

**Simultaneous determination of thiacloprid and its five metabolites in vegetables
and flowers via QuEChERS combined with HPLC-MS/MS**

Shaodong Pan^a, Xianpeng Song^a, Dan Wang^a, Yajie Ma^a, Xiangliang Ren^{a,b}, Hongyan Hu^{a,b}, Yongpan Shan^a, Xiaoyan Ma^{a,b}, Junyu Luo^{a,b}, Changcai Wu^{a,b,*}, Yan Ma^{a,b,*}, Jinjie Cui^{a,b,*},

^a State Key Laboratory of Cotton Biology, Institute of Cotton Research, Chinese Academy of Agricultural Sciences, Anyang, Henan, 455000, China;

^b Zhengzhou Research Base, State Key Laboratory of Cotton Biology, School of Agricultural Sciences, Zhengzhou University, 450001 Zhengzhou, China;

^c College of Horticulture, Anhui Agriculture University, Hefei City, China;

* Correspondence: Changcai Wu, Yan Ma, and Jinjie Cui

E-mail: ccwu00@163.com; aymayan@126.com; cuijinjie@126.com

Table of Contents:

Fig.

Fig. S1. Typical HPLC-MS/MS MRM chromatograms of thiacloprid and its five metabolites of (A) blank cowpea sample, (B) blank cucumber sample, (C) blank muskmelon sample, (D) blank tomato sample, (E) blank cotton flower sample, (F) blank chrysanthemum flower sample, (G) blank mixed flower sample, (a) spiked cowpea samples ($100 \mu\text{g kg}^{-1}$), (b) spiked cucumber sample ($100 \mu\text{g kg}^{-1}$), (c) spiked muskmelon sample ($100 \mu\text{g kg}^{-1}$), (d) spiked tomato sample ($100 \mu\text{g kg}^{-1}$), (e) spiked cotton flower sample ($100 \mu\text{g kg}^{-1}$), (f) spiked chrysanthemum flower sample ($100 \mu\text{g kg}^{-1}$), (g) spiked mixed flower sample ($100 \mu\text{g kg}^{-1}$).

Fig. S2. Effects of six different sorbent kits on extraction recovery of thiacloprid and its five metabolites in different flowers.

Table.

Table S1. The MS basic instrument parameters setting. Declustering Potential (DP), Entrance Potential (EP), Collision Cell Exit Potential (CPX) and Collision Energy (CE). All expressed in V.

Table S2. Calibration curve, linearity, matrix effect and LOQ of thiacloprid and its metabolites.

Table S3. Average recoveries ($n = 15$, %), RSD_F (%), and RSD_R (%) for target compounds from different flower matrices at three spiked levels.

Table S4. The detection result of thiacloprid and its metabolites in flower samples collected from the farm.

