Fate and transport of antimicrobials and antimicrobial resistance genes in soil

and runoff following land application of swine manure slurry

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This SI file includes:

17 pages, 8 tables, and 4 figures.

MATERIALS AND METHODS

Swine Manure Slurry

Manure slurry samples were collected from the sampling locations labeled in Figure S1. The wet weight and dry weight of the manure solids in the manure slurries from the finisher (BAC-manure), grower (CTC-manure), and sow and gilts (TYL-manure) were measured using gravimetric methods.

Field Site

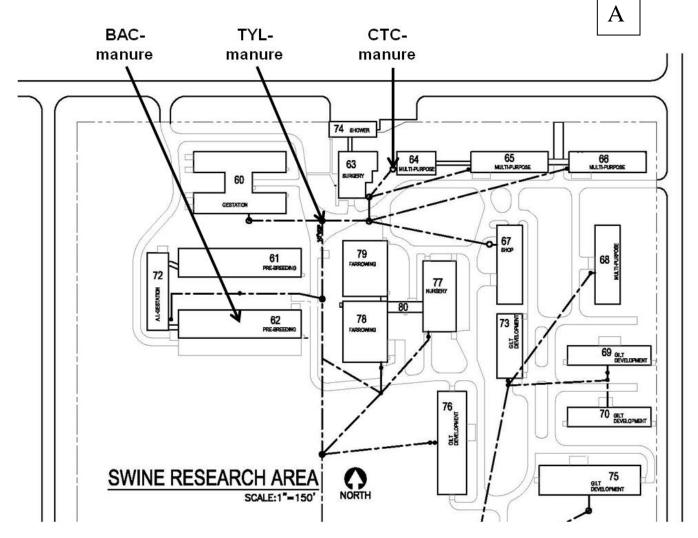
This field study was conducted in May and June 2011 at the University of Nebraska Rogers Memorial Farm located 18 km east of Lincoln, Nebraska. The study site had been cropped using a long-term no-till management system with controlled wheel traffic. Soybeans (*Glycine max*) were planted during the 2010 season and herbicide (glyphosate) was applied as needed to control weed growth. The soil at the site developed in loess under prairie vegetation, and is the Aksarben silty clay loam (fine, smectitic, mesic Typic Argiudoll) containing 15% sand, 57% silt, and 28% clay¹.

Soil samples for site characterization were obtained from the surface down to 2 cm just prior to manure application, and were air dried following collection. The study site had a mean slope gradient of 5.8%, an electrical conductivity (EC) of 0.38 dS m⁻¹ and a pH of 6.8. The organic matter and total carbon content of the soil was 4.7% and 2.62%, respectively. Mean measured concentrations of Bray and Kurtz No. 1 P, water-soluble P, NO₃-N, and NH₄-N were 43, 5.2, 8, and 4 mg kg⁻¹, respectively. The initial soil moisture condition prior to swine slurry application was not measured.

Chemicals

Standards for roxithromycin, doxycycline, bactracin A, and fenbendazole were purchased from Sigma-Aldrich (Fluka Chemicals). Oleandomycin, tylosin A and chlortetracycline were obtained from ThermoFisher Scientific (ICN Biomedicals and MP Biomedicals). Roxithromycin and doxycycline were used as internal standards and oleandomycin was used as a surrogate. Analytes were chlortetracycline, bactracin A, tylosin, and fenbendazole. Because bacitracin A is rapidly hydrolyzed in water at near neutral pH, a standard for bacitracin F (one of its degradation products) was synthesized and used to quantify this compound in the manure, soil, and runoff samples ².

