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## **Supplemental Methods**

### **Additional Sample Details**

Our sample for this report is composed of 87 subjects with bipolar disorder (“BP”) originally collected as part of the Chicago, Hopkins, National Institute of Mental Health Intermural Program (“CHIP”) BP study and 931 BP subjects originally collected as part of the NIMH Genetics Initiative Bipolar Disorder Collaborative study [1-3]. There are 119 male and 268 female attempter subjects, and 291 male and 340 female non-attempter subjects present in the study population.

### **Target Library Creation**

NimbleGen SeqCap EZ (NimbleGen, Madison WI) paired-end libraries were prepared for each subject following standard protocols. Briefly, 1-3µg fragmented whole genomic DNA was prepared for each subject using a Covaris S2 (Covaris, Woburn MA) with a 10% duty cycle, an intensity setting of 5, and 200 cycles per burst for 90s at 4° C. Each fragmented sample was then end-repaired and ligated with either paired-end barcoded DNA adapters from Integrated DNA Technologies (IDT, Coralville IA) or with NEXTflex DNA barcode adapters from Bioo Scientific (Austin TX). Barcoded samples were then filtered for size via a 2% agarose gel in 1X TAE buffer and either 0.5 µg/mL ethidium bromide or 0.01% SYBR Gold (Life Technologies, Grand Island NY). ~300bp DNA fragments were sliced and extracted from the gel. Libraries were then amplified via 8 cycles of PCR using Phusion High-Fidelity PCR Master Mix (New England Biolabs, Ipswich MA). Annealing temperatures for the PCR were set at 65° C for samples prepared with full-length paired-end primers or at 60° C for partial-length primers.

Following preparation, a 400bp unimodal sample peak was confirmed via a Bioanalyzer DNA 1000 kit (Agilent Technologies, Santa Clara CA). Samples were quantified via a NanoDrop 1000 Spectrophotometer (Thermo Scientific, Waltham MA) with the requirement that at least 1µg of

sample to be present for non-multiplexed capture or 500/250/166ng DNA per subject in multiplexed 2/4/6 subject capture experiments, respectively.

### **Target Design and Capture**

Our 1,018 BP subjects were each hybridized to one of three different arrays. The SeqCap EZ Exome v1 (NimbleGen, Madison WI) array was used to capture the first 30 attempter and 57 non-attempter BP subject samples. The version 2 SeqCap EZ array was used to capture an additional 15 attempter and 56 non-attempter BP subject samples. For the third and fourth years of our sequencing experiments, version 2 SeqCap EZ (NimbleGen, Madison WI) was used with the addition of custom targets designed to capture regulatory regions (1kb promoters/full UTR regions) for 1,422 post-synaptic density genes [4] and 57 genes that have been previously associated with BP disorder. The customized version 2 SeqCap EZ array was used to capture the remaining 342 attempter and 518 non-attempter BP subjects. Overall target coverage of the SeqCap EZ Exome (NimbleGen, Madison WI) kits was approximately 26.2 Mb (~16,000 transcripts), 36.0 Mb (~30,000 transcripts), and 54.3 Mb (~30,000 transcripts) for SeqCap EZ version 1, 2, and customized 2, respectively.

All hybridizations were performed with either 1µg of the DNA library from a single sample (initial non-multiplexed experiments) or with 1µg total pooled DNA libraries equally distributed by mass among the pooled samples followed by post-hybridization following the instructions in the SeqCap EZ Exome guide (NimbleGen, Madison WI). Briefly, the post hybridization products were washed and enriched in duplicate 18 cycle PCR reactions. Duplicate PCR products were then pooled together, washed, and quality tested with Bioanalyzer DNA 1000 kits (DNA quantity and fragment size).

### **Sequencing and Initial Quality Control**

Two different sequencing platforms were used in this project: samples prepared with SeqCap EZ Exome version 1 (NimbleGen, Madison WI) were sequenced at one sample per lane using a GA-IIx sequencer (Illumina, San Diego CA), while samples prepared with the version 2 and version 2+custom capture targets were sequenced using HiSeq 2000 sequencers (Illumina, San Diego CA) using 2, 4, or 6 pooled samples. All samples were sequenced using paired-end 76-cycle runs. Only samples assessed to have  $\geq 20X$  coverage depth in  $\geq 70\%$  of the capture targets were included within the final dataset. It was noted that samples failing to meet the 20X/70% threshold that were sequenced with the Illumina GA-IIx could often be pooled together and reanalyzed to gain greater depth of coverage (thus allowing re-inclusion of these samples in downstream analysis). Samples failing the 20X/70% metric when sequenced with the increased data collection capacity of the HiSeq 2000 generally did not see improvement if pooling/re-sequencing attempts were applied. An elevated level of detected PCR duplicates and/or diminished 2X sequencing depth coverage of targets detected in these samples was strongly suggestive of low library complexity, and such samples were either excluded from the study or re-sequenced from recreated libraries.

### **Sample Alignment, Calling, and Quality Control Pipeline**

A semi-automated sample processing pipeline was constructed to prepare all samples for analysis. Individual samples were separated by barcode through the CASAVA version 1.8 software package (Illumina, San Diego CA). Sample FASTQ output files were individually aligned against the hg19/GRCh37 build via the Burrows Wheeler Aligner [5] version 0.6.2 aln module with a seed length of 30. Resultant SAM files were converted to BAM files and processed via SAMtools [6] version 0.1.18. BAMtools [7] version 2.1.1 was used to remove unpaired reads and Picard version 1.88 (<http://broadinstitute.github.io/picard>) was used to remove duplicate reads from the BAM files. Reads with low mapping quality were then removed

by BAMtools [7] followed by indexing via SAMtools [6]. The genome analysis toolkit (GATK) [8] version 2.6-5 was then used to realign sequences around suspected and known DNA insertion and deletion (indel) sites and to perform base recalibration of predicted variant sites based on dbSNP version 138 and 1000 Genomes known sites.

Following the initial data preparation, the GATK Reducereads tool generated reduced BAM files. The reduced files were then simultaneously genotyped as a single group (all study subjects genotyped at once) for SNPs and indels using the GATK unified genotyper package. The GATK was used for group variant recalibration around called SNPs and indels. Initial flagging and removal of low-quality SNPs was performed based on the combined examination of quality-depth (QD), mapping quality rank sum (MQRankSum), read position rank sum (ReadPosRankSum), fisher based strand bias (FS), mapping quality (MQ), and haplotype score (HaplotypeScore) based assessments during the recalibration step. In addition, all detected indels and tri/quad allelic sites were removed from the final analyzed dataset due to the technological limitations involved with accurately calling these complex alleles.

### **Variant-Level Quality Control Measures**

Several additional levels of quality control were employed to remove questionable variant calls from the dataset. All subject genotype calls that had a call depth of less than 10 or a genotype quality ("GQ") score of 20 or less were removed from the dataset by setting them to missing in the variant call file ("VCF"). All variants that were flagged during GATK [8] variant recalibration checks for any reason were removed. Variants failing a Hardy-Weinberg check via a p-value of  $< 1 \times 10^{-6}$  were also removed. Finally, any variants missing calls in  $\geq 10\%$  of all subjects post call QC were removed from the dataset.

Systematic bias in variant calling between study groups (attempters and non-attempters) was assessed by examining the average number of variants that arose in a single subject within the

study population (“singletons”). On average, 227.7 singletons were observed per attempter and 224.0 singletons per non-attempter. Performing a t-test on these distributions led to a p-value of 0.29 when assuming equal variance and a p-value of 0.29 if assuming unequal variance. These results demonstrate no evidence of significant calling bias between the groups.

Finally, assessments of the final quality-checked dataset were made via two methods. First, the distribution of individual variant test results was plotted against the expected results for a sample of this size as a QQ-plot (Fig S2). This QQ assessment showed no broad deviations from the expected p-values, suggesting little or no underlying population structure contribution to our results. Second, exome genotypes were compared against overlapping available GWAS genotypes [9, 10] for all subjects in the study. 10,251 variants were covered in the exome that were also genotyped within the GWAS study. A call mismatch rate of 0.33% (16,379,480 matched calls, 54,170 mismatched calls) was noted in the comparison of these studies, suggesting very high quality calls within the prepared exome data.

### **Subject Quality Assessment**

Subjects were assessed for population stratification using principal component analysis via the software packages EIGENSTRAT and SMARTPCA within the EIGENSOFT package [11]. Only variants with a minor allele frequency (“MAF”)  $>0.05$  which were successfully typed in  $\geq 95\%$  of the study sample and were not in linkage disequilibrium (LD) were used for this analysis. The first two principal components for the subjects are represented within Fig S1 and demonstrate an equal distribution of each subject population with no other discernible outliers.

In addition, subjects were assessed for correct sex via the PLINK [12] v1.07 “--check-sex” method. This method assesses the ratio of X chromosome homozygote SNP calls to all X chromosome calls against expected values for males and females against the labeled sex of each subject in the data. We used a set of X chromosome variants with  $MAF > 0.05$  and which

were successfully called in  $\geq 95\%$  of our subject population to perform this check and flag any mislabeled subjects for removal from the final dataset.

### Statistical Analyses Details

All variants detected in the dataset, regardless of allele frequency, were assessed for association with suicidal behavior via the logistic regression package “logistf” [13] which included the correction for the covariates of sex, assay/platform, and the first five components of the principal component analysis for each subject (for a single variant result overview see Table S6). Functional variants were assessed using two minor allele frequency (“MAF”) thresholds for all gene and pathway tests: functional variants with  $MAF \leq 0.05$  (“MAF05”) and functional variants with  $MAF \leq 0.01$  (“MAF01”). The MAF of any given variant was determined by examining the frequency within our complete dataset in addition to the listed frequency in the European 1000 genomes October 2014 release dataset [14], the non-Finnish European Exome Aggregation Consortium version 0.3 (Exome Aggregation Consortium (“ExAC”), Cambridge, MA (URL: <http://exac.broadinstitute.org>) (accessed March, 2015)), and/or the European NHLBI GO Exome Sequencing Project (<http://evs.gs.washington.edu/EVS/>; version SIV2) subjects, where available.

Variants were also selected for inclusion in gene- and pathway-level analyses based on a variety of functional annotations. Variants were annotated for coding regions via ANNOVAR [15] (version 2015Mar22). Coding functional variants were classified into one of two groups: disruptive coding and broad coding functional variants [16]. The disruptive coding classification refers to any identified stopgain/nonsense, frameshift, or essential splice site mutations. The broad coding classification encompasses the complete set of disruptive variants as well as any non-synonymous variants predicted to be functional by at least one of the following packages: SIFT [17], polyphen2 (HVAR and HDIV) [18], LRT [19], Mutation Taster [20], and VEST [21].

Variants to be included within the regulatory gene- and pathway-level analyses were selected based on functional classification within the “regulome” database (“regDB”) [22]. Variant selection was based on two described score thresholds that represent variants “likely to affect the binding of regulatory elements” (regDB scores of 1 and 2; “regulatory narrow”) and those that are “less likely to affect binding” due to less overlapping evidence (regDB scores of 3-6) [22]. For consistency, we refer to these groups as regulatory narrow and regulatory broad. The regulatory broad classification includes all regulatory narrow variants (regDB scores 1-6).

