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Controlling the assembly of C₂-symmetric molecular tectons using a thiocarbamate appended carbocyclic cleft molecule analogous to Tröger's base

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ABSTRACT

By way of appending the C₂-symmetric carbocyclic cleft diol with thiocarbamates with varying substituents, significant control of the hydrogen bonded network can be achieved. Smaller alkyl substituents lead to the formation of stacked columns of components with the apex of one molecule suitably aligned in the cleft of a second. Aryl substituents however, lead to the formation of ribbons via an H-bonding network. Additionally, the packing of these ribbons into networks is considerably different between the enantiopure and racemic clefts, with the latter giving rise to channels within the crystal structure.

INTRODUCTION

The carbocyclic diol cleft molecule **1**¹⁻³ has become a surrogate for Tröger's base (**2**) in catalysis,⁴ supramolecular chemistry⁵⁻⁷ and chiral recognition (figure 1).⁸⁻¹¹ It is analogous to Tröger's base as it contains a C₂-symmetric axis, a chiral cavity with a defined geometry and a rigid, predictable structure. This give **1** a predictable V-shape, whose angle (θ) can vary between 83.4° to 104.3° depending on the substitution on the aromatic ring.^{12,13} The advantages of **1** over Tröger's base lie in the diol, with the two hydroxyl groups being pointed into the interior of the cleft giving the ability, via hydrogen bonding, to orientate and organize substrates into this uniquely chiral cavity. We have previously emphasized the practicality of **1** by appending the hydroxyl groups with pyridyl functionality, as well as highlighting the self-assembly of these pyridyl adducts with phenylboronic acid to yield bis-boroxine substrates, with the 4-pyridyl analogue being fully characterized by single crystal X-ray analysis emphasizing the chiral space.^{14,15}

Figure 1. The C₂-centrosymmetric chiral cleft diol **1**.

The capacity to be able to confidently predict crystal packing to facilitate the design of new solids with sought after physical properties has been central to the evolution of crystal engineering.¹⁶⁻¹⁸ Furthermore, the ability to control secondary and tertiary structure within crystal packing has enormous potential in the development of new functional materials and devices. Molecular tectonics¹⁹⁻²² is the analysis of molecular crystals and their subsequent formation of networks, and as a consequence the tecton is defined as a construction unit with

defined structural and energetic attributes that can control assembly via programmed molecular recognition processes. Consequently, the identification of new 3-dimensional chiral molecules, or tectons, with rigid, predictable structures opens further prospects in expanding the scope of crystal engineering.

Akin to Troger's base (**2**),^{23,24} diol **1** can pack, in the solid state, in four possible forms **M1**, **M2**, **M3** and **M4**, all of which are primarily driven by weak van der Waals, and π - π interactions (figure 2).²⁵

Figure 2. Control of crystal packing in chiral cleft diol **1**.

Conversely, thiocarbamates (**3**) like amides, form networks based on stronger hydrogen bonding, which is, in part, controlled by their configuration, with the *cisoid* form **3a** leading to synthon **4**, and the *transoid* form **3b** leading to synthon **5** (scheme 1).^{26,27}

Scheme 1. Control of crystal packing in thiocarbamate **3**.

In an effort to regulate the crystal packing of chiral cleft **1**, we envisaged appending the diol with a variety of thiocarbamates (alkyl and aryl, scheme 2, **6**). The crystal packing of these derivatives would then be primarily defined by the orientation of the thiocarbamate (*cisoid* and/or *transoid*) that is accurately described by **6a** $\leftrightarrow$ **6b** $\leftrightarrow$ **6c**. This would serve two purposes: (a) to allow the confident prediction of the crystal packing of **6**, therefore demonstrating this scaffold as a potential tecton; (b) to demonstrate that novel crystalline solids with predictable chiral environments can be formed from analogues of **1**.

Scheme 2. Targeted thiocarbamates.

