## (Supporting Information)

# Chemical Constituents from the Peels of Citrus sudachi 

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## Known Compounds from the Peels of Citrus sudachi

Following 27 known compounds were isolated from the peels of $C$. sudachi; isoobacunoic acid (8), ${ }^{1}$ limonin (9), ${ }^{2}$ methyl deacetylnomilinate (10), ${ }^{3}$ nomilinic acid (11), ${ }^{4}$ vanillic acid (12), ${ }^{5} 1 S, 2 S, 4 R$-limonene-1,2-diol (13), ${ }^{6}(+)-4 S$-7-hydroxypiperitone (14), ${ }^{7}$ methyl ferulate (15), ${ }^{8}$ ferulic acid (16), ${ }^{9}$ citrusin III (17), ${ }^{10}$ citrusin IX (18), ${ }^{11}$ sudachitin (6), ${ }^{12} 3^{\prime}$-demethoxysudachitin (7), ${ }^{12} 7$-methylsudachitin (19), ${ }^{13}$ xanthomicrol (20), ${ }^{13}$ jaceosidin (21), ${ }^{14}$ sudachiin $\mathrm{B}(\mathbf{2 2}),{ }^{15}$ sudachiin $\mathrm{C}(\mathbf{2 3}),{ }^{15}$ prunin (24), ${ }^{16}$ narirutin (25), ${ }^{17}$ naringin (26), ${ }^{17}$ hesperidin (27), ${ }^{17}$ neohesperidin (28), ${ }^{18}$ eriocitrin (29), ${ }^{19}$ poncirin (30), ${ }^{20}$ hesperetin 7-O-(2",6"-di- $O$ - $\alpha$-rhamnopyranosyl)- $\beta$-glucopyranoside (31), ${ }^{17}$ naringenin $7-O-\left(2^{\prime \prime}, 6^{\prime \prime}-\mathrm{di}-O-\alpha\right.$-rhamnopyranosyl)- $\beta$-glucopyranoside (32). ${ }^{21}$ Their structure were identified by the analysis of their NMR spectral data, and then by comparison with those of literature listed below, respectively. Eleven compounds (8,10, 12-18, 24, 32) were the first isolation from C. sudachi.

## Isolation of Known Compounds

The EtOAc soluble fraction ( 54 g , the extraction and the partion were described in text) was subjected to silica gel column chromatography ( $1 \mathrm{~kg}, 11 \times 100 \mathrm{~cm}$ ). The column was eluted with solvents of increasing polarity (n-hexane-EtOAc, EtOAc, EtOAc-MeOH, MeOH) to give 15 major fractions (frs. 1-15). Fraction 1 ( 460 mg ) was separated on GPC $\left(\mathrm{CHCl}_{3}\right)$, $\mathrm{Si} \mathrm{HPLC}\left(\mathrm{CHCl}_{3}-\mathrm{MeOH}, 95: 5\right)$ to give $15(5 \mathrm{mg})$. Fraction 3 ( 477 mg ) was separated by $\mathrm{Si} \mathrm{HPLC}\left(\mathrm{CHCl}_{3}-\mathrm{MeOH}\right)$, $\mathrm{GPC}(\mathrm{MeOH})$ to give 12 (3 $\mathrm{mg}), \mathbf{1 3}(15 \mathrm{mg}), \mathbf{1 9}(6 \mathrm{mg}), \mathbf{2 0}(10 \mathrm{mg})$. Fraction $4(3.7 \mathrm{~g})$ was chromatographed on a silica gel column $\left(\mathrm{CHCl}_{3}-\mathrm{MeOH}\right)$ to give three fractions (frs 2.1-2.3) and $7(300 \mathrm{mg})$. Fraction 2.3 was separated by a Toyopearl HW-40 column ( $\mathrm{CHCl}_{3}-\mathrm{MeOH}, 1: 1$ ), Si

HPLC ( $\left.\mathrm{CHCl}_{3}-\mathrm{MeOH}\right)$, GPC (MeOH) to give $\mathbf{1 4}(7 \mathrm{mg}), \mathbf{1 6}(8 \mathrm{mg}), \mathbf{2 1}(5 \mathrm{mg})$. Fraction $5(1.1 \mathrm{~g})$ was subjected on $\mathrm{Si} \mathrm{HPLC}\left(\mathrm{CHCl}_{3}-\mathrm{MeOH}\right)$ to give $6(360 \mathrm{mg})$. Fraction $6(2.4$ g) was chromatographed on a Toyopearl HW-40 column $\left(\mathrm{CHCl}_{3}-\mathrm{MeOH}\right)$ to give four fractions (frs 6.1-6.4). Fraction 6.3 was separated by Si HPLC $\left(\mathrm{CHCl}_{3}-\mathrm{MeOH}\right)$, GPC $(\mathrm{MeOH})$ to give $\mathbf{8}(13 \mathrm{mg}), \mathbf{1 0}(44 \mathrm{mg})$. Fraction $7(19.0 \mathrm{~g})$ was chromatographed on a silica gel column $\left(\mathrm{CHCl}_{3}-\mathrm{MeOH}\right)$ to give seven fractions (frs 7.1-7.7). Fraction 7.3 was subjected on recrystallization $(\mathrm{MeOH})$; fraction 7.4 subjected on Si HPLC (hexaneEtOAc) to give $\mathbf{9}(1.0 \mathrm{~g}), \mathbf{1 1}(5 \mathrm{mg})$, respectively. Fraction $11(3.0 \mathrm{~g})$ was separated by a Sephadex LH-20 (MeOH) column, GPC ( MeOH ), ODS $\left(\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}\right)$ to give 24 (2 $\mathrm{mg})$. Fractions $12(1.9 \mathrm{~g})$ was recrystallized from MeOH to give $22(144 \mathrm{mg})$. Fraction 13 (1.9 g) was separated by a Sephadex LH-20 (MeOH) column, recrystallization $(\mathrm{MeOH}), \mathrm{GPC}(\mathrm{MeOH})$ to give $25(138 \mathrm{mg}), 29(60 \mathrm{mg})$. Fraction $14(8.3 \mathrm{~g})$ was separated by a Sephadex LH-20 (MeOH) column to give seven fractions (frs 14.1-14.7) and a mixture of 27, $\mathbf{2 8}(2.5 \mathrm{~g}, 3: 2)$. Fraction 14.7 was separated by GPC (MeOH), ODS ( $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}$ ) to give $27(193 \mathrm{mg}), \mathbf{3 0}(1 \mathrm{mg})$, a mixture of $\mathbf{2 5}, 26(85 \mathrm{mg}, 1: 1)$ and that of 26, 29 ( $63 \mathrm{mg}, ~ 2: 5$ ). Fraction $15(1.5 \mathrm{~g})$ was separated by a Sephadex LH-20 (MeOH) column, GPC ( MeOH ), ODS $\left(\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}\right)$ to give $17(63 \mathrm{mg}), \mathbf{1 8}(29 \mathrm{mg}), \mathbf{2 3}(13 \mathrm{mg})$, 31 ( 11 mg ).