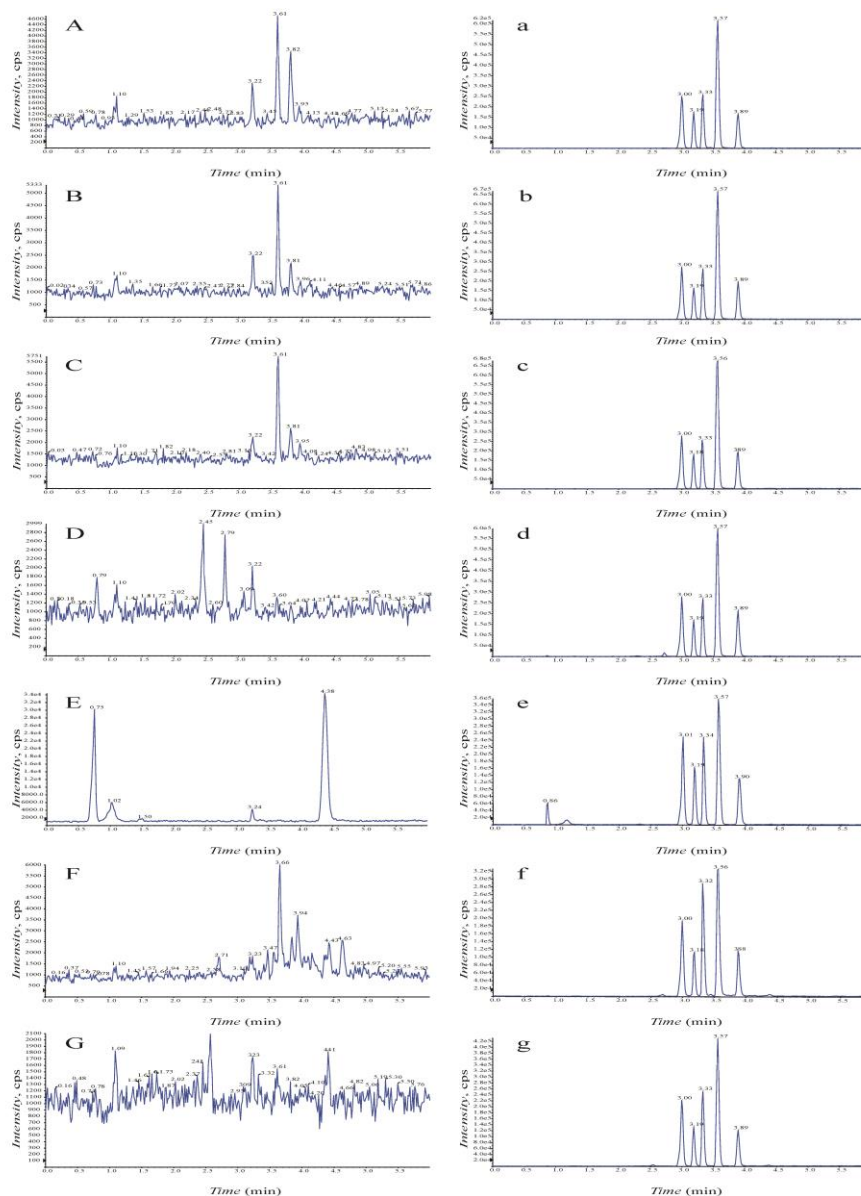


Fig. S1. Typical HPLC-MS/MS MRM chromatograms of thiacloprid and its six metabolites of (A) blank cowpea sample, (B) blank cucumber sample, (C) blank muskmelon sample, (D) blank tomato sample, (E) blank cotton flower sample, (F) blank chrysanthemum flower sample, (G) blank mixed wildflower sample, (a) spiked cowpea samples ($100 \mu\text{g kg}^{-1}$), (b) spiked cucumber sample ($100 \mu\text{g kg}^{-1}$), (c) spiked muskmelon sample ($100 \mu\text{g kg}^{-1}$), (d) spiked tomato sample ($100 \mu\text{g kg}^{-1}$), (e) spiked cotton flower sample ($100 \mu\text{g kg}^{-1}$), (f) spiked chrysanthemum flower sample ($100 \mu\text{g kg}^{-1}$), (g) spiked mixed wildflower sample ($100 \mu\text{g kg}^{-1}$).