RESULTS AND DISCUSSION

Preparation of thiocarbamate appended clefts

The synthesis of the thiocarbamate cleft derivatives began with obtaining multi-gram quantities of the parent cleft diol **1**, and this was achieved as reported previously (scheme 3).² This racemic diol could then be converted to the *bis*-thiocarbamate by treatment with NaH and excess isothiocyanate. This was accomplished with phenyl, tolyl, 3,5-ditrifluoromethylphenyl, methyl, and butyl isothiocyanates giving the desired compounds ($\pm$)-**10a-e** in moderate to good yields. In addition to the racemic thiocarbamates, enantiopure tolyl

thiocarbamate was also prepared (scheme 3). This was achieved by synthesizing the enantiopure di-acid **(+)-8** via resolution of its racemate with quinine. This enantiopure diacid was then converted to the thiocarbamate, delivering enantiopure **(+)-10b**.

Scheme 3. Preparation of thiocarbamates **(±)-10a-e** and **(+)-10b**.

X-ray crystallographic analysis

Single crystal X-ray analysis was performed on phenyl **(±)-10a**, tolyl **(±)-10b**, 3,5-difluoromethylphenyl **(±)-10c**, methyl **(±)-10d**, and butyl **(±)-10e** thiocarbamates. Additionally, single crystal X-analysis was also performed on the enantiopure tolyl **(+)-10b** analogue as a comparison with its racemic analogue. Crystal data, H-bond geometry, and 50% ORTEP plots are presented in the SI, and crystallographic procedures are described in the experimental section.²⁸

The crystal structure of the racemic phenyl-amino-thiocarbamate cleft **(±)-10a** illustrates the rigid cleft framework and the position of the two appending phenyl-amino-thiocarbamate

units that are orientated towards the interior of the cleft.²⁸ Interestingly the sulfur and thiocarbamate N–H groups are pointing outwards away from the cleft cavity such that only the aromatic phenyl groups are directed inwards towards the cleft. Molecules of ($\pm$)-**10a** were shown to interact with each other according to the packing plot shown in figure 3(a). Thus, molecules of ($\pm$)-**10a** are linked together in a 1D polymeric H-bonded chain with amino-thiocarbamate groups forming pairs of $R^2_2(8)$, centro-symmetric N–H $\cdots$ S H-bond interactions to neighboring molecules.^{26,27,29}

Figure 3. X-ray analysis and the H-bonded chain of ($\pm$)-**10a**, (+)-**10b** and ($\pm$)-**10b**, respectively.

(a) ($\pm$)-**10a**

(b) (+)-**10b**

(c) ($\pm$)-**10b**

The X-Ray crystal structure³⁰ of the (+)-enantiomer of the *p*-tolyl-amino-thiocarbamate cleft (**+**)-**10b**, shows that the substituted amino-thiocarbamate groups are also directed into the cavity of the cleft bicycle with the thiocarbamate sulfur and N–H groups pointing outwards away from the center of the cavity.²⁸ Molecules of (**+**)-**10b** were bound to each other by a very similar H-bonding network as displayed in the crystal structure of ($\pm$)-**10a**. Thus molecules are linked together via pairs of $R^2_2(8)$, centro-symmetric N–H $\cdots$ S H-bonds forming chains of molecules (figure 3(b)).³¹ Essentially the molecular structure of ($\pm$)-**10b** is exactly the same as the (+)-enantiomer with complexing *p*-tolyl-amino-thiocarbamate groups pointing into the cavity of the cleft bicycle and both the N–H and sulfur groups of the appending amino-thiocarbamate groups pointing away from the interior of the cavity.³⁰ Like in (**+**)-**10b**, this arrangement of functional groups allowed molecules to form chains linked by pairs of $R^2_2(8)$, centro-symmetric N–H $\cdots$ S H-bonds (figure 3(c)).³²

However, unlike the crystal structure of (**+**)-**10b**, which was found to contain crystallographically independent molecules with no solvent of crystallization, the crystal structure of the racemic compound ($\pm$)-**10b** contained solvate molecules of methanol which lie disordered over a two-fold axis *exo*- to the cleft molecules. In addition, an acetonitrile solvate molecule bound via a pair of C–H $\cdots$ π interactions resides in the cleft cavity (figures 3c and 4b). Interestingly, because the cleft cavity is occupied with solvent molecules bound

to the cleft by moderate C–H $\cdots\pi$ interactions, neighboring H-bonded chains made up of molecules of ($\pm$)-**10b** are unable to interact as they did in (+)-**10b** where the terminal *p*-tolyl group from one chain penetrates into the cavity of the molecule in the next chain forming an ‘above / below’ type packing pattern (type **M2** figure 4(a)). Instead chains of H-bonded molecules arrange into groups of six molecules which interact to form rings with the internal clefts filled with acetonitrile solvate molecules (figure 4(b)).