One percent of $n-\mathrm{BuOH}$ soluble fraction $(3.8 \mathrm{~g})$ was chromatographed on a Sephadex LH-20 (MeOH) column chromatography to give five fractions (frs 1-5). Fraction 2 (1.3 g) was subjected on GPC ( MeOH ) to give $31(4 \mathrm{mg}), \mathbf{3 2}(21 \mathrm{mg})$. Fraction $3(460 \mathrm{mg})$ was separated by GPC $(\mathrm{MeOH})$, ODS $\left(\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}\right)$ to give a mixture of $\mathbf{2 5}, 26$ (104 $\mathrm{mg}, 1: 1)$ and that of $\mathbf{2 7}, \mathbf{2 8}(106 \mathrm{mg}, 1: 2)$. Fractions $4(155 \mathrm{mg})$ was subjected on GPC
$(\mathrm{MeOH})$ to give a mixture of $\mathbf{2 5}, \mathbf{2 6}(36 \mathrm{mg}, 3: 2)$ and that of $\mathbf{2 7}, \mathbf{2 8}(22 \mathrm{mg}, 1: 1)$, that of 26, 29 ( $37 \mathrm{mg}, 1: 3$ ).

## References and Notes

[1] Raymond, D.B.; Hasegawa, S.; Zareb, H. Phytochemistry 1989, 28, 2777-2781.
[2] Ming, K.; Gray A.I.; Waterman, P.G. Journal of Natural Products 1987, 50, 11601163.
[3] Bennet, R.D.; Hasegawa, S. Tetrahedron 1981, 37, 17-24.
[4] Ellis, B.E.; Amrhein, N. Phytochemistry 1971, 10, 3069-3072.
[5] Sakushima, A.; Coskun, M.; Maoka, T. Phytochemistry 1995, 40, 257-261..
[6] Demyttenaere, J.C.R.; Belleghem, K.V.; Kimpe, N.D. Phytochemistry 2001, 57, 199208.
[7] Delgado, G.; Rios, M.Y. Phytochemistry 1991, 30, 3129-3131.
[8] Fujita, M.; Inoue, T; Nagai, M. Yakugaku zasshi 1985, 105, 240-248.
[9] Ternai, B.; Markham, K.R. Tetrahedron 1976, 32, 565-569.
[10] Matsubara, Y.; Yusa, T.; Sawabe, A.; Iizuka, Y.; Takekuma, S.; Yoshida, Y. Agric. Biol. Chem. 1991, 55, 2923-2929.
[11] Matsumoto, T.; Shishido, A.; Takeya, K. Tennen Yuki Kagobutsu Toronkai Koen Yoshishiu 2001, 43th, 407-412.
[12] Greenham, J.; Vassiliades, D.D.; Harborne, J.B.; Williams, C.A.; Eagles, J.; Grayer, R.J.; Veitch, N.C. Phytochemistry 2001, 56, 87-91.
[13] Horie, T; Nakayama, M. Phytochemistry 1981, 20, 337-338.
[14] Martinez, V.; Barbera, O.; Parareda, J.S.; Marco, J.A. Phytochemistry 1987, 26,

2619-2624.
[15] Horie, T.; Tsukayama, M.; Yamada, T.; Miura, I.; Nakayama, M. Phytochemistry 1986, 25, 2621-2624.
[16] Lewinsohn, E.; Berman, E.; Mazur, Y.; Gressel, J. Phytochemistry 1986, 25, 25312535.
[17] Kumamoto, H.; Matsubara, Y.; Iizuka, Y.; Okamoto, K.; Yokoi, K. Nippon Nogeikagaku Kaishi 1985, 59, 683-687.
[18] Matsubara, Y.; Kumamoto, H.; Yonemoto, H.; Iizuka, Y.; Okamoto, K.; Yokoi, K. Nippon Nogeikagaku Kaishi 1985, 59, 405-410.
[19] Miyake, Y.; Yamamoto, K.; Osawa, T. Food Sci. Technol. Int. Tokyo 1997, 3, 8489.
[20] Kim, D.H.; Bae, E.A.; Han, M.J. Biol. Pharm. Bull. 1999, 22, 422-424.
[21] Kim, H.K.; Jeon, W.K.; Ko, B.S. Planta Medica 2001, 67, 548-549.