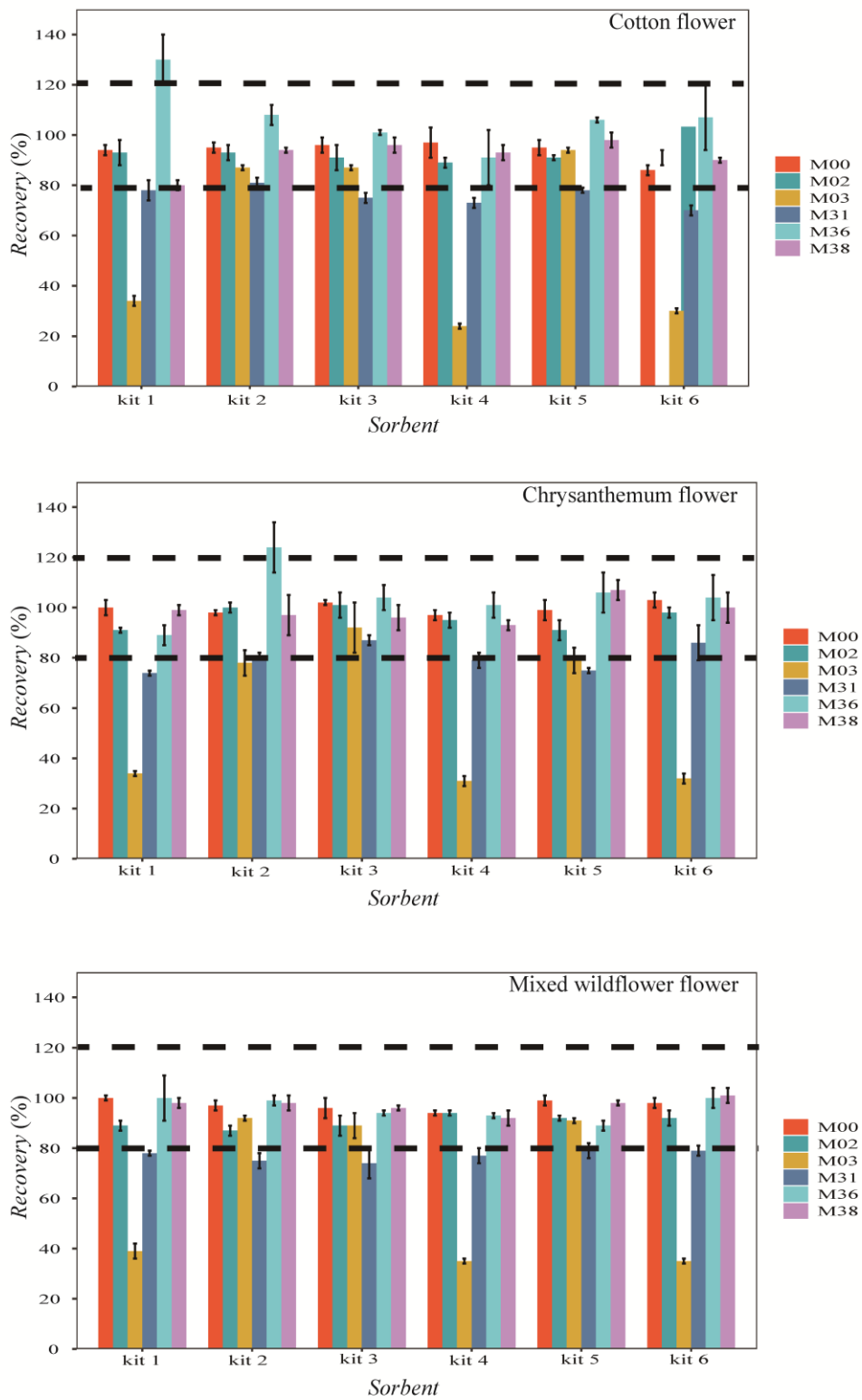


Fig. S2. Effects of six different sorbent kits on extraction recovery of thiacloprid and its five metabolites in different flowers

Table S1. The MS basic instrument parameters setting

Instrumental parameters	ESI mode	
	ESI+	ESI-
ion spray voltage (V)	5500	-4500
ESI source temperature (°C)	500	500
Ion Source Gas 1 (GS 1)	50	50
Ion Source Gas 2 (GS 2)	50	50
Curtain Gas (CUR)	40	40
collision gas (CAD)	9	9
DP	-40	-40
EP	-10	-10
CE	38.0	-15
CXP	6.0	-11

Declustering Potential (DP), Entrance Potential (EP), Collision Cell Exit Potential (CPX), and Collision Energy (CE). All expressed in Volt (V).

Table S2. Calibration curve, linearity, matrix effect and LOQ of thiacloprid and its metabolites

