare proteins ^{TF}

no silent changes

RFLP

- mostly snps, 25% indels, some large TEs

- Kreitman, high variation 10X alowtyme

many silent/synonymous

SNP density is metric

expected 39 AA poly, see 1

\rightarrow most aa changes are 'del'

silent/noncoding may be governed by drift

- Tj & D same when $n=3$, necessary mathematically

- E coli genome 0.05

D. mel (Af) 0.02

H. sep 0.001

- genome studies are as unbiased as possible

- Forms of selection: purifying, directional, balancing

- divergence, multiple hits JC is HAMM

- saturation, $D \sim .75$ random sequences
- $K = 2\mu T$, neutral evolution
- Kimura 1968, Wright derived but no data
- yes: μ from obs K & poor T
- $K < 2\mu T$, purifying selection
- $N_e, 10^6$, don't need strong selection to prevent aa from fixing
 - \Rightarrow explains why biochemist can μ many aa w/ no effect but seqs stay similar over time
- similar conclusion for directional selection
- $\bar{U} = 0.80$, deleterious
- 4×10^{-9} point sub μ rate, same as electrophoretic
 - 2-3x times molecular clock
- no real evidence for $\bar{D} \neq 2\mu$ rates
- most sel is purifying
- genetic death per generation e^{-u}
- Malik & Henikoff (2001) $K_a/K_s < 1$ but MK fit \bar{U}
- selective sweeps, distance $\sim r/s$
- Kim & Stephan simulated sweeps, recent ($.25N_e$)
 - $10^6 \times .25 \times 10 \text{ gen/yr} = 5,000 \text{ yrs ago}$
 - frequency distributions, assumes derived state known
 - bottleneck, $A_f \rightarrow E_n$, $D > 0$ if partial
 $D < 0$ if complete
- diff parts of genome with diff s/μ D , and \uparrow variance in T_{ajD}
- sel sweep \sim bottleneck at one locus
- avoid species w/ bottlenecks, $D. miranda$
 - no \times sweep at E_n !
- Bordeaux mixture, nebulothione
- O netto, bottleneck model based approach.

Selfish DNA

- 1 obvious x-drive, Genshemion
- why are they are there? TEs & Seq dist-ten
- most families seq sim, but w/defective
- adaptive vs. selfish DNA
- some cases of adaptation, but cannot explain prevalence
- most visibles are TE insertions
- n ↑ transp ↓ excision
- few solo LTRs (ectopic recomb in Dros is rare)
- rates of movement, by in situ hyl.
- 21 lines, 27 new insertions, 1 excision
- trans \gg excision of pt 5×10^{-9}
 1.2×10^{-4} 4.0×10^{-6}
- 1 new mutant in DHPLC screen
- biased insitu towards
- n per family, additive quant trait
- $\bar{n}(\mu-v) \Rightarrow$ all sites that can be occupied, should be
but most sites are at low freq
- occupancy distribution < 0.5
RFLP unique
- 23000 gen \sim 2000 yrs (10x increase in abundance)
- Forces controlling element abundance
- neutral model rejected $N_e S \gg 1$, equilibrium (12.5)

a - self-regulation. P-element

b - sel. against insertions, few in genes, but μ in lab
so nothing stopping insertion into genes

c - ectopic exchange

self regulation, works on intergenic regions

d - direct effect on fitness

a - no $U > V$

b - inadequate in site excision, $S \sim U$

- Langley / Charlesworth \uparrow in low x-over
- \rightarrow \overline{B} , w ^{dup} little work
- ectopic exchange - ^{could be} pre meiotic
can occur b/w sisters
- X-chrom c.f. - mn 7-10
distal most 2-5th

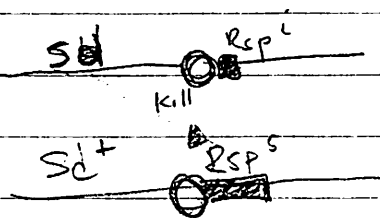
SD - B. Charlesworth

- D-drive allele violates Mendel's law
- no genes required for sperm (all but 4th ^{4th not essential})
- all SD in σ , overproduction of sperm
- some fertility loss, but if D gains it will drive
- Hiraizumi wild flies to c/w/+ -
small fraction produced all wt
- 1-2% of w.f. chrom are SD in wild
2nd chr

- SD homozygotes are sterile \neq

- Hartl, Phil SD is 2 genes $Sd \neq Rsp$
 $Sd^-/+$ and

- Sd , RanGap dup, truncated protein Ganetzky
- Rsp , CIWu, 2R, repeat 120 bp, \uparrow repeat, \uparrow sens




^{chrom}
 SD_n are $Sd Rsp^L$
 $Sd Rsp^S$ is suicide

50% $Sd^+ Rsp^L$
50% $Sd^+ Rsp^S$ in wild

- SD chroms have paric
paracentric inversions

S.H. \rightarrow = add recessive lethal to prevent homozygosity of SD
B.C. not always true, (true to + allele)

- Sex ratio distorters (SR) all in ♂
no mechanisms
- 1st in *D. obscura*
- - Short & Dob in *D. pse* involves an X chrom Xr, such that sperm w/ y die
- Xr, inversions
- XR (3L) ST pse is same arrangement SR ^{primary} CINs
- recens driving X is non recomb
- Dyer, one haplotype, complex of inversions not paired in polyploid chrom
- Gen - balancer ~~FA7~~
- cryptic distortion in sim, X linked
Y & A have suppressors
- distortion causes distortion
- double m to get SD, requires close linkage
- primary disadvantage is sterility
- recens X is "balancer"
- approach to eq. is 
- multiple genes, adapt, suppress X-over
- Fisher → Dob (inversions), SD is good example

Evol of Y

- Muller 1914 proposed proto-X / proto-Y model
- some human X-Y gene pairs in *extremus* is autosomal in *manusial*
- Darwin proposed model for sex evol in plants
 - hermaphrodite → ♀ knockout male for "female"
 - need to be closely linked otherwise neuter
 - linkage constraint
- *C. elegans*, secondarily hermaphroditic males are produced by nondisjunction outgroups are dioecious

- no hormonal sex determination in flies, not entirely true

- if \downarrow X-over, het sex (ϕ in lepidoptera)

- Degeneration of Y

Muller
N: - μ on Y can never become homozygous
- 3x:1y pop size for Y
- lack of recomb.

- Hill-Robertson

- 1st proposed by Muller & Fisher

Greg Gibson

- background mol genet. Antp/Scr

- what abt genetic background?

