Supplementary Materials for

Electrochemically Driven, Ni-Catalyzed Aryl Amination: Scope, Mechanism, and Applications

Yu Kawamata,[1,7] Julien C. Vantourout,[1] David P. Hickey,[2,7] Peng Bai,[3,7] Longrui Chen,[4] Qinglong Hou,[4] Wenhua Qiao,[4] Koushik Barman,[2,7] Martin A. Edwards,[2,7] Alberto F. Garrido-Castro,[1] Justine N. deGruyter,[1] Hugh Nakamura,[1] Kyle Knouse,[1] Chuanguang Qin,[1] Khalyd J. Clay,[1] Denghui Bao,[4] Chao Li,[1] Jeremy T. Starr,[5] Carmen Garcia-Irizarry,[5] Neal Sach,[6] Henry S. White,[2,7] Matthew Neurock,*[3,7] Shelley D. Minteer,*[2,7] Phil S. Baran*[1,7]

[1] Department of Chemistry, The Scripps Research Institute, 10550 North Torrey Pines Road, La Jolla, California 92037, United State

[2]Department of Chemistry, University of Utah, 315 South 1400 East, Salt Lake City, Utah 84112, United States

[3]Department of Chemical Engineering and Materials Science, University of Minnesota, Minneapolis, MN 55455, United States

[4]Asymchem Life Science (Tianjin), Tianjin Economic-Technological Development Zone, Tianjin 300457, China

[5]Discovery Sciences, Medicine Design, Pfizer Global Research and Development, 445 Eastern Point Road, Groton, Connecticut 06340, United States

[6]Department of Chemistry, La Jolla Laboratories, Pfizer, 10770 Science Center Drive, San Diego, CA 92121, USA.

[7]NSF Center for Synthetic Organic Electrochemistry

Correspondence to:
Matthew Neurock: mneurock@umn.edu
Shelley D. Minteer: minteer@chem.utah.edu
Phil S. Baran: pbaran@scripps.edu

Table of Contents

General Methods

Instruments

Reagents were purchased at the highest commercial quality grade and used without further purification, unless otherwise stated. Yields refer to chromatographically and spectroscopically (^{1}H NMR) homogeneous material, unless otherwise stated. Reactions were monitored by thin layer chromatography (TLC) carried out on 0.25 mm E. Merck silica plates (60F-254), using shortwave UV light and $KMnO_4$ for visualization. Flash column chromatography was performed using E. Merck silica gel (60, particle size 0.043 – 0.063 mm). NMR spectra were recorded on Bruker AVIII-600 instruments and Bruker AV400 for ^{19}F were calibrated using residual undeuterated solvent as an internal reference ($CDCl_3$: 7.26 ppm ^{1}H NMR, 77.16 ppm ^{13}C NMR. The following abbreviations were used to explain NMR peak multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, m = multiplet, br = broad. High–resolution mass spectra (HRMS) were recorded on an Agilent LCMS TOF mass spectrometer using electrospray ionization time-of-flight (ESI-TOF) reflectron experiments. The enantiomeric excesses were determined with Waters UPC^2 SFC equipped with a photodiode array detector or an Agilent Technologies 1220 Infinity II LC HPLC. Optical rotations were recorded on a Rudolph Research Analytical Autopol III Automatic Polarimeter.

Electrode materials and dimensions

The RVC electrodes were purchased from commercial RVC block (purchased from ULTRAMET, 80 ppi, 14.40” x 13.86” x 8”) and Ni foam electrodes were furnished from Nickel Foam (1.5 mm x 100 mm x 250 mm for Battery, Electric Capacity etc. purchased from eBay).

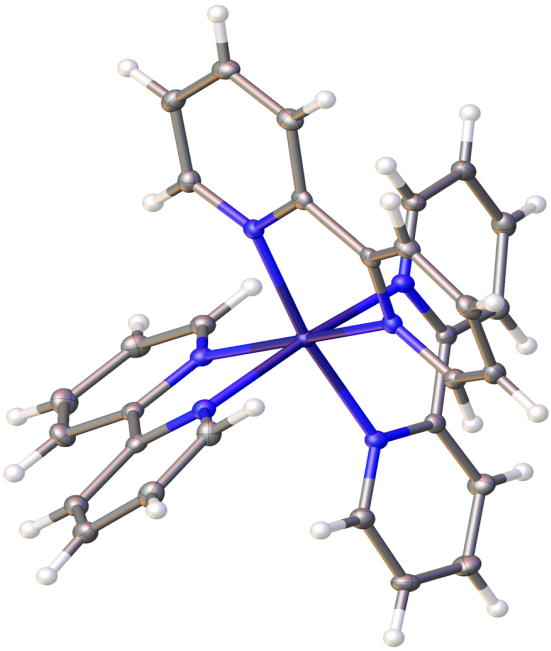
For experiments using 5 mL IKA ElectraSyn vial, the dimensions of the RVC anode were 47 mm x 8 mm x 2 mm; the dimensions of Ni foam cathode were 0.8 cm × 4.7 × 0.1 cm (the submerged exterior surface areas of the anode and cathode were approximately 0.8 cm × 1.5 cm on 0.1 mmol scale). For experiments on larger scales, dimensions of electrodes have been specified in the relevant experimental section.

$Ni(bpy)_3Br_2$ catalyst

Preparation

To a 1 L round bottom flask was added a saturated solution of 2,2'-bipyridine (450 mmol) in MeOH (100 mL) followed by the addition of a solution of $NiBr_2 \bullet 3H_2O$ (150 mmol) in MeOH (100 mL). The mixture immediately turned red and a pink precipitate was formed (*note: the reaction is exothermic*). After cooling to ambient temperature, acetone (150 mL) was added to induce more precipitation and the pink solid was filtered. This solid was used as catalyst without further purification (88 g, 87% yield). Aldrich catalog number ALD00608.

X-ray crystallographic data of compound 17



The structure of $[Ni(bpy)_3]Br_2$ and $[Ni(bpy)_3](ClO_4)_2$ were determined by single crystal X-ray diffraction. It was confirmed that nickel ion is ligated by three bpy ligands in both structures. However, due to the disorder of bromide counter aninon, structural refinement was not completely successful. Accordingly, the graphic and cif file of $[Ni(bpy)_3](ClO_4)_2$ were provided for the publication.

Empirical formula	C30 H24 Cl2 N6 Ni O8	
Molecular formula	C30 H24 N6 Ni, 2(Cl O4)	
Formula weight	726.16	
Temperature	100.0 K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	C 1 2/c 1	
Unit cell dimensions	a = 17.0095(9) Å	α= 90°.
	b = 10.7266(6) Å	β= 91.4250(10)°.
	c = 15.9498(9) Å	γ = 90°.
Volume	2909.2(3) $Å^3$	
Z	4	
Density (calculated)	1.658 Mg/m^3	
Absorption coefficient	0.916 mm^{-1}	
F(000)	1488	
Crystal size	0.125 x 0.1 x 0.09 mm^3	
Crystal color, habit	pink block	
Theta range for data collection	2.245 to 27.102°.	
Index ranges	-16<=h<=21, -13<=k<=13, -20<=l<=20	
Reflections collected	10973	
Independent reflections	3212 [R(int) = 0.0371]	
Completeness to theta = 25.242°	100.0 %	
Absorption correction	Semi-empirical from equivalents	

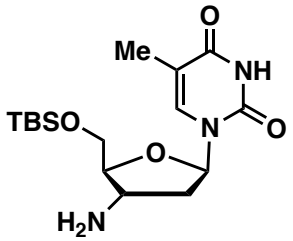
Max. and min. transmission	0.6468 and 0.5945
Refinement method	Full-matrix least-squares on F^2
Data / restraints / parameters	3212 / 0 / 213
Goodness-of-fit on F^2	1.052
Final R indices [I>2sigma(I)]	R1 = 0.0283, wR2 = 0.0711
R indices (all data)	R1 = 0.0322, wR2 = 0.0737
Largest diff. peak and hole	0.381 and -0.437 e.$Å^{-3}$

Table 2. Atomic coordinates (x 10^4) and equivalent isotropic displacement parameters ($Å^2$x 10^3)

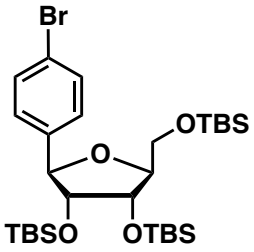
	x	y	z	U(eq)
Ni(1)	5000	5319(1)	2500	9(1)
N(1)	4396(1)	3905(1)	1859(1)	11(1)
N(2)	3993(1)	5138(1)	3216(1)	11(1)
N(3)	5438(1)	6800(1)	3192(1)	11(1)
C(1)	4617(1)	3347(2)	1148(1)	13(1)
C(2)	4223(1)	2336(2)	802(1)	15(1)
C(3)	3569(1)	1885(2)	1206(1)	17(1)
C(4)	3335(1)	2445(2)	1941(1)	15(1)
C(5)	3764(1)	3456(2)	2254(1)	11(1)
C(6)	3559(1)	4115(2)	3036(1)	11(1)
C(7)	2970(1)	3704(2)	3560(1)	14(1)
C(8)	2822(1)	4379(2)	4281(1)	15(1)
C(9)	3238(1)	5467(2)	4445(1)	16(1)
C(10)	3818(1)	5813(2)	3897(1)	14(1)
C(11)	5816(1)	6715(2)	3942(1)	13(1)
C(12)	5937(1)	7735(2)	4458(1)	14(1)
C(13)	5665(1)	8890(2)	4191(1)	15(1)
C(14)	5300(1)	8999(2)	3405(1)	14(1)
C(15)	5199(1)	7931(2)	2920(1)	11(1)
Cl(1)	6763(1)	9937(1)	1374(1)	14(1)

O(1)	6649(1)	9248(1)	2130(1)	26(1)
O(2)	6177(1)	10907(1)	1292(1)	27(1)
O(3)	7536(1)	10499(1)	1393(1)	24(1)
O(4)	6691(1)	9114(1)	660(1)	29(1)

Substrate Synthesis and Characterization



3-amino-thymidine derivative was prepared following a known procedure.[1]



1-(4-bromophenyl)-ribose derivative was prepared following a known procedure.[2]

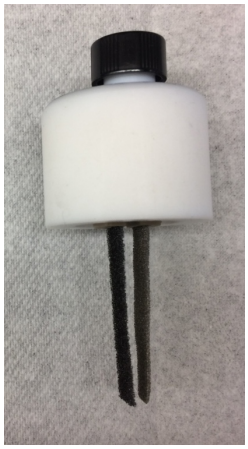
General and Graphical Procedure

General procedure for small scale (0.05-0.2 mmol) reactions

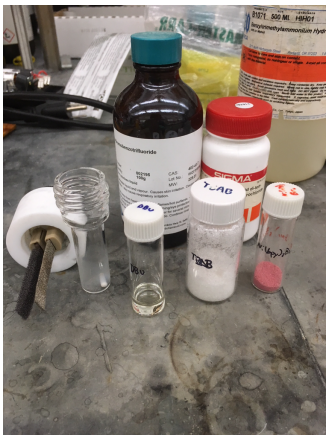
An ElectraSyn vial (5 mL) with a stir bar was charged with $Ni(bpy)_3Br_2$ (10 mol%), TBAB (0.4 mmol, 0.2 M), aryl halide, amine, DBU and DMA (2.0 mL). [Liquid compounds were added after the addition of DMA. The order of the addition does not affect the result. The amount of aryl halide, amine and DBU in individual case are indicated in the characterization section] The ElectraSyn vial cap equipped with anode (RVC) and cathode (Ni foam) were inserted into the mixture (See below for the graphical guide. If electrode area submerged into the solution is less than 1.0 cm, add more solvent.). The vial was then evacuated and backfilled with an argon balloon. This cycle was repeated twice. The reaction mixture was electrolyzed under a constant current of 4 mA until complete consumption of the starting material as judged by TLC. After the reaction, the ElectraSyn vial cap was removed and electrodes were rinsed with a mixture of organic solvents (EtOAc:hexanes = 1:1), which was combined with the crude mixture. Aqueous sat. NH_4Cl was then added to the combined solutions; the resulting solution was extracted with a mixture of organic solvents (EtOAc:hexanes = 1:1). The combined organic layers were dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The crude material was purified by column chromatography or preparative thin-layer chromatography (PTLC) to furnish the desired product.

Graphical guide

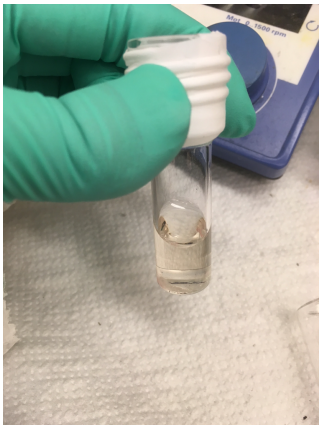
Photos were taken from the coupling between 4-bromobenzotrifluoride and glutamic acid di-tert-butyl ester.

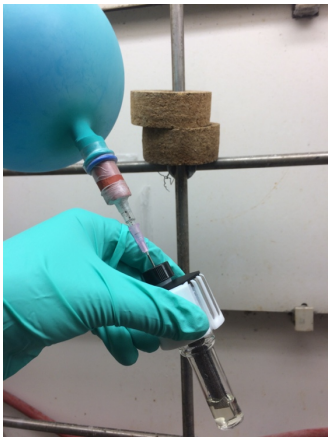



(Left) ElectraSyn 2.0. ***(Middle)*** ElectraSyn 2.0 vial (5 mL). ***(Right)*** ElectraSyn 2.0 cap equipped with RVC (left side) and Nickel electrodes (right side).

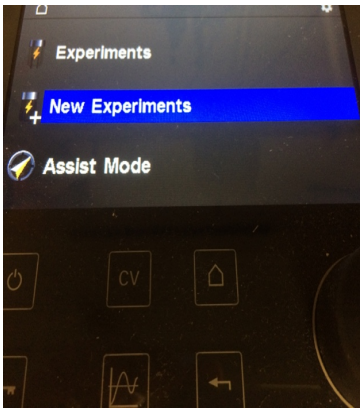


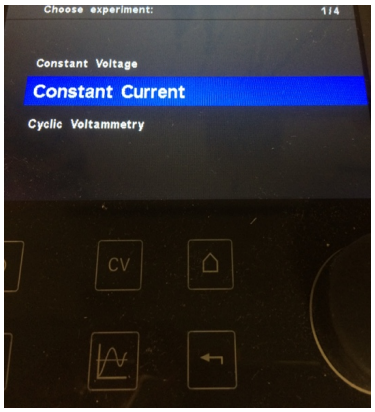
(Left) Materials used in the reaction. ***(Middle)*** Wrapping Teflon tape helps to keep a good sealing during a reaction. ***(Right)*** The appearance of the mixture after the addition of all solid components.



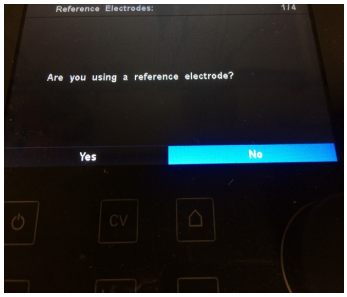


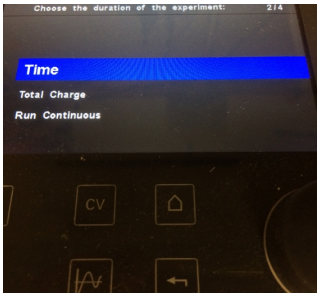
(Left) After the addition of DMA. ***(Middle)*** The cap was tightly screwed into the vial. Ensure > 1cm of the electrodes are submerged into the solution**.** If not, please add more solvent. ***(Right)*** The reaction vessel was connected to a vacuum line through a needle. The reaction vessel was backfilled with an argon balloon [***Right*** was repeated for three cycles].

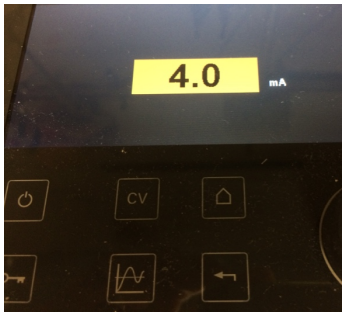




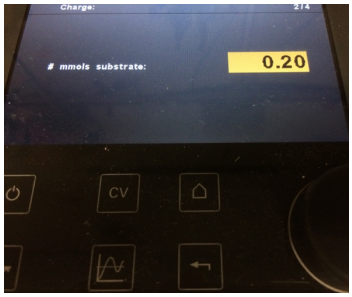
(Left) The electrochemical cell was plugged into ElectraSyn 2.0. ***(Middle)*** Select New experiments. ***(Right)*** Select Constant Current.

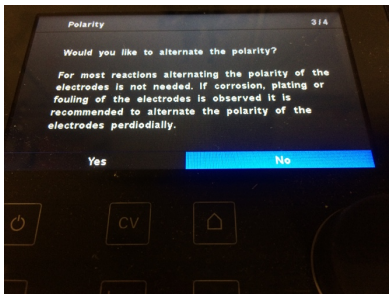






(Left) No use of reference electrode. ***(Middle)*** Select Time. ***(Right)*** Adjust the current value.





(Left) Define the “time”. ***(Middle)*** Indicate the “mmol” of the substrate. ***(Right)*** No alternate polarity.

Peptide Synthesis, general amination procedure and purification

Methods for peptide synthesis

HPLC analysis were conducted on a Waters Autopurification LC with a Waters XBridge C18 column (4.6x150 mm, 3.5 ⌈m). Fractionation was triggered by a Waters QDa single quadruple mass spec in ESI^+ single ion or ESI^- single ions recoding modes. UV detection was monitored at 261 nm.

Solvent A: 0.1 M aqueous triethylammonium acetate Solvent B: acetonitrile
1.5 mL/min, 25 °C.
Gradient: 5–90% B over 11 minutes

Preparative HPLC were conducted on the same instrument as above and were based on the HPLC analysis using the methods describe below:

RT: Method: %B:
0-2 min Narrow 5-20 %B
2-4 min Narrow 1 10-25 %B
4-5 min Narrow 2 15-35 %B
5-6 min Narrow 3 25-45 %B
6-7 min Narrow 4 35-55 %B
7-8 min Narrow 5 45-65 %B
8-9 min Narrow 6 55-75 %B
9-11 min Narrow 7 65-95 %B

Materials

Commercial materials were used as received unless otherwise noted. Amino acids and coupling reagents were obtained from Novabiochem or Combi-blocks. 2-CTC resin was purchased from Chem Impex (1.0 – 2.0 mmol/g). Solid-phase reaction vessels and pressure caps were purchased from Torviq.

Solid phase peptide synthesis

General procedure. 2-CTC resin

2-CTC resin (1.0 equiv., substitution = 1.0 – 2.0 mmol/g) was swollen in dry DCM for 30 min then washed with DCM (5 x 3 mL) and DMF (5 x 3 mL). A solution of the Fmoc-AA-OH (2.0 equiv.) and *N*,*N*-diisopropylethylamine (DIPEA, 4.0 equiv.) in DMF (final concentration 0.1 M) was added to the resin (1.0 equiv.) and agitated at room temperature. After 16 h, the resin was washed with DMF (5 x 3 mL), DCM (5 x 3 mL), and DMF (5 x 3 mL). A capping step was performed as described below and the resin-bound residue was submitted to iterative peptide assembly (Fmoc-SPPS).

The loading efficiency was evaluated through treatment of the resin with 20% piperidine/DMF (3 mL, 2 × 3 min) to deprotect the Fmoc group. The combined deprotection solutions were diluted to 10 mL with 20% piperidine/DMF. An aliquot of this mixture (50 μL) was diluted 200-fold with 20% piperidine/DMF and the UV absorbance of the piperidine-fulvene adduct was measured (λ = 301 nm, ε = 7800 M^{-1} cm^{-1}) to quantify the amount of amino acid loaded onto the resin. The theoretical maximum for the reported yields of all isolated peptides are based on the numerical value obtained from the resin loading.

General iterative peptide assembly (Fmoc-SPPS)

Peptides were elongated using iterative Fmoc-solid-phase peptide synthesis (Fmoc-SPPS), according to the following general protocols:

Deprotection: The resin was treated with 20% piperidine/DMF (3 mL, 2 x 3 min) and washed with DMF (5 x 3 mL), DCM (5 x 3 mL) and DMF (5 x 3 mL).

General amino acid coupling: A preactivated solution of protected amino acid (4 equiv.), PyBOP (4 equiv.), and *N*-methylmorpholine (NMM) (8 equiv.) in DMF (final concentration 0.1 M) was added to the resin. After 1 h, the resin was washed with DMF (5 x 3 mL), DCM (5 x 3 mL) and DMF (5 x 3 mL).

Capping: Acetic anhydride/pyridine (1:9 v/v) was added to the resin (3 mL). After 3 min the resin was washed with DMF (5 x 3 mL), DCM (5 x 3 mL) and DMF (5 x 3 mL).

Cleavage: A mixture of DCM and HFIP (95:5 v/v) was added to the resin. After 2 h, the resin was washed with DCM (3 x 2 mL).

Work-up: The combined cleavage solution and DCM washes were concentrated under a stream of nitrogen. The residue was treated with cold Et_2O to precipitate the crude peptide, which was subsequently dissolved

in water/acetonitrile containing 0.1% TFA, filtered and used crude in the next esterification or amidation steps except otherwise stated.

Esterification: The crude peptide was dissolved in MeOH (0.25 M) and was titrated with TMS diazomethane solution until yellow color remains. Solvent was removed under *vacuo* and the residue was treated with cold Et_2O to precipitate the crude peptide, which was subsequently dissolved in water/acetonitrile, filtered and purified by reverse-phase HPLC.

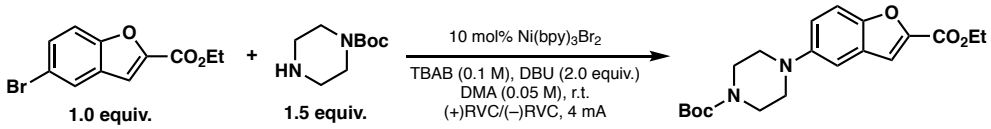
General procedure for the coupling of side chain protected peptides – 0.05 mmol scale.

An ElectraSyn vial (5 mL) with a stir bar was charged with $Ni(bpy)_3Br_2$ (0.3 or 1 equiv.), LiBr (4 equiv.), side chain protected *N*-acetylated methyl ester/acid or amide peptide (1 equiv.), amine (3 equiv.), and DMA (2.0 mL). [Liquid compounds were added after the addition of DMA. The order of the addition does not affect the result]. The ElectraSyn vial cap equipped with anode (RVC) and cathode (Ni foam) was inserted into the mixture. The vial was then evacuated and backfilled with an argon balloon. This cycle was repeated twice. The reaction mixture was electrolyzed under a constant current of 4 mA for 6 hours. After the reaction, the ElectraSyn vial cap was removed and electrodes were rinsed with the mixture of organic solvents (EtOAc:hexane = 2:1), which was combined with the crude mixture. Aqueous sat. NH_4Cl was then added to the combined solution; the resulting solution was extracted with the mixture of organic solvents (EtOAc:hexane = 2:1). The combined organic layer was dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The crude material was purified by preparative HPLC to furnish the desired product.

Mechanistic investigation (Figure 2)

Kinetic experiments

Method

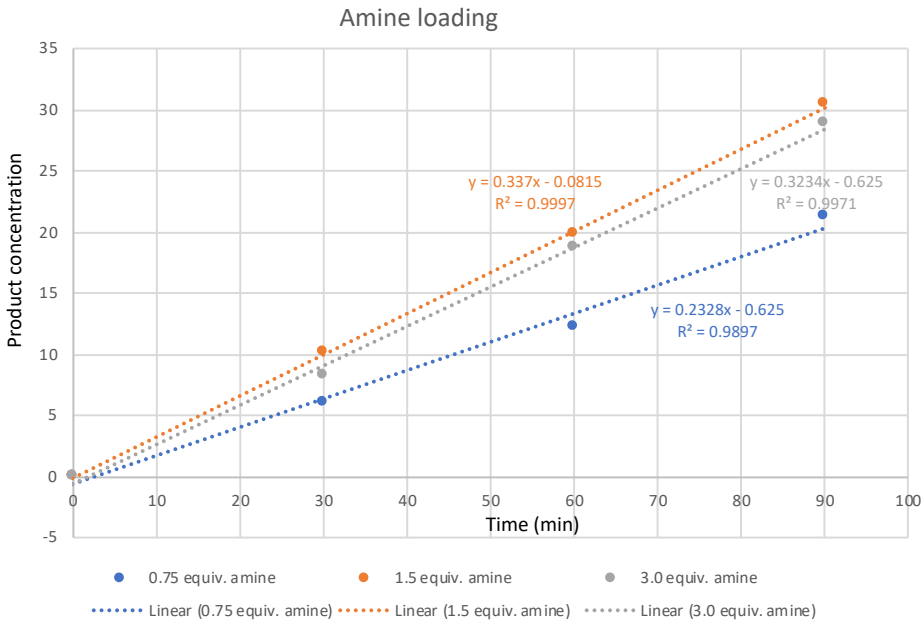


Kinetic experiments were carried out using LCMS analysis following the general e-amination procedure described above. Aliquots were taken at different times (please see graphics for more details) and the conversion was measured against an internal standard (4,4-di-*tert*-butylbiphenyl).

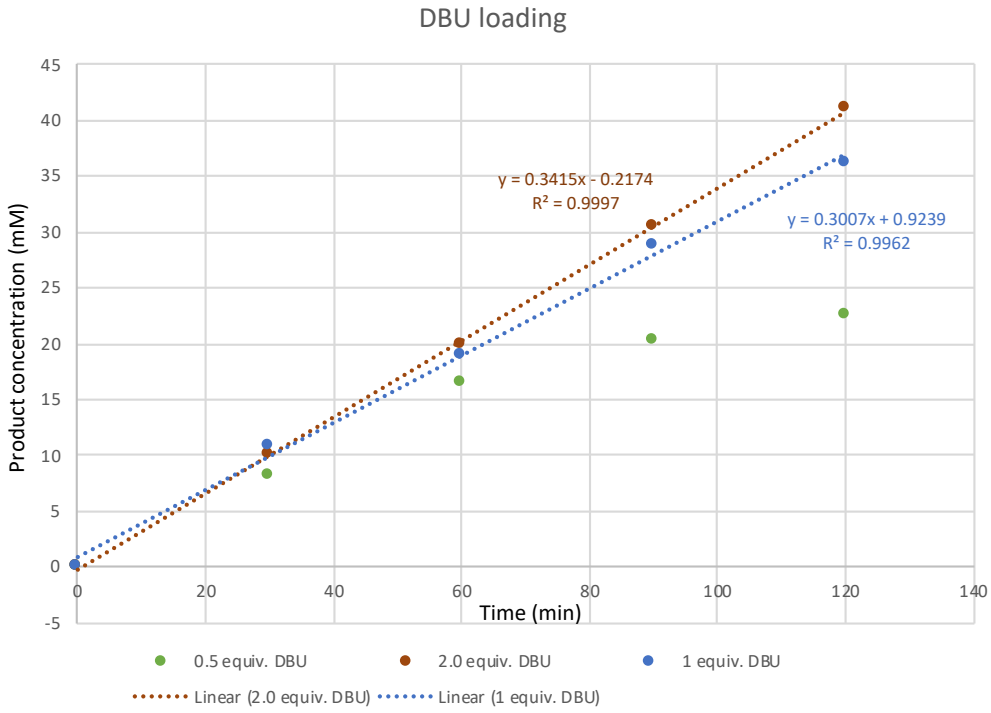
Results

The product formation was monitored by varying one of the parameters under the above conditions. The results showed that the rate of product formation was largely independent of the loading of amine, DBU and Ni catalyst, whereas current value (mA) had large effect on the rate of product formation. This indicates that, if all chemical steps are fast enough, the rate determining step is electron transfer between an electrode and an intermediate. However, the slower rate with 0.75 equiv. anime indicates that the rate determining step in this case is likely a chemical step, rather than electron transfer step. Under the conditions with 0.75 equiv. amine, the rate increase was observed with higher Ni loading.

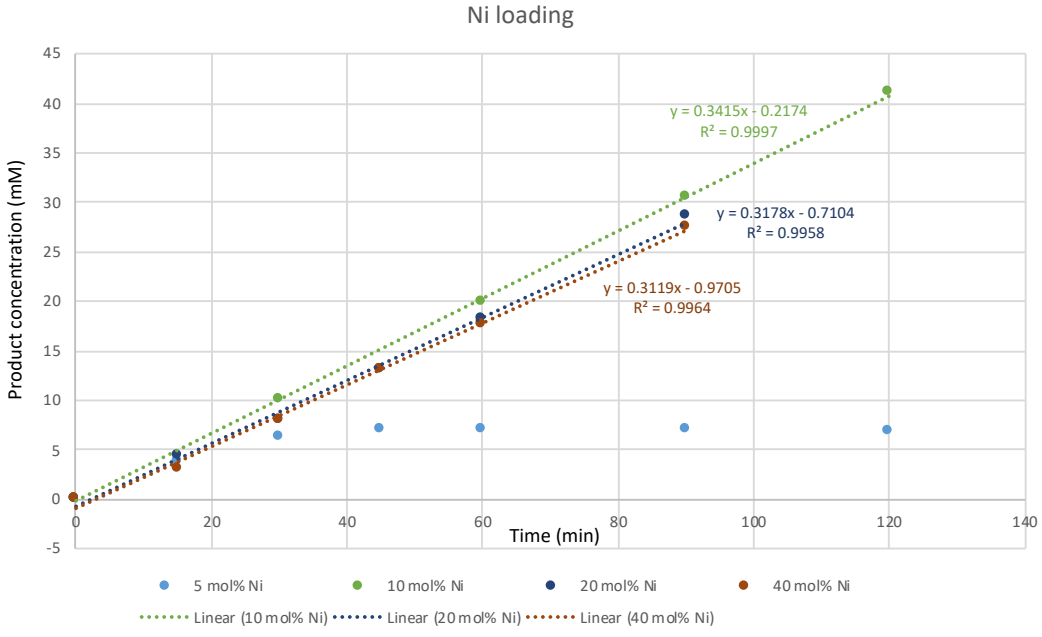
Amine loading



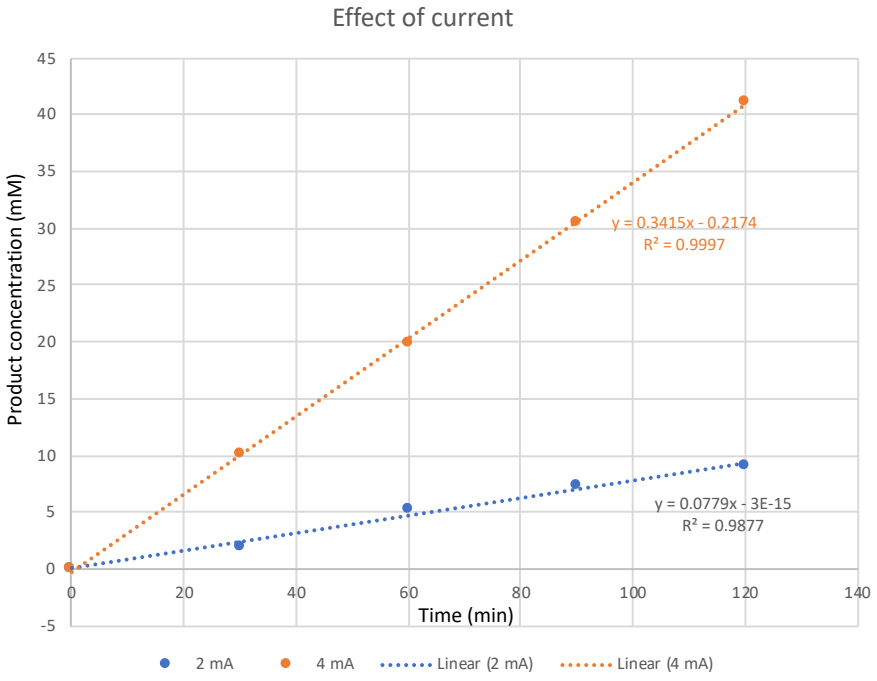
Base loading



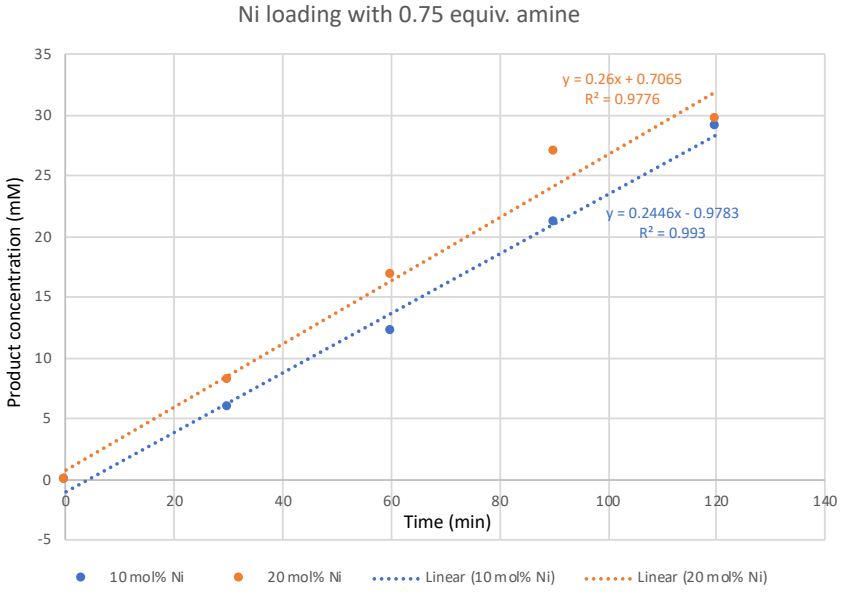
Nickel catalyst loading



Effect of current



Effect of Nickel catalyst loading at low concentration of amine



UV-Vis Analysis

Method

Spectrophotometric experiments were carried out using a Thermo Scientific Evolution 260Bio UV-Vis Spectrophotometer. Absorbance was monitored between 450 and 750 nm with a path length of 1 cm using a solution of 100 mM NBu_4Br in dry DMF as a background solution. Background-subtracted spectra were obtained for solutions of 1 mM $NiBr_2•3H_2O$ in the presence of 0, 0.25, 0.5, 0.75, 1, 1.5, 2, 2.5, 3, 3.5, and 4 mM 4,4'-dimethyl-2,2'-bipyridine.

Electrochemical Analysis

Method

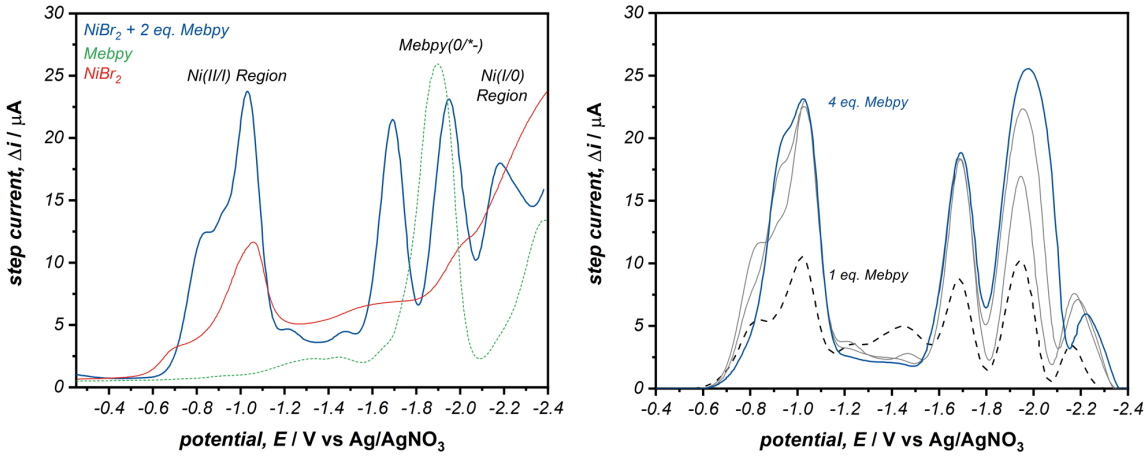
Electrochemical experiments for mechanistic analysis were carried out using a Biologic SP-150 potentiostat using a 3-electrode cell with a 3 mm glassy carbon working electrode, a platinum mesh counter electrode, and a $Ag/AgNO_3$ (10 mM) reference electrode unless otherwise noted. Cyclic voltammetry and square wave voltammetry experiments were performed using1 mM $NiBr_2•3H_2O$ as a Ni(II) species and 4,4'-dimethyl-2,2'-bipyridine (Mebpy) as a ligand with 100 mM NBu_4Br in dry DMF as a supporting electrolyte/solvent system under an atmosphere of Ar with 3.2% H_2 and less than 3 ppm O_2. All glassware for electrochemical analysis were flame dried prior to use. Unless otherwise noted, SWVs were performed from positive to negative potential with an amplitude of 20 mV, frequency of 40 Hz, and a step potential of 5 mV. Throughout the manuscript, positive current corresponds to reduction while negative current corresponds to oxidation. Electrochemical oxidative addition experiments were performed using solutions containing 1 mM $NiBr_2•3H_2O$ with 1 mM Mebpy, where CVs were run at 100 mV s^{-1} in the absence of any electrophile, then again in the presence of 30 mM p-bromoanisole ($MeOC_6H_4Br$). Electrochemical experiments to determine the effect of amine on the oxidation window of Ni(II)($MeOC_6H_4$)(Mebpy)Br were carried out using 1 mM Ni complex in the absence and presence of 100 mM hexylamine at 100 mV s^{-1} and 20 mV s^{-1}. For experiments used to determine the diffusion coefficient and number of electrons transferred; prior to electrochemical measurements, the Pt microelectrode was cleaned by polishing in 50 nm alumina slurry and rinsed with acetone. Electrochemical data were recorded with a CH Instruments 760E bipotentiostat (1 kHz sampling frequency) in a two-electrode configuration, employing the 25 μm diameter Pt microdisk as the working electrode and a leakless Ag/AgCl electrode (eDAQ) as the reference/counter electrode. The entire electrochemical cell was inside a home-built Faraday cage.

Results

Voltammetric Analysis of $Ni(Mebpy)_nBr_2$ Complexes

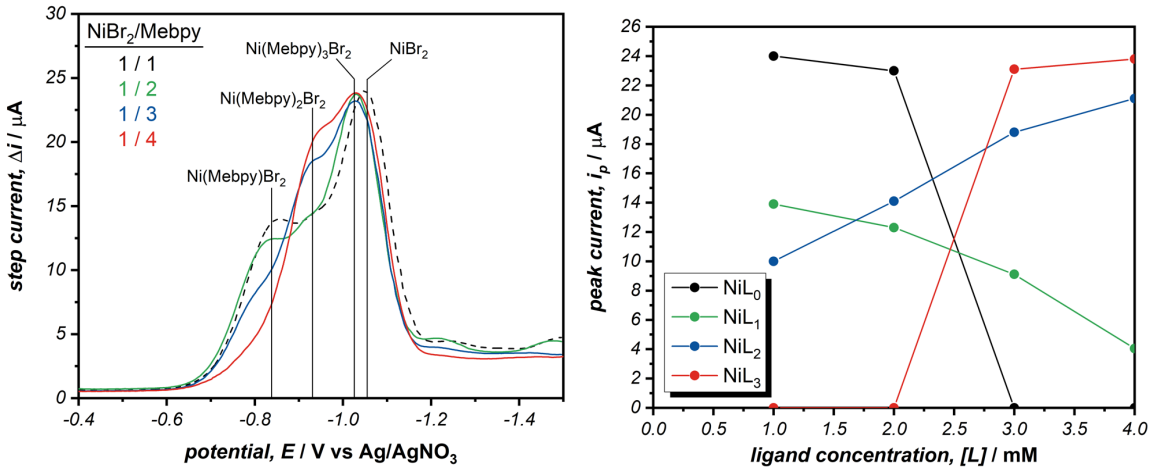
Cyclic voltammetry (CV) and square wave voltammetry (SWV) were used to characterize the reversibility and ligation state of each Ni(II) complex. SWVs of $NiBr_2$ in the presence of various concentrations of Mebpy reveal a complex series of peaks that are generally clustered around -1 V, -1.7 V and -1.9 V.

Comparing this with SWVs of $NiBr_2$ and Mebpy individually, suggests that the peaks around -1 V are consistent with a Ni(II/I) redox couple, while the peak at -1.9 V matches closely the authentic peak for Mebpy reduction, leaving the peak at -1.7 V as likely to resulting from a second electrochemical reduction of the $Ni(Mebpy)_nBr_2$ complex. Upon application of -2.1 V to a solution of 1 mM $NiBr_2$, Ni(0) could be observed precipitating out of solution (coincides with increased reductive current at an onset potential of -1.95 V, below).



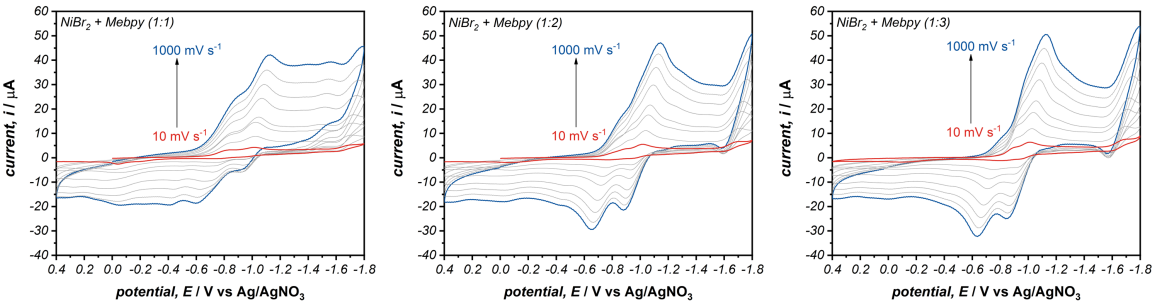
Representative SWVs of ***(left)*** 1mM $NiBr_2 \cdot 3H_2O$ (–), 1 mM Mebpy (---), and $NiBr_2$/Mebpy 1:2 (–); or ***(right)*** 1 mM $NiBr_2 \cdot 3H_2O$ in the presence of 1, 2, 3, or 4 equivalents of Mebpy.

The peak step current (i_p) for a SWV corresponds to the concentration of a species in solution (directly comparable assuming equivalent diffusion coefficients and number of electrons transferred). Therefore, i_p was measured as a function of Mebpy concentration to generate a profile of ligation states for $Ni(Mebpy)_nBr_2$. The resulting profiles were then correlated to spectrophotometric data (primary text) to assign the reduction potentials of $Ni(Mebpy)Br_2$, $Ni(Mebpy)_2Br_2$, and $Ni(Mebpy)_3Br_2$ as -0.82 ± 0.02 V, -0.94 ± 0.02 V, and -1.03 ± 0.01 V respectively.



(*Left*) Representative SWVs 1 mM $NiBr_2$•$3H_2O$ in the presence of 1 (---), 2 (–), 3 (–), or 4 (–) equivalents of Mebpy, and **(*right*)** profiles of the peak step current (i_p) for the four observed redox features corresponding to various ligation states of $Ni(Mebpy)_nBr_2$.

Cyclic voltammetry of $NiBr_2$ in the presence of either one, two or three equivalents of Mebpy (below) reveal the variably reversible nature of each ligation state. The loss of reversibility for Ni(II) species with one equivalent of Mebpy is largely attributed to the formation of insoluble Ni(0) species at the edge of the CV window. While isolation of peaks is complicated slightly by their overlapping nature, peak currents for all observed peaks are linear with square root of the corresponding scan rate. This suggests that adsorbed species are not playing a significant role in the catalytic cycle. Increased concentration of Mebpy alters the shape and number of reductive peaks without dramatically altering the profile of corresponding oxidative peaks. The primary oxidative features include a pair of peaks at -0.65 V and -0.85 V that most closely correspond to the reductive peaks of $Ni(Mebpy)Br_2$ and $Ni(Mebpy)_3Br_2$ (potentials determined by SWV and correlated by peak separation, where E_{ipc}-E_{ipa} approaches 81 mV for the peaks assigned to $Ni(Mebpy)Br_2$ and 64 mV for the peaks assigned to $Ni(Mebpy)_3Br_2$). The precise determination of ligation states is experimentally challenging; however, electrochemical analysis suggests that ligation state is dynamic and dependent on the oxidation state of Ni.



Representative CVs at variable scan rates of 1 mM $NiBr_2 \bullet 3H_2O$ in the presence of either 1 mM ***(left)***, 2 mM ***(center)*** or 3 mM ***(right)*** Mebypy. CVs were performed at scan rates of 10, 50, 100, 200, 400, 600, 800, and 1000 mV s^{-1}.

Previous studies have employed microelectrode analysis to suggest that the initial electrochemical reduction of bpy-ligated Ni(II) species is a two-electron transfer resulting in Ni(0).[3] Similar analysis of $Ni(Mebpy)_nBr_2$ under synthetically relevant conditions using a two-step process. First, the diffusion coefficient was calculated from a temporal decay of the current at a microelectrode upon the application of a potential step. Next, the number of electrons transferred was calculated from the steady-state diffusion-limited current at the electrode, through rearrangement of the analytical expression for this current, which includes the diffusion coefficient, determined in the first step.