Gene-level tests were performed using all variants within the four specified classifications based on gene boundaries as defined by the ANNOVAR annotation using Ensembl transcripts (Ensembl version 75, Feb 2014) [23] on the hg19/GRCh37 assembly. The genes were then assessed via two methods: gene burden tests and sequence kernel association tests (SKAT). Gene burden tests were accomplished by collapsing variant signals using the CMC variant collapsing method over subjects as has been described elsewhere [24]. Briefly, each gene locus was assessed for the presence or absence of functional variants. Any subject with one or more functional variants within the given gene locus was classified as a “1” for the burden test of that gene, whereas subjects with no functional variants in the given gene locus were classified as a “0” for the burden test of the gene. The collapsed values were then assessed along with all covariates using Firth’s corrected penalized logistic regression [25] via the R package “logistf” [13] version 1.21 for all genes. SKAT tests utilized the SKAT R package, version 1.1.2, using the “davies” method [26] for all autosomal genes. The SKAT examines each variant site within a multivariate model to allow individual sites to have varying directional effects on the test result [26]. All gene-level analyses included the covariates of sex, the first five components from our sample PCA analysis, and the sample processing platform/array group.

Pathway-level analyses were performed on a primary selection of 33 existing pathways defined in the literature and online resources. These pathways were selected based on suspected



importance within suicidal behavior risk and/or due to general importance to brain structure and function (Table S4). In addition, a secondary set of pathway analyses were performed on 3,621 pathways selected primarily from the MSigDB v5.1 [27]. These pathways were selected to represent well-described biological pathways (collections H and C2), shared motifs (collection C3), and gene ontology pathways (collection C5).

The complete pathways (including sex chromosome genes) were assessed for direct genetic association with suicidal behavior via the PLINK/SEQ v0.10 package “SMP” method. This method utilized all dataset genes assessed using the PLINK/SEQ “burden” test with 1 million permutations and correcting for platform/assay via the `–strata` PLINK/SEQ command. The SMP method includes a correction for the background variation distribution across all genes for a given analysis. Top results were also corrected for past diagnoses of alcohol dependence within all subjects in a secondary analysis.

Finally, the SMP analyses were corrected for multiple testing via a 500 attempter/non-attempter label swapping permutations. This involved repeating all SMP analyses across each functional variant set and MAF threshold 500 times with randomly swapped attempter/non-attempter labels for each subject and comparing these randomized results against the primary SMP analysis results, generating an empirical p-value for each result. Note that only 500 permutations were run due to the computational burden of performing these analyses.

### **Power Calculations**

Individual and gene-based power calculations were performed using Quanto version 1.2.4 [28]. Program settings were as follows: a mismatched case/control ratio of 1:1.6 with 387 cases, “gene only” hypothesis, log-additive mode of inheritance with allele frequencies of 0.01 to 0.05 by 0.01 steps for single variants and allele frequencies of 0.02 to 0.1 by 0.02 steps for gene based tests were used. P0 and kP were placed at 0.046 (4.6% estimated population disease

prevalence [29]) with RG being plotted from values of 1-5 for genes and 1-8 for single variants. Significance levels were estimated at the liberal  $2.5 \times 10^{-6}$  and standard  $5.0 \times 10^{-8}$  for genes and single variants, respectively. Plots of the resulting power calculations are available in Figs S3 and S4 for single variant and gene tests, respectively.

### **Sample Set Acknowledgements**

Data and biomaterials for the CHIP sample were collected at Johns Hopkins, NIMH intramural program, and University of Chicago from 1988 to 2010, under the direction of Ray DePaulo, along with James Potash, Francis McMahon, and Elliot Gershon. This work was supported by NIH grants R01 MH-042243 (Drs. DePaulo and Potash) and R01 MH-061613 (Dr. Gershon); by the National Alliance for Research on Schizophrenia and Depression; by the Stanley Medical Research Institute; and by the NIMH Intramural Research Program (Dr. McMahon).

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## Supplemental Materials and Methods

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Data and biomaterials were collected in four projects that participated in the National Institute of Mental Health (NIMH) Bipolar Disorder Genetics Initiative. From 1991-98, the Principal Investigators and Co-Investigators were: Indiana University, Indianapolis, IN, U01 MH46282, John Nurnberger, M.D., Ph.D., Marvin Miller, M.D., and Elizabeth Bowman, M.D.; Washington University, St. Louis, MO, U01 MH46280, Theodore Reich, M.D., Allison Goate, Ph.D., and John Rice, Ph.D.; Johns Hopkins University, Baltimore, MD U01 MH46274, J. Raymond DePaulo, Jr., M.D., Sylvia Simpson, M.D., MPH, and Colin Stine, Ph.D.; NIMH Intramural Research Program, Clinical Neurogenetics Branch, Bethesda, MD, Elliot Gershon, M.D., Diane Kazuba, B.A., and Elizabeth Maxwell, M.S.W.

Data and biomaterials were collected as part of ten projects that participated in the National Institute of Mental Health (NIMH) Bipolar Disorder Genetics Initiative. From 1999-03, the Principal Investigators and Co-Investigators were: Indiana University, Indianapolis, IN, R01 MH59545, John Nurnberger, M.D., Ph.D., Marvin J. Miller, M.D., Elizabeth S. Bowman, M.D., N. Leela Rau, M.D., P. Ryan Moe, M.D., Nalini Samavedy, M.D., Rif El-Mallakh, M.D. (at University of Louisville), Hussein Manji, M.D. (at Wayne State University), Debra A. Glitz, M.D. (at Wayne State University), Eric T. Meyer, M.S., Carrie Smiley, R.N., Tatiana Foroud, Ph.D., Leah Flury, M.S., Danielle M. Dick, Ph.D., Howard Edenberg, Ph.D.; Washington University, St. Louis, MO, R01 MH059534, John Rice, Ph.D., Theodore Reich, M.D., Allison Goate, Ph.D., Laura Bierut, M.D. ; Johns Hopkins University, Baltimore, MD, R01 MH59533, Melvin McInnis M.D. , J. Raymond DePaulo, Jr., M.D., Dean F. MacKinnon, M.D., Francis M. Mondimore, M.D., James

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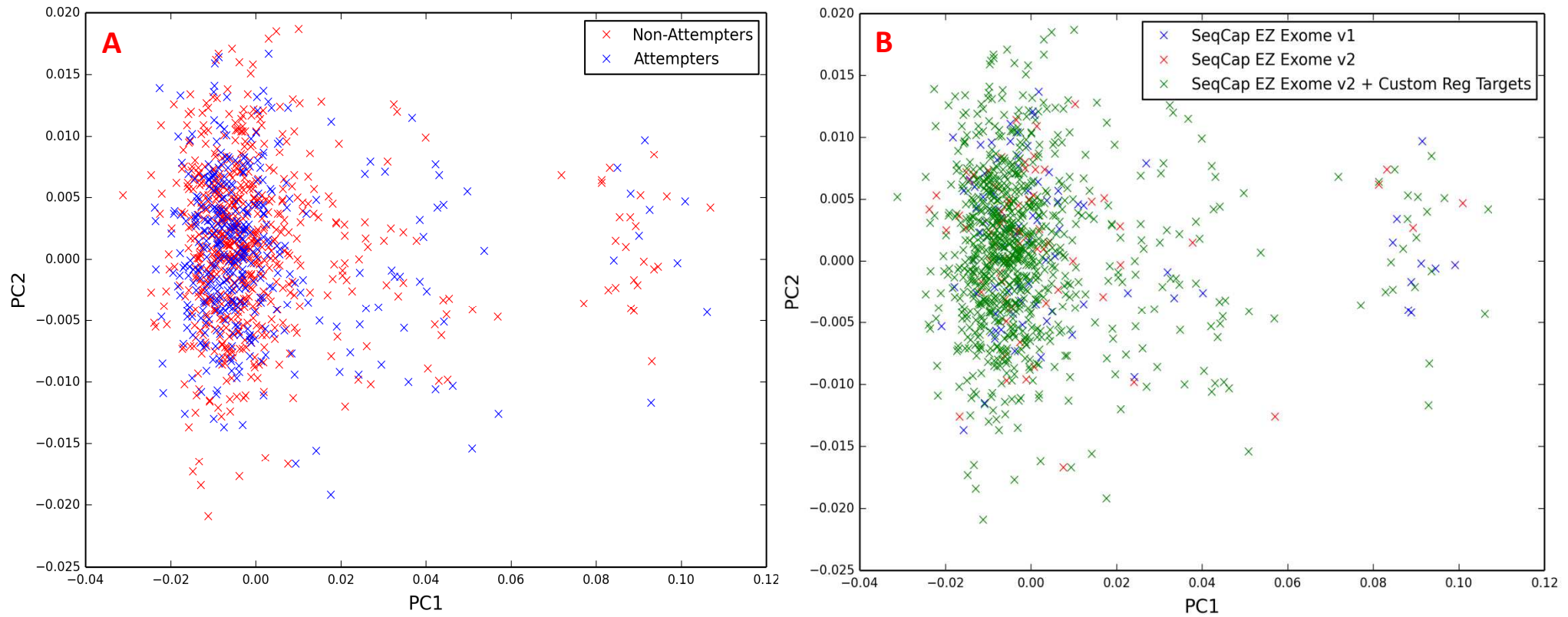
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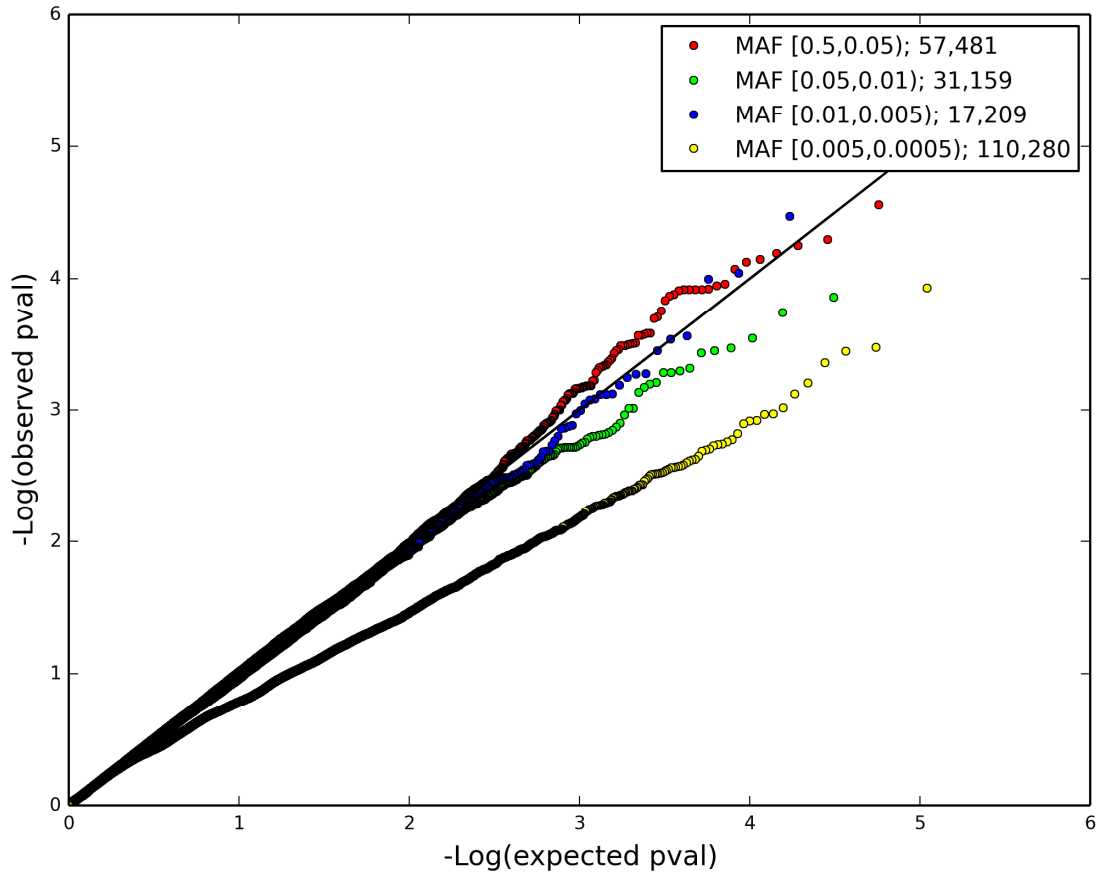
# **Supplemental Figures**