FIGURES AND TABLES



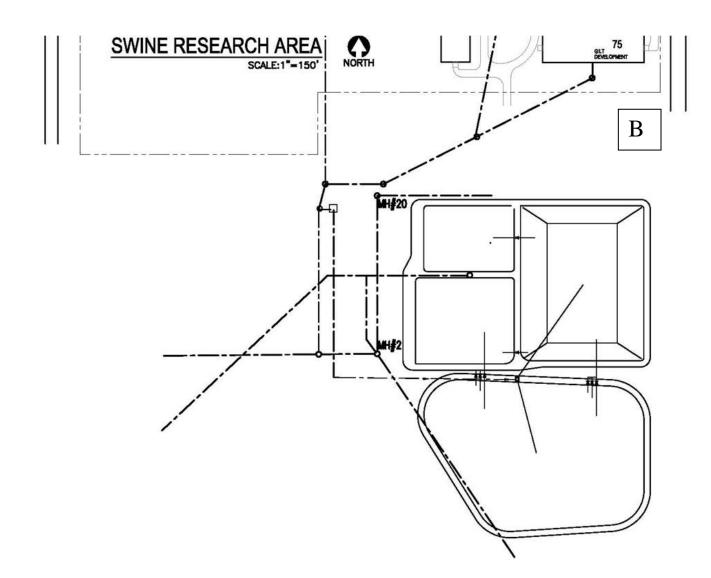


Figure S1. Map showing the locations of the barns and sampling points (A) and the lagoon systems (B) in the swine research area in the USDA Meat Animal Research Center.

Broadcast BAC	Broadcast CTC	Broadcast CONTROL	Broadcast TYL				
Incorporation	Incorporation	Incorporation	Incorporation	Injection	Injection	Injection	Injection
CONTROL	CTC	TYL	BAC	TYL	BAC	CONTROL	CTC
Injection	Injection	Injection	Injection	Incorporation	Incorporation	Incorporation	Incorporation
CTC	BAC	TYL	CONTROL	TYL	CTC	CONTROL	BAC
Broadcast	Broadcast	Broadcast	Broadcast	Incorporation	Incorporation	Incorporation	Incorporation
BAC	CTC	CONTROL	TYL	BAC	TYL	CTC	CONTROL
Broadcast	Broadcast	Broadcast	Broadcast	Injection	Injection	Injection	Injection
TYL	BAC	CONTROL	CTC	TYL	CONTROL	BAC	CTC

Figure S2. Randomized block design used in the field experiment. Plots in each row were constructed in the same week.

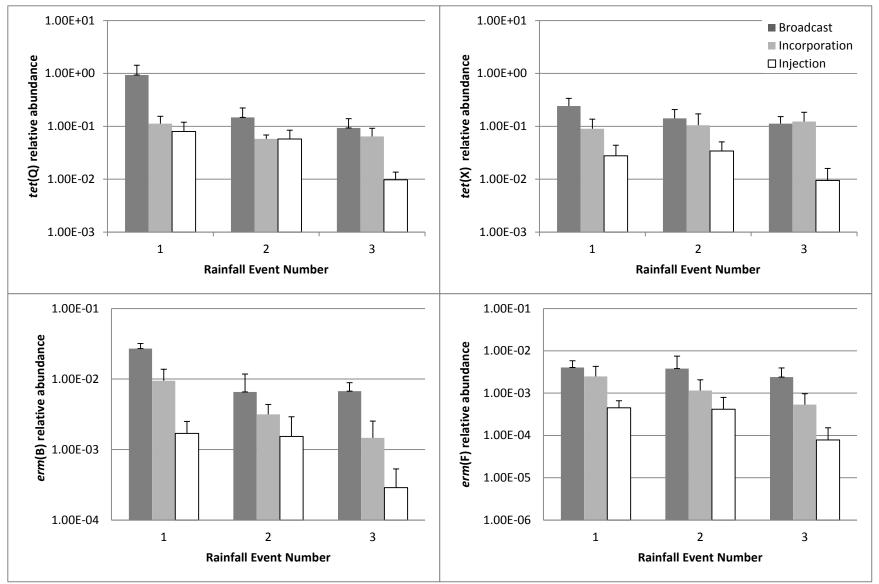


Figure S3. Relative abundance of tet(Q), tet(X), erm(B), and erm(F) in runoff from control and amended plots receiving broadcast, incorporation, or injection treatment. Error bars represent standard errors from triplicate field experiments.