Determination of Diffusion coefficient (D)

The diffusion coefficient of the $Ni(Mebpy)_nBr_2$ complex in DMF (D) was calculated from the current response of a microelectrode to a potential step, as described previously.[4] Briefly, a 25 µm diameter Pt microdisk electrode (CH Instruments) was initially held at a potential of -0.4 V vs Ag/AgCl in a DMF solution containing the $Ni(Mebpy)_nBr_2$ complex with excess supporting electrolyte (0.1 M TBAB). At this potential, no oxidation or reduction of the complex occurs (Fig. 1, left). The potential was then stepped to -1.25 V vs Ag/AgCl, a potential sufficient to reduce the complex at a diffusion-limited rate, while the current response, i_d, was recorded (Fig 1, right).

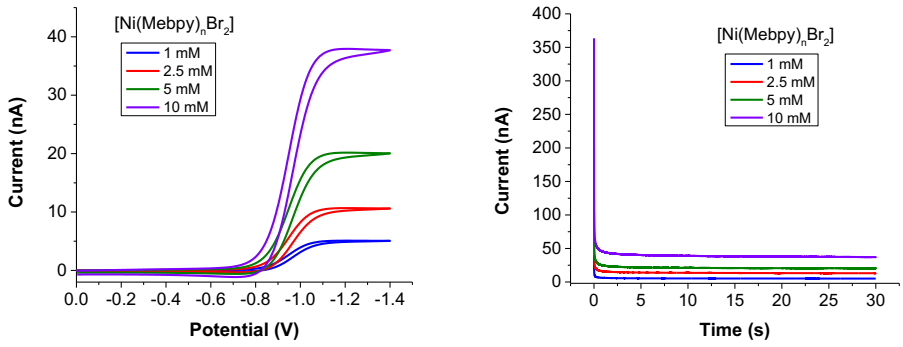


Figure 1. Cyclic voltammetry ***(left)*** and chronoamperometry ***(right)*** of the $Ni(Mebpy)_nBr_2$ complex (concentrations as given in legend) at a 25 µm diameter disk Pt microelectrode. The solution contained 0.1 M TBAB in DMF, and the concentration of 4,4′ -dimethyl-2,2′ -dipyridyl was selected to be at 2 equivalents w.r.t. Ni. In the chronoamperometric experiment, the potential was stepped from -0.4 V to -1.25 V vs Ag/AgCl at t = 0 s. For the voltammetry experiment, the scan rate is 20 mV s^{-1}.

A plot of i_d, normalized by dividing by the steady-state diffusion limited current, i_{ss}, vs $1/\sqrt{t}$ was then made (Figure 2), where i_{ss} was obtained from the limit of the current response at long times (30 s). A fit of a straight line gives a slope (S) that is related to the diffusion coefficient through:

$$D = \pi a^2/16S^2$$

where *a* is the electrode radius. Note, normalizing i_{ss} means this method can be used to determine *D* without knowledge of the number of electrons transferred during the electrochemical process (*n*) or the concentration of redox species (*C*), as noted by their absence in the above equation. From the slope measured from Figure 2, we calculate $D = 0.5(\pm 0.1) \times 10^{-5}$ cm^2 s^{-1}.

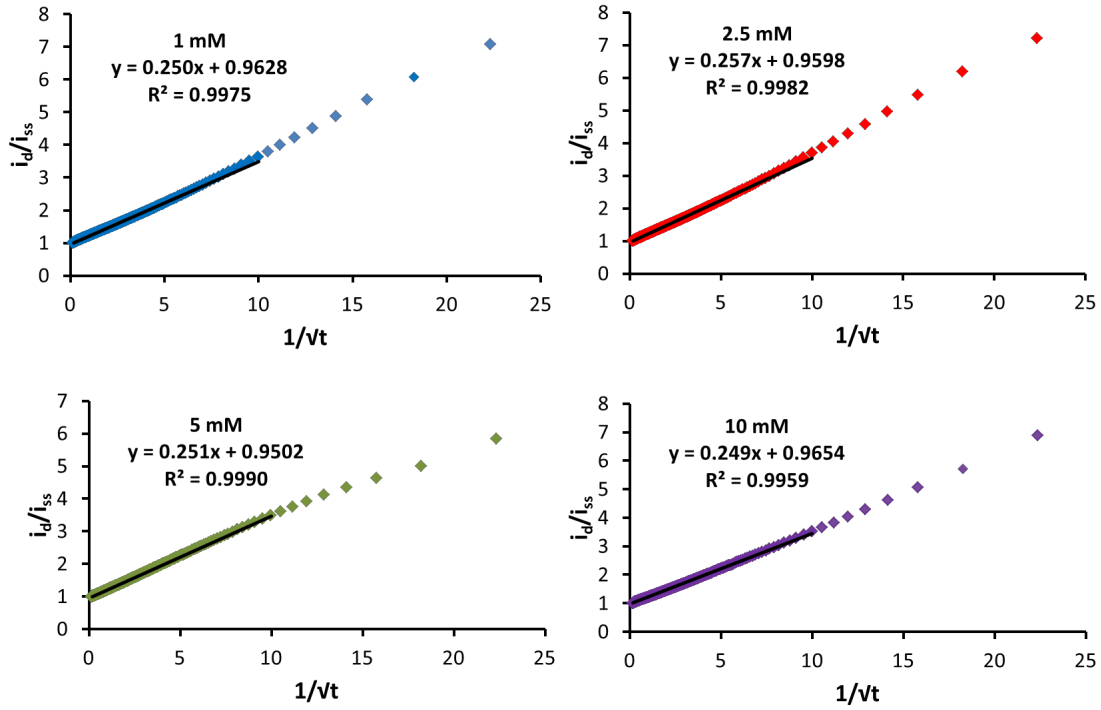


Figure 2. The plot of i_d /i_{ss} *vs* $1/\sqrt{t}$ from the chronoamperometric response for different concentration of the Ni(Mebpy)$_n$Br$_2$ complex. Solid lines show least-squares best fits to the data (''<10.0 s$^{-1/2}$'') to obtain D. Conditions as described in caption of **Figure 1**.

Measurement of the *D* in solutions containing 4 different concentrations of Ni(Mebpy)$_n$Br$_2$, all gave diffusion coefficients that agreed within error.

Number of Electrons Transferred (n)

The steady-state diffusion-limited current at an inlaid micro-disk electrode is described by:[5]

$i_{ss}=4nFCDa$

where F is Faraday's constant and the other variables are as described above. Rearrangement of this equation gives the number of electrons transferred during the reduction as a function of the measured steady-state current, $n = i_{ss}/naFCD$. From i_{ss}, obtained from cyclic voltammetry (Fig. 1, right), and the value of $D = 0.5(\pm0.1) \times 10^{-5}$ cm^2 s^{-1}, we obtain $n = 1.9 \pm 0.2$, i.e., the reduction of the Ni(Mebpy)$_n$Br$_2$ complex is a 2-electron process.

While the precise nature of the Ni oxidation state at the electrode interface remains unclear, several previous studies have demonstrated both that Ni(I) can be generated in appreciable quantities at the electrode surface (e.g. by comproportionation of Ni(0)/Ni(II)),[6] and that oxidative addition to Ni(I)(bpy)$_n$ species is thermodynamically preferential to oxidative addition to Ni(0)(bpy) complexes.[7]

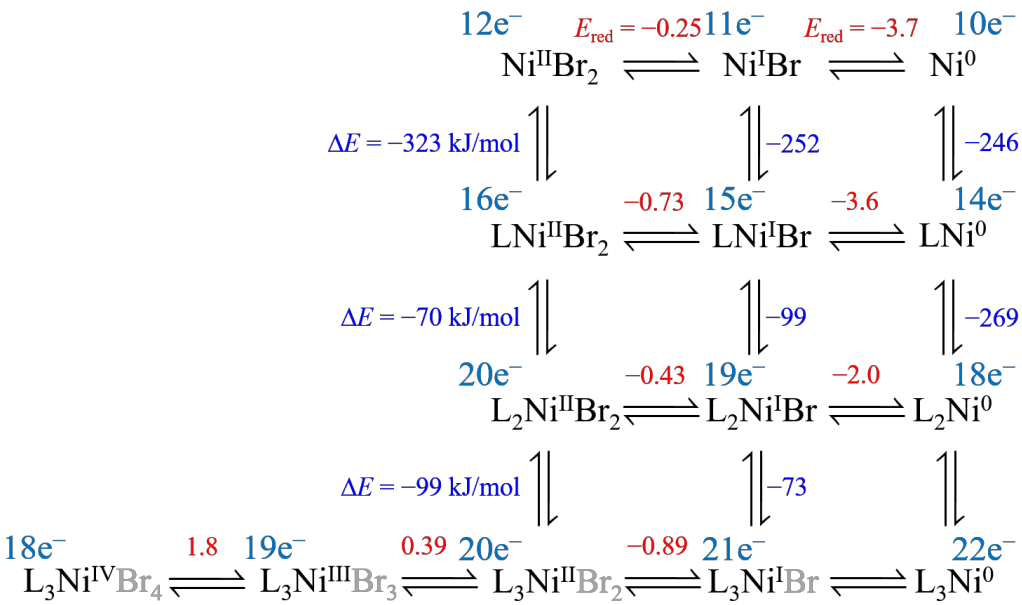
DFT calculations

Computational method

All density-functional theory calculations were performed using the Gaussian 16 software package, revision A.03, with the M06-L exchange-correlation functional, and 6-31+G(d,p) basis set.[8,9] The Stuttgart/Dresden effective core potentials ECP10MDF[10] and ECP28MWB[11] were used on Ni and Br atoms, and solvation effects were taken into account using the SMD model[12] with DMF as the implicit solvent. SCF cycles were converged to 10^{-8} Ha while geometry optimizations were stopped after maximum force, root-mean-square force, maximum displacement, and root-mean-square displacement dropped below 4.5×10^{-4} Ha/a_0, 3.0×10^{-4} Ha/a_0, 1.8×10^{-3} a_0, and 1.2×10^{-3} a_0, respectively. The reported reaction energetics correspond to the spin states with the lowest energies, and the nudged elastic band method[13,14] was used to locate transition states. The experimental solvation free energies of H^+ and Br^- in DMF[15] were used for the calculation of standard reduction potentials (vs. SHE), and the resulting values were then converted to the Ag/AgNO$_3$ (10 mM) reference by subtracting 0.5 V.[16]

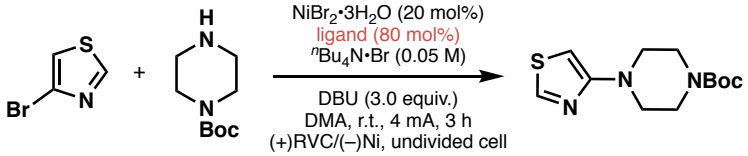
Ligand binding and reduction potential

Calculated ligand binding and reduction potentials in DMF (vs. 10 mM $Ag/AgNO_3$). L = 4,4'-Mebpy.

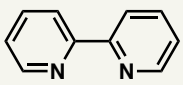


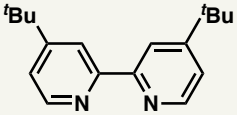
Ligand Screening (Table 1)

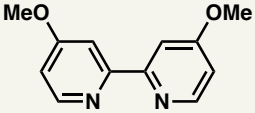
Initial ligand screening for the arylation of 4-bromothiazole

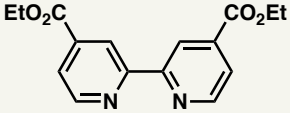


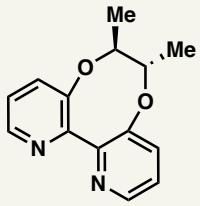
Effective ligands

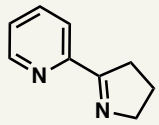

27%

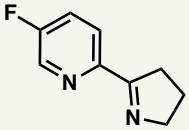

26%

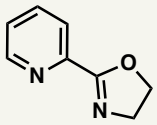

1%

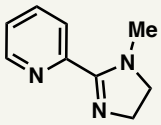

0%

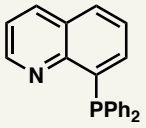

10%


28%

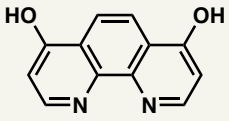

10%

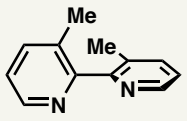

22%

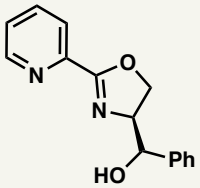

8%

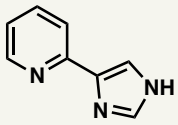

4%

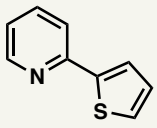
Ineffective ligands (0% yield)

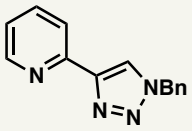


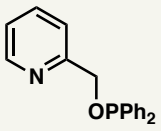


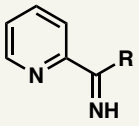


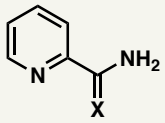




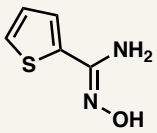


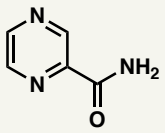


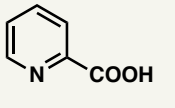


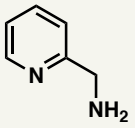


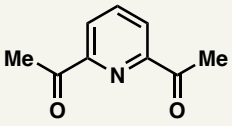
R = H, Me, NH_2 X = O, S

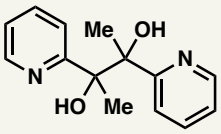


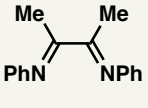


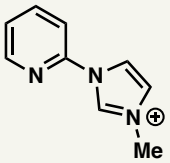




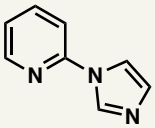


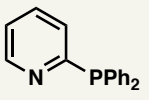


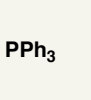


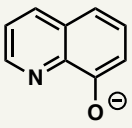


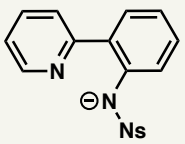
as bidentate NHC

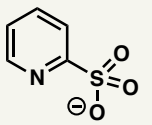


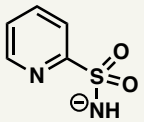


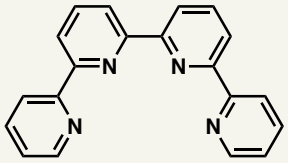

PPh_3



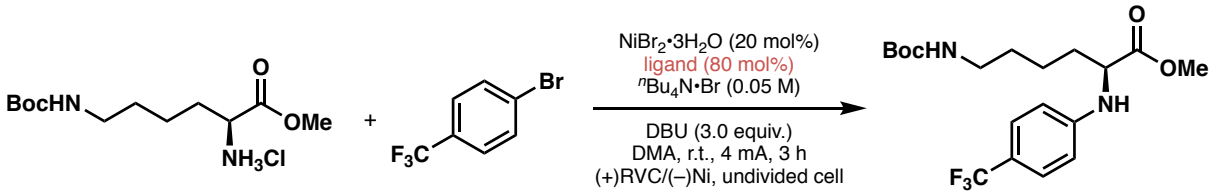


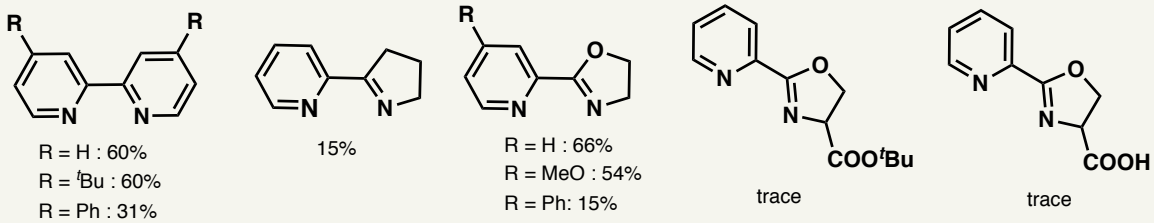


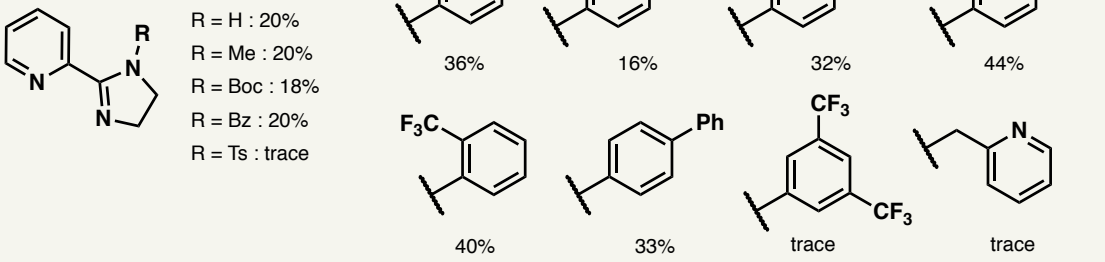


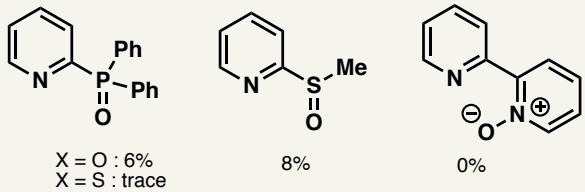


Second ligand screening focusing on electronic/steric tuning of effective types of ligands









Trouble shooting & FAQ

Q. What can I do if the yield of the reaction is not reproducible?

A. Check the voltage of the reaction. From our experiences, if the cell voltage is too high (above 6 V at 4 mA on 0.2 mmol scale without a reference electrode), this reaction does not proceed well. Normal operational voltage range for this reaction is around 2.5V–5V.

When a high voltage is noted:
• Ensure there is sufficient contact between the electrode and the reaction solution. If not, please add more solvent containing 0.2 M TBAB to decrease the resistance in the circuit.
• Increasing the concentration of electrolyte helps to reduce the resistance of the cell.
• Reducing the current value also helps to decrease the reaction potential. Please note that longer reaction time is required in this case to ensure the passage of the same amount of electrical charge.

If the reaction voltage is unlikely to be the cause of irreproducibility, make sure that the reaction is degassed and the cap is screwed tightly. Though the reaction is not extremely sensitive to either oxygen or water, irreproducible results have been observed by insufficient sealing of the reaction cell. One easy way to improve sealing is to wrap Teflon tape around the rim of the vial, as described in Graphical guide.

Q. What are the suitable current values for reactions on different scales?

A. We typically use 4 mA on 0.2 mmol scales with exceptions noted in the procedure of each coupling reaction. The submerged exterior anode area is approximately 1.5 cm × 1 cm; the submerged exterior cathode area is approximately 1.5 cm × 1 cm.

Q. Where can I get the materials to construct the electrochemical cell?

A. Although in this work we used materials that we purchased previously, everything required for setting up this reaction can be obtained from IKA (https://www.ika.com/fr/Produits-Lab-Eq/Electrochemistry-Kit-csp-516/).

Q. Is this reaction sensitive to water?

A. A small amount of water (100–300 mol%) does not significantly affect the outcome.

Q. How air sensitive is the reaction?

A. The reaction is not particularly air sensitive as it proceeds without freeze-pump-thaw. However, evacuation-argon backfill cycle is still required as running the reaction under air results in much lower yield.

Q. How stable is $Ni(bpy)_3Br_2$?

A. This catalyst is indefinitely bench-stable, free-flowing pink powder. In addition, no sign of hygroscopic nature has been observed.

Q. Do other ligands work?

A. Please see the ligand screening section above

Q. Is stirring crucial for this reaction?

A. Stirring is critical—without stirring, the potential of the reaction could increase, leading to low yields. Our preferred stirring rate is from 500 to 1000 rpm.

Q. What is the byproduct of this reaction?

A. The major side reactions were homocoupling and proto-dehalogenation of the aryl halide. Occasionally, phenols (from the oxidation of aryl halides) could be detected, albeit in small quantities.

Q. What can I do if lots of starting materials remain after electrolysis?

A. One can increase the reaction time, use a higher current. If black deposit is observed at the Ni cathode, increasing ligand loading helps to slow down this undesired Ni deposition.

Q. How can I optimize the reaction if lots of homocoupling dimer or dehalogenation byproducts were detected at the end of the reaction?

A. One can change the ratio of amine and aryl halide. Alternatively, one can also increase the amount of DBU for the enhancement of amine nucleophilicity.

Q. Would it affect the yield if the electrolysis is conducted for longer, after the consumption of starting materials?

A. Longer reaction times can lead to the over-oxidation of amination products. Therefore, stopping the reaction after disappearance of starting materials by TLC/GC is recommended.

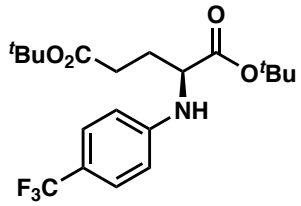
Q. What are the essential reaction features to further develop the e-amination?

A. The reaction has already been carefully optimized and processes cleanly. The only detected by-products are unreacted starting material and dehalogenated. Screening of ligands could increase the conversion and reduce the amount of dehalogenated by-product. Current and reaction time could also be adjusted to increase the conversion.

Characterization Data

Table 2-a – Scope of Amino acid esters

Compound 15



L-glutamic acid di-*tert*-butyl ester hydrochloride (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 10:1) to give the **15** as a white solid (63% yield).

R_f = 0.5 (hexanes:EtOAc = 5:1)

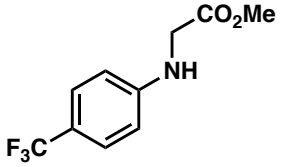
^{1}H NMR (600 MHz, $CDCl_3$) δ 7.39 (d, *J* = 8.4 Hz, 2H), 6.62 (d, *J* = 8.5 Hz, 2H), 4.59 (s, 1H), 4.03 (s, 1H), 2.31-2.43 (m, 2H), 2.01-2.14 (m, 2H), 1.45 (s, 9H), 1.44 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 172.4, 172.3, 149.7, 126.8 (q, *J* = 3.8 Hz), 125.0 (q, *J* = 270.4 Hz), 119.8 (q, *J* = 32.6 Hz), 112.6, 82.5, 80.9, 56.2, 31.6, 28.2, 28.1, 27.8 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.39 ppm.

HRMS (ESI-TOF): calc'd for $C_{20}H_{29}F_3NO_4$ ($[M+H]^+$) 404.2049, found 404.2062.

Compound 18



Glycine methyl ester hydrochloride (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 3:1) to give the **18** as a white solid (54% yield).

R_f = 0.5 (hexanes:EtOAc = 3:1)

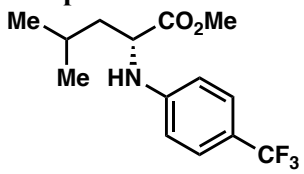
^{1}H NMR (600 MHz, $CDCl_3$) δ 7.43 (d, *J* = 8.4 Hz, 2H), 6.61 (d, *J* = 8.5 Hz, 2H), 4.62 (s, 1H), 3.94 (d, *J* = 5.3 Hz, 2H), 3.81 (s, 3H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 171.1, 149.5, 126.9 (q, *J* = 3.8 Hz), 125.0 (q, *J* = 270.4 Hz), 120.0 (q, *J* = 32.6 Hz), 112.3, 52.6, 45.2 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.41 ppm.

HRMS (ESI-TOF): calc'd for $C_{10}H_{11}F_3NO_2$ ($[M+H]^+$) 234.0742, found 234.0743.

Compound 19



D-leucine methyl ester hydrochloride (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 5:1) to give the **19** as a colorless oil (70% yield).

R_f = 0.6 (hexanes:EtOAc = 5:1)

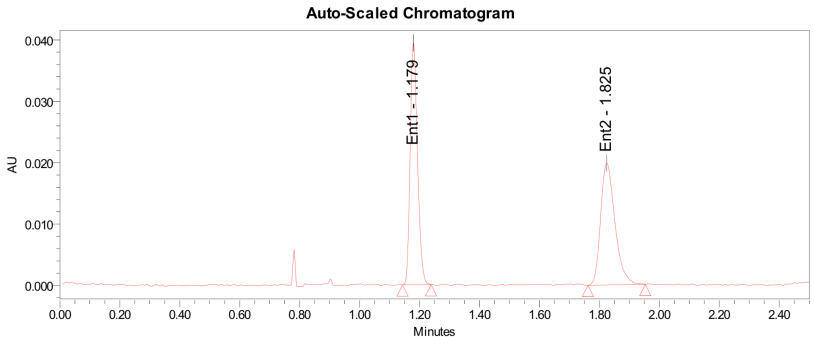
^{1}H NMR (600 MHz, $CDCl_3$) δ 7.40 (d, *J* = 8.4 Hz, 2H), 6.62 (d, *J* = 8.5 Hz, 2H), 4.30 (d, *J* = 8.9 Hz, 1H), 4.12 (td, *J* = 8.5, 6.1 Hz, 1H), 3.73 (s, 3H), 1.78 (dp, *J* = 13.2, 6.6 Hz, 1H), 1.73 – 1.62 (m, 2H), 1.00 (d, *J* = 6.6 Hz, 3H), 0.95 (d, *J* = 6.5 Hz, 3H) ppm.

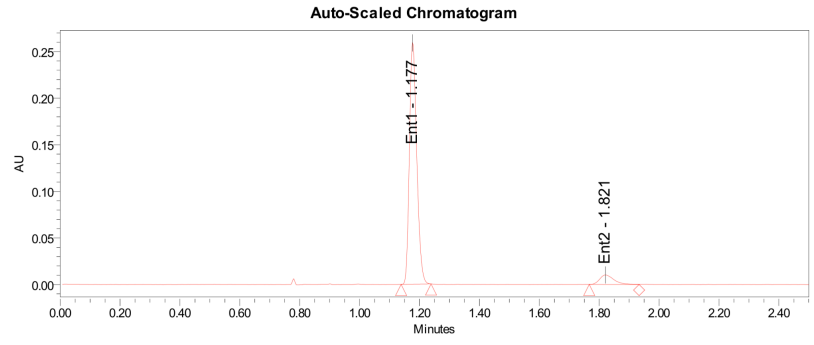
^{13}C NMR (151 MHz, $CDCl_3$) δ 174.6, 149.6, 126.9 (q, *J* = 3.8 Hz), 124.8 (q, *J* = 270.4 Hz), 120.0 (q, *J* = 32.7 Hz), 112.6, 54.8, 52.4, 42.2, 25.0, 22.8, 22.3 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.45 ppm.

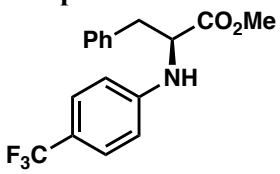
HRMS (ESI-TOF): calc'd for $C_{14}H_{19}F_3NO_2$ ($[M+H]^+$) 290.1368, found 290.1369.

Chiral SFC: IG column (3 μm, 4.6x250 mm) under isocratic conditions [3% MeOH / CO_2 (4 mL/min), 1600 psi backpressure] at 30 °C. The enantiomers were detected by UV light (260 nm). t_R (major) = 1.179 min, t_R (minor) = 1.825 min, 86% *ee*.





Compound 20



L-phenylalanine methyl ester hydrochloride (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 5:1) to give the **20** as a white solid (53% yield).

R_f = 0.5 (hexanes:EtOAc = 5:1)

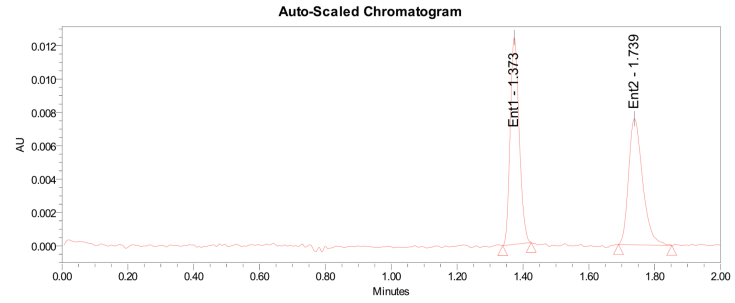
^{1}H NMR (600 MHz, $CDCl_3$) δ 7.41 (d, *J* = 8.6 Hz, 2H), 7.31 (t, *J* = 7.3 Hz, 2H), 7.26 (t, *J* = 7.3 Hz, 1H), 7.15 (d, *J* = 7.2 Hz, 2H), 6.60 (d, *J* = 8.6 Hz, 2H), 4.49 (d, *J* = 7.9 Hz, 1H), 4.43 – 4.40 (m, 1H), 3.71 (s, 3H), 3.20 (dd, *J* = 13.7, 5.8 Hz, 1H), 3.12 (dd, *J* = 13.7, 6.2 Hz, 1H) ppm.

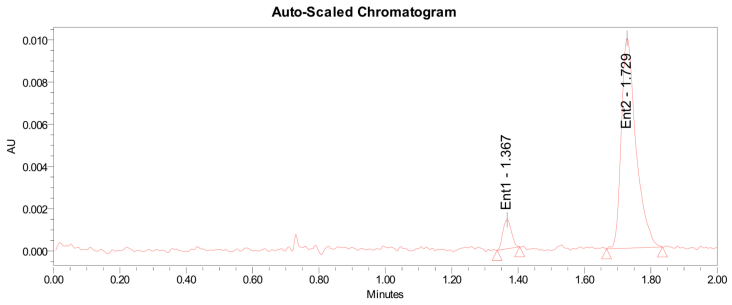
^{13}C NMR (151 MHz, $CDCl_3$) δ 173.0, 149.0, 135.94, 129.4, 128.8, 127.4, 126.9 (q, *J* = 3.7 Hz), 125.5 (q, *J* = 270.3 Hz) 120.1 (q, *J* = 32.6 Hz), 112.7, 57.20, 52.43, 38.5 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.43 ppm.

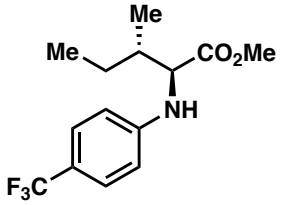
HRMS (ESI-TOF): calc'd for $C_{17}H_{17}F_3NO_2$ ($[M+H]^+$) 324.1211, found 324.1218.

Chiral SFC: IG column (3 μm, 4.6x250 mm) under isocratic conditions [3% MeOH / CO_2 (4 mL/min), 1600 psi backpressure] at 30 °C. The enantiomers were detected by UV light (260 nm). t_R (minor) = 1.329 min, t_R (major) = 1.739 min, 85% *ee*.





Compound 21



L-isoleucine methyl ester hydrochloride (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 10:1) to give the **21** as a colorless liquid (47% yield).

R_f = 0.6 (hexanes:EtOAc = 10:1)

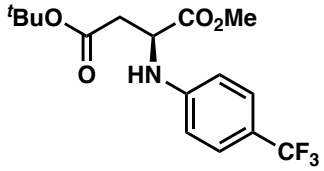
^{1}H NMR (600 MHz, $CDCl_3$) δ 7.39 (d, *J* = 8.4 Hz, 2H), 6.62 (d, *J* = 8.5 Hz, 2H), 4.47 (br, 1H), 4.00 (d, *J* = 5.8 Hz, 1H), 3.73 (s, 3H), 1.85-1.92 (m, 1H), 1.58-1.65 (m, 1H), 1.27-1.34 (m, 1H), 0.96-0.98 (m, 6H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 173.5, 149.7, 126.9 (q, *J* = 3.8 Hz), 125.0 (q, *J* = 270.3 Hz), 119.8 (q, *J* = 32.7 Hz), 112.6, 60.7, 52.2, 38.1, 25.8, 15.6, 11.6 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.42 ppm.

HRMS (ESI-TOF): calc'd for $C_{14}H_{19}F_3NO_2$ ($[M+H]^+$) 290.1368, found 290.1375.

Compound 22



H-Asp(O^{t}Bu)-OMe•HCl (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 5:1) to give the **22** as a white solid (49% yield).

R_f = 0.5 (hexanes:EtOAc = 10:1)

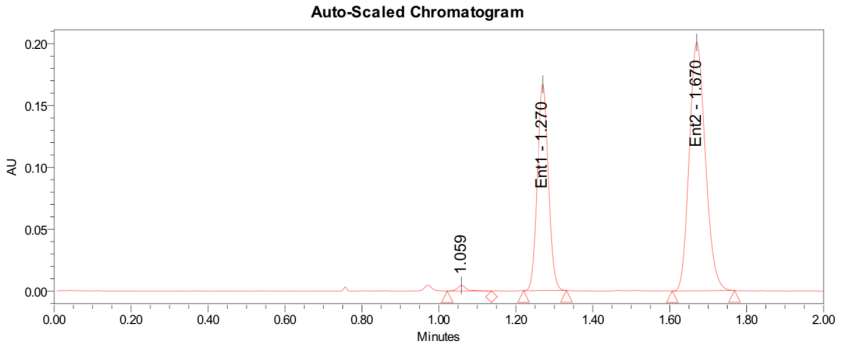
^{1}H NMR (600 MHz, $CDCl_3$) δ 7.42 (d, J = 8.4 Hz, 2H), 6.66 (d, J = 8.4 Hz, 2H), 4.83 (d, J = 8.4 Hz, 1H), 4.43 (dt, J = 8.5, 5.5 Hz, 1H), 2.82 (dd, J = 15.9, 5.6 Hz, 1H), 2.77 (dd, J = 15.9, 5.5 Hz, 1H), 1.44 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 172.5, 169.6, 149.0, 126.9 (q, J = 3.8 Hz), 124.9 (q, J = 270.6 Hz), 120.3 (q, J = 32.7 Hz), 112.8, 82.0, 53.0, 52.8, 38.4, 28.14 ppm.

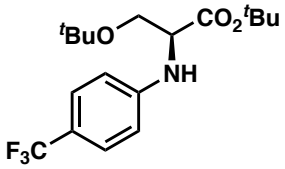
^{19}F NMR (376 MHz, $CDCl_3$) δ -61.49 ppm.

HRMS (ESI-TOF): calc'd for $C_{16}H_{21}F_3NO_4$ ($[M+H]^+$) 348.1423, found 348.1429.

Chiral SFC: IG column (3 μm, 4.6x250 mm) under isocratic conditions [3% MeOH / CO_2 (4 mL/min), 1600 psi backpressure] at 30 °C. The enantiomers were detected by UV light (260 nm). t_R (minor) = 1.270 min, t_R (major) = 1.670 min, 82% *ee*.



Compound 23



H-Ser(O^{t}Bu)-O^{t}Bu•HCl (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 20:1) to give the **23** as a white solid (69% yield).

R_f = 0.6 (hexanes:EtOAc = 10:1)

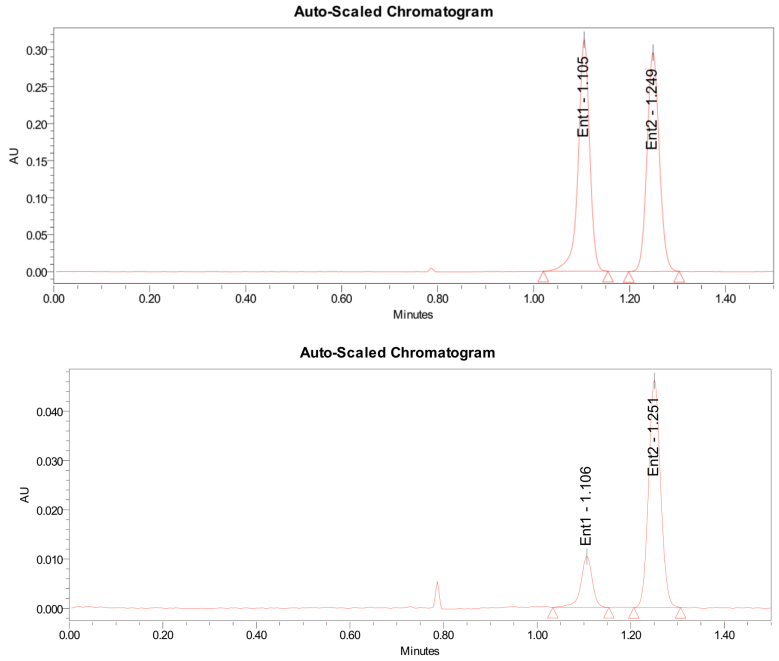
^{1}H NMR (600 MHz, CDCl$_3$) δ 7.39 (d, *J* = 8.4 Hz, 2H), 6.62 (d, *J* = 8.5 Hz, 2H), 4.82 (d, *J* = 8.8 Hz, 1H), 4.09 (dt, *J* = 8.8, 3.8 Hz, 1H), 3.78 (dd, *J* = 8.6, 3.7 Hz, 1H), 3.65 (dd, *J* = 8.6, 3.8 Hz, 1H), 1.45 (s, 9H), 1.17 (s, 9H) ppm.

^{13}C NMR (151 MHz, CDCl$_3$) δ 170.7, 149.9, 126.7 (q, *J* = 3.8 Hz), 125.1 (q, *J* = 270.3 Hz), 119.6 (q, *J* = 32.6 Hz), 112.7, 82.0, 73.5, 62.4, 57.0, 28.2, 27.5 ppm.

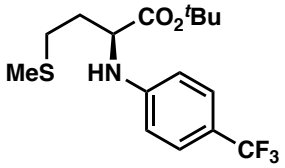
^{19}F NMR (376 MHz, CDCl$_3$) δ -61.32 ppm.

HRMS (ESI-TOF): calc'd for $C_{16}H_{27}F_3NO_3$ ($[M+H]^+$) 362.1943, found 362.1946.

Chiral SFC: IG column (3 μm, 4.6x250 mm) under isocratic conditions [3% MeOH / CO2 (4 mL/min), 1600 psi backpressure] at 30 °C. The enantiomers were detected by UV light (260 nm). t_R (minor) = 1.105 min, t_R (major) = 1.249 min, 64% *ee*.



Compound 24



L-Methionine *tert*-butyl ester hydrochloride (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 4:1) to give the **24** as a white solid (59% yield).

R_f = 0.5 (hexanes:EtOAc = 3:1)

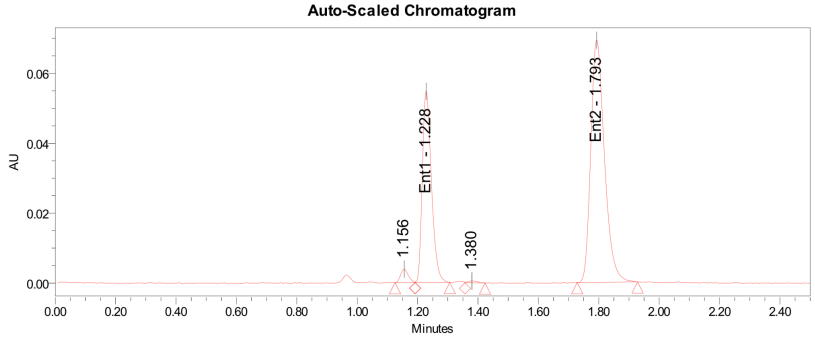
^{1}H NMR (600 MHz, $CDCl_3$) δ 7.40 (d, *J* = 8.5 Hz, 2H), 6.65 (d, *J* = 8.5 Hz, 2H), 4.54 (d, *J* = 8.4 Hz, 1H), 4.17 (q, *J* = 7.1 Hz, 1H), 2.60 (t, *J* = 7.4 Hz, 2H), 2.07-2.16 (m, 4H), 2.00 (dq, *J* = 14.1, 7.1 Hz, 1H), 1.45 (s, 9H) ppm.

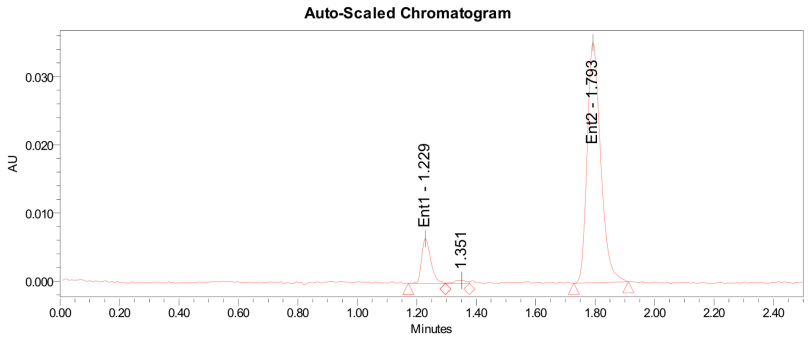
^{13}C NMR (151 MHz, $CDCl_3$) δ 172.3, 149.6, 126.8 (q, *J* = 3.7 Hz), 125.0 (q, *J* = 270.3 Hz), 119.9 (q, *J* = 32.7 Hz), 11287, 82.6, 55.7, 32.2, 30.3, 28.2, 15.7 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.41 ppm.

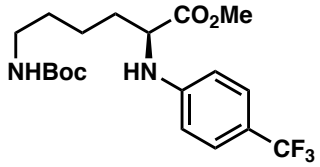
HRMS (ESI-TOF): calc'd for $C_{16}H_{23}F_3NO_2S$ ($[M+H]^+$) 350.1402, found 350.1408.

Chiral SFC: IG column (3 μm, 4.6x250 mm) under isocratic conditions [3% MeOH / CO_2 (4 mL/min), 1600 psi backpressure] at 30 °C. The enantiomers were detected by UV light (260 nm). t_R (minor) = 1.228 min, t_R (major) = 1.793 min, 78% *ee*.





Compound 25



H-Lys(Boc)-OMe•HCl (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 2:1) to give the **25** as a white solid (54% yield).

R_f = 0.3 (hexanes:EtOAc = 3:1)

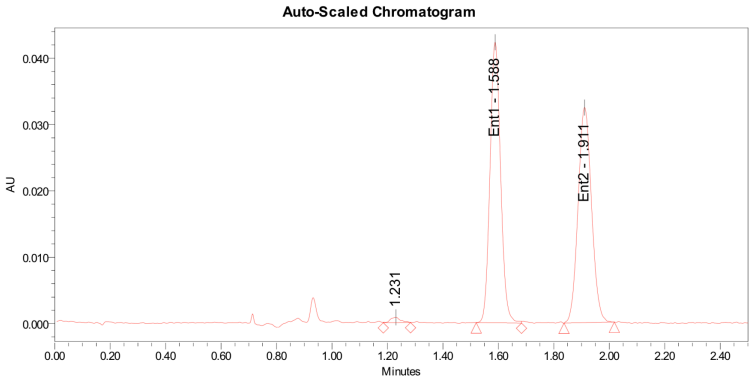
^{1}H NMR (600 MHz, $CDCl_3$) δ 7.39 (d, *J* = 8.5 Hz, 2H), 6.60 (d, *J* = 8.6 Hz, 2H), 4.54 (s, 2H), 4.09 (q, *J* = 7.1 Hz, 1H), 3.73 (s, 3H), 3.19 – 3.04 (m, 2H), 1.86-1.92 (m, 1H), 1.77-1.83 (m, 1H), 1.39-1.54 (m, 13H) ppm.

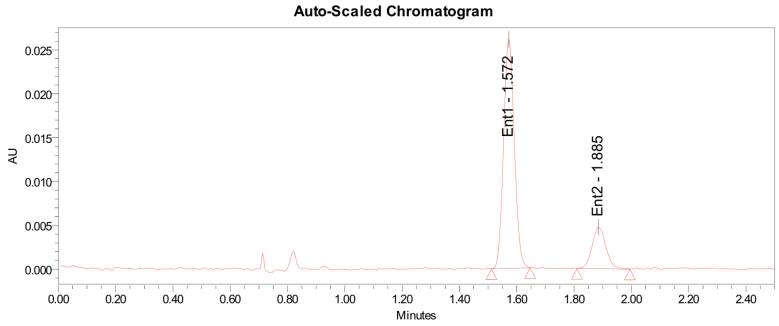
^{13}C NMR (151 MHz, $CDCl_3$) δ 185.7, 174.0, 156.2, 149.4, 126.9 (q, *J* = 3.8 Hz), 124.9 (q, *J* = 270.5 Hz), 119.9 (q, *J* = 33.4, 32.5 Hz), 112.5, 56.0, 52.5, 40.2, 32.4, 30.0, 28.5, 22.8 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.42 ppm.

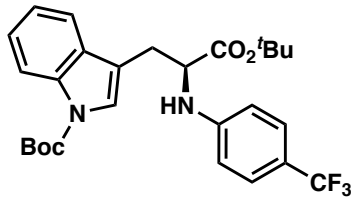
HRMS (ESI-TOF): calc'd for $C_{19}H_{28}F_3N_2O_4$ ($[M+H]^+$) 405.2001, found 405.2004.

Chiral SFC: IG column (3 μm, 4.6x250 mm) under isocratic conditions [3% MeOH / CO_2 (4 mL/min), 1600 psi backpressure] at 30 °C. The enantiomers were detected by UV light (260 nm). t_R (major) = 1.558 min, t_R (minor) = 1.885 min, 62% *ee*.





Compound 26



H-Trp(Boc)-O^{t}Bu•HCl (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 10:1) to give the **26** as a pale yellow liquid (57% yield).

