

Figure S1. Subcellular localization of human ADAR isoforms, and verification of samples prior to RIP-seq by western blot and silver staining. (A) Fluorescence microscopy showing GFP signal of each overexpressed mycGFP-tagged ADAR isoform or mycGFP expression constructs in HeLa cells 48h post-transfection. Green indicates GFP signal, and blue indicates the nucleus stained by DAPI. Bar, 20 μ m. Western blot (B) and silver staining (C) of input samples and IP samples immunoprecipitated with anti-GFP antibody (GFP) or control IgG from cells transfected with each ADAR-expressing construct (pmGFP-ADAR1-150, pmGFP-ADAR1-110, and pmGFP-ADAR2) or pmGFP control. RIP-seq was carried out using input samples and IP samples with anti-GFP antibody. Red arrowheads indicate the positions of the expected molecular weights for each mGFP-tagged protein.

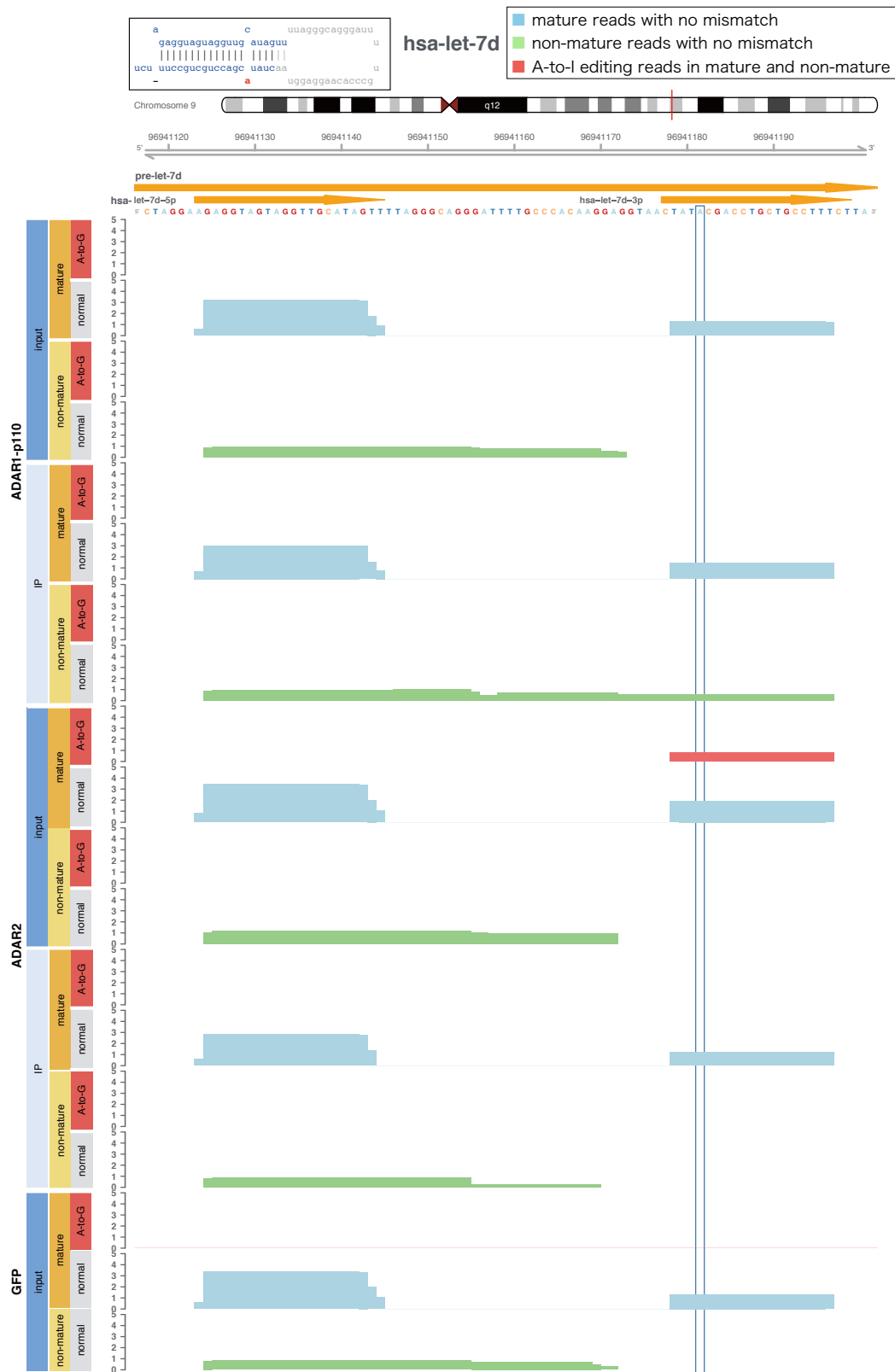
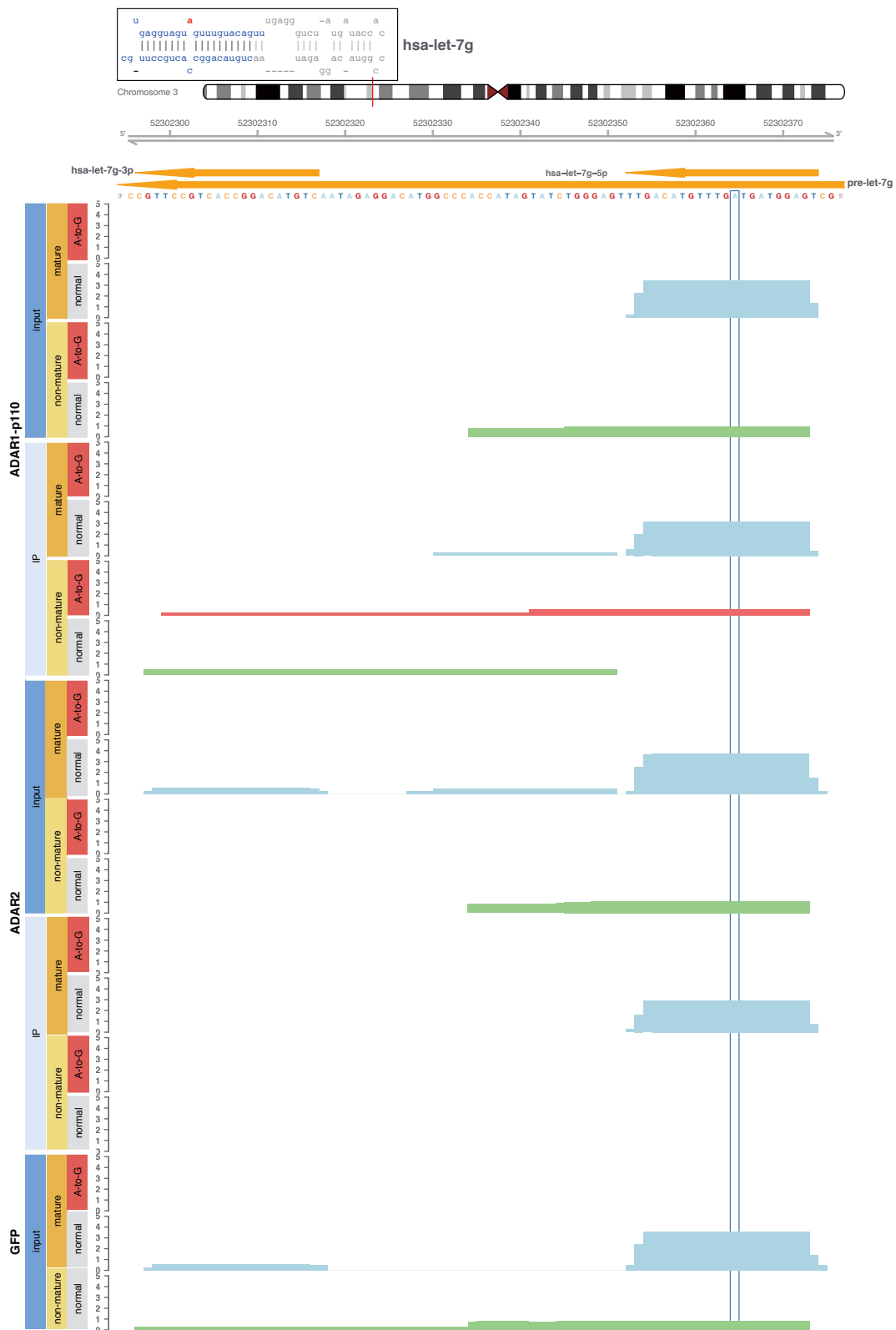
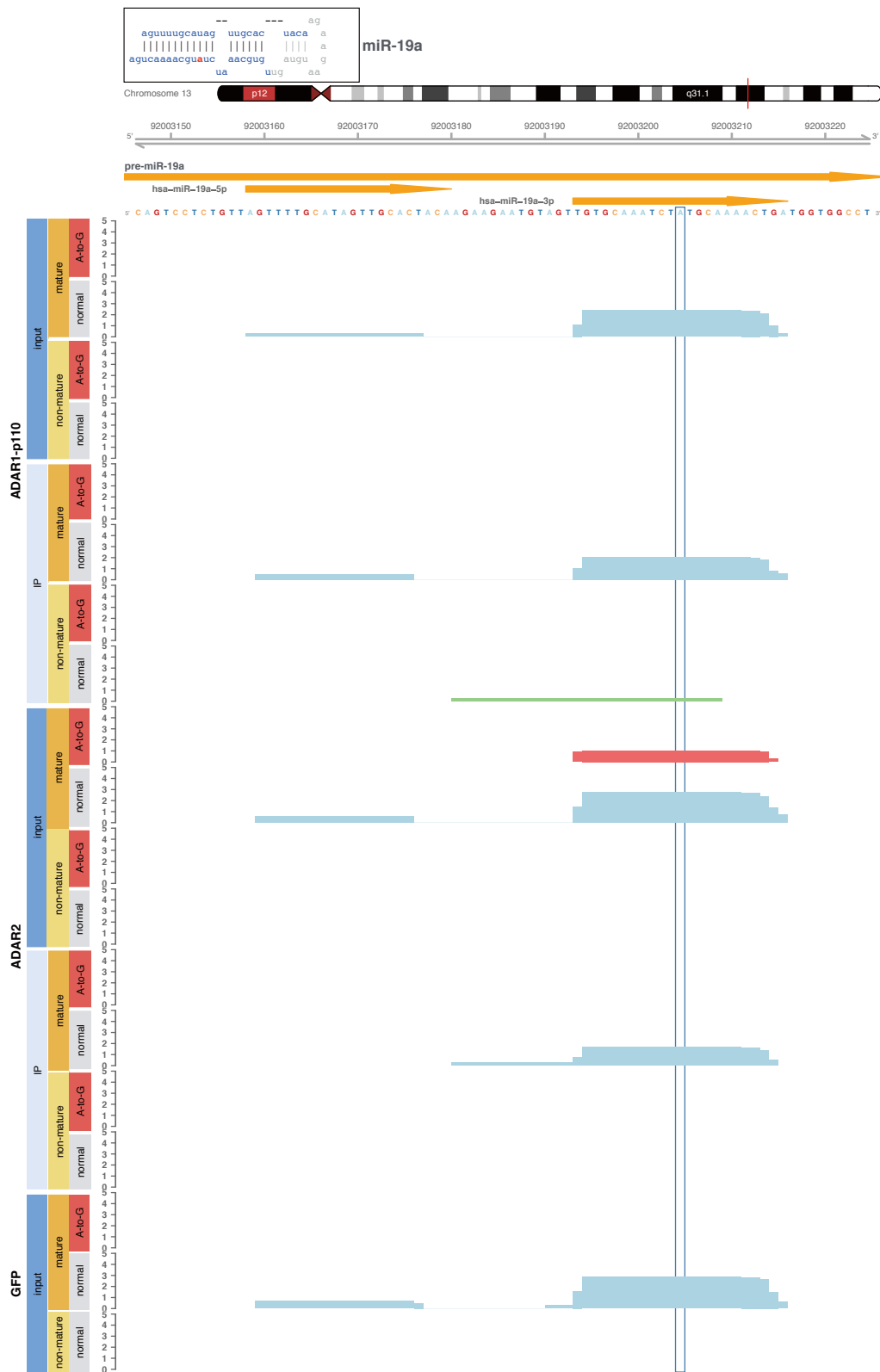
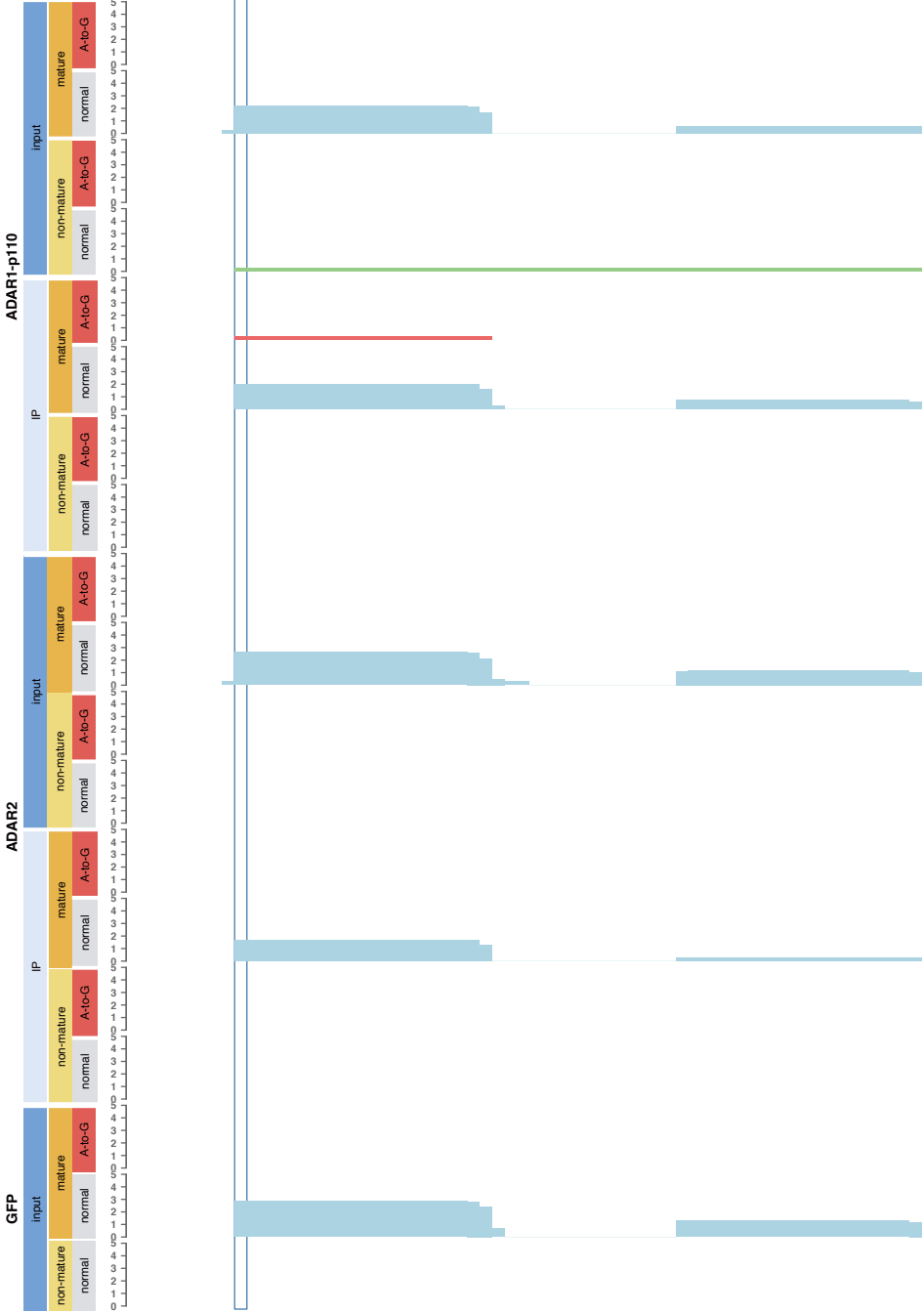
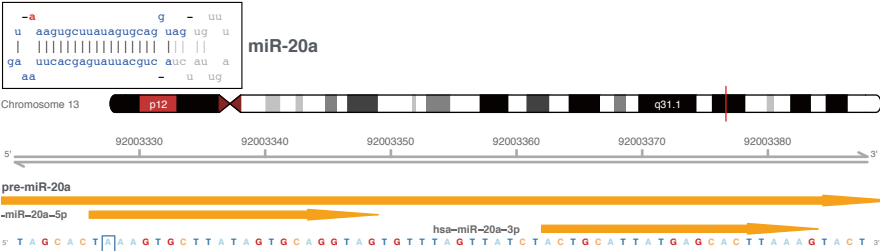
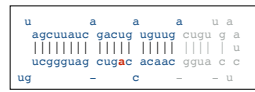


Figure S2. Genome browser view of all normal and collapse reads mapped to miRNA annotations. The editing positions detected in this study are shown by dark blue boxes for A-to-I edited miRNAs and pre-miRNAs. The secondary structures of the corresponding pre-miRNAs are shown in the upper left region as they appear in miRBase. The location of the miRNA on the chromosome is indicated in red. The genome coordinates are indicated on a gray axis under the chromosome. The mature and pre-miRNA annotations registered in miRBase are shown in orange, the arrows indicating direction of transcription. Mature miRNA reads with no mismatch are in light blue, non-mature miRNA reads with no mismatch in light green, and mature and non-mature miRNA reads with A-to-I edited reads in red. The y-axis of all tracks is in the log10 scale.