Figure 4. Comparison of the packing pattern of enantiopure and racemic **10b**, both of which illustrate **M2**-type pack of the C_2 -centrosymmetric cleft.

(a) (+)-**10b**

(b) ($\pm$)-**10b**

In the crystal structure of ($\pm$)-**10c**, the fluorine atoms which make up the CF_3 groups were modeled as split over two sets of positions with one of the $C_6H_3CF_3$ groups²⁸ showing the potential to be more disordered than the others.³³ Like the other *N*-aryl functionalized amino-thiocarbamate clefts the $Ph(CF_3)_2$ substituted amino-thiocarbamate groups in ($\pm$)-**10c** are directed into the cleft cavity with the thiocarbamate sulfur and N–H groups directed away from the interior of the cleft. It would appear from X-Ray packing plots that the bulky pendant fluorinated aromatic groups block the cleft cavity, preventing any interactions between the π -systems of neighboring cleft molecules. As a result, the only significant non-

covalent interactions observed are H-bonds in ($\pm$)-**10c** between the thiocarbamate N–H groups and the sulfur atom of a neighboring molecule resulting in a 2-dimensional H-bonded sheet via one $R^2_2(8)$, centro-symmetric motif and several chain linkages (figure 5).

Figure 5. X-ray H-bonding analysis of ($\pm$)-**10c**. The 3,5-ditrifluoromethylphenyl groups have been omitted for clarity.

The length of the H-bonds between the sulfur and N–H groups³⁴ are very similar to those observed in (+)-**10b** and ($\pm$)-**10b**, however the overall packing pattern displayed in this structure is considerably more complex due to the fluoroaryl groups.

As in the *p*-tolyl, phenyl and fluoroaryl substituted amino-thiocarbamate groups the appending methyl-thiocarbamate groups of ($\pm$)-**10d**³⁰ are directed into the cavity of the cleft.²⁸ However, unlike the *N*-aryl substituted thiocarbamate clefts mentioned previously, the thiocarbamate N–H groups in ($\pm$)-**10d** are shown to be directed into the cavity of the cleft; presumably the less bulky methyl groups facilitate this orientation. This removes the ability of the thiocarbamate N–H groups to form centro-symmetric H-bonds with neighboring clefts, as demonstrated by the clefts (+)-**10b** and ($\pm$)-**10b** (figures 3 and 4), which appears to influence the way in which molecules of ($\pm$)-**10d** interact with each other. The packing pattern displayed by ($\pm$)-**10d** showed that molecules are linked via N–H $\cdots$ S H-bonds into

chains / ladders, thus the apex of one molecule is accommodated in the cleft of the next to form columns of apex-to-base stacked molecules, like a stack of party hats (type **M1**).³⁵ The two hydrogens in the CH₂ at the bridgehead of the cleft at C(9); C(30) and C(51) (figure 6(a)) make up pairs of C–H··· π interactions with the cleft of the molecule ‘above’.³⁶ In addition to this molecular stacking pattern, this cleft presents a highly unusual cooperative packing system, where each group of three molecules is H-bonded to the next group of three molecules but with a 180° rotation between one group and the next.³⁷

Weakly diffracting crystals of ($\pm$)-**10e** were obtained and analyzed using synchrotron radiation.^{28,30} There were difficulties in uniquely identifying the space group which are described in the experimental section. Despite these problems the identification of the compound and the nature of the stacking was clearly established (figure 6(b)). The stacking adopted is the same **M1** motif adopted by the methyl analog ($\pm$)-**10d**.

Figure 6. X-ray analysis of ($\pm$)-**10d** and ($\pm$)-**10e**, respectively illustrating the H-bonded ladder network.

