1	The interaction of anti-inflammatory and anti-tumor
2	components in the traditional Chinese medicine Solanum
3	<i>lyratum</i> Thunb
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24 Abstract

25 Solanum lyratum Thunb is a traditional Chinese medicinal with a significant clinical 26 outcome for tumor treatment; however, chemicals or fractions separated from the herb 27 did not exhibit strong and comparable efficacy. To investigate the potential synergy or 28 antagonism among chemicals in the extract, we obtained the compounds solavetivone 29 (SO), tigogenin (TI), and friedelin (FR) from the herb. The anti-tumor effects of these 30 three monomer compounds alone or in combination with the anti-inflammatory 31 compound DRG were also tested in this study. SO, FR, and TI used alone did not inhibit the proliferation of A549 and HepG2 cells, but the combination of the three 32 33 achieved 40% inhibition. In vitro anti-inflammatory analysis showed that DRG had a 34 stronger anti-inflammatory effect than TS at the same concentration, and the 35 combination of DRG with SO, FR, or TI inhibited the anti-tumor effect of DRG. This is the first study that documented the synergistic and antagonistic interactions 36 between different compounds in a single herb. 37

38 Key words: Solanum lyratum Thunb; anti-tumor; anti-inflammation; synergism;
39 antagonism.

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41 Experimental

42 Plant materials

43 The herb of Solanum lyratum Thunb was used in this study as herbal material. 44 The traditional Chinese medicine Solanum lyratum Thunb was purchased from Hubei Tianji traditional Chinese medicine decoction Co., Ltd. and identified as Solanum 45 lyratum Thunb of Solanaceae by Professor Xuebo Hu of the Institute of medicinal 46 47 plants of Huazhong Agricultural University, Wuhan China. The sample was stored in 48 the laboratory of natural medicine and molecular engineering of Huazhong University. The voucher specimen number was 20170202001. The reference specimen is WUK 49 0198806 from plant plus of China. 50

51 Extraction and isolation

52 The air-dried whole plant of S. lyratum (10.0 kg) was cut and extracted three times (1 h each time) with refluxing EtOH (80 L each time). Evaporation of the 53 solvent under reduced pressure provided the ethanolic extract (1.88 kg), which was 54 dissolved and suspended in $H_2O(5 L)$ and then partitioned with $CH_2Cl_2(5\times 5 L)$ and 55 56 EtOAc (5 \times 5 L). The CH₂Cl₂ fraction (250.0 g) was subjected initially to silica gel column (120×15 cm) chromatography (100-200 mesh, 2.5 kg) and eluted with 57 petroleum ether-ethyl acetate at 100:0, 95:5, 90:10, 85:15, 82:20, 75:25, 60:40, 50:50, 58 46:60, 30:70, 20:80, and 0:100 to produce 38 fractions, which were numbered 59 60 DA.1-38. Fraction DA.9 (5 g) was separated over silica gel (200-300 mesh) and eluted by petroleum ether-ethyl acetate (20:1, 15:1, 10:1, and 0:1) to obtain DA.9 61 (1-5). Fraction DA.9-5 (1.3 g) was separated by CC over silica gel (200-300 mesh) 62 and eluted by petroleum ether-ethyl acetate (30:1, 20:1, 10:1, and 0:1) to obtain 63 64 compound 2 (20 mg). Fraction DA.12 (3.4 g) was separated by CC over silica gel 65 (200-300 mesh) and eluted by petroleum ether-ethyl acetate (6:1, 4:1, 2:1, and 0:1) to yield DA.12 (1-3). Fractions DA.12-2 and 12-3 was separated by Sephadex LH-20 66 (40 g, eluted with DCM-EtOH, 50:50, v/v) to produce compound 3 (30 mg). We 67 mixed fraction DA.5-6 (9.2 g) and eluted by petroleum ether-ethyl acetate (100:0, 68 69 80:1, 50:1, 20:1, 10:1, 5:1, and 0:100) to obtain compound 14 (15 mg).

70 Extract total saponin and alkaloids

71 The air-dried whole plant of S. lyratum (500 g) was powdered and soaked in 95% 72 ethanol overnight and extracted three times with refluxing EtOH at 60 °C. The solvent was filtered and ethanol was removed by a rotary evaporator to give the crude ethanol 73 74 extract. was We dissolved and suspended the extract in purified water, added HCl (5%) 75 until the pH was 3, stirred well, and let stand overnight to precipitate insoluble acid components. After filtration and removal of insoluble matter, petroleum ether was 76 77 added three times to remove fat-soluble impurities. We modulated the acid solution 78 was with ammonium hydroxide until the pH was 10, extracted with dichloromethane 5-10 times until the organic phase had no obvious color, and removed 79 80 dichloromethane at 45°C to obtain fat-soluble alkaloids. The alkaline aqueous solution 81 was extracted with n-butanol and n-butanol was recovered under decompression to 82 obtain water-soluble alkaloids.