- Hagness & Waddington work

- use Dros to study complex traits

- disease, animal/plant breeding

- 2 flies more diff than human-primate

- classical: many genes of small effect (R.A. Fisher)

- QTL: few genes of large effect (10-12)

- QTL \rightarrow QTN (goal)

- 5 million SNPs in fly genome

- many questions can be studied at higher level than QTN, reduction not always necessary

- "emergent properties"

- oligogenic discrete \rightarrow continuous

- environment blurs distinction b/w classes

- all under additive contribution

- "discrete" complex trait disease/no disease

- threshold

- mendelian

- penetrance - % of genotypes w/ phenotype
expressivity - severity given penetrant.

- if Mendel's traits were a part of human experience then ancient Greeks would have discovered it
- heritability is not taught, maybe political bad name
- what are most "heritable" traits

skin, hair, height

opera, religion

← not genetic

$$h^2 = \frac{V_G}{V_G + V_E}$$

- heritability says nothing abt diff btwn populations

since it is a w/o populations

- Af. Am IQ Fallacy assumes V_E is same

- h^2 is not a property of individual

can't infer from diff btw pop & know of genet \Rightarrow genet diff.

- h^2 valid for single pop, single "point" in time

- vertical (genetics) vs horizontal (child-child)

- height n=10, need large sample

- 25-30% of genes expressed in egg, maternal effect

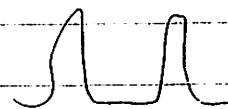
- variance among lines

- 150 female line (1-3 σ^2 sperm) \rightarrow single 15 σ^2 sib-pair mating

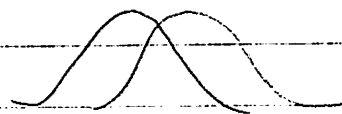
- isogenic by balancers, double x overs, gene on

- ANOVA

| | | |
|----|----|-----|
| 90 | | 100 |
| 89 | vs | 99 |
| 91 | | 101 |



| | | | |
|------------|-----|----|-----|
| same means | 90 | | 100 |
| | 110 | vs | 170 |
| | 70 | | 80 |



- eval diff means by measuring dispersion around mean

- $R/s = h^2$

- estimate # genes, F2 analysis



narrow - more genes

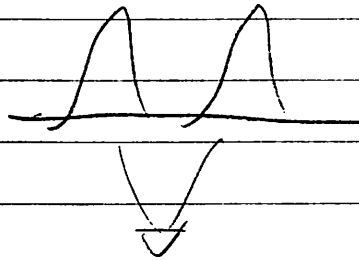
wide - fewer genes

100,000
20,000

5000 generations human

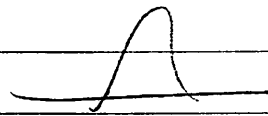
100,000

- wide variation in est # of genes
- modifier changes expression of highly penetrant
- transgressive segregation
grand children more diff than grand parents



P if pure breeding

if same env.



#1 should have same variance

- canalization
- 17-18 whiskers on mouse / dog
tabby & whiskers ~ 6-20, $\bar{n} = 12$
- perturb trait: change mean & variance \rightarrow decanalization
- additive vs multiplicative model
- epistasis, interaction in Q&A terms
diff usage of word from mol. genetics
 \rightarrow btwn g w/in loci
- dominance, under & over
- "molecular existentialism" - epistatic effects
- eg, eyeless, humidity dependent
miniature, temperature
- if N.S. powerful, why not identical?
diff environments
- maybe mutational effects ("quantum" force)
- 20 yrs of M.A. ~ standing variation
100 generations (not same at mol level) \Rightarrow pleiotropy
- $\frac{V_M}{V_E}$ is $1/1000$ V_E $V_G \sim .5$
- muscular dystrophy from new alleles

2020...
complementation on
rescue

270 M size genome

- phenotypic - p.m. embryoogenesis 3 markers not
- may explain upper limit
- P screen for brittle
- introgression, backcross to males
- QTL, lower limit 5% effect on trait
- similar to linkage maps but not perfect association

- mapping multiple genes simultaneously

- QTL limits 5 cM ~ 100+ genes

eg wing shape 10-15, 3 expts, each diff
add all up ~ 250 cM

limit of
100-200

- statistical problem since phen not fully penetrant

- not # markers or sample size

- RIL allows more accurate QTLs since

you can make multi measurements, reduce V_E

- dCAPs - few ϕ per SNP

- all markers can be used to map Mandelians

- LD mapping - a la human genetics

use recomb history

linkage mapping in F1000 not F2

- hap map, 100,000 blocks 200K

- flies 50 bp hap blocks, need to type all
snps to get QTN

- association studies infer causality from correlation

- variation rec. rates in flies (selectable)

- Schlötterer paper diversity/pop struct in Dros

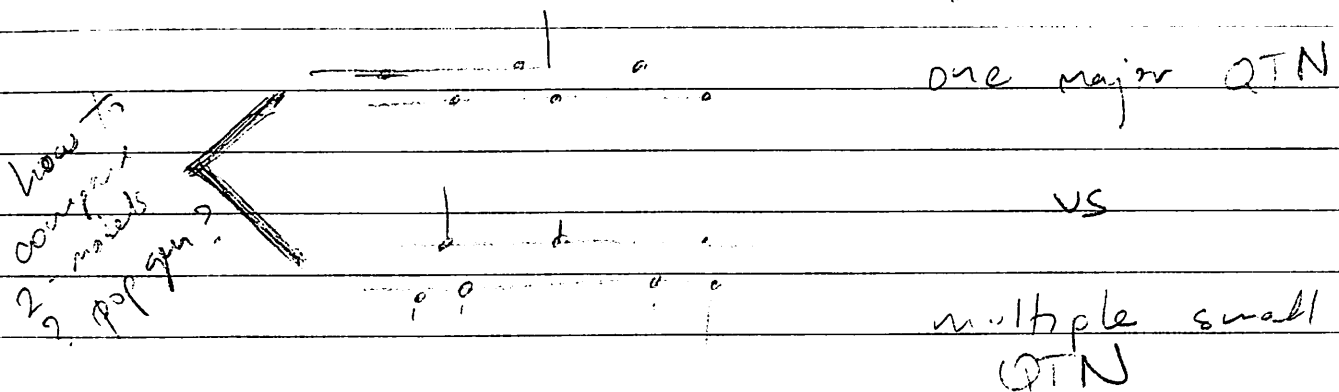
- can't use assoc. study in flies (too much recomb)