(a) ($\pm$)-**10d**

(b) ($\pm$)-**10e**

From the X-ray data it is clear that the size of the R group on the thiocarbamate effects the orientation and packing of the cleft. In the case of the R being alkyl (methyl or *n*-butyl) both the thiocarbamates take on the *cisoid* conformation (scheme 4, **6a**) and this promotes *synthon 4* giving ($\pm$)-**11d** and ($\pm$)-**11e**, respectively. This is further reinforced by weak, non-bonding interactions forming columns of two or more components with the apex of one molecule suitably aligned in the cleft of a second, and can be defined as **M1** type packing.

Scheme 4. The role of the thiocarbamate in determining crystal structure and the H-bonded network.

However, when R is an aryl group the preferred conformation of the thiocarbamate is *transoid* (**6c**) which promotes *synthon 5* giving ribbons of clefts within the crystal structures ($\pm$)-**11a,b** and (+)-**11b**. The packing of these ribbons in the enantiopure clefts (+)-**11b** is then

controlled by the insertion of an aryl group of one cleft into the cleft of another via endo-face to endo-face interactions, and can be defined as **M2** type packing. In contrast, in the racemic series ($\pm$)-**11a,b** the packing of these ribbons gives rise to a new motif **M5**, which promotes the formation of rings with the internal clefts filled with acetonitrile solvate molecules. Clearly this latter network provides the opportunity to form other host:guest complexes using these channels. Finally, the 3,5-CF₃Ph thiocarbamate ($\pm$)-**10c**, which contains bulky pendant fluorinated aromatic groups that block the cleft cavity, presumably forms networks using the *cisoid* (**6a**), *transoid* (**6c**) or mixtures of both forms (**6b**), giving a disordered H-bonded network.

CONCLUSIONS

Through using a combination of a novel chiral cleft molecule and thiocarbamates we have demonstrated that we can control the crystal packing by simple variation of the size of the substituent on the thiocarbamate. This has been achieved due to the tendency of the C₂-symmetric cleft to form molecule associations within the solid state described as **M1–M4**, and thiocarbamates to form synthons via their *cisoid* and *transoid* conformers. Smaller substituents lead to H-bonding in the thiocarbamate in its *cisoid* form, and this is further strengthened by type **M1** interactions. Larger substituents such as phenyl and tolyl lead to H-bonding ribbon type networks via the *transoid* form of the thiocarbamate. However, we have demonstrated that the packing of these ribbons differs significant between the racemic and enantiopure clefts, with the racemic series giving rise to channels within the crystal structure, via a new type of interaction **M5**. Moreover, the crystal structure of ($\pm$)-**10b** provided evidence of the ability of these novel amino-thiocarbamate clefts to bind guest molecules with complementary size and binding features. Interestingly, binding within the cleft cavity was facilitated by C–H $\cdots\pi$ H-bond interactions originating from the core carbocyclic cleft

framework, and not by H-bonds involving the thiocarbamate groups. Given that cleft **1** is C₂-symmetric, this simple but effective control of crystallization within the solid state has significant potential to be expanded upon using other hydrogen bond synthons.

EXPERIMENTAL

General Procedure for the Preparation of Amino-thiocarbamate Clefts. Sodium hydride (60 % mineral oil, 4.0 mmol) was washed with hexane (3 x 10 mL) and dried under vacuum, THF (10 mL) was added and the suspension was cooled to 0°C. Racemic (±)-**1** or (+)-**1** (1.0 mmol) was dissolved in THF (5 mL) and added to the sodium hydride suspension *via* a cannula under nitrogen and the reaction was stirred at 0°C for 25 min. After this time a solution of substituted isothiocyanate (3.0 mmol) in THF (5 mL) was added dropwise to the reaction mixture. The solution was allowed to warm to room temperature and then stirred for a further 18 h; reactions were monitored by TLC. After this time the reaction was quenched by the careful addition of water (20 mL) and extracted with dichloromethane (3 x 25 mL). The combined organic layers were washed with brine (25 mL), dried over MgSO₄, filtered and concentrated to dryness in vacuo. The crude compound was purified by silica column chromatography.