R_f = 0.5 (hexanes:EtOAc = 10:1)

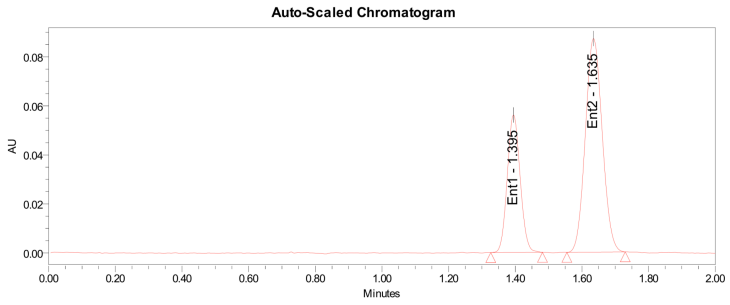
^{1}H NMR (600 MHz, $CDCl_3$) δ 8.14 (s, 1H), 7.44 (m, 4H), 7.33 (t, *J* = 7.4 Hz, 1H), 7.23 (t, *J* = 7.5 Hz, 1H), 6.61 (d, *J* = 8.6 Hz, 2H), 4.59 (d, *J* = 7.9 Hz, 1H), 4.37 (dd, *J* = 8.0, 4.0 Hz, 1H), 3.27 (dd, *J* = 14.6, 5.9 Hz, 1H), 3.20 (dd, *J* = 14.7, 5.9 Hz, 1H), 1.65 (s, 9H), 1.39 (s, 9H) ppm.

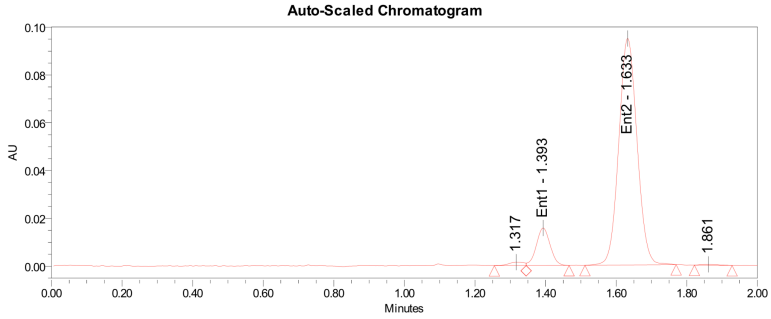
^{13}C NMR (151 MHz, $CDCl_3$) δ 171.7, 149.7, 149.2, 135.5, 130.7, 126.8 (q, *J* = 3.7 Hz), 125.0 (q, *J* = 270.3 Hz), 124.7, 124.4, 122.7, 119.8 (q, *J* = 32.7 Hz), 119.1, 115.4, 115.2, 112.7, 82.6, 56.7, 28.3, 28.1 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.36 ppm.

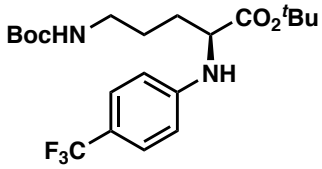
HRMS (ESI-TOF): calc'd for $C_{27}H_{32}F_3N_2O_4$ ($[M+H]^+$) 505.2314, found 505.2310.

Chiral SFC: IG column (3 μm, 4.6x250 mm) under isocratic conditions [3% MeOH / CO2 (4 mL/min), 1600 psi backpressure] at 30 °C. The enantiomers were detected by UV light (260 nm). t_R (minor) = 1.385 min, t_R (major) = 1.633 min, 78% *ee*.




Auto-Scaled Chromatogram
AU
Minutes
1.317
Ent1 - 1.393
Ent2 - 1.633
1.861

Compound 27



H-Orn(Boc)-O^{t}Bu•HCl (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 5:1) to give the **27** as a white solid (70% yield).

R_f = 0.3 (hexanes:EtOAc = 5:1)

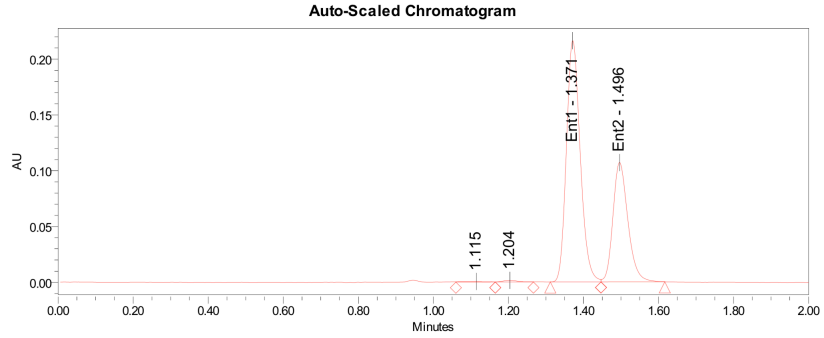
^{1}H NMR (600 MHz, $CDCl_3$) δ 7.27-7.48 (m, 2H), 6.57-6.92 (m, 2H), 4.84-5.15 (m, 1H), 4.02-4.23 (m, 2H), 3.18-3.22 (m, 2H), 1.84-1.91 (m, 1H), 1.61-1.80 (m, 3H), 1.35-1.53 (m, 18H) ppm.

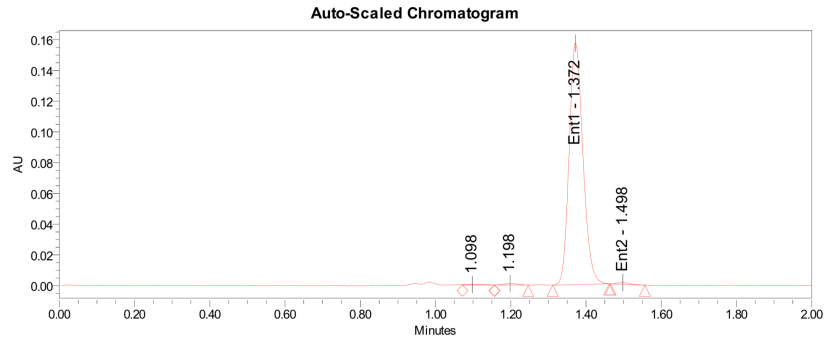
^{13}C NMR (151 MHz, $CDCl_3$) δ 171.8, 159.3, 155.7, 150.7, 126.7 (q, J = 3.7 Hz), 125.1 (q, J = 270.4 Hz), 118.8 (q, J = 33.5, 32.6 Hz), 115.6, 111.9, 82.4, 80.2, 53.70, 43.0, 30.8, 28.5, 28.1, 24.9 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.21, -61.65 ppm.

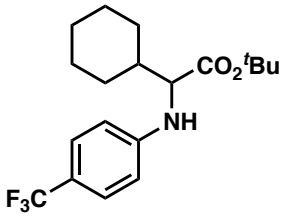
HRMS (ESI-TOF): calc'd for $C_{21}H_{32}F_3N_2O_4$ ($[M+H]^+$) 433.2314, found 433.2320.

Chiral SFC: IG column (3 μm, 4.6x250 mm) under isocratic conditions [3% MeOH / CO2 (4 mL/min), 1600 psi backpressure] at 30 °C. The enantiomers were detected by UV light (260 nm). t_R (major) = 1.371 min, t_R (minor) = 1.485 min, 99% *ee*.





Compound 28



tert-Butyl 2-amino-2-cyclohexylacetate (0.1 mmol), 4-trifluormethylbromobenzene (0.2 mmol), DBU (0.3 mmol). Electrolysis was conducted for 3 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 20:1) to give the **28** as a white solid (51% yield).

R_f = 0.6 (hexanes:EtOAc = 10:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 7.38 (d, *J* = 8.8 Hz, 2H), 6.62 (d, *J* = 8.5 Hz, 2H), 4.47 (d, *J* = 8.4 Hz, 1H), 3.79 (dd, *J* = 8.2, 5.7 Hz, 1H), 1.66-1.81 (m, 6H), 1.44 (s, 9H), 1.11-1.30 (m, 5H) ppm.

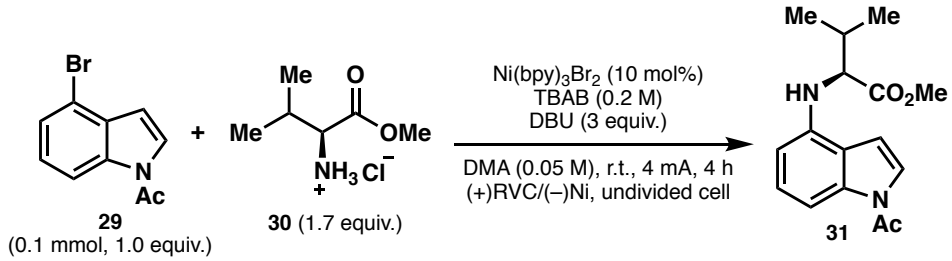
^{13}C NMR (151 MHz, $CDCl_3$) δ 172.2, 150.2, 126.7 (q, *J* = 3.8 Hz), 125.0 (q, *J* = 270.3 Hz), 119.4 (q, *J* = 32.6 Hz), 112.6, 82.1, 61.9, 41.4, 29.6, 29.2, 26.3, 26.3, 26.2 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.33 ppm.

HRMS (ESI-TOF): calc'd for $C_{19}H_{27}F_3NO_2$ ($[M+H]^+$) 358.1994, found 358.1997.

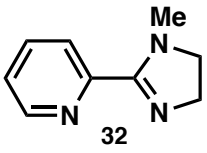
Table 2-b – Application to total synthesis

Optimization table



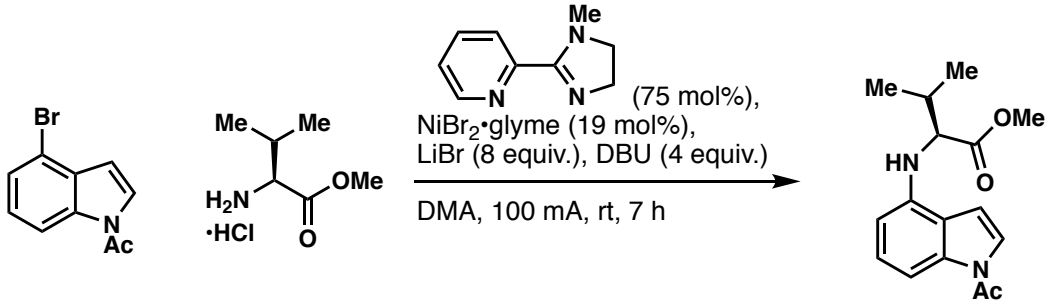
Optimization was conducted following the general procedure. The results are summarized below:

Entry	Change from standard conditions	Yield
1	First generation conditions (conditions A)	0%
2	None	16%
3	LiBr (0.2 M)	11%
4	$NiBr_2 \cdot 3H_2O$ (10 mol%), Ligand = bpy (30 mol%)	20%
5	$NiBr_2 \cdot 3H_2O$ (10 mol%), Ligand = **16** (30 mol%)	6%
6	$NiBr_2 \cdot 3H_2O$ (10 mol%), Ligand = **32** (30 mol%)	10%
7	$NiBr_2$•glyme (19 mol%), Ligand = **32** (75 mol%), LiBr (0.2 M)	32%
8	3.0 mmol scale with $NiBr_2$•glyme (19 mol%) and **32** (75 mol%)	51%[a]

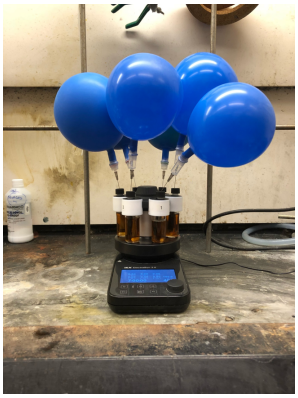


[a] LiBr (0.89 M), DMA (0.1 M), 100 mA, 7 h.

Large scale experiment (entry 8)



To a cell were added compound **29** (735 mg, 3.09 mmol), **30** (991 mg, 5.87 mmol, 1.9 equiv.), **32** (375 mg, 2.32 mmol, 75 mol%), LiBr solution (14 mL, 2.0 M in DMA, 8 equiv.), DBU (1.75 mL, 11.7 mmol, 4 equiv.) and DMA (13.9 mL). RVC anode and Ni form cathode were inserted into the mixture. A solution of $NiBr_2$•glyme (181 mg, 0.587 mmol, 19 mol%) in DMA (3.6 mL) was added then the reaction mixture was electrolyzed under a constant current of 100 mA for 7 h at room temperature. After the reaction, the reaction was quenched with water. The resulting mixture was extracted with Et_2O (x3). The combined organic layer was dried over anhydrous $MgSO_4$ and concentrated under reduced pressure. The crude material was purified by column chromatography (silica, 10:1 to 3:2 hexane:EtOAc) to afford 475 mg (51%) of compound **31**.



Physical State: yellow oil.

^{1}H NMR (600 MHz, $CDCl_3$): δ 7.83 (d, *J* = 8.3 Hz, 1H), 7.34 (d, *J* = 3.8 Hz, 1H), 7.19 (t, *J* = 8.1 Hz, 1H), 6.65 (d, *J* = 3.9 Hz, 1H), 6.44 (d, *J* = 7.9 Hz, 1H), 4.41 (s, 1H), 4.02 (d, *J* = 5.8 Hz, 1H), 3.72 (s, 3H), 2.62 (s, 3H), 2.19 (dq, *J* = 13.4, 6.5 Hz, 1H), 1.10 (d, *J* = 6.8 Hz, 3H), 1.05 (d, *J* = 6.7 Hz, 3H).

^{13}C NMR (151 MHz, $CDCl_3$): δ 174.33, 140.36, 136.50, 126.62, 123.39, 119.10, 105.28, 105.27, 62.57, 52.09, 31.82, 24.26, 18.94.

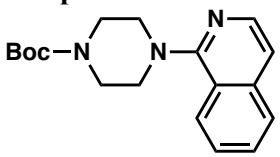
HRMS (ESI-TOF): calcd for $C_{16}H_{20}N_2O_3$ $[M+H]^+$: 289.1547, found: 289.1555.

TLC: R_f = 0.33 (3:2 hexanes:EtOAc, $Ce_2(SO_4)_3$ in phosphomolybdic acid).

$[\alpha]_D^{20}$ = −3.3 (c 0.392, $CHCl_3$).

Table 3 – Amination of heteroaryl halides with N-Boc-piperazine

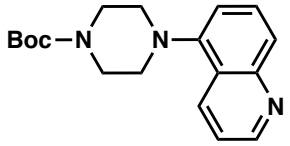
Compound 33



N-Boc-piperazine (0.3 mmol), 1-bromoisoquinoline (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 2:1) to give the **33** as a white solid (28% yield). The spectrum matched with the reported values.[17]

R_f = 0.4 (hexanes:EtOAc = 2:1)

Compound 34



N-Boc-piperazine (0.3 mmol), 5-bromoquinoline (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 6 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 1:1) to give the **34** as off-white solid (59% yield).

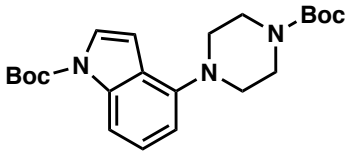
R_f = 0.4 (hexanes:EtOAc = 2:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 8.89 (d, *J* = 4.1 Hz, 1H), 8.51 (d, *J* = 8.5 Hz, 1H), 7.83 (d, *J* = 8.5 Hz, 1H), 7.62 (t, *J* = 8.0 Hz, 1H), 7.39 (dd, *J* = 8.5, 4.2 Hz, 1H), 7.11 (d, *J* = 8.3 Hz, 1H), 3.69 (br, 4H), 3.03 (br, 4H), 1.50 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 155.0, 150.4, 149.6, 149.6, 132.1, 129.5, 125.3, 124.2, 120.5, 115.5, 80.1, 53.3, 44.2 (br), 28.6 ppm.

HRMS (ESI-TOF): calc'd for $C_{18}H_{24}N_3O_2$ ($[M+H]^+$) 314.1869, found 314.1879.

Compound 35



N-Boc-piperazine (0.3 mmol), *N*-Boc-4-bromoindole (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 7 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 5:1) to give the **35** as off-white solid (68% yield).

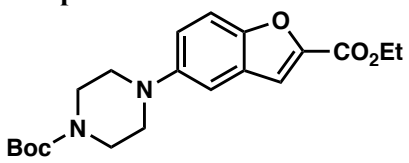
R_f = 0.5 (hexanes:EtOAc = 5:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 7.85 (d, *J* = 7.9 Hz, 1H), 7.56 (d, *J* = 3.6 Hz, 1H), 7.23 (t, *J* = 8.0 Hz, 1H), 6.74 (d, *J* = 7.7 Hz, 1H), 6.58 (d, *J* = 3.7 Hz, 1H), 3.66 (br, 4H), 3.11 (br, 4H), 1.67 (s, 9H), 1.50 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 155.0, 149.9, 145.8, 136.4, 125.0, 124.8, 124.3, 110.6, 110.4, 105.4, 83.9, 79.9, 51.8, 44.1 (br), 28.6, 28.3 ppm.

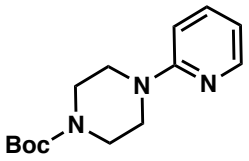
HRMS (ESI-TOF): calc'd for $C_{22}H_{32}N_3O_4$ ($[M+H]^+$) 403.2393, found 403.2400.

Compound 36



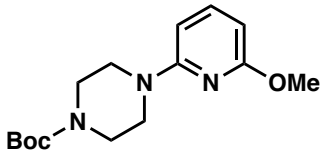
N-Boc-piperazine (0.3 mmol), ethyl 5-bromobenzofuran-2-carboxylate (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 6 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 3:1) to give the **36** as pale yellow solid (79% yield). The spectrum matched with the reported values.[18]

Compound 37



N-Boc-piperazine (0.3 mmol), 2-bromopyridine (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 3:1) to give the **37** as a white solid (51% yield). The spectrum matched with the reported values.[17]

Compound 38



N-Boc-piperazine (0.3 mmol), 6-methoxy-2-chloropyridine (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 4:1) to give the **38** as off-white solid (56% yield).

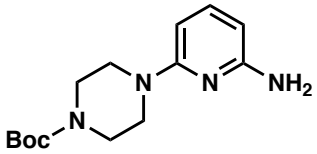
R_f = 0.4 (hexanes:EtOAc = 4:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 7.40 (t, *J* = 7.9 Hz, 1H), 6.15 (d, *J* = 8.0 Hz, 1H), 6.09 (d, *J* = 7.9 Hz, 1H), 3.85 (s, 3H), 3.48-3.53 (m, 8H), 1.48 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 163.2, 158.2, 154.9, 140.3, 98.7, 98.4, 80.0, 53.1, 45.2, 43.4 (br), 28.5 ppm.

HRMS (ESI-TOF): calc'd for $C_{15}H_{24}N_3O_3$ ($[M+H]^+$) 294.1818, found 294.1823.

Compound 39



N-Boc-piperazine (0.3 mmol), 2-amino-6-bromopyridine (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 1:1) to give the **39** as off-white solid (45% yield).

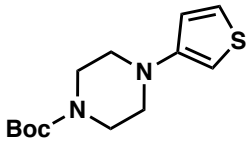
R_f = 0.4 (hexanes:EtOAc = 1:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 7.27 (t, *J* = 7.9 Hz, 1H), 5.98 (d, *J* = 8.1 Hz, 1H), 5.88 (d, *J* = 7.7 Hz, 1H), 4.21 (br, 2H), 3.43-3.50 (m, 8H), 1.47 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 158.9, 157.4, 155.0, 139.5, 97.8, 96.6, 79.9, 45.2, 43.5 (br), 28.6 ppm.

HRMS (ESI-TOF): calc'd for $C_{14}H_{23}N_4O_2$ ($[M+H]^+$) 279.1821, found 279.1827.

Compound 40



N-Boc-piperazine (0.3 mmol), 3-chlorothiophene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 6 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 5:1) to give the **40** as a white solid (60% yield).

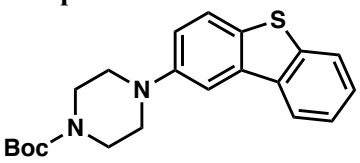
R_f = 0.6 (hexanes:EtOAc = 5:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 77.24 (dd, *J* = 5.2, 3.1 Hz, 1H), 6.86 (dd, *J* = 5.3, 1.5 Hz, 1H), 6.21 (dd, *J* = 3.0, 1.5 Hz, 1H), 3.55-3.57 (m, 4H), 3.03-3.05 (m, 4H), 1.47 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 154.8, 152.3, 125.7, 120.3, 101.2, 80.0, 50.6, 43.5 (br), 28.5 ppm.

HRMS (ESI-TOF): calc'd for $C_{13}H_{21}N_2O_2S$ ($[M+H]^+$) 269.1324, found 269.1329.

Compound 41



N-Boc-piperazine (0.3 mmol), 2-bromodibenzothiophene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 7 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 5:1) to give the **41** as a white solid (68% yield).

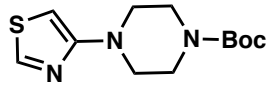
R_f = 0.5 (hexanes:EtOAc = 5:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 8.10 (dd, *J* = 6.0, 3.2 Hz, 1H), 7.83 (dd, *J* = 5.8, 3.2 Hz, 1H), 7.72 (d, *J* = 8.7 Hz, 1H), 7.66 (s, 1H), 7.43 (dd, *J* = 6.0, 3.2 Hz, 2H), 7.16 (dd, *J* = 8.7, 2.3 Hz, 1H), 3.66 (br, 4H), 3.22 (br, 4H), 1.52 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 154.9, 149.4, 140.5, 136.6, 135.6, 131.7, 126.7, 124.3, 123.3, 123.1, 121.5, 118.7, 109.2, 80.1, 50.7, 43.8 (br), 28.6 ppm.

HRMS (ESI-TOF): calc'd for $C_{21}H_{25}N_2O_2S$ ($[M+H]^+$) 369.1637, found 369.1641.

Compound 42



N-Boc-piperazine (0.3 mmol), 4-bromothiazole (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 3:1) to give the **42** as a white solid (33% yield).

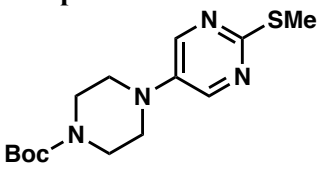
R_f = 0.5 (hexanes:EtOAc = 3:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 8.60 (d, *J* = 2.1 Hz, 1H), 5.96 (d, *J* = 2.1 Hz, 1H), 3.57-3.58 (m, 4H), 3.25-3.27 (m, 4H), 1.47 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 162.7, 154.9, 151.4, 90.6, 80.1, 48.82, 43.36 (br), 28.55 ppm.

HRMS (ESI-TOF): calc'd for $C_{12}H_{20}N_3O_2$ ($[M+H]^+$) 270.1276, found 270.1275.

Compound 43



N-Boc-piperazine (0.3 mmol), 4-bromo-1-methylthiopyrimidine (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:CH_2Cl_2:EtOAc = 1:1:0.5) to give the **43** as a white solid (71% yield).

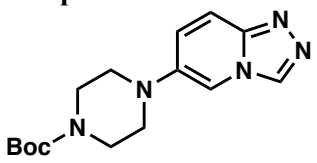
R_f = 0.6 (hexanes:CH_2Cl_2:EtOAc = 1:1:0.5)

^{1}H NMR (600 MHz, $CDCl_3$) δ 8.23 (s, 2H), 3.57-3.59 (m, 4H), 3.09-3.08 (m, 4H), 2.53 (s, 3H), 1.46 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 162.9, 154.6, 146.3, 141.3, 80.3, 48.8, 43.3 (br), 28.5, 14.4 ppm.

HRMS (ESI-TOF): calc'd for $C_{14}H_{23}N_4O_2S$ ($[M+H]^+$) 311.1542, found 311.1545.

Compound 44



N-Boc-piperazine (0.3 mmol), 6-bromo-[1,2,4]triazolo[4,3-a]pyridine (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, CH_2Cl_2:MeOH = 20:1) to give the **44** as pale-blue liquid (67% yield).

$\boldsymbol{R_f}$ = 0.2 (CH_2Cl_2:MeOH = 10:1)

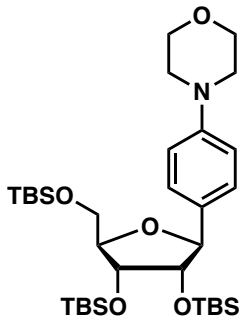
^{1}H NMR (600 MHz, $CDCl_3$) δ 8.71 (s, 1H), 7.69 (d, *J* = 9.7 Hz, 1H), 7.45 (s, 1H), 7.18 (d, *J* = 9.9 Hz, 1H), 3.62 (br, 4H), 3.04 (br, 4H), 1.49 (s, 9H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 154.7, 147.4, 141.1, 136.0, 125.5, 116.4, 107.7, 80.5, 50.2, 43.4 (br), 28.6 ppm.

HRMS (ESI-TOF): calc'd for $C_{15}H_{22}N_5O_2$ ($[M+H]^+$) 304.1773, found 304.1777.

Table 4-A – C-N bond formation on nucleosides

Compound 45



TBS-protected 1-(4-bromophenyl)ribose **45** (0.10 mmol), morpholine (0.15 mmol), DBU (0.2 mmol). Electrolysis was conducted for 4 h (4 mA) following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 5:1) to give the **46** as a colorless oil (50% yield).

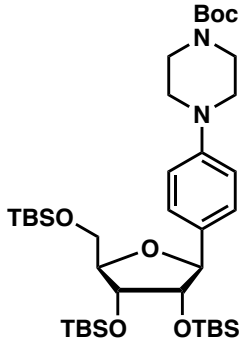
R_f = 0.4 (hexanes:EtOAc = 5:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 7.31 (d, *J* = 8.5 Hz, 2H), 6.85 (d, *J* = 8.7 Hz, 2H), 4.71 (d, *J* = 7.5 Hz, 1H), 4.13 (dd, *J* = 4.5, 2.1 Hz, 1H), 3.99-4.01 (m, 1H), 3.84-3.87 (m, 5H), 3.75-3.80 (m, 2H), 3.10-3.16 (m, 4H), 0.95 (s, 9H), 0.93 (s, 9H), 0.80 (s, 9H), 0.12 (s, 3H), 0.11 (s, 3H), 0.09 (s, 6H), -0.13 (s, 3H), -0.39 (s, 3H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 151.3, 132.4, 128.0, 115.7, 85.7, 83.0, 79.5, 73.9, 67.1, 63.9, 49.9, 26.2, 26.1, 26.0, 18.6, 18.3, 18.1, -4.3, -4.4, -5.1, -5.2, -5.4 ppm.

HRMS (ESI-TOF): calc'd for $C_{33}H_{64}NO_5Si_3$ ($[M+H]^+$) 638.4092, found 638.4096.

Compound 46



TBS-protected 1-(4-bromophenyl)ribose **45** (0.10 mmol), N-Boc-piperazine (0.15 mmol), DBU (0.2 mmol). Electrolysis was conducted for 4 h (4 mA) following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 10:1) to give the **47** as a colorless oil (71% yield).

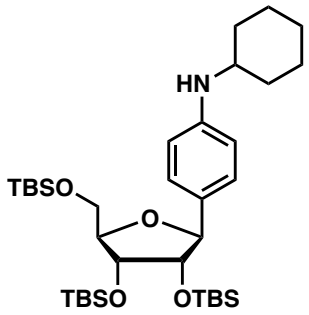
R_f = 0.6 (hexanes:EtOAc = 5:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 7.31 (d, *J* = 8.6 Hz, 2H), 6.87 (d, *J* = 8.7 Hz, 2H), 4.70 (d, *J* = 7.5 Hz, 1H), 4.12 (dd, *J* = 4.4, 2.0 Hz, 1H), 4.00 (q, *J* = 3.4 Hz, 1H), 3.84 (dd, *J* = 7.5, 4.5 Hz, 1H), 3.77 – 3.78 (m, 1H), 3.57 – 3.58 (m, 4H), 3.06 – 3.12 (m, 4H), 1.48 (s, 9H), 0.94 (s, 9H), 0.93 (s, 9H), 0.79 (s, 9H), 0.12 (s, 3H), 0.11 (s, 3H), 0.09 (s, 6H), -0.13 (s, 3H), -0.41 (s, 3H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 154.9, 151.3, 132.7, 128.0, 116.6, 85.8, 82.9, 80.0, 79.5, 73.9, 63.90, 50.0, 43.7, 28.6, 26.2, 26.1, 26.0, 18.5, 18.2, 18.1, -4.3, -4.4, -5.1, -5.2, -5.4 ppm.

HRMS (ESI-TOF): calc'd for $C_{38}H_{73}N_2O_6Si_3$ ($[M+H]^+$) 737.4776, found 737.4773.

Compound 47



TBS-protected 1-(4-bromophenyl)ribose **45** (0.10 mmol), cyclohexylamine (0.3 mmol), no DBU. Electrolysis was conducted for 4 h (4 mA) following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 20:1) to give the **48** as a colorless oil (34% yield).

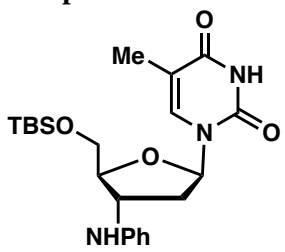
R_f = 0.7 (hexanes:EtOAc = 5:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 7.17 (d, *J* = 8.5 Hz, 2H), 6.53 (d, *J* = 8.5 Hz, 2H), 4.65 (d, *J* = 7.4 Hz, 1H), 4.12 (dd, *J* = 4.4, 2.3 Hz, 1H), 3.97 (d, *J* = 9.6 Hz, 1H), 3.83 (dd, *J* = 7.4, 4.5 Hz, 1H), 3.76 (d, *J* = 3.7 Hz, 2H), 3.25 (tt, *J* = 10.1, 3.6 Hz, 1H), 2.03-2.05 (m 2H), 1.73-1.76 (m, 2H), 1.63-1.66 (m, 1H), 1.32-1.40 (m, 2H), 1.18-1.25 (m, 1H), 1.08-1.15 (m, 2H), 0.94 (s, 9H), 0.93 (s, 9H), 0.80 (s, 9H), 0.12 (s, 3H), 0.10 (s, 3H), 0.09 (s, 3H), 0.08 (s, 3H), -0.13 (s, 3H), -0.35 (s, 3H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 147.2, 129.1, 128.3, 113.4, 85.4, 83.3, 79.4, 73.9, 63.9, 52.1, 33.6, 33.5, 26.2, 26.1, 26.1, 26.1, 25.2, 25.2, 18.6, 18.6, 18.6, -4.3, -4.3, -4.4, -5.0, -5.2, -5.4 ppm.

HRMS (ESI-TOF): calc'd for $C_{35}H_{68}NO_4Si_3$ ($[M+H]^+$) 650.4456, found 650.4467.

Compound 48



TBS-protected 3-amino thymidine **49** (0.05 mmol), bromobenzene (0.1 mmol), DBU (0.15 mmol) with TMSCl (0.05 mmol). Electrolysis was conducted for 2 h (4 mA) following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 1:1.5) to give the arylated product in 56% yield as a mixture of **50** and DMA-adduct **SI-1** (**50**:**SI-1** = **1.4**:**1**)

Major isomer **50** was characterized below.

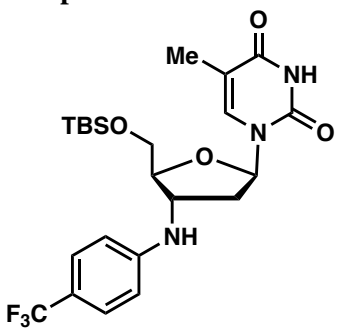
R_f = 0.5 (hexanes:EtOAc = 1:2)

^{1}H NMR (600 MHz, $CDCl_3$) δ 9.09 (br, 1H), 7.56 (s, 1H), 7.19 (t, *J* = 7.8 Hz, 2H), 6.76 (t, *J* = 7.3 Hz, 1H), 6.61 (d, *J* = 7.8 Hz, 2H), 6.36 (t, *J* = 6.7 Hz, 1H), 4.16-4.18 (m, 1H), 4.01-3.97 (m, 2H), 3.87 (dd, *J* = 11.3, 2.2 Hz, 1H), 2.27-2.37 (m, 2H), 1.95 (s, 3H), 0.96 (s, 9H), 0.15 (s, 3H), 0.14 (s, 3H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 163.9, 150.6, 146.6, 135.4, 129.6, 118.4, 113.4, 111.2, 86.0, 85.0, 63.8, 54.0, 39.3, 26.21, 18.6, 12.8, -5.2 ppm.

HRMS (ESI-TOF): calc'd for $C_{22}H_{34}N_3O_4Si$ ($[M+H]^+$) 432.2319, found 432.2320.

Compound 49



TBS-protected 3-amino thymidine **49** (0.05 mmol), 4-trifluoromethylbromobenzene (0.2 mmol), DBU (0.1 mmol). Electrolysis was conducted for 2 h (4 mA)following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 1:1) to give the **51** as a white solid (62% yield).

R_f = 0.4 (hexanes:EtOAc = 1:1)

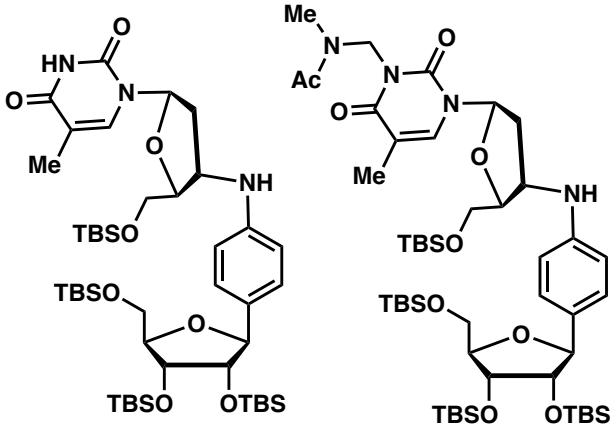
^{1}H NMR (600 MHz, $CDCl_3$) δ 9.32 (br, 1H), 7.54 (s, 1H), 7.42 (d, *J* = 8.0 Hz, 2H), 6.65 (d, *J* = 8.1 Hz, 2H), 6.39 (s, 1H), 4.76 (br, 1H), 4.18 (s, 1H), 4.04 (s, 1H), 3.97 (d, *J* = 11.2 Hz, 1H), 3.88 (d, *J* = 10.8 Hz, 1H), 2.27-2.40 (m, 2H), 1.96 (s, 3H), 0.96 (s, 9H), 0.16 (s, 3H), 0.15 (s, 3H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 163.8, 150.6, 149.3, 135.2, 126.9, 124.9 (q, *J* = 270.3 Hz), 119.8 (q, *J* = 32.8 Hz), 112.4, 111.4, 85.9, 85.2, 63.8, 53.9, 39.0, 26.1, 18.5, 12.8, -5.2, -5.2 ppm.

^{19}F NMR (376 MHz, $CDCl_3$) δ -61.40 ppm.

HRMS (ESI-TOF): calc'd for $C_{23}H_{33}F_3N_3O_4Si$ ($[M+H]^+$) 500.2192, found 500.2192.

Compound 50a and 50b



TBS-protected 3-amino thymidine **49** (0.05 mmol), TBS-protected 1-(4-bromophenyl)ribose **45** (0.05 mmol), DBU (0.1 mmol). Electrolysis was conducted for 3 h (4 mA) following the general procedure. The crude material was purified by PTLC (silica gel, hexanes:EtOAc = 1:1) to give the arylated product in 62% yield as a mixture of **52a** and DMA-adduct **52b** (**a**:**b** = 1:2.6).

*Characterization of **52a***

R_f = 0.5 (hexanes:EtOAc = 1:1)

^{1}H NMR (600 MHz, $CDCl_3$) δ 8.05 (s, 1H), 7.56 (s, 1H), 7.25 (d, *J* = 9.1 Hz, 2H), 6.55 (d, *J* = 8.4 Hz, 2H), 6.32 (t, *J* = 6.8 Hz, 1H), 4.67 (d, *J* = 7.6 Hz, 1H), 4.1-4.20 (m, 1H), 4.12-4.13 (m, 1H), 3.97-3.99 (m, 2H), 3.82-3.86 (m, 2H), 3.75-3.78 (m, 2H), 2.31 (t, *J* = 5.8 Hz, 2H), 1.95 (s, 3H), 0.95 (s, 9H), 0.94 (s, 9H), 0.93 (s, 9H), 0.80 (s, 9H), 0.14 (s, 3H), 0.13 (s, 3H), 0.12 (s, 3H), 0.11 (s, 3H), 0.09 (s, 6H), -0.12 (s, 3H), -0.38 (s, 3H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 163.5, 150.2, 146.1, 135.5, 130.9, 128.5, 113.3, 111.1, 86.0, 85.7, 84.9, 83.1, 79.5, 73.9, 63.9, 63.8, 54.1, 39.4, 29.7, 26.2, 26.1, 26.1, 26.0, 18.6, 18.6, 18.3, 18.1, 12.7, -4.3, -4.3, -5.0, -5.2, -5.2, -5.2, -5.3 ppm.

HRMS (ESI-TOF): calc'd for $C_{45}H_{83}N_3O_8Si_4$ ($[M+H]^+$) 905.5257, found 906.5334.

*Characterization of **52b** (Ligation product DMA adduct)*

R_f = 0.2 (hexanes:EtOAc = 1:1)

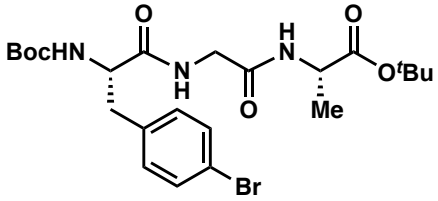
^{1}H NMR (600 MHz, $CDCl_3$) δ 7.56-7.60 (m, 1H), 7.23-7.25 (m, 2H), 6.54-6.55 (m, 2H), 6.30-6.36 (m, 1H), 5.49-5.61 (m, 2H), 4.67 (d, *J* = 7.4 Hz, 1H), 4.12-4.19 (m, 2H), 3.96-3.99 (m, 3H), 3.82-3.86 (m, 2H), 3.74-3.79 (m, 2H), 3.00 (s, 0.43H), 2.87-2.88 (m, 2H), 2.43 (s, 2H), 2.28-2.43 (m, 2H), 2.07 (s, 0.60 H), 1.96 (s, 3H), 0.93-0.95 (m, 30H), 0.80 (s, 9H), -0.08-0.04 (m, 18H), -0.12 (s, 3H), -0.38 (s, 3H) ppm.

^{13}C NMR (151 MHz, $CDCl_3$) δ 171.9, 171.2, 163.7, 163.6, 151.1, 150.8, 146.0, 134.4, 134.1, 130.9, 128.5, 128.5, 113.3, 110.3, 110.2, 86.1, 85.9, 85.7, 85.6, 85.6, 85.5, 83.1, 83.0, 79.5, 73.9, 63.9, 63.7, 55.5, 54.1, 54.0, 52.6, 39.5, 35.2, 31.7, 26.2, 26.1, 26.1, 26.0, 22.2, 21.5, 18.6, 18.5, 18.2, 18.1, 13.4, -4.3, -4.3, -4.3, -5.0, -5.0, -5.2, -5.2, -5.3, -5.4 ppm.

HRMS (ESI-TOF): calc'd for $C_{49}H_{91}N_4O_9Si_4$ ($[M+H]^+$) 991.5863, found 991.5861.

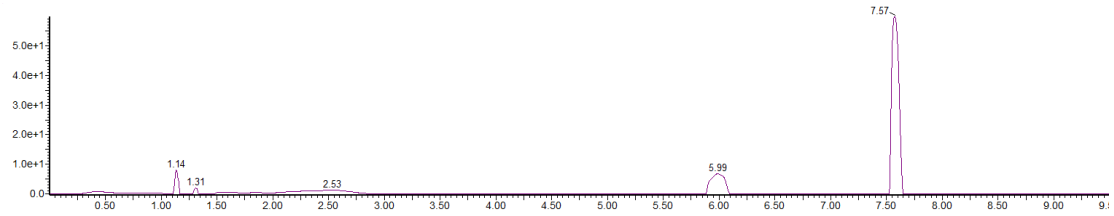
Table 4-B – C-N bond formation on oligopeptides

Peptide SI-2

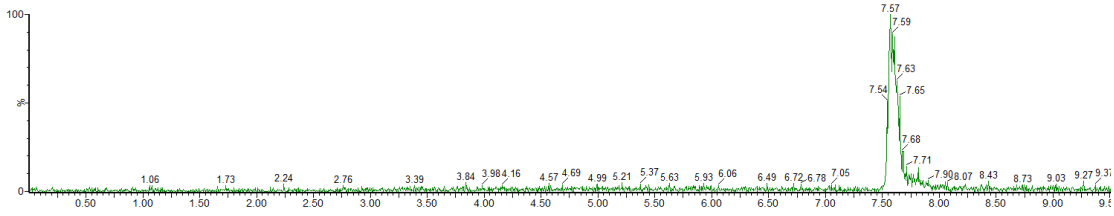


Peptide **SI-1** was synthesized according to a reported procedure on 2 mmol scale.[19] The crude Peptide **SI-2** was purified by preparative reverse-phase HPLC method Narrow 5 to afford peptide **SI-2** (642 mg, 60%) as a white solid following lyophilization.

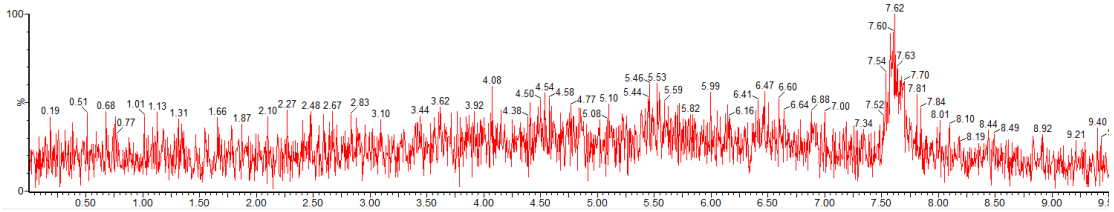
HPLC trace

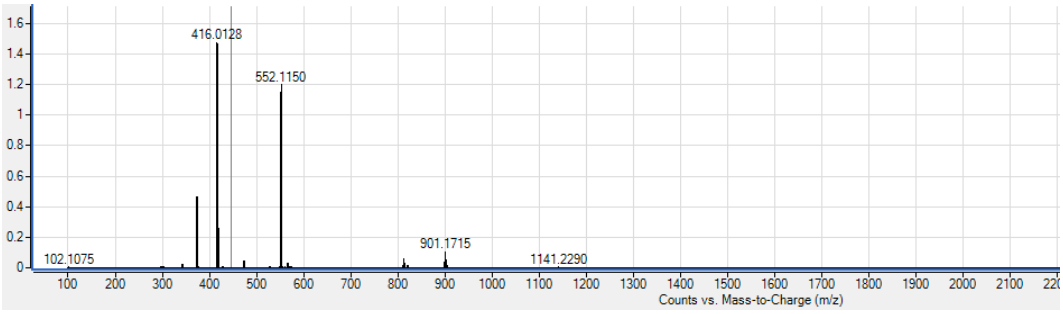


ESI$^-$



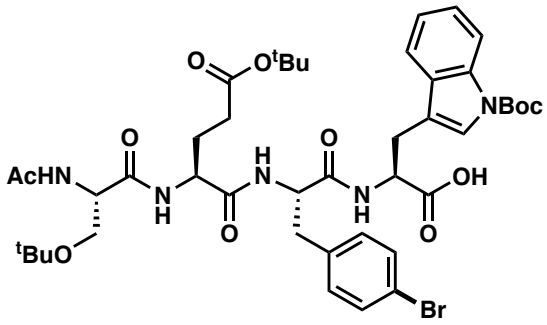
ESI$^+$





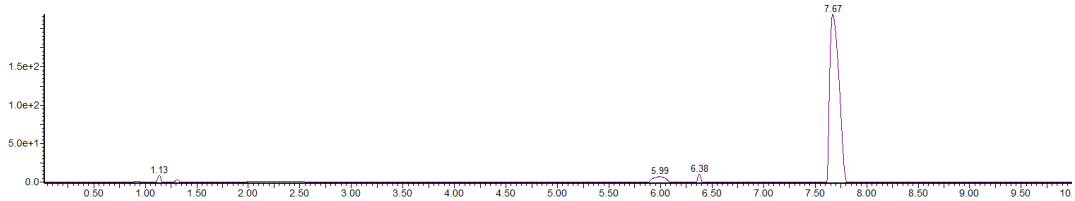
LRMS (ESI-TOF): calc'd for $C_{23}H_{34}BrN_3O_6Na$ $[M+Na]^+$ 552.16; found 552.12.