Fig S1



**Fig S1:** Principal component analysis plots of the first two components for each subject. Panel A presents the subjects labeled by suicide attempt status where red = BP subjects with no history of suicide attempt (631 subjects); dark blue = BP subjects with a history of suicide attempt (387 subjects). Panel B labels subjects by the sequencing array they were genotyped with where blue = BP subjects processed in the first year of sequencing using the Illumina SeqCap EZ Exome Array v1 (87 subjects), red = BP subjects processed in the second year of sequencing using the SeqCap EZ Exome v2 (71 subjects), and green = BP subjects processed in the third and fourth years of sequencing using the SeqCap EZ Exome v2 + custom regulatory sequence targets (860 subjects).

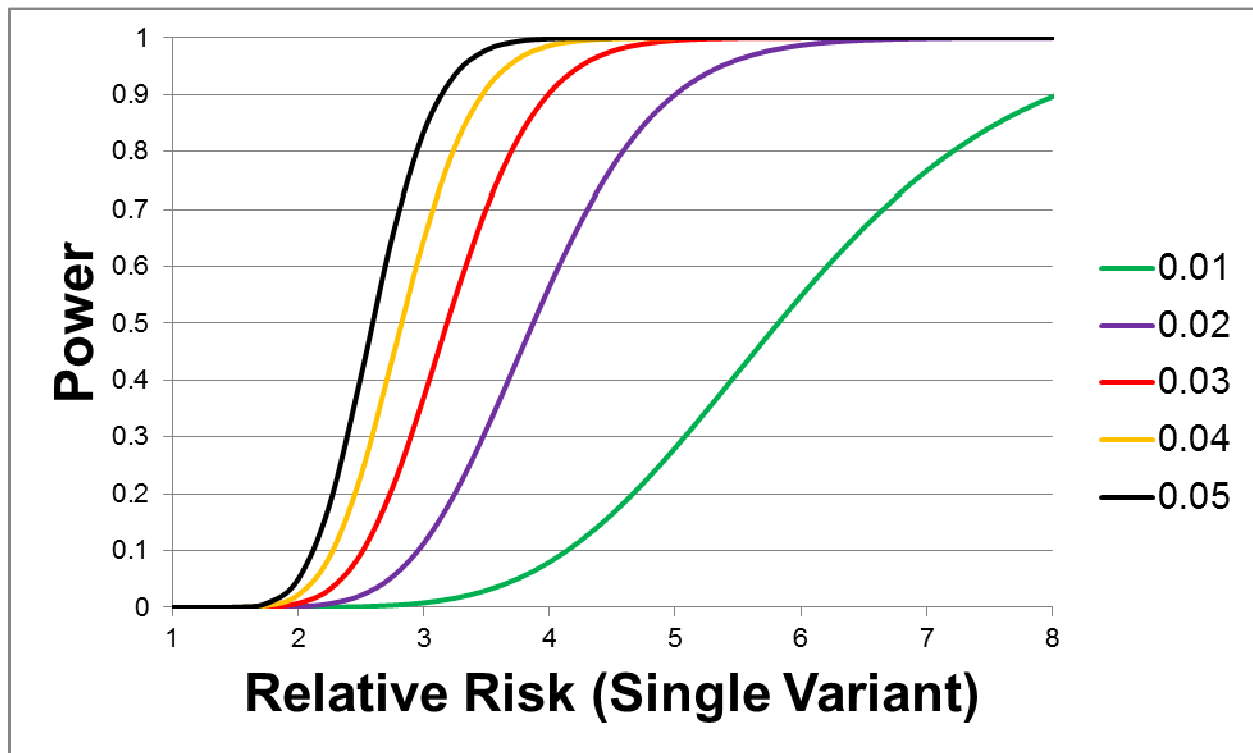
Fig S2



**Fig S2:** Individual variant QQ-plot for all subjects with a history of suicide attempt (387 subjects) versus all non-attempter subjects (631 subjects). The solid line represents the expected p-value distribution and the colored points represent observed single variant data at different minor allele frequency (MAF) ranges, as labeled. The value at the right of each MAF range is the number of unique sites that were identified within the dataset in the MAF range.

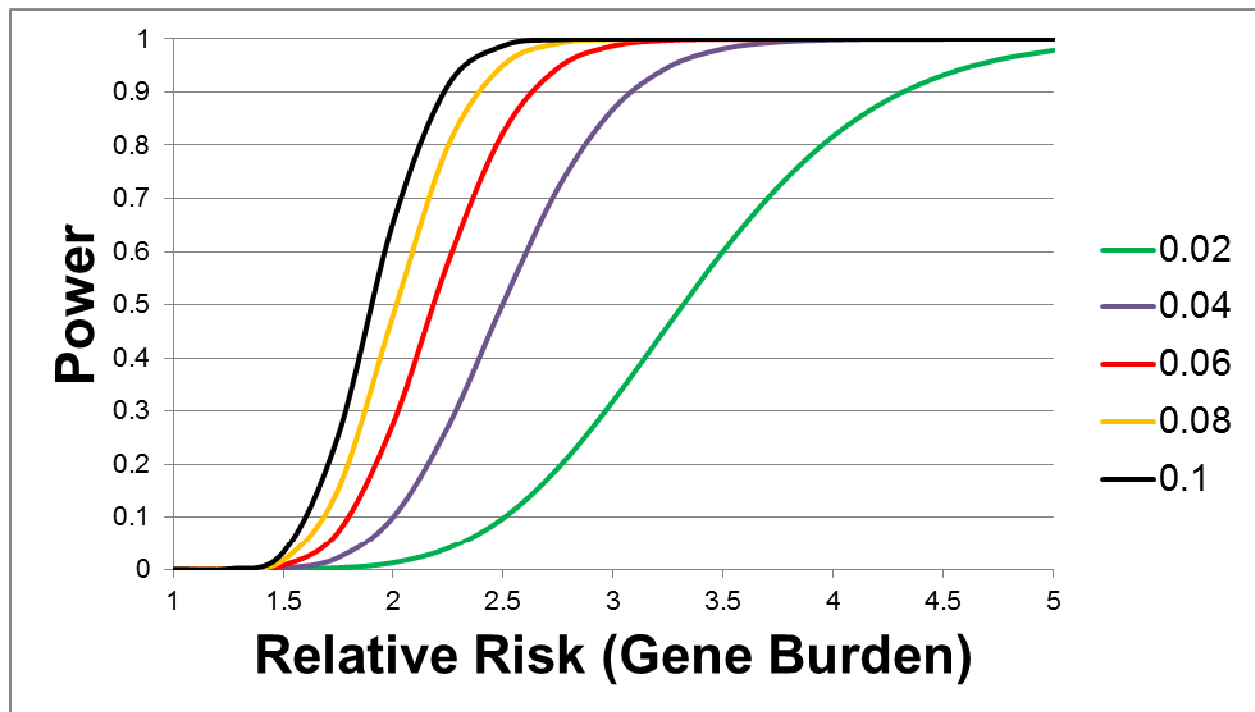


Fig S3



**Fig S3:** Power calculations for 387 attempters and 631 non-attempters for single variant test significance (Bonferroni  $p=5 \times 10^{-8}$ ). Each colored line represents a different risk allele minor allele frequency, the X axis representing the relative risk of the variant to be detected, and the Y axis representing the power for detection of the given variant rarity and relative risk.

Fig S4



**Fig S4:** Power calculations for 387 attempters and 631 non-attempters for gene burden test significance (Bonferroni  $p=2.6 \times 10^{-6}$ ). Each colored line represents a different collapsed risk allele minor allele frequency (the cumulative frequenting of all rare functional variants collapsed by subject), the X axis represents the relative risk of the collapsed variants to be detected, and the Y axis represents the power for detection of the given collapsed variant rarity and relative risk.