→ use candidate gene approach

about can get to QTN

- eg for specifying size/number development

- w. shape is less variable than size "wire-string"
- \therefore likely that "bowing up a shape" so veins are impr
- replication
- multiple testing
- excess of sites associated w/ trait even though few (ϕ) sites are associated
- common alleles, large effect "Bevi's effect"
genetic effect + sampling effect
- human gen. only believes after several studies
- hitchhiking mapping 10^{-5}
association mapping (diff scale)
- Bullhead effect false negative effect

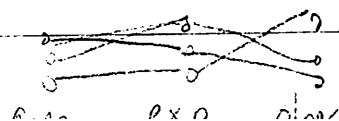
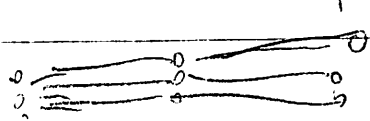


- same problem as QTL at diff. scale
 - no modularity of EGF
 - Second site vs. out crossing to find modifiers
↑ hard to map
 - decline in ins. signaling for vas heart failure
problems
 - reversion, mapping, single or compound
 - only certain class of μ , don't forget all genes
-
- canalization vs. homeostasis (buffering)
(evolved)
 - redundancy is a mech for hom., but evolved for it

- "creed" Waddington 1st systems biologist?
- Andreas Wagner "Robustness & Evolvability in Biological Systems"
- Laszlo Barabasi "Linked"
- "Erekanomics"
- Schmalhausen
 - open up genotype \rightarrow phenotype "black box"
 - is h^2 contribution of h^2 of expression
 - mixed model analysis of expression
 - genotype \times sex \times age? what is biggest contribution
 - age $\leq 10\%$
 - sex $\leq 10\%$
- eQTLs & QTL (quant. trait transcripts)
- "genetical genomics" eQTL
- Illumina: exp & genotype

Schadt

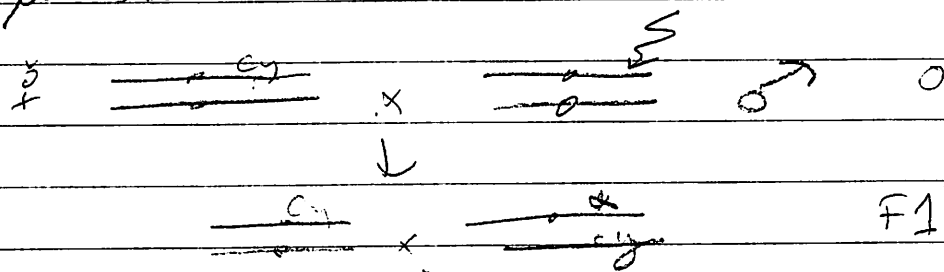
- 15% of 23,000 genes w/ $\sim 25\%$ variance
- stronger effects tend to be in "cis"
- eQTLs clustered
- most genes controlled by ≥ 1 QTL
- structure of eQTLs is unknown
- much non-additivity in eQTLs (Rusself, later 06)
- nicotine sensitivity
- ornithine amino transferase
- transcriptional cliques, groups of genes w/ similar bimodal profiles
- TFC, transcript frequency classes
- plasticity \sim global responses by class of individuals (worker bees etc)
- more divers. in MA than WT \Rightarrow stab sel
- Laabo \sim many genes dev b/c or reprod



w/ cond.

D. St. Johnston

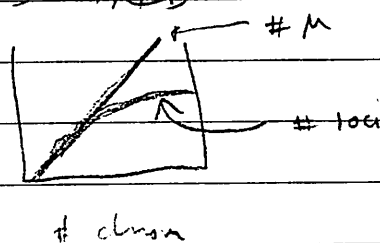
- Lewis screened adult phenotypes
- Heidelberg screens 1980
 - saturation
 - embryonic phenotypes
 - whole genome
- F3 μ screen



- DTS T1 kills non balancer μ flies
- don't need to sort flies at F2
- similar to HS-HID

- saturation

• misnomer, 0.1
class frequent



- 80% sat, 5000 chrom.

N-V, Roux Archive

- not all genes are equally available (assumption of Piss)
- chumpy, 1/20 EMS μ synthetic phenotypes
- rapid devel \Rightarrow if can be provided maternally, it will
- Heid. screen was successful since many essential genes were not found, just spatially restricted μ
- Itzyer's is a horse backbite
- exoskeleton \rightarrow whole \rightarrow phen (key to success)
- what did they not find (new technology)?
- zygotic frame at mid-blast to term, so upstream can't be zygotic \rightarrow look for maternal μ

- maternal effect screen F4, same as zygotic but take viable F3 & look at progeny

- frang only sides of special rubber band

- zygotic - mostly unique phen

maternal - large groups of common phenotype

→ 4 classes of genes & signals

- what did mat not find?

- μ for internal tissues (only routine)

→ Tear & Goodman Heid screen on fixed embryos

- Gas et al 1999 dendritic phenotypes

- wry pathway components (not spatially restricted)
only looked for AP (pattern) defects

- only find 1st essential fn but if supplied maternally it can be missed

- Weischaus attached X & B X-Y translocation

↳ Dev, 1988 essential fn location on X

- deficiency kit

- 7 cellular blastoderm
& gastrulate) zygotic

- Simon et al (1991) cell 67: 710

- enh of sevenless (RTK, R7)

- sensitized background, μ rescue transgene

- tissue specific

- assay is pseudo pupal

- F1 screen, need to recover μ in fly assayed

- RTK, src, exciting, pathway & substrate not known

- biochemists using RIK doesn't have targets besides itself, recruits RAS

- each ommatidia is "replicate" exp

- eye is dispensable

- Banerjee Dm suppressors (opposite) ras GAC
Hafen

100% is for
a few genes

2-3 = few degrees of exp.

- enh/sup screens w/ sev did I find genes
of enh/sup in ETS (lethal in μ)
- use eye-specific sev enh to drive
 ETS , only in eye, post mitotic cells
- X -ray used since EMS hits one of 2
strands \rightarrow mosaics (see in eye but not in germline)
- 1 more generation to get rid of mosaicism
- cytologically visible, breakpoint in Southern
- EMS: order of mag more efficient, but mult
hits, TS 100 alleles, some % small Δ
diff types of allele (allelic series)
- sperm storage \uparrow Δ from EMS
- most enh/sup in eye
 - non essential
 - easy read out
 - multiple indep events in each annotation
 - can use molecules not normally expressed in eye
- adv: \$1, same time 1000 more flies
screen whole genome, not whole brain
only what's even in eye, not from
drivers $sev < GMF < excess$
 \hookrightarrow post mitotic control for cell div phen
- $chradv$: suppress expr. of construct, not product
eg mod of GAL4, don't use GAL-UTS
(make proper construct, secondary screen necessary)
not all genes are dosage sensitive, eg. cat
some μ affect unrelated process (noisy)
many μ give no phen in wildtype background
(redundancy?)
don't know where μ is & only are expressed
in sensitized background

- get around 1st essential fun?