(±)-2,8-Dibenzobicyclo [b,f] [3.3.1]nona-5a, 6a-dien-6,12-phenyl amino thiocarbamate (±)-10a. (0.2 g, 57 %), fine white solid; Mp 111.6 – 114.7 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.74 – 7.00 (m, 20 H), 3.91 – 3.88 (m, 2H), 2.60 – 2.58 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 188.41, 136.86, 136.65, 134.61, 133.96, 130.67, 130.48, 129.04, 127.66, 127.47, 126.77, 125.26, 121.42, 83.74, 72.59, 39.22, 35.62, 29.08; IR CH₂Cl₂ max/cm⁻¹: 3943, 3584, 3361 (N–H), 3054, 2986 (aromatics), 2305, 1526, 1265, 1160, 1034, 743, 705; HRMS *m/z* found 545.1327 C₃₁H₂₆O₂N₂S₂ (M⁺ Na) expected 545.1328 (M⁺ Na); Elemental Analysis found C 71.06; H 4.98; N 5.41 % C₃₁H₂₆N₂O₂S₂ requires C 71.24; H 5.01; N 5.36 %.

(±)-2,8-Dibenzobicyclo [b,f] [3.3.1]nona-5a, 6a-dien-6,12-*p*-tolyl amino thiocarbamate

(±)-10b. (0.4 g, 70 %) fine white solid; Mp 209.9 – 211.2 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.58 (d, *J* = 8.4 Hz, 2H), 7.29 – 7.15 (m, 7H), 7.09 (d, *J* = 7.2 Hz, 2H), 7.04 – 6.84 (m, 7H), 3.89 (m, 2H), 2.58 (m, 2H), 2.41 (s, 3H), 2.28 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 188.30, 136.32, 135.36, 134.92, 134.83, 133.85, 131.05, 129.72, 129.54, 127.39, 127.11, 126.55, 123.59, 121.47, 83.37, 79.45, 46.00, 35.36, 20.93; IR CH₂Cl₂ max/cm⁻¹: 3944 (s, N–H), 3689 (s, N–H), 2986 (aromatics), 2305, 1520, 1265, 739, 704; HRMS *m/z* found 573.1639 C₃₃H₃₀O₂N₂S₂ (M⁺ Na) expected 573.1641 (M⁺ Na); Elemental analysis found C 71.55; H 5.46; N 5.13 %; C₃₃H₃₀N₂O₂S₂ requires C 71.97; H 5.49; S 5.09 %.

(+)-2,8-Dibenzobicyclo [b,f] [3.3.1]nona-5a, 6a-dien-6,12-*p*-tolyl amino thiocarbamate

(+)-10b (0.26 g, 24 %), fine white solid; Mp 116.2 – 124.7 °C; [α]_D²¹ + 174.3 (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.47 (d, *J* = 7.6 Hz, 2H), 7.17 – 7.08 (m, 7H), 6.95 (d, *J* = 5.2 Hz, 2H), 6.89 – 6.75 (m, 7H), 3.79 (m, 2H), 2.46 (m, 2H), 2.30 (s, 3H), 2.14 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 188.21, 136.36, 134.84, 134.30, 131.08, 129.61, 129.54, 127.14, 126.56, 123.63, 121.52, 83.34, 79.44, 35.42, 28.82, 20.99; IR CH₂Cl₂ max/cm⁻¹: 3944, 3054 (N–H), 2986 (aromatics), 2305, 1522, 1265, 1171, 1034, 739, 704; HRMS *m/z* found 573.1642 C₃₃H₃₀O₂N₂S₂ (M⁺ Na) expected 573.1641 (M⁺ Na); Elemental analysis found C 71.79; H 5.57; N 5.16 %; C₃₃H₃₀N₂O₂S₂ requires C 71.97; H 5.49; N 5.09 %.