For total saponin extraction, the air-dried whole plant of *S. lyratum* (100 g) was soaked in 95% ethanol overnight and extracted three times (30 min each time) using an ultrasonic machine (60 °C, 90 W). Evaporation of the solvent under reduced pressure provided the ethanolic extract. The extract was dissolved and suspended in H_2O and partitioned with petroleum ether first and then with n-butanol. Finally, total saponin was obtained after evaporating n-butanol under reduced pressure.

89 HPLC analysis of DRG content in total saponin

90 The of content 91 diosgenin3-O- α -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosiduronic acid (DRG) in the total saponins, which is a steroidal saponin that was extracted in our previous 92 research (to be published), was determined by high performance liquid 93 94 chromatography (HPLC). DRG was dissolved and diluted into a series of solutions 95 that covered the concentrations from 25 ppm to 100 ppm. Total saponins powder (100 mg) was dissolved in 1 ml methanol and configured into a 100 mg/ml sample solution. 96 97 The separation was done on a ZORBAX SB-C18 column (4.6 mm×250 mm, 5 micron) with a C_{18} guard column. The column temperature was set at 40 °C with a detection 98 99 wavelength of 210 nm and an injection volume of 10 µL. Two eluents were used for 100 the mobile phase. Eluent A was methanol and eluent B was water. The program of

101 eluent A was $10\% (0 \text{ min}) \rightarrow 30\% (10 \text{ min}) \rightarrow 40\% (25 \text{ min}) \rightarrow 60\% (35 \text{ min}) \rightarrow 80\%$

102 $(40 \text{ min}) \rightarrow 100\%$ (65 min). The flow rate was 1.0 mL/min.

103 Cell culture

HepG2 and A549 cells were purchased from the American Type Culture 104 Collection (Manassas, VA, USA). The cells were cultured separately in 25 cm² culture 105 flasks in DMEM medium (GibcoTM, Invitrogen Corp. USA) that were supplemented 106 with 5% FBS, 100 U/ml penicillin, and 100 µg/mL streptomycin at 37 °C in an 107 atmosphere of 95% humidity and 5% CO2. HMEC-1 cells were cultured in 108 MCDB131 medium that included 10% FBS, 1 µg/ml hydrocortisone, 10 ng/ml 109 110 recombinant human epidermal growth factor, 100 U/ml penicillin, and 1 µg/mL streptomycin at 37°C in 5% CO₂. 111

Determination of cell viability

The viability of cells after treatment with various chemicals was evaluated using 113 MTT assay. Cells were plated at a density of 2×10^4 /well into 96-well plates with 114 DMEM medium. Subsequent to 24 h incubation, the cells were treated with different 115 116 concentrations of compounds alone or in combination. Twenty-four hours after 117 treatment, the drug-containing medium was removed and replaced by MTT solution, and the cells were cultured at 37°C in 5% CO₂ for 4 h. MTT solution was then 118 removed and 150 µl DMSO was added to each well. The optical density was then 119 120 determined at 570 nm and 630 nm with a microplate reader.

121 Determination of the anti-inflammatory activity of DRG and total saponins

A validation model based on the ICAM-1 signaling pathway has been established in our laboratory (Zhang, et al., 2018). With the same DRG concentration, we used this model to determine the anti-inflammatory effects of total saponins and DRG.

125 Statistical analysis

126 All data were presented as the mean \pm standard deviation. A one-way analysis of 127 variance using GraphPad Prism 6 (GraphPad Software Inc., San Diego, CA, USA) 128 determined whether the results showed statistical significance between groups. The 129 significance level was expressed as * if *P* was <0.05, expressed as ** if *P* was 0.01-130 0.001, and expressed as *** if *P* was <0.001.