Peptide SI-3

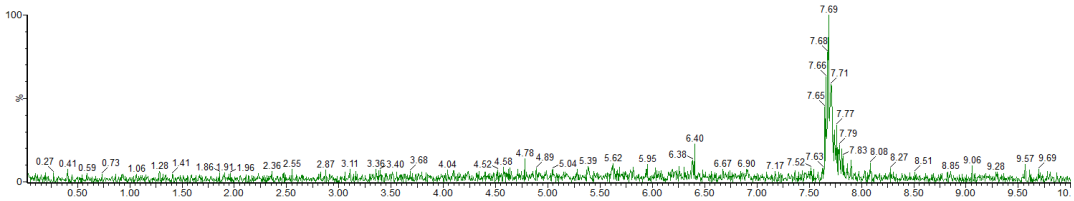


Peptide **SI-3** was prepared on a 300 µmol scale on 2-CTC resin using standard Fmoc-SPPS according to the general procedure. Following cleavage from the resin and ether precipitation, the crude peptide was purified by preparative reverse-phase HPLC method Narrow 5 to afford peptide **SI-3** (110 mg, 41% yield based on the original resin loading) as a white solid following lyophilization.

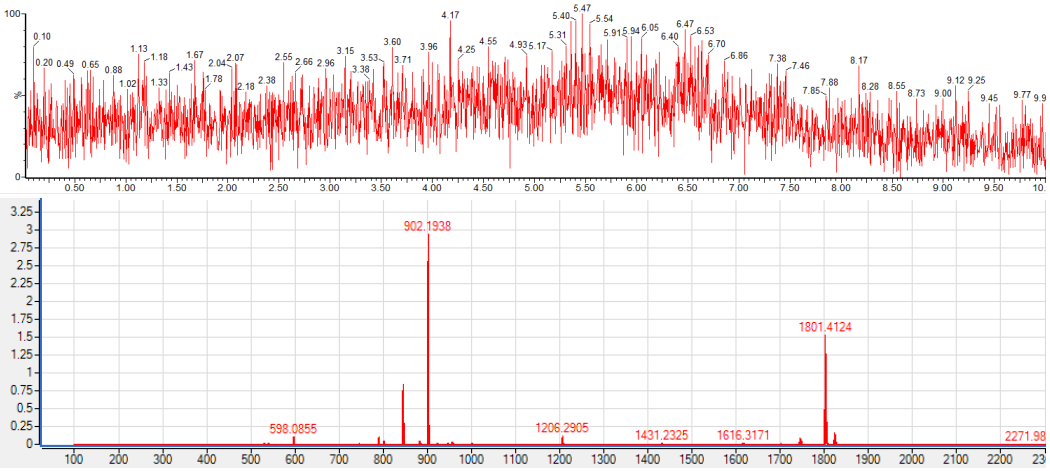
HPLC trace



ESI$^-$

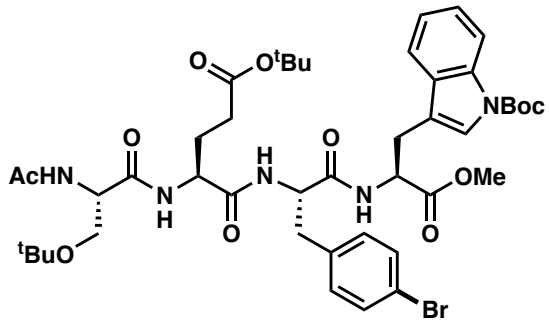


ESI$^+$



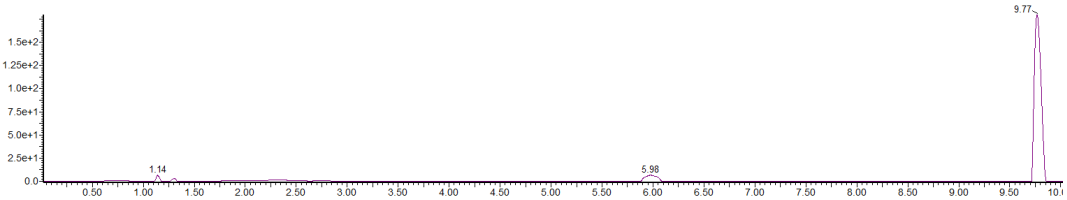
LRMS (ESI-TOF): calc'd for $C_{43}H_{59}BrN_5O_{11}$ $[M+H]^+$ 900.34; found 900.31.

Peptide SI-4

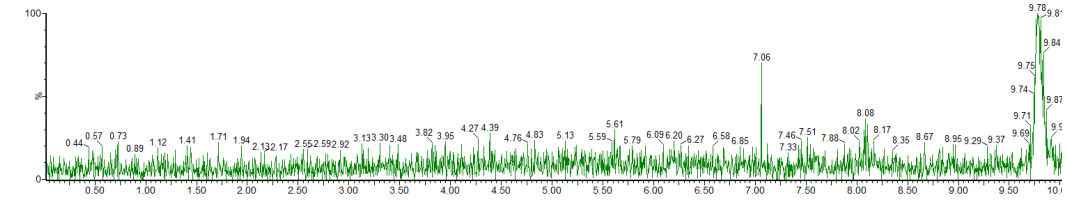


Peptide **SI-4** was prepared on a 300 µmol scale on 2-CTC resin using standard Fmoc-SPPS according to the general procedure esterification step added. Following cleavage from the resin and ether precipitation, the crude peptide was purified by preparative reverse-phase HPLC method Narrow 7 to afford peptide **SI-4** (103 mg, 31% yield based on the original resin loading) as a white solid following lyophilization.

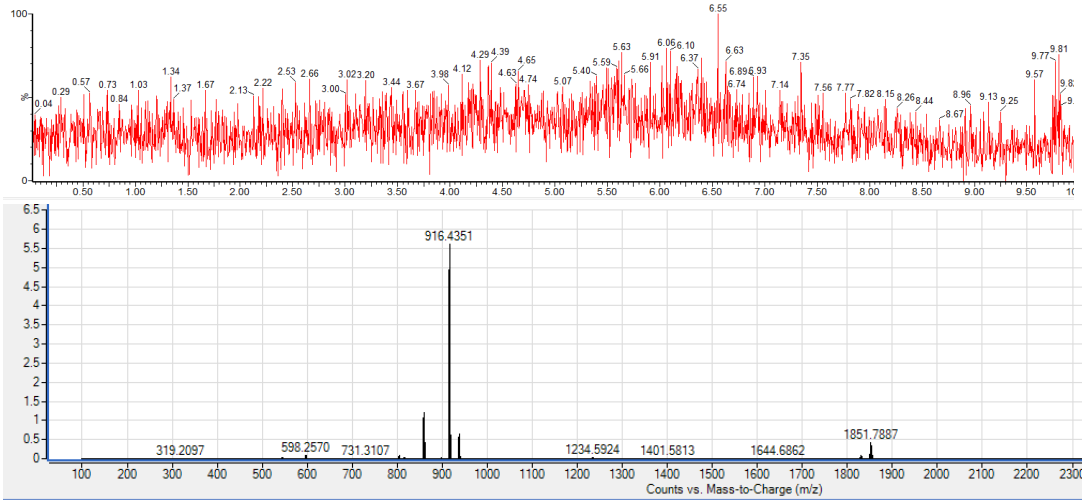
HPLC trace



ESI-

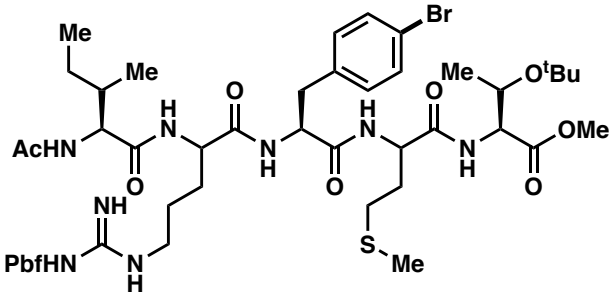


ESI+



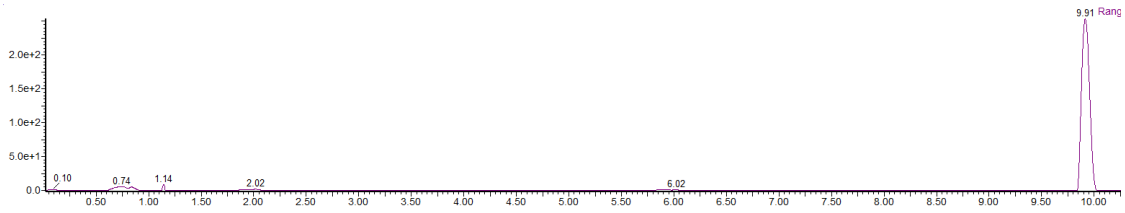
LRMS (ESI-TOF): calc'd for $C_{43}H_{61}BrN_5O_{11}$ $[M+H]^+$ 914.36; found 914.40.

Peptide SI-5

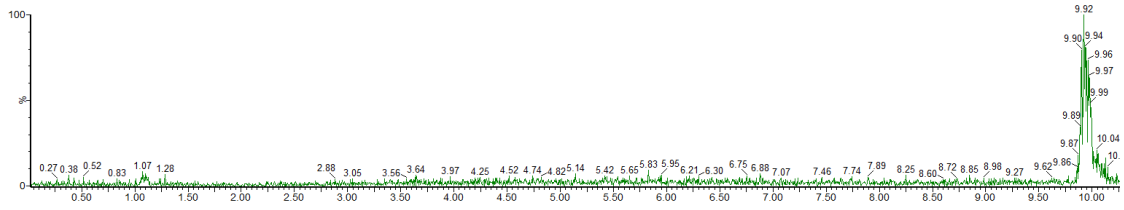


Peptide **SI-5** was prepared on a 300 μmol scale on 2-CTC resin using standard Fmoc-SPPS according to the general procedure esterification step added. Following cleavage from the resin and ether precipitation, the crude peptide was purified by preparative reverse-phase HPLC method Narrow 7 to afford peptide **SI-5** (153 mg, 46% yield based on the original resin loading) as a white solid following lyophilization.

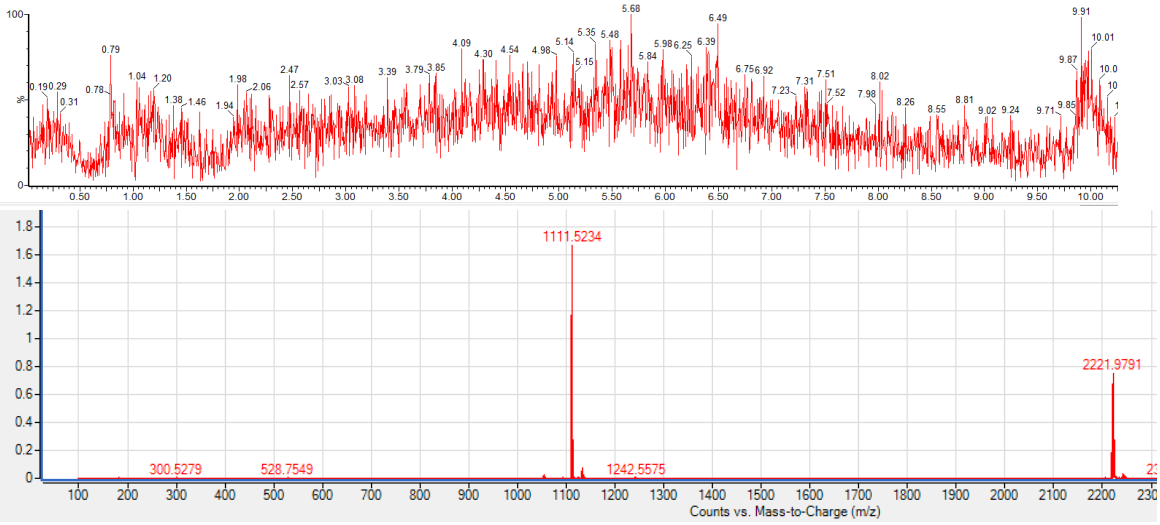
HPLC trace



ESI$^-$

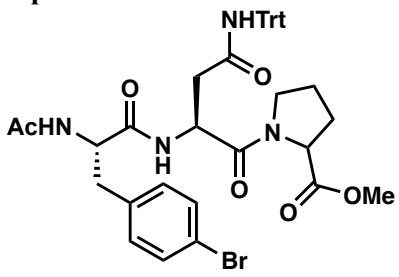


ESI$^+$



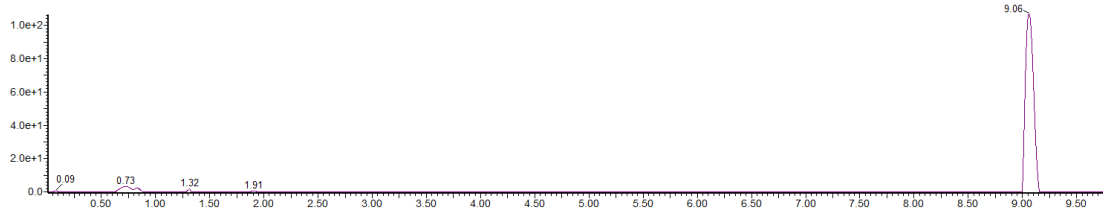
LRMS (ESI-TOF): calc'd for $C_{50}H_{78}BrN_8O_{11}S_2$ $[M+H]^+$ 1109.43; found 1109.32.

Peptide SI-6

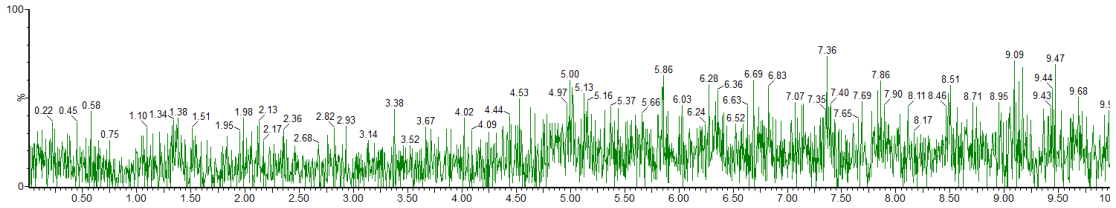


Peptide **SI-56** was prepared on a 300 μmol scale on 2-CTC resin using standard Fmoc-SPPS according to the general procedure esterification step added. Following cleavage from the resin and ether precipitation, the crude peptide was purified by preparative reverse-phase HPLC method Narrow 7 to afford peptide **SI-6** (126 mg, 56% yield based on the original resin loading) as a white solid following lyophilization.

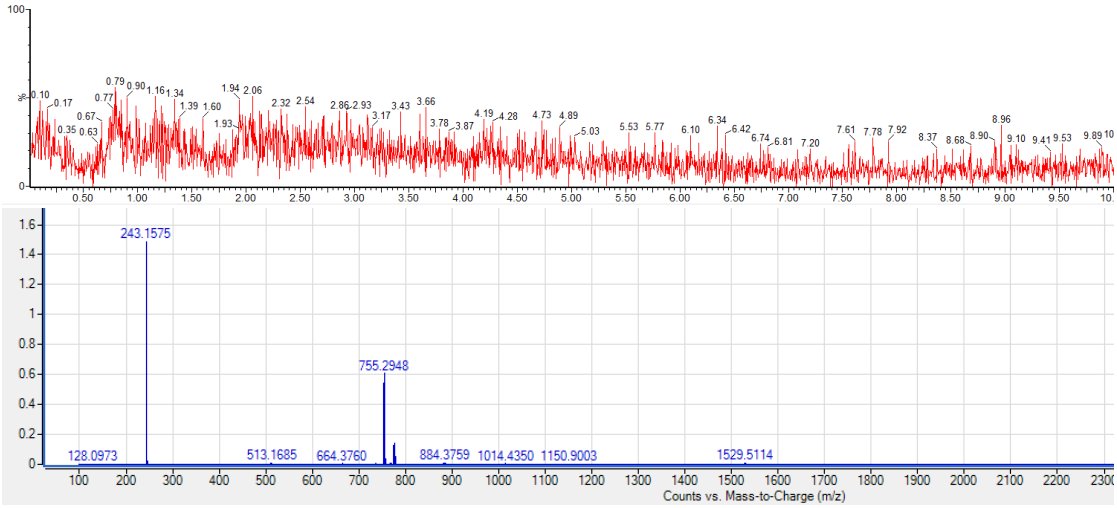
HPLC trace



ESI$^-$

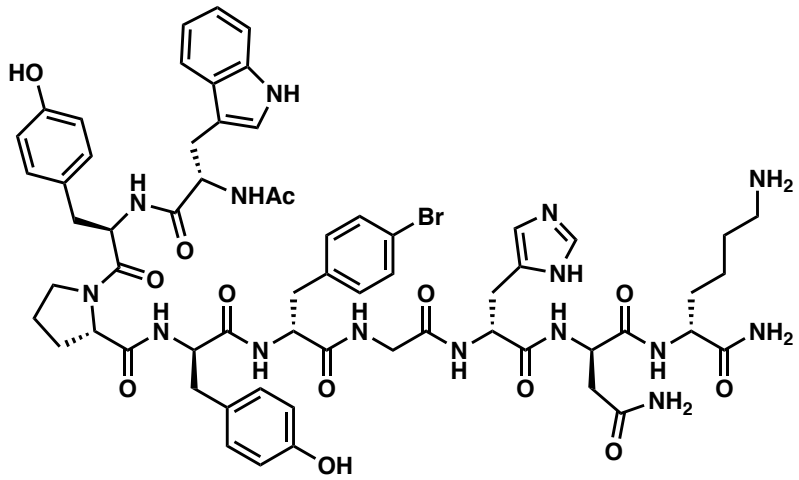


ESI+



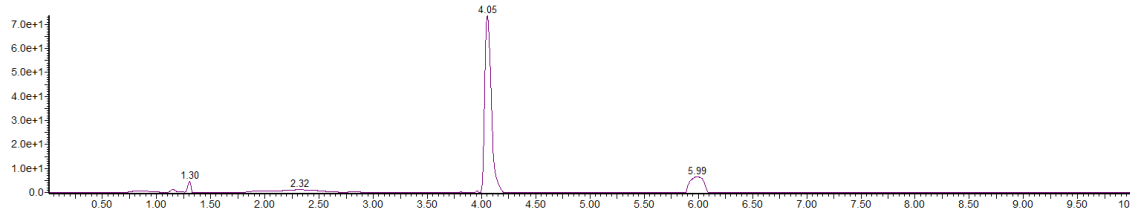
LRMS (ESI-TOF): calc'd for $C_{50}H_{78}BrN_8O_{11}S_2$ $[M+H]^+$ 753.23; found 753.24.

Peptide SI-7

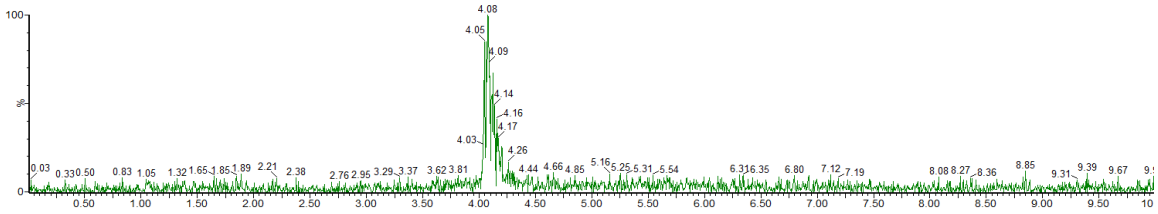


Peptide **SI-7** was provided by Pfizer and was prepared on Rink amide resin using standard Fmoc-SPPS strategy. HPLC and LCMS traces are described below.

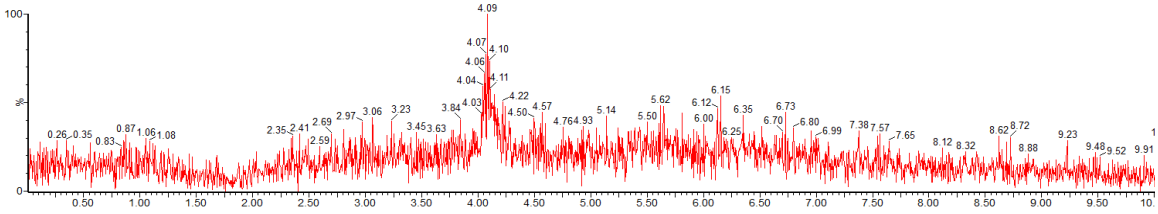
HPLC trace

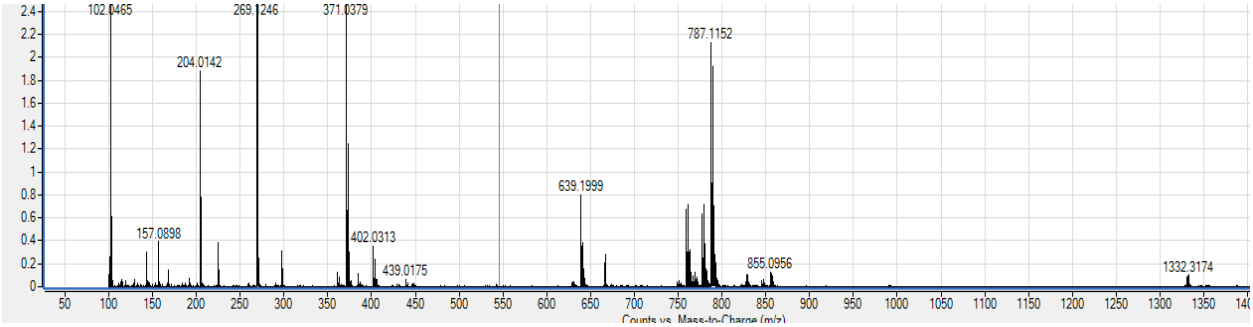


***ESI*$^{-}$**



***ESI*$^{+}$**

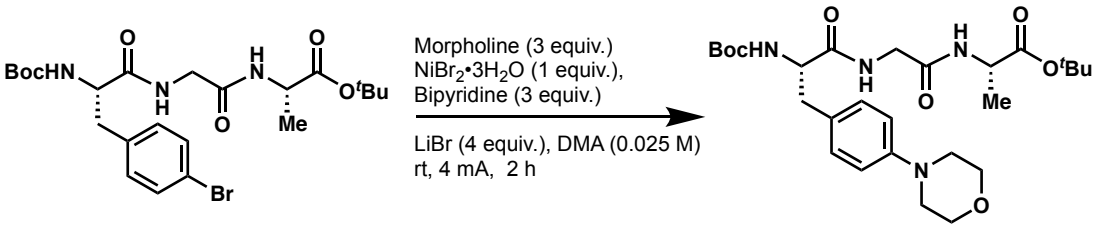




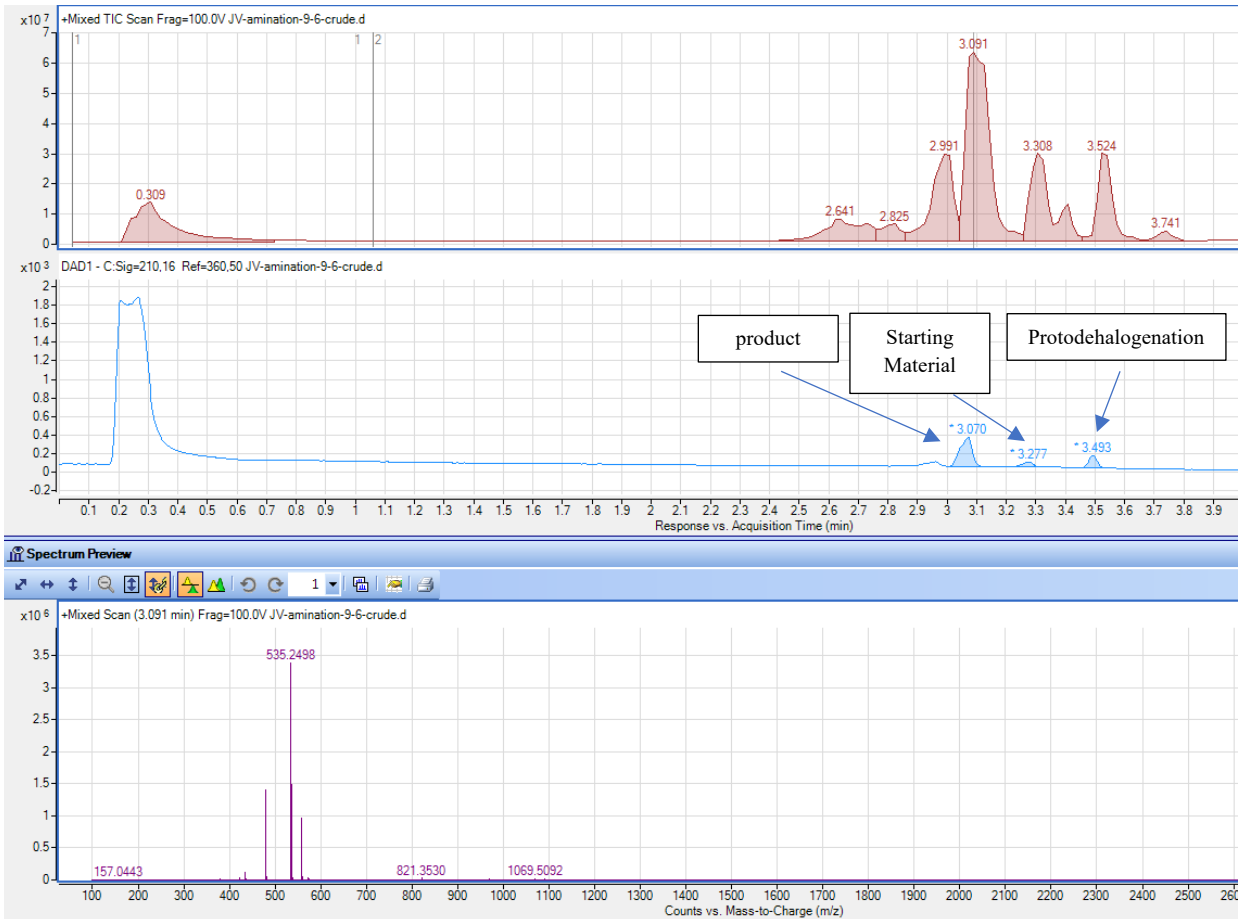
LRMS (ESI-TOF): calc'd for $C_{63}H_{77}BrN_{15}O_{13}$ $[M+H]^+$ 1330.50; found 1330.53.

Peptide – Optimization table

The optimization of the coupling reaction between morpholine and peptide **SI-2** was conducted following the general procedure for peptides.



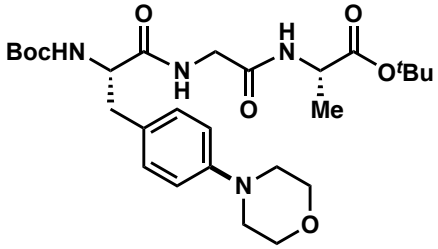
The crude reaction mixtures were analyzed by LCMS and the conversion to the desired product was based on relative integration.



The table below summarized the deviations from standard conditions that led the optimized set of condition:

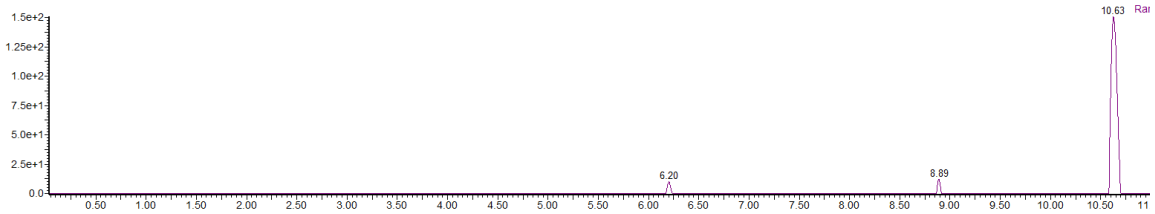
#	Deviation from standard conditions	Yield %
1	none	37 (32)
2	8 equiv. amine	51
3	2 equiv LiBr	29
4	8 equiv LiBr	40
5	20 equiv LiBr	38
6	8 mA, 1h	29
7	2 mA, 8h	36
8	4 mA, 4h	56
9	4 mA, 6h	63 (57)
10	8 equiv LiBr, 4 mA, 6h	61
11	30 mol% [Ni], 4 mA, 6h	57 (51)
12	30 mol% [Ni] complex, 4 mA, 6h	61 (55)

Compound 51

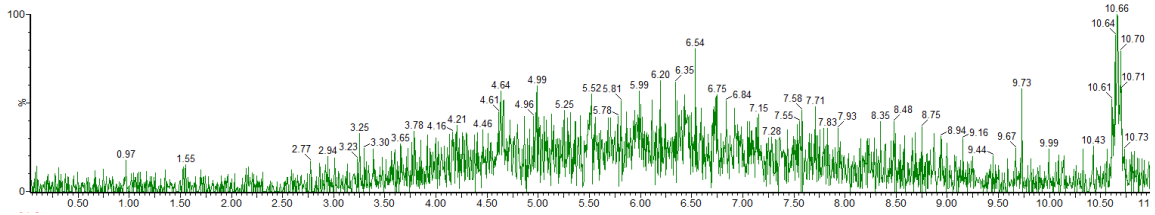


Morpholine (0.15 mmol), peptide **SI-2** (0.05 mmol). Electrolysis was conducted for 6 h following the general peptide procedure. The crude peptide was purified by preparative reverse-phase HPLC method Narrow 7 to afford peptide **53** (13.6 mg, 51%) as a white solid following lyophilization.

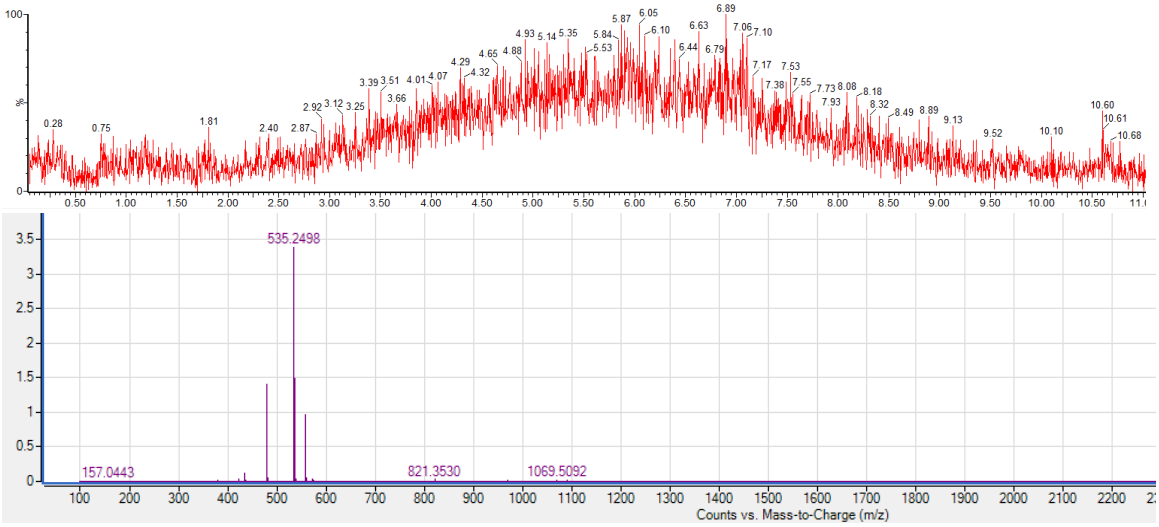
HPLC trace



ESI$^-$

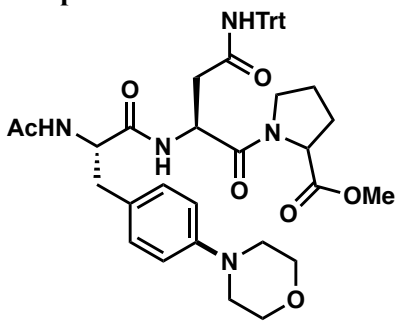


ESI$^+$



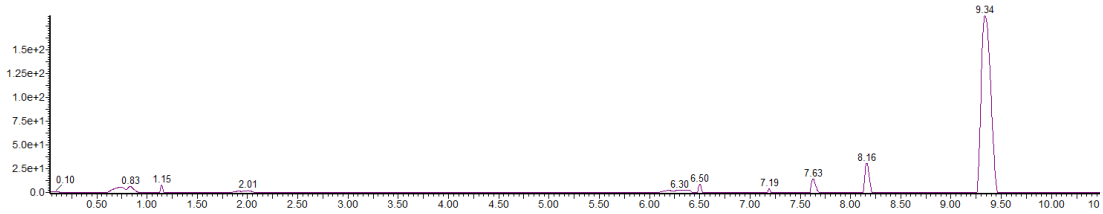
LRMS (ESI-TOF): calc'd for $C_{27}H_{43}N_4O_7$ $[M+H]^+$ 535.31; found 535.25.

Compound 52

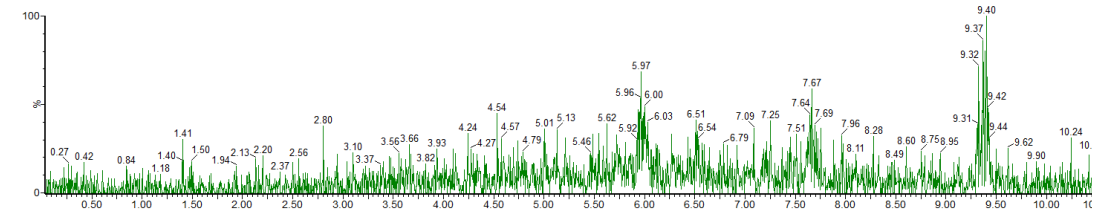


Morpholine (0.15 mmol), peptide **SI-6** (0.05 mmol). Electrolysis was conducted for 6 h following the general peptide procedure. The crude peptide was purified by preparative reverse-phase HPLC method Narrow 7 to afford peptide **54** (21.5 mg, 57%) as a white solid following lyophilization.

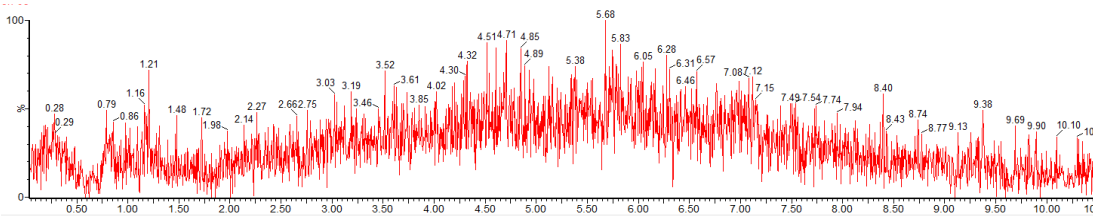
HPLC trace

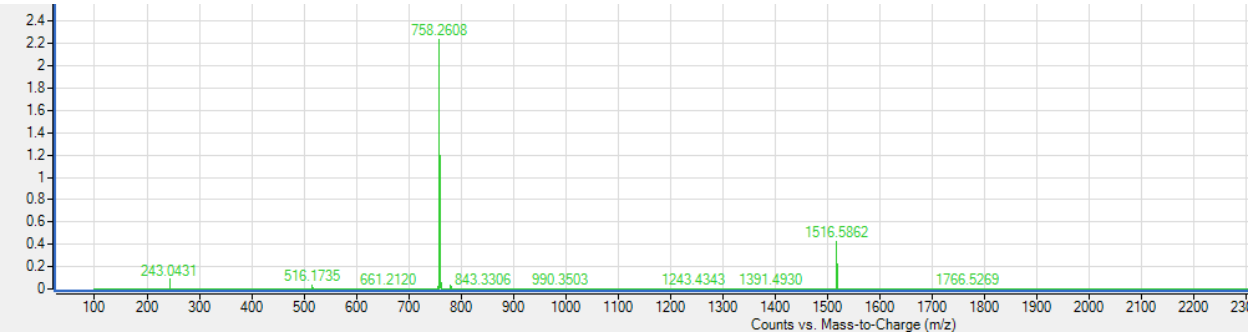


ESI$^-$



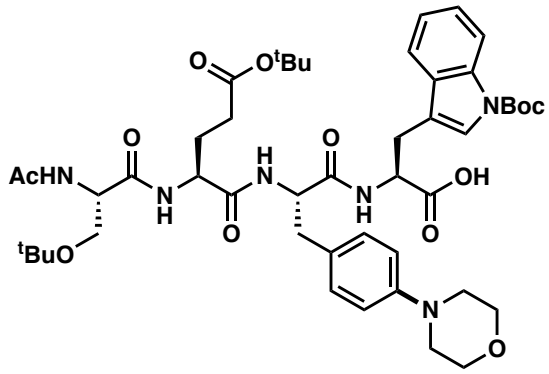
ESI$^+$





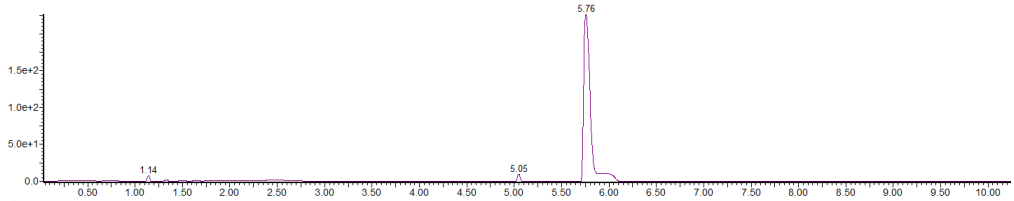
LRMS (ESI-TOF): **calc'd for $C_{44}H_{50}N_5O_7$ $[M+H]^+$ 758.28; found 758.26.**

Compound 53

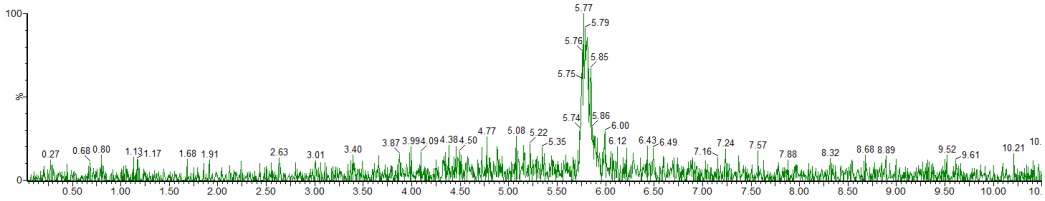


Morpholine (0.15 mmol), peptide **SI-3** (0.05 mmol). Electrolysis was conducted for 6 h following the general peptide procedure. The crude peptide was purified by preparative reverse-phase HPLC method Narrow 5 to afford peptide **55** (10.6 mg, 23%) as a white solid following lyophilization.

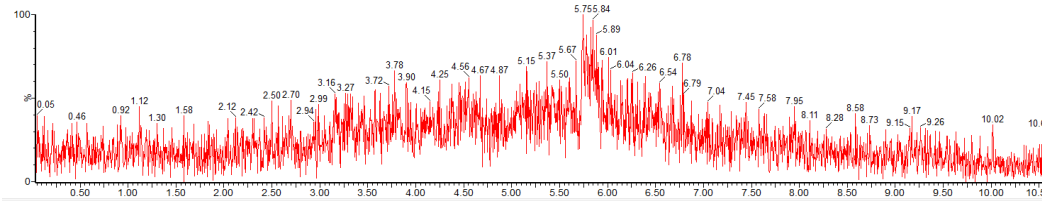
HPLC trace

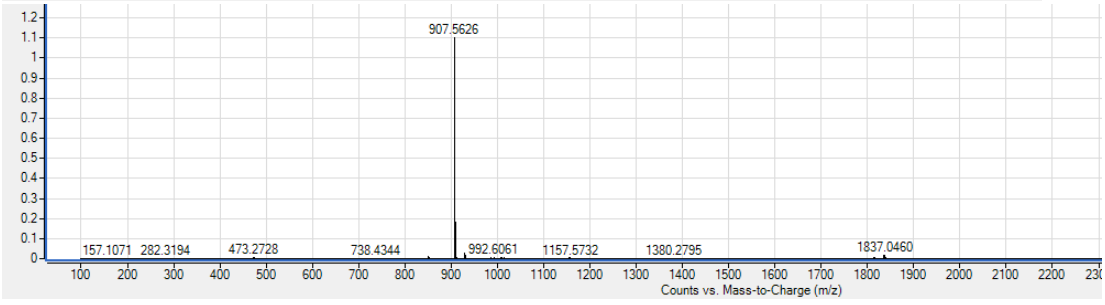


ESI$^-$



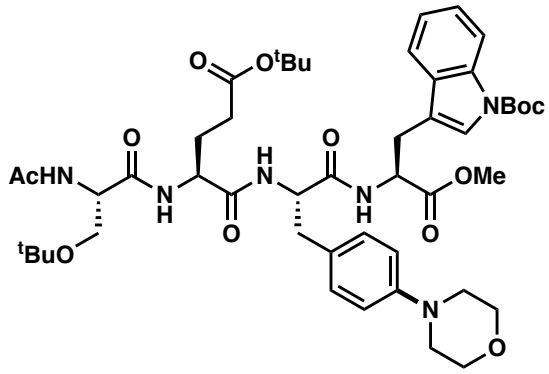
ESI$^+$





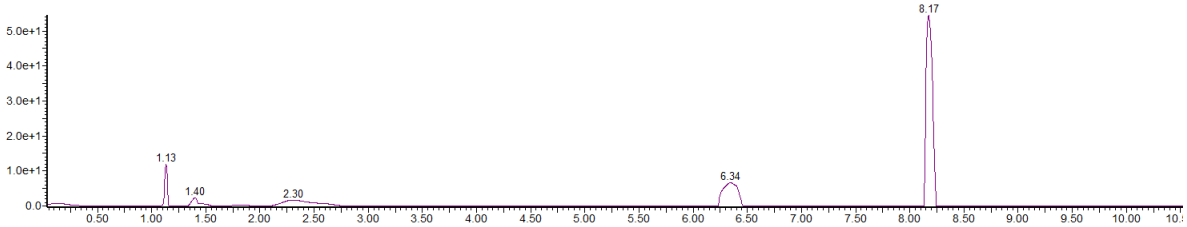
LRMS (ESI-TOF): calc'd for $C_{47}H_{67}N_6O_{12}$ $[M+H]^+$ 907.48; found 907.56.

Compound 54

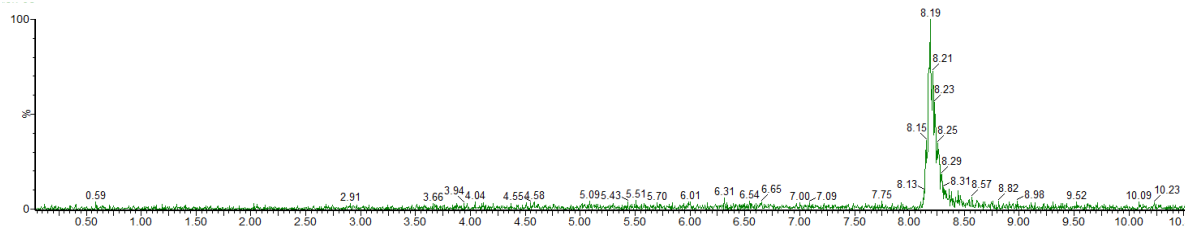


Morpholine (0.15 mmol), peptide **SI-4** (0.05 mmol). Electrolysis was conducted for 6 h following the general peptide procedure. The crude peptide was purified by preparative reverse-phase HPLC method Narrow 6 to afford peptide **56** (14.7 mg, 32%) as a white solid following lyophilization. <u>*Note:*</u> *Peptide **56** was obtained in 45% when using 100 mol% [Ni].*

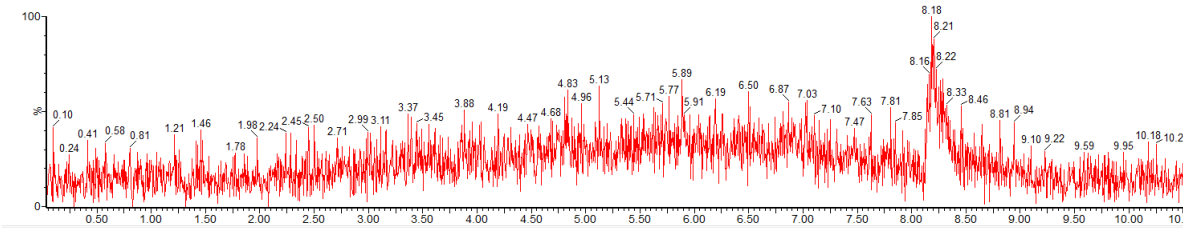
HPLC trace

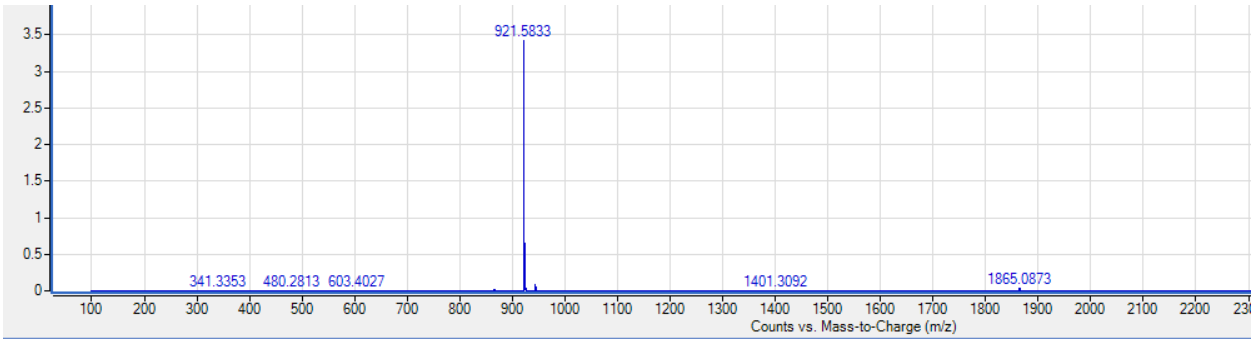


ESI$^-$



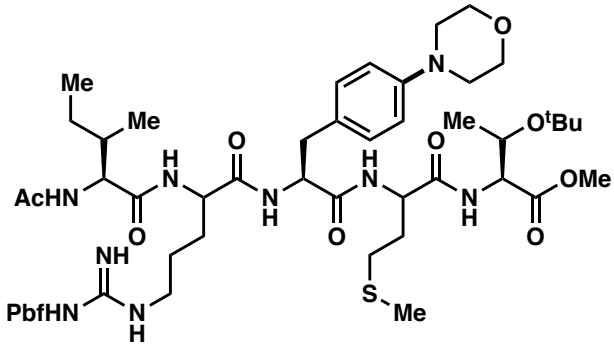
ESI$^+$





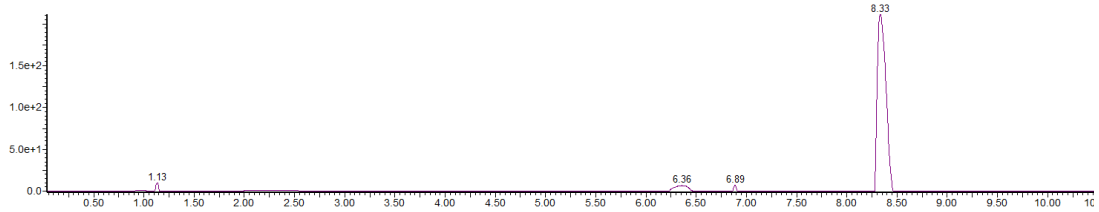
LRMS (ESI-TOF): calc'd for $C_{48}H_{69}N_6O_{12}$ $[M+H]^+$ 921.60; found 921.58.