- Dom en / sup screens

- clonal screens

Garver 10 - FLP/FRT Galic Science 1991

exchange in G2

somatic pairing → makes system efficient

- Xu & Rubin, adult

- F1 screen, since only a clone, recover from mutant directly

- wing bubbles, integrin mediated adhesion

- germ line clones, look for embryonic phenotypes (orig 1985 (GFP))

can't screen for specific phen on X

- Brand & Perrimon, GAL4-UAS

GAL80 inhibits GAL4, enh traps are expressed in mult cell types, need to switch off combinatoric GAL4 / GAL80 → more specific ubiquitous GAL80 TS

Two standard? - MARCM, Reuter et al Dev 130.1203

- only mark m clones, single cell

- neuronal pathfinding m

- living flies

- F2 screen, lines guaranteed to recover m (avoid mosaicism)

mapping FRT know arm, but each arm separately

DES-FLP (dominant female sterile) Chow & Perrimon 1996

- stage 2-6 kill egg chambers

- clone ovo^D m

- select for germline clones rather than mark clone

- 100% of progeny from mitotic clones

- cross is from ♂ can't recombine onto DES-FLP

- Perrimon et al (1996) 50% of F1 have maternal effect, 4 new seg pol m

regression 3 ... FET chrom.

- many mod of ng signalling found
- $OU^D + GFP$ select & mark F2
- inducing clones via $HS-FLP$ is random in space but controllable in time,
- clones require mitosis, in adults many tissues not dividing

Bunny
Dickson
lab.

- driver - FLP , specific gene production
- mark chrom w/mini white on non- μ chrom
- kill transposon w/recessive lethals
- slow growth with minute μ in non-recomb cells, get out-competed.
- more reproducible, less unintended phen, less work than consequences
- more specific than MARCM
- get all cells of a type, rather than random
- "eye as an independent organism" to find μ Natl. Cel. Biol.
- pinhead screen Stalker (2003) 5:559

FLP/FRT Screens Summary

- ad: find μ , regardless of other processes
F1 since find recessives
- dis: 5 sep screens, 1/awn
4th of cen proximal to FRT sites
all cell-autonomous effects

Mutagens

- EMS $1/1000$ efficiency 74 per chrom
mosaicism in F1
Makes genes harder to clone
- X-rays $1/12,000$ ragged ends
no mosaicism
rearrangements
- P-elements $1/50,000 - 1/100,000$
tgs genes, IPCK
D-old cells \rightarrow mosaic

Practical Tips

- isogenize chrom you are going to μ
 - need independence of μ
 - need to know SNP haplotype
- do a pilot screen 1st
- balance, map, complementation test are more work than screen
- specific & laborious are better than non-specific easy
Dickson (section & screen mutants) vs. Fipovsky (behavior)

SNP mapping

- visible & SNPs on same mapping chrom
- cleans up chromosome in mapping process
- 2 stage visibles \rightarrow SNPs
- ~ 50 kb

Forward Genetics w/ RNAi screens

- S2 cells, good for basic cellular processes
- via RNAi

- | | |
|---------|---|
| adv. | - guarantee "saturation" post-mitotic |
| | - know the relevant gene |
| | - less persistence (RNA not DNA) <small>problem in clonal</small> |
| disadv. | - don't require cell division |
| | - 20,000 lines hard to cross into complex backgrounds |
| | - not loss of fxn (down expression) |
| | - some cells refractory to RNAi (germline & CNS?) |
| | - still need a μ |
| | - can't make rescue constructs to map full domain |

some working

maternal RNAi not working

- better, or
- targeted screens
- mapping

DST

? is bcd RNA model of?
in vitro shows same

localized RNA

- B-actin polarized cell behavior
- mother/daughter Ascl1
- macho-4 D-V

bed/osk - AP

grk - D/V

- ~~general~~ bed is sufficient to organize Anterior

- bed localization stage & oogen.

- Molecular no x-link shift expts

- many proteins found, general RNA bind prot.

- but unlikely to be imp.

- proper complex of many, low specificity

- even necessary to make bed competent to localize

- Swm, Dgrip, γ -tub 37C, Mops -

indirect role in bed local

anterior microtubule function

- similar needed to stabilize bed localization

- bed gets localized to microtub spindles

- contrast to osk where 20-30 genes are known

- mech of localization changes over time

- show required for efficient ctrl of bed

- multistage process covers components of process
in indirect screen for head defect

- larson (lsn) vps 22 part of complex

ESCRT - sorting pathway multivesicular body

μ in vps cause tumors bc receptors are

kept on since not packaged into multives body

- other members of ESCRT complex not
involved in bed localization

- yeast 3-hybrid RNA acts as bridge

- UV x-linking, competition assays

Vps 22 has point to complex but doesn't bind

1. 454
2. Super 10
3. Accus
4. Inverto

- bed recruits Vps 3b to Ant
- shafken not required to get bed - Vps 3b
- can't X-link DS RNA to protein
- CSCR has 2 func
- Vps 3b has 2D phen. to vps 22
- ask localization requires 1st intron splicing and uses Exon Intron complex
- several mechanisms
 - 1) + end transport
 - 2) flow
 - 3) movement away from center

Kent Golie

1-element (2.9 Kb)

- 31 bp IR, 150 bp terminal required
- 2-3 intron only spliced in germline
- engineered elements are non-autonomous
- > 1 TE allows mobilization of 1 w/o moving others
- helper, eg wings clipped
- prior to 90 min, prior to cell membrane formation
- kill old embryos on slide by blowing up.
- 8 bp BSD - bent & Rio 1997
- imprecise excision - exonuclease after excision
- S' preference hypo-morphs
- fuzzy back, no bias, but no imprecise excision
- revert 17 bp left + 8 bp

in cis

17/8

or

anneal single strand & w/ compl.