# **Supplemental Tables**

Table S1 Previously Associated Genes

**Table S1: Best Results for Genes Found to be Associated with Suicide in Published Studies**

Gene Symbol (References)	Variant Set	Set P-value	Burden Odds Ratio	Predicted Gene Function
<i>ABI3BP</i> [30]	Code Broad MAF05 SKAT	0.060	0.77	Poorly characterized; may be involved in cell fate determination
<i>ACE</i> [31-33]	Code Broad MAF01 SKAT	0.32	1.3	Initiate vasopressin (increase in blood pressure)
<i>ACP1</i> [34]	Reg Broad MAF01 SKAT	0.066	1.1	Acid phosphatase that may act in regulating WNT signaling
<i>ADRA2A</i> [35-40]	-	-	-	Alpha-2a adrenergic; paralog of HTR2A
<i>AKT1</i> [41, 42]	Code Broad MAF01 SKAT	0.82	1.1	Mediator of growth factor induced neuronal survival
<i>BDNF</i> [43-55]	Reg Broad MAF01 Burden	0.0045	1.8	Promotes survival and differentiation of neurons
<i>CCKBR</i> [56, 57]	Code Broad MAF01 Burden	0.59	1.5	Receptor for the CCK neuro peptide
<i>CLOCK</i> [58]	Code Broad MAF05 SKAT	0.093	0.52	A core mediator of the circadian clock cycle
<i>COMT</i> [59-68]	Code Broad MAF05 SKAT	0.44	1.4	Degradation of dopamine, norepinephrine, and epinephrine
<i>CREB1</i> [69]	Code Broad MAF01 SKAT	0.82	1.2	DNA transcription factor
<i>CRHBP</i> [70, 71]	Code Broad MAF01 Burden	0.92	1.1	Suppresses CRH (and thus HPA) activity
<i>CRHR1</i> [70, 72-75]	-	-	-	Mediates numerous CRH responses
<i>CRHR2</i> [75, 76]	Code Broad MAF05 Burden	0.27	0.74	Mediates numerous CRH responses
<i>CRP</i> [77]	Code Disrupt MAF01 Burden	0.33	4.4	Mediates several inflammatory processes
<i>DISC1</i> [78]	Reg Broad MAF05 Burden	0.035	0.72	Mediates cortical development processes
<i>DNMT3B</i> [79]	Code Broad MAF05 SKAT	0.13	1.4	DeNovo methyltransferase activity
<i>DRD2</i> [80-83]	Code Broad MAF01 SKAT	0.45	1.6	Mediates dopamine response by opposing DRD1
<i>EFEMP1</i> [84]	Code Broad MAF01 Burden	0.090	0.14	Activates EGFR signaling, with possible roles in cell migration/adhesion
<i>FAM110C</i> [34]	Code Broad MAF01 Burden	0.87	0.76	Poorly characterized; may play a role in cell spread/migration
<i>FAM150B</i> [34]	-	-	-	Poorly characterized
<i>FKBP5</i> [71, 85-91]	Reg Broad MAF01 Burden	0.094	1.4	Immune regulation mediation
<i>FTO</i> [92]	Code Broad MAF01 Burden	0.24	0.37	Alkylated DNA and RNA repair
<i>GABRG2</i> [93]	Code Broad MAF01 SKAT	0.82	1.1	Receptor for Gamma-Aminobutyric Acid
<i>HTR1A</i> [94-96]	Code Broad MAF01 Burden	0.74	0.87	Serotonergic response mediation
<i>HTR2A</i> [97-103]	Code Broad MAF01 Burden	0.50	2.2	Serotonergic response mediation

Table S1 Previously Associated Genes

<i>LSAMP</i> [104]	Reg Broad MAF05 SKAT	0.071	4.1	Guides neuronal connections in development
<i>MAOA</i> [105-108]	Reg Broad MAF05 Burden	0.084	2.2	Degrades serotonin and other monoamines
<i>MAPK1</i> [69]	Reg Broad MAF05 Burden	0.15	0.68	Regulation of cellular division and life-cycle
<i>NGFR</i> [109]	-	-	-	Mediates neuronal survival
<i>NOS1</i> [110-112]	Code Broad MAF01 Burden	0.84	0.95	Smooth muscle and neurotransmission regulation
<i>NTRK2</i> [40, 54, 113-118]	Code Broad MAF01 Burden	0.095	0.42	Central/peripheral nervous development
<i>ODC1</i> [119]	Code Broad MAF01 Burden	0.40	0.62	A central enzyme in the urea cycle
<i>SAT1</i> [120-124]	-	-	-	Core enzyme in polyamine catabolism
<i>SH3YL1</i> [34]	Code Broad MAF05 Burden	0.49	0.60	Poorly characterized; may regulate cell migration
<i>SKA2</i> [125]	Code Broad MAF05 SKAT	0.41	0.64	Mediates microtubule binding for chromosome organization in mitosis
<i>SLC6A4</i> [59, 103, 106, 126-151]	Code Broad MAF01 Burden	0.12	3.4	Serotonin uptake from synapse
<i>TAAR6</i> [152]	Code Broad MAF05 SKAT	0.83	0.95	Trace amine reception
<i>TBC1D25 (OATL1)</i> [123]	Code Broad MAF05 Burden	0.35	0.70	Autophagosome maturation
<i>TIMELESS</i> [58]	Code Broad MAF05 Burden	0.42	1.4	Regulates DNA replication and the circadian cycle
<i>TPH1</i> [98, 150, 153-170]	Code Broad MAF05 SKAT	0.30	0.60	Core enzyme for serotonin biosynthesis
<i>TPH2</i> [166, 171-183]	Code Disrupt MAF01 Burden	0.19	7.3	Core enzyme for serotonin biosynthesis

Table S1: This table outlines the results from our data of the gene level analyses. The best p-values from all analyses are presented and have been corrected for the covariates of sex, analysis platform, and the first five principal component analyses. Comparisons were all made of suicide attempters (n=387) vs non-attempters (n=631). References represent papers with positive evidence for association of the gene/region with suicidal behavior. Any value listed as “-” could not be calculated due to insufficient detected functional variation for analysis within the gene locus. All listed genes were captured by the arrays.

Table S2 Sample Demographics

<b>Table S2: Sample Demographics</b>				
<b>Attribute</b>	<b>Group</b>	<b>Attempters</b>	<b>Non-Attempters</b>	<b>P-value</b>
Sex	Female	268 (69.3%)	340 (53.9%)	$1.1 \times 10^{-6}$
	Male	119 (30.7%)	291 (46.1%)	$1.1 \times 10^{-6}$
Age	Mean Interview Age	42.0	41.4	0.45
Education	Mean Years of Schooling	14.4	15.0	$5.2 \times 10^{-4}$
Diagnosis	BPI	343 (88.6%)	599 (94.9%)	$3.2 \times 10^{-4}$
	SABP	44 (11.4%)	31 (4.9%)	$1.8 \times 10^{-4}$
	BP Otherwise Affected	0 (0.0%)	1 (0.16%)	1.0
Comorbidities	Alcohol Dependence	160 (41.3%)	157 (24.9%)	$4.9 \times 10^{-8}$
	Substance Abuse	84 (21.7%)	75 (11.9%)	$3.9 \times 10^{-5}$

Table S4 Primary Pathways Hypothesized to be of Importance to Suicidal Behavior Risk

**Table S4: Primary Pathways Hypothesized to be of Importance to Suicidal Behavior Risk**

Pathway	Source	Number of Genes	Best Burden P-value	Odds Ratio	Variant Set
Glutamatergic Synapse	KEGG [184]	115	0.52	0.91	Coding Disruptive MAF01
Hypothalamic-Pituitary-Adrenal (HPA) Axis	Breen et al. [185]	22	0.040	6.5	Coding Disruptive MAF01
Ion-Channels: All Ion Channels	Purcell et al. [16]	237	0.35	0.99	Coding Broad MAF01
Ion-Channels: Cholinergic Receptor Genes	Purcell et al. [16]	16	0.41	0.89	Coding Disruptive MAF01
Ion-Channels: GABA Receptor Genes	Purcell et al. [16]	20	0.19	1.0	Coding Broad MAF05
Ion-Channels Ionotropic Glutamate Receptor Genes	Purcell et al. [16]	14	0.12	1.0	Coding Broad MAF05
Ion-Channels: Potassium Inwardly Rectifying Channel Genes	Purcell et al. [16]	14	0.81	0.98	Coding Broad MAF05
Ion-Channels: Serotonin Receptor Genes	Purcell et al. [16]	16	0.46	0.88	Coding Broad MAF01
Ion-Channels Twin Pore Potassium Channel Genes	Purcell et al. [16]	14	0.48	1.1	Coding Broad MAF05
Ion-Channels Voltage-Gated Calcium Channel Genes	Purcell et al. [16]	26	0.11	2.4	Coding Disruptive MAF01
Ion-Channels Voltage-Gated Potassium Channel Genes	Purcell et al. [16]	50	0.24	1.0	Coding Broad MAF01
Ion-Channels Voltage-Gated Sodium Channel Genes	Purcell et al. [16]	14	0.22	0.97	Coding Broad MAF01
Neuronal ARC	Purcell et al. [16]	28	0.94	0.78	Coding Broad MAF01
Neuronal Cytoplasm	Purcell et al. [16]	271	0.86	0.86	Coding Disruptive MAF01
Neuronal Early Endosomes	Purcell et al. [16]	17	0.71	0.90	Coding Broad MAF01
Neuronal Endoplasmic Reticulum	Purcell et al. [16]	97	0.095	1.4	Coding Disruptive MAF01
Neuronal ER Golgi-Derived Vesicles	Purcell et al. [16]	94	0.12	1.6	Coding Disruptive MAF01
Neuronal Golgi	Purcell et al. [16]	31	0.54	1.1	Coding Disruptive MAF01
Neuronal Mglur5	Purcell et al. [16]	39	0.76	1.0	Coding Broad MAF05
Neuronal Mitochondrion	Purcell et al. [16]	197	0.70	0.95	Coding Broad MAF01
Neuronal NMDAR Network	Purcell et al. [16]	61	0.13	1.2	Coding Broad MAF01
Neuronal Nucleus	Purcell et al. [16]	167	0.098	1.0	Coding Broad MAF05
Neuronal Plasma Membrane	Purcell et al. [16]	50	0.26	1.1	Coding Disruptive MAF01
Neuronal Pre-Synapse	Purcell et al. [16]	431	0.72	1.0	Coding Disruptive MAF01
Neuronal Pre-Synaptic Active Zone	Purcell et al. [16]	173	0.60	1.1	Coding Broad MAF01
Neuronal PSD	Purcell et al. [16]	685	0.48	1.1	Coding Disruptive MAF01
Neuronal PSD-95	Purcell et al. [16]	65	0.66	0.82	Coding Disruptive MAF01
Neuronal Recycling Endosomes Trans-Golgi Network	Purcell et al. [16]	68	0.50	1.1	Coding Broad MAF01
Neuronal Synaptic Vesicle	Purcell et al. [16]	344	0.54	1.0	Coding Broad MAF01
Primary Brain-expressed	Pirooznia et al. [186]	13,128	0.48	1.0	Coding Broad MAF05
Serotonergic Synapse	Judy et al. [187], KEGG [184]	20	0.51	1.6	Coding Disruptive MAF01
Synaptome	Bayes et al. [4]	1,461	0.18	1.0	Regulatory Broad MAF05
WNT Signaling	KEGG [184]	154	0.31	1.2	Coding Disruptive MAF05

Table S5 Average Sample Quality Metrics

**Table S5: Average On-Target Sample Quality Metrics**

Metric	Sequencing Platform / Array	Attempters	Non-Attempters
Average Sequenced Sites Per Sample	GA-IIx / SeqCap EZ v1	26,400,929	26,401,298
	HiSeq / SeqCap EZ v2	36,172,446	36,172,543
	HiSeq / SeqCap EZ v2+	47,656,051	47,655,952
Average per Site Read Depth	GA-IIx / SeqCap EZ v1	56.9	56.5
	HiSeq / SeqCap EZ v2	72.0	66.3
	HiSeq / SeqCap EZ v2+	75.0	74.3
Sites with Read Depth $\geq 1$	GA-IIx / SeqCap EZ v1	95.7%	95.7%
	HiSeq / SeqCap EZ v2	95.5%	95.7%
	HiSeq / SeqCap EZ v2+	97.9%	97.9%
Sites with Read Depth $\geq 10$	GA-IIx / SeqCap EZ v1	88.4%	88.4%
	HiSeq / SeqCap EZ v2	92.0%	91.8%
	HiSeq / SeqCap EZ v2+	94.7%	94.6%
Average Minor Allele Calls	GA-IIx / SeqCap EZ v1	15,709	15,664
	HiSeq / SeqCap EZ v2	23,742	23,386
	HiSeq / SeqCap EZ v2+	27,589	27,415
Average Singleton Variants	GA-IIx / SeqCap EZ v1	160	152
	HiSeq / SeqCap EZ v2	197	204
	HiSeq / SeqCap EZ v2+	235	234
Average Ti/Tv	GA-IIx / SeqCap EZ v1	3.08	3.06
	HiSeq / SeqCap EZ v2	2.96	2.96
	HiSeq / SeqCap EZ v2+	2.83	2.83

Table S5: Reported sample metrics averaged over all individuals in Attempter (N=387) and Non-Attempter (N=631) subjects. Ti/Tv represents the ratio of transition variants to transversion variants. "Sites" represent single on-target base pairs identified via sequence alignment and analysis. The referenced sequence platforms and arrays are described above within the supplemental materials and methods.