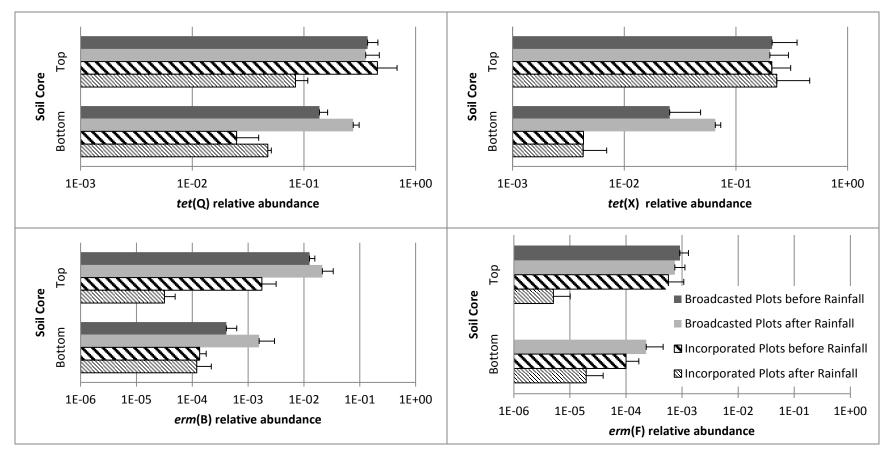


Figure S4. Relative abundance of tet(Q), tet(X), erm(B), and erm(F) in top and bottom soil in amended plots before and after three rainfall simulation tests. Error bars represent standard errors from triplicate field experiments.

Antimicrobial	Chemical Structure	Properties
Chlortetracycline	HCI	$K_d = 501 - 3715 \text{ L/kg}$
		Solubility = 500 mg/L $t_{1/2} = 21$ days ⁴
Tylosin		$K_d = 1,300 L/kg^5$
		Solubility = 6,000 mg/L
		$t_{1/2} = 6-8 \text{ days}^{-4, 6}$
	I''' O' OH O' Q	
Bacitracin		Environmental fate
(Bacitracin A)		data for Bacitracin A are not available in the literature
	I I NH2	

Table S1. Properties of the antimicrobials used in this study.

Analyte	Molecular weight	Retention time (min)	MRM Transition (m/z)
Bacitracin A	1422.7	9.82	712.10->86.20
Bacitracin F	1419.64	10.05	710.19->281.26
Chlortetracycline	478.88	8.71	478.90->444.00
Fenbendazole	299.35	10.63	300.20->268.20
Tylosin	916.10	10.40	916.9->174.2
Doxycycline (IS)	444.4	8.63	445.05->428.05
Oleandomycin (S)	687.86	10.51	688.35->544.10
Roxythromycin (IS)	837.05	11.58	837.55->679.50

Table S2. Molecular weight, retention times, and MRM transition of antimicrobials, internal standards (IS), and surrogate (S) compound.

Target ARG	Primer	Sequence (5'-3')	Annealing Temp (°C)	Linear Range (copies/20µL)	\mathbf{R}^2	Efficiency (%)	Reference
tet(Q) TetQ-FV	TetQ-FW	AGAATCTGCTGTTTGCCAGTG					
iei(Q)	TetQ-RV	CGGAGTGTCAATGATATTGCA	63	$10^2 - 10^9$	0.996	104.4	7
TetX-	TetX-FW	AGCCTTACCAATGGGTGTAAA					
tet(X)	TetX-RV	TTCTTACCTTGGACATCCCG	70	10^{1} - 10^{9}	0.997	81.8	8
<i>erm</i> (B) ErmB-FW	ErmB-FW	GGTTGCTCTTGCACACTCAAG					
erm(D)	ErmB-RV	CAGTTGACGATATTCTCGATTG	65	10^{1} - 10^{9}	0.978	111.1	9
erm(F)	ErmF-FW	TCTGGGAGGTTCCATTGTCC					
	ErmF-RV	TTCAGGGACAACTTCCAGC	65	10^{1} - 10^{9}	0.978	89.4	9
bceA*	BceA-FW	GCTACGACAGCACTTAATCA					
DUCA	BceA-RV	CACCTTCAGTTAGTCCATCA	55				10
bceR*	BceB-FW	TTAACCAACATCAACCTCAG					
Beek	BceB-RV	CCCCATTTGTATTGCCAT	55				10
bcrA	BcrA-FW	AAGTGGCAAGGCTTTTGAGA					
DUTA	BcrA-RV	CTCAGGATCAATCGGCAAAT	60				11
bcrB	BcrB-FW	AAGTGGCAAGGCTTTTGAGA					
UCID	BcrB-RV	AAATCACCGGGGGAATTAAG	60				11
harC	BcrC-FW	AAGTGGCAAGGCTTTTGAGA					
bcrC	BcrC-RV	CTCAAGTTCCCCAGTTTCCA	60				11