⇒ precise excision

= P goes in lots of places - good (diversity of μ) & bad (variability)

- revertants often leave a few bp behind even if phen is reverted, (what are implications?)
- repair off sister \rightarrow restore, often incomplete
- repair off homology \rightarrow gene conversion \rightarrow precise
- repair off non homologous position to introduce eg into a position by gene conversion (need a marker)
- site specific recombination
 - control for μ

- FLP - FRT yeast 2 μ plasmid

Cre - lox

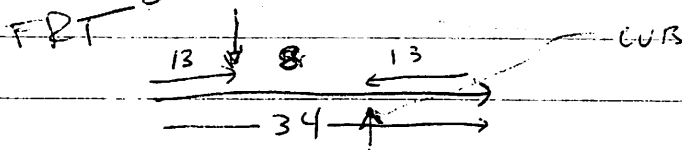
phage ϕ 1 bact

integrase - attP/s

ϕ C31

(constit. expr of Cre causes lethality)
 ϕ lox sites?

- all target sites have orientation, not symmet



- excision \leftrightarrow integration, equilibrium away from integration.
limited recombines for transformation system

- can change FRT to lox sites still work but can't work w/ wt or each other

- close inverted FRTs will create acentric/dicentric

direct repeats \rightarrow duplication

- use 2 diff FRT sites to make transposons

RMCE - recomb mediated cassette exchange

FLP - Handler

Cre-lox - Small

\rightarrow integrates whole plasmid

integrase - (in 1984) / Ting Wu - \rightarrow cassette exchange leaves vector behind

Bakeman et al 2006

- as efficient as ϕ -element transformation

- < 1% have integrated elements, so efficient that loss of white is sufficient, so no marker needed in construct

- orientation of construct not specified. since same attP sites used at both ends

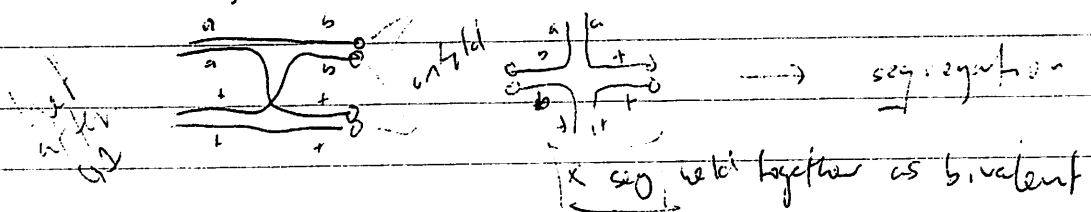
- exchange on one end puts whole plasmid in but get out w/ one more round of recomb.
- BAC ? may not be plug possible because of shearing during injection process
 - need ATT markers
- Mosaics - 2 genotypes
 - chimeras, transplantation
 - genetic
 - 1) lineage analysis
 - 2) time & place of gene action
 - 3) cell autonomy
 - 4) cellular effect of lethals

Baker et al 1978 events that summarize events to form μ clones

- twin spots are evidence for mitotic exchange
- C. Stern 1936 X vs Z segregation
 - conjugation: more imp when X rays used
 - 1000 rads, $1/2$ cell death, embryos die
- PRT recomb can occur b/w homologs
 - \rightarrow eye color 2 pathways

anmochromes (brown) x sc
pteridines (red) x bw

- use induced recombination to investigate models of segregation (X vs Z) in mitotic exchange
- $\Rightarrow Z$ seg essentially never happens



- exceptions when exchange near ho
- sister cohesion converted to homolog cohesion in X over

- exchange in g1 or exchange in g2 w/ x seg
- similar to meiosis
- multiple exchange leads to 3 genotypes (parent/roth)
- MAREM requires FRT to control exchange
- gal80 & μ & FRT on same chrom arm, other components elsewhere in genome

lec 8
LWO
1/199

= Minute - dominant slow growing, ^{nonstop} recessive lethal
Nounsime et al' 2000

very useful if μ has growth disadvantage

- Maternal effects & germline clones

Perrimon
1998

DFS dominant female sterile, ovo^D , early defect
used to study early lethal genes

- 3 genotypes, 2 DFS, 1 viable - has μ homozygous
- >1 ovo^D required for full ϕ sterility
- difficult chrom to build get ovo^D onto FRT, recomb in ϕ

- how to distinguish twins w/out molecular markers?

- site & shape of clones are dependent of time in development and cell type

- very little cell mixing so most clones are from different events

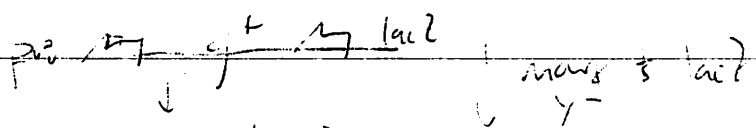
- 60h wing clones large, 96h small Baker & Basler

- ^{clonal} use of "incident" markers to mark clones
- Q. influence process? control by reversing markers

- cloned gene analysis to produce conditional dominant lethal, etc.

- FRTs inserted in intron & 3' inverted, reversible
- excise or invert, part or all of gene & ^{marker} cassette

- FLP - out



1/5 powder
not covered

- allows island of activated marker in background of no marker activity.
- later in dev \uparrow # clones \downarrow size of clones in eye
- small w⁻ clones can look dark w/ direct light

Turning seg into μ

- random vs. targeted (RNAi & homologous recomb)

- yeast 1978

- Hinnen, Hicks, Fink

- transformation leads to homologous recomb.

- mouse 1987

- Capecchi

- ES cells

- positive & negative selection, random insert mostly

- Dros - Giken Ruvy 2000

bypass cultured cell requirement

rare cutting endonucleases

- ~~targeting~~ precedent, \uparrow gap repair by gene conversion
DSB is recombinogenic

- need 3 components ISCE-1, flipped out of cycle
& 2 element μ construct to produce \uparrow

- targeting is predicted to make tandem dup

- not always observed

- 2 copies integrate

- repair by double

- deletions

- need to do Southern to verify structure of integrated region (PCR can be fooled)

- \uparrow \rightarrow \uparrow efficiency, 1/500 \uparrow gametes

\uparrow random insertion $>$ targeted as ratio \downarrow rate

- no real explanation for differences, practically \uparrow

- ? extent of homology $>$ 5 Kb is OK

- construct μ
- express FLP
- μ -F-P

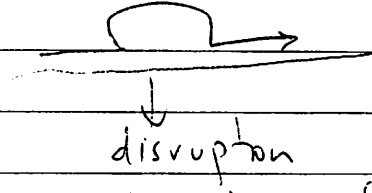
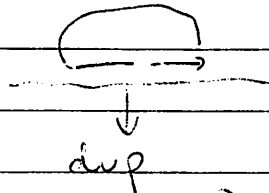
? both homologs?

- other questions?

is this generally useful?

can we produce μ ?