Table S6 Top Individual Variant Results

**Table S6: Top Individual Variants (P<0.0001); Suicide Attempters versus Non-Attempters**

SNP ID	hg19 Location	P-Value	Odds Ratio	Gene	Position/Function	Attempter MAF	Non-Attempter MAF
rs2215955	chr7:38469039	$2.8 \times 10^{-5}$	0.61	<i>AMPH</i>	Non-Synonymous (V→F)	0.24	0.33
rs11214703	chr11:113561025	$3.4 \times 10^{-5}$	8.9	<i>TMPRSS5</i>	Synonymous	0.023	0.0028
rs45585336	chr14:20528682	$5.1 \times 10^{-5}$	2.0	<i>OR4L1</i>	Non-Synonymous (I→T)	0.12	0.066
rs45584133	chr14:20528250	$5.6 \times 10^{-5}$	1.9	<i>OR4L1</i>	Non-Synonymous (G→V)	0.12	0.066
rs3752252	chr20:59829847	$6.4 \times 10^{-5}$	1.5	<i>CDH4</i>	Intronic	0.43	0.33
rs17851444	chr3:9920138	$7.2 \times 10^{-5}$	0.54	<i>CIDEA</i>	Synonymous	0.079	0.14
rs9677004	chr19:58117083	$7.5 \times 10^{-5}$	1.6	<i>ZNF530</i>	Non-Synonymous (T→A)	0.21	0.14
rs3742732	chr14:77942316	$8.5 \times 10^{-5}$	1.9	<i>ISM2</i>	Synonymous	0.12	0.067
rs151147550	chr21:31768847	$9.1 \times 10^{-5}$	15	<i>KRTAP13-1</i>	Non-Synonymous (R→H)	0.013	0.00079

Table S7 Top Genes

Table S7: All Top Gene Results (P&lt;0.01)

Gene	Chromosome	hg19 Start	hg19 Stop	Cytogenetic Band	Uncorrected P-value	Odds Ratio	Variant Set
<i>CDK11A</i>	1	1634169	1655766	p36.33	0.0061	1.0	MAF01 Coding Broad SKAT
<i>TNFRSF8</i>	1	12123434	12204264	p36.22	0.0046	1.1	MAF05 Coding Broad SKAT
<i>AADACL4</i>	1	12704566	12727097	p36.21	0.0087	3.0	MAF01 Coding Broad Burden
<i>PRDM2</i>	1	14026693	14151574	p36.21	0.0086	2.7	MAF01 Coding Broad Burden
<i>WNT4</i>	1	22443798	22470462	p36.12	0.0031	4.3	MAF01 Coding Broad SKAT
<i>FUCA1</i>	1	24171567	24194784	p36.11	0.0046	3.5	MAF01 Coding Broad Burden
<i>PPT1</i>	1	40538379	40563375	p34.2	0.0052	4.5	MAF01 Coding Broad SKAT
<i>LRRC41</i>	1	46726868	46769280	p34.1	0.0071	2.3	MAF01 Coding Broad SKAT
<i>RPF1</i>	1	84944942	84963473	p22.3	0.0019	2.0	MAF05 Coding Broad Burden
<i>OLFM3</i>	1	102268130	102462586	p21.1	0.0074	1.0	MAF01 Coding Broad SKAT
<i>PROK1</i>	1	110993822	110999976	p13.3	0.0083	3.6	MAF01 Coding Broad SKAT
<i>CHIA</i>	1	111833484	111863188	p13.2	0.0015	2.0	MAF01 Coding Broad SKAT
<i>CCT3</i>	1	156278759	156337664	q22	0.0080	8.1	MAF01 Coding Broad SKAT
<i>OR6Y1</i>	1	158516918	158517895	q23.1	0.0093	0.39	MAF05 Coding Broad Burden
<i>OLFML2B</i>	1	161952982	161993644	q23.3	0.0048	0.067	MAF01 Coding Broad Burden
<i>PRDX6</i>	1	173446405	173457946	q25.1	0.0070	2.0	MAF05 Regulatory Broad Burden
<i>RC3H1</i>	1	173900352	173991435	q25.1	0.0090	1.6	MAF05 Coding Broad SKAT
<i>IER5</i>	1	181057638	181059977	q25.3	0.0059	3.7	MAF01 Coding Broad SKAT
<i>COLGALT2</i>	1	183898796	184006863	q25.3	0.00039	3.1	MAF05 Coding Broad Burden
<i>RGS21</i>	1	192286122	192336415	q31.2	0.0095	1.5	MAF05 Coding Broad SKAT
<i>C1orf106</i>	1	200860176	200884863	q32.1	0.00020	1.9	MAF01 Coding Broad SKAT
<i>C1orf35</i>	1	228288427	228293112	q42.13	0.0096	1.1	MAF05 Coding Broad SKAT
<i>GNPAT</i>	1	231376953	231413719	q42.2	0.00043	27	MAF01 Regulatory Broad Burden
<i>CEP170</i>	1	243287730	243418650	q43	0.0091	16	MAF01 Regulatory Broad Burden
<i>C1orf100</i>	1	244515937	244552965	q44	0.00076	5.6	MAF05 Coding Broad SKAT
<i>SMYD3</i>	1	245912642	246670614	q44	0.0056	1.6	MAF01 Coding Broad SKAT
<i>ROCK2</i>	2	11319887	11488456	p25.1	0.0032	0.66	MAF05 Regulatory Broad SKAT
<i>OTOF</i>	2	26680071	26781566	p23.3	0.0064	1.6	MAF05 Coding Broad Burden
<i>CAPN14</i>	2	31395924	31456724	p23.1	0.0081	5.1	MAF01 Coding Broad SKAT
<i>HEATR5B</i>	2	37195526	37311485	p22.2	0.0043	2.2	MAF01 Coding Broad SKAT
<i>CDKL4</i>	2	39402787	39456729	p22.1	0.0049	2.0	MAF01 Coding Broad SKAT
<i>SRBD1</i>	2	45615819	45839304	p21	0.0058	2.3	MAF01 Coding Broad Burden
<i>NRXN1</i>	2	50145643	51259674	p16.3	0.0071	1.2	MAF01 Regulatory Broad SKAT
<i>PROKR1</i>	2	68870721	68882708	p13.3	0.0099	2.4	MAF01 Coding Broad SKAT
<i>PCBP1</i>	2	70314585	70316332	p13.3	0.00095	2.5	MAF05 Regulatory Broad SKAT
<i>RNF181</i>	2	85822848	85824736	p11.2	0.0047	4.2	MAF01 Coding Broad SKAT
<i>GNLY</i>	2	85912298	85925977	p11.2	0.0034	4.7	MAF01 Coding Broad Burden
<i>SULT1C3</i>	2	108863651	108881807	q12.3	0.0011	2.1	MAF05 Coding Broad SKAT
<i>KIF5C</i>	2	149632819	149883273	q23.1	0.0040	5.4	MAF01 Coding Broad SKAT
<i>HDLBP</i>	2	242166679	242256476	q37.3	0.0070	0.95	MAF01 Regulatory Narrow SKAT
<i>EDEM1</i>	3	5229331	5261642	p26.1	0.0094	1.6	MAF01 Coding Broad SKAT