(±)-2,8-Dibenzobicyclo [b,f] [3.3.1]nona-5a, 6a-dien-6,12-bis-3,5- bis (trifluoromethyl)

phenyl amino thiocarbamate (±)-10c. (0.70 g, 44 %), off-white solid; Mp 120.1 – 127.4 °C;

¹H NMR (400 MHz, CDCl₃): δ 8.00 – 6.20 (m, 14H), 3.85 (s, 2H), 1.44 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 194.42, 140.05, 134.52, 128.85, 138.83, 128.79, 128.30, 48.82, 32.33; IR CH₂Cl₂ max/cm⁻¹: 3204, 3055 (N–H), 2987 (aromatics), 2306, 1551, 1139, 1265, 1031, 738, 703; HRMS *m/z* found 817.0777 C₃₅H₂₂O₂N₂S₂F₁₂ (M⁺ Na) expected 817.0823 (M⁺ Na);

Elemental analysis found C 52.34; H 2.77; N 3.72 % $C_{35}H_{22}F_{12}N_2O_2S_2$ requires C 52.90; H 2.79; N 3.53 %.

(±)-2,8-Dibenzobicyclo [b,f] [3.3.1]nona-5a, 6a-dien-6,12-methyl amino thiocarbamate

(±)-10d. (0.60 g, 75 %), colorless solid; Mp 212.1 – 218.2 °C; 1H NMR (400 MHz, $CDCl_3$): δ 7.12 – 7.04 (m, 8H), 6.79 (dd, J = 6.8 Hz, 1H), 6.35 (dd, J = 6.0 Hz, 1H), 3.74 (m, 1H), 3.66 (m, 1H), 3.13 (t, J = 5.2 Hz, 6H) 2.64 (d, J = 5.2 Hz, 1H), 2.53 (d, J = 5.2 Hz, 1H), 1.97 (m, 2H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 194.1 90.09, 135.52, 135.43, 135.26, 134.88, 130.98, 127.36, 127.31, 127.20, 126.99, 126.81, 126.77, 126.71; IR CH_2Cl_2 max/ cm^{-1} : 3399 (N–H), 3052 (aromatics), 1528, 1265, 1068, 737, 703; HRMS m/z found 421.1013 $C_{21}H_{22}O_2N_2S_2$ (M^+ Na) expected 421.1015 (M^+ Na); Elemental analysis found C 63.07; H 5.58; N 6.99 % $C_{21}H_{22}N_2O_2S_2$ requires C 63.29; H 5.56; N 7.03 %.

(±)-2,8-Dibenzobicyclo [b,f] [3.3.1]nona-5a, 6a-dien-butyl amino thiocarbamate (±)-10e.

(0.19 g, 20 %), colorless solid; Mp 213.3 – 214.4 °C; 1H NMR (400 MHz, $CDCl_3$): δ 7.46 – 6.70 (m, 12H), 5.01 (m, 2H), 3.74 (s, 1H), 3.31 (s, 1H), 2.49 – 2.36 (m, 2H), 2.25 (s, 3H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 188.51, 130.59, 129.56, 127.63, 127.44, 126.73, 121.50, 83.63, 72.60, 39.22, 35.61, 29.09, 21.31; IR CH_2Cl_2 max/ cm^{-1} : 3257 (N–H), 2957, 2931 (aromatics), 1519, 1179, 1038, 752; HRMS m/z found 505.1954 $C_{27}H_{34}O_2N_2S_2$ (M^+ Na) expected 505.1954 (M^+ Na); Elemental analysis found C 66.96; H 7.02; N 5.73 % $C_{25}H_{23}NO_2S$ requires C 67.18; H 7.10; N 5.80 %.

X-ray Crystallography

Diffraction data for (±)-10a· $CHCl_3$ and (±)-10b· $MeOH \cdot CH_3CN$, and were collected on a Bruker APEX 2 CCD diffractometer equipped with graphite-monochromated Mo- $K\alpha$ X-radiation. Diffraction data for (+)-10b, (±)-10c·0.25 C_5H_{12} , and (±)-10d and were collected on