- ends in vs ends out



- no HSP70, also complicated events, partial deletions

2 why

- different donor μ 's have diff efficiency on targeting (pick >1 transposon)

- what abt effects on neighbors - yes, need to use W+ transgene to complement Δ

ends in

- add μ to donor \rightarrow distance dependent

> 800 bp both recipient copies

- ends out is quickest way to a μ (dirty)

- ends-in gives specific μ (pt sub etc)
2 step allelic substitution (target reduction)

- I-CREI used to generate second DSB

" " has endogenous recog sites in rDNA

cross onto fresh rDNA to avoid changes in rDNA

- use water bath for HTS-FLP step

- rapid denaturation 2 rounds of HTS-FLP gets rid of flres where donor didn't move

Kent Golik - ends of DNA

- telomere: prevent end-to-end fusions

" rearrangements

" check-point response

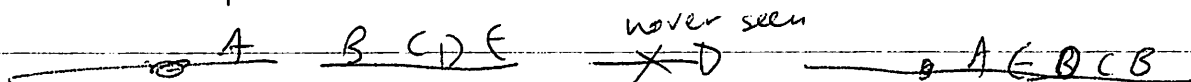
solve end-replication

overlaid
informs?

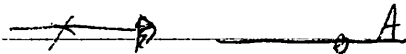
Fun

p - ^{eventual} spinning

Muller & telomeres



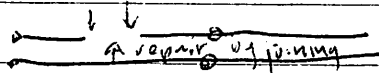
no terminal inversions



" " deficiencies

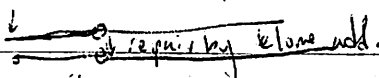
- telomere is a functional definition
- telomeres are polymorphic
- since telomere repeats are not essential
⇒ telomere ~~fun~~ is not seq specific (same as centromere)
- telomere loss → soma - cell cycle arrest
germline - telomere addition
- DC chroms are engineered to permit dicentric bridge formation w/ no phenotypic effects
- fates in embryo can be breakage or not but nuclei are fated to go to fork
- still see phenotypes when dicentric bridge occurs in non-essential chrom
- same phenotypes observed in hypomorphic ATM, etc mutants
- see apoptosis in cells that are being instructed to differentiate, when go thro cell cycle again
- apoptosis occurs after cell cycle arrest that can last for days
- telomeres from broken ends are capped w/ hope, but don't have het-A
- resulting broken chrom are short or long short are viable, long are senescent
- why diff from muller?

X-ray



2 broken ends

dicentric bridge



one broken end

- some cells can escape cell cycle arrest & differentiate

R7 receptor is good example of gene expression
not strictly to gene function

Druce Baker

- EMS, mitotic x-over, late maps in early 70s \rightarrow dev. genet.
- XAA δ^1
XXAA φ
XXAAA intersex } Bridges balance theory

- tra, etc were available
- tra an example of Bridges' male factor? no tra $\rightarrow \varphi$
- from classical perspective, define the null phen \rightarrow ut tra
- is tra cell autonomous? what is null phen?
- what is epistatic relation of diff genes? when/where tra?

1. Null? cross to deficiency Muller's definition of a null
? a/a to a/ Δ , equal null, ok for recessive
- only Minutes are het dom., usually dom are g.o.f.
? what abt dom, revert dsx² to null by x-rays
& cross to recessive

2. autonomous? use mitotic clones

- if cell has δ^1 in φ , then autonomous

3. when tra?

- make clones at diff times
- temp. shift w/ TS alleles - usually thermostable in protein tra
- (tra is active ~ puparium formation)
- (# sex comb bristles, indep from morphology)

4. how μ relative to each other?

cp single & double μ for genes w/ diff phen

- Sxl, dies as embryos, how do you bypass early defect \rightarrow clones

- dsx, unlike other genes in pathway, had tra in δ^1 & φ , precedent w/ mating type in yeast

- cline schedl, Baker, Balogh, Nothinger

- result, all genes in sex det hierarchy use alternative splicing

- do all splices act directly or indirectly

- constitutive activation construct

Sxl \downarrow o. Sxl \downarrow make tra const.

- constitutive expression $Up^{-}, down^{++} = down^{+}$

Up works first down only

$Up^{-} down^{++} = Up^{-}$, branch in pathway or order of pathway, no real conclusion

- constitutive mutant test is powerful way to order μ , but not a symmetrical test

- Sxl only in flies

dsx (mab-3 in worms, Dmrt 1 in mouse)

ix conserved, interspecific gal4 UAS rescue

? what about pantherogenic species?

Dosage Compensation

- problem of gene dose on sex chromosomes

- Muller ϕ w/ 2 leaky recessives = σ w/ 1 leaky

- show both X active? = eq w/ 10^{-}

het cell autonomous marker \rightarrow mosaic = inactivated
no mosaic, both expressed in all cells

- what is phenotype of dosage comp μ ? lethal, sex sp.

Lucchesi - Bolate F3 sex specific lethal screen

- screens 3 intellectual goal - ideal

practical goal - lab

incongruent get μ that aren't

\Rightarrow need second screen !!

eg sex determination μ all $\phi \rightarrow \sigma$ looks like dos. comp.

- msl's male specific lethals (all autosomal)

- how do we know these are involved dos. comp.

- could be Y, male sp. physiological, one X region

XX tra σ , msl $^{-}$, ok, mal male physiology

XO σ , msl $^{-}$, die not Y

X^{dup} σ , msl $^{-}$, die not 2 X

polytene, X same diam as 2n
thinner in msl $^{-}$

- Sxl Lucchesi

- hypertranscription of σ X

- MLE (cloned in Baker lab)
- all present at MBLs bind to same regions, complex
- not simply DNA condensation dependent
- compensasome, msl 1 & 2 form core
 - modify chromatin of X
- Sxl represses msl 2
- vax transgene \rightarrow autoregulate, induce compensasome formation & secondary sites
- spreading model: 30-40 initial sites \rightarrow complete compensasome \rightarrow spreading
- direct evidence for spreading on X
 - X \rightarrow A dup, always see X banding pattern with or without entry sites
 - A \rightarrow X no spreading into A
- entry site model replaced by affinity model
- vax transgene may have been nucleating since vax RNA is where compens. assemble.
- neo-X attract msl by changes in cis
- evolution is piecemeal, gene by gene

open questions { - What is seq on X?

what is biochemistry of histones?

how do you build a band?