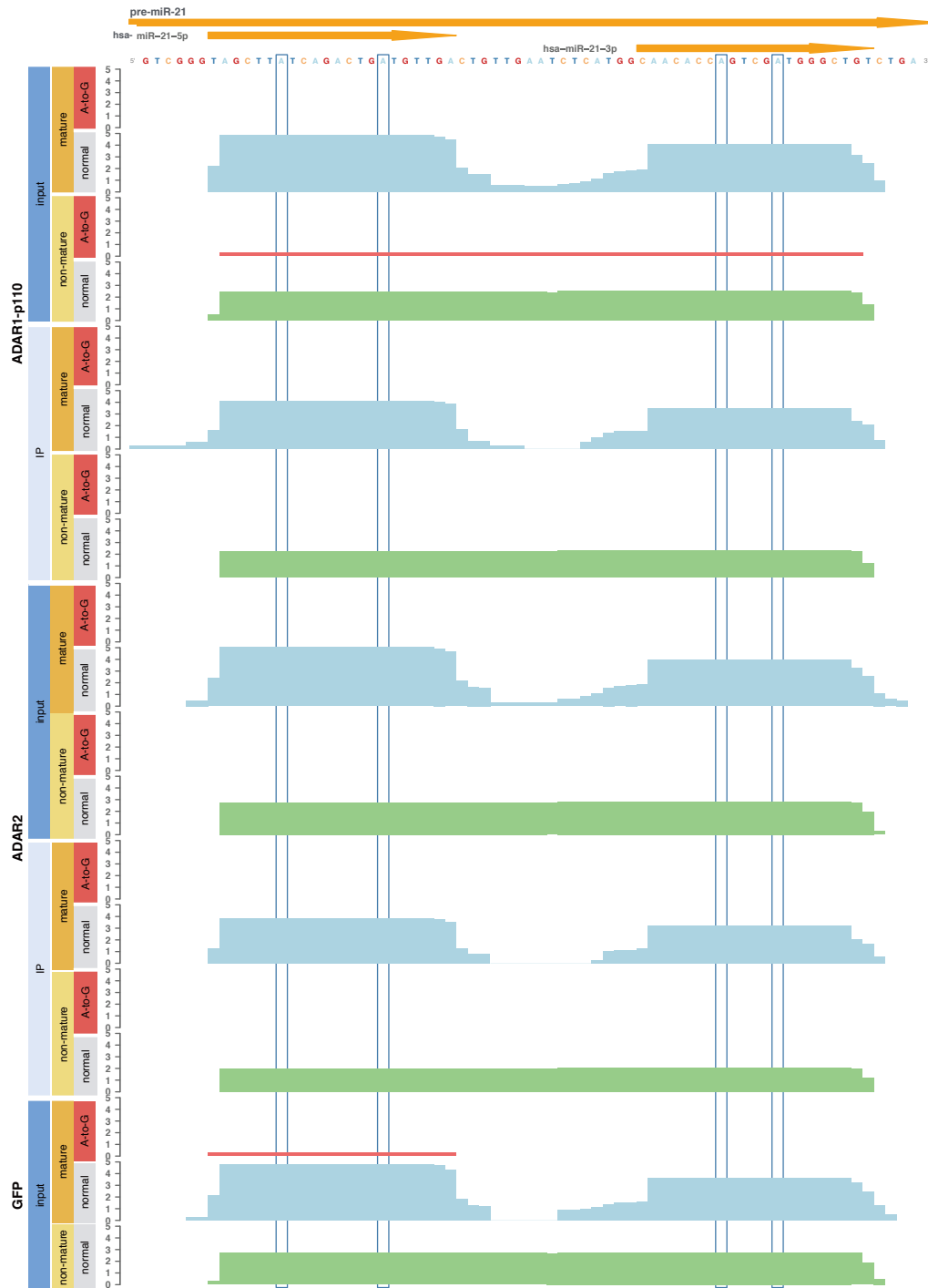
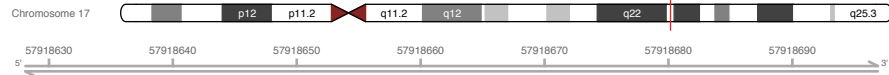


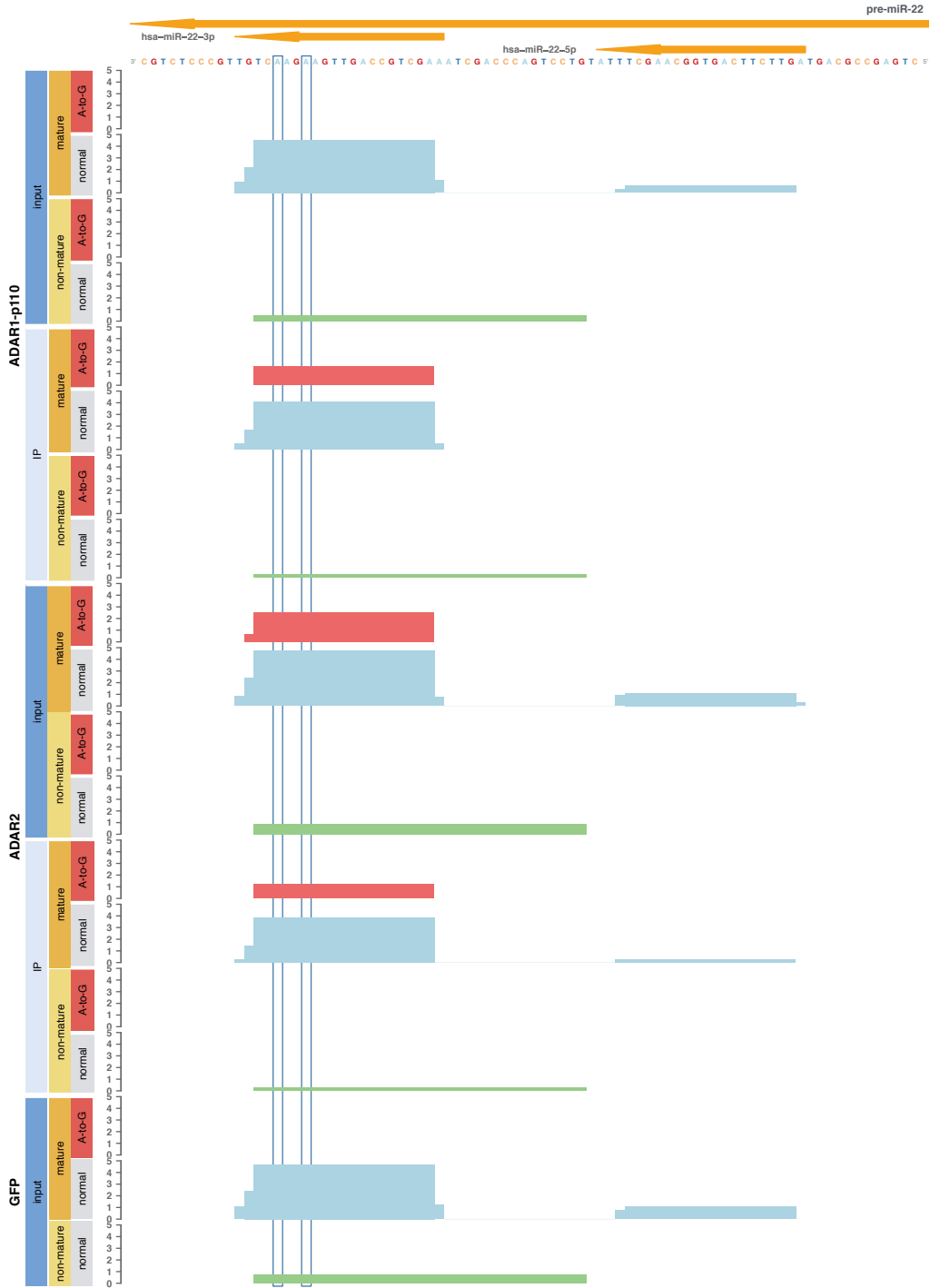
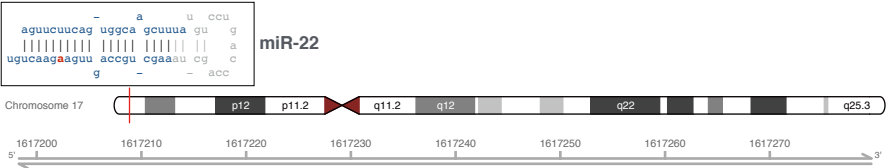


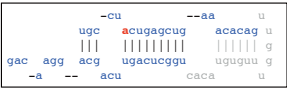




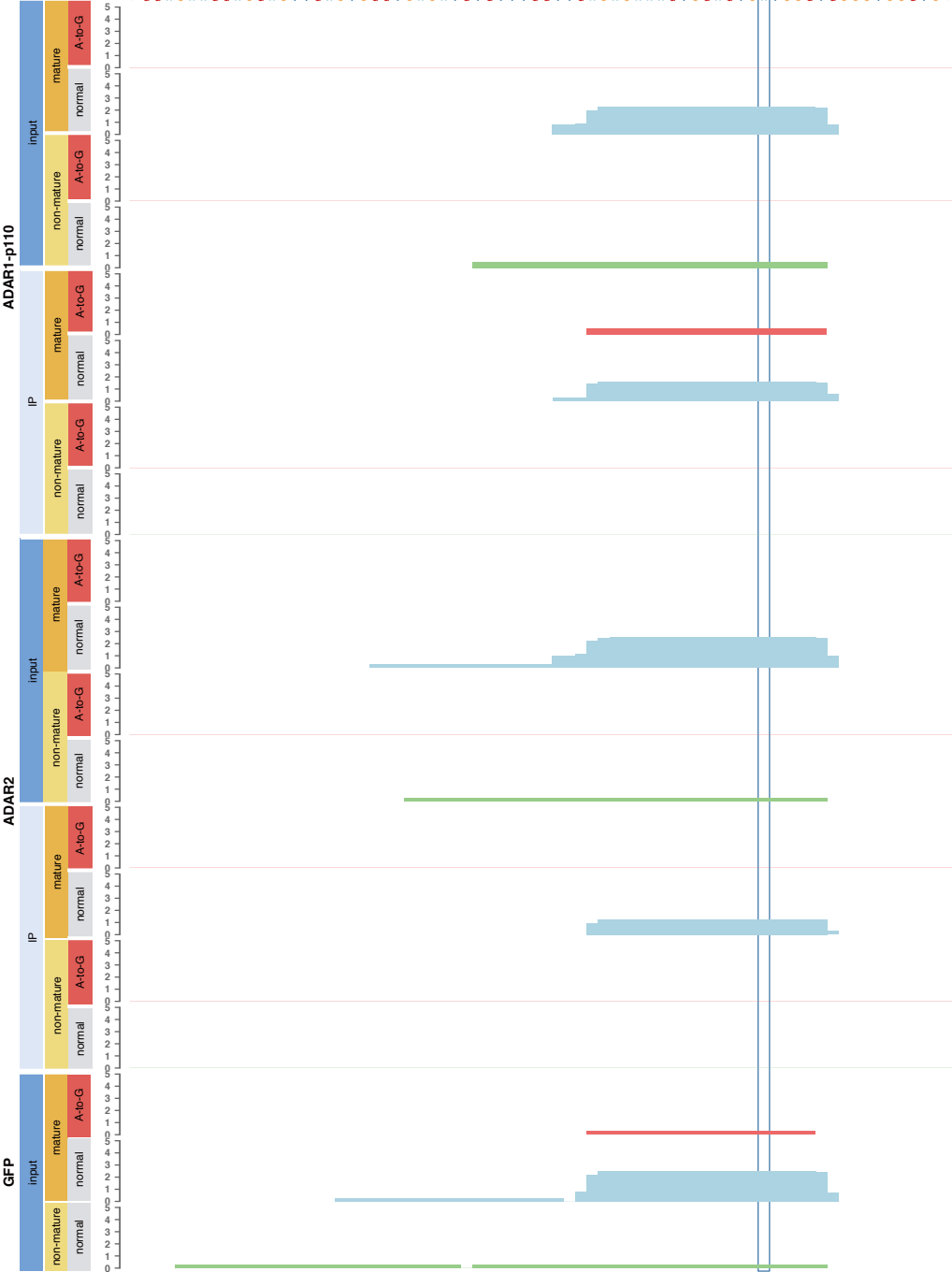
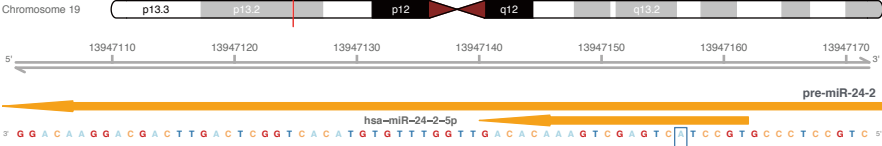
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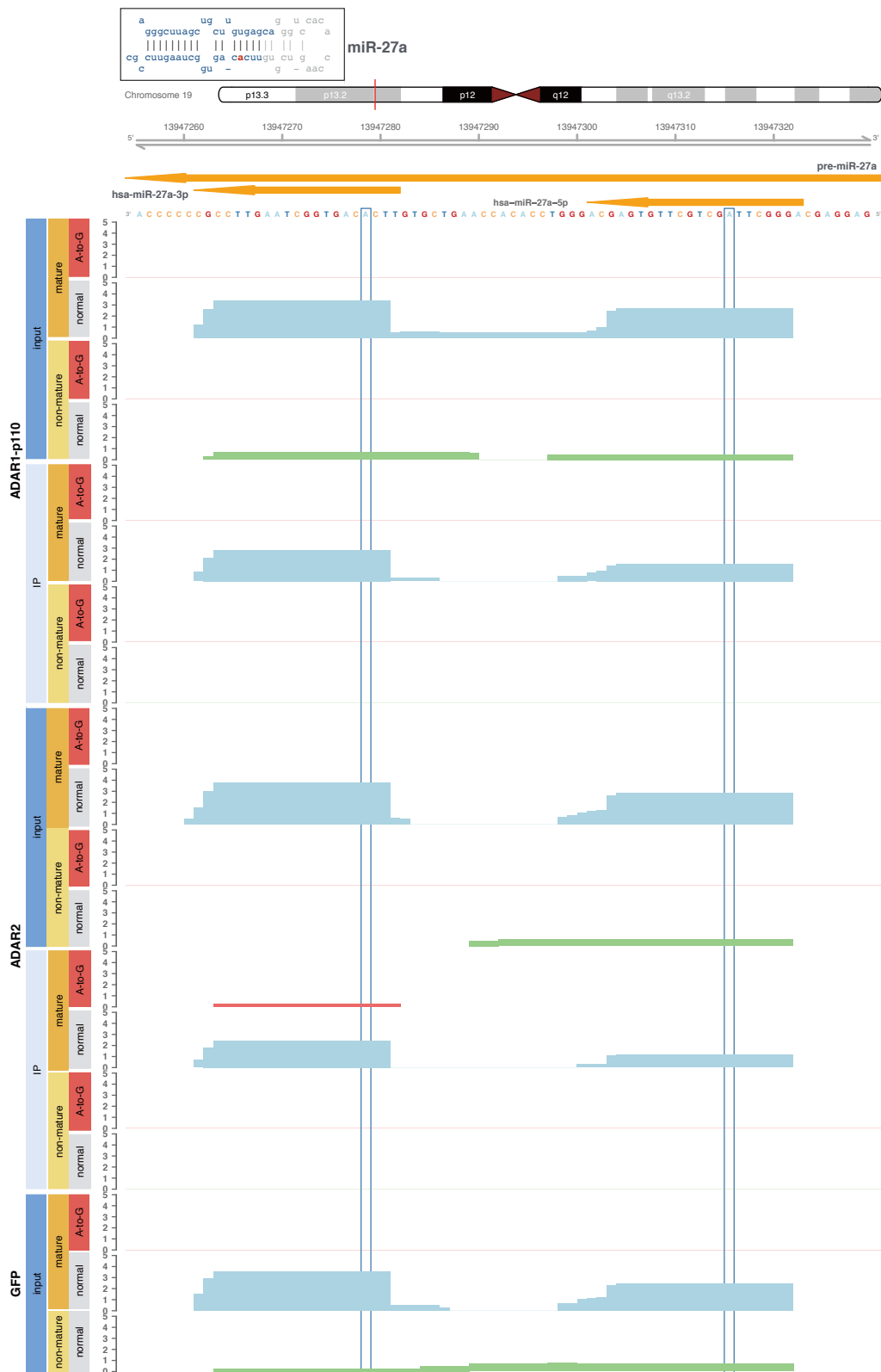