Compound 55

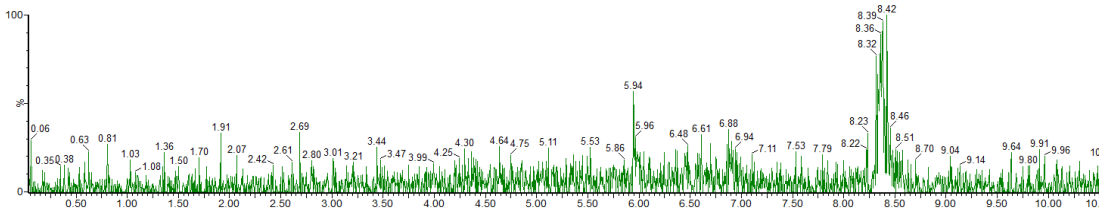


Morpholine (0.15 mmol), peptide **SI-5** (0.05 mmol). Electrolysis was conducted for 6 h following the general peptide procedure. The crude peptide was purified by preparative reverse-phase HPLC method Narrow 6 to afford peptide **57** (21.7 mg, 39%) as a white solid following lyophilization.

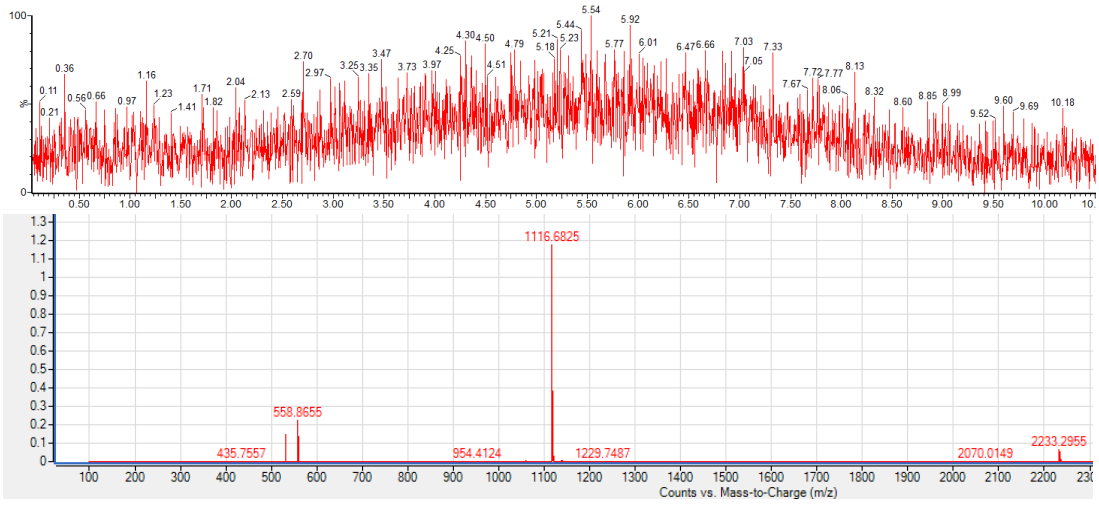
HPLC trace



ESI$^-$

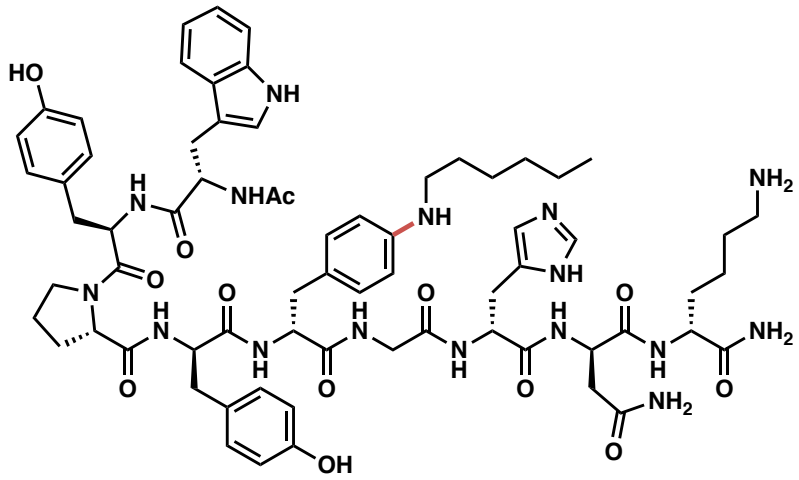


ESI$^+$



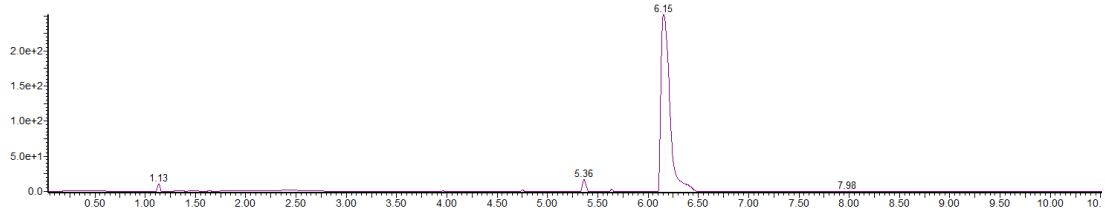
LRMS (ESI-TOF): calc'd for $C_{54}H_{88}N_9O_{12}S_2$ $[M+H]^+$ 1116.56; found 1116.58.

Compound 56

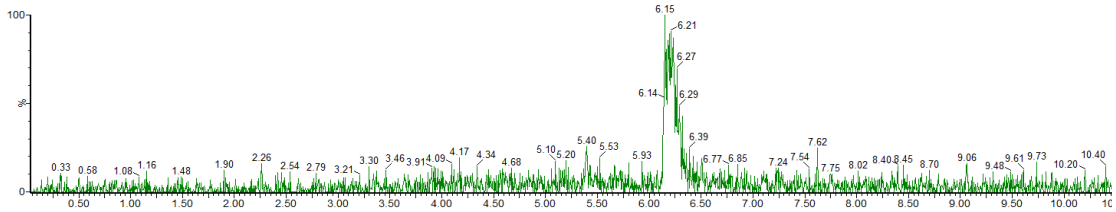


Hexalamine (3mmol), peptide **SI-7** (0.05 mmol), [Ni] (100 mol%). Electrolysis was conducted for 6 h following the general peptide procedure. The crude peptide was purified by preparative reverse-phase HPLC method Narrow 6 to afford peptide **58** (7.4 mg, 11%) as a white solid following lyophilization.

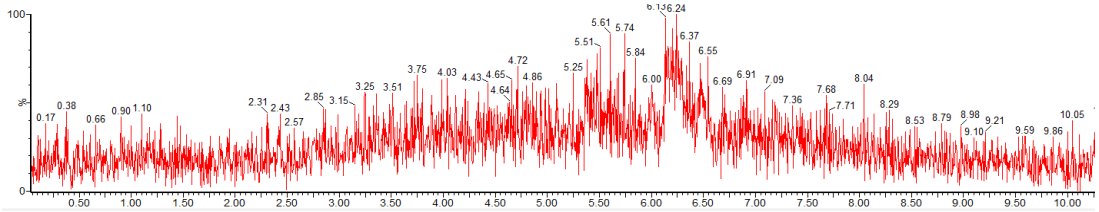
HPLC trace

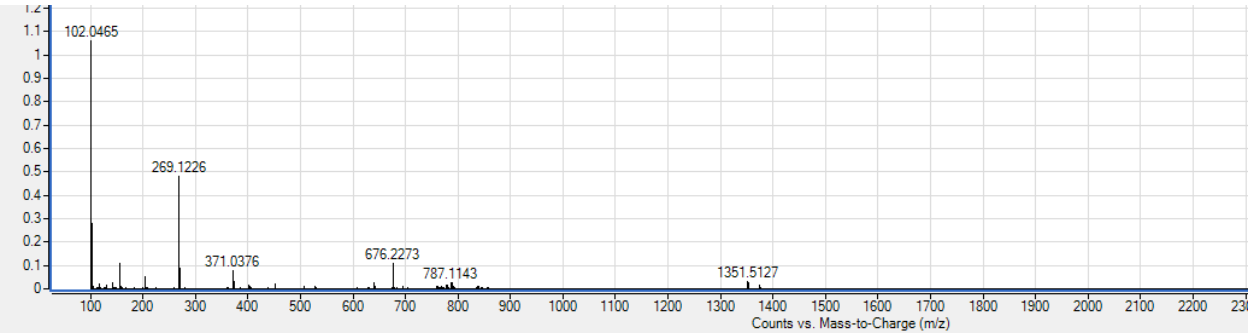


ESI$^-$



ESI$^+$



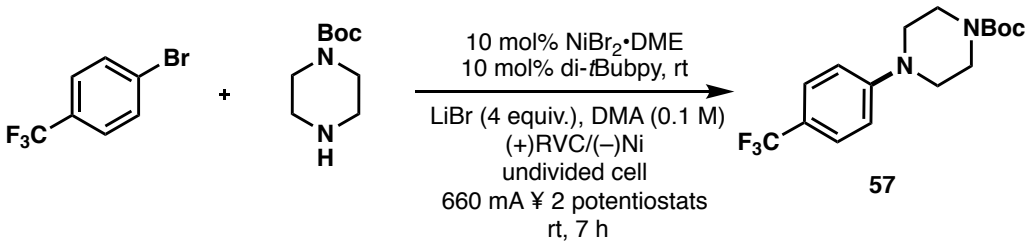


LRMS (ESI-TOF): calc'd for $C_{44}H_{50}N_5O_7$ $[M+H]^+$ 1351.70; found 1351.51.

Figure 4-C – Large scale amination

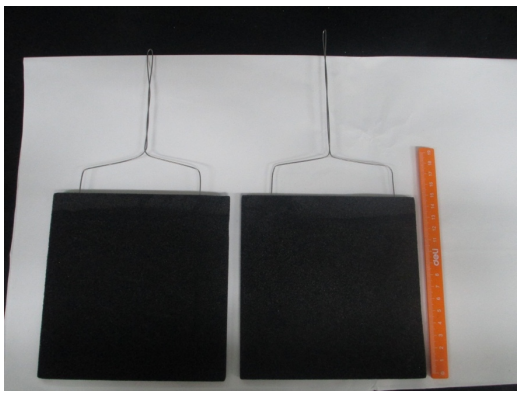
Experimental detail of 22.5 g scale reaction – Compound 57

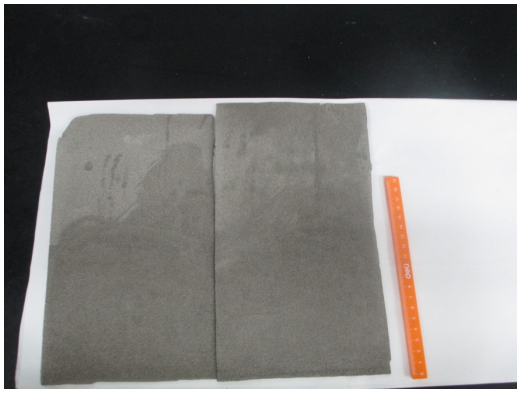
Procedure of decagram scale reaction (Performed at Asymchem)



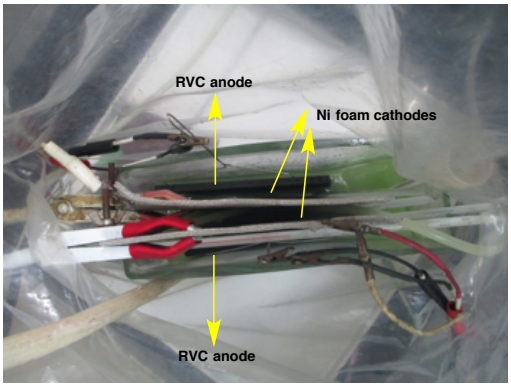
To a clean and dry glass chamber was added DMA (1.5 L), LiBr (52 g, 600 mmol, 4 equiv.), $NiBr_2$•DME (3.08g, 10 mmol, 10 mol%), 4,4'-di-*tert*-butyl-2,2'-bipyridine (2.68g, 10 mmol, 10 mol%), 1-Boc-piperazine (55.88g, 0.3mol, 3 equiv.), and 4-bromobenzotrifluoride (22.50g, 0.1 mol, 1 equiv.). The resulting suspension was stirred until the dissolution of all solids. The RVC anode (two plates) the Ni-foam cathode (two plates) were inserted into the solution; each set of anode/cathode was connected to a potentiostat (EZ-stat pro). The submerged surface area of each electrode was adjusted to 11cm × 15cm. The reaction mixture was deoxygenated by sparging with nitrogen for 30 mins when constant current electrolysis (660 mA for both potentiostats)* was conducted for 7 h under a nitrogen atmosphere. The resulting mixture was poured onto water (7.5 L) and extracted with MTBE (3.4 L × 3). The combined organic layer was concentrated *in vacuo*; the resulting residue was purified by flash column chromatography (silica gel) to afford the desired product **61** as a white solid (22.1 g, 66%).

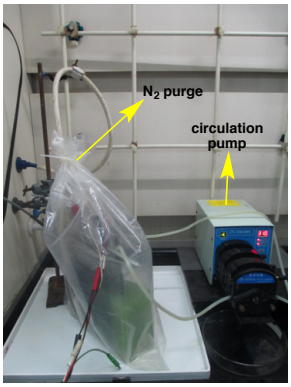
*Two potentiostats were used as the maximum current output of each unit is 1.0 A.



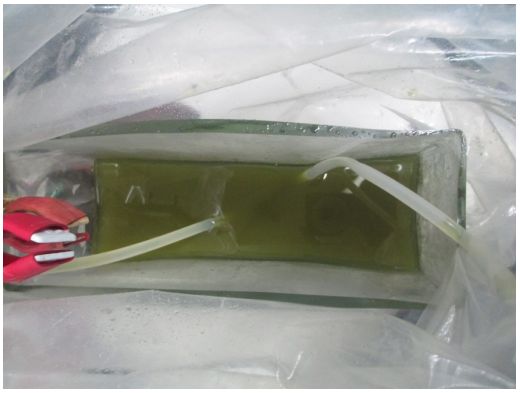


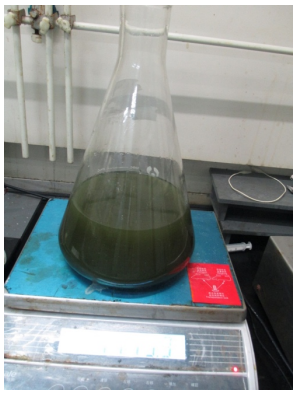
(***Left***) RVC anodes used in the reaction (2 plates, dimensions of each plate: 15 cm × 15 cm × 0.5 cm) (the submerged surface area was 11 cm × 15 cm for each plate). (***Right***) Ni-foam cathodes used in the reaction (2 plates, dimensions of each plate: 15 cm × 25 cm × 0.5 cm) (the submerged exterior surface area was 11 cm × 15 cm for each plate).



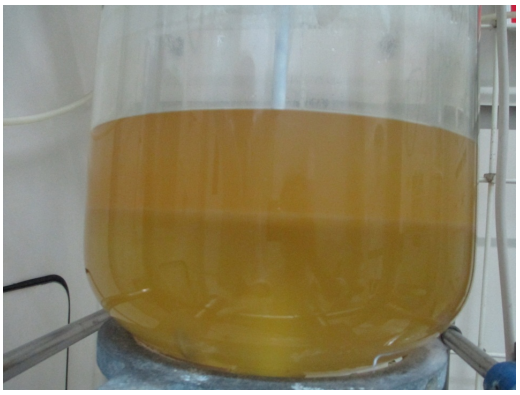


(***Left***) and (***Right***) Electrolysis setup.





(***Left***) and (***Right***) Reaction mixture after the electrolysis.

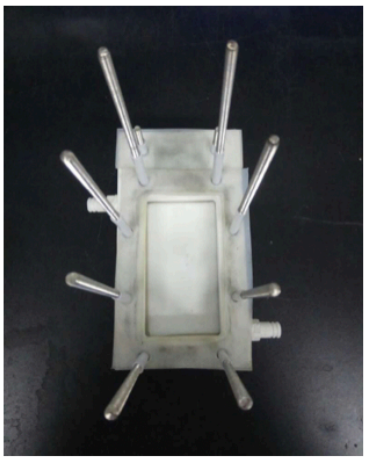


(***Left***) and (***Right***) Extraction of the reaction mixture with MTBE.

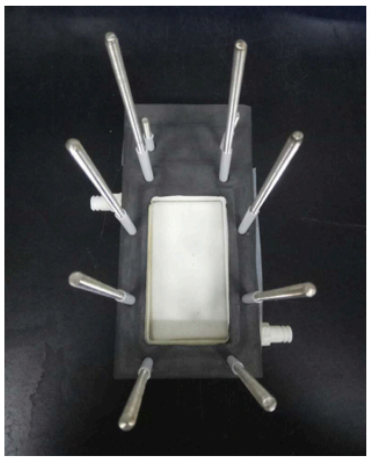
Experimental detail of 100 g scale reaction – Compound 36

Multi-Frame Cell setup:

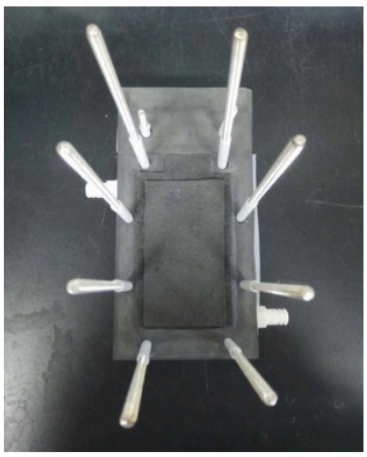
10 stainless steel screws with cap (length: 25.0 cm, diameter: 5.0 mm) were threaded through a base part of multi-frame cell constituted from one Teflon plate (length: 18.0 cm, width: 12.0 cm, thickness: 2.0 cm, with eight holes each has a diameter of 1.0 cm), one Teflon frame block (length: 22.0 cm, width: 12.0 cm, thickness: 2.0 cm, with ten holes each has a diameter of 1.0 cm) and a silicone pad (length: 18.0 cm, width: 12.0 cm, thickness: 2.0 mm, with ten holes each has a diameter of 1.0 cm) between them. The base part with stainless steel screws was then added components topped each layer followed a sequence of silicone pad, graphite plate (as electrode cover, length: 25.0 cm, width: 12.0 cm, thickness: 2.0 mm, with ten holes each has a diameter of 1.0 cm), carbon felt (as electrode, both anode and cathode, length: 15.0 cm, width: 9.0 cm, thickness: 5.0 mm, with an immersion surface area of 13.0 cm×7.0 cm), silicone pad, Teflon frame and carbon felt to form one single working cell. Repeated the adding sequence until a multi-frame cell with total four working cells were packed, ended by putting another base part on the very top layer. The multi-frame cell with a total of 6 frame blocks, 2 Teflon plates, 5 graphite covers, 5 carbon felts and 12 silicone pads were then locked by 10 screw nuts tightened with appropriate force to complete the entire construction. Each side of Teflon frame was screwed a Teflon joint which connected to a rubber tube (6 mm in diameter), the upper joint of four middle frames was connected to the lower joint of its sideward frame, and the joint of two terminal frames was connected to rubber tubes (6 mm in diameter) which through two Teflon tubes (6 mm in diameter) linked with peristaltic pump (as in flow) and external reservoir (as out flow).



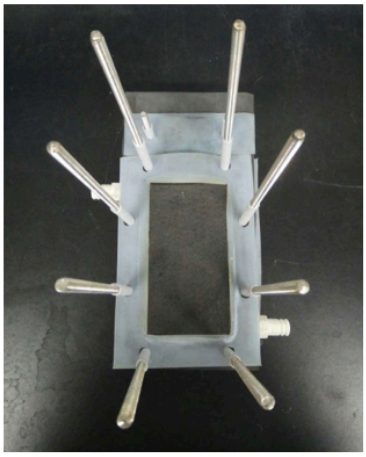
Step 1
Silicon plate on frame



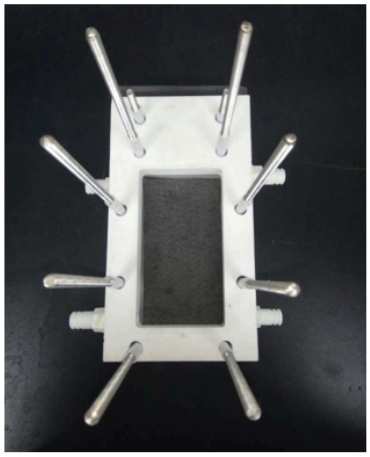
Step 2
Graphite plate on top



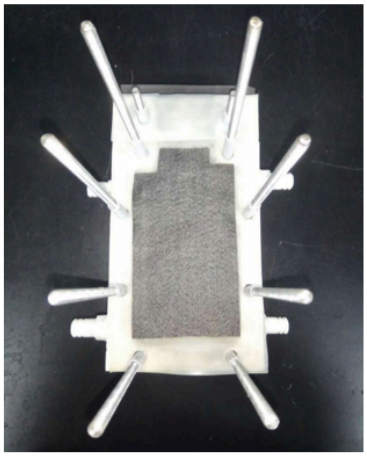
Step 3
Carbon felt on top



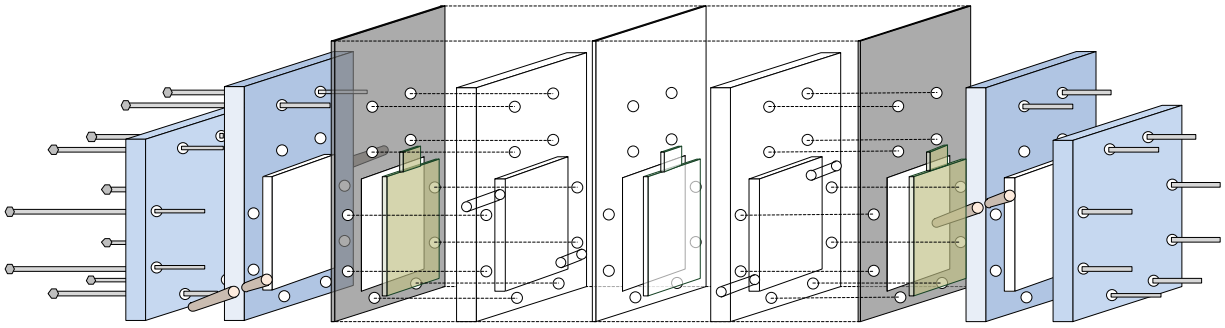
Step 4
Silicon plate on top



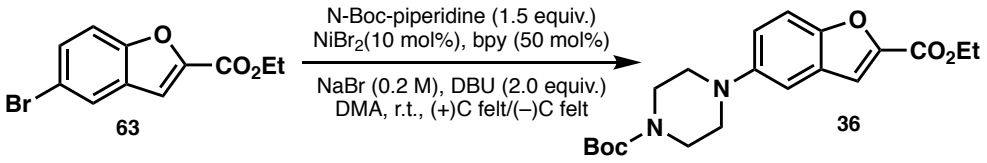
Step 5
Frame on top



Step 6
Carbon felt on top



Experimental procedure：



A clean and dry 5.0 L Erlenmeyer flask with a stir bar was charged with $NiBr_2$ (37.2 mmol, 8.13 g, 10 mol%), DMA (3.0 L) was then added and stirred vigorously at room temperature for 10 min until all solid were dissolved and obtained a green color solution. 2,2'-bipyridinyl (186 mmol, 29.0 g, 50 mol%) was added and stirred for another 30 min until solution turn to a pink color. NaBr (0.2 M, 600 mmol, 61.8 g), DBU (744 mmol, 113.1 g, 2.0 equiv.), 5-bromo-benzofuran-2-carboxylic acid ethyl ester **63** (372 mmol, 100.0 g, 1.0 equiv.) and 1-t-Butoxycarbonylpiperazine (558 mmol, 104.3 g, 1.5 equiv.) were added successively under stirring, the obtained solution followed by transferring into a 3.0 L four-necked round bottom as external reservoir. A peristaltic pump was connected to multi-frame cell (4 working cells) and external reservoir (3.0 L four-necked round bottom) by a peristaltic tube (length: 19.0 cm, thickness: 0.8 mm) and two Teflon tubes (length: 15.0 cm to the multi-frame cell and 45.0 cm to the external reservoir, diameter: 6.0 mm) formed a circulatory system. The loop system was then purged with nitrogen for 10 minutes followed by bubbling the reaction mixture with nitrogen for 20 minutes. The reaction mixture was then pumped into multi-frame cell with a flow rate of 500 rpm from external reservoir, set an automatic switch of polarity on all electrodes every 5.0 minutes and electrolyzed under a constant current of 7.2 A provided from a direct current power until the complete consumption of 5-bromo-benzofuran-2-carboxylic acid ethyl ester judged by HPLC. After reaction, all mixture was drove into a 5.0 L Erlenmeyer flask from both multi-frame cell and external reservoir, followed by adding DMA (2x 2.0 L) to the loop and circulated for 10 min to wash the multi-frame cell and flush the residue in tubes twice. The combined solution was poured into three 5.0 L four-necked round bottom with 4.0 L ice water in each and blended vigorously by a two-blade mechanic stir for 30 min. The precipitate was then filtered by a Buchner funnel, the filter cake was crude product as a light brown solid (109.0 g) and was purified by column chromatography (200-300 mesh silica gel, 25.0 cm height in column), washed with eluent (n-Hexane/EtOAc = 10/1) to afford desired product **36** as a light yellow solid (88.9 g, 64 % isolated yield).

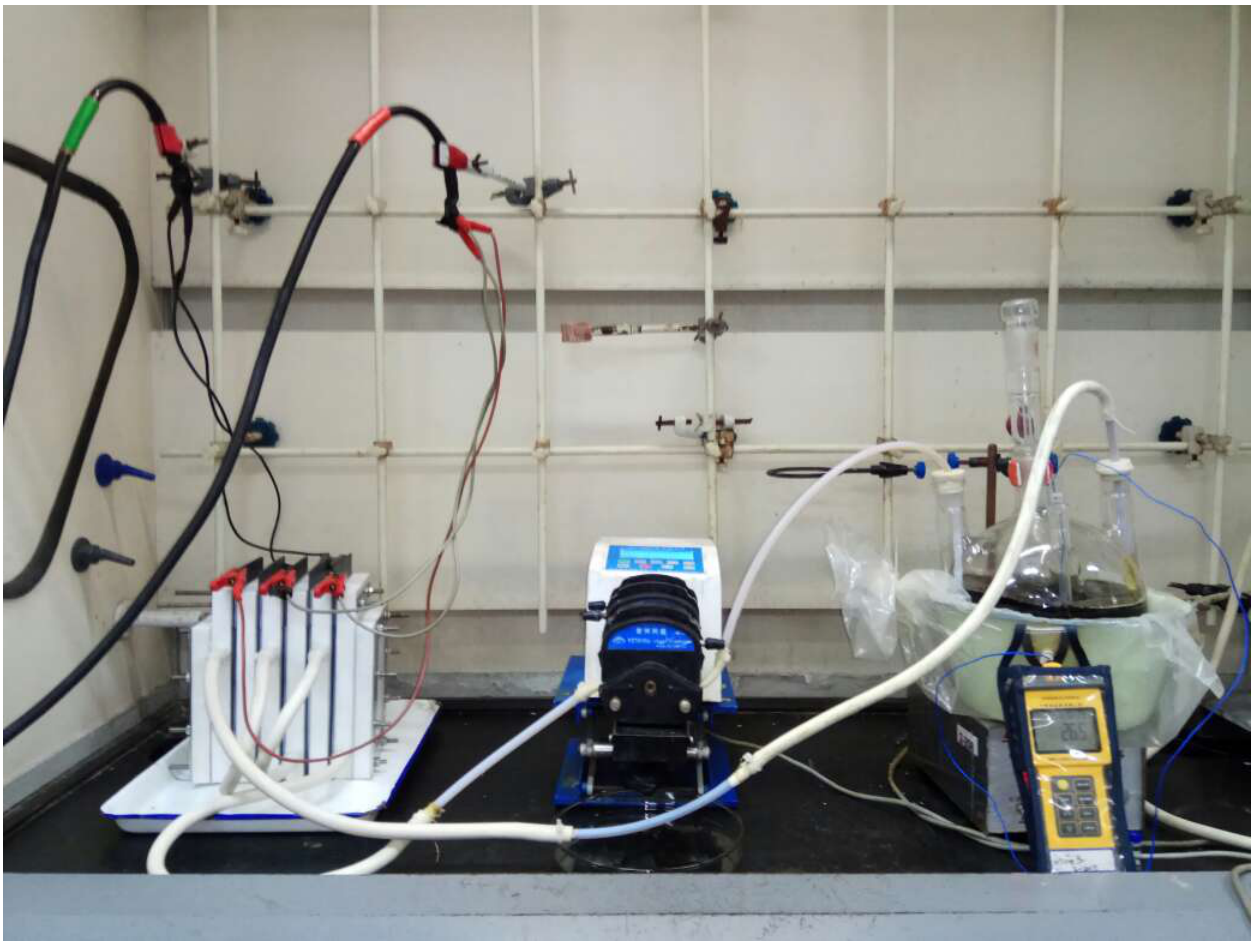
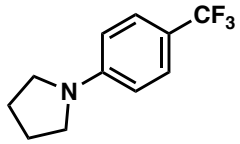


Table 5 – Applicability to previously successful substrates and limitations.

Compound 60



From aryl bromide

Pyrrolidine (0.3 mmol), 1-bromo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, Et_2O:pentane = 1:10) to give the **64** as a colorless oil (54% yield). The spectrum matched with the reported values.[20]

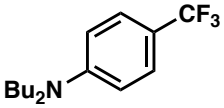
From aryl chloride

Pyrrolidine (0.3 mmol), 1-chloro-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, Et_2O:pentane = 1:10) to give the **64** as a colorless oil (69% yield).

From iodide

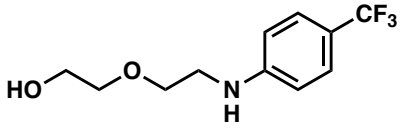
Pyrrolidine (0.3 mmol), 1-iodo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, Et_2O:pentane = 1:10) to give the **64** as a colorless oil (82% yield).

Compound 61



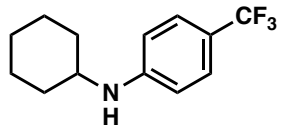
Dibutylamine (0.3 mmol), 1-bromo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, hexanes) to give **65** as a colorless liquid (61% yield). The spectrum matched with the reported values.[20]

Compound 62



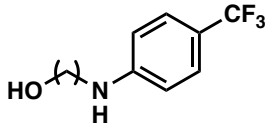
2-(2-aminoethoxy)ethan-1-ol (0.3 mmol), 1-bromo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, Et_2O) to give **66** as a colorless liquid (41% yield). The spectrum matched with the reported values.[20]

Compound 63



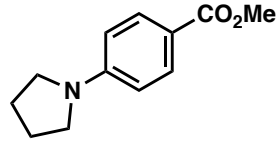
Cyclohexylamine (0.3 mmol), 1-bromo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, EtOAc:hexanes = 1:9) to give the **67** as a colorless liquid (78% yield). The spectrum matched with the reported values.[20]

Compound 64



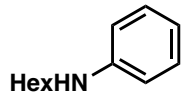
6-aminohexan-1-ol (0.3 mmol), 1-bromo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by flash column chromatography (silica gel, EtOAc:hexanes = 3:7) to give **68** as a colorless oil (53% yield). The spectrum matched with the reported values.[20]

Compound 65



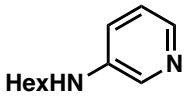
Pyrrolidine (0.3 mmol), methyl 4-(((trifluoromethyl)sulfonyl)oxy)benzoate (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, Et_2O:hexanes = 3:7) to give the **69** as a white solid (75% yield). The spectrum matched with the reported values.[20]

Compound 66



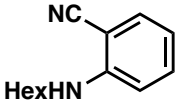
Hexalamine (0.3 mmol), bromophenyl (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, Et_2O:pentane = 1:15) to give **70** as a colorless liquid (76% yield). The spectrum matched with the reported values.[20]

Compound 67



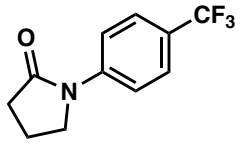
Hexalamine (0.3 mmol), 3-brompyridine (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by flash column chromatography (silica gel, EtOAc:hexanes = 1:3) to give **71** as a white solid (54% yield). The spectrum matched with the reported values.[20]

Compound 68



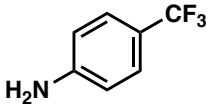
Hexalamine (0.3 mmol), 2-bromobenzonitrile (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, EtOAc:hexanes = 1:6) to give **72** as a colorless liquid (29 mg, 65% yield). The spectrum matched with the reported values.[20]

Compound 70



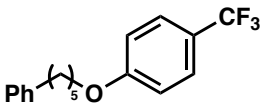
Pyrrolidin-2-one (0.3 mmol), 1-bromo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, EtOAc:hexanes = 2:3) to give **74** as a white solid (29 mg, 67% yield). The spectrum matched with the reported values.[20]

Compound 71



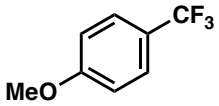
Ammonium hydroxide (2 mmol), 1-bromo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.6 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, EtOAc:hexanes = 1:1) to give **75** as a yellow solid (14% yield). The spectrum matched with the reported values.[21]

Compound 73



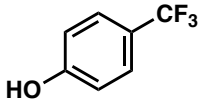
6-phenylhexan-1-ol (0.3 mmol), 1-bromo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.4 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, EtOAc:hexanes = 1:20) to give **77** as a colorless oil (37% yield). The spectrum matched with the reported values.[20]

Compound 74



Methanol (2 mmol), 1-bromo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.6 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, EtOAc:hexanes = 1:20) to give **78** as a white solid (32% yield). The spectrum matched with the reported values.[22]

Compound 75



Water (4 mmol), 1-bromo-4-(trifluoromethyl)benzene (0.2 mmol), DBU (0.6 mmol). Electrolysis was conducted for 4 h following the general procedure. The crude material was purified by PTLC (silica gel, EtOAc:hexanes = 1:2) to give **79** as a white solid (43% yield). The spectrum matched with the reported values.[23]

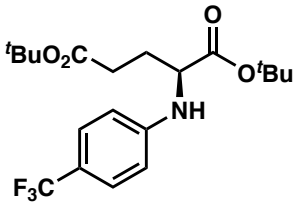
References

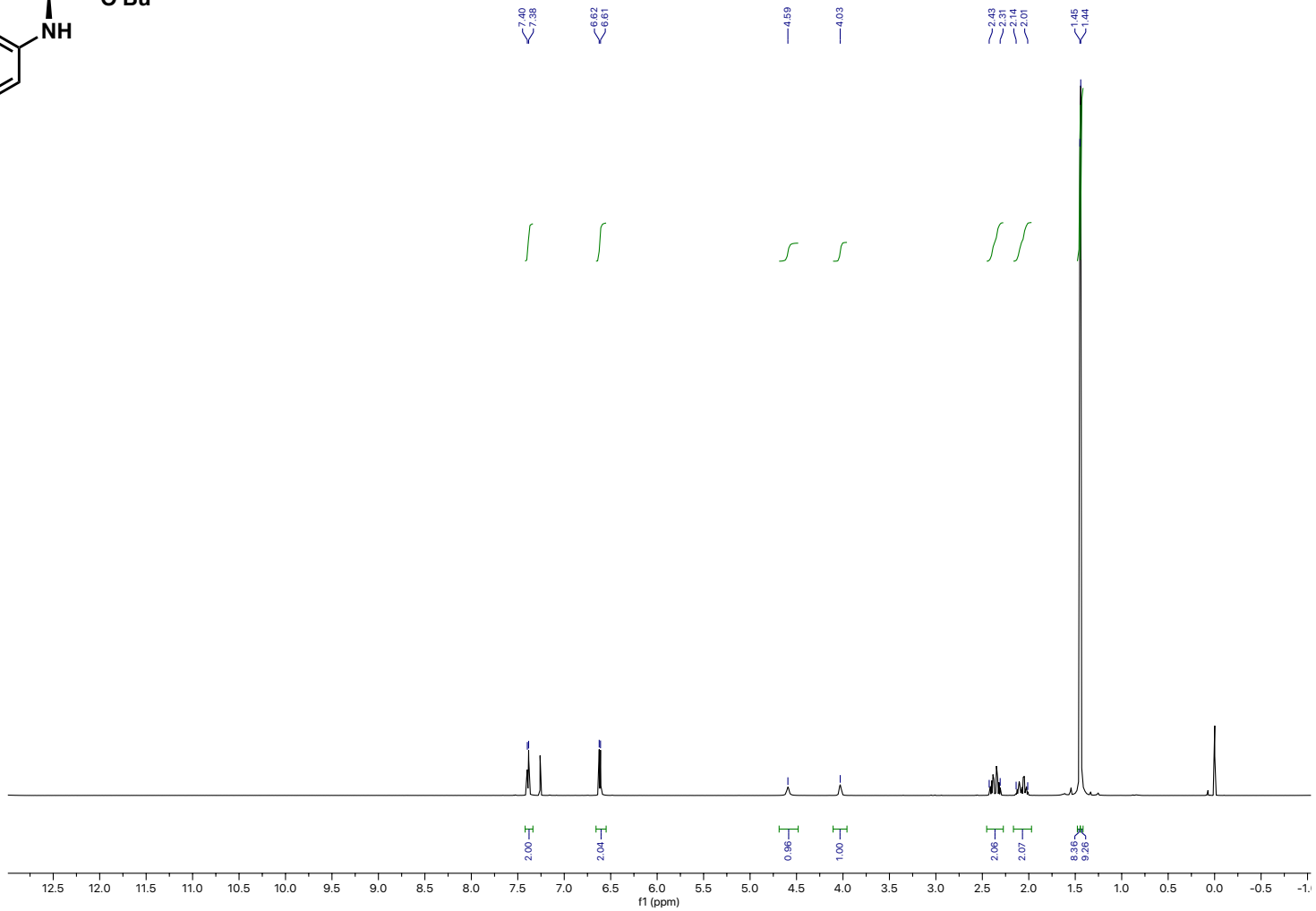
1. Varizhuk, A. M.; Kochetkova, S. V.; Kolganova, N. A.; Timofeev, E. N.; Florent'ev, V. L. *Russ. J. Bioorg. Chem.* **2010**, *36*, 199–206.
2. Štefko, M.; Pohl, R.; Klepetářová, B.; Hocek, M. *Eur. J. Org. Chem.* **2008**, *2008*, 1689–1704.
3. Amatore, C.; Azzabi, M.; Calas, P.; Jutand, A.; Lefrou, C.; Rollin, Y., *J. Electroanal. Chem. and Interfacial Electrochem.* **1990,** *288*, 45–63.
4. Denuault, G.; Mirkin, M. V.; Bard, A. J. *Eletroanal. Chem.* **1991**, *308*, 27–38.
5. Newman, J. *J. Electrochem. Soc.* **1966**, *113*, 501–502.
6. Gutierrez, O.; Tellis, J. C.; Primer, D. N.; Molander, G. A.; Kozlowski, M. C., *J. Am. Chem. Soc.* **2015,** *137*, 4896–4899.
7. Kalvet, I.; Guo, Q.; Tizzard, G. J.; Schoenebeck, F. *ACS Cat.* **2017,** *7*, 2126-2132.
8. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman, and D. J. Fox, Gaussian, Inc., Wallingford CT, **2016**.
9. Zhao, Y.; Truhlar, D. G. *J. Chem. Phys.* **2006**, *125*, 19410–194118.
10. Dolg, M.; Wedig, U.; Stoll, H.; Preuss, H. *J. Chem. Phys.* **1987**, *86*, 866–872.
11. Bergner, A.; Dolg, M.; Kuechle, W.; Stoll, H.; H, Preuss, H. *Mol. Phys.* **1993**, *80*, 1431–1441.
12. Marenich, A. V.; Cramer, C. J.; Truhlar, D. G. *J. Phys. Chem.* **2009**, *113*, 6378–6396.
13. Henkelman, G.; Uberuaga, B.P., Jónsson, H. J. *Chem. Phys.* **2000**, *113*, 9901–9904.
14. Henkelman, G.; Jónsson, H. *J. Chem. Phys.* **2000**, *113*, 9978–9985.
15. Marcus, Y. Ions in solutions and their solvation, **2015** John Wiley & Sons, Inc.
16. Chowdhury, D. R.; Spiccia, L.; Amritphale, S. S.; Paul, A.; Singh, A. *J. Mater. Chem. A*, **2016**, *4*, 3655–3660.
17. Hendrick, C. E.; Bitting, K. J.; Cho, S.; Wang, Q. *J. Am. Chem. Soc.* **2017**, *139*, 11622–11628.
18. Shao, Q.-L.; Jiang, Z.-J.; Su, W.-K. *Tetrahedron Lett.* **2018**, *59*, 2277–2280.
19. Sommerfeld, T. L.; Seebach, D. *Helv Chim Acta.* **2003**, *76*, 1702–1714.

20. Li, C.; Kawamata, Y.; Nakamura, H.; Vantourout, J. C.; Liu, Z.; Hou, Q.; Bao, D.; Starr, J. T.; Chen, J.; Yan, M.; Baran, P. S. *Angew. Chem. Int. Ed.* **2017**, *56*, 13088–13093.
21. Han-Sem, K. *Tetrahedron Lett.* **2018**, 59, 4597–4601.
22. Song-Lin, Z.; Wen-Feng, B. *RSC Advances*, **2016**, *6*, 70902–70906.
23. Gao, X.; Geng, Y.; Han, S.; Liang, A.; Li, J.; Zou, D.; Wu, Y.; Wu, Y. *Org. Lett.* **2018**, *20*, 3732–3735.