Table S7 Top Genes

<i>RP11-438J1.1</i>	3	10291056	10327480	p25.3	0.0099	1.6	MAF01 Coding Broad SKAT
<i>ENTPD3</i>	3	40428647	40470110	p22.1	0.0071	3.3	MAF01 Coding Broad SKAT
<i>NBEAL2</i>	3	47021173	47051193	p21.31	0.0036	2.4	MAF01 Coding Broad Burden
<i>SMARCC1</i>	3	47626762	47823596	p21.31	0.0052	1.3	MAF01 Coding Broad SKAT
<i>CDHR4</i>	3	49828165	49837268	p21.31	0.0091	3.1	MAF05 Coding Broad Burden
<i>TRAIIP</i>	3	49866034	49894007	p21.31	0.0099	0.076	MAF01 Coding Broad Burden
<i>LRTM1</i>	3	54952264	55001115	p14.3	0.0074	8.7	MAF01 Coding Broad Burden
<i>OR5H2</i>	3	98001732	98002676	q11.2	0.0043	3.7	MAF01 Coding Broad Burden
<i>KIAA1524</i>	3	108268716	108308491	q13.13	0.0083	1.1	MAF05 Coding Broad SKAT
<i>GAP43</i>	3	115342171	115440337	q13.31	0.0046	2.0	MAF05 Regulatory Broad SKAT
<i>MYLK</i>	3	123328896	123603178	q21.1	0.0079	0.97	MAF01 Coding Broad SKAT
<i>OSBPL11</i>	3	125247702	125313934	q21.2	0.0074	3.3	MAF01 Coding Broad SKAT
<i>SLCO2A1</i>	3	133651540	133771028	q22.2	0.0025	2.7	MAF01 Coding Broad SKAT
<i>PRR23B</i>	3	138737873	138739768	q23	0.0084	23	MAF01 Coding Broad Burden
<i>MED12L</i>	3	150803484	151154860	q25.1	0.0014	0.13	MAF01 Coding Broad Burden
<i>IFT80</i>	3	159974774	160117668	q25.33	0.0046	2.8	MAF01 Coding Broad SKAT
<i>NAALADL2</i>	3	174156363	175523428	q26.31	0.0051	4.9	MAF01 Coding Broad Burden
<i>RTP1</i>	3	186915274	186919253	q27.3	0.0041	0.064	MAF01 Coding Broad Burden
<i>LRCH3</i>	3	197518097	197615307	q29	0.0070	3.7	MAF01 Coding Broad Burden
<i>HTT</i>	4	3076408	3245676	p16.3	0.0025	1.9	MAF01 Regulatory Broad Burden
<i>LRPAP1</i>	4	3508103	3534286	p16.3	0.0074	0.50	MAF05 Coding Broad Burden
<i>CLNK</i>	4	10488019	10686489	p16.1	0.0028	2.9	MAF01 Coding Broad SKAT
<i>RFC1</i>	4	39289076	39367995	p14	0.0080	2.8	MAF01 Coding Broad SKAT
<i>TMPRSS11B</i>	4	69092371	69111438	q13.2	0.0036	3.5	MAF01 Coding Broad SKAT
<i>ART3</i>	4	76932337	77033955	q21.1	0.0080	3.4	MAF01 Coding Broad SKAT
<i>PRDM5</i>	4	121606074	121844025	q27	0.0055	2.9	MAF01 Coding Broad SKAT
<i>GRIA2</i>	4	158125334	158287227	q32.1	0.0034	23	MAF01 Coding Broad SKAT
<i>MLF1IP</i>	4	185615772	185655287	q35.1	0.0071	1.7	MAF01 Coding Broad SKAT
<i>ACSL1</i>	4	185676749	185747972	q35.1	0.0031	2.5	MAF01 Coding Broad SKAT
<i>FAT1</i>	4	187508937	187647876	q35.2	0.0069	1.5	MAF01 Coding Broad SKAT
<i>PDZD2</i>	5	31639517	32111037	p13.3	0.0069	1.5	MAF05 Coding Broad Burden
<i>ADAMTS12</i>	5	33523640	33892297	p13.2	0.0024	0.54	MAF05 Coding Broad Burden
<i>RXFP3</i>	5	33936491	33939023	p13.2	0.0025	3.0	MAF01 Coding Broad SKAT
<i>SLC1A3</i>	5	36606457	36688436	p13.2	0.0017	1.1	MAF05 Regulatory Broad SKAT
<i>NNT</i>	5	43602794	43707507	p12	0.00067	2.8	MAF01 Coding Broad SKAT
<i>MAST4</i>	5	65892176	66465423	q12.3	0.0044	1.7	MAF05 Coding Broad Burden
<i>BHMT</i>	5	78407602	78428108	q14.1	0.0014	2.6	MAF01 Coding Broad SKAT
<i>ZFYVE16</i>	5	79703832	79775169	q14.1	0.0045	2.6	MAF01 Coding Broad SKAT
<i>RASGRF2</i>	5	80256491	80525975	q14.1	0.0049	1.5	MAF01 Regulatory Broad SKAT
<i>REEP5</i>	5	112212084	112258236	q22.2	0.0035	2.7	MAF01 Regulatory Broad Burden
<i>SEMA6A</i>	5	115779312	115910630	q23.1	0.0077	0.41	MAF01 Coding Broad Burden
<i>SLC12A2</i>	5	127419458	127525380	q23.3	0.0057	2.7	MAF01 Coding Broad SKAT
<i>SFXN1</i>	5	174904065	174956745	q35.2	0.00081	1.7	MAF01 Regulatory Broad SKAT
<i>BTN3A1</i>	6	26402465	26415444	p22.2	0.0046	2.3	MAF01 Coding Broad SKAT
<i>UHRF1BP1</i>	6	34759857	34850915	p21.31	0.0054	3.3	MAF01 Coding Broad Burden
<i>ENPP5</i>	6	46126924	46138708	p21.1	0.0072	0.21	MAF01 Coding Broad Burden

Table S7 Top Genes

<i>DDX43</i>	6	74104471	74127292	q13	0.0028	4.8	MAF01 Coding Broad SKAT
<i>LIN28B</i>	6	105404923	105531207	q16.3	0.0065	6.1	MAF01 Coding Broad Burden
<i>SESN1</i>	6	109307640	109416022	q21	0.0037	6.7	MAF01 Coding Broad SKAT
<i>TRAF3IP2</i>	6	111877657	111927481	q21	0.0082	0.16	MAF01 Coding Broad Burden
<i>PHACTR2</i>	6	143857982	144152322	q24.2	0.0062	4.3	MAF01 Coding Broad Burden
<i>GRM1</i>	6	146348782	146758734	q24.3	0.00023	2.8	MAF01 Coding Broad SKAT
<i>PLEKHG1</i>	6	150920999	151164799	q25.1	0.0064	2.1	MAF05 Coding Broad SKAT
<i>SYNE1</i>	6	152442819	152958936	q25.2	0.00058	1.9	MAF01 Regulatory Broad Burden
<i>FNDC1</i>	6	159590429	159693141	q25.3	0.0063	1.3	MAF01 Coding Broad SKAT
<i>DGKB</i>	7	14184674	15014402	p21.2	0.0089	1.7	MAF01 Regulatory Broad Burden
<i>GPNMB</i>	7	23275586	23314727	p15.3	0.0089	2.8	MAF01 Coding Disrupt SKAT
<i>HOXA1</i>	7	27132612	27135615	p15.2	0.0043	4.0	MAF01 Coding Broad SKAT
<i>PLEKHA8</i>	7	30067020	30170096	p14.3	0.0095	3.3	MAF01 Coding Broad SKAT
<i>NT5C3A</i>	7	33053742	33102409	p14.3	0.0087	6.1	MAF01 Coding Broad SKAT
<i>BBS9</i>	7	33168856	33645680	p14.3	0.0029	0.41	MAF05 Regulatory Broad Burden
<i>ZNF680</i>	7	63980262	64023484	q11.21	0.0063	0.15	MAF01 Coding Broad Burden
<i>TRIM50</i>	7	72726535	72742085	q11.23	0.0053	6.2	MAF01 Coding Broad Burden
<i>CCDC146</i>	7	76751751	76958850	q11.23	0.00090	2.2	MAF01 Coding Broad SKAT
<i>GNAT3</i>	7	80087987	80141336	q21.11	0.0084	0.078	MAF01 Coding Broad Burden
<i>ARPC1B</i>	7	98971872	98992424	q22.1	0.0077	0.52	MAF05 Coding Broad Burden
<i>CYP3A43</i>	7	99425636	99463718	q22.1	0.0055	2.0	MAF05 Coding Broad Burden
<i>GPC2</i>	7	99767229	99774995	q22.1	0.0078	3.2	MAF01 Coding Broad SKAT
<i>SRPK2</i>	7	104751151	105039755	q22.3	0.0033	3.1	MAF01 Coding Broad SKAT
<i>LAMB1</i>	7	107564244	107643700	q31.1	0.0034	1.2	MAF05 Coding Broad SKAT
<i>CEP41</i>	7	130033612	130082274	q32.2	0.0058	1.8	MAF01 Coding Broad SKAT
<i>AGK</i>	7	141250989	141355044	q34	0.0076	0.44	MAF01 Regulatory Broad SKAT
<i>SLC4A2</i>	7	150754297	150773614	q36.1	0.0043	4.0	MAF01 Coding Broad SKAT
<i>ESYT2</i>	7	158523686	158622944	q36.3	0.0078	15	MAF01 Regulatory Narrow SKAT
<i>ZDHHC2</i>	8	17013538	17082308	p22	0.0032	2.6	MAF01 Coding Broad SKAT
<i>PBK</i>	8	27667137	27695612	p21.1	0.0039	22	MAF01 Coding Broad Burden
<i>WHSC1L1</i>	8	38127215	38239790	p11.23	0.0080	0.32	MAF05 Coding Broad Burden
<i>PXDNL</i>	8	52232138	52722005	q11.23	0.0053	2.8	MAF05 Coding Disrupt Burden
<i>KCNB2</i>	8	73449626	73850584	q13.3	0.0065	20	MAF01 Regulatory Broad Burden
<i>GDAP1</i>	8	75233365	75401107	q21.11	0.0071	7.9	MAF01 Coding Broad Burden
<i>CDH17</i>	8	95139399	95229531	q22.1	0.0070	1.2	MAF01 Coding Broad SKAT
<i>TBC1D31</i>	8	124054208	124164393	q24.13	0.0014	1.4	MAF01 Coding Broad SKAT
<i>NDRG1</i>	8	134249414	134314265	q24.22	0.0048	2.4	MAF05 Regulatory Broad Burden
<i>IL33</i>	9	6215805	6257983	p24.1	0.0060	5.5	MAF01 Coding Broad SKAT
<i>LURAP1L</i>	9	12775020	12822130	p23	0.0016	3.4	MAF01 Coding Broad SKAT
<i>C9orf3</i>	9	97488983	97849441	q22.32	0.0080	1.8	MAF01 Coding Broad SKAT
<i>ANKS6</i>	9	101493611	101559247	q22.33	0.0029	2.8	MAF01 Coding Broad SKAT
<i>STOM</i>	9	124101355	124132531	q33.2	0.0049	0.89	MAF05 Regulatory Broad SKAT
<i>GLE1</i>	9	131266979	131304567	q34.11	0.0019	4.1	MAF05 Coding Broad Burden
<i>NELFB</i>	9	140149625	140167998	q34.3	0.0065	5.1	MAF01 Coding Broad SKAT
<i>TAF3</i>	10	7860467	8058590	p14	0.0037	19	MAF01 Coding Broad Burden
<i>OTUD1</i>	10	23728198	23731308	p12.2	0.0077	0.40	MAF05 Coding Broad Burden