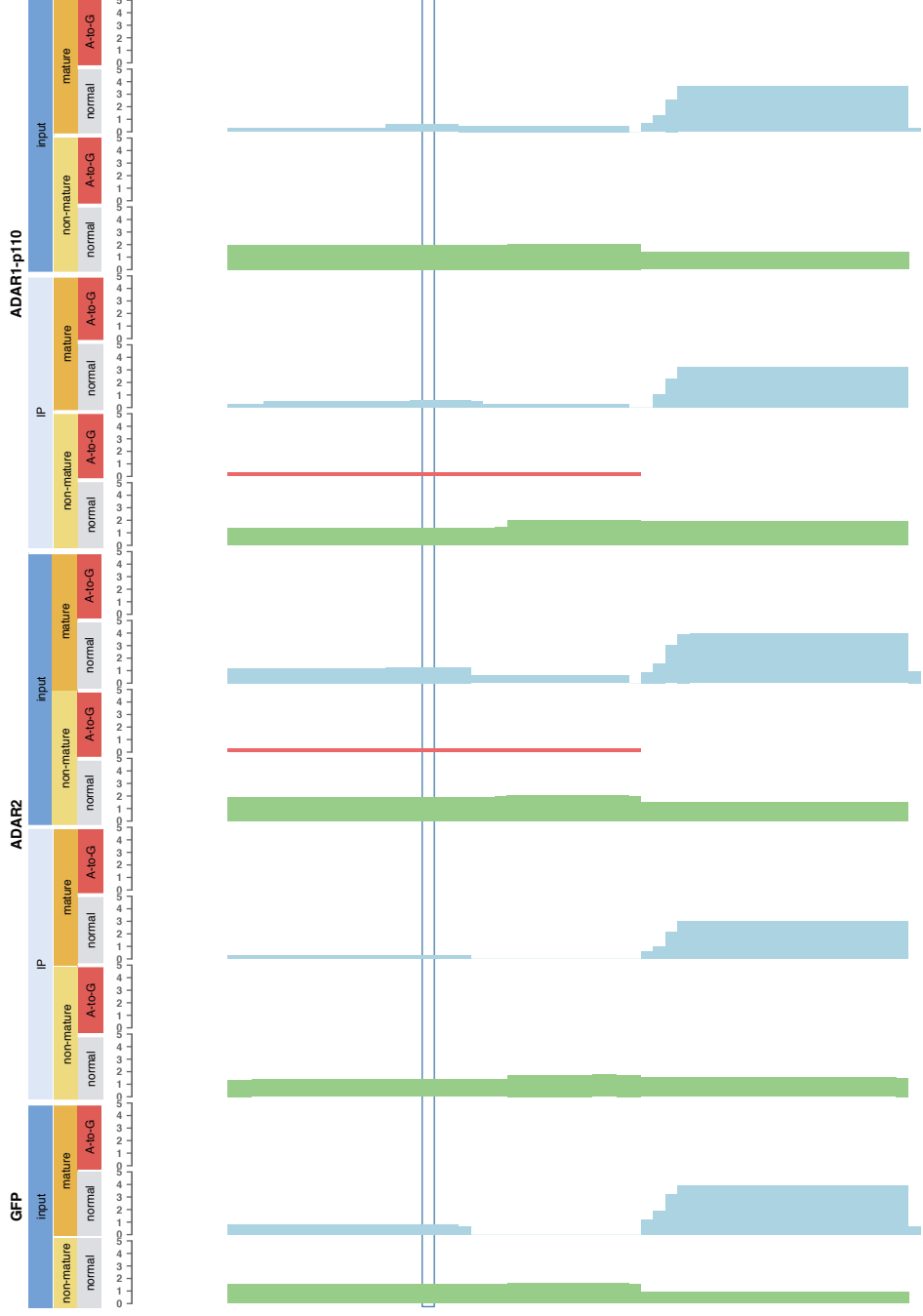
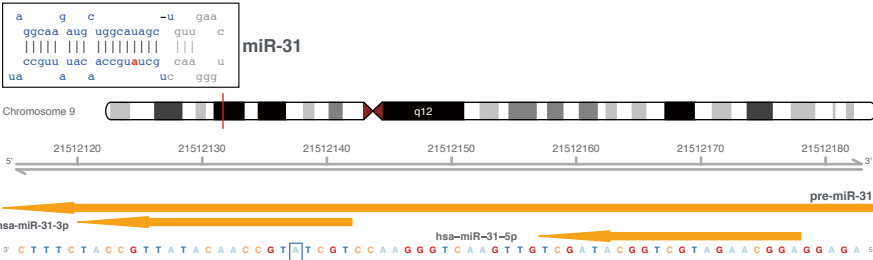


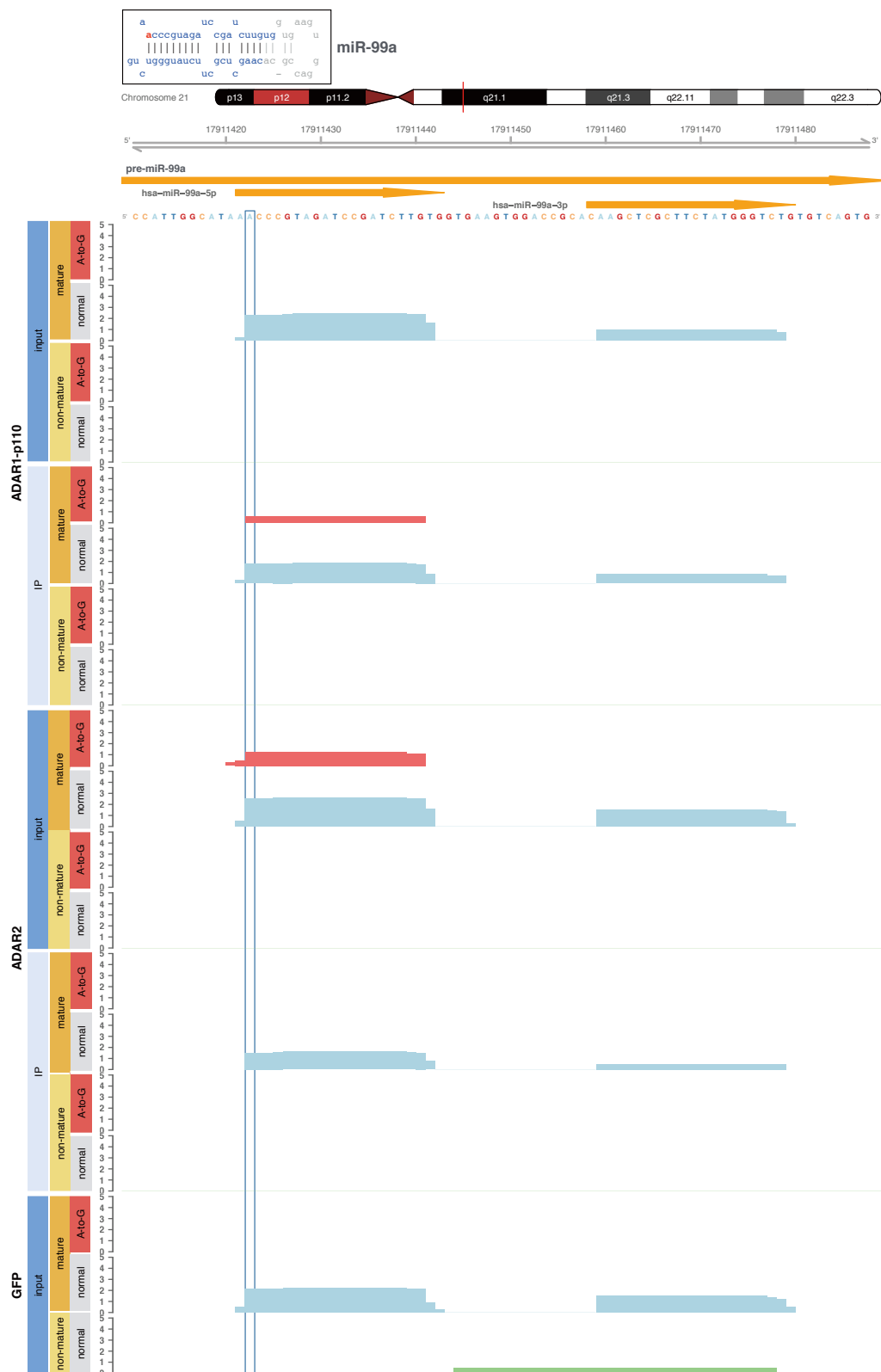


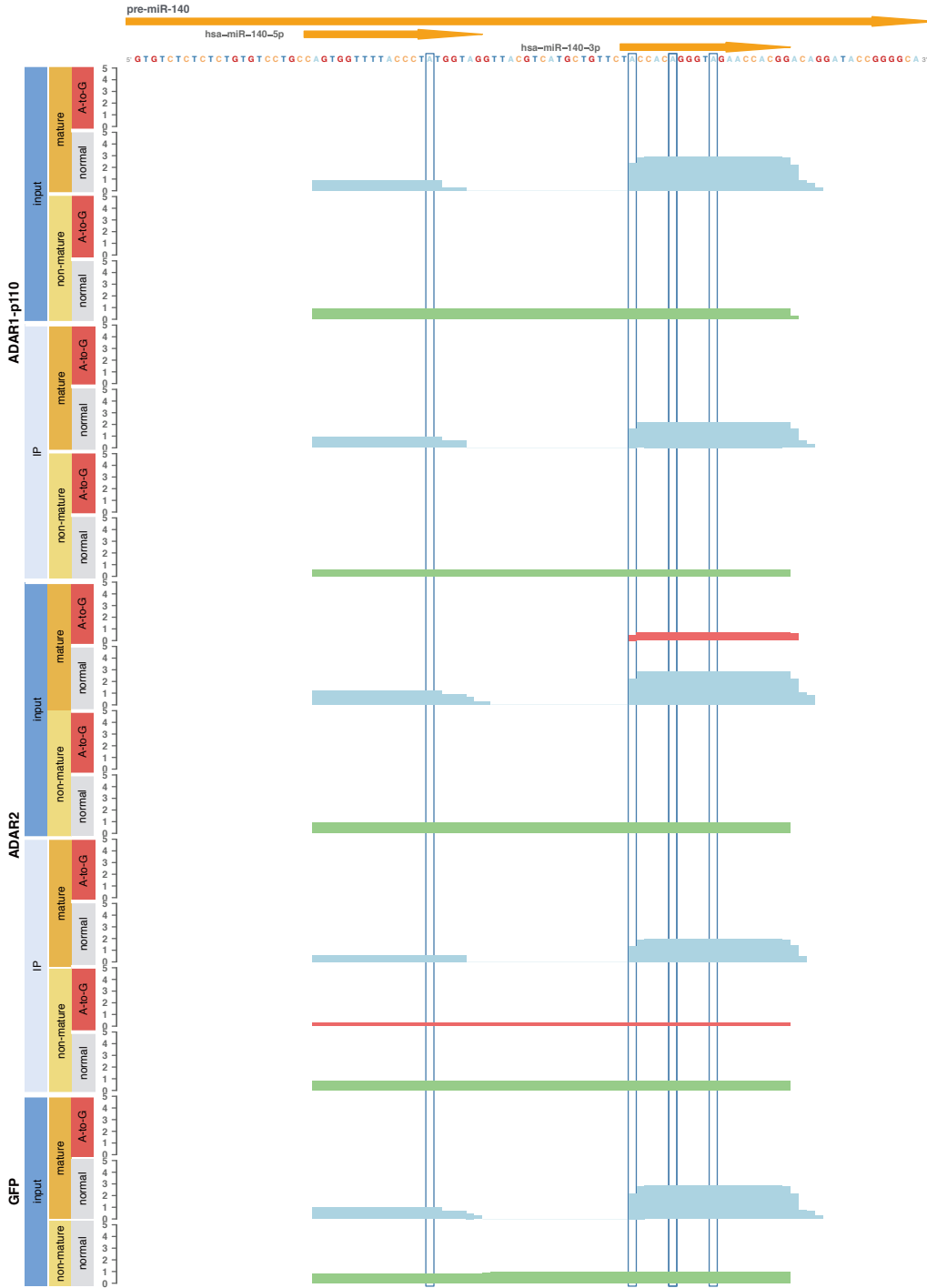
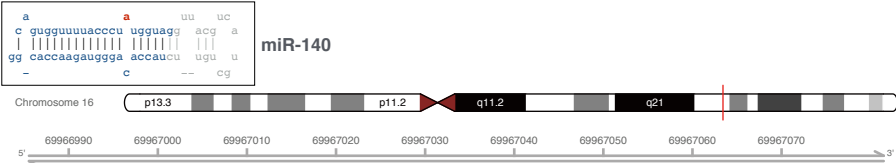
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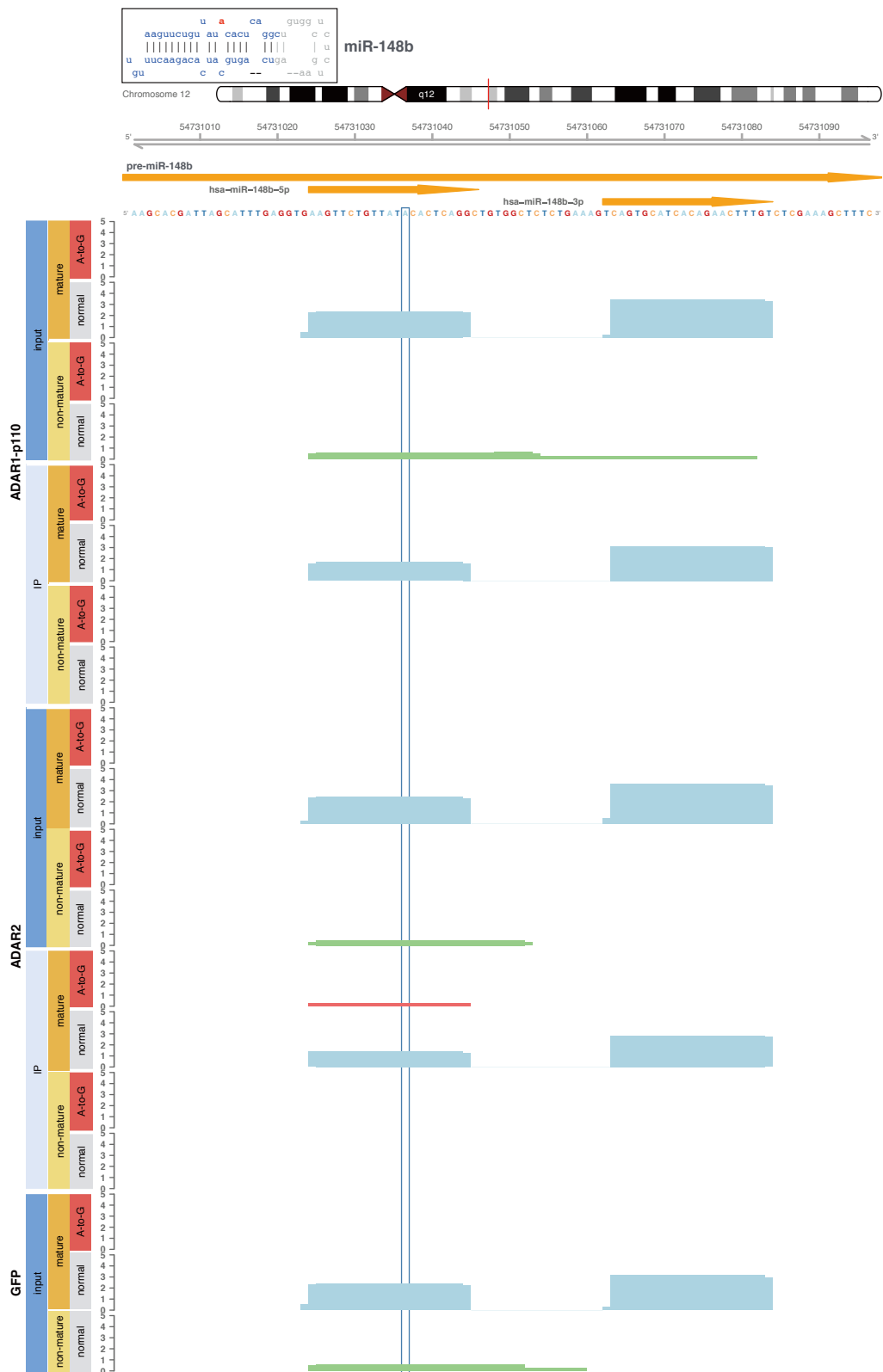