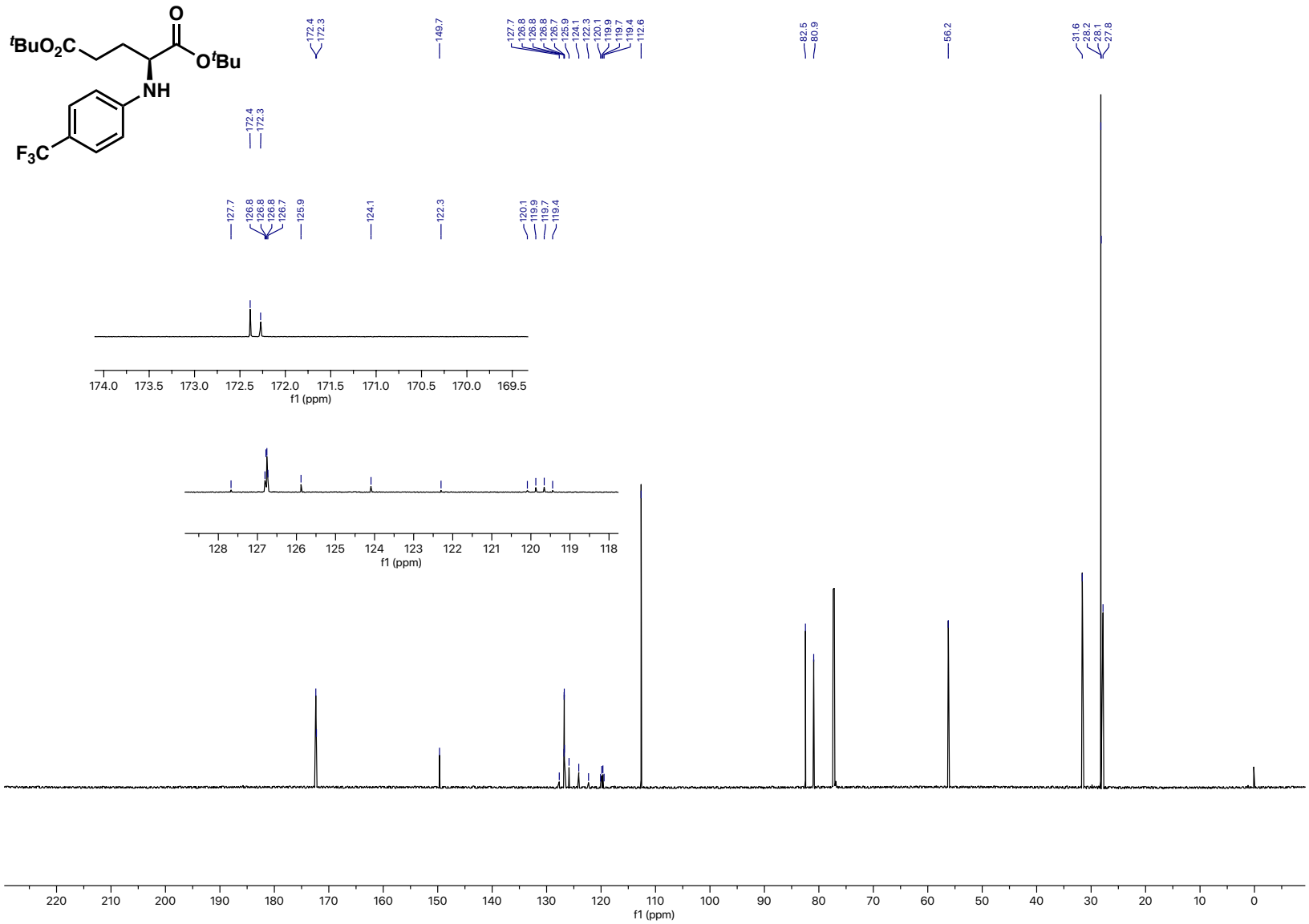
Compounds Spectra

Compound 15 ^{1}H NMR

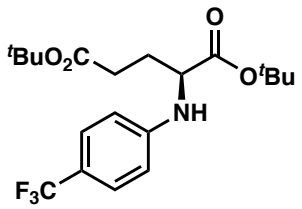




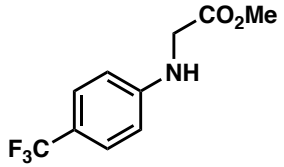
Compound 15 ^{13}C NMR



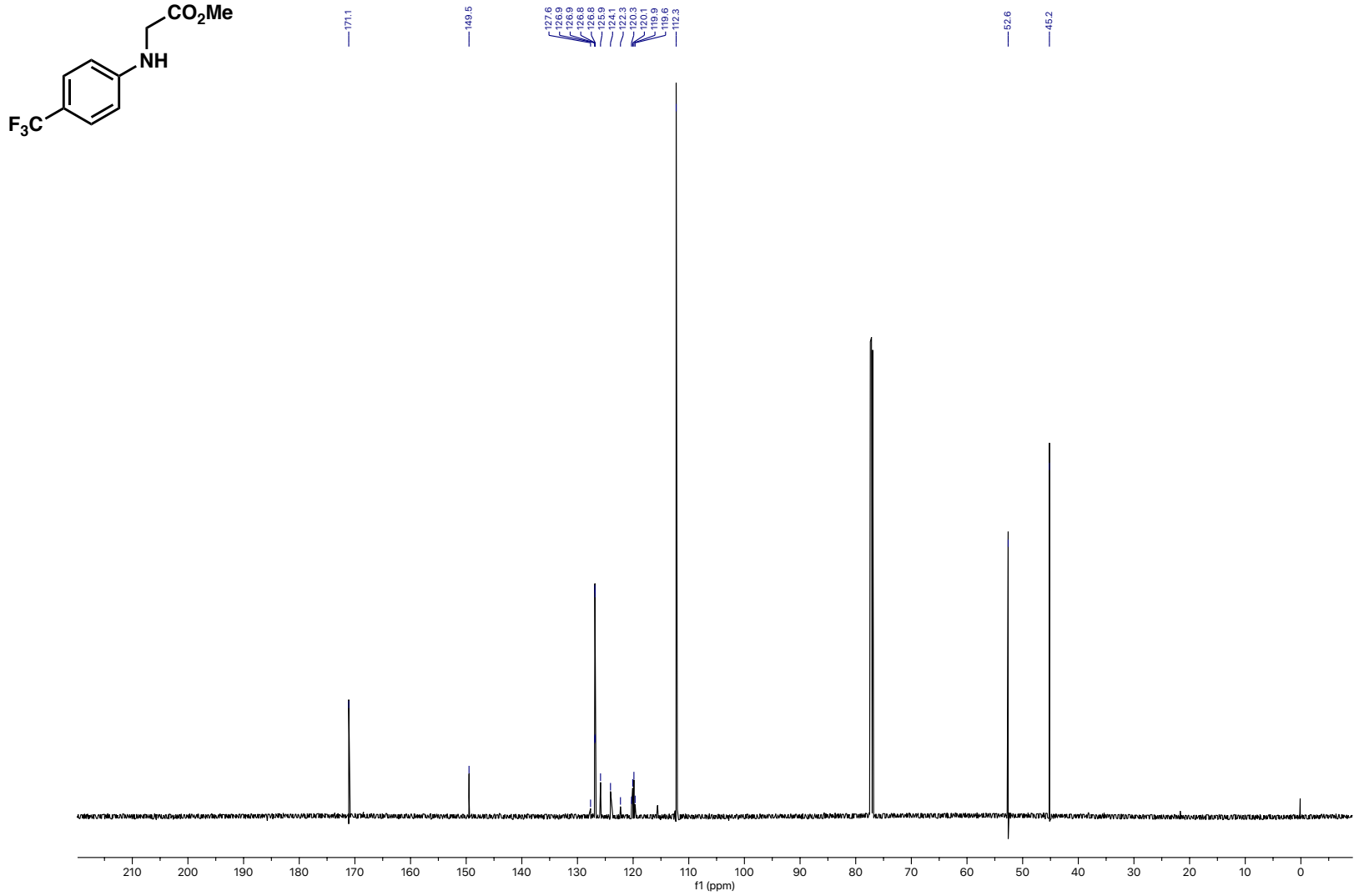
Compound 15 ^{19}F NMR



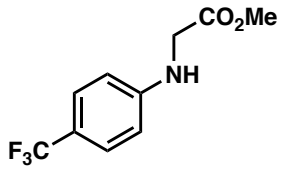
Compound 18 ^{1}H NMR

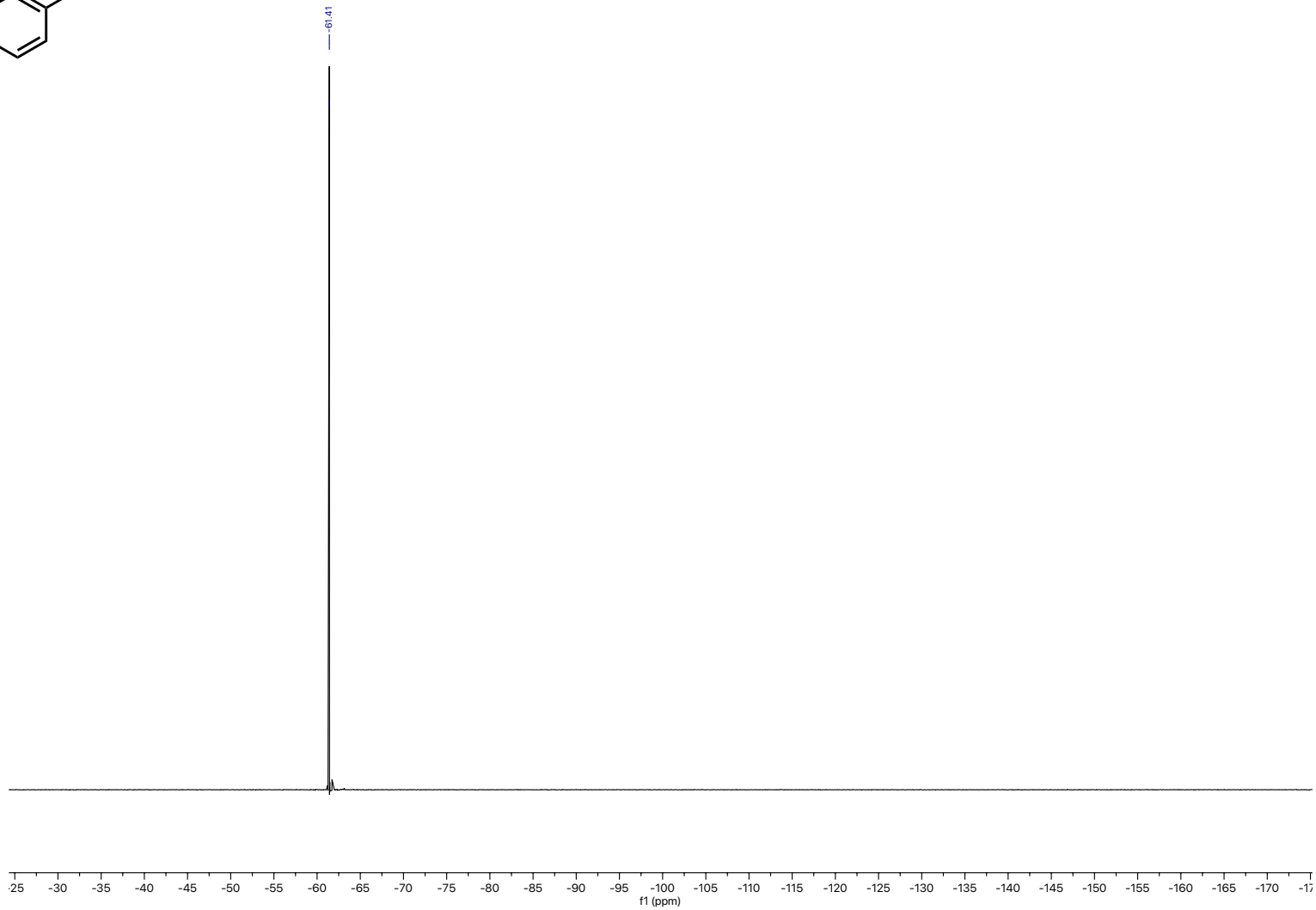


Compound 18 ^{13}C NMR

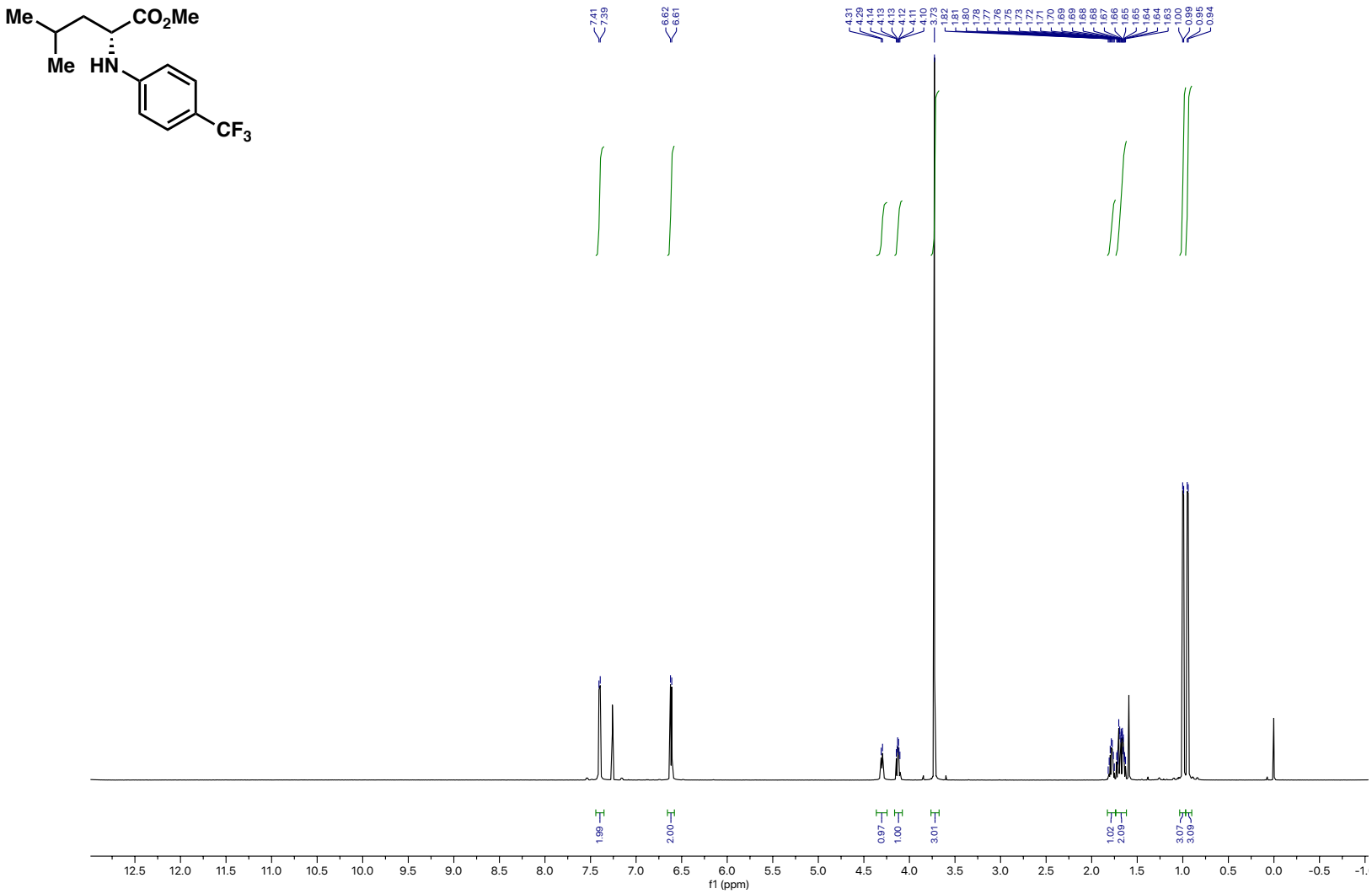


Compound 18 ^{19}F NMR

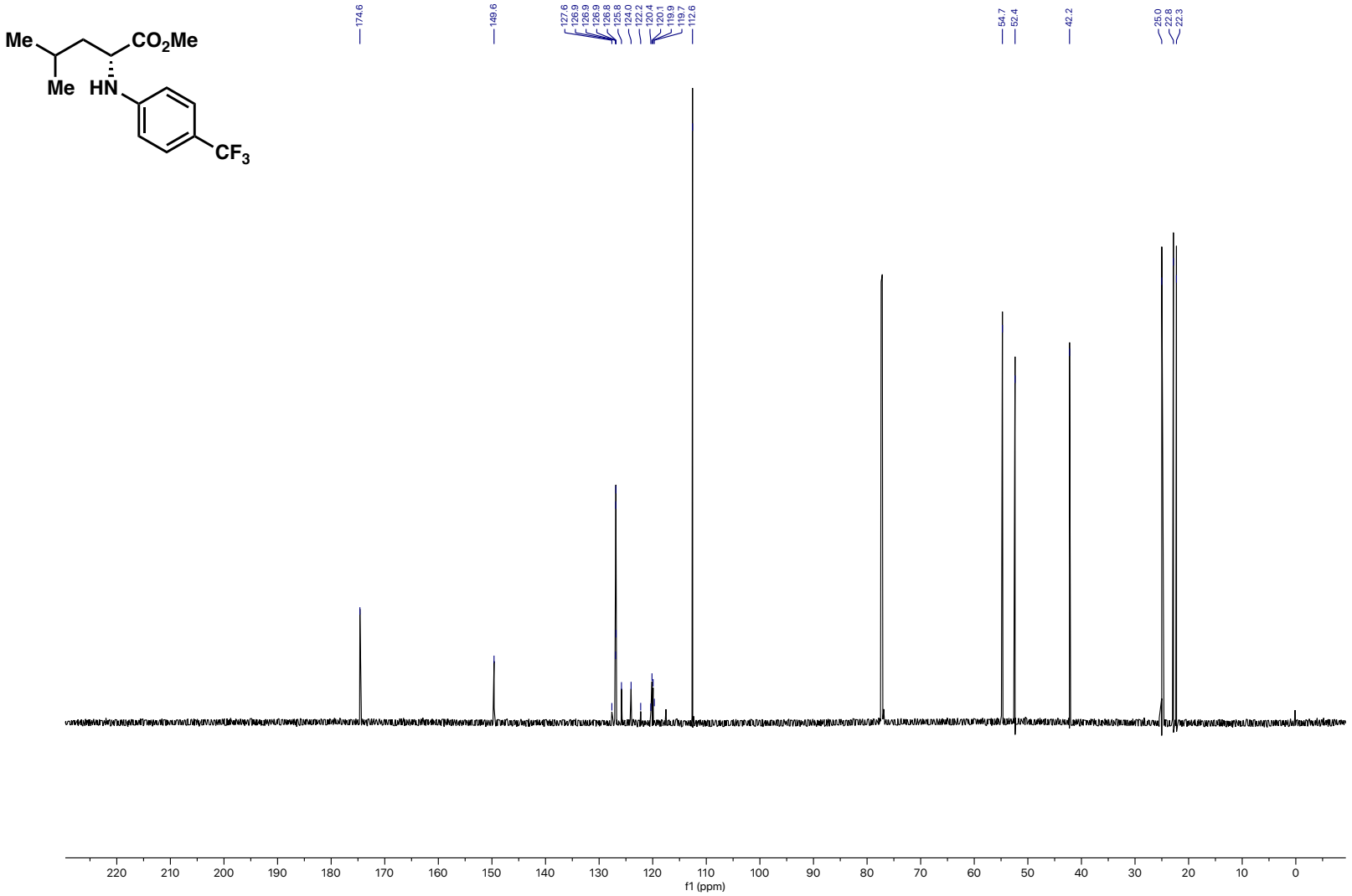




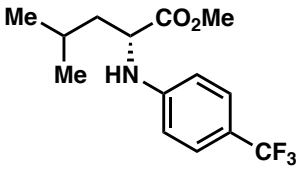
Compound 19 ^{1}H NMR



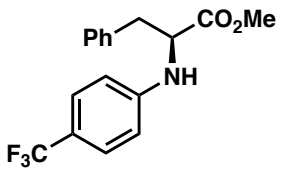
Compound 19 ^{13}C NMR



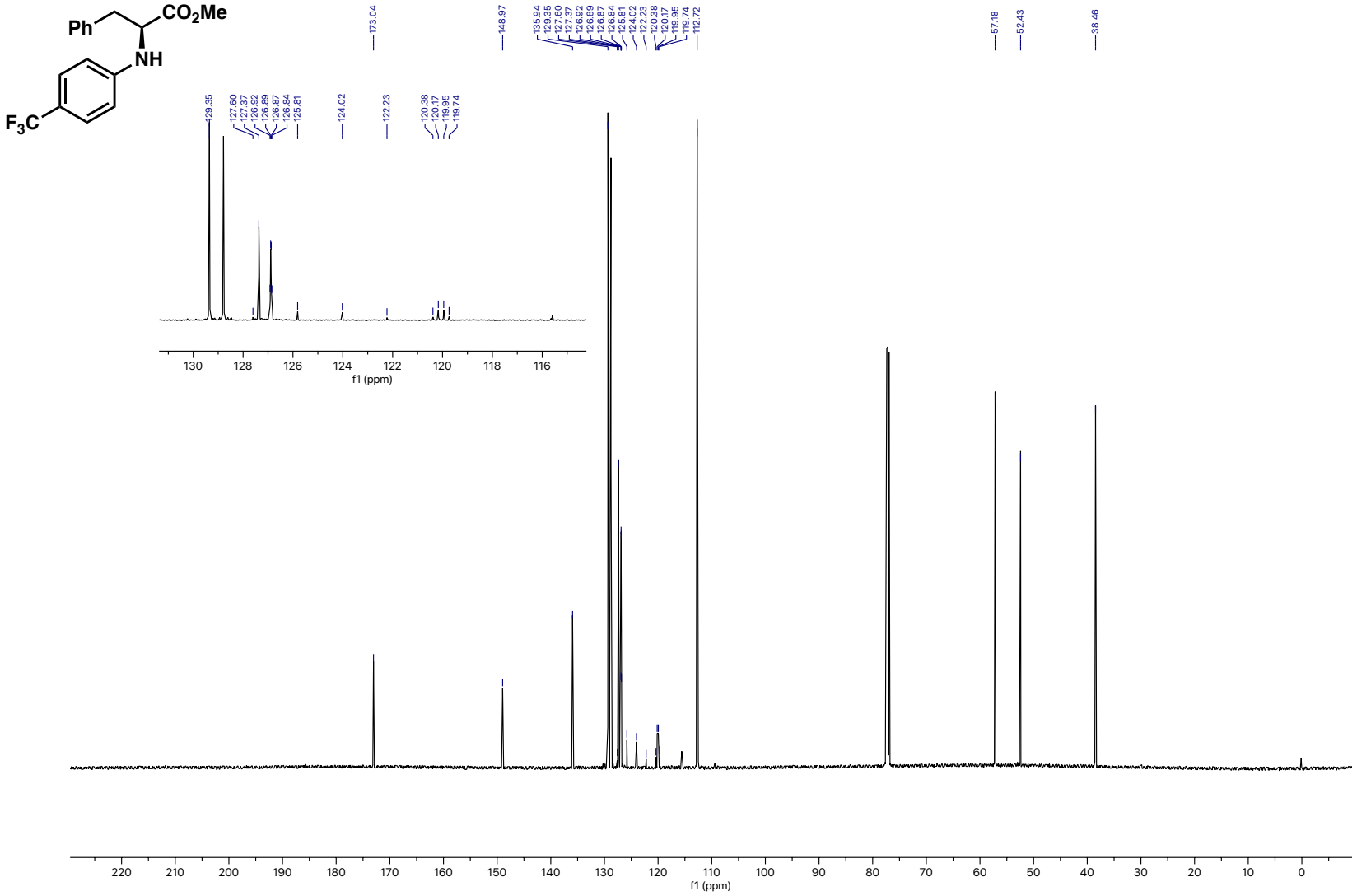
Compound 19 ^{19}F NMR



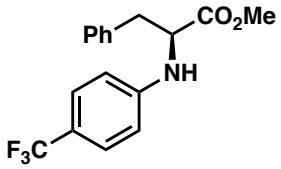
Compound 20 ^{1}H NMR



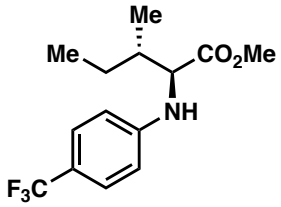
Compound 20 ^{13}C NMR

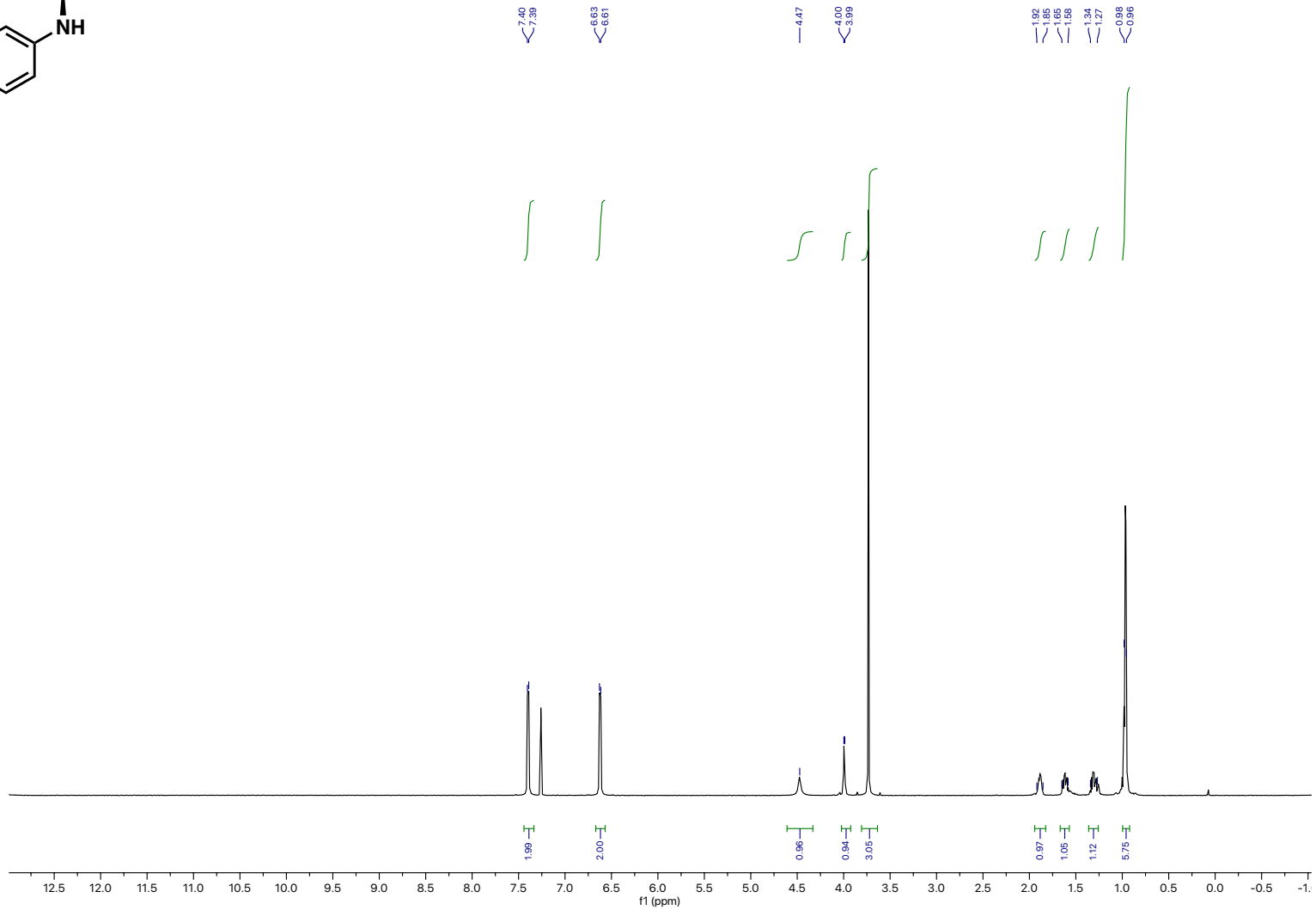


Compound 20 ^{19}F NMR

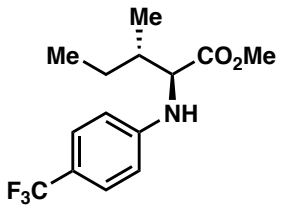


Compound 21 ^{1}H NMR

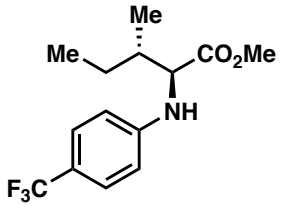




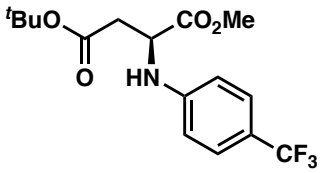
Compound 21 ^{13}C NMR

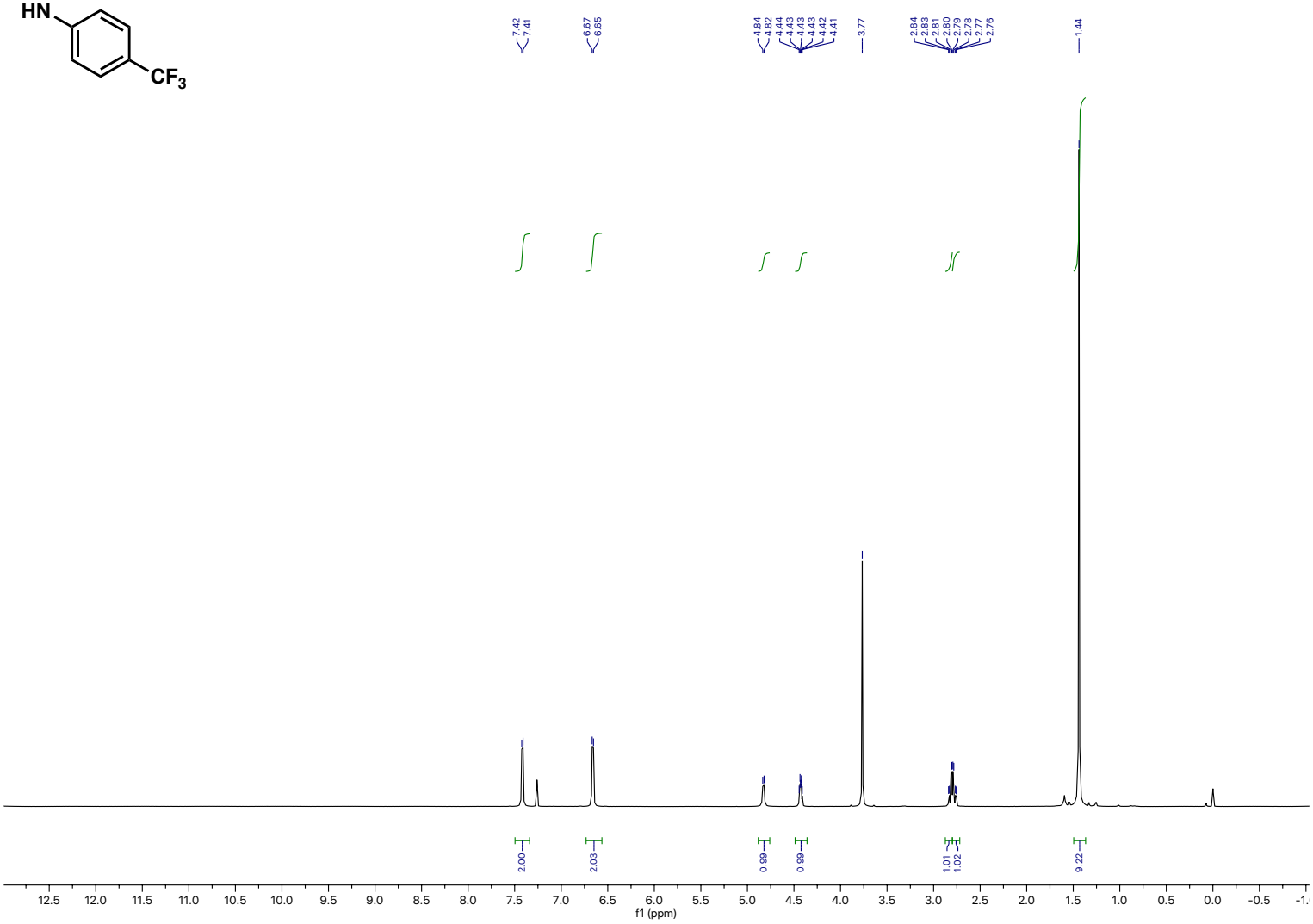


Compound 21 ^{19}F NMR

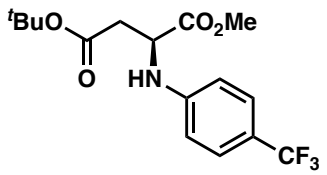


Compound 22 ^{1}H NMR

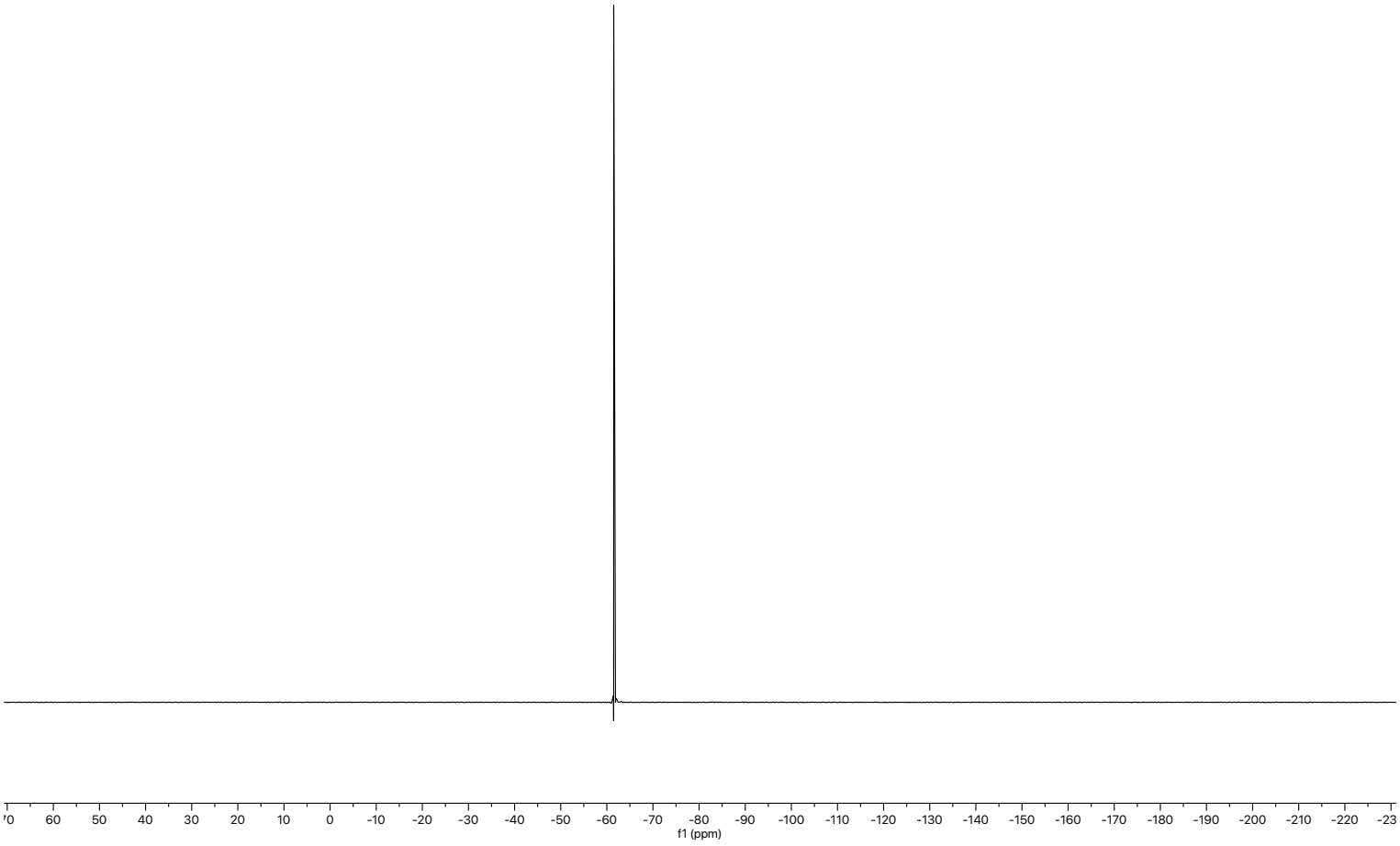




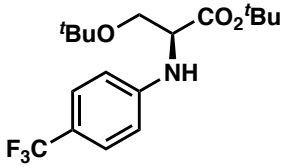
Compound 22 ^{13}C NMR



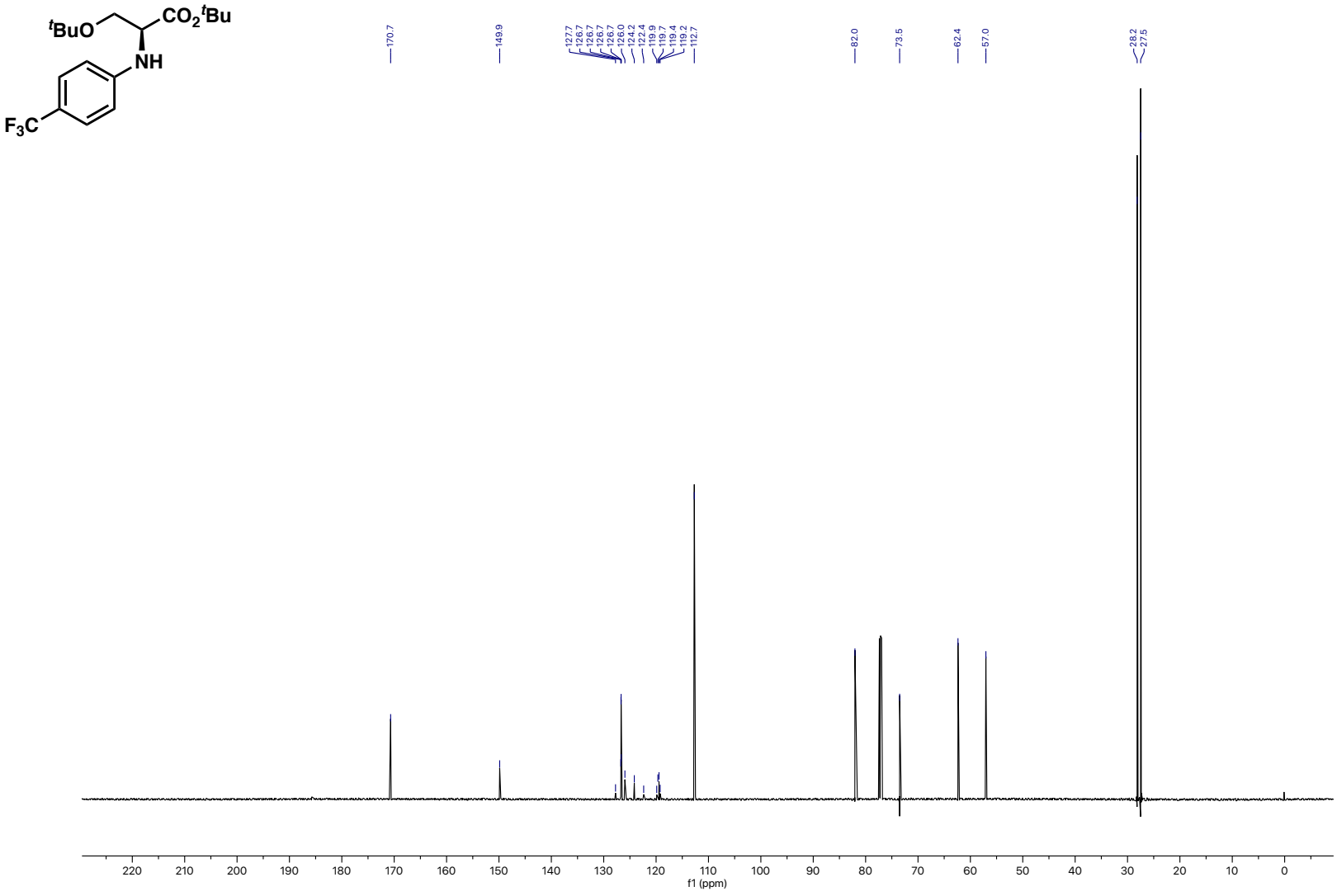
Compound 22 ^{19}F NMR



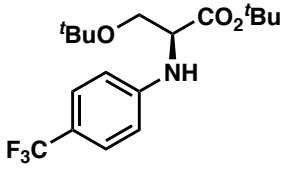
Compound 23 ^{1}H NMR



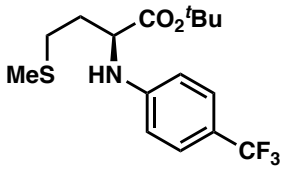
Compound 23 ^{13}C NMR



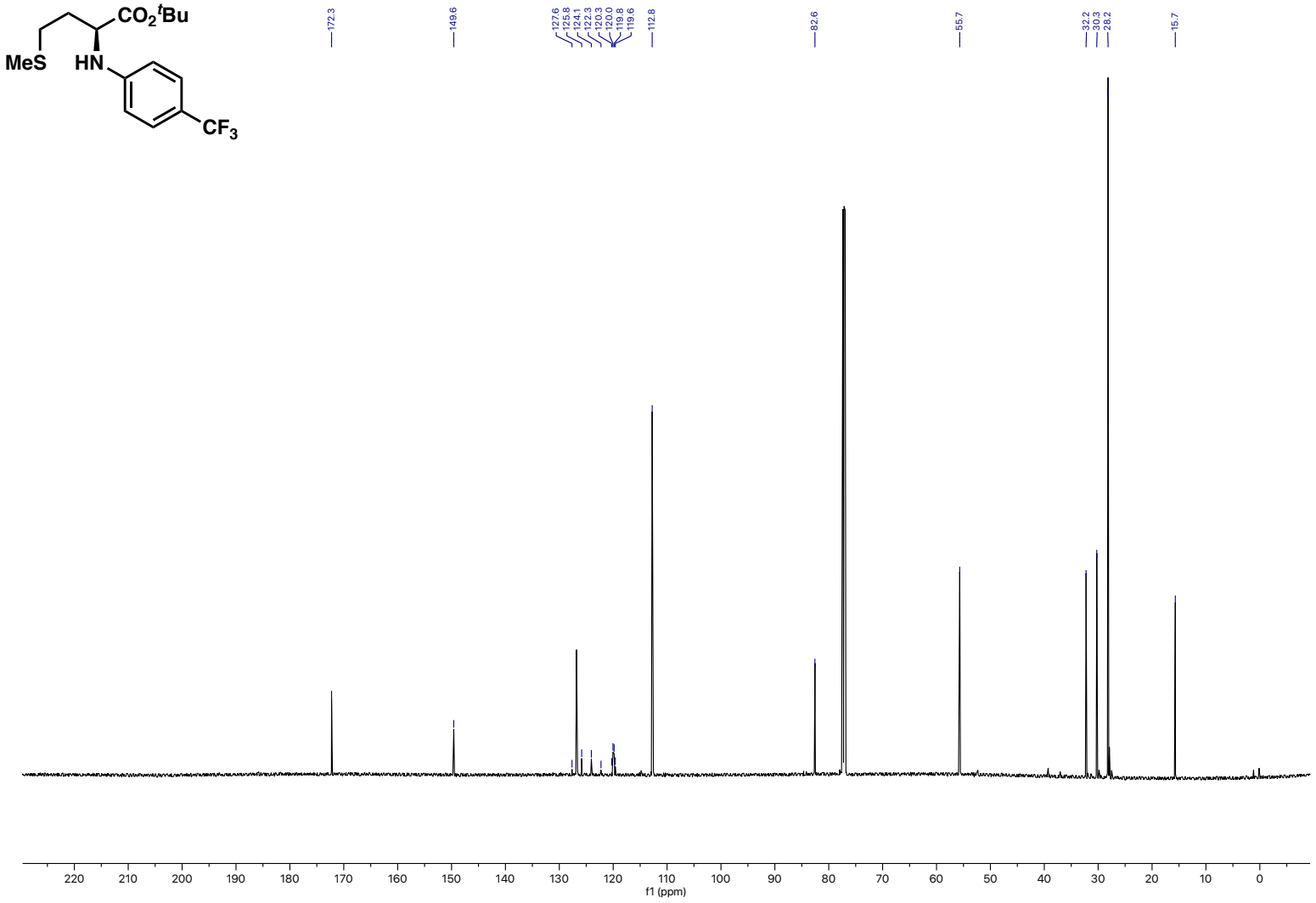
Compound 23 ^{19}F NMR



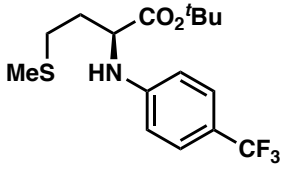
Compound 24 ^{1}H NMR