- lack of pairing \rightarrow not likely
- transgenes are difficult to recapitulate binding

Targets of Sex determination

- direct
- YP male in fat body, hemolymph \rightarrow egg
 - need sex hier. to activate & maintain YP genes
- gene level
- sex hier. need to form access. gland, transcripts w/in acc. gland are regulated w/in the organ
 - genital imaginal disk

control
of Sxl + h
germline not
cored

? pathway controlling
Ejak. Duct ?

tra - should
switch series to
see if sex
regulated

- dsx is instructive in ♂ / ♀ genital disc
primordia

- clones of ♀ in ♂ proliferate, thus not
cell autonomous

- sex of A-P organizer determines fate of genital disc

- sex & segmentation genes involved in differential

- targets of dsx: cardinals, enh traps, microway

- bab, post pigmentation: $dsx^F \rightarrow bab$
Abd-A \rightarrow bab

- branchless (FGF) & breathless (FGFR) male ^{disc} only

- expression of FGFR allows migration by recruitment

- sex hierarchy controls FGF expression

$dsx^F \rightarrow$ branchless

- sex is fixing at same level as Hox genes

(- modulation doesn't incur genomic cost)
since same binding sites

- microway, somatic differences

former - genes diff in some + can germline

- all internal genitalia gene expression is

tissue specific, not directly by sex

- 12 microway targets, none direct

- gal4-UAS screen, sex disc in live animals

Sex & Behavior

- Sxl & tra control sex & behavior

- dsx, controls sex but not behavior

- pulled out for by probe to 13bp x 6 repeat
which tra recognizes

- null for P1 transcript no courtship behavior

Greene
1995

behavior in cell autonomous?

- leaky & defective in recognition
- fru is BTB-ZF TF
- Pf tr has 100 aa & unique to all
- 2000 cells in CNS
- fru expressed at highest level during larval → adult remaining
- fru necessary from earliest stage → innervation of ♂ sex organs

Get fru from CNS tag

- insert GAL4 by homolog recomb into fru⁺ Pf ⇒ express in every cell where fru is
- UAS-GFP persists in cells where fru is transient.

- ♂ & ♀ same pattern of fru expression
- is fru sufficient for courtship behavior
- UAS-tra drives tra in fru cells & make ♀ ✓ / ♂ behavior

⇒ Fru is sufficient to make male

- visual, olfactory, auditory, & tactile
- antibodies don't penetrate cuticle
- multiple fru⁺ cells in a row ⇒ circuit
- male - male habituation, fru required
- courtship conditioning, ♂ → ♀ ↓ receptivity → ♂² less likely to court for ~3 hrs
- do fru⁺ cells have other fxn.

drive TS behavior

- only recognize male when sex circuitry is fxn

- use UAS-RNAi to 100 AA fru⁺

- knock in ♂ by galic, put in others (GAL4, LexA) by ♂

- fru tr how to set up a circuit

b. Kyrniacou

- genetics of behavior, 1960's started, selection & strain comparisons, didn't yield many results
- Students of Tinbergen & von Frisch
- early 1970's, Benzer, EMS: single pt m in genes which have behav. phenotype
- Benzer was interested in neurosystem not behavior
- "time, love, memory", J. Weisner ³ major phen. of interest
- Konopka ⁽¹⁹⁷¹⁾, screened for flies w/ clock p
- flies sleep, 12 hrs

attached X, first 60 lines found per

per^L 29 per^S 19 per^O - arrhythmic

- per is not a vital locus, del is viable

- Hall & Kyrniacou picked up per work

- 160-200 wing beats/sec in flight

- hum & pulse

- interpulse intervals ^(IPI) vary in cyclical pattern

Shows neurogenic effect (not myogenic)

- song cycles & IPI, sympathetic character displacement
(\Rightarrow female recognizes (but no independence or phylogeny))

two components are independent & impt.

- recip. mel-sim hybrids ^{arrhythm} maps to X, IPI maps to A

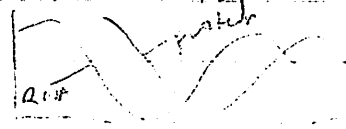
- rhythm of 1 min & 24 hrs, are they same?

- yes, per^L long IPI per^S short IPI

- cloning of per, Hall/Robash & Young - 1984

- per rescue in circadian & song domain

- RNA cycles in fly head, & protein cycle



neg. auto-regulation

- 2 features: need delay for \oplus feedback

need short half-life of protein

- photoreceptors have per, 100-1000 mRNA are per

7 hrs delay
clock p effect
on wingbeat
500

- 3 grps of lateral neurons, 3 grps of dorsal vent in photoreceptors
- LNd - evening > gal4/gal80 expts
LNv - morning
- ZTO, per nuclear localized
12, cytoplasmic
- LN determining behavioral clocks, not peripheral
- per - luciferase, cycles apparent in ^{isolated} leg, etc
- no master clock, only for behavior
- per protein, thr/gly repeats
founding member of PAS domain, dimerization dom
Sim is another, has bHLH
- PAS / LOV, respond to light oxygen & voltage
- T/G repeat → remove → sim like song rhythm
- mel/sim differences map to T/G region of per
interspecific transgenics mel 20, sim 24
- single gene of large effect, adaptive divergence.
- T/G ↑, ↑ song rhythm length
- ring X, tra all male w/ clones of per⁺
- brain⁺ thor⁻ 24 hrs, song messed up > decoupled
thor⁺ brain⁻ song ok, 19 hrs
- how many genes cycle, many, 2 groups
50% concordance, diff expl. design
- per⁺, other genes don't cycle
650 genes, non cyclical, tra +/-
- bouton size varies day/night, cellular change
- thr overgrowth at bouton, no effect on 24 hr clock
song is rhythmic but long
- ? is neuronal branching related to song rhythm?
- is song only major gene effect
- D. psc transgene per shows dom. affect
& maps to 3' end of gene

^M per^{1st} ID using
on population assay

- per. transformants show associative learning
- dom. single locus, affect both ♂ & ♀
- nonA, no-on-transient exp potential
- dissonance maps to nonA, song gene maps to eye physiology defect
- every song μ is visual μ , but not vice versa
- vir nonA trans rescue mel nonA
- optomotor response is fine in transformant
so not a "partial rescue", but real
- major genes common? morphology? bristles
- timeless⁰, per mRNA affected & vice versa
same delay ^{mRNA + protein} cycles as per
- neg reg - tim & per bind via PAS domain, mutually
repress to nucleus
- clock & cycle are bHLH - PAS, E boxes
CACGTC in per/tim promoters
- clock cycles in a loop, clock is gene
that links clock loop & per/tim loop
- native cycling enhanced by cryptochrome
induced per degradation
- > PER genes in mammal 3 + 1 ψ , poorly conserved
- human per gene has same phos site μ
- conclusion { - clocks historically initiated neurogenesis
- impt mech in gene regulation
- impt for evol. speciation
- ^{molecular} mechanism for circadian rhythm is not used
in courtship song