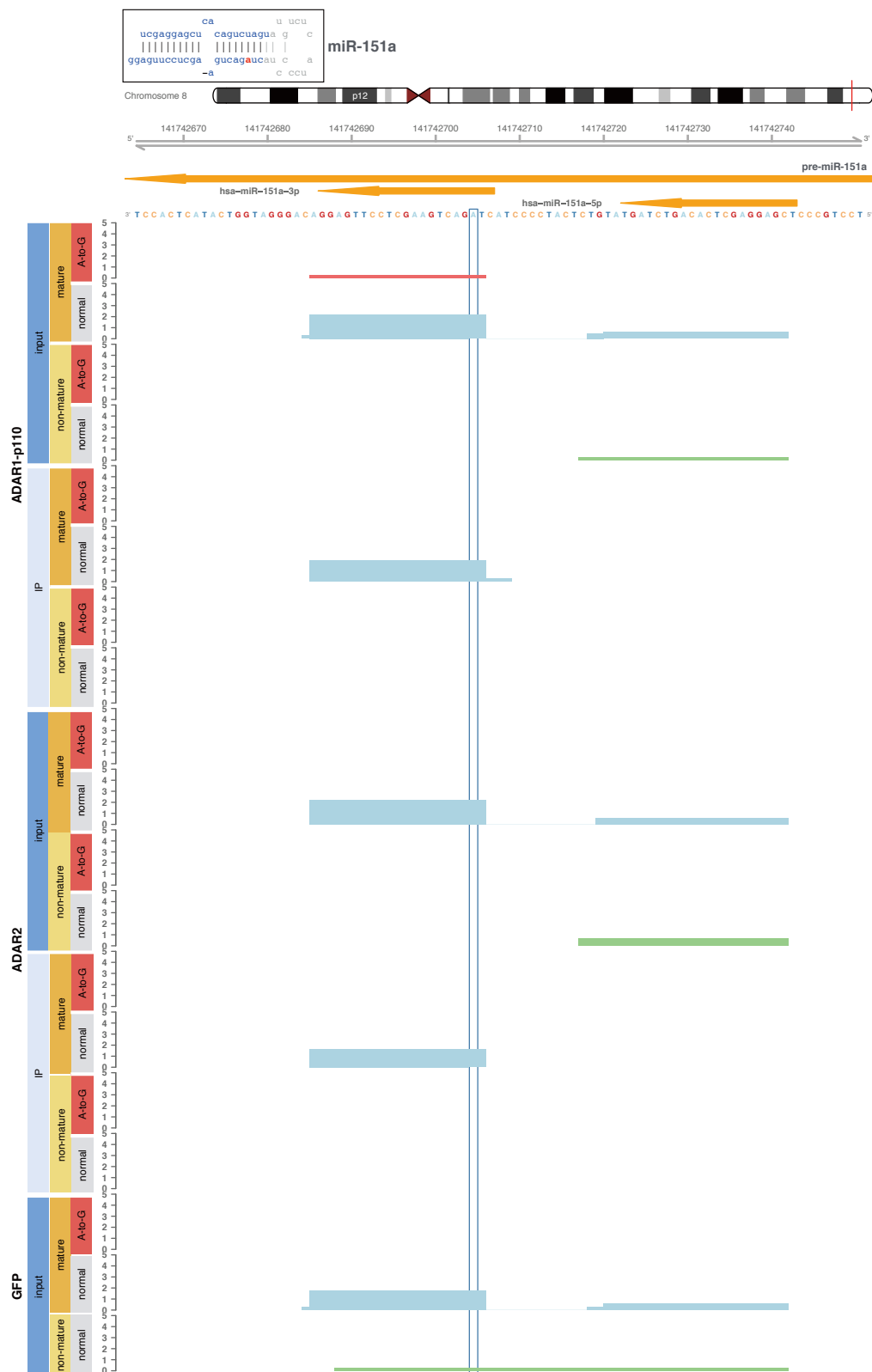


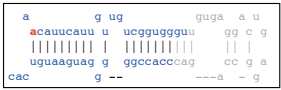




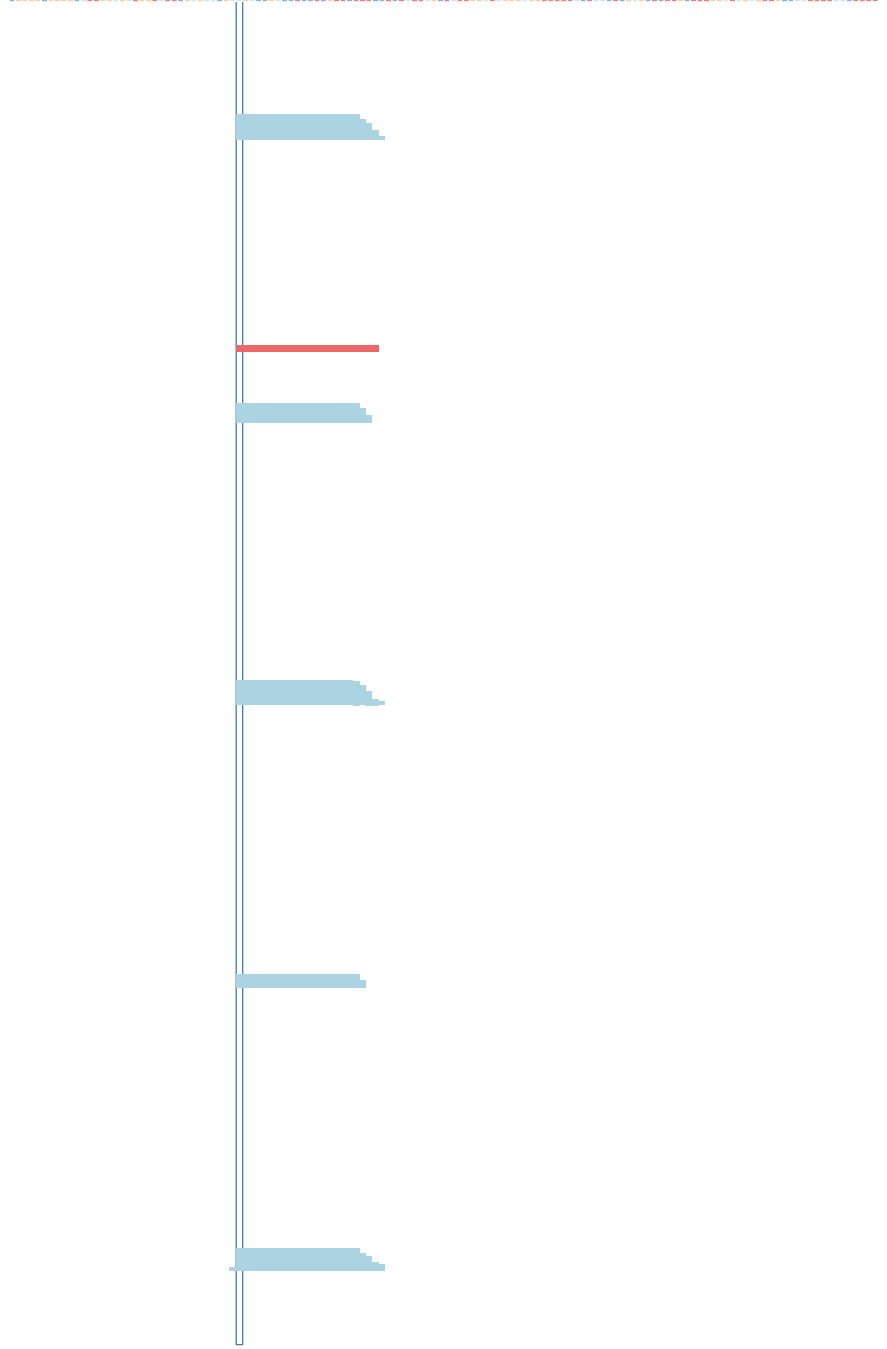
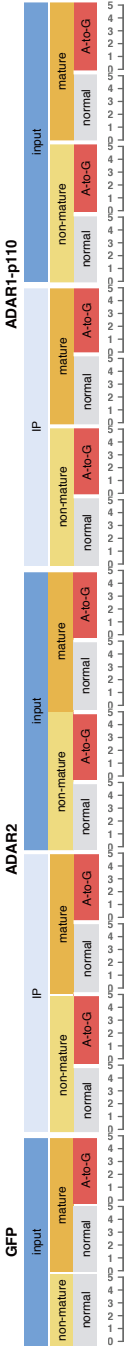
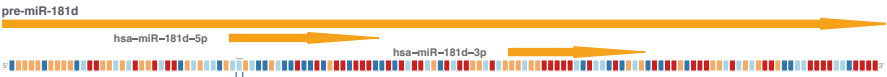