Table S3. Relevant information of the qPCR and PCR reactions used in this study.

* Regular PCR reactions.

	ARG	Relative Abundance
CTC Manuna	<i>tet</i> (Q)	1.33±0.26
CTC-Manure	tet(X)	0.078 ± 0.018
TVI Mamura	<i>erm</i> (B)	0.12 ± 0.024
TYL-Manure	<i>erm</i> (F)	0.0022 ± 0.0004

Table S4. Relative abundance (average \pm standard error) of ARGs tet(Q), tet(X), erm(B), and erm(F) in manure slurry.

	Rainfall	CTC	TYL	BAC
	Event	(ng/µL)	(ng/µL)	$(ng/\mu L)$
	1	<mdl< td=""><td>0.006^{*}</td><td><mdl< td=""></mdl<></td></mdl<>	0.006^{*}	<mdl< td=""></mdl<>
Broadcast	2	<mdl< td=""><td>0.011^{*}</td><td><mdl< td=""></mdl<></td></mdl<>	0.011^{*}	<mdl< td=""></mdl<>
	3	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>
	1	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>
Incorporation	2	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>
	3	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>
	1	<mdl< td=""><td>0.007^{a}</td><td><mdl< td=""></mdl<></td></mdl<>	0.007^{a}	<mdl< td=""></mdl<>
Injection	2	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>
	3	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>

Table S5. Aqueous antimicrobial concentrations in runoff from control plots (average \pm standard error). MDL was 0.005 ng/µL.

* Values are from one of the triplicate field experiments. No antimicrobials were detected in the other replicates.

		$tet(\mathbf{Q})$	tet(X)	<i>erm</i> (B)	<i>erm</i> (F)
		(copy/mL)	(copy/mL)	(copy/mL)	(copy/mL)
	Run 1	< MDL	< MDL	< MDL	< MDL
Broadcast	Run 2	423 ± 416	233 ± 231	25 ± 25	16 ± 16
	Run 3	< MDL	< MDL	< MDL	< MDL
	Run 1	< MDL	< MDL	< MDL	< MDL
Incorporation	Run 2	< MDL	< MDL	< MDL	< MDL
	Run 3	< MDL	< MDL	< MDL	< MDL
	Run 1	< MDL	< MDL	< MDL	< MDL
Injection	Run 2	< MDL	< MDL	< MDL	< MDL
	Run 3	< MDL	< MDL	< MDL	< MDL

Table S6. Concentrations of ARGs in runoff from control plots (average \pm standard error). Standard errors were calculated based on triplicate field experiments. The MDL for each ARG is reported in Table S2.

		Broadcast		Incorporation		
		Before Rainfalls	After Rainfalls	Before Rainfalls	After Rainfalls	
4-4(0)	Top soil	<mdl< td=""><td><mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>	
tet(Q)	Bottom soil	<mdl< td=""><td><mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>	
$4 - 4(\mathbf{V})$	Top soil	<mdl< td=""><td><mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>	
tet(X)	Bottom soil	<mdl< td=""><td><mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>	
	Top soil	<mdl< td=""><td><mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>	
<i>erm</i> (B)	Bottom soil	<mdl< td=""><td><mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>	
a(F)	Top soil	<mdl< td=""><td><mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>	
<i>erm</i> (F)	Bottom soil	<mdl< td=""><td><mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""><td><mdl< td=""></mdl<></td></mdl<></td></mdl<>	<mdl< td=""><td><mdl< td=""></mdl<></td></mdl<>	<mdl< td=""></mdl<>	

Table S7. No ARGs were detected in the top and bottom soil of control plots before and after the rainfall events in triplicate field experiments.