Fly learning

- Bower student
- 1974 Quinn's olfactory learning paradigm
 - classical conditioning, smell & shock, pop assay
 - 1976 Duncie isolated

Parlov's Flies 1988

- 1980 Tim Tully, enhanced Quinn's assay 90% flies condition

- Robo Trainer

- dunce - cAMP phosphodiesterase

amnesiac - neuropeptide

rutabaga - adenylyl cyclase

- learning μ affect acoustic systems, central pathway

- σ are more receptive if hear song, memory index

dCREB - cAMP pathway components implicated in learning & memory

- mushroom body has memory genes

- over-express $G\alpha$ (rutabaga), learning index \uparrow in mushroom body but not other parts of brain like central complex

- amnesiac expressed in TBM neuron, modulates memory in mushroom body

- memory trace ~~obs~~ \rightarrow obs comprised of short, medium, amnesiac, long memory components

Tully training spaced increases memory

- LTM depends on protein synthesis for spaced training but not for cramming

- dCREB2, 2 isoforms, hyp-act form \rightarrow photographic memory (Yin Cell 1994, controversial)

stop codon in transgene \uparrow Parvizon 2004

Parvizon

- nalyot affects LTM but not STM

- LTM localized to α -lobes of mushroom body

- Hebbian process - memory is built up by connections of synapses

- Shibire^{TS}, disrupt during learning or retrieval

- gene expression \rightarrow synaptic growth \rightarrow circuitry

\uparrow no strong evidence for molecular growth

Steve Russell

"probe vs. target" reversed

- functional genomics of gene expression.
- high throughput changes the way we deal w/ data
- reverse northern → microarrayed → microarray
- ordered set of frags
- single channel - comparison difference
- dual channel - competitive differences
- PCR - full length cDNA, hetero^{trans} probes, cross-hybs
 - 3' frag from genomic DNA, good if no cDNAs, requires genome sequence/annotation

oligo - 70 - operon
50 - compugen } rely on gen. seq.
30 - amersham
25 - affy

- 40 K element on 1 slide
- inkjet oligo synthesis, in situ, programmable, not in house
- phosphoramidite vs photolithography
- maskless micromirror oligo synthesis - Nimblegen
- 400 000 elements

- efficiency determines quality, off-line you can purify
in situ, take what you get 799%

⇒ problems w/ background & sensitivity

- intern length probe is most specific

- sensitivity ↑ w/ length

- noise / bias ↓ w/ length

- 50-100 bp are ^{probes} ^{good} trade-off b/w spec. & sens.

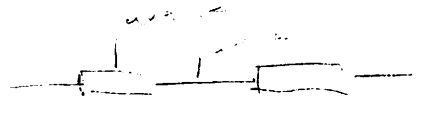
- ^{equivalent} optimize binding energy across oligos w/ T_m

- 18 labs print INDAC

- sacrifice sensitivity for specificity, increase sample

- 150 E INDAC, Affy 300-600 E

- Affy ~25 bp, lack spec/sens, multi probe per gene
- match/mismatch design



- most people ignore mismatch data
- what abt polymorphism?
- subtract mismatch from match to avg over probes per gene
- must amplify to detect mRNAs
- label - 1st strand synthesis (Arberman)

3' end of RNA

2 what is label?

- Eberwine at 7' pol in 1st strand synth
- 10-1000 fold amplification Klebs 2002 "overamplified"
- 1 amplification \Rightarrow 1 bias
- PCR amplification RT + 2 primers

SMART cDNA synth.

- end labelling 1 mRNA \rightarrow 1 label
- use of branched oligo dendrimers
- Estrada (2005) Plos Genetics, gfp + cell sorting
- recombining gal4/gfp onto μ clone

- Experimental design (Yang & Speed Nat Rev Genet 2002)

- time course T_1 vs T_2 vs T_3 vs. $T_1 + T_2 + T_3$
- biological replication is most imp't, pool to save \$
- 3 smooth biol replication

- wt vs μ 4 biol 3 dye swap \sim 8 slides

- spot finding, normalisation

- idea behind norm is that majority of genes are unchanged

- M^{diff} vs A^{norm} \log_2 scale

- Affy norm MAS5, Speed GCRMA is best

- affy estimates are poor for genes at low expression levels

- good correlation btwn Affy & Spotted results

- ~~Stolc~~ N. Bleaen, \sim 40% of non-exon detect expression, genome-wide

- Affy genome tile array, 25 bp every 35 bp
- 106 Mb

lecture

share from
path to chips

email at
pages 3-4 (11)

C. No. 3

- J. Manak

- | | | |
|----------------|------------------|-----------------|
| 60% known exon | } - significance | |
| 9% EST | | - unannot exons |
| 23% intron | | - in mammals |
| 7% intergenic | | |

IGB ~ integrated genome browser

- Slamon & Janschavsky chip
- chromatin immunoprecipitation
- chip array reproducibility higher than expt array
- good correspondence between 1 kb tiling vs. Affy whole genome tiling array
- couple w/ cell sorting
- depends on quality of antibodies
- DAMID, leaky promoters or overexpression leads to no resolution, try to couple processivity
⇒ can't drive in tissue specific manner
- similar results DAMID vs Chip
- Andreea Brand Nimble gen data

proteomics

- yeast 2 hybrid, 9% in common in 2 expts
- very noisy data, why? dimeric, foreign conditions
- Mass spec:

Peptide Mass Fingerprints

need genome & good annotation

post translational mod change mass

- MALDI-TOF → peptides

bottom up proteomics fragmentation tandem mass spec → get AA seq from lab

- 2 D gels

- DIGE Góng (2004) Development 2D gel w/ Cy5 & Cy3

2.1 PG 97.8/110

- membrane proteins don't go into gels
- number of ~2000 genes
- ICAT, Rudi Abersold, Moseley 2001 Trends Biochem
- 2 labels, heavy water
- iTRAQ, more labels
- Peptide Atlas www.peptideatlas.org Tandem MS
- ~5% of spectra can be interpreted

RNAi knockdown

- R. Paro's amplicons had T7 & T3 incorporated!
- Perrimon Lab
- reproducibility is good
- second generation Michael Boutros
- UAS constructs
- Echeverri & Perrimon NRC 7:373
- cell arrays, print dsRNA