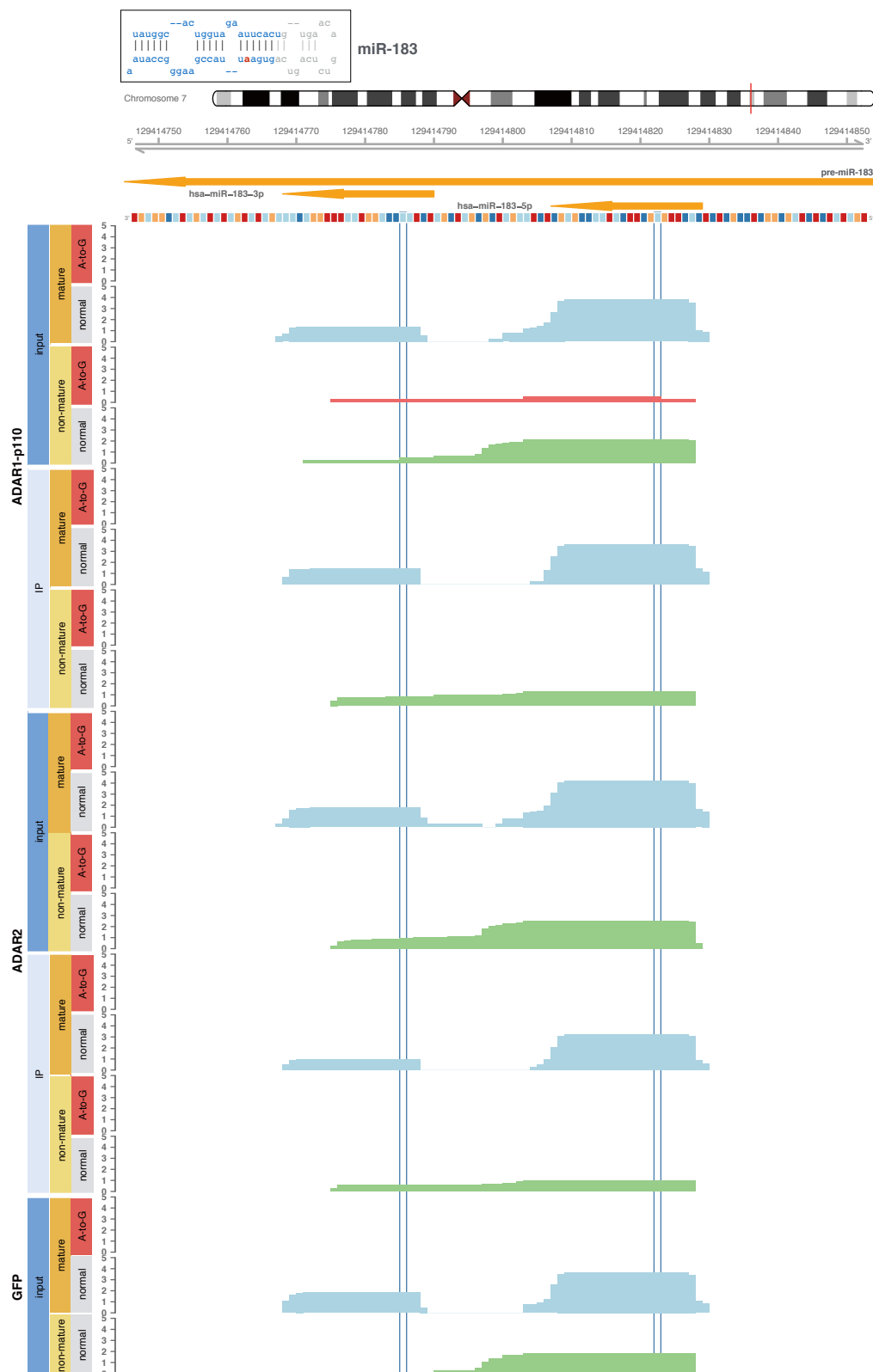


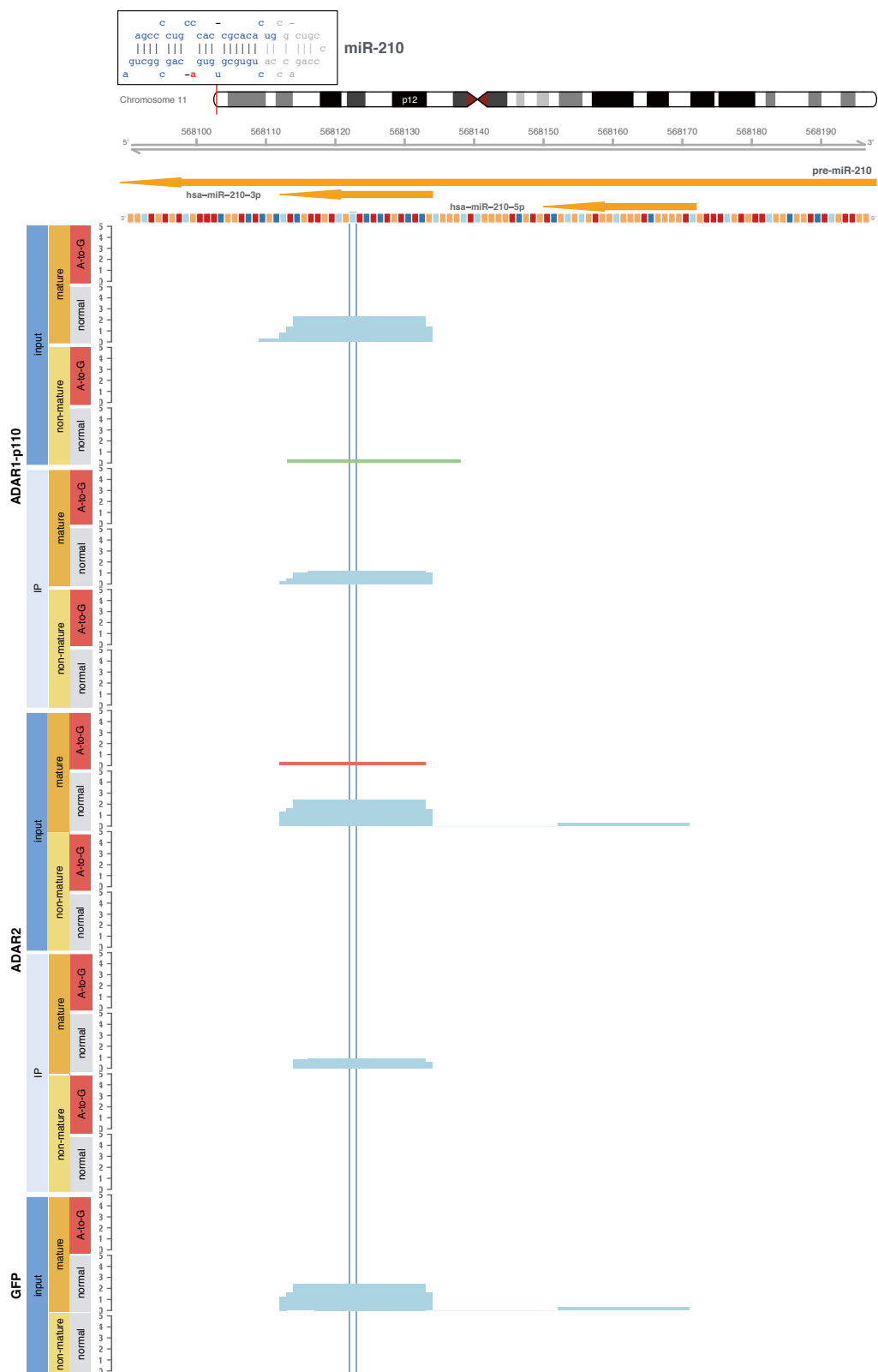


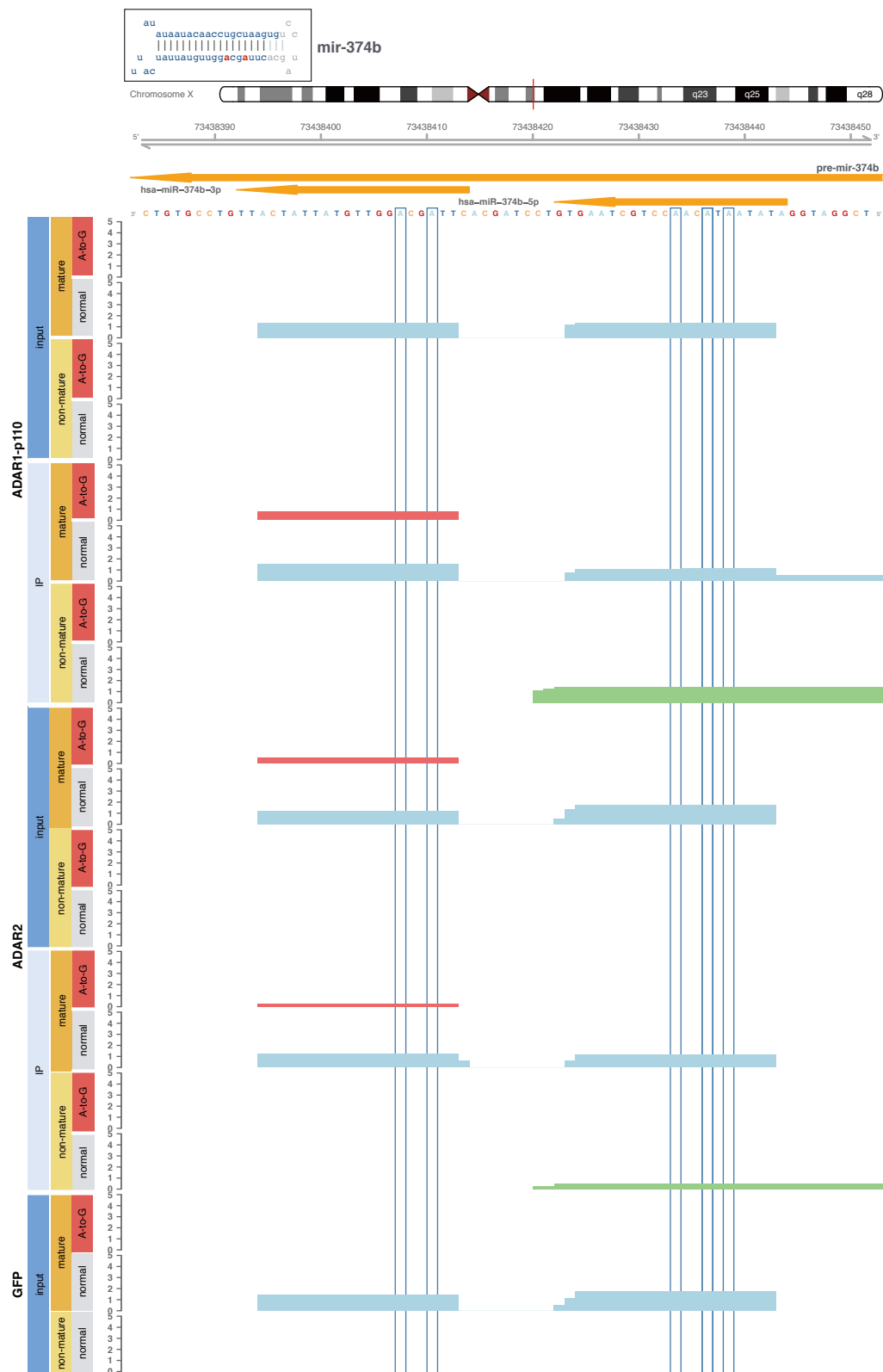


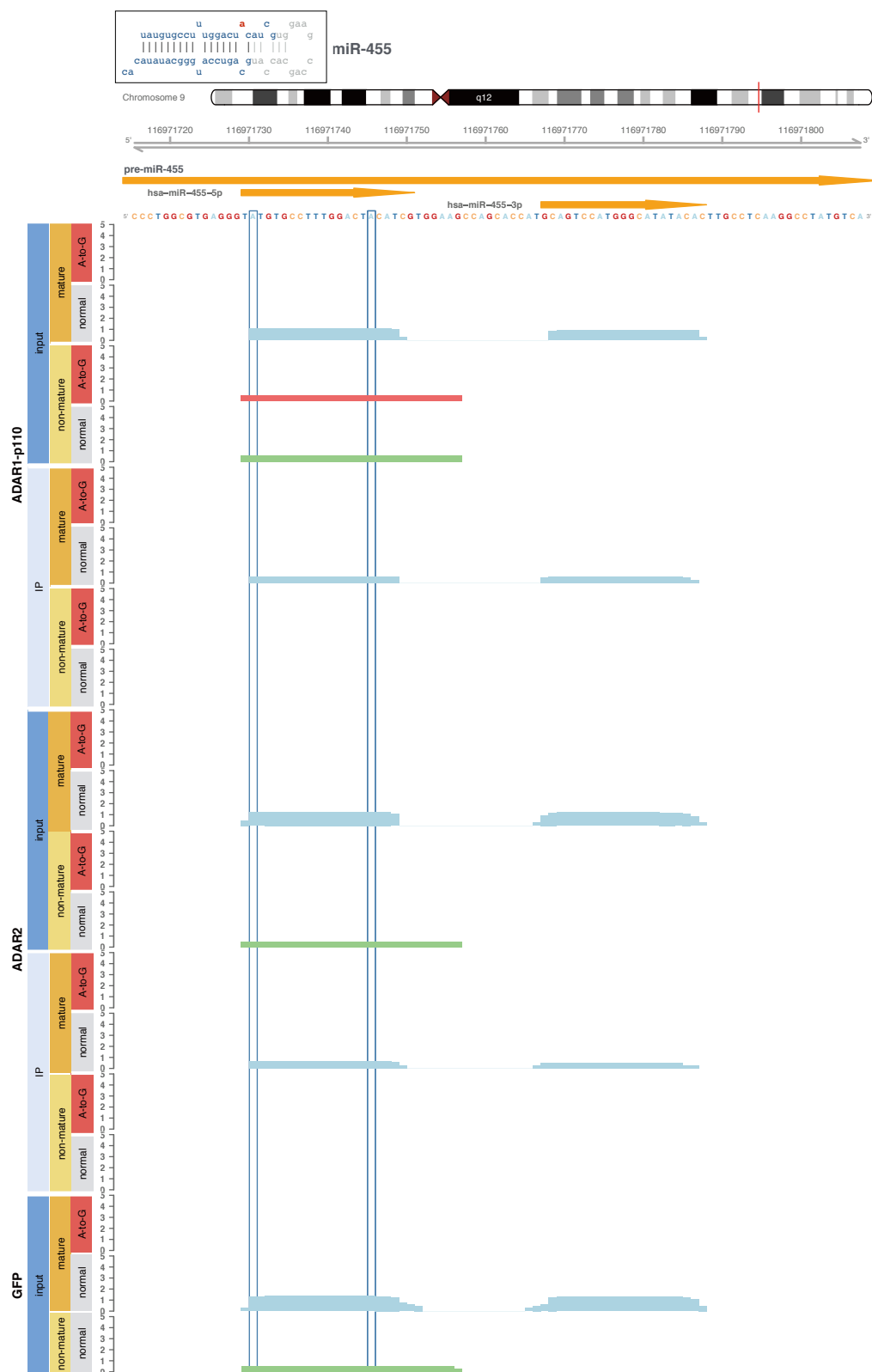
miR-181d

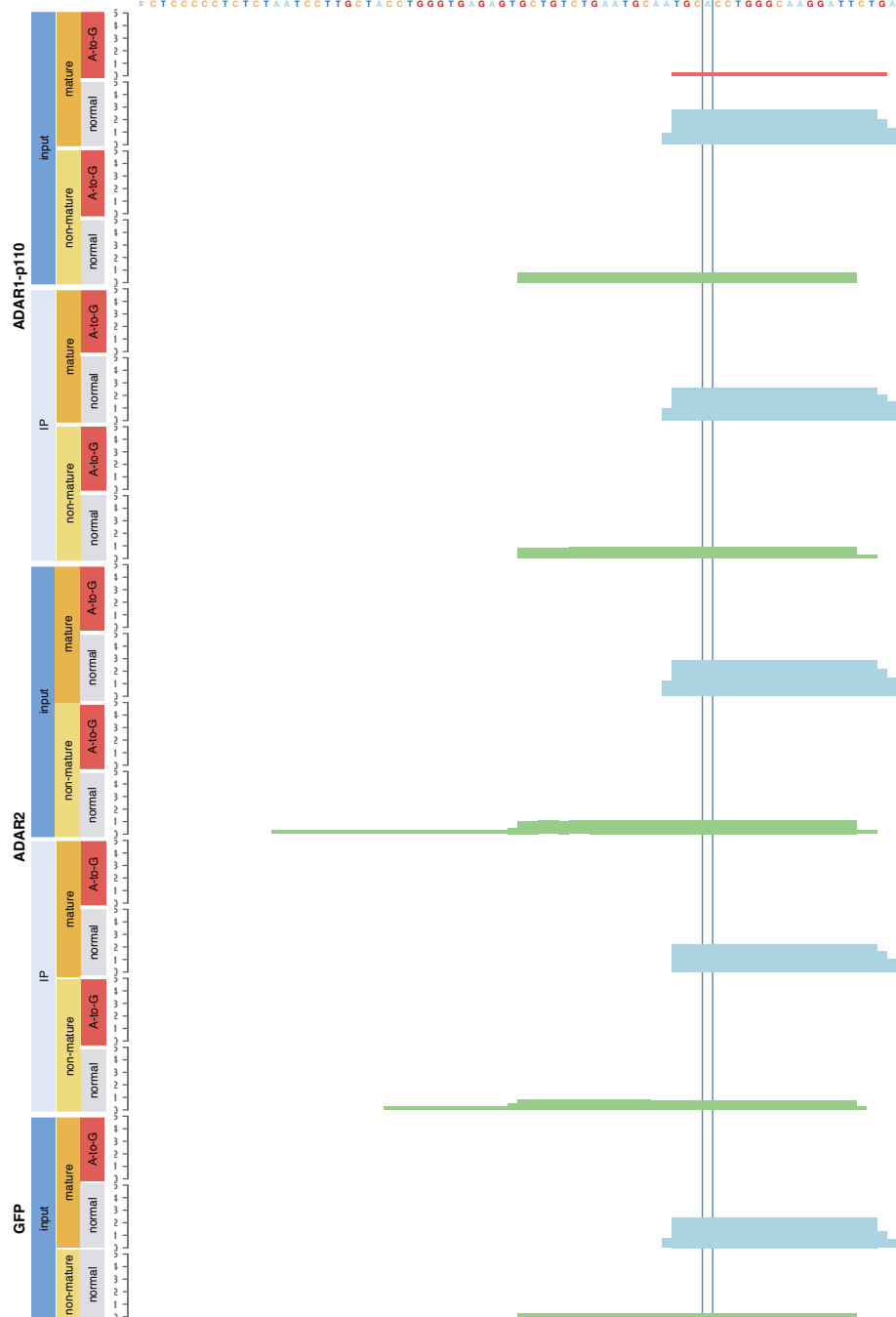
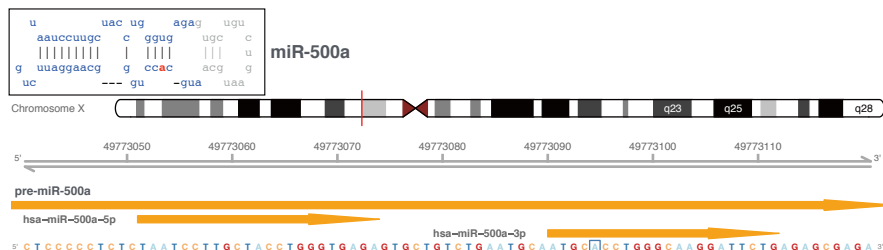