Compound	Matrix	Regression equation	R ²	ME	LOQ
M00	Acetonitrile	$y = 5613.15638x + 3677.10769$	0.99895	–	–
	cowpea	$y = 4672.98788x + 36377.2$	0.99935	0.83	2
	cucumber	$y = 4860.71864x + 37825.5$	0.99753	0.87	2
	muskmelon	$y = 5233.85370x + 48319.3$	0.99883	0.93	2
	tomato	$y = 4892.50419x + 7724.99921$	0.99902	0.87	2
	cotton flower	$y = 2767.74035x + 3061.67211$	0.99965	0.49	2
	chrysanthemum flower	$y = 2918.89839x + 2685.13282$	0.99998	0.52	2
	mixed flower sample	$y = 3372.72101x + 5528.79500$	0.99983	0.60	2
	M02	Acetonitrile	$y = 2620.43615x - 1038.31920$	0.99669	–
cowpea		$y = 2709.45896x + 3094.92423$	0.99895	1.03	2
cucumber		$y = 2626.04274x + 2310.73125$	0.99989	1.00	2
muskmelon		$y = 2665.25264x + 1880.85410$	0.99993	1.02	2
tomato		$y = 2636.73450x + 2267.78149$	0.99977	1.01	2
cotton flower		$y = 2381.46472x + 1605.46503$	0.99976	0.91	2
chrysanthemum flower		$y = 2829.76786x + 2167.59524$	0.99991	1.08	2
mixed flower		$y = 2588.53848x + 2263.95870$	0.99946	0.99	2

	sample				
M03	Acetonitrile	$y = 2525.57993x + 5.86330$	0.99992	–	–
	cowpea	$y = 2936.98705x + 40251.1$	0.9986	1.16	2
	cucumber	$y = 3013.12483x + 16259.74807$	0.99832	1.19	2
	muskmelon	$y = 3270.28509x + 28969.67781$	0.99882	1.29	2
	tomato	$y = 3006.54985x + 6714.58132$	0.99949	1.19	2
	cotton flower	$y = 2162.13846x + 346752$	0.99802	0.86	2
	chrysanthemum flower	$y = 2466.09673x + 8119.31849$	0.99568	0.98	2
	mixed flower sample	$y = 1104.04749x + 2705.83436$	0.99998	0.44	2
M31	Acetonitrile	$y = 4052.30722x - 908.14449$	0.99944	–	–
	cowpea	$y = 4115.59092x + 2143.89003$	0.99767	1.02	2
	cucumber	$y = 3893.04414x + 3573.60829$	0.99955	0.96	2
	muskmelon	$y = 3993.52346x + 1820.16819$	0.99996	0.99	2
	tomato	$y = 4064.50745x + 3996.14392$	0.9991	1.00	2
	cotton flower	$y = 3202.22460x + 4080.79169$	0.99997	0.79	2
	chrysanthemum flower	$y = 3367.92637x + 2380.51881$	0.99972	0.83	2
	mixed flower	$y = 3317.10934x + 3855.96736$	0.9998	0.82	2

	sample				
M36	Acetonitrile	$y = 2518.07115x - 378.84128$	0.99883	–	–
	cowpea	$y = 2362.82302x + 18016.17213$	0.99884	0.82	2
	cucumber	$y = 2359.09446x + 16130.43826$	0.99837	0.99	2
	muskmelon	$y = 2397.23825x + 40465.8$	0.99711	1.15	2
	tomato	$y = 2448.97453x + 17040.45634$	0.9995	0.81	2
	cotton flower	$y = 2135.18631x + 35578.3$	0.99964	0.97	2
	chrysanthemum flower	$y = 1879.95484x + 6423.57558$	0.99943	0.95	2
	mixed flower sample	$y = 2021.37028x + 8946.11665$	0.99967	0.78	2
M38	Acetonitrile	$y = 4896.17489x + 2630.41867$	0.99928	–	–
	cowpea	$y = 4015.12258x + 4907.09713$	0.99897	0.82	2
	cucumber	$y = 4150.65842x + 7673.27139$	0.99719	0.85	2
	muskmelon	$y = 4578.73116x + 6817.59279$	0.9971	0.94	2
	tomato	$y = 4344.34306x + 6621.92223$	0.99807	0.89	2
	cotton flower	$y = 2208.30379x + 1607.18794$	0.99925	0.45	2
	chrysanthemum flower	$y = 2544.04051x + 2367.41175$	0.99979	0.52	2

mixed flower

$$y = 2730.20975x + 4101.70149$$

1

0.56

2

sample

Table S3. Average recoveries (n = 15, %), RSD_T (%), and RSD_R (%) for target compounds from different flower matrices at three spiked levers.