Compound 3: Tigogenin, white crystals, $C_{27}H_{44}O_3$ ¹H-NMR (400MHz, CDCl₃): 131 δ_H 0.75 (3H, s, 18-CH₃), 0.77 (3H, d, J=6.3Hz, 27-CH₃), 0.81 (3H, s, 19-CH₃), 0.95 132 (3H, d, J=6.0Hz, 21-CH₃), 3.38-3.40 (1H, m, H-26α), 3.56-3.58 (1H, m, H-26β), 4.41 133 (1H, dd, J=7.2, 7.8Hz, H-16). ¹³C-NMR (150MHz, CDCl₃): 12.37 (C-19), 14.51 134 (C-21), 16.51 (C-18), 17.15 (C-27), 21.08 (C-11), 28.62 (C-6), 28.81 (C-24), 30.31 135 (C-25), 31.38 (C-2), 31.52 (C-23), 31.78 (C-15), 32.26 (C-7), 35.13 (C-8), 35.59 136 (C-10), 36.97 (C-1), 38.21 (C-4), 40.08 (C-12), 40.58 (C-13), 41.62 (C-20), 44.84 137 (C-5), 54.36 (C-9), 56.32 (C-14), 62.19 (C-17), 66.85 (C-26), 71.31 (C-3), 80.86 138 (C-16), 109.26 (C-22). 139

Compound 2: Solavetivone, C₁₅H₂₂O, yellow oily liquid, EI-MS: m/z 219 [M+H]⁺. ¹H-NMR (400MHz, CDCl₃): $\delta_{\rm H}$ 5.69 (1H, brs, H-1), 2.15 (1H, dd, J =16.7, 4.4Hz, H-3α), 2.59 (1H, dd, J=16.7, 4.4Hz, H-3β), δ 4.69 (1H, s, H-12α), 4.67(1H, s, H-12β), 1.70 (3H, brs, H-13), 1.88 (3H, brs, H-14), 0.93 (3H, d, J=6.9Hz, H-15). ¹³C-NMR (150MHz, CDCl₃): 15.9 (C-15), 20.92 (C-13), 21.29 (C-14), 32.77 (C-8), 34.4 (C-9), 39.27 (C-4), 40.83 (C-6), 42.99 (C-3), 46.57 (C-7), 50.12 (C-5), 109.4 (C-12), 125.52 (C-1), 147.16 (C-11), 166.64 (C-10), 199.12 (C-2).

147 Compound 1: Friedelin, $C_{30}H_{50}O$, white crystals; EI-MS: m/z 427 [M+H]⁺. ¹H-NMR (400MHz, CDCl₃): $\delta_{\rm H}$ 0.73 (3H, s), 0.87 (3H, s), 0.88 (3H, d), 0.95 (3H, s), 148 1.00 (3H, s), 1.01 (3H, s), 1.05 (3H, s), 1.18 (3H, s), 2.39 (1H, dd, J = 13.4, 3.7 Hz), 149 2.30 (1H, dd, J =13.4, 5.9Hz), 2.25 (1H, q). ¹³C-NMR (150MHz, CDCl₃): 6.84 (C-23), 150 151 14.67 (C-24), 17.96 (C-25), 18.25 (C-7), 18.68 (C-27), 20.27 (C-26), 22.29 (C-1), 28.18 (C-20), 30 (C-17), 30.51 (C-12), 31.79 (C-30), 32.1 (C-28), 32.43 (C-15), 32.77 152 (C-21), 35.04 (C-29), 35.35 (C-19), 35.63 (C-11), 36.01 (C-16), 37.45 (C-9), 38.3 153 (C-14), 39.26 (C-22), 39.71 (C-13), 41.29 (C-6), 41.54 (C-2), 42.16 (C-5), 42.79 154 (C-18), 53.11 (C-8), 58.23 (C-4), 59.48 (C-10), 213.27 (C-3). 155

156 Figures

Fig S1. Effect of whole plant extract (WP), total saponin (TS), water-soluble alkaloid
(WA), and hydrophobic alkaloid (HA) from *Solanum lyratum* Thunb. on the viability

159 of HMEC-1 cells. Asterisks denote (** p < 0.01 and *** p < 0.001, n=6) statistically

- significant differences between the treatment groups and the blank control; nsindicates no significant difference.
- 162 **Fig S2**. Effect of the monomeric compounds separated from *Solanum lyratum* Thunb.
- 163 on the viability of HepG2 and A549 cells. Asterisks denote (* p < 0.05 and *** p < 0.05
- 164 0.001, n=6) statistically significant differences between the treatment groups and the
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- 166 **Fig S3**. Combinatorial action of solavetivone, tigogenin, and friedelin on the viability
- 167 of HepG2 and A549 cells. Asterisks denote (* p < 0.05, ** p < 0.01 and *** p < 0.001,
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- 169 control; ns indicates no significant difference.
- 170 Fig S4. A) Inflammation index of DRG and TS. B) Anti-tumor activity analysis of
- 171 DRG on the proliferation of HepG2 cells after drug exposure for 24h. C) Anti-tumor
- activity analysis of DRG alone or combined with SO, TI, or FR on HepG2 cells.
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