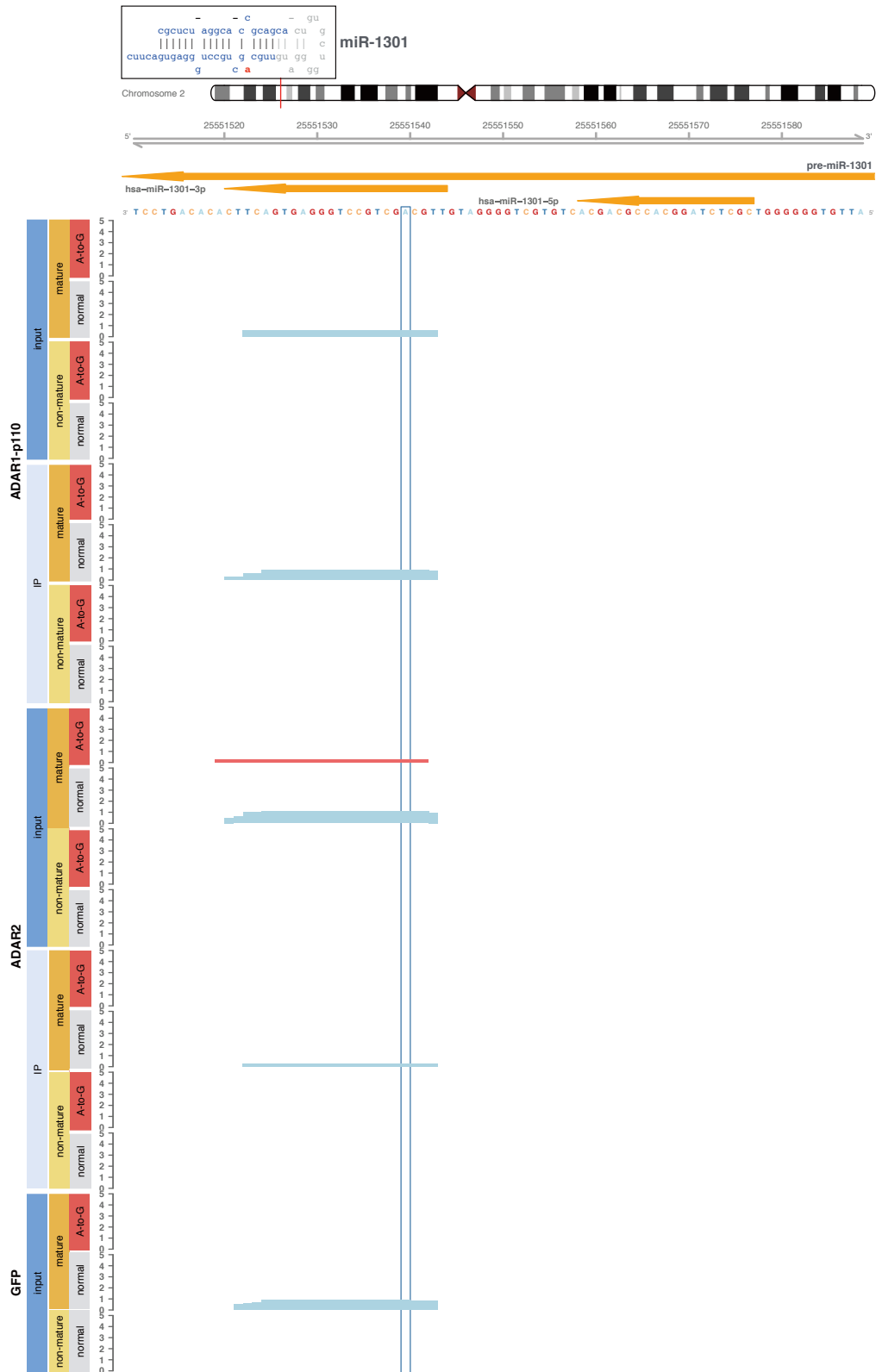


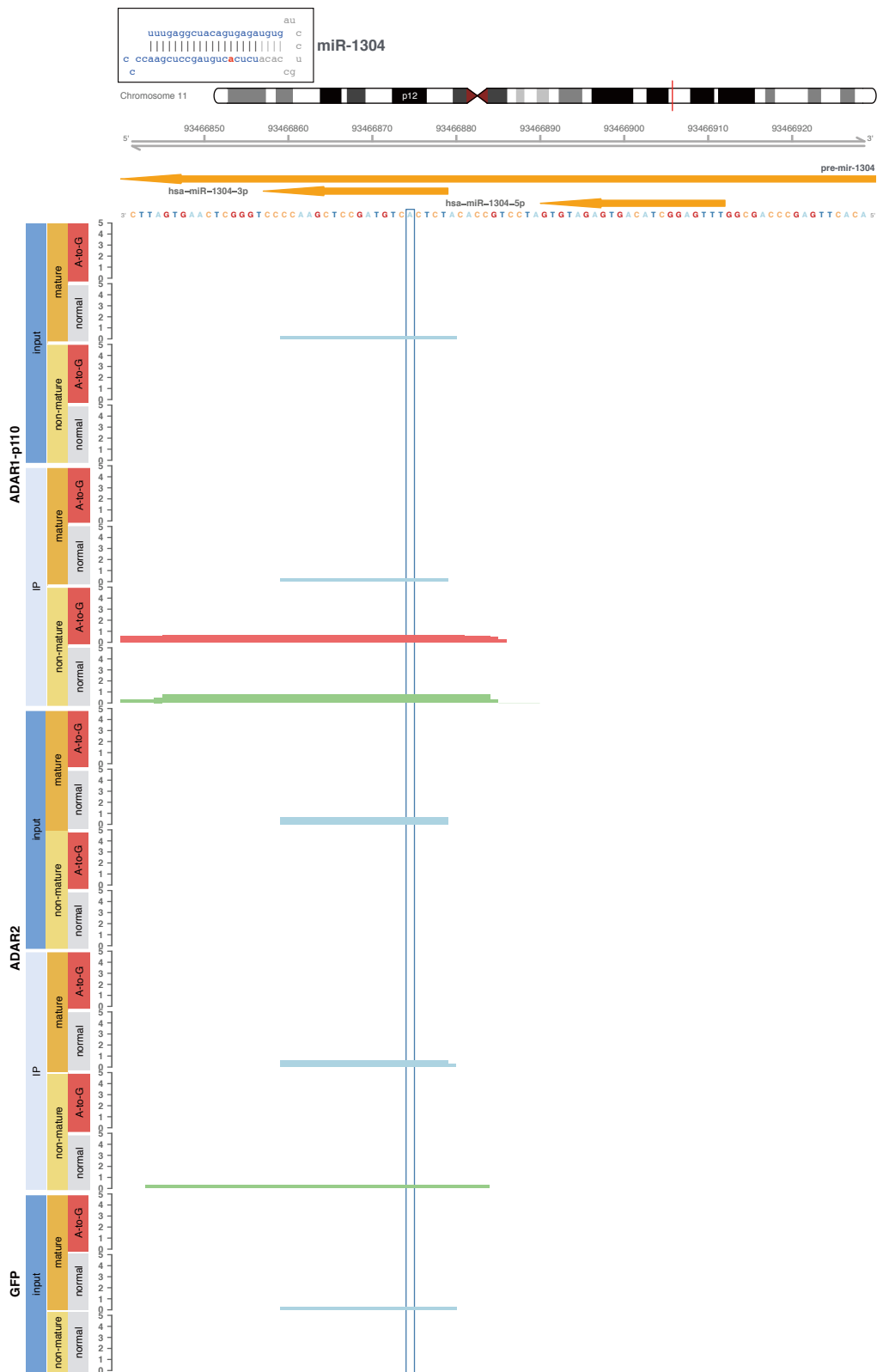


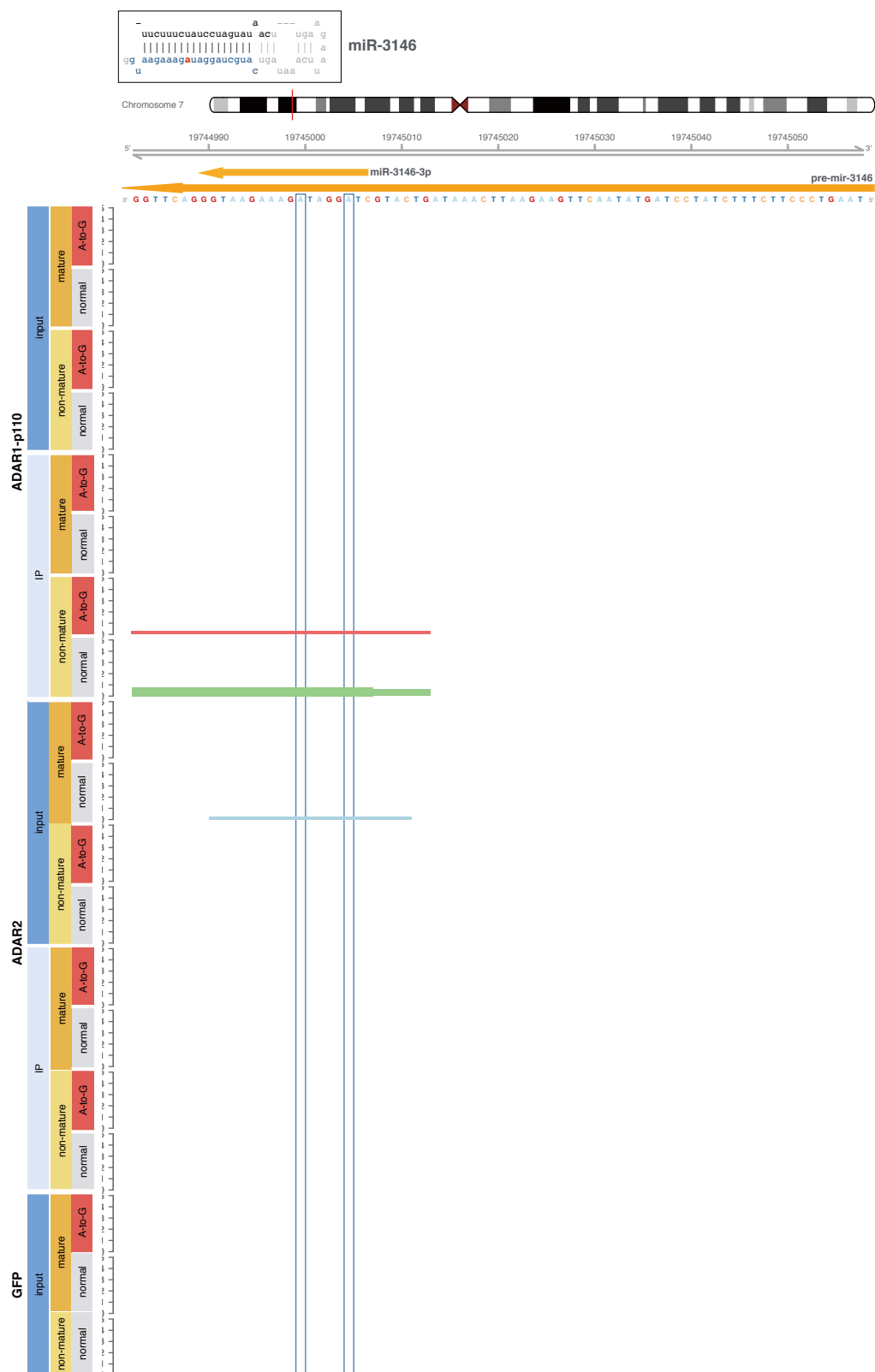


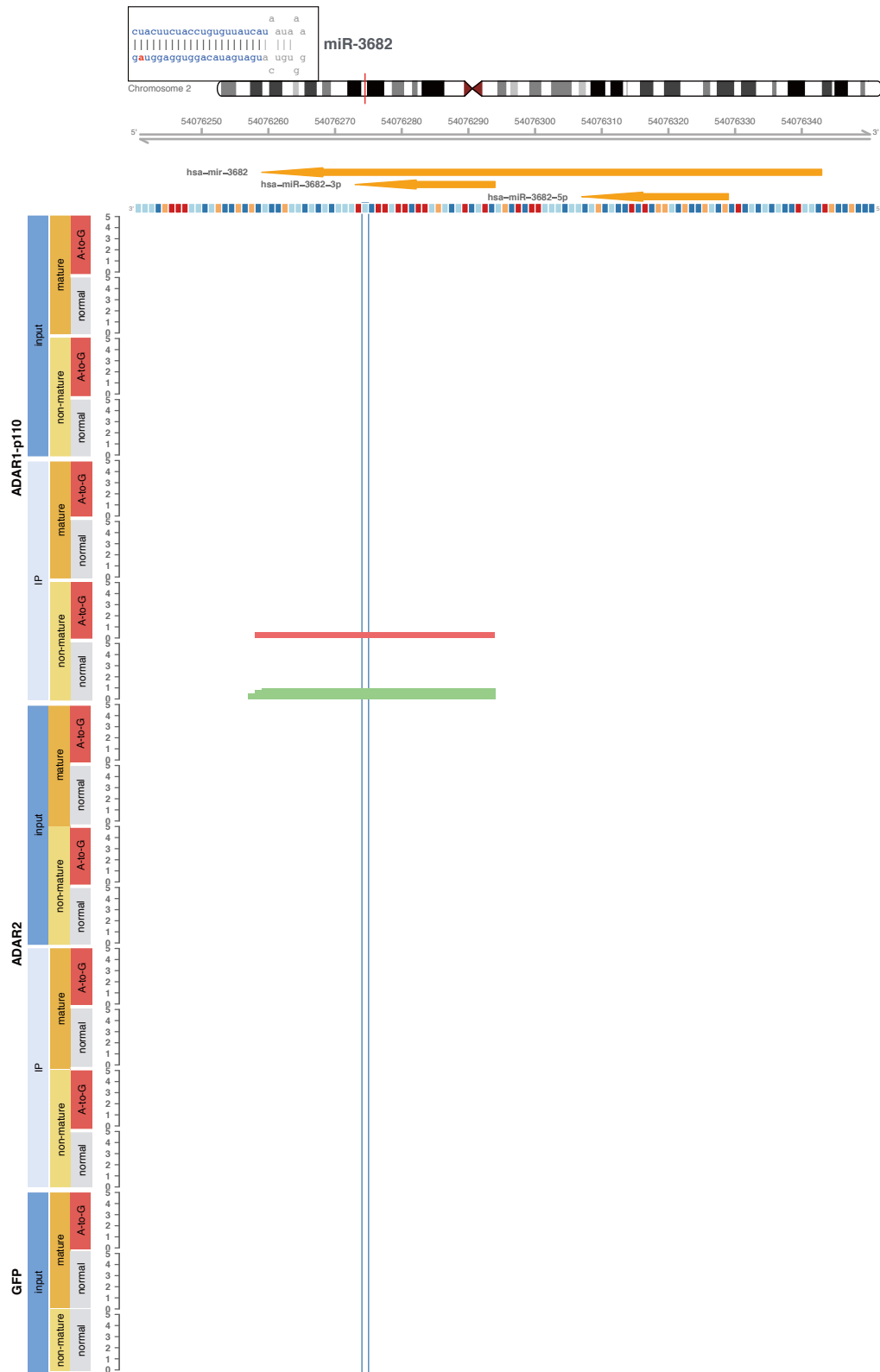


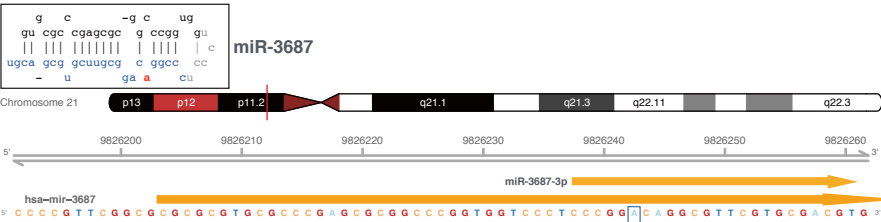


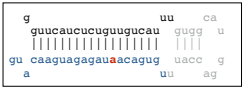












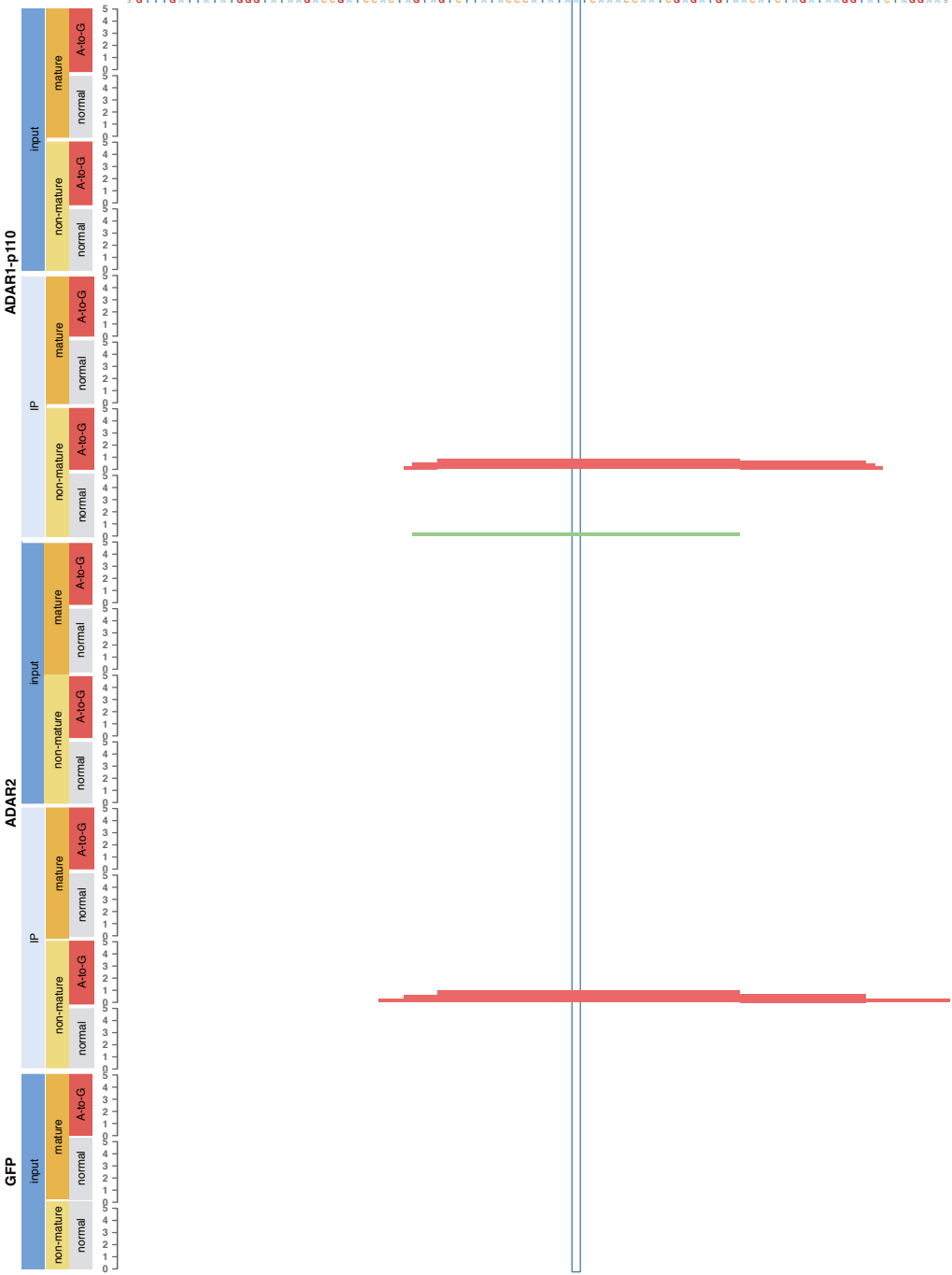
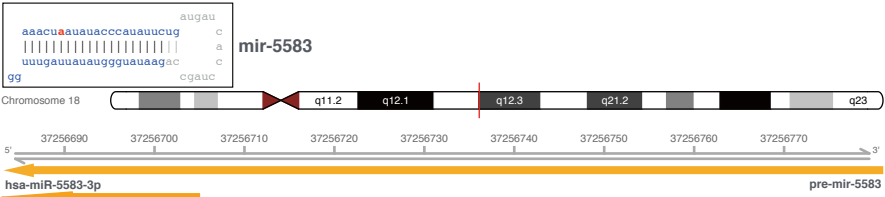
miR-4504



ADAR1-p110			
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non-mature	mature	non-mature	mature
normal	A-to-G	normal	A-to-G
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2	2	2	2
3	3	3	3
4	4	4	4
5	5	5	5
6	6	6	6
7	7	7	7
8	8	8	8
9	9	9	9
10	10	10	10
11	11	11	11
12	12	12	12
13	13	13	13
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94	94	94	94
95	95	95	95
96	96	96	96
97	97	97	97
98	98	98	98
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100	100	100	100

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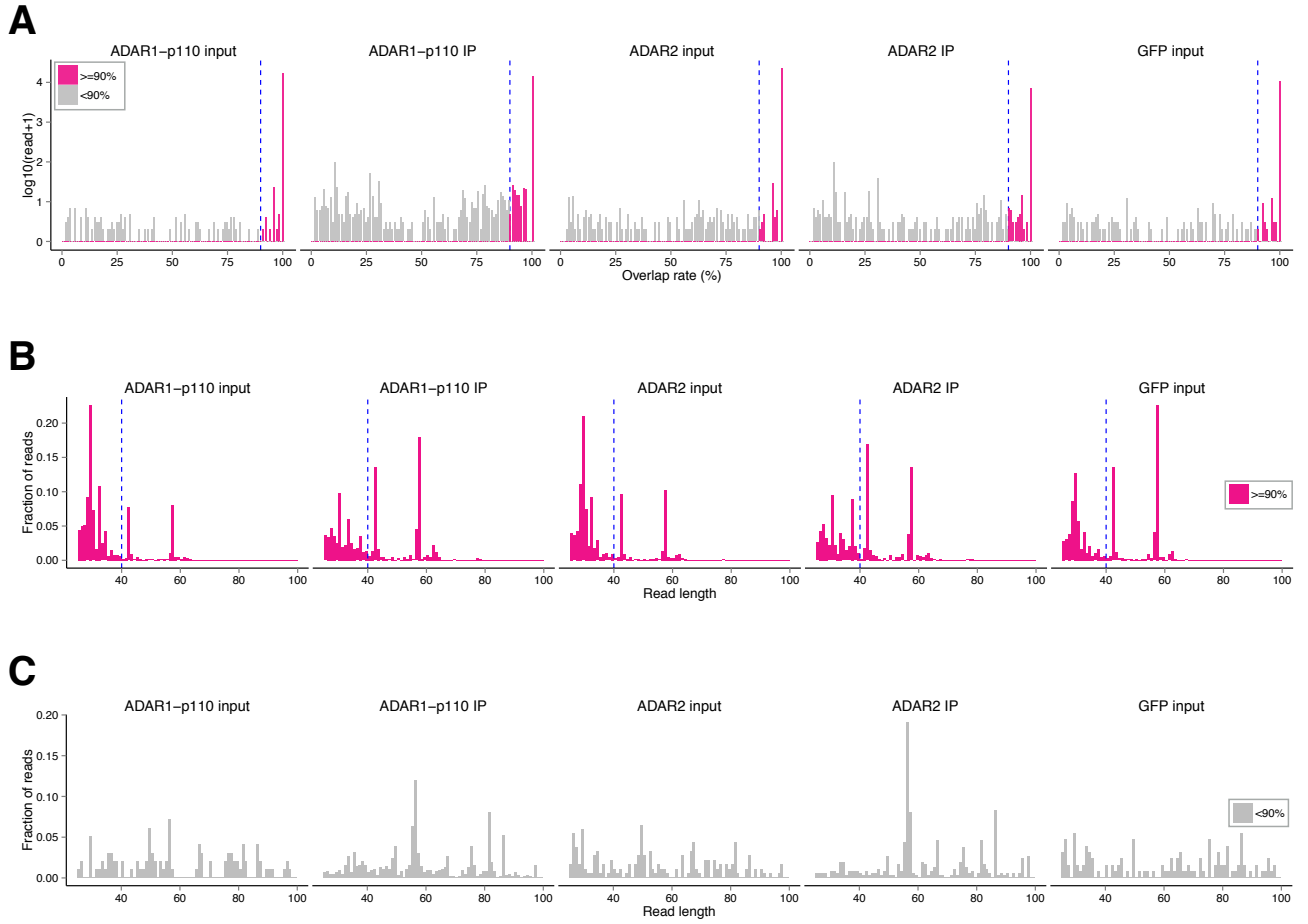


Figure S3. Length distribution of longer reads (≥ 25 nt) mapped to miRNA regions and their overlap with pre-miRNA annotation. (A) Percentage of overlap between each longer read (25 nt) and its pre-miRNA annotation in ADAR1-p110 input and IP, and ADAR2 input and IP samples, respectively. The horizontal axis indicates the percentage of overlap with the corresponding pre-miRNA annotation, and the vertical axis indicates \log_{10} (raw read count+1). The dashed line indicates 90% overlap. The pink and gray lines indicate the reads overlapped with $\geq 90\%$ and $< 90\%$ with pre-miRNA annotations, respectively. The length distribution of the reads overlapped $\geq 90\%$ (B) and $< 90\%$ (C) with annotated miRNAs.