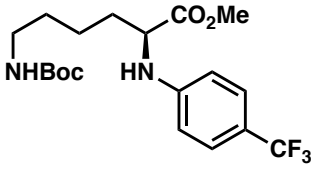
Compound 24 ^{13}C NMR



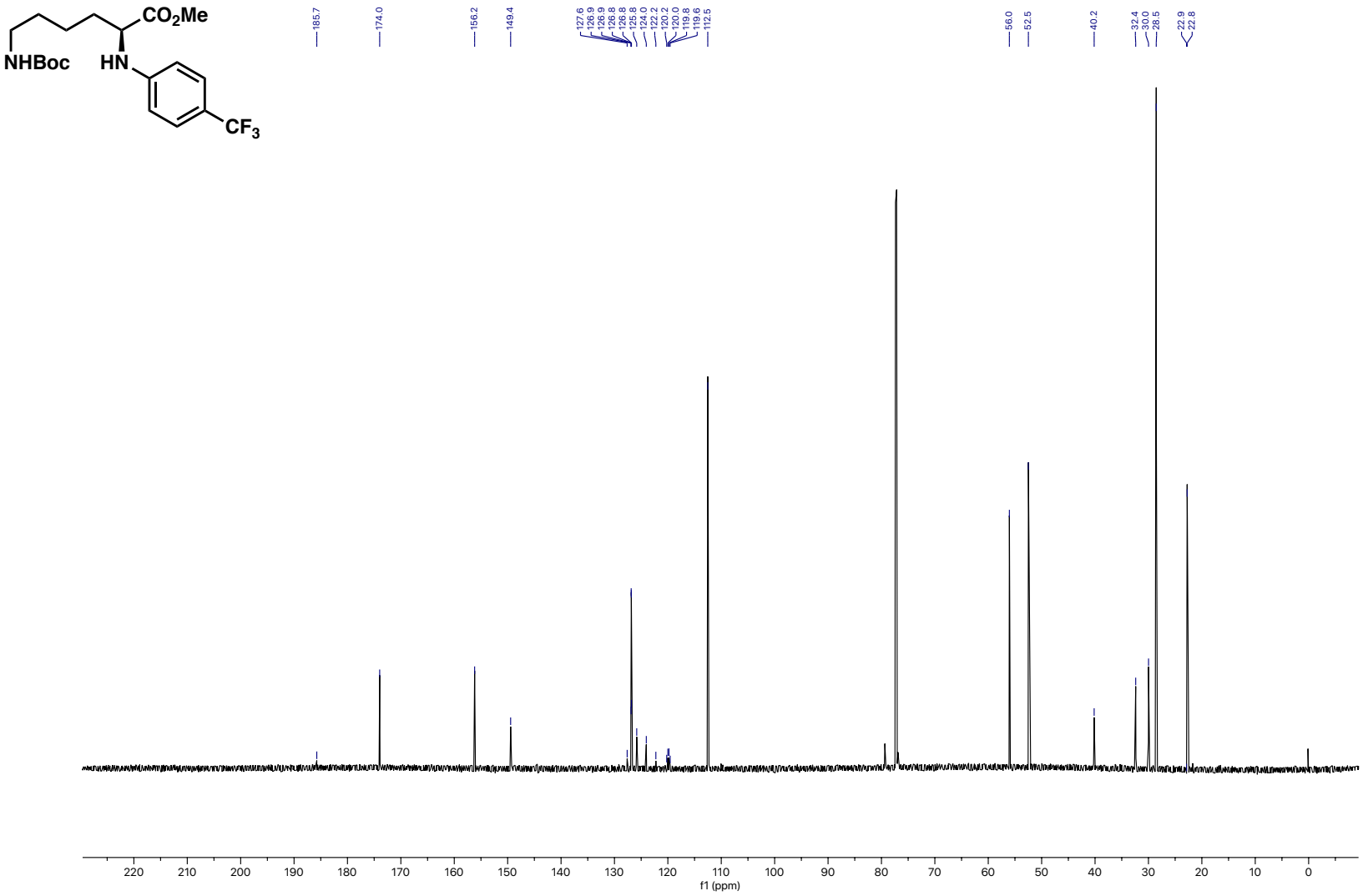
Compound 24 ^{19}F NMR



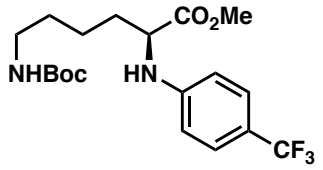
Compound 25 ^{1}H NMR



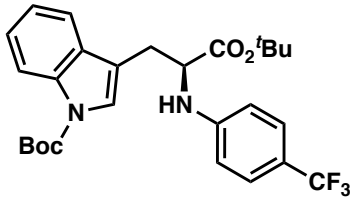
Compound 25 ^{13}C NMR

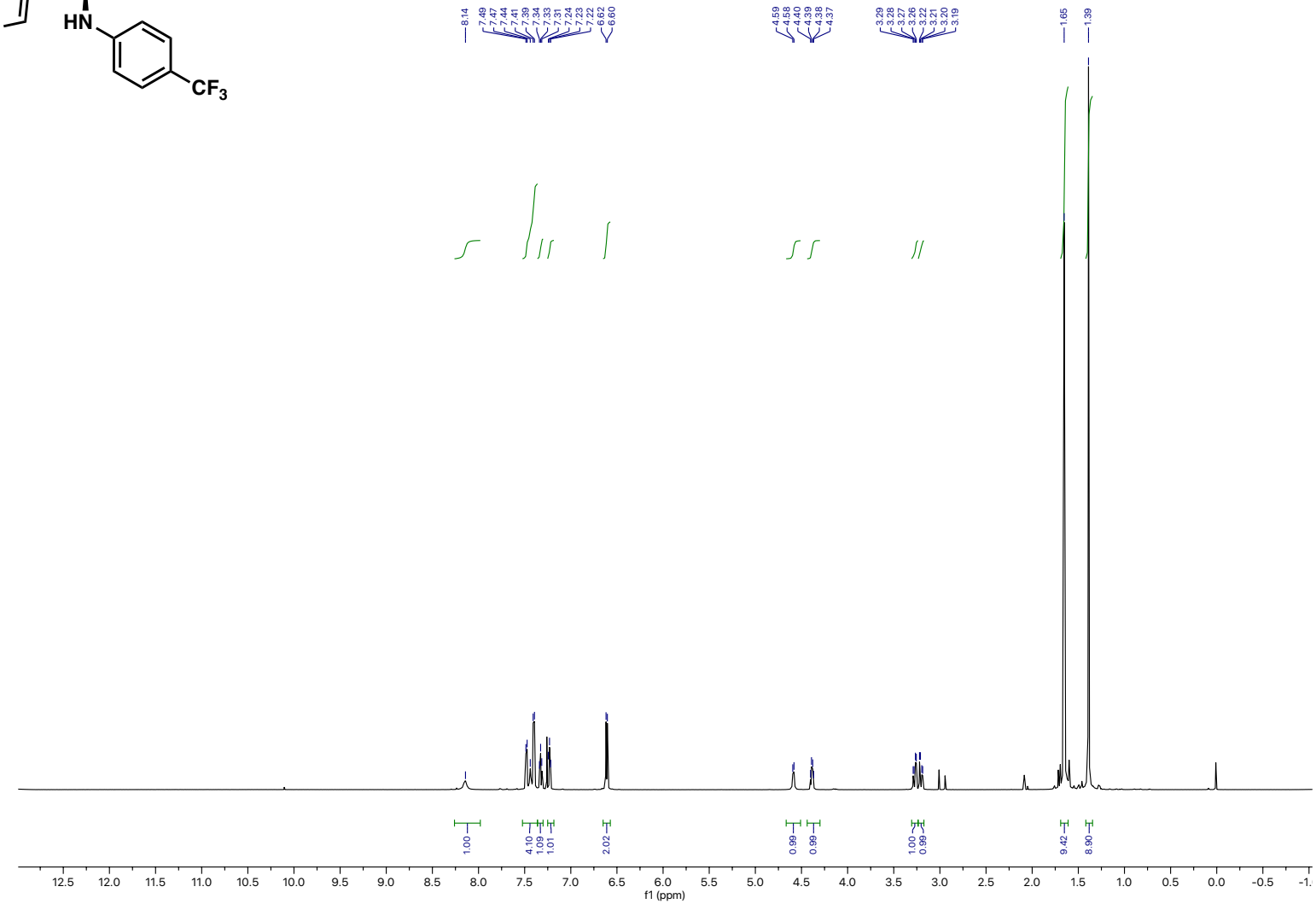


Compound 25 ^{19}F NMR

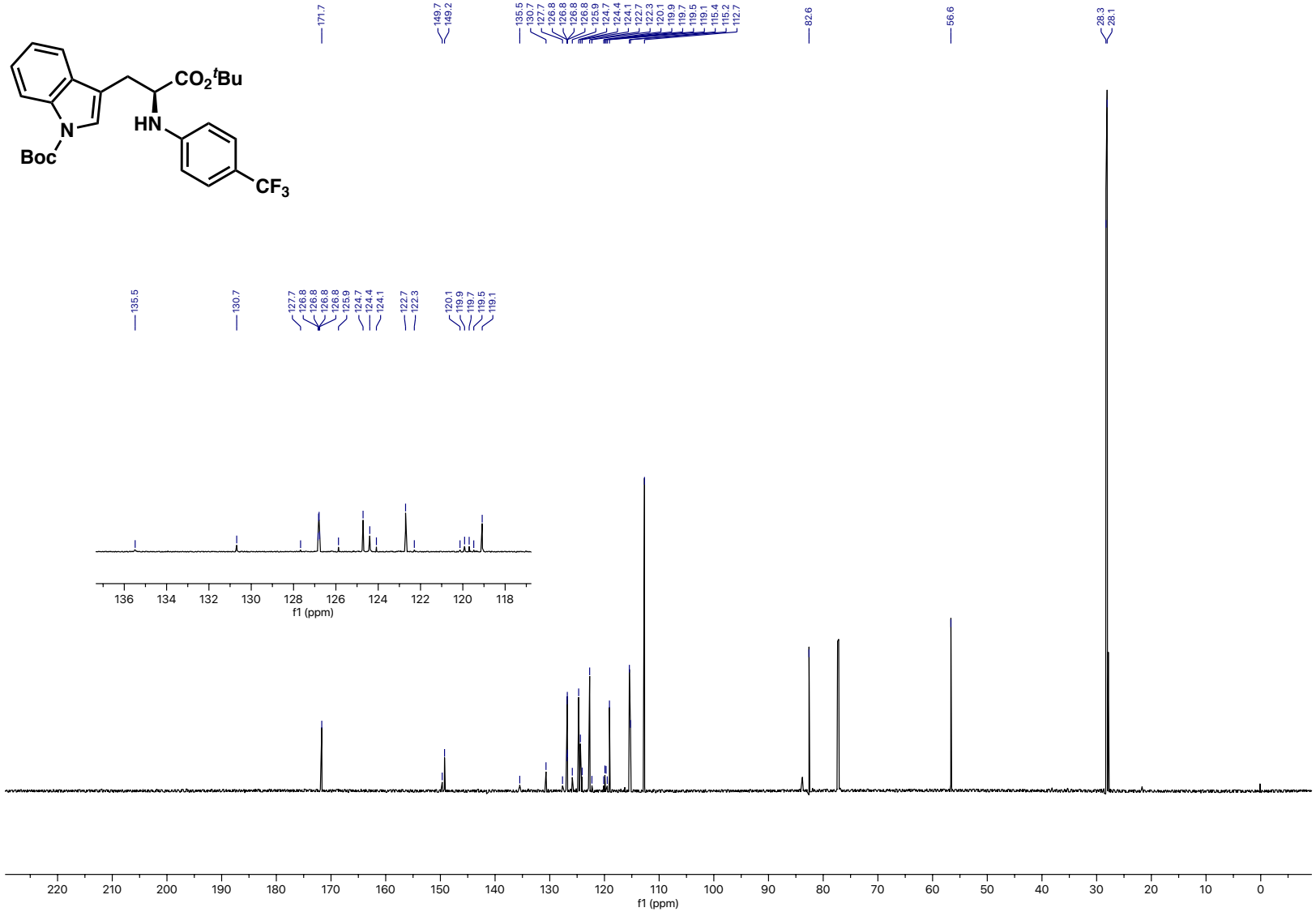


Compound 26 ^{1}H NMR

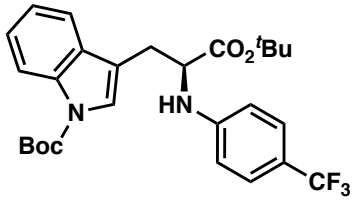




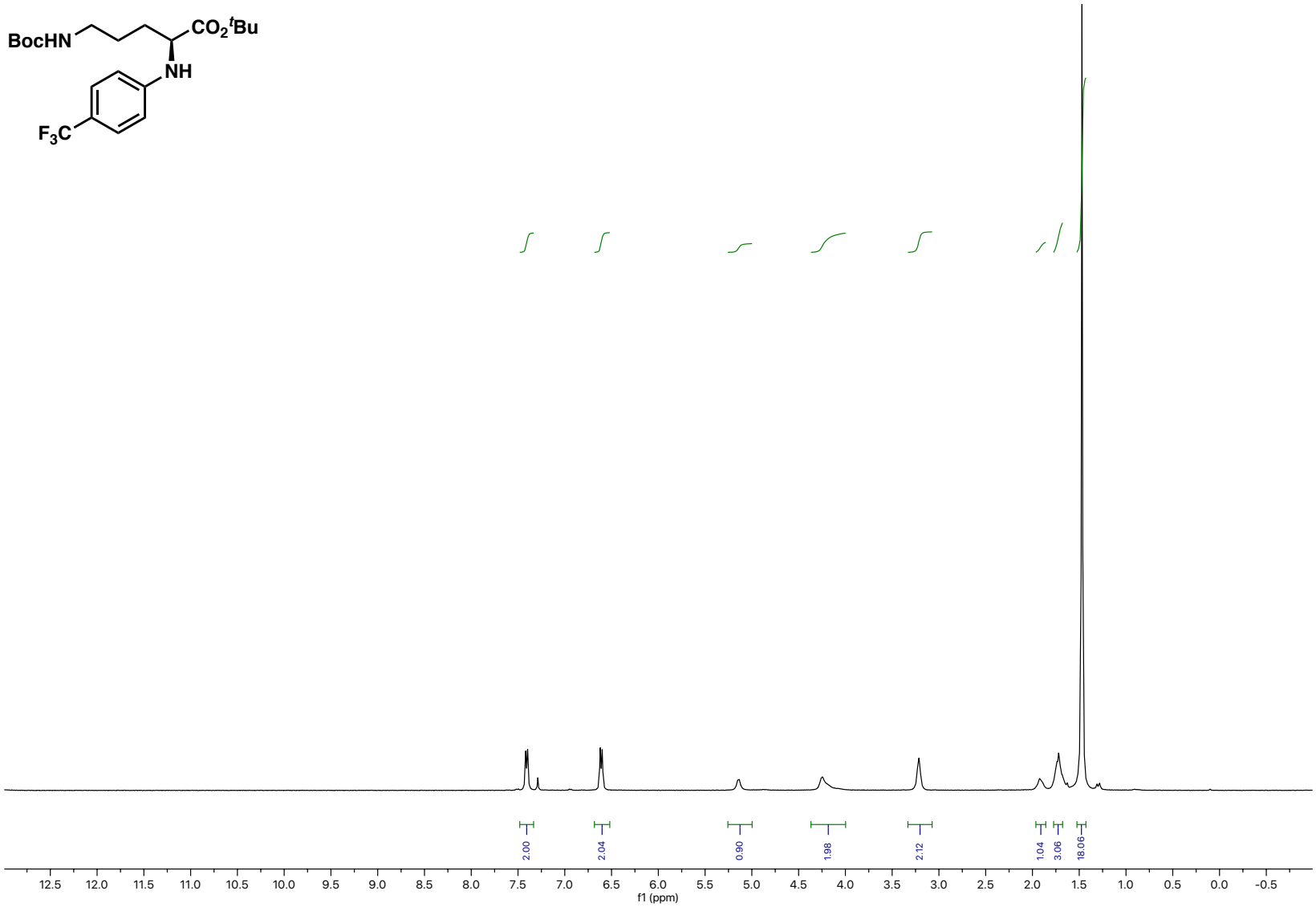
Compound 26 ^{13}C NMR



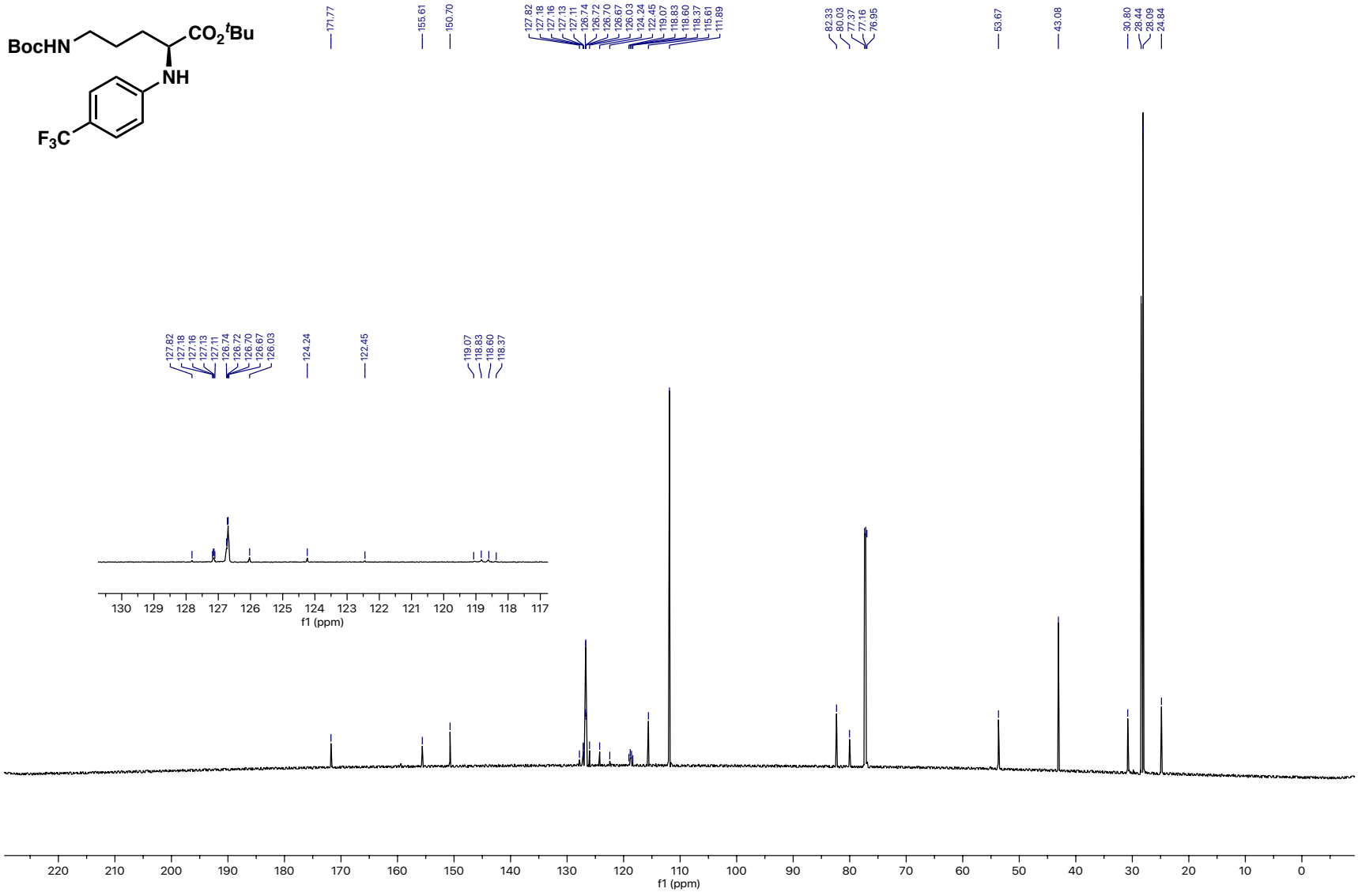
Compound 26 ^{19}F NMR



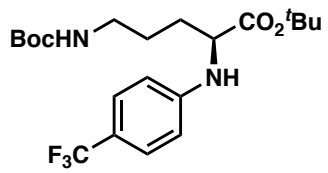
Compound 27 ^{1}H NMR



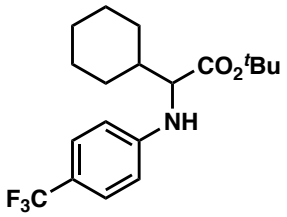
Compound 27 ^{13}C NMR



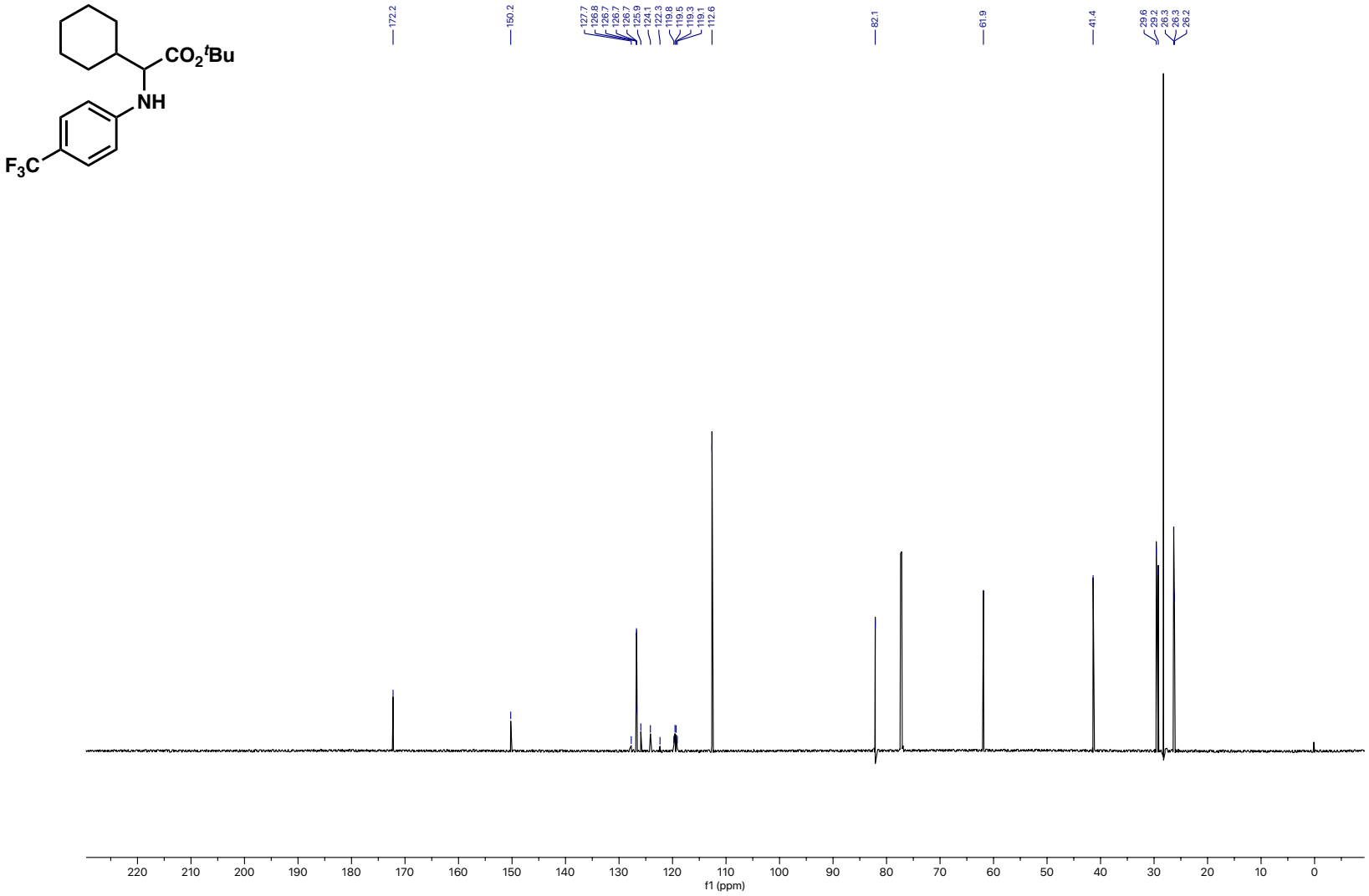
Compound 27 ^{19}F NMR



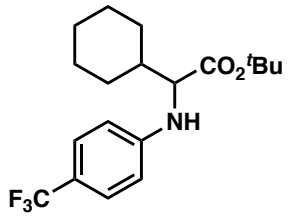
Compound 28 ^{1}H NMR



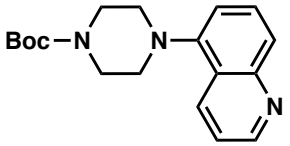
Compound 28 ^{13}C NMR

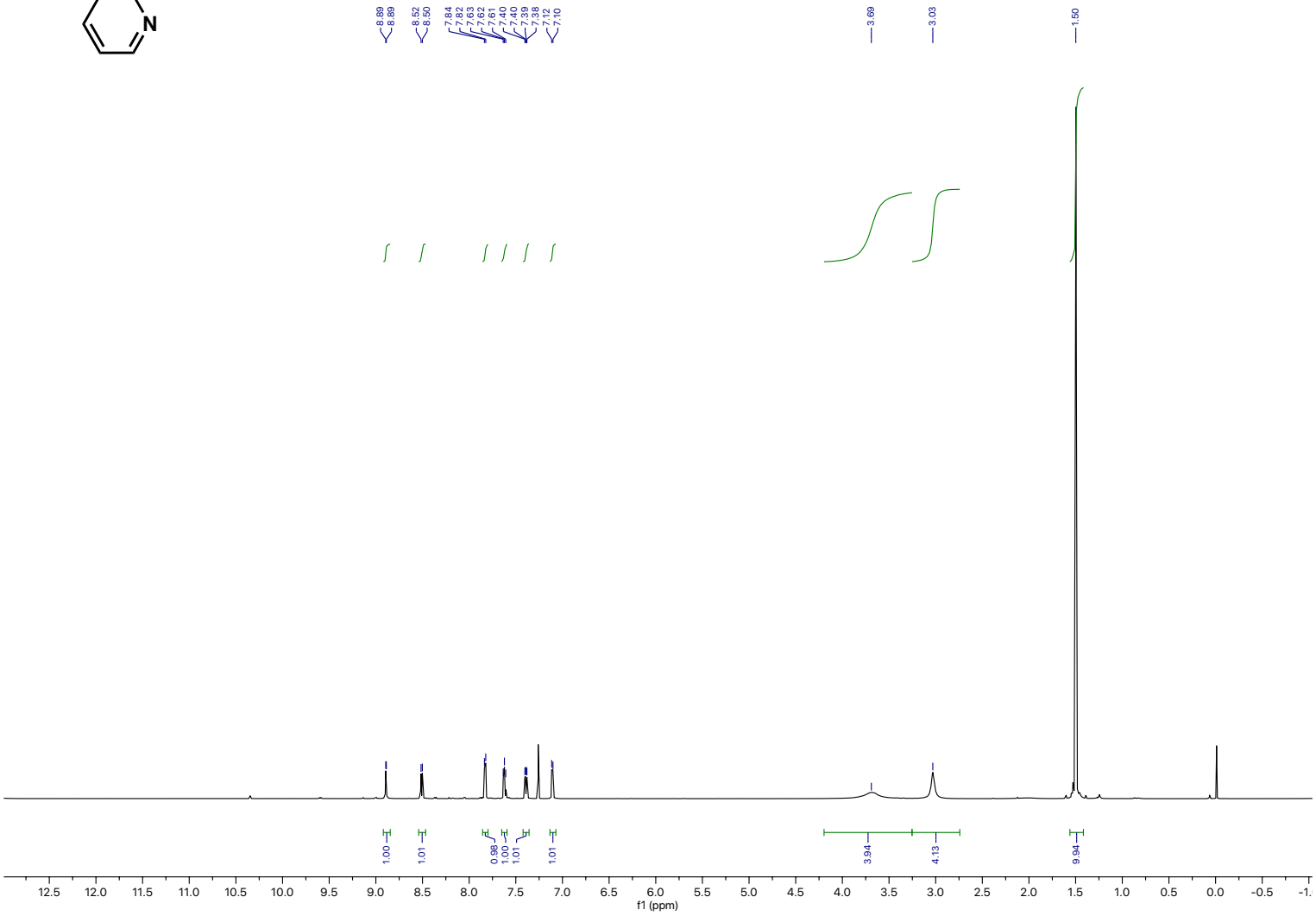


Compound 28 ^{19}F NMR

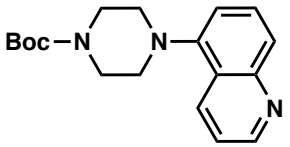


Compound 34 ^{1}H NMR

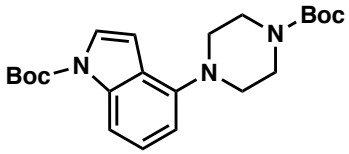


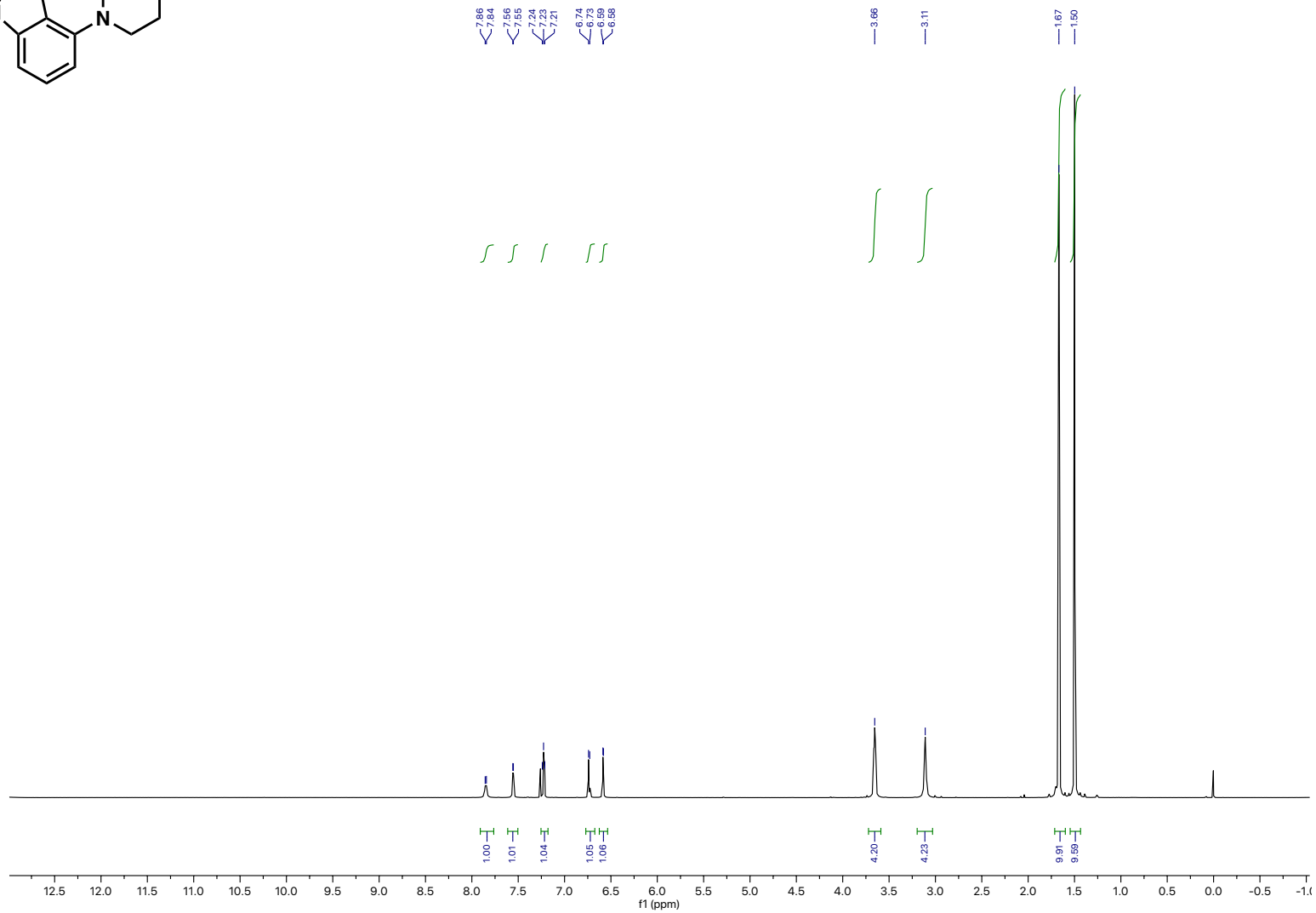


Compound 34 ^{13}C NMR

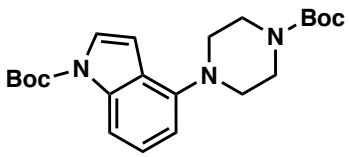


Compound 35 ^{1}H NMR

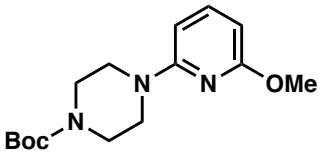


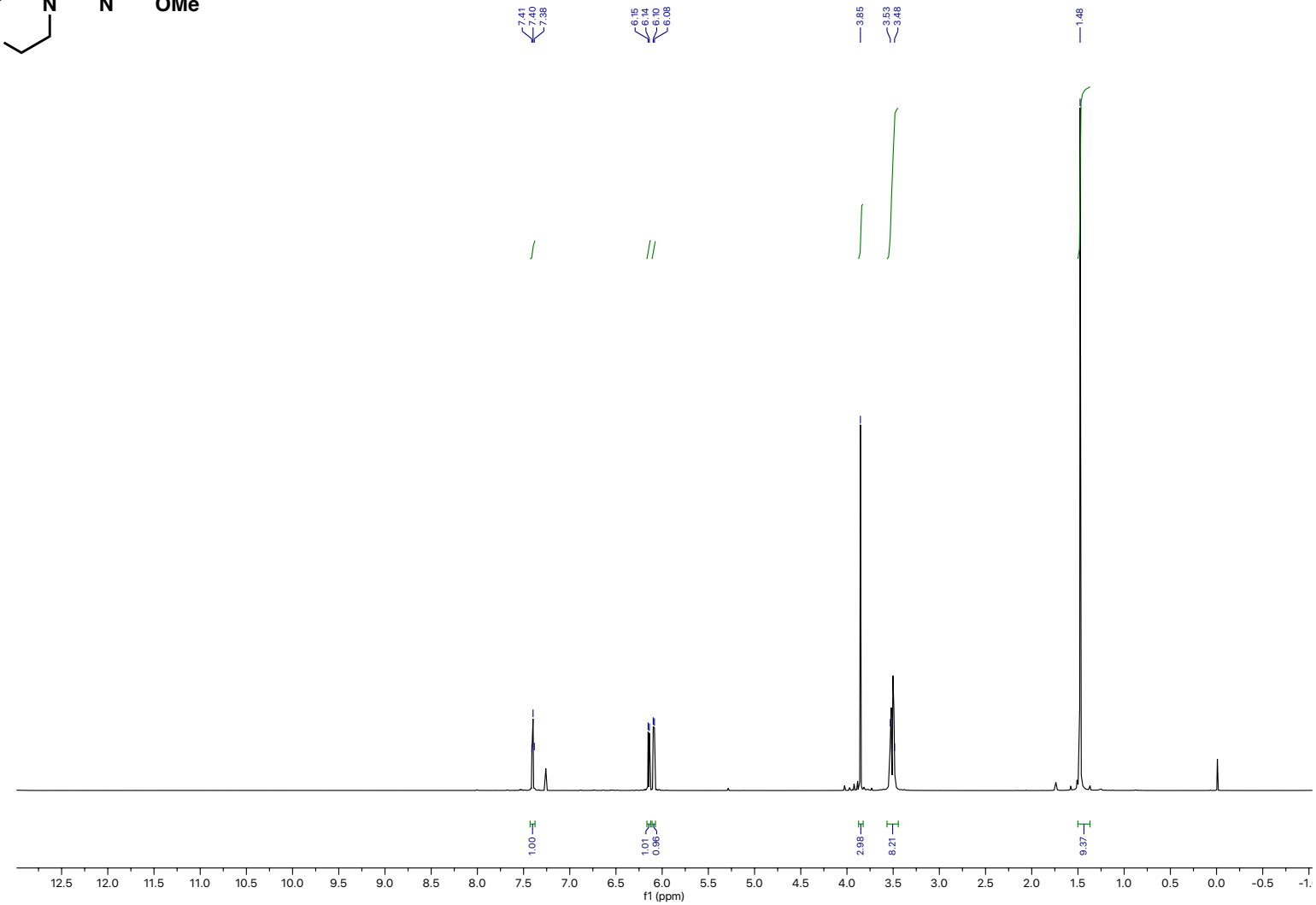


Compound 35 ^{13}C NMR

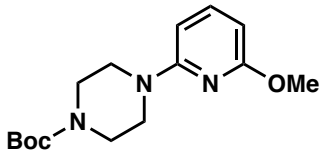


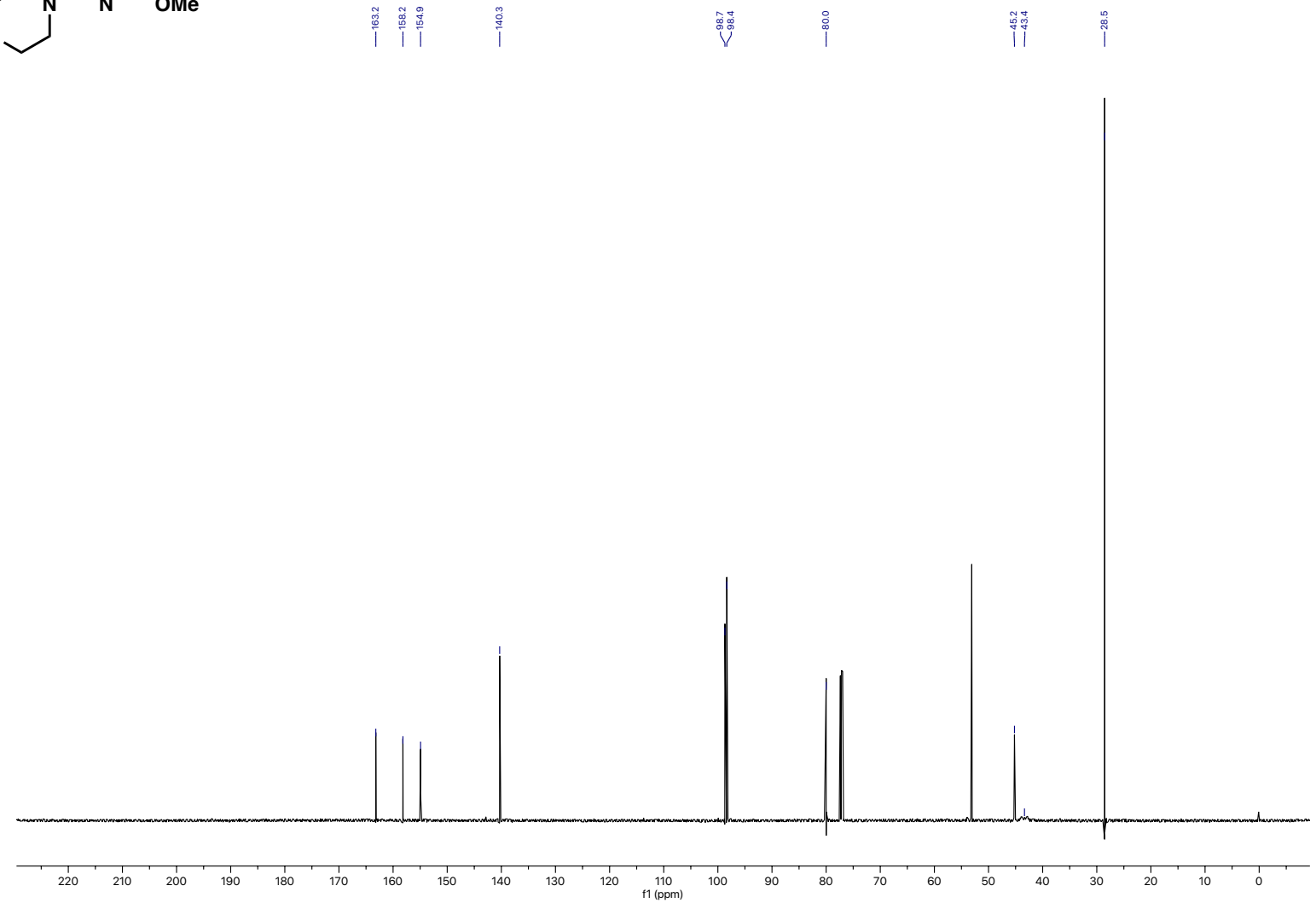
Compound 38 ^{1}H NMR



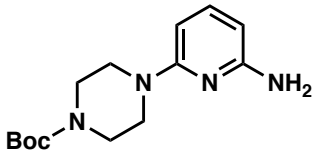


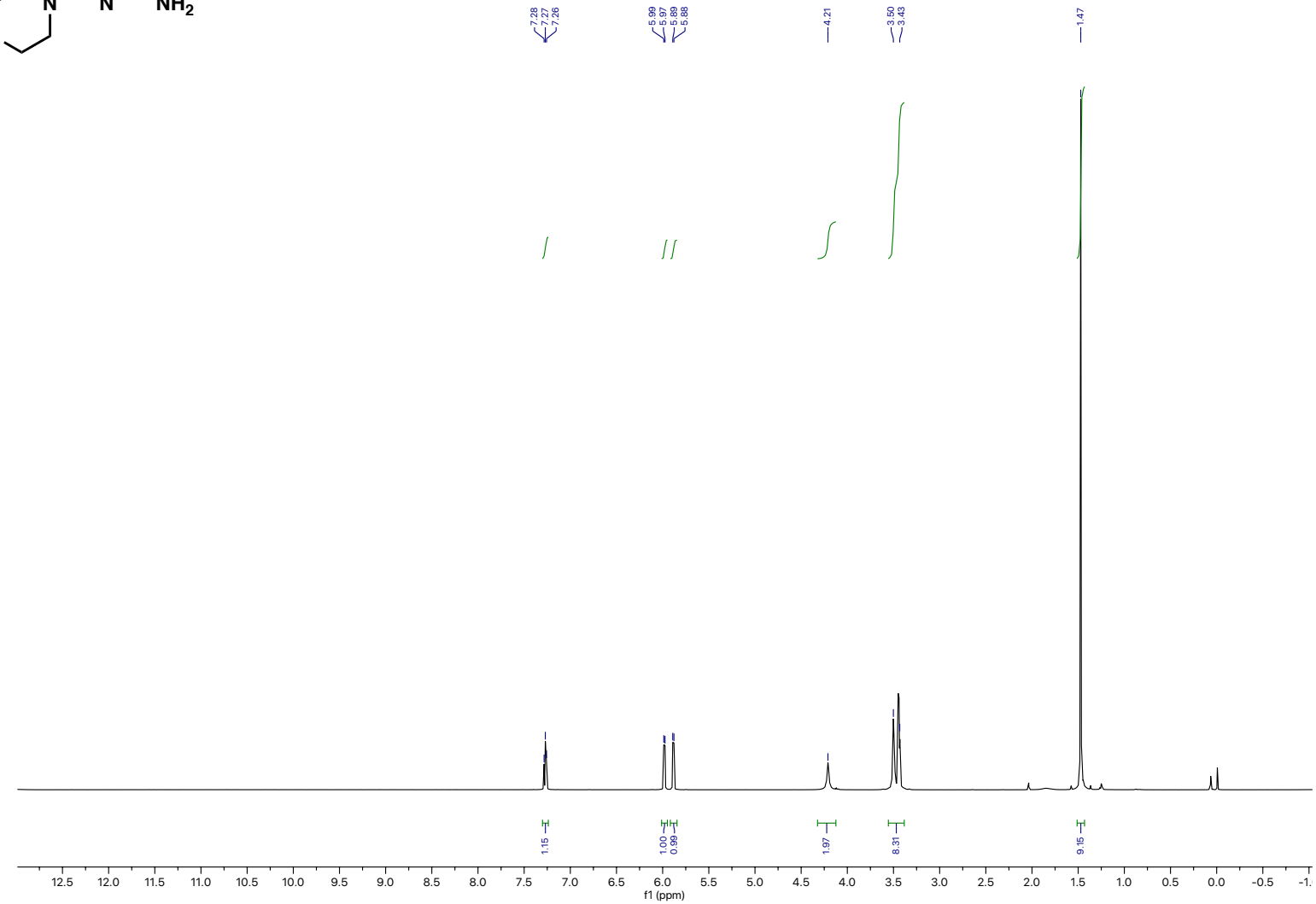
Compound 38 ^{13}C NMR



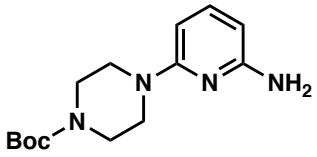


Compound 39 ^{1}H NMR

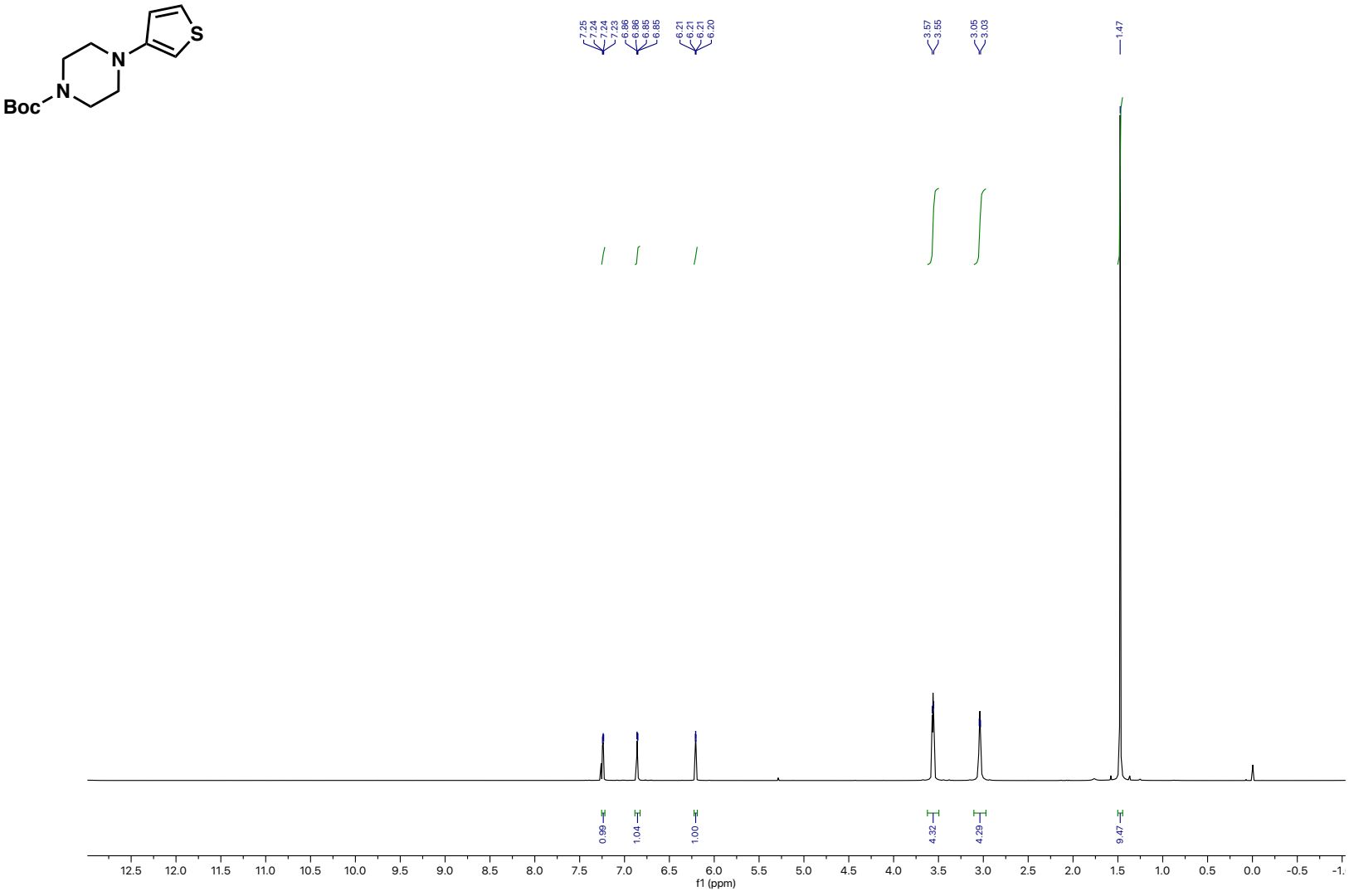