a Bruker APEX 2 CCD diffractometer equipped with a silicon 111 monochromator using narrow frame ω -scans on Station 11.3.1 at the ALS using synchrotron radiation.³⁸ Data for ($\pm$)-**10e** were also collected at the ALS but with a Bruker D8 diffractometer with a CMOS detector using shutterless ω -scans. Data were corrected for absorption and Lp effects.³⁸ Structures were solved by direct methods or charge flipping algorithms^{39,40} and refined by full-matrix least squares on F^2 .⁴¹ Further details are given in Table 7 in the ESI. In ($\pm$)-**10a**·CHCl₃ the CHCl₃ molecule was modeled as disordered over two sets of positions with major component 63.6(6)%. In ($\pm$)-**10b**·MeOH·CH₃CN the MeOH molecule is disordered over a 2-fold axis and the OH position could not be reliably determined, so was not refined. In ($\pm$)-**10c**·0.25C₅H₁₂ there are two cleft molecules in the asymmetric unit, both exhibiting disorder. CF₃ groups at C(26), C(34), C(35), C(69) and C(70) were all modeled as two-fold disordered, as were the ring atoms C(54) > C(57) and the CF₃ group at C(60). The badly disordered partial pentane of crystallization was modeled as a diffuse area of electron density by the Platon Squeeze procedure.⁴² The structure of ($\pm$)-**10e** is presented as provisional due to the severe crystallographic problems encountered associated with space group ambiguity. Data sets were collected both in the home laboratory and at the ALS synchrotron. In both cases the same unit cell parameters were obtained. Crystal data for ($\pm$)-**10e**: C₂₇H₃₄N₂O₂S₂, $M = 482.68$, monoclinic, Pc (or $P2_1/c$), $a = 12.2150(11)$, $b = 26.553(2)$, $c = 24.029(2)$ Å, $\beta = 99.357(5)^\circ$, $V = 7690.0(11)$ Å³, $Z = 12$, $\mu = 0.234$ mm⁻¹, 77800 reflections measured, 25961 unique, $R_{\text{int}} = 0.0749$, $R_1[F^2 > 2\sigma(F^2)] = 0.2110$. Similarity restraints were applied to the geometry of all molecules and global restraints were required for anisotropic displacement parameters. Six molecules in the asymmetric unit in two groups of three for space group Pc , which gave the best result (one group of three in space group $P2_1/c$). N–H···S H-bonds in the range 3.28(2) – 3.541(13) Å and C–H··· π distances in the range 2.41 – 2.75 Å.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website. Experimental procedures for the synthesis of (+)-**9**, (-)-**9**, (±)-**9**, as well as the HPLC traces of (+)-**9**, (-)-**9** and (±)-**9**. Spectroscopic data for all reported compounds. Table S1 contains the crystallographic data for (±)-**10a**, (+)-**10b**, (±)-**10b**, (±)-**10c** and (±)-**10d**. Tables S2-S7 contain selected H-bonded geometries for (±)-**10a**, (+)-**10b**, (±)-**10b**, (±)-**10c** and (±)-**10d**, respectively. Figures S1-S6 contains ORTEP plots of (±)-**10a**, (+)-**10b**, (±)-**10b**, (±)-**10c** and (±)-**10d** (two molecules), respectively. Figure S7 depicts the H-bonding network of (±)-**10c**, and Figure S8 the H-bonding network of (±)-**10e**.

Accession Codes

CCDC 1441482-1441486 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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Notes

The authors declare no competing financial interests.

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28. ORTEP plots of compound ($\pm$)-**10a-e** and (+)-**10b** can be found with in the Supporting information.
29. See table S1 with the Supporting Information.
30. Due to small crystal dimensions this data set was collected using synchrotron radiation at the ALS.
31. See table S2 with the Supporting Information.
32. See table S3 with the Supporting Information.
33. Modeled with atoms C(54) > C(57), C(60) and F(13) > F(15)
34. See table S4 with the Supporting Information.
35. See table S5 with the Supporting Information for the geometry of the C–H... π H-bond interactions between stacked molecules of ($\pm$)-**10d**.
36. See table S6 with the Supporting Information.
37. See figure S7 within the Supporting information.
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Controlling the assembly of C₂-symmetric molecular tectons using a thiocarbamate appended carbocyclic cleft molecule analogous to Tröger's base

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By appending the C₂-symmetric carbocyclic cleft diol, an analogue of Tröger's base, with thiocarbamates with varying substituents (both alkyl and aryl) gives significant control of the H-bonded network. This leads to unique solid-state structures that have been fully characterized by crystallographic analysis. Additionally, a new type of intermolecular stacking interaction of the carbocyclic cleft molecules (M5), has been observed.