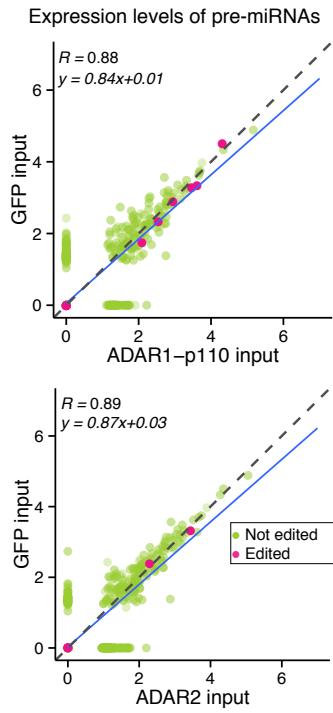
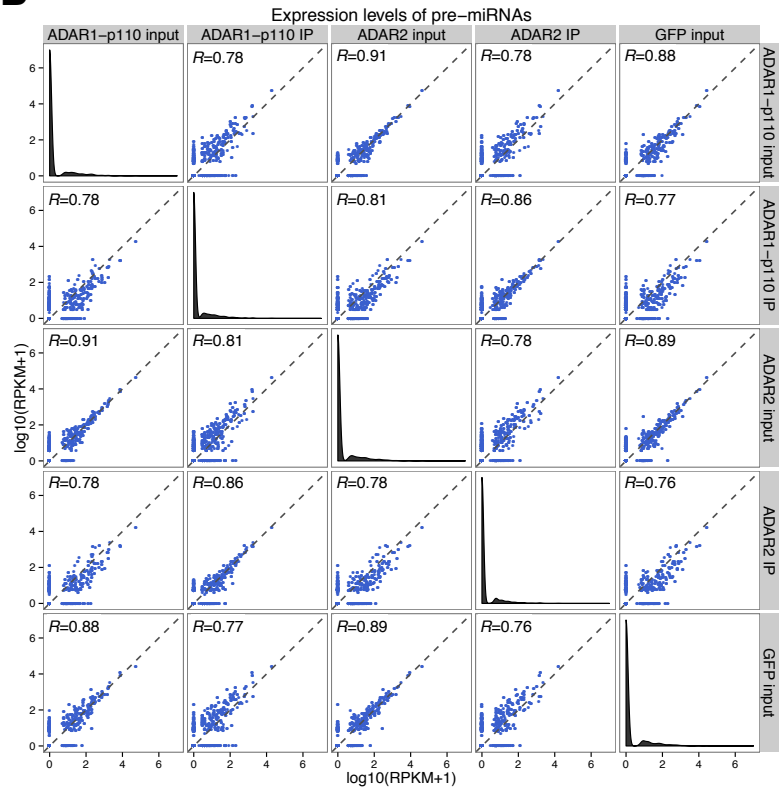
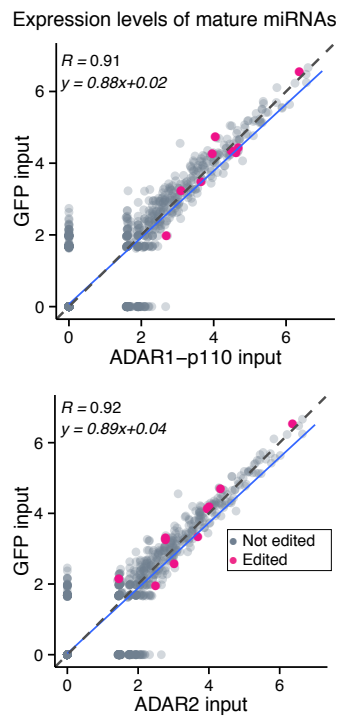
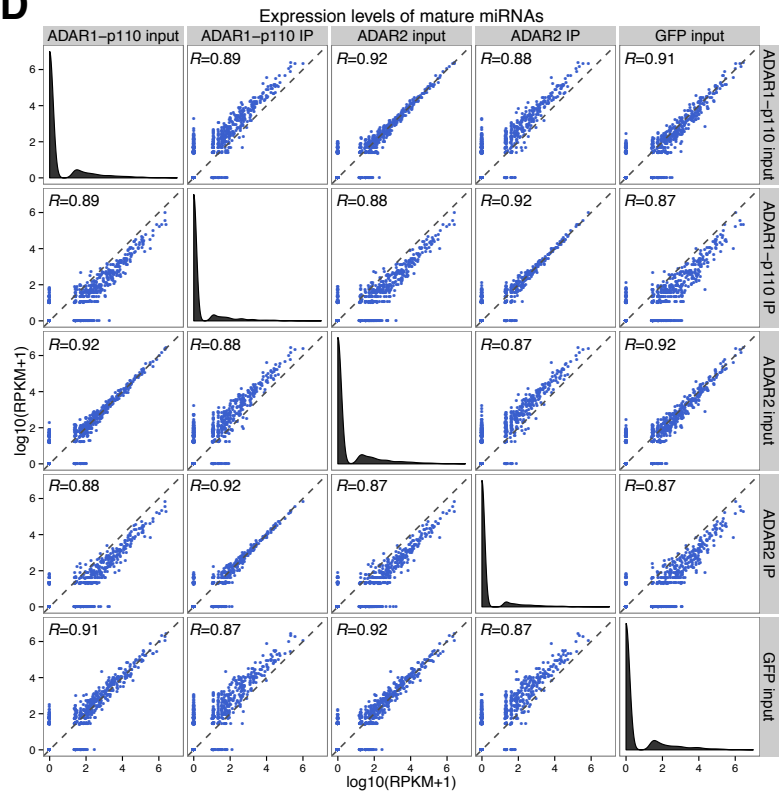
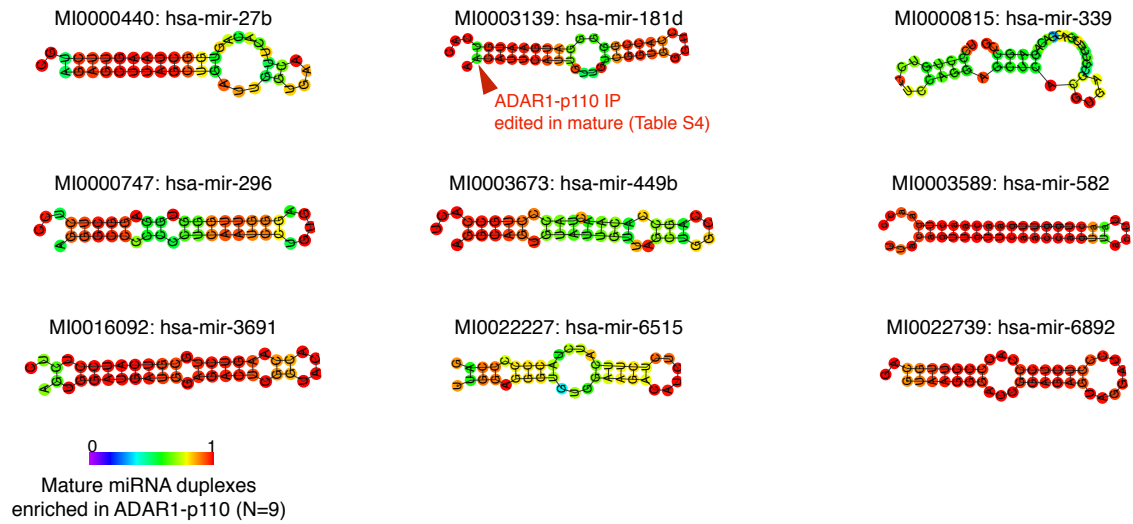
A**B****C****D**

Figure S4. miRNA expression levels and binding preferences of each ADAR isoform to miRNAs. Correlation between the mean expression levels of pre-miRNAs (A) or mature miRNAs (C) in ADAR1-p110 (upper) or ADAR2 (lower) input sample and those of GFP control input. Round-robin comparison of expression levels of pre-miRNAs (B) and mature miRNAs (D) across all samples sequenced. X-axis and Y-axis are $\log_{10}(\text{RPKM}+1)$. R indicates the Pearson correlation coefficient and y represents the regression line. The dashed gray line corresponds to $y=x$. Pink dots in (A, C): edited pre-miRNAs (A) and mature miRNAs (C). The histogram filled in black in (B, D) in the diagonally aligned panels shows the distribution of expression levels (RPKM) in each sample.

A



B

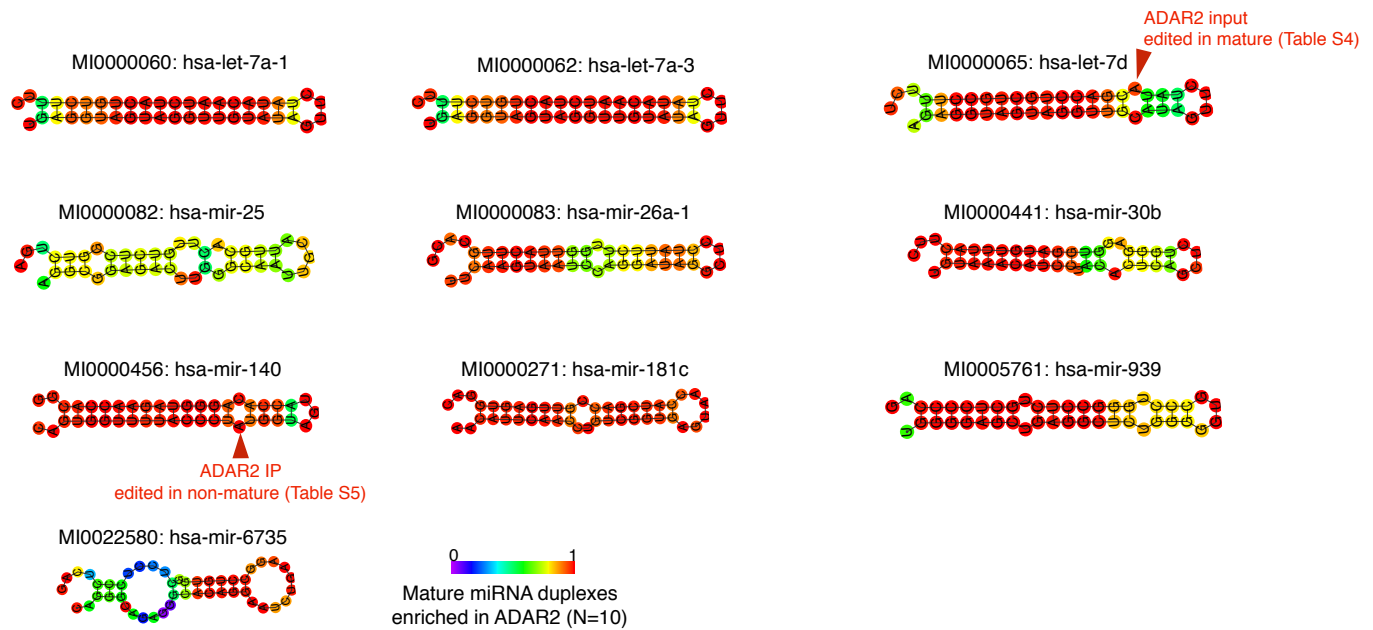


Figure S5. Predicted secondary structures of pre-miRNA that were enriched in the IP samples of ADAR1-p110 (A) versus ADAR2 (B) as shown in Figure 3C and D. Individual secondary structures were predicted by CentroidFold with default parameters. Gradient color corresponds to base-pairing and loop probability for each base.

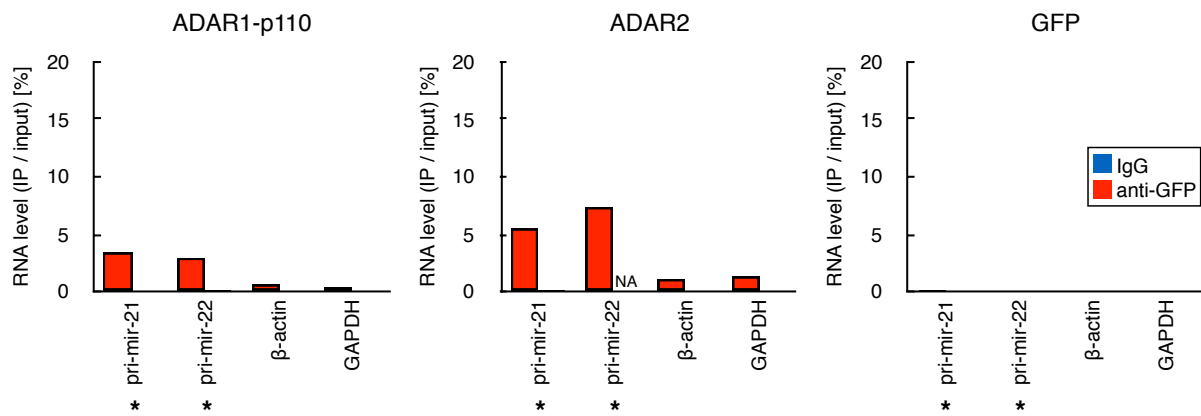
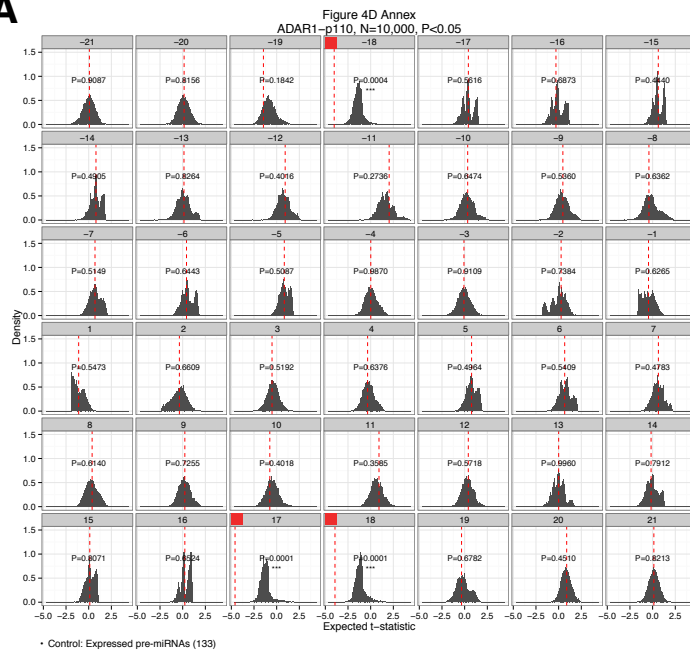
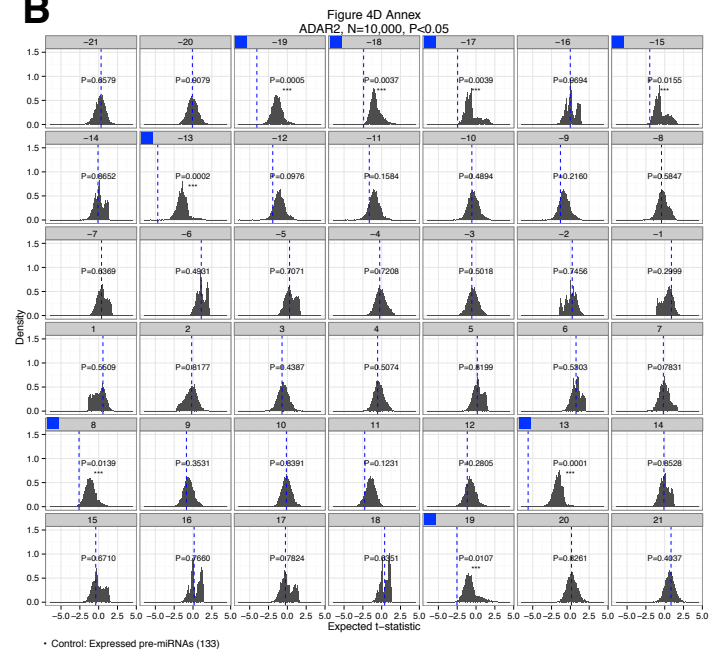
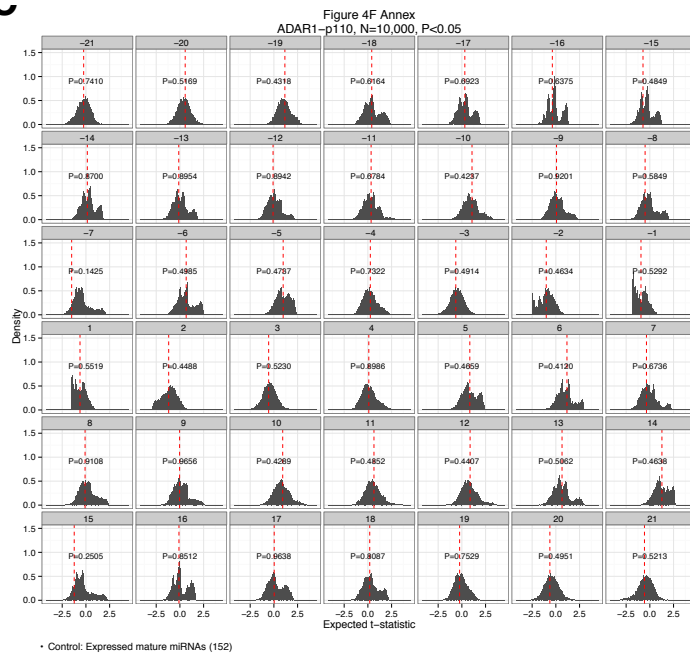
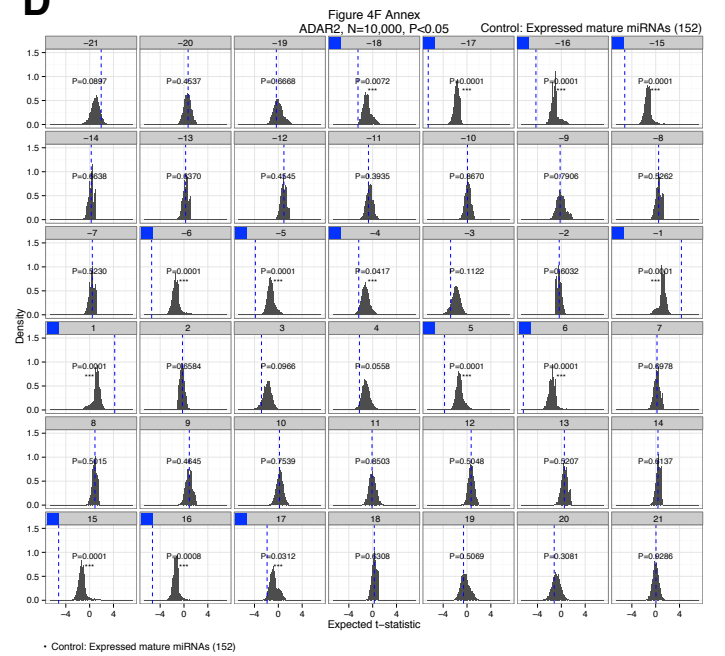


Figure S6. Results of real-time PCR for pri-miRNAs using purified total RNA from IP and input samples of cells overexpressing mGFP-ADAR1-p110 (left), mGFP-ADAR2 (center), and mGFP (right). The results for anti-GFP and control IgG IP samples antibody are shown in red and blue bars, respectively. The ratio of IP to input indicates relative RNA expression levels. Stars (pri-mir-21 and pri-mir-22) show endogenous pri-miRNAs. β-actin and GAPDH are used as controls.