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<i>CREM</i>	10	35415719	35501886	p11.21	0.0059	3.6	MAF01 Coding Broad SKAT
<i>CHAT</i>	10	50817141	50901925	q11.23	0.0058	3.8	MAF01 Coding Broad Burden
<i>A1CF</i>	10	52559169	52645435	q11.23	0.0083	2.0	MAF01 Coding Broad SKAT
<i>CFAP70</i>	10	75013517	75118617	q22.2	0.000065	4.1	MAF01 Coding Broad SKAT
<i>POLR3A</i>	10	79734907	79789303	q22.3	0.0010	0.45	MAF05 Coding Broad SKAT
<i>ANXA11</i>	10	81910645	81965328	q22.3	0.0076	0.74	MAF05 Coding Broad SKAT
<i>KIF20B</i>	10	91461367	91534700	q23.31	0.0081	0.36	MAF01 Regulatory Narrow Burden
<i>LZTS2</i>	10	102756375	102767593	q24.31	0.0029	4.4	MAF01 Coding Broad SKAT
<i>TECTB</i>	10	114043493	114064793	q25.2	0.0062	1.6	MAF01 Coding Broad SKAT
<i>AFAP1L2</i>	10	116054583	116164515	q25.3	0.0074	5.5	MAF01 Coding Broad Burden
<i>RAB11FIP2</i>	10	119764427	119806114	q26.11	0.0050	3.5	MAF01 Coding Broad Burden
<i>PTPRE</i>	10	129705325	129884119	q26.2	0.0033	0.23	MAF01 Coding Broad Burden
<i>PSMD13</i>	11	236546	252983	p15.5	0.0044	8.8	MAF01 Coding Broad Burden
<i>MUC5B</i>	11	1244296	1283406	p15.5	0.0060	1.6	MAF01 Coding Broad Burden
<i>KCNQ1</i>	11	2465914	2870339	p15.5	0.0044	1.2	MAF01 Regulatory Narrow SKAT
<i>OR51A4</i>	11	4967355	4968356	p15.4	0.00093	0.46	MAF05 Coding Broad Burden
<i>OR52N4</i>	11	5775923	5776959	p15.4	0.0094	2.5	MAF01 Coding Broad Burden
<i>OR10A6</i>	11	7949180	7950209	p15.4	0.0049	0.51	MAF05 Coding Broad Burden
<i>IPO7</i>	11	9406169	9469673	p15.4	0.0076	2.3	MAF01 Regulatory Broad Burden
<i>TSG101</i>	11	18489883	18548779	p15.1	0.0038	6.8	MAF01 Coding Broad SKAT
<i>BDNF</i>	11	27676440	27743605	p14.1	0.0045	1.8	MAF01 Regulatory Broad Burden
<i>KIAA1549L</i>	11	33563618	33695648	p13	0.0063	2.5	MAF01 Coding Broad SKAT
<i>CD59</i>	11	33719807	33757991	p13	0.0030	1.6	MAF05 Regulatory Broad Burden
<i>PACSIN3</i>	11	47199076	47207994	p11.2	0.0039	3.1	MAF01 Coding Broad SKAT
<i>FOLH1</i>	11	49168187	49230222	p11.12	0.0046	2.5	MAF01 Coding Broad SKAT
<i>OR5M11</i>	11	56309746	56310757	q12.1	0.0031	5.5	MAF01 Coding Broad SKAT
<i>OR5B3</i>	11	58169937	58170882	q12.1	0.0056	0.54	MAF01 Coding Broad SKAT
<i>TMEM132A</i>	11	60691935	60704631	q12.2	0.0053	0.49	MAF05 Coding Broad Burden
<i>PPP1R32</i>	11	61248592	61258403	q12.2	0.0055	2.0	MAF05 Coding Broad Burden
<i>KLC2</i>	11	66024765	66035331	q13.2	0.0051	1.5	MAF01 Regulatory Broad SKAT
<i>BRMS1</i>	11	66104804	66112596	q13.2	0.0058	6.7	MAF01 Coding Broad SKAT
<i>CABP4</i>	11	67219877	67226699	q13.2	0.0087	0.074	MAF01 Coding Broad Burden
<i>SHANK2</i>	11	70313961	70963623	q13.4	0.00022	2.2	MAF05 Coding Broad Burden
<i>FAT3</i>	11	92085262	92629618	q14.3	0.0060	1.5	MAF05 Coding Broad Burden
<i>ATM</i>	11	108093211	108239829	q22.3	0.0097	0.63	MAF05 Coding Broad Burden
<i>TMPRSS5</i>	11	113558272	113577095	q23.2	0.0099	2.5	MAF01 Coding Broad SKAT
<i>TREH</i>	11	118528026	118550399	q23.3	0.0022	3.0	MAF05 Coding Broad Burden
<i>ABCG4</i>	11	119019722	119033360	q23.3	0.0041	21	MAF01 Coding Broad Burden
<i>ZNF202</i>	11	123594885	123612383	q24.1	0.0058	0.21	MAF05 Coding Broad Burden
<i>VWA5A</i>	11	123986069	124018428	q24.2	0.0038	1.3	MAF01 Coding Broad SKAT
<i>SRPR</i>	11	126132814	126139039	q24.2	0.0047	2.3	MAF05 Coding Broad SKAT
<i>SLC6A13</i>	12	329789	372039	p13.33	0.000061	3.6	MAF01 Coding Broad SKAT
<i>FKBP4</i>	12	2904119	2914576	p13.33	0.0079	15	MAF01 Regulatory Broad Burden
<i>MFAP5</i>	12	8789942	8815484	p13.31	0.0095	0.076	MAF01 Coding Broad Burden
<i>GOLT1B</i>	12	21654715	21671342	p12.1	0.0078	22	MAF01 Coding Broad Burden
<i>CNTN1</i>	12	41086244	41466220	q12	0.0087	2.5	MAF01 Coding Broad SKAT

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<i>FMNL3</i>	12	50031724	50101948	q13.12	0.0078	0.073	MAF01 Regulatory Narrow Burden
<i>KRT79</i>	12	53215194	53228079	q13.13	0.0078	0.61	MAF05 Coding Broad Burden
<i>SOAT2</i>	12	53497302	53518322	q13.13	0.0085	2.9	MAF01 Coding Broad SKAT
<i>OR6C68</i>	12	55886147	55887100	q13.2	0.0089	0.16	MAF05 Coding Broad Burden
<i>MARS</i>	12	57869228	57911352	q13.3	0.0076	0.15	MAF01 Coding Broad Burden
<i>USP15</i>	12	62654119	62811211	q14.1	0.0023	0.30	MAF05 Regulatory Broad Burden
<i>LEMD3</i>	12	65563351	65642107	q14.3	0.0017	3.1	MAF01 Coding Broad SKAT
<i>PPP1R12A</i>	12	80167343	80329240	q21.31	0.00089	3.5	MAF01 Regulatory Broad SKAT
<i>EEA1</i>	12	93164413	93323107	q22	0.0056	1.6	MAF01 Coding Broad SKAT
<i>FGD6</i>	12	95470525	95611258	q22	0.0070	0.35	MAF01 Coding Broad Burden
<i>APAF1</i>	12	99038919	99129204	q23.1	0.0084	1.9	MAF05 Coding Broad SKAT
<i>ALDH2</i>	12	112204691	112247782	q24.12	0.0012	10	MAF01 Coding Broad Burden
<i>MORN3</i>	12	122089024	122110537	q24.31	0.0012	11	MAF01 Coding Broad Burden
<i>RILPL1</i>	12	123955925	124018265	q24.31	0.0036	8.7	MAF01 Coding Broad SKAT
<i>ZNF140</i>	12	133656424	133684130	q24.33	0.0084	17	MAF01 Coding Broad Burden
<i>TPTE2</i>	13	19997017	20110903	q12.11	0.0041	2.1	MAF01 Coding Broad SKAT
<i>DCLK1</i>	13	36345478	36705443	q13.3	0.00039	4.9	MAF01 Coding Broad SKAT
<i>TNFSF11</i>	13	43136872	43182149	q14.11	0.0024	2.4	MAF01 Coding Broad SKAT
<i>COG3</i>	13	46039060	46110765	q14.13	0.0053	3.5	MAF01 Coding Broad SKAT
<i>LRCH1</i>	13	47127303	47327175	q14.13	0.0093	1.4	MAF01 Coding Broad SKAT
<i>DACH1</i>	13	72012098	72441330	q21.33	0.0046	8.7	MAF01 Coding Broad Burden
<i>KCTD12</i>	13	77454312	77460540	q22.3	0.00077	1.6	MAF01 Regulatory Broad SKAT
<i>OR4K2</i>	14	20344391	20345468	q11.2	0.0050	3.1	MAF01 Coding Broad SKAT
<i>METTL17</i>	14	21457929	21465189	q11.2	0.0034	0.23	MAF05 Coding Broad Burden
<i>EAPP</i>	14	34985135	35008916	q13.1	0.0067	8.0	MAF01 Coding Broad Burden
<i>RP11-407N17.3</i>	14	39703106	39820397	q21.1	0.0015	0.24	MAF01 Coding Broad Burden
<i>CTAGE5</i>	14	39734488	39856156	q21.1	0.0033	0.26	MAF01 Coding Broad Burden
<i>GPX2</i>	14	65405870	65409531	q23.3	0.0033	0.34	MAF05 Coding Broad SKAT
<i>ACTN1</i>	14	69340860	69446157	q24.1	0.0079	0.60	MAF01 Regulatory Broad SKAT
<i>ZFYVE1</i>	14	73436159	73493920	q24.2	0.0032	2.5	MAF01 Coding Broad SKAT
<i>NUMB</i>	14	73741815	73930348	q24.3	0.0069	8.0	MAF01 Coding Broad Burden
<i>WARS</i>	14	100800125	100843142	q32.2	0.0071	2.5	MAF01 Coding Broad SKAT
<i>ASPG</i>	14	104552016	104579098	q32.33	0.0041	7.1	MAF01 Coding Broad SKAT
<i>AHNAK2</i>	14	105403581	105444694	q32.33	0.0013	1.0	MAF01 Coding Broad SKAT
<i>AVEN</i>	15	34158428	34331377	q14	0.00079	1.1	MAF05 Coding Broad SKAT
<i>DUOX2</i>	15	45384848	45406542	q21.1	0.0083	23	MAF01 Coding Disrupt Burden
<i>CORO2B</i>	15	68871308	69020145	q23	0.0051	1.4	MAF01 Coding Broad SKAT
<i>GRAMD2</i>	15	72452148	72490126	q23	0.0061	4.3	MAF01 Coding Broad Burden
<i>FBXO22</i>	15	76196200	76227609	q24.2	0.0014	0.056	MAF01 Coding Broad Burden
<i>CHRNA5</i>	15	78857862	78887611	q25.1	0.0028	0.061	MAF01 Coding Broad Burden
<i>KLHL25</i>	15	86302554	86338261	q25.3	0.0082	2.6	MAF01 Coding Broad SKAT
<i>C16orf13</i>	16	684429	686358	p13.3	0.0016	6.7	MAF01 Coding Broad SKAT
<i>STUB1</i>	16	730224	732870	p13.3	0.0038	2.6	MAF01 Regulatory Broad Burden
<i>TSC2</i>	16	2097466	2138716	p13.3	0.0089	0.63	MAF05 Regulatory Broad Burden
<i>CASKIN1</i>	16	2227184	2246526	p13.3	0.0084	2.7	MAF01 Regulatory Broad Burden
<i>PAM16</i>	16	4381550	4405608	p13.3	0.0080	1.1	MAF05 Regulatory Broad SKAT