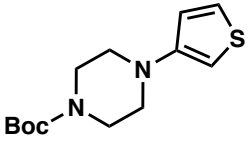
Compound 39 ^{13}C NMR

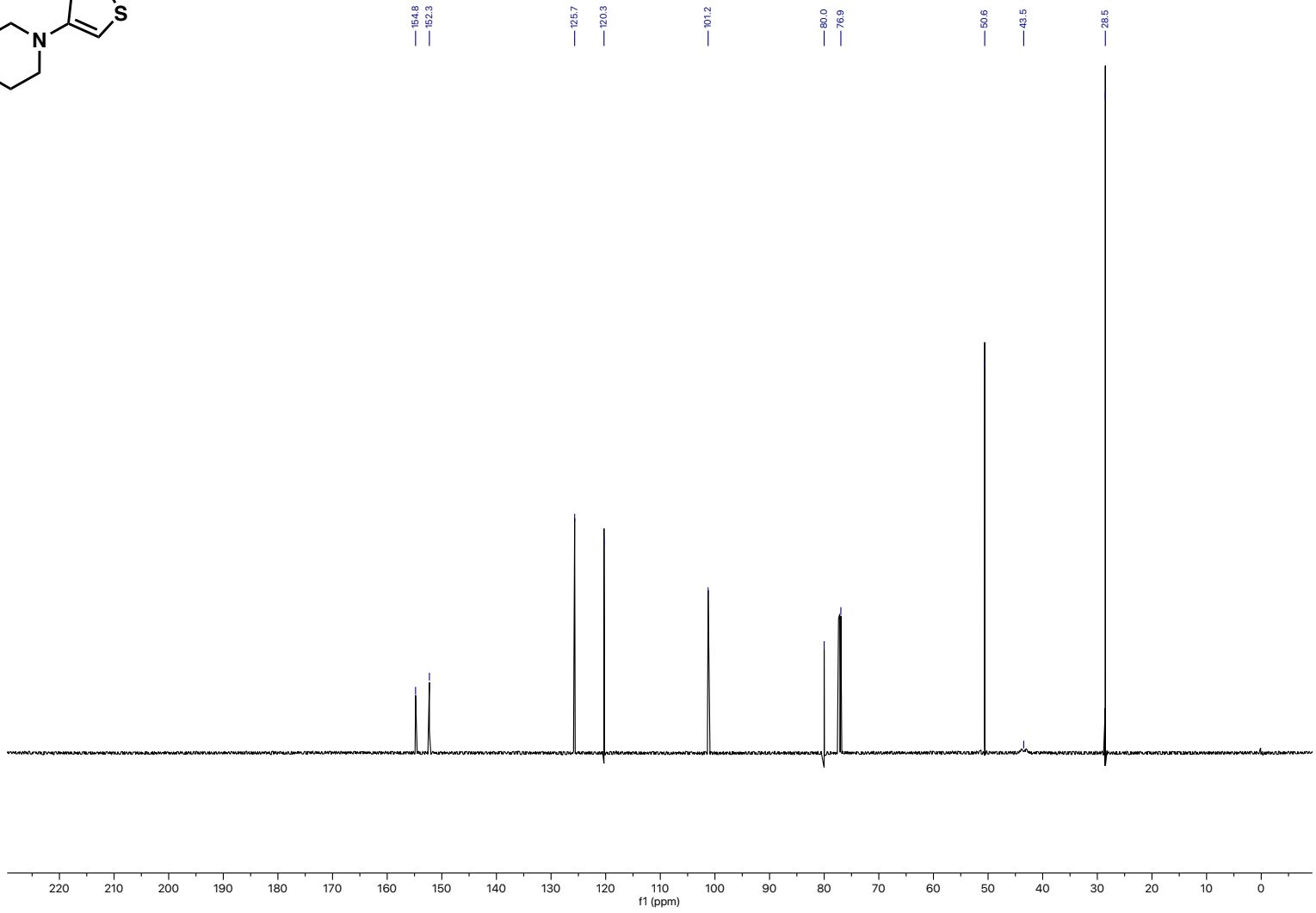


Compound 40 ^{1}H NMR

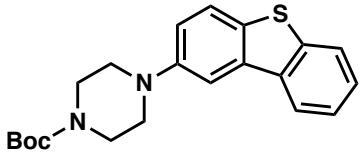


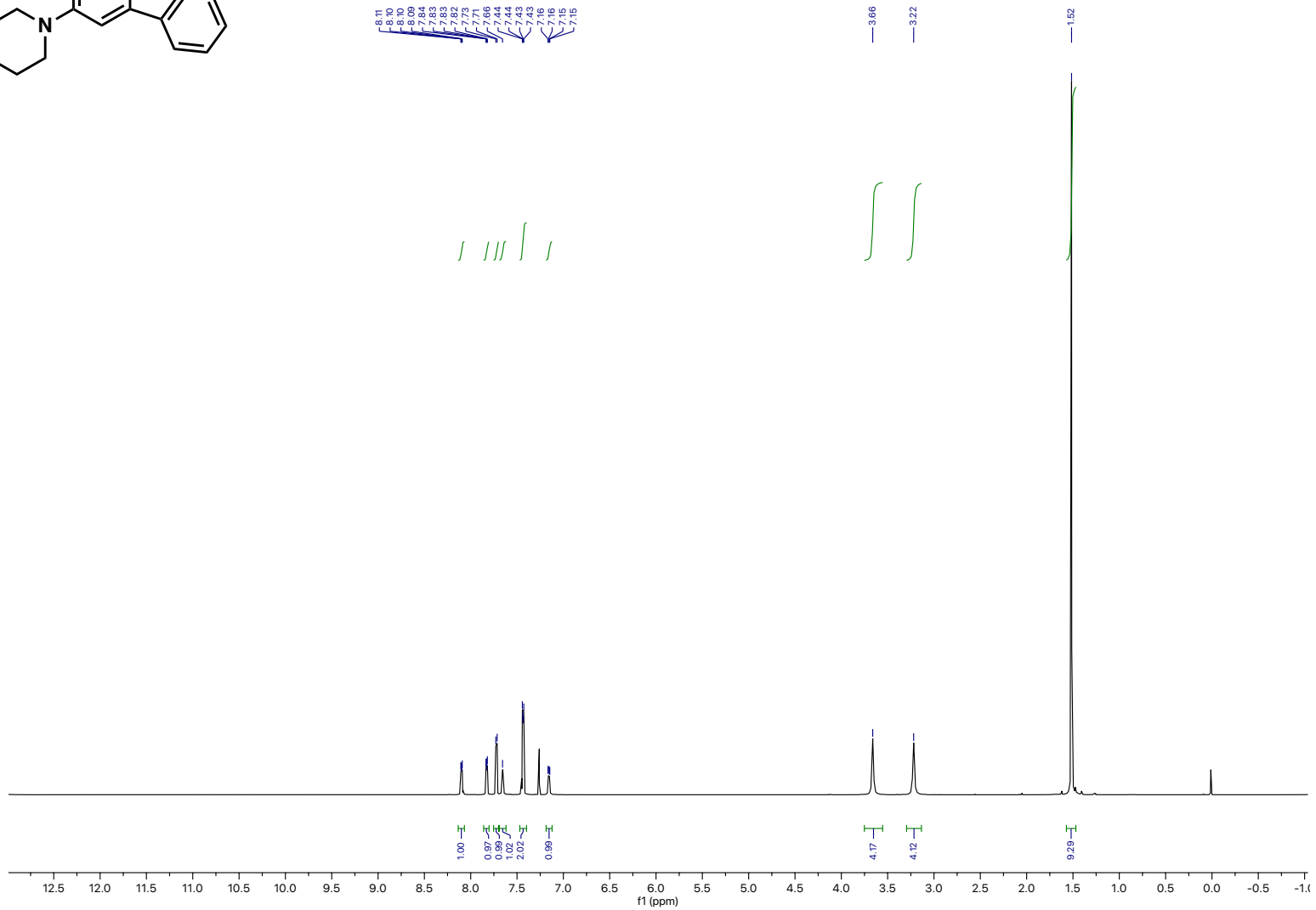
Compound 40 ^{13}C NMR



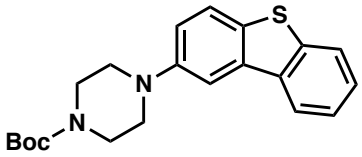


Compound 41 ^{1}H NMR

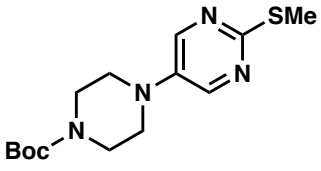




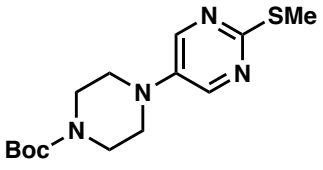
Compound 41 ^{13}C NMR



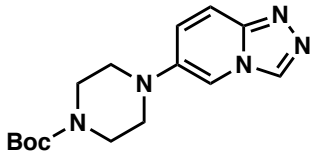
Compound 43 ^{1}H NMR

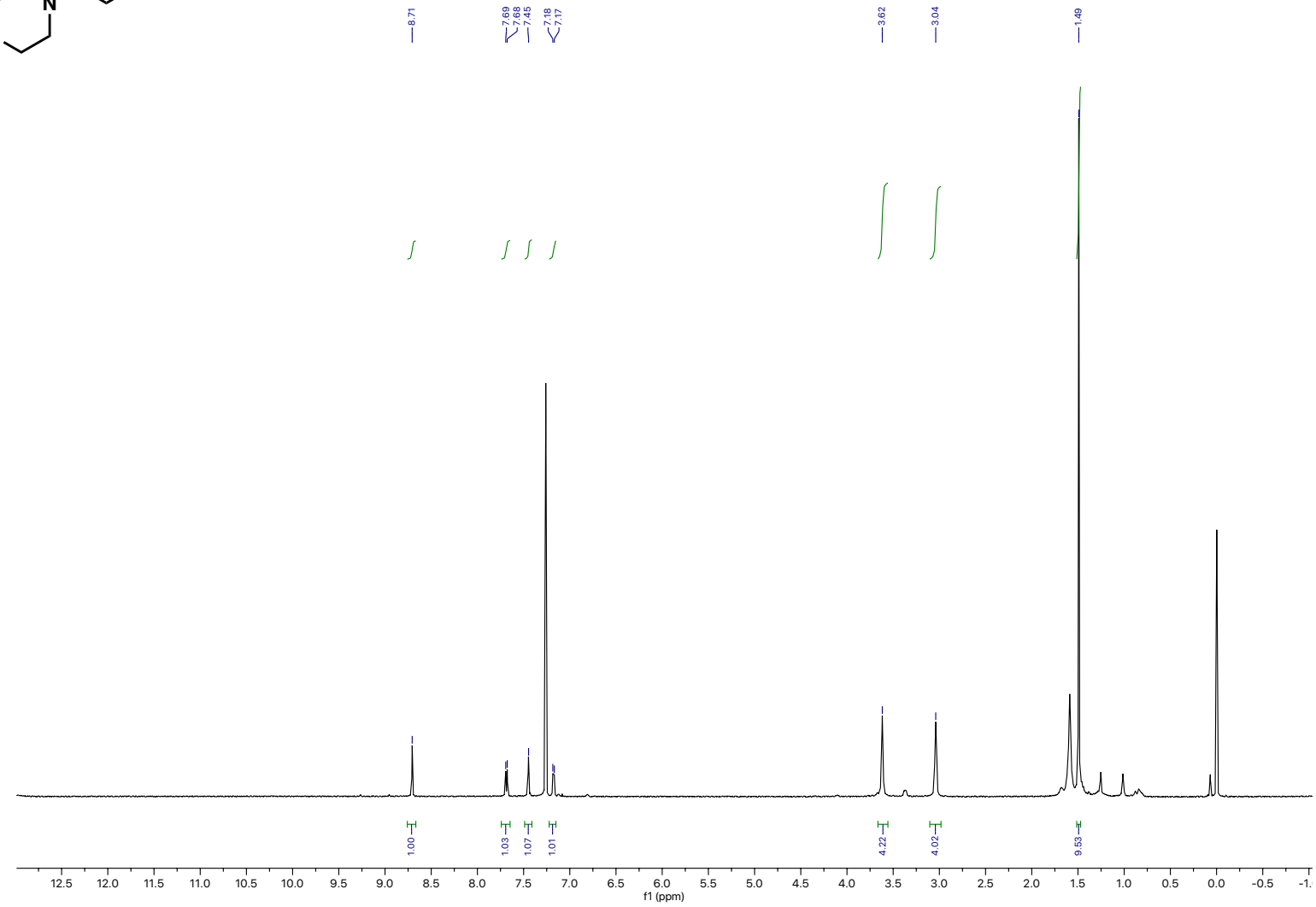


Compound 43 ^{13}C NMR

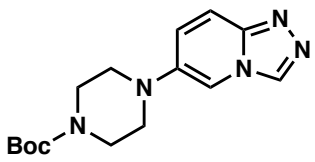


Compound 44 ^{1}H NMR

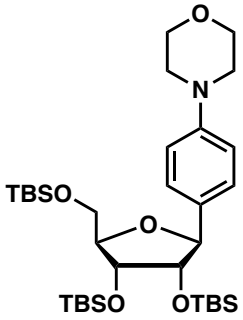


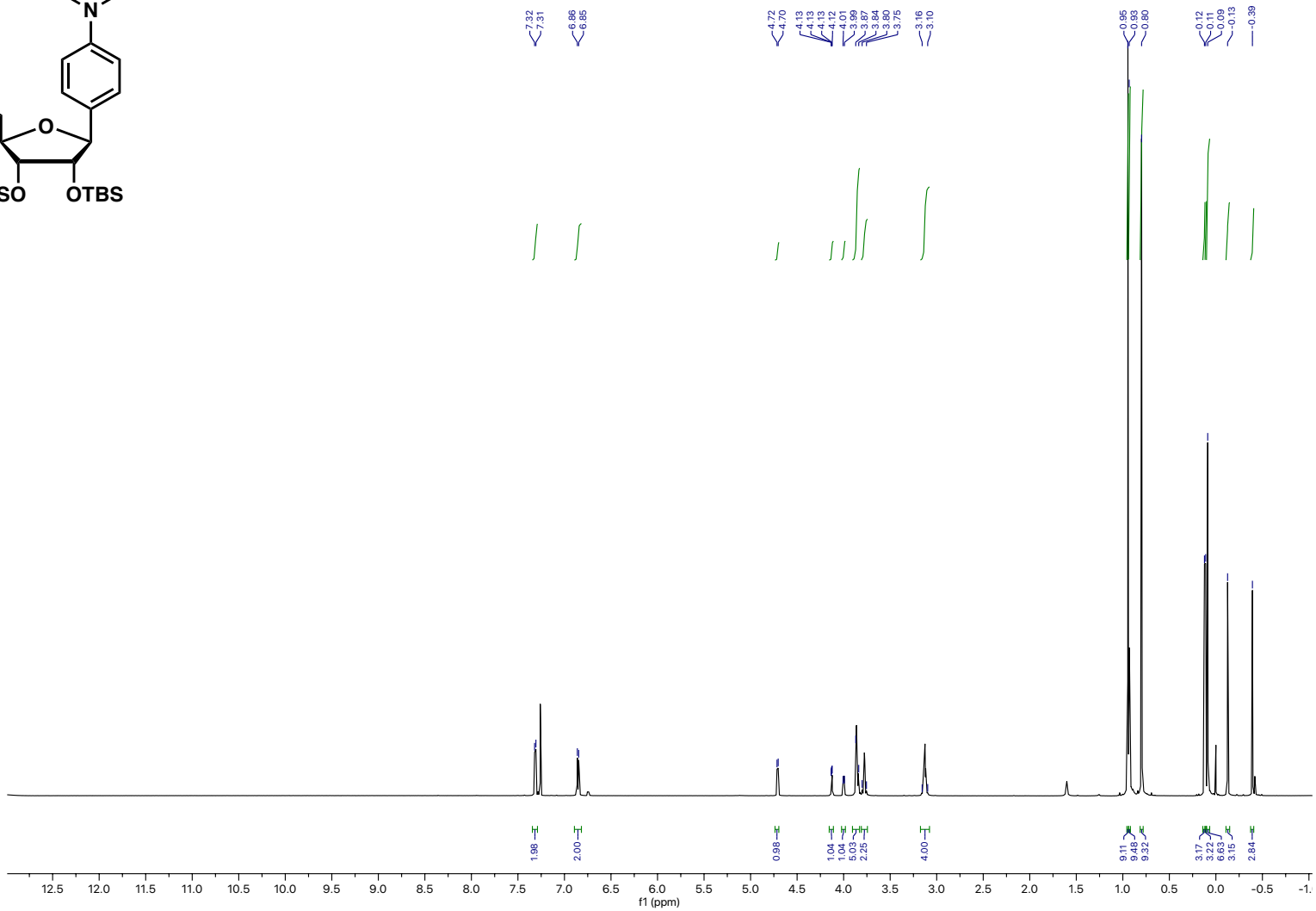


Compound 44 ^{13}C NMR

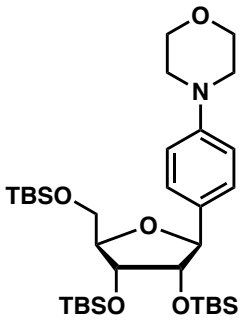


Compound 45 ^{1}H NMR

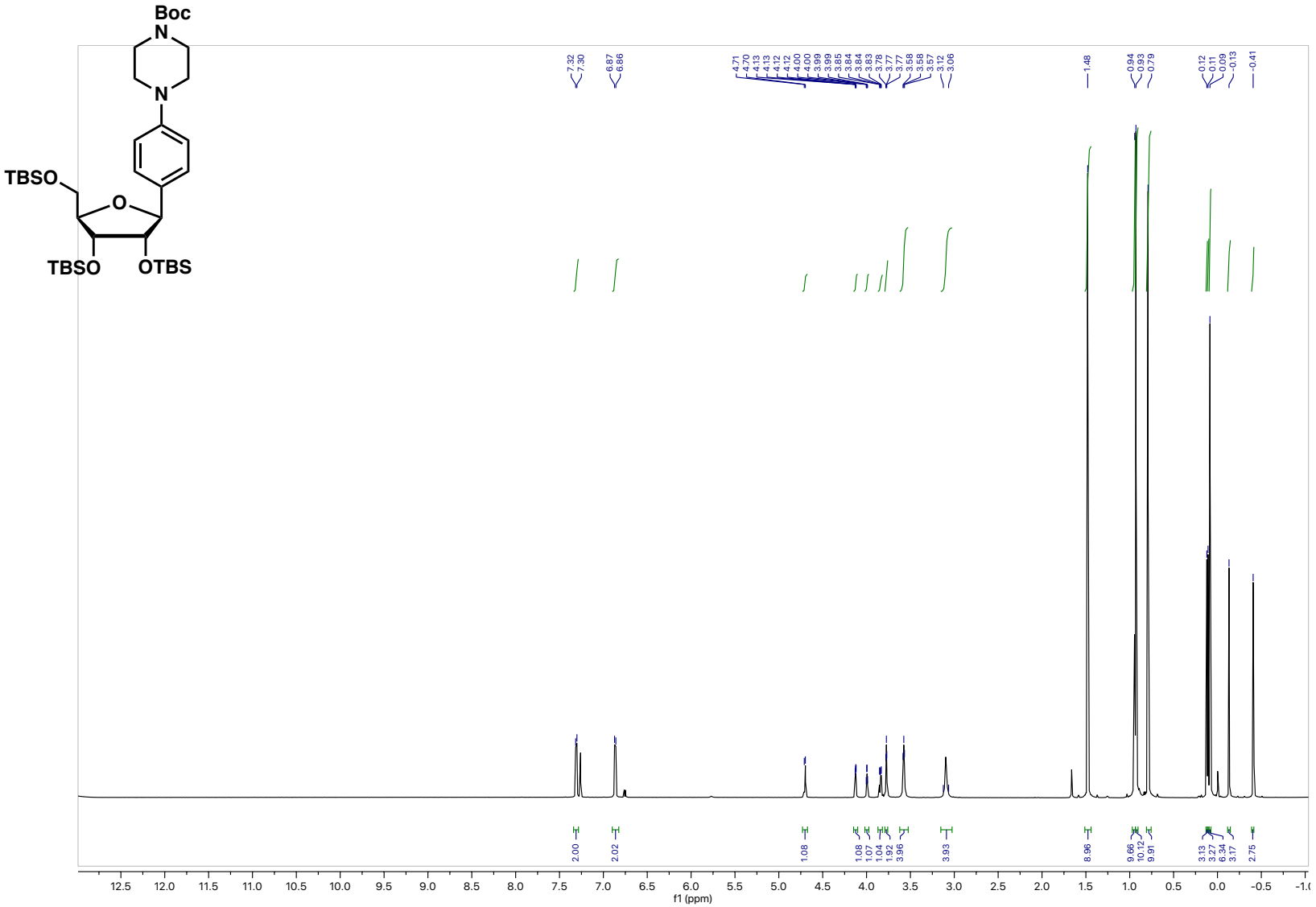




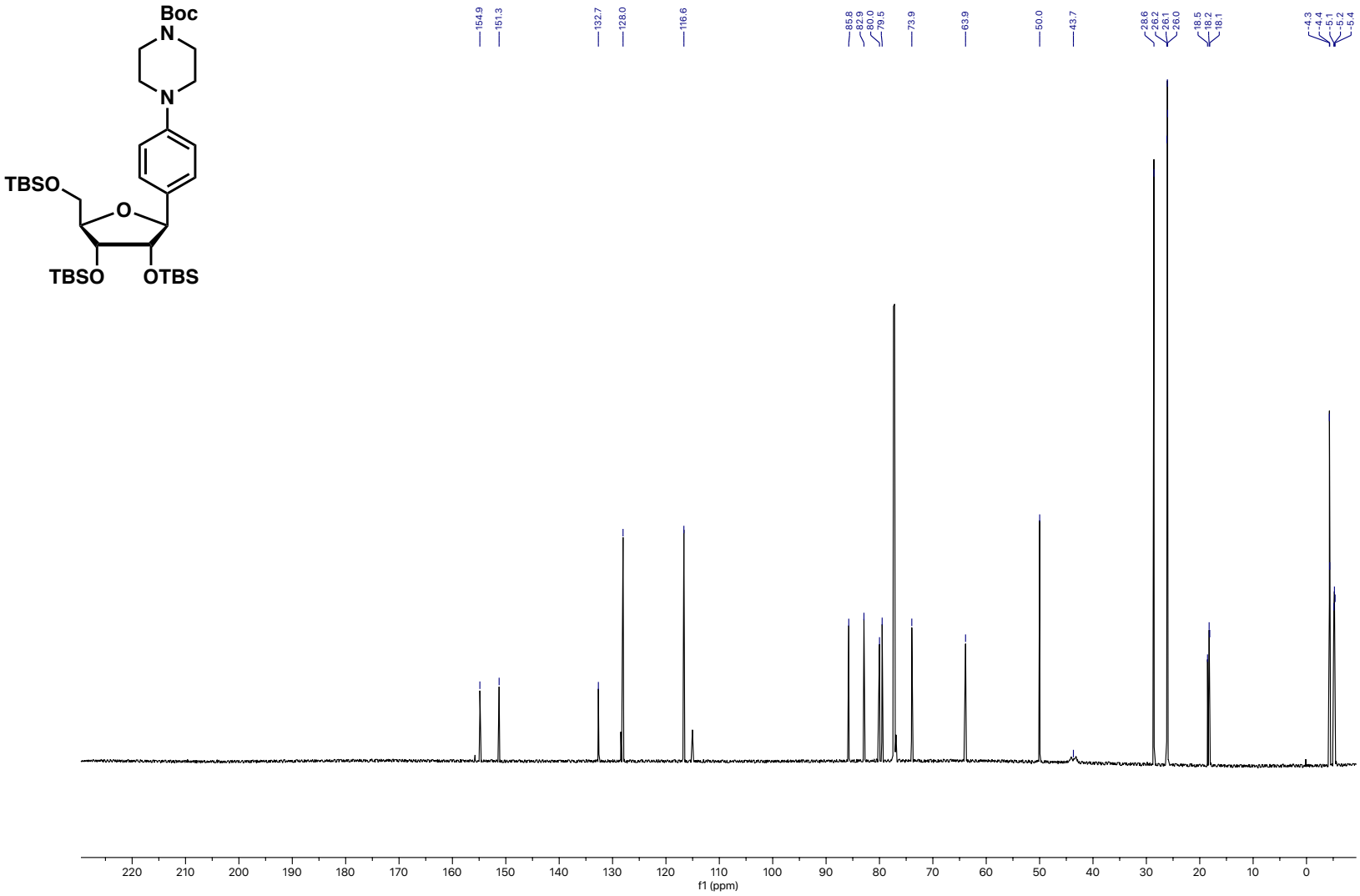
Compound 45 ^{13}C NMR



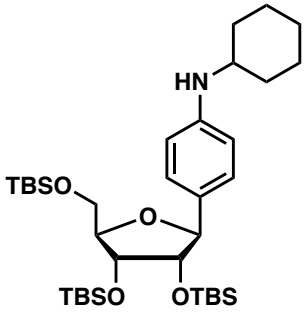
Compound 46 ^{1}H NMR



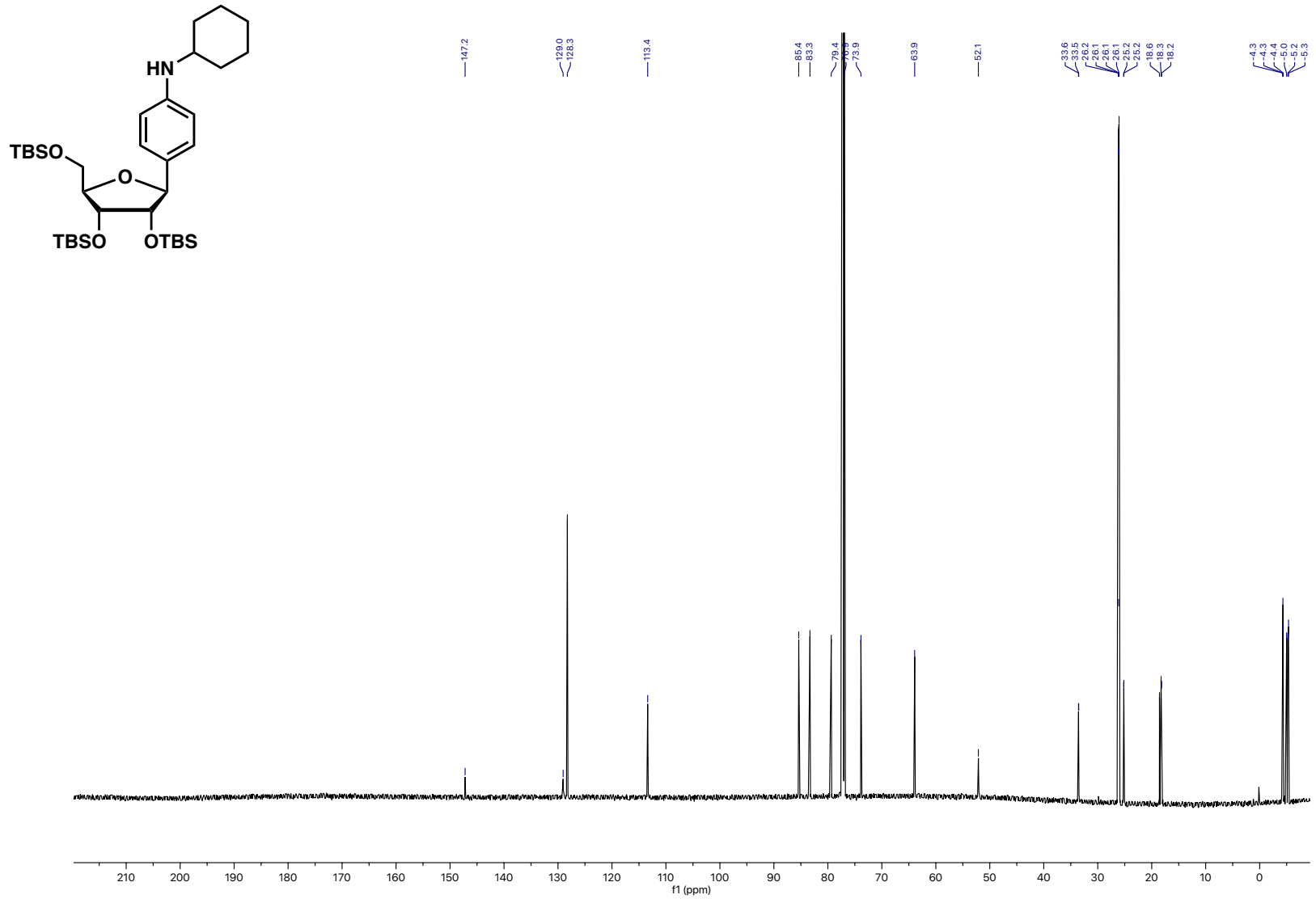
Compound 46 ^{13}C NMR



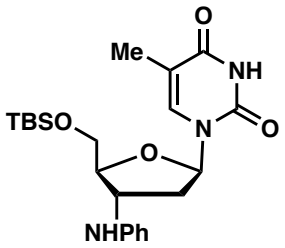
Compound 47 ^{1}H NMR



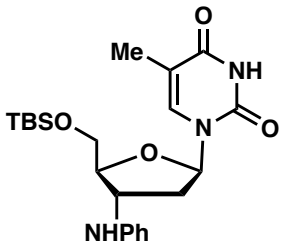
Compound 47 ^{13}C NMR



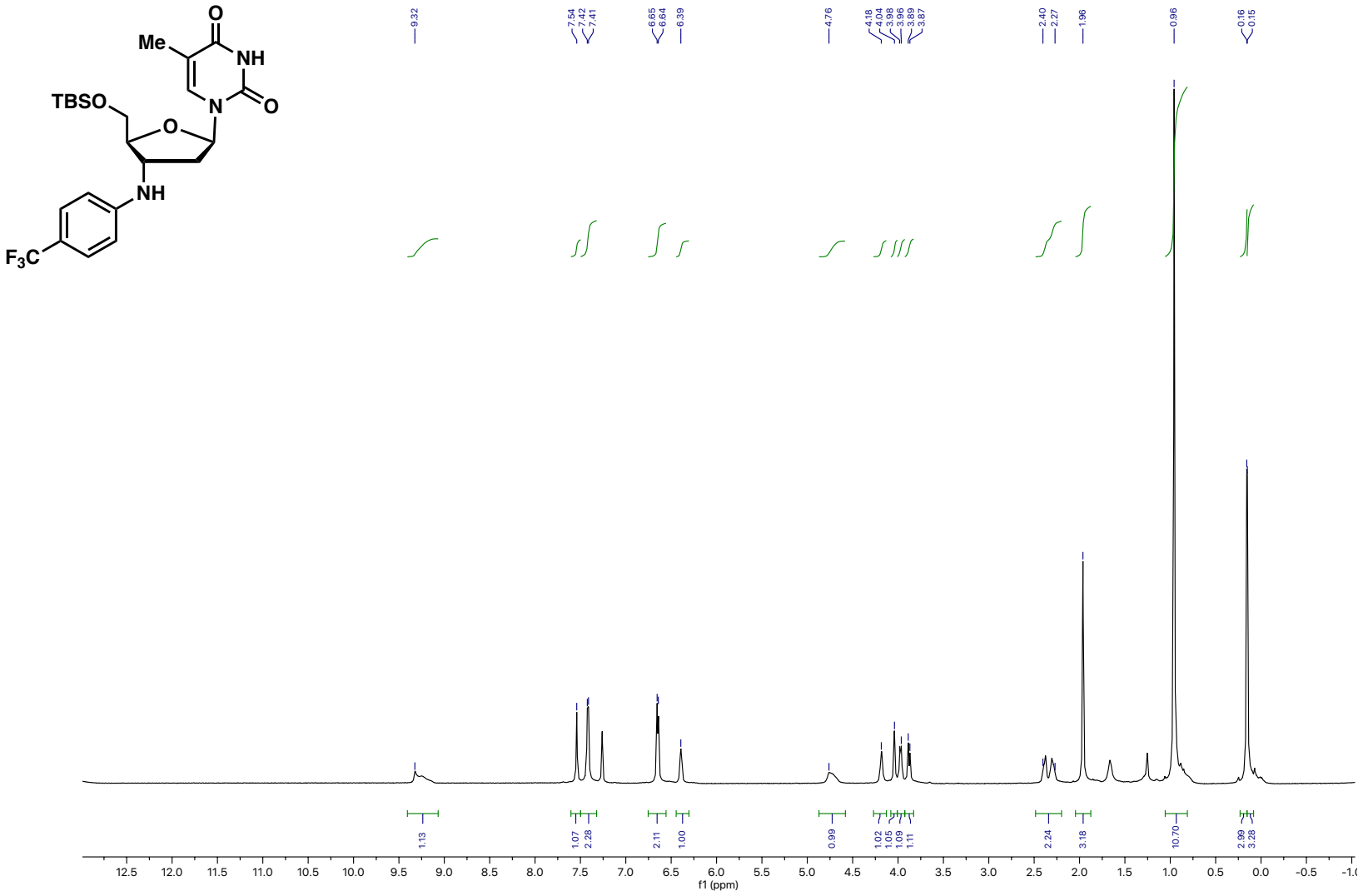
Compound 48 ^{1}H NMR



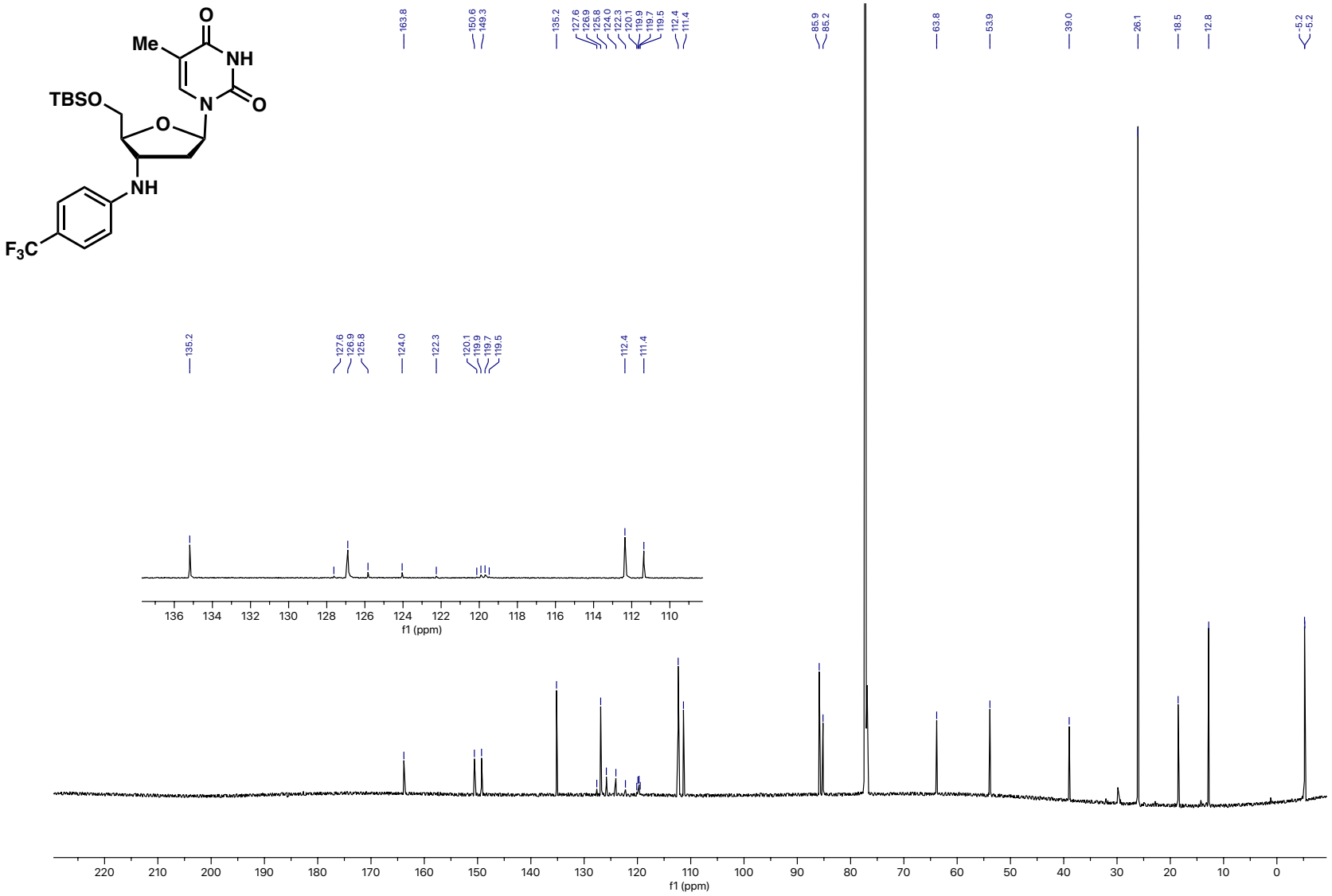
Compound 48 ^{13}C NMR



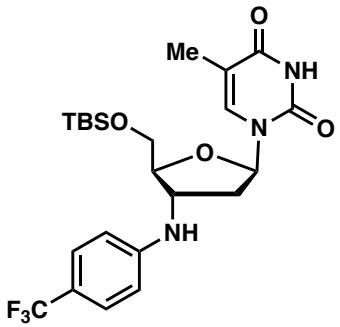
Compound 49 ^{1}H NMR



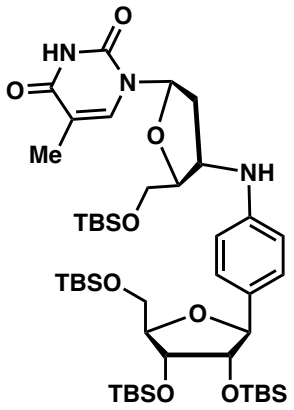
Compound 49 ^{13}C NMR



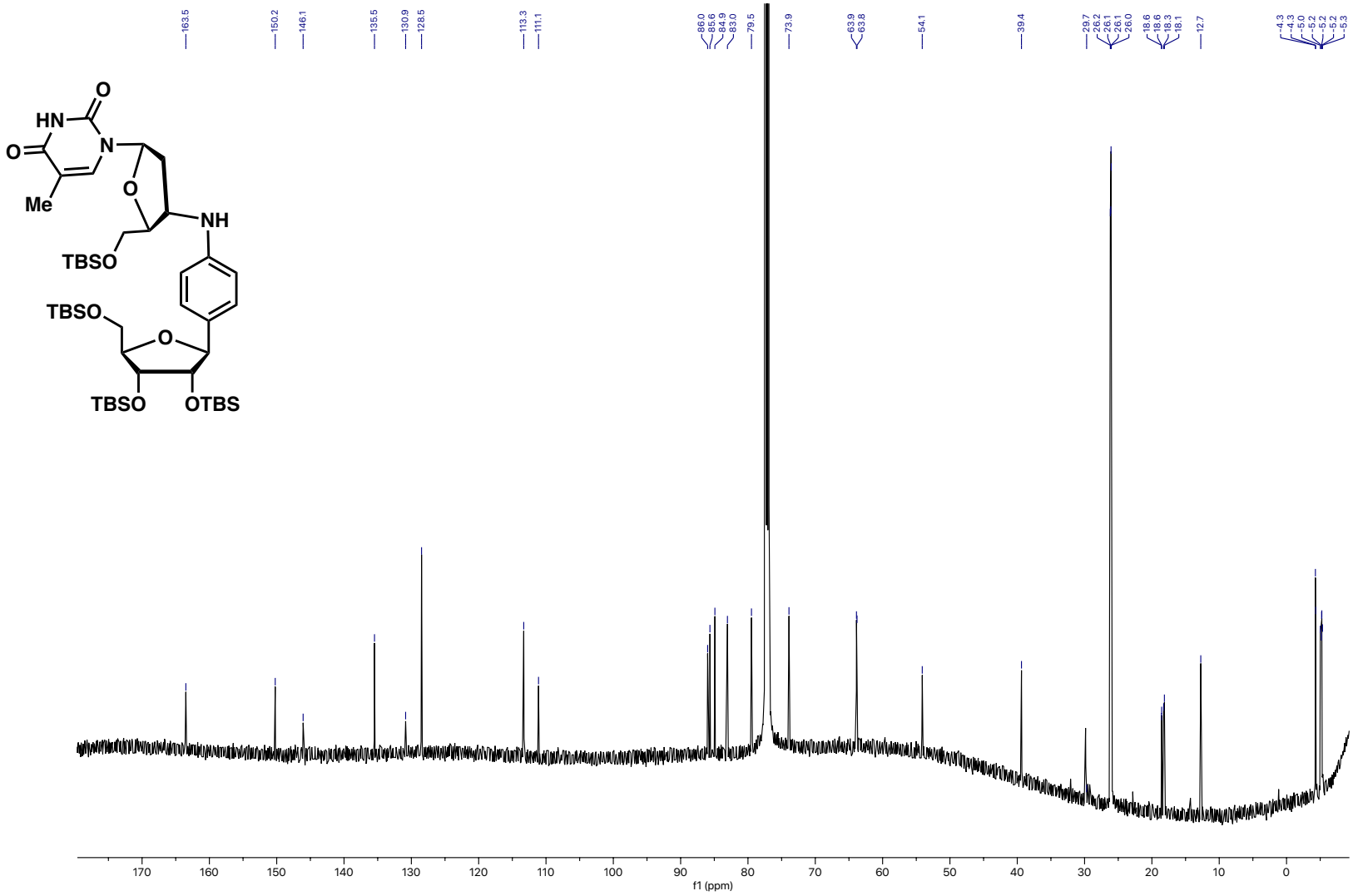
Compound 49 ^{19}F NMR



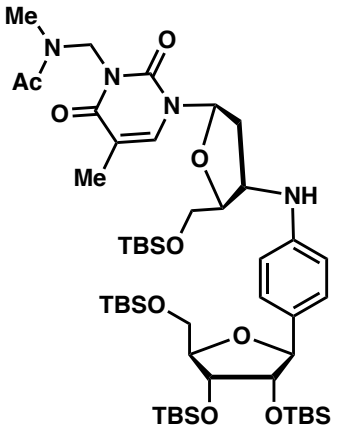
Compound 50a ^{1}H NMR



Compound 50a ^{13}C NMR



Compound 50b ^{1}H NMR



Compound 50b ^{13}C NMR