	Chlortetracyline	Tylosin (ng/g solid ww)	Bacitracin
CTC-manure	$3,324 \pm 1,560$	2 ± 1	172 ± 164
TYL-manure	102 ± 40	287 ± 124	7 ± 7
BAC-manure	16 ± 9	124 ± 68	777±753

Table S8. Antimicrobial concentrations in swine manure slurry (average \pm standard error). Standard errors were calculated based on five weekly manure slurry samples.

- 1. Kettler, T. A.; Doran, J. W.; Gilbert, T. L., Simplifed method for soil particle-size determination to accompany soil-quality analyses. *Soil Sci. Soc. Am. J.* **2001**, *65*, (3), 849-852.
- 2. Pavli, V.; Kmetec, V., Pathways of chemical degradation of polypeptide antibiotic bacitracin. *Biol. Pharm. Bull.* **2006**, *29*, (11), 2160-2167.
- 3. Teixido, M.; Granados, M.; Prat, M. D.; Beltran, J. L., Sorption of tetracyclines onto natural soils: data analysis and prediction. *Environmental Science and Pollution Research* **2012**, *19*, (8), 3087-3095.
- 4. Carlson, J. C.; Mabury, S. A., Dissipation kinetics and mobility of chlortetracycline, tylosin, and monensin in an agricultural soil in Northumberland County, Ontario, Canada. *Environ. Toxicol. Chem.* **2006**, *25*, (1), 1-10.
- 5. Clay, S. A.; Liu, Z.; Thaler, R.; Kennouche, H., Tylosin sorption to silty clay loam soils, swine manure, and sand. *Journal of Environmental Science and Health Part B-Pesticides Food Contaminants and Agricultural Wastes* **2005**, *40*, (6), 841-850.
- 6. Hu, D.; Coats, J. R., Aerobic degradation and photolysis of tylosin in water and soil. *Environ. Toxicol. Chem.* **2007**, *26*, (5), 884-889.
- Koike, S.; Krapac, I. G.; Oliver, H. D.; Yannarell, A. C.; Chee-Sanford, J. C.; Aminov, R. I.; Mackie, R. I., Monitoring and source tracking of tetracycline resistance genes in lagoons and groundwater adjacent to swine production facilities over a 3-year period. *Appl. Environ. Microbiol.* 2007, *73*, (15), 4813-4823.
- 8. Ghosh, S.; Ramsden, S. J.; LaPara, T. M., The role of anaerobic digestion in controlling the release of tetracycline resistance genes and class 1 integrons from municipal wastewater treatment plants. *Appl. Microbiol. Biotechnol.* **2009**, *84*, (4), 791-796.
- Koike, S.; Aminov, R. I.; Yannarell, A. C.; Gans, H. D.; Krapac, I. G.; Chee-Sanford, J. C.; Mackie, R. I., Molecular ecology of Macrolide-Lincosamide-Streptogramin B methylases in waste lagoons and subsurface waters associated with swine production. *Microb. Ecol.* 2010, *59*, (3), 487-498.
- Yoshida, Y.; Matsuo, M.; Oogai, Y.; Kato, F.; Nakamura, N.; Sugai, M.; Komatsuzawa, H., Bacitracin sensing and resistance in Staphylococcus aureus. *FEMS Microbiol. Lett.* 2011, 320, (1), 33-39.
- 11. Murphy, T.; Roy, I.; Harrop, A.; Dixon, K.; Keshavarz, T., Elicitation effects of oligosaccharides on the transcriptional level of bacitracin ABC transporter genes in Bacillus licheniformis. *Biotechnol. Lett.* **2008**, *30*, (9), 1665-1670.