A**B****C****D**

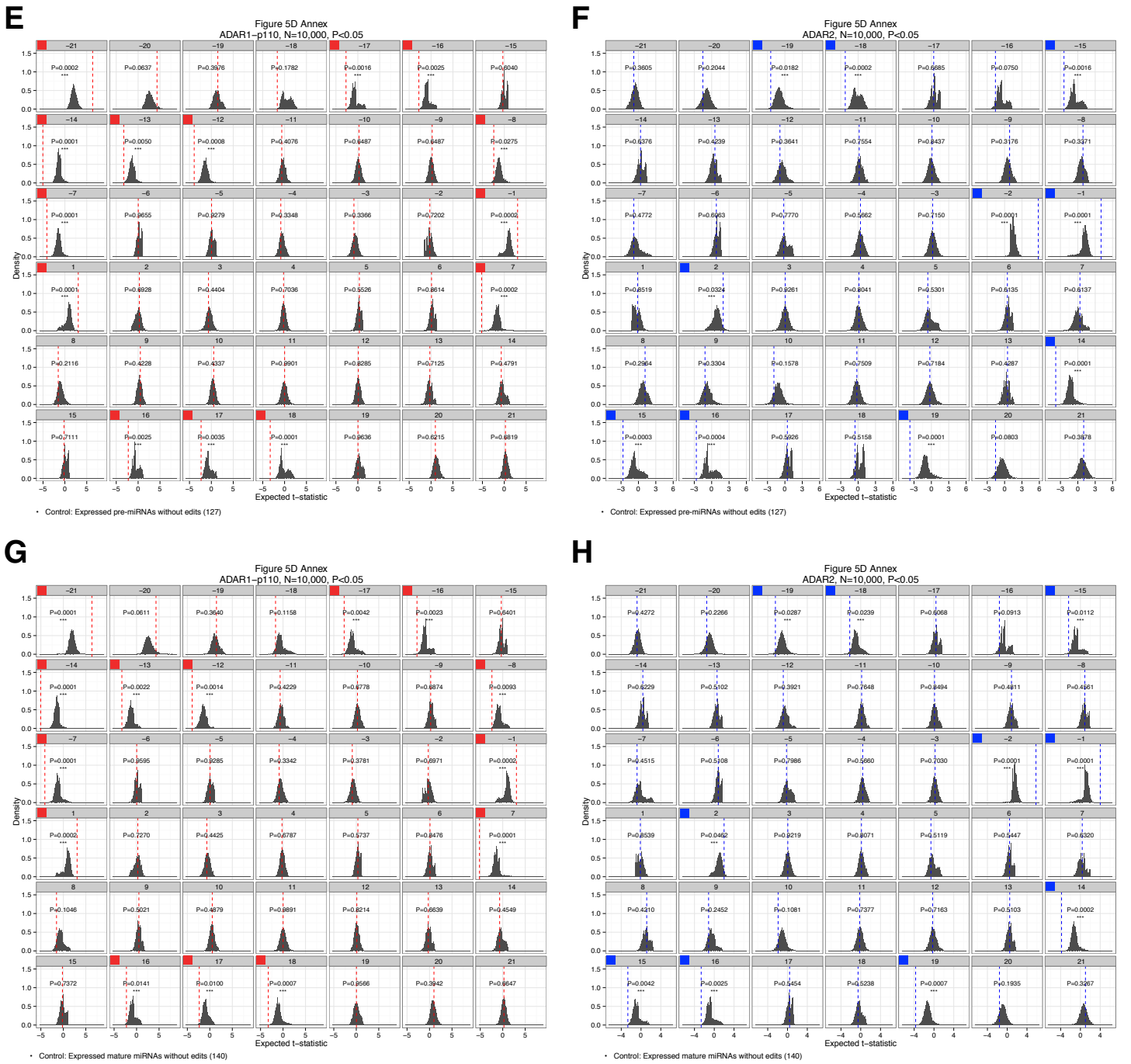


Figure S7. Bootstrap t-statistical test. Distribution of bootstrap t-statistic at each position on pre-miRNAs enriched in ADAR1-p110 IP (A) or ADAR2 IP (B) compared to all expressed pre-miRNAs, or candidate mature miRNA duplexes enriched in ADAR1-p110 IP (C) or ADAR2 IP (D) compared to all candidate mature miRNA duplexes expressed in both input samples, and their levels of significance for each position. Distribution of bootstrap t-statistic at each position in pre-miRNAs enriched in ADAR1-p110 IP (E,F) or ADAR2 IP (G,H) compared to the pre-miRNAs (E,G) or candidate mature miRNA duplexes expressed in both input samples (F,H), and their level of significance for each position. Vertical axis: density; horizontal axis: expected t-statistic. Red or blue dashed lines: observed t-statistic; red or blue squares: positions for which the difference was significant.

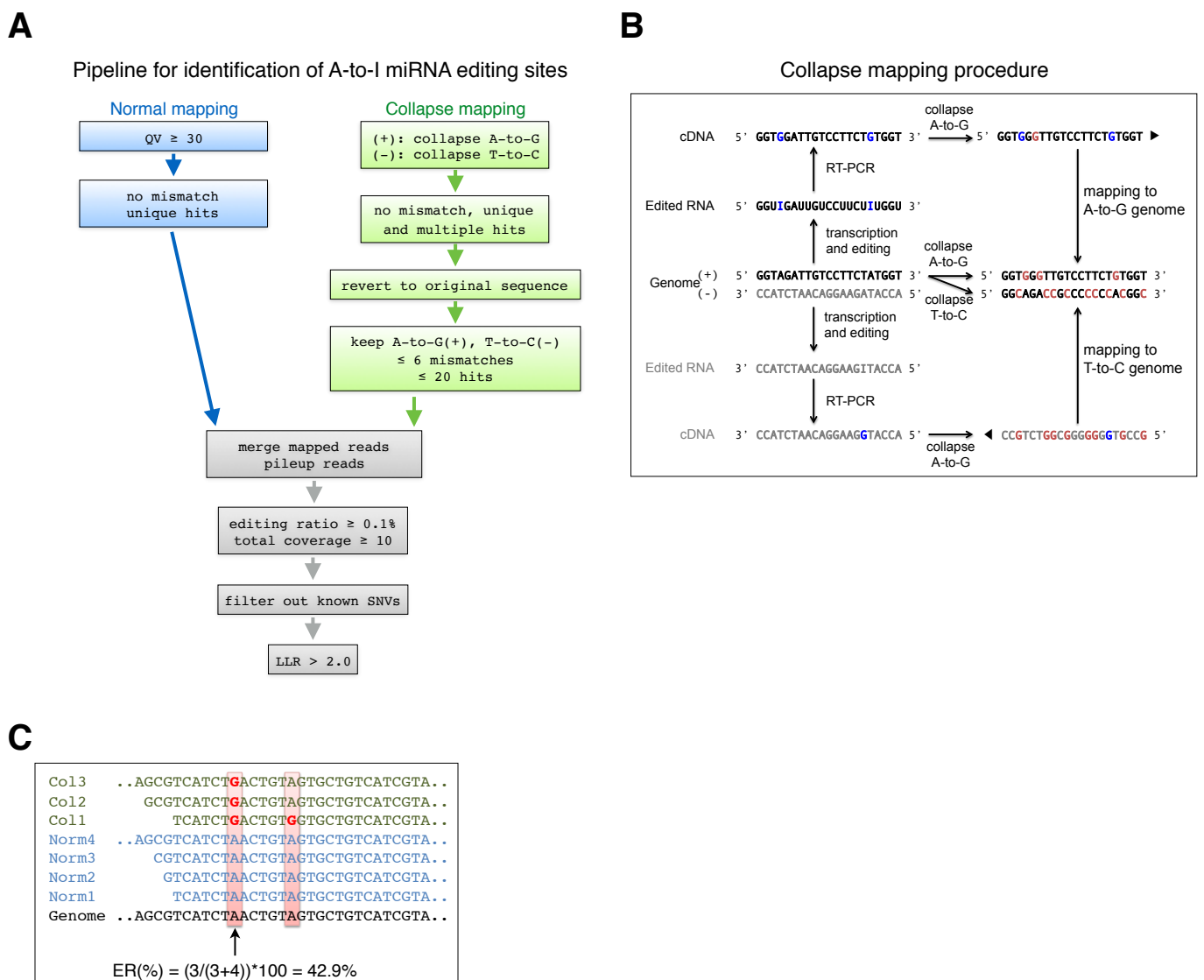


Figure S8. (A) Computational pipeline for identification of A-to-I miRNA editing sites by normal and collapse mapping procedures. Blue (normal mapping): reads which had at least one nucleotide with a quality value (QV) of less than 30 were discarded; mapping was performed with the output of “unique hits with no mismatches”. Green (collapse mapping): mapping was performed with the output of “multiple hits with no mismatches”, the remainder (no hits) and the reference genome were collapsed (A-to-G for forward reads and (+) strand genome; T-to-C for reverse reads and (-) strand genome); the collapsed reads were mapped again to output “unique and multiple hits with no mismatches”; collapsed sequences were reverted back to their original sequence to identify mismatches against the reference genome; reads with mismatches other than A-to-G for (+) reads and T-to-C for (-) reads were discarded; reads with more than 6 mismatches were discarded; collapsed reads which had mapped to more than 20 locations in the genome were discarded. Gray (identification of editing sites): merge “normal” and “collapse” uniquely mapped reads; output per-nucleotide coverage; calculate A/(A+G) editing ratio (ER) for each A-to-G site; sites with $ER \geq 0.1\%$ and with a coverage of 5 or more were validated as editing sites. (B) Procedure for A-to-G collapse mapping. After RT-PCR, the positions corresponding to an A-to-I editing site carry a “G”. All “A”s in (+) reads were collapsed to “G”s. All “A”s in (-) reads were collapsed to “G”s. All “A”s in the (+) strand of the genome were collapsed to “G”s. All “T”s on the (+) strand of the genome were collapsed to “C”s. After alignment of the collapsed (+) and (-) reads to the A-to-G and T-to-C collapsed genome sequences, respectively, the reads and the genome were reverted back to their original sequence and compared. See Materials and Methods section for further details. (C) Illustration of the visual output of computational analysis for normal and collapse reads aligned against a reference genome. Non-edited reads (blue) are obtained by normal mapping as described in (A), and potentially edited reads containing A-to-G mismatches (green) are selected by collapse mapping, as described in (A).

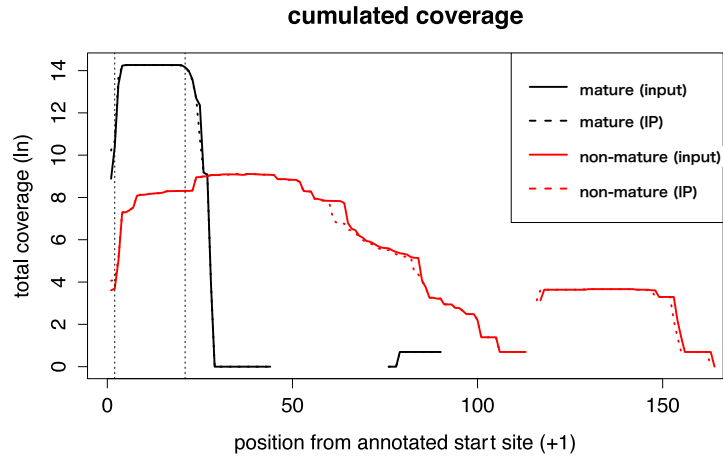


Figure S9. Cumulated coverage for all annotated mature (black) or non-mature miRNAs (red) given by the sum of all input (full lines) and IP (dotted lines) samples. X-axis: nucleotide position from the 5' end of miRNA or pre-miRNA annotation (starting from +1); y-axis: \log_2 scale of the per-nucleotide sum of coverages.

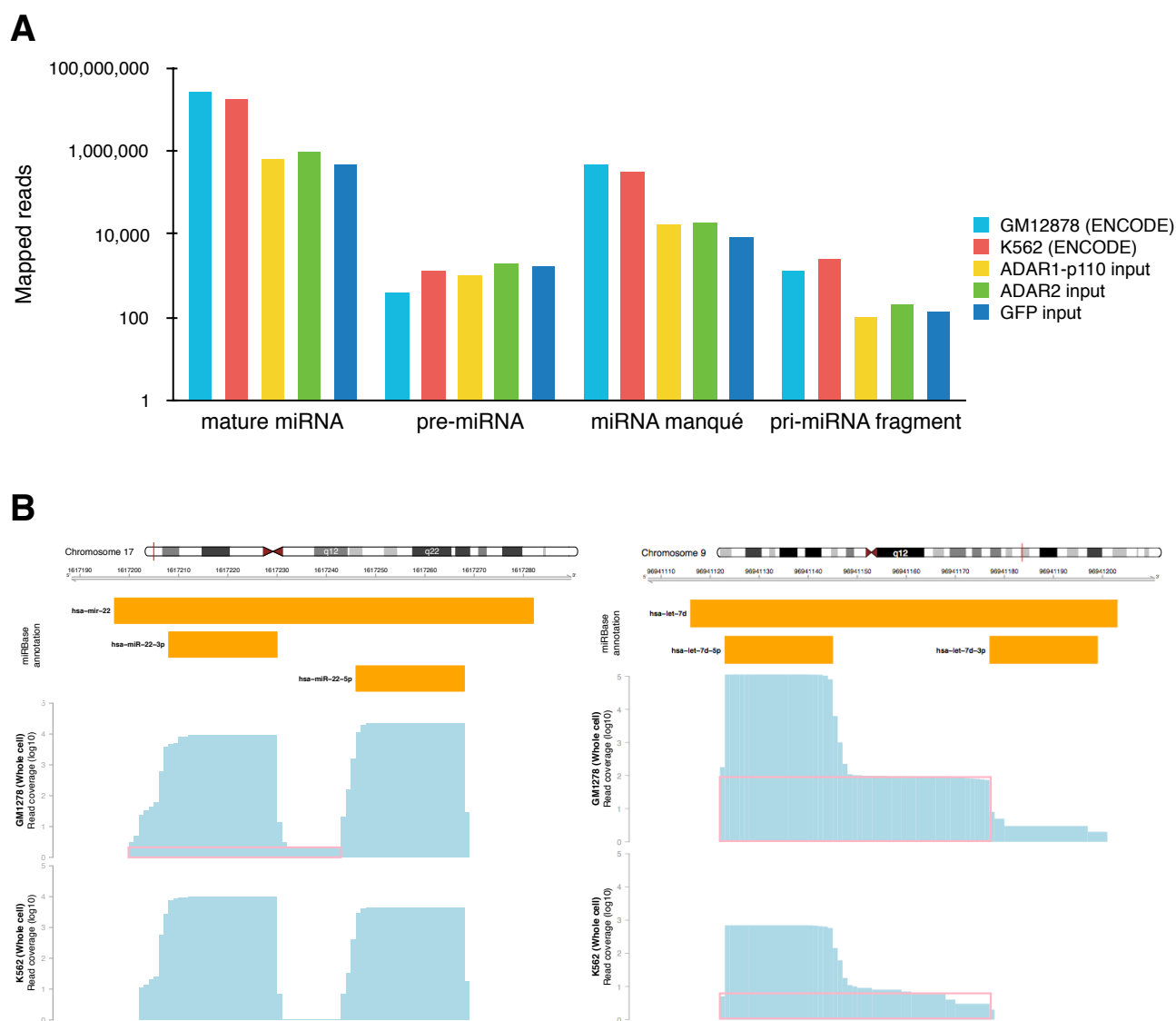


Figure S10. ENCODE small RNA-seq data analysis confirming the expression of miRNA manqué. (A) Total mapped reads of our ADAR sequencing data alongside ENCODE small RNA-seq data of GM12878 (light blue) and K562 cells (red). X-axis shows the total mapped reads for each type of miRNA species. (B) Example of miRNA manqué transcripts found in miR-22 and let-7d loci from different two cell lines, GM1278 and K562, were highlighted (light pink).

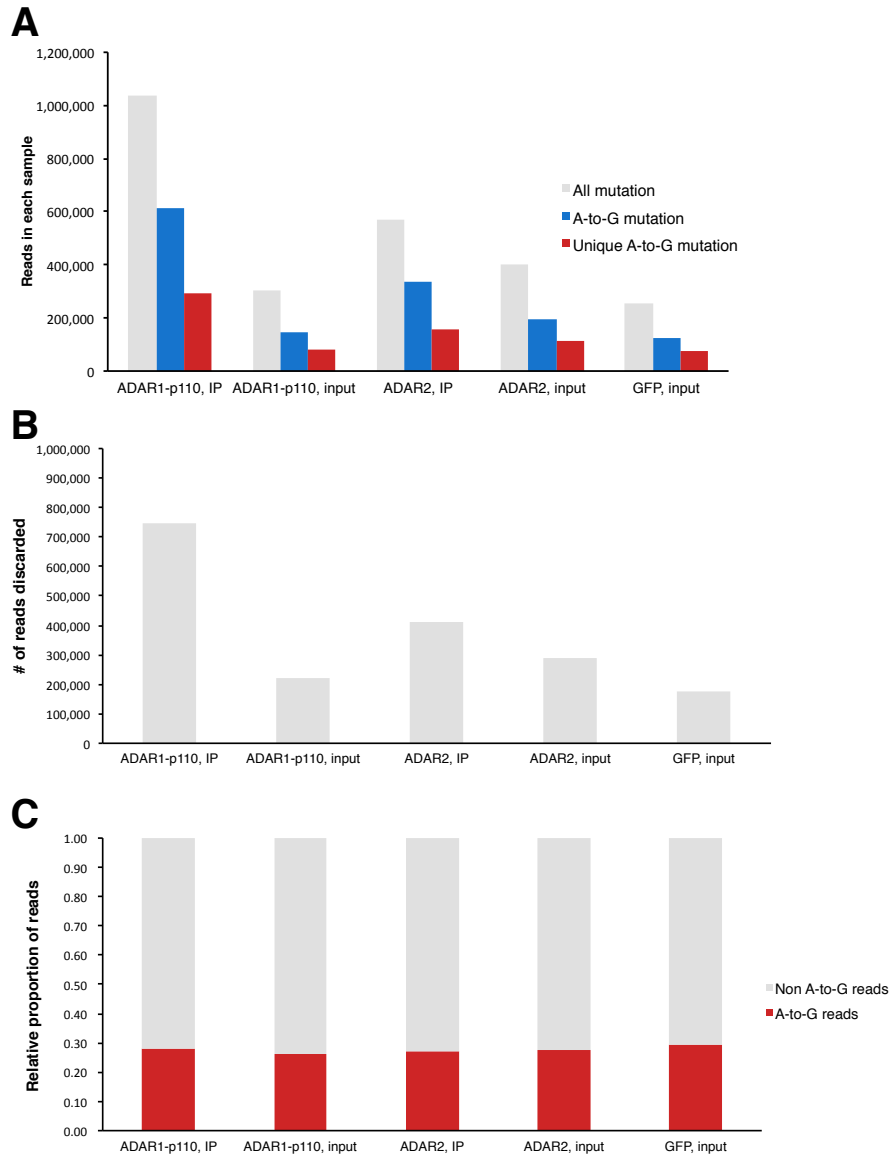


Figure S11. The numbers and percentages of A-to-G mismatched reads among all types of mismatches. (A) The number of reads of all types of mismatches containing A-to-G, A-to-C, A-to-U, G-to-C, G-to-U, G-to-A, C-to-U, C-to-A, C-to-G, U-to-A, U-to-G, and U-to-C in ADAR1-p110 IP and input, and ADAR2 IP and input, and GFP input samples, respectively. Gray bar indicates the number of reads with all types of mismatches. Blue bar, the number of reads with A-to-G mismatches in single reads. Red bar, the number of reads with unique A-to-G mismatches in paired reads. The numbers of reads that have non A-to-G mismatches in each library, which are the subtracted numbers of blue bar from gray bar, are ADAR1-p110 IP: 427,316 reads, ADAR1-p110 input: 155,175 reads, ADAR2 IP: 232,075 reads, ADAR2 input: 207,307 reads, GFP input control: 125,611 reads, respectively. (B) The numbers of discarded reads which have non A-to-G mismatches in each sample. (C) The percentages of discarded reads in each sample.