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<i>UBN1</i>	16	4896666	4932361	p13.3	0.0071	0.21	MAF01 Coding Broad Burden
<i>AC004381.6</i>	16	20817751	20860987	p12.3	0.0027	1.3	MAF01 Coding Broad SKAT
<i>TMEM219</i>	16	29952206	29984373	p11.2	0.0074	5.5	MAF01 Coding Broad SKAT
<i>CCDC102A</i>	16	57546090	57570511	q21	0.0075	7.7	MAF01 Coding Broad Burden
<i>TEPP</i>	16	58010339	58022020	q21	0.00027	31	MAF01 Coding Broad Burden
<i>ZNF319</i>	16	58028572	58034357	q21	0.0056	4.8	MAF01 Coding Broad Burden
<i>CDH11</i>	16	64977656	65160015	q21	0.0084	0.30	MAF01 Coding Broad Burden
<i>CDH3</i>	16	68670092	68756519	q22.1	0.0059	3.2	MAF01 Coding Broad Burden
<i>TAF1C</i>	16	84211458	84220669	q24.1	0.0071	3.4	MAF01 Coding Broad Burden
<i>ZNF778</i>	16	89284118	89295363	q24.3	0.0040	1.0	MAF01 Coding Broad SKAT
<i>NXN</i>	17	702553	883010	p13.3	0.0039	2.5	MAF01 Coding Broad SKAT
<i>MYO1C</i>	17	1367392	1396106	p13.3	0.0099	1.8	MAF05 Coding Broad Burden
<i>TMEM88</i>	17	7758383	7759417	p13.1	0.0043	3.6	MAF01 Coding Broad SKAT
<i>MPRIIP</i>	17	16945859	17120993	p11.2	0.0075	2.0	MAF01 Coding Broad SKAT
<i>ALDH3A2</i>	17	19551449	19580911	p11.2	0.0071	3.4	MAF01 Coding Broad SKAT
<i>MYO18A</i>	17	27400528	27507430	q11.2	0.0065	2.3	MAF01 Coding Broad Burden
<i>GIT1</i>	17	27900487	27921072	q11.2	0.0070	2.6	MAF01 Regulatory Broad Burden
<i>MYO1D</i>	17	30819540	31204195	q11.2	0.0055	0.28	MAF01 Coding Broad Burden
<i>CCL16</i>	17	34303529	34308532	q12	0.0090	0.42	MAF05 Coding Broad Burden
<i>ATP6V0A1</i>	17	40610862	40674629	q21.2	0.0044	2.3	MAF01 Regulatory Broad Burden
<i>MTMR4</i>	17	56566898	56595266	q22	0.0093	0.98	MAF01 Coding Broad SKAT
<i>CBX2</i>	17	77751931	77761782	q25.3	0.0069	0.96	MAF01 Coding Broad SKAT
<i>TBC1D16</i>	17	77906142	78009647	q25.3	0.0011	2.1	MAF05 Coding Broad Burden
<i>EMILIN2</i>	18	2847028	2915991	p11.32	0.0030	1.6	MAF05 Coding Broad SKAT
<i>DSG1</i>	18	28898052	28936992	q12.1	0.0051	1.6	MAF01 Coding Broad SKAT
<i>DSG3</i>	18	29027758	29058665	q12.1	0.00018	2.7	MAF01 Coding Broad SKAT
<i>SLC25A52</i>	18	29339525	29340843	q12.1	0.0096	3.1	MAF01 Coding Broad SKAT
<i>LOXHD1</i>	18	44056935	44236996	q21.1	0.00046	1.0	MAF05 Coding Broad SKAT
<i>ZNF516</i>	18	74069644	74207146	q23	0.0090	1.9	MAF01 Coding Broad SKAT
<i>PPAP2C</i>	19	281040	291393	p13.3	0.0097	0.29	MAF01 Coding Broad Burden
<i>THEG</i>	19	361750	376013	p13.3	0.0094	0.074	MAF01 Coding Broad Burden
<i>POLRMT</i>	19	617223	633597	p13.3	0.0012	26	MAF01 Coding Broad Burden
<i>PRSS57</i>	19	685521	695460	p13.3	0.0093	2.4	MAF01 Coding Broad SKAT
<i>FSD1</i>	19	4304597	4323840	p13.3	0.0052	3.4	MAF01 Regulatory Broad Burden
<i>ZNF559</i>	19	9434448	9461838	p13.2	0.0023	5.9	MAF01 Coding Broad SKAT
<i>ICAM5</i>	19	10400657	10407454	p13.2	0.0039	0.063	MAF01 Regulatory Broad Burden
<i>ZNF823</i>	19	11832080	11849824	p13.2	0.0036	1.9	MAF05 Coding Broad SKAT
<i>CCDC130</i>	19	13842574	13874110	p13.2	0.0014	11	MAF01 Coding Broad Burden
<i>CC2D1A</i>	19	14017014	14041692	p13.12	0.0073	1.6	MAF01 Coding Broad SKAT
<i>OR7A17</i>	19	14991138	14992264	p13.12	0.0099	0.51	MAF05 Coding Broad Burden
<i>AP1M1</i>	19	16308389	16346160	p13.11	0.0030	1.6	MAF01 Regulatory Broad SKAT
<i>SUGP1</i>	19	19386827	19431653	p13.11	0.0060	22	MAF01 Coding Broad SKAT
<i>ZNF429</i>	19	21679484	21739072	p12	0.0090	1.2	MAF05 Coding Broad SKAT
<i>RHPN2</i>	19	33469499	33555794	q13.11	0.00079	2.2	MAF05 Coding Broad Burden
<i>ZNF780B</i>	19	40534167	40562116	q13.2	0.0076	3.3	MAF05 Coding Disrupt Burden
<i>CNTD2</i>	19	40728115	40732597	q13.2	0.0057	6.9	MAF01 Coding Broad SKAT

Table S7 Top Genes

<i>IRGQ</i>	19	44088521	44100287	q13.31	0.0081	2.3	MAF05 Regulatory Narrow SKAT
<i>ZNF221</i>	19	44455375	44471861	q13.31	0.0015	3.6	MAF01 Coding Broad SKAT
<i>ZNF222</i>	19	44529494	44537265	q13.31	0.0099	2.9	MAF01 Coding Broad Burden
<i>TRPM4</i>	19	49660998	49715093	q13.33	0.0073	0.87	MAF01 Coding Broad SKAT
<i>TSKS</i>	19	50243010	50266587	q13.33	0.0055	1.7	MAF01 Coding Broad SKAT
<i>SIGLEC12</i>	19	51994611	52005043	q13.41	0.0081	0.32	MAF01 Coding Broad Burden
<i>PPP2R1A</i>	19	52693292	52730687	q13.41	0.0072	2.7	MAF01 Regulatory Broad Burden
<i>ZNF534</i>	19	52932440	52955568	q13.41	0.0087	0.79	MAF05 Coding Broad SKAT
<i>ZNF835</i>	19	57174020	57183151	q13.43	0.0087	1.9	MAF05 Coding Broad SKAT
<i>ZNF530</i>	19	58111253	58124090	q13.43	0.0028	1.2	MAF05 Coding Broad SKAT
<i>CSNK2A1</i>	20	459116	524465	p13	0.0028	6.0	MAF01 Regulatory Narrow SKAT
<i>PYGB</i>	20	25228705	25278650	p11.21	0.00047	0.11	MAF01 Coding Broad Burden
<i>ZNF341</i>	20	32319463	32380075	q11.22	0.0064	0.93	MAF01 Coding Broad SKAT
<i>CPNE1</i>	20	34213953	34252878	q11.22	0.0092	2.0	MAF01 Coding Broad SKAT
<i>SLC32A1</i>	20	37353105	37358015	q11.23	0.0099	5.8	MAF01 Coding Broad Burden
<i>YWHAB</i>	20	43514317	43537173	q13.12	0.0049	1.5	MAF05 Regulatory Broad Burden
<i>KCNB1</i>	20	47980414	48099184	q13.13	0.0077	19	MAF01 Coding Broad Burden
<i>PHACTR3</i>	20	58152564	58422766	q13.32	0.0087	1.9	MAF05 Coding Broad SKAT
<i>EEF1A2</i>	20	62119366	62130505	q13.33	0.00047	0.047	MAF01 Regulatory Broad Burden
<i>ZBTB21</i>	21	43406940	43430496	q22.3	0.0067	0.15	MAF01 Coding Broad Burden
<i>UBASH3A</i>	21	43824008	43867791	q22.3	0.0029	1.6	MAF01 Coding Broad SKAT
<i>MYO18B</i>	22	26138111	26427007	q12.1	0.0079	1.4	MAF01 Coding Broad SKAT
<i>PDXP</i>	22	38054734	38062941	q13.1	0.0062	0.81	MAF05 Coding Broad SKAT
<i>MCHR1</i>	22	41074754	41078818	q13.2	0.0022	0.30	MAF01 Coding Broad SKAT
<i>MEI1</i>	22	42095503	42195460	q13.2	0.0092	2.3	MAF01 Coding Broad SKAT
<i>PACSIN2</i>	22	43231418	43411151	q13.2	0.0047	1.5	MAF01 Regulatory Broad SKAT
<i>PNPLA3</i>	22	44319619	44360368	q13.31	0.00055	2.8	MAF01 Coding Broad SKAT
<i>HDAC10</i>	22	50683612	50689834	q13.33	0.0093	7.8	MAF01 Coding Broad Burden
<i>PASD1</i>	X	150732094	150845211	q28	0.0035	0.42	MAF05 Coding Broad Burden
<i>LCA10</i>	X	153146127	153154444	q28	0.0038	0.14	MAF05 Coding Broad Burden

Table S7: P-values are corrected for the covariates of sex, platform, and the first 5 PCA components but are not corrected for multiple testing.



Table S8 Top Pathway Contributing Variants

**Table S8: Top Variants Contributing to the Limonene and Pinene Degradation Pathway Result**

SNP ID	hg19 Location	P-Value	Odds Ratio	Gene	Position/Function	Attempter MAF	Non-Attempter MAF
rs61737992	chr17:19575096	0.013	3.3	<i>ALDH3A2</i>	Non-Synonymous (P→S)	0.014	0.0032
rs61757684	chr5:125880710	0.033	3.4	<i>ALDH7A1</i>	Non-Synonymous (T→A)	0.012	0.0032
rs556650006	chr5:125930919	0.057	12	<i>ALDH7A1</i>	Non-Synonymous (L→P)	0.0028	0.0
rs199916968	chr3:184910322	0.086	9.6	<i>EHHADH</i>	Non-Synonymous (R→C)	0.0026	0.0
rs147086207	chr12:112221070	0.11	8.6	<i>ALDH2</i>	Non-Synonymous (D→H)	0.0026	0.0

Table S9 Top Pathway Contributing Genes

**Table S9: MAF01 Coding Broad Gene Burden Results Contributing to the Limonene and Pinene Degradation Pathway Result**

Gene	Chromosome	hg19 Start	hg19 Stop	P-value	Odds Ratio	Attempter Collapsed Frequency	Non-Attempter Collapsed Frequency
<i>ALDH2</i>	12	38390661	38400658	0.0012	10	0.023	0.0016
<i>ALDH3A2</i>	17	19549449	19582911	0.011	3.4	0.031	0.0095
<i>ALDH7A1</i>	5	125875533	125933110	0.039	2.3	0.041	0.017
<i>ALDH9A1</i>	1	165629453	165670100	0.066	0.24	0.0026	0.013
<i>ECHS1</i>	10	135173984	135189193	0.12	4.0	0.010	0.0016
<i>EHHADH</i>	3	184906412	185001778	0.14	1.8	0.034	0.019
<i>ALDH1B1</i>	9	38390661	38400658	0.20	1.6	0.041	0.025
<i>NAT6</i>	3	50331833	50338852	0.26	0.39	0.0031	0.012
<i>YOD1</i>	1	207215194	207228325	0.30	2.6	0.0078	0.0016
<i>HADHA</i>	2	26411504	26469594	0.91	1.0	0.031	0.030

Collapsed frequencies refer to the frequency of individuals with at least one variant meeting the variant set criteria across the complete gene locus; P-values and odds ratios corrected for sex, platform, and the first 5 principal components for each subject.  $p < 1.0 \times 10^{-6}$  (conservative) or  $p < 2.5 \times 10^{-6}$  (liberal) required for significance via Bonferroni correction.

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