Matrix	Spike level	cotton			chrysanthemum			mixed wildflower		
		Reco	intra-	inter-	Reco	intra-	inter-	Reco	intra-	inter-
comp	($\mu\text{g kg}^{-1}$)	very	RSD	RSD	very	RSD	RSD	very	RSD	RSD
M00	2000	95	6.8	4.8	95	3.8	4.2	100	3.5	7.2
	200	95	5.2	2.8	98	1.8	2.9	100	5.4	8.2
	2	97	10.5	6.5	105	11.0	10.9	93	10.2	4.8
M02	2000	91	2.1	4.1	94	2.6	6.7	96	3.1	3.7
	200	97	2.5	1.1	99	1.5	3.7	90	4.3	1.2
	2	90	4.5	11.8	105	12.0	5.6	102	7.3	7.4
M31	2000	78	3.0	7.1	83	1.6	3.7	85	2.2	6.6
	200	79	6.9	8.8	83	3.0	1.8	80	6.8	7.5
	2	84	5.5	5.9	77	11.8	14.4	81	5.6	5.1
M38	2000	96	3.2	5.4	98	2.4	3.3	102	2.2	8.3
	200	99	5.8	3.8	99	1.6	2.4	100	8.6	9.4
	2	94	9.9	14.1	94	1.0	12.4	94	6.2	12.1
M36	2000	97	3.4	4.0	93	4.5	5.6	100	4.3	4.0
	200	98	4.5	4.2	98	2.3	6.0	95	4.7	4.6
	2	105	11.5	2.9	108	11.7	11.6	119	12.8	9.8
M03	2000	77	5.7	8.9	91	4.2	3.4	88	5.0	10.7
	200	89	13.4	9.1	93	3.6	6.4	94	3.4	7.2
	2	104	4.6	14.0	86	13.1	6.4	97	1.0	3.0

Table S4. The detection result of thiacloprid and its metabolites in flower samples collected from the farm.

Latin name of species (flower samples)	Thiacloprid ($\mu\text{g kg}^{-1}$)	M02 ($\mu\text{g kg}^{-1}$)	M03 ($\mu\text{g kg}^{-1}$)	M31 ($\mu\text{g kg}^{-1}$)	M36 ($\mu\text{g kg}^{-1}$)	M38 ($\mu\text{g kg}^{-1}$)
<i>Polygonum viscosum</i> Buch.-Ham.ex D.Don.	ND	ND	ND	ND	ND	ND
<i>Gerbera jamesonii</i> Bolus	ND	ND	7.3	ND	13.8	ND
<i>Erigeron annuus</i> (Linn.) Pers.	ND	ND	ND	ND	3.3	ND
<i>Punica granatum</i> Linn.	ND	ND	1.8	ND	2.1	ND
<i>Dendranthema indicum</i> (Linn.) Des Moul	ND	ND	ND	ND	ND	ND
<i>Hypericum monogynum</i> Linn.	ND	ND	ND	ND	5.6	ND
<i>Sambucus williamsii</i> Hance	ND	ND	ND	ND	2.1	ND
<i>Hibiscus syriacus</i> Linn.	ND	ND	55.9	ND	3.7	ND
<i>Trifolium repens</i> Linn.	ND	ND	122.4	ND	3.4	ND
<i>Rosa chinensis</i> Jacq.	ND	ND	ND	ND	5.0	ND
<i>Verbena officinalis</i> Linn.	ND	ND	ND	ND	4.8	ND
<i>Daucus carota</i> Linn.	ND	ND	35.1	ND	2.2	ND

<i>Cosmos bipinnata</i> Cav.	ND	ND	4.5	ND	2.6	ND
<i>Campsis grandiflora</i> (Thunb.) Schum.	ND	ND	ND	ND	2.4	ND
<i>Ligustrum lucidum</i> Ait.	ND	ND	ND	ND	2.6	ND
<i>Alternanthera</i> <i>philoxeroides</i> (Mart.) Griseb.	ND	ND	ND	ND	5.2	ND
<i>Helianthus annuus</i> Linn.	ND	ND	ND	ND	ND	ND
<i>Cichorium intybus</i> Linn.	ND	ND	10.6	ND	2.1	ND
<i>Brassica campestris</i> L.	ND	ND	42.5	ND	162.4	ND

All flowers were collected from this farm and their species names are expressed